THE EFFECT OF ABRUS PRECATORIUS METHANOL LEAVES EXTRACT IN INHIBITING GLUCOSE ABSORPTION IN IN VITRO STUDY

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Dedicated with love: To my beloved daddy; Ahmed Abdullah Al-moalemi To my adore mom; Najmah Omar To my lovely wife To my dear brothers and sisters To my two beautiful daughters; Rawa and Rafah

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ABSTRACT

Abrus precatorius leaves have been used traditionally for treatment of Type II diabetes in Malaysia. In this study, the potential of the 80% methanolic extract of Abrus *precatorius* leaves has been studied for its α -glucosidase inhibitory activity and its glucose diffusion effect. The extract of this plant at varying concentrations (6.25, 12.5, 25, 50 mg/ml) were tested using *in vitro* methods. An inhibitory α -glucosidase activity was determined spectrophotometrically in a 96-well microtiter plate. The results of α glucosidase inhibition assay revealed that Abrus precatorius leaves extract showed strong inhibition (65.4% and 84.6%) at the concentrations of 25 and 50 mg/ml, respectively, and represent less inhibition (25% and 28.2%) at the concentration of 6.25 and 12.5 mg/ml respectively when compared to control. Whereas, the effects of Abrus precatorius leaves extract on glucose movement across the sealed dialysis tube was measured spectrophotometrically using the glucose oxidase method. The results of inhibition of glucose diffusion in vitro assay indicated that, Abrus precatorius leaves extract shows a slight inhibitory effect on glucose diffusion at the concentrations of 12.5, 25 and 50 mg/ml after 16 hours, and the concentration of 50 mg/ml showed more effect after 24 hours. Nonetheless, the plant extract did not show any statistically significant effect on glucose movement after 0, 4, 8, 12, 16, 20 and 24 hours when compared to control. Therefore, it is considered that, the concentrations higher than 50 mg/ml may show a significant inhibitory effect on glucose diffusion. Overall, these results suggest that, Abrus precatorius leaves extract have significant inhibitory effects on glucose absorption due to the ability of plant to inhibit intestinal α -glucosidase enzyme.

ABSTRAK

Daun Abrus precatorius digunakan secara tradisional di Malaysia untuk merawat penyakit diabetes jenis II. Dalam kajian ini, 80% ekstrak metanol daun Abrus *precatorius* telah dikaji untuk potensinya bagi menghalang aktiviti enzim α -glucosidase dan kesan penyerapan glukosa. Pada kepekatan yang berbeza (6.25, 12.5, 25, 50 mg/ml), ekstrak tumbuhan ini telah diuji menggunakan kaedah in vitro. Spektrofotometrik telah digunakan untuk mengenalpasti aktiviti pembantutan aktiviti α-glucosidase pada plat 96microtiter. Hasil kajian aktiviti pambantutan α -glucosidase menunjukkan ekstrak Abrus precatorius memberi kesan yang kuat (65.4% dan 84.6%) pada kepekatan 25 dan 50 mg/ml masing-masing, dan kesan pembantutan berkurangan (25% dan 28.2%) pada kepekatan 6.25 dan 12.5 mg/ml masing-masing berbanding dengan kawalan. Manakala, kesan ekstrak Abrus precatorius pada pergerakan glukosa merentasi tiub dialisis telah diukur menggunakan alat spektrofotometrik dengan kaedah "glukosa oxidase". Hasil kesan pembantutan penyerapan glukosa *in vitro* menunjukkan kesan pembantutan yang kurang pada kepekatan 12.5, 25 dan 50 mg/ml selepas 16 jam dan kesan yang lebih tinggi telah ditunjukkan pada kepekatan 50 mg/ml selepas 24 jam. Walau bagaimanapun, ekstrak tumbuhan tidak menunjukkan sebarang kesan yang berbeza secara statistiknya pada pergerakan glukosa selepas 0, 4, 8, 12, 16, 20 dan 24 jam apabila dibandingkan dengan kawalan. Oleh itu, kepekatan yang lebih tinggi daripada 50 mg/ml mungkin menunjukkan kesan yang ketara pada tindak balas pembantutan penyerapan glukosa. Berdasarkan hasil kajian ini, ekstrak daun Abrus precatorius mempunyai kesan yang efektif terhadap pembantutan penyerapan glukosa disebabkan oleh tumbuhan ini mempunyai keupayaan untuk menghalang aktiviti enzim αglucosidase di dalam usus.

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LIST OF ABBREVIATION AND SYMBOLS

ADA American Diabetes Association _ IDDM -Insulin-dependent diabetes mellitus NIDDM Non-Insulin-dependent diabetes mellitus -GDM **Gestational Diabetes Mellitus** -HDL High-density lipoprotein _ Peroxisome proliferator-activated receptor gamma PPAR- γ -IRS1 Insulin receptor substrate 1 -SUR Sulphonylurea receptor SUR -DKA Diabetic ketoacidosis -Non-ketotic hyperosmolar NKHS _ SGLT 1 Na⁺/glucose co-transporter _ SGLT 2 Glucose transporter 2 -CNS Central nervous system - Na^+ Sodium ions -PNPG p-nitrophenyl-a-D-glucopyranoside -

World Health Organization

WHO

-

GOD	-	Glucose oxidase
POD	-	Peroxidase
NaCl	-	Sodium Chloride
Na2Co3	-	Sodium Carbonate
KH2PO4	-	Potassium dihydrogen phosphate
K2HPO4	-	Potassium hydrogen phosphate
STD	-	Standard
S	-	Sample
В	-	Blank
SD	-	Standard deviation
°C	-	Degree Celcius
mL	-	Millilitre
mM	-	Millimolar
μl	-	Microliter
g	-	Gram
mg	-	Milligram

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Diabetes mellitus is a commont chronic disease characterised by high blood glucose levels (Hyperglycemia) with disturbances of carbohydrate, protein, and fat metabolism (West, 2000). Approximately, 285 million adults worldwide suffered from diabetes mellitus in 2010. It is anticipated that the number will increase to 439 million by 2030 (Shaw *et al.*, 2010). The World Health Organization (WHO) reported that some 347 million people worldwide suffer from diabetes mellitus and more than 80% are from low-and middle income countries (WHO, 2013). Diabetes mellitus occurs when the pancreas fails to yield sufficient insulin, or when the body cannot use the insulin it produced efficiently (Association, 2010). The insulin hormone is responsible for decreasing the glucose levels in the blood circulation (Association, 2010).

The classification of this disease is dependent on the pathogenesis of the diabetes mellitus. The first classification of diabetes was published in 1997 by a team of experts from the American Diabetes Association. Their findings were

approved by the World Health Organization (WHO) in 1998 (Alberti *et al.*, 1998a). In 2003, it was updated and refined by the ADA and updated again by the WHO in 2006 to the current classification which includes Insulin-dependent diabetes mellitus (IDDM), Non-Insulin-dependent diabetes mellitus (NIDDM), Gestational Diabetes Mellitus (GDM), and other specific types of diabetes (Alberti and Zimmet, 1998; Genuth *et al.*, 2003; Organization, 2006). Among these types of diabetes mellitus, NIDDM (Type II) is a metabolic disorder characterised by hyperglycaemia, which often results from insulin resistance or β -cell dysfunction due to the interaction between environmental and genetic factors (Stumvoll *et al.*, 2005). Diabetes mellitus, NIDDM (Type II), has been recognized as a global health problem of the 21st century, and represents 90% of all cases of diabetes mellitus (Chen *et al.*, 2012; Scully, 2012).

The medical symptoms of diabetes mellitus are excessive thirst, hunger, muscular weakness, weight loss, and continuous urination. These symptoms were first recorded by the ancient Egyptians in the Ebers papyrus some 3500 years ago, and by the Greek physician Aretaens the Cappadocian (A.D 30-90) and Galen (A.D 130-200) (Farnsworth and Seligman, 1971). An increase in blood glucose levels for a sustained periods can lead to several complications such as kidney failure, cardiovascular diseases, hepatic, nerve damage, and blindness (Chen, *et al.*, 2012; Hussain *et al.*, 2008). Diagnosis of diabetes is dependent on the demonstration of increased concentrations of glucose in the blood. This can be determined through the use of the oral glucose tolerance test which has been used as a diagnostic criteria (Association, 2008).

Treatment of diabetes mellitus aims to reduce levels of blood glucose to normal or close to normal in the patients. One of the most effective ways to control Type II diabetes is to reduce glucose absorption through inhibiting carbohydrate digestion, which is an important source for glucose in the human body. Digestion of carbohydrate occurs in the gastrointestinal tract, and requires two enzymes, pancreatic α -amylase and intestinal α -glucosidases to release absorbable glucose (Jones *et al.*, 2011).

The function of α -glucosidases enzymes is by breaking down the disaccharides that come from food or produced through digestion of carbohydrate by α -amylases to the monosaccharides (glucose) (Lieberman and Marks, 2009). The inhibition of α -glucosidase can help delay the digestion of carbohydrates in the intestine thereby better managing the Type II diabetes (Heacock *et al.*, 2005). Current medications of α -glucosidase inhibitors are comparatively expensive and have toxic side-effects such as acute hypoglycemia at higher doses, liver problems, diarrhea, nausea, lactic acidosis, and weight gain (Avery *et al.*, 2008; Bastaki, 2005). This has encoraged people to use plants and herbs as alternative source of α -glucosidase inhibitors because it is more acceptable and cheaper (Kee *et al.*, 2013).

There are many dietary supplements of plant origins, which have been studied and have therapeutic potential agents for the treatment of diabetes and its complications (Mohamed *et al.*, 2012). One of the local herbs in Malaysia, locally called Akar Saga, scientifically known as *Abrus precatorius*, is used as an antidiabetic remedy (Monago and Alumanah, 2005, Bhatia *et al.*, 2013). *Abrus precatorius* is a common herb belonging to the *Fabaceae* family and is widely available in the forests of Malaysia, Indonesia, India, and Nigeria to name a few (Monago and Alumanah, 2005, Bhatia *et al.*, 2013). Several studies have reported that *Abrus precatorius* has a wide range of medical effects (Bhatia *et al.*, 2013).

In this study, the potential of the 80% methanolic extract of *Abrus precatorius* leaves has been studied for its α -glucosidase inhibition and its glucose diffusion effect. The extract of these leaves at different concentrations were tested using *in vitro* methods. Inhibitory α -glucosidase activities have been determined spectrophotometrically in a 96-well Microtiter plate. Whereas, the effects of *Abrus precatorius* leaves extract on glucose movement across the sealed dialysis tube have been measured spectrophotometrically using the glucose oxidase assay.

1.2 Problem statement

Alpha-glucosidase is involved in the breakdown of complex carbohydrate molecules to glucose in the small intestine before they can be absorbed into the bloodstream. Patients with Type II diabetes suffer from an increase in blood glucose levels after eating complex carbohydrates. The inhibition of the glucose absorption through inhibiting α -glucosidase or inhibiting glucose transport would help to reduce the impact of rising postprandial blood glucose and better manage Type II diabetes. In the current study, the potential of the *Abrus precatorius* plant methanolic extract has been studied for its α -glucosidase inhibiting and its effect on glucose diffusion.

1.3 Objectives of the study

- i. To study the potential of *Abrus precatorius* plant methanolic extract as α -glucosidase inhibitors.
- ii. To investigate the glucose entrapment study by using *Abrus precatorius* plant methanolic extract.

1.4 Significance of the study

In this study, the potential of *Abrus precatorius* leaves methanolic extract to inhibit α -glucosidase and glucose diffusion has been studied *in vitro* by using α -

glucosidase inhibitor assay and the glucose entrapment assay. This helps to provide a natural source of the glucose absorption inhibitors that will eventually have an impact on the treatment and management of Type II diabetes.

1.5 Scope of the study

The *Abrus precatorius* leaves methanolic extract at different concentrations were tested using *in vitro* methods for their anti-diabetic effects on α -glucosidase inhibition and glucose diffusion. Inhibitory α -glucosidase activities have been determined spectrophotometrically in a 96-well Microtiter plate. Whereas, the effects of *Abrus precatorius* leaves extract on glucose movement across the sealed dialysis tube have been measured spectrophotometrically using the glucose oxidase method.

REFERENCES

- Abo, K., Fred-Jaiyesimi, A. and Jaiyesimi, A. (2008). Ethnobotanical studies of medicinal plants used in the management of diabetes mellitus in South Western Nigeria. *Journal of ethnopharmacology*, 115(1), 67-71.
- Adelowotan, O., Aibinu, I., Adenipekun, E. and Odugbemi, T. (2008). The in-vitro antimicrobial activity of Abrus precatorius (L) fabaceae extract on some clinical pathogens. *The Nigerian postgraduate medical journal*, 15(1), 32-37.
- Ahmad, M., Qureshi, R., Arshad, M., Khan, M. A. and Zafar, M. (2009). Traditional herbal remedies used for the treatment of diabetes from district Attock (Pakistan). *Pakistan Journal of Botany*, 41(6), 2777-2782.
- Alagesan, K., Thennarasu, P., Kumar, V., Sankarnarayanan, S. and Balsamy, T. (2012). Identification of α-glucosidase inhibitors from Psidium guajava leaves and Syzygium cumini Linn. seeds. *International Journal of Pharma Sciences and Research*, 3(2), 316-322.
- Alberti, K., Davidson, M. B., DeFronzo, R. A., Drash, A., Genuth, S., Harris, M. I., et al. (1998). Report of the expert committee on the diagnosis and classification of diabetes mellitus. *Diabetes care*, 21, S5-S19.
- Alberti, K. and Zimmet, P. (1998). Report of a WHO Consultation. Definition, Diagnosis and Classification of Diabetes Mellitus and its Complications Part 1: Diagnosis and Classification of Diabetes Mellitus. World Health Organization Department of Noncommunicable Disease Surveillance Geneva. Diabet Med, 15(7), 539-553.

- Association, A. D. (2008). Diagnosis and classification of diabetes mellitus. *Diabetes care*, 31(Supplement 1), S55-S60.
- Association, A. D. (2010). Diagnosis and classification of diabetes mellitus. *Diabetes care*, 33(Supplement 1), S62-S69.
- Avery, M. A., Mizuno, C. S., Chittiboyina, A. G., Kurtz, T. W. and Pershadsingh, H.
 A. (2008). Type 2 diabetes and oral antihyperglycemic drugs. *Current medicinal chemistry*, 15(1), 61-74.
- Balandrin, M. F., Klocke, J. A., Wurtele, E. S. and Bollinger, W. H. (1985). Natural plant chemicals: sources of industrial and medicinal materials. *Science*, 228(4704), 1154-1160.
- Balfour, J. A. and McTavish, D. (1993). Acarbose. Drugs, 46(6), 1025-1054.
- Basha, S. K. and Kumari, V. S. (2012). In vitro antidiabetic activity of psidium guajava leaves extracts. *Asian Pacific Journal of Tropical Disease*, 2, S98-S100.
- Bastaki, A. (2005). Diabetes mellitus and its treatment. International Journal of Diabetes and Metabolism, 13(3), 111-134.
- Bhatia, M., Siddiqui, N. and Gupta, S. (2013). Abrus Precatorius (L.): An Evaluation of Traditional Herb. *Journal of Pharmaceutical Research*, 3(4).
- Bhutia, S. K., Mallick, S. K., Maiti, S. and Maiti, T. K. (2009). Inhibitory effect of Abrus abrin-derived peptide fraction against Dalton's lymphoma ascites model. *Phytomedicine*, 16(4), 377-385.
- Bnouham, M., Ziyyat, A., Mekhfi, H., Tahri, A. and Legssyer, A. (2006). Medicinal plants with potential antidiabetic activity-A review of ten years of herbal medicine research (1990-2000). *International Journal of Diabetes and Metabolism*, 14(1), 1-25.

Bolen, S., Feldman, L., Vassy, J., Wilson, L., Yeh, H.-C., Marinopoulos, S., et al. (2007). Systematic review: comparative effectiveness and safety of oral medications for type 2 diabetes mellitus. *Annals of internal medicine*, 147(6), 386-399.

Capasso, F. (2003). Phytotherapy: a quick reference to herbal medicine: Springer.

- Catalano, P. M., McIntyre, H. D., Cruickshank, J. K., McCance, D. R., Dyer, A. R., Metzger, B. E., et al. (2012). The Hyperglycemia and Adverse Pregnancy Outcome Study Associations of GDM and obesity with pregnancy outcomes. *Diabetes care*, 35(4), 780-786.
- Chen, L., Magliano, D. J. and Zimmet, P. Z. (2012). The worldwide epidemiology of type 2 diabetes mellitus—present and future perspectives. *Nature Reviews Endocrinology*, 8(4), 228-236.
- Chistokhodova, N., Nguyen, C., Calvino, T., Kachirskaia, I., Cunningham, G. and Howard Miles, D. (2002). Antithrombin activity of medicinal plants from central Florida. *Journal of ethnopharmacology*, 81(2), 277-280.
- Choi, Y.-H., Kinghorn, A. D., Shi, X., Zhang, H. and Teo, B. K. (1989). AbrusosideA: a new type of highly sweet triterpene glycoside. *Journal of the Chemical Society, Chemical Communications*, (13), 887-888.
- Da Silva Pinto, M. and Shetty, K. (2010). Health Benefits of Berries for Potential Management of Hyperglycemia and Hypertension. Proceedings of the 2010 ACS symposium series, 121-137.

Das, D. (1978). Biochemistry: Academic Publishers.

DeFronzo, R. A. (2004). Pathogenesis of type 2 diabetes mellitus. *Medical Clinics of North America*, 88(4), 787-835.

- DeFronzo, R. A. and Ferrannini, E. (1991). Insulin resistance: a multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes care*, 14(3), 173-194.
- Dehghan-Kooshkghazi, M. and Mathers, J. C. (2004). Starch digestion, large-bowel fermentation and intestinal mucosal cell proliferation in rats treated with the α -glucosidase inhibitor acarbose. *British journal of nutrition*, 91(03), 357-365.
- Derosa, G. and Maffioli, P. (2012). Efficacy and safety profile evaluation of acarbose alone and in association with other antidiabetic drugs: a systematic review. *Clinical therapeutics*, 34(6), 1221-1236.
- Deshpande, A. D., Harris-Hayes, M. and Schootman, M. (2008). Epidemiology of diabetes and diabetes-related complications. *Physical therapy*, 88(11), 1254-1264.
- Edwards, C., Blackburn, N., Craigen, L., Davison, P., Tomlin, J., Sugden, K., et al. (1987). Viscosity of food gums determined in vitro related to their hypoglycemic actions. *The American journal of clinical nutrition*, 46(1), 72-77.
- Edwards, C., Johnson, I. and Read, N. (1988). Do viscous polysaccharides slow absorption by inhibiting diffusion or convection? *European Journal of Clinical Nutrition*, 42(4), 307-312.
- El-Kaissi, S. and Sherbeeni, S. (2011). Pharmacological management of type 2 diabetes mellitus: an update. *Current diabetes reviews*, 7(6), 392-405.
- Farnsworth, N. and Seligman, A. (1971). Hypoglycemic plants. *Tile and Till*, 57(3), 52-56.
- Gallagher, A., Flatt, P., Duffy, G. and Abdel-Wahab, Y. (2003). The effects of traditional antidiabetic plants on in vitro glucose diffusion. *Nutrition research*, 23(3), 413-424.

- Ganong, W. F. and Barrett, K. E. (2005). *Review of medical physiology* (Vol. 21): McGraw-Hill Medical New York:.
- Gbolade, A. A. (2009). Inventory of antidiabetic plants in selected districts of Lagos State, Nigeria. *Journal of ethnopharmacology*, 121(1), 135-139.
- Genuth, S., Alberti, K., Bennett, P., Buse, J., Defronzo, R., Kahn, R., et al. (2003). Follow-up report on the diagnosis of diabetes mellitus. *Diabetes care*, 26(11), 3160.
- Gerich, J. E. (2003). Clinical significance, pathogenesis, and management of postprandial hyperglycemia. *Archives of internal medicine*, 163(11), 1306-1316.
- Gupta, R., Gigras, P., Mohapatra, H., Goswami, V. K. and Chauhan, B. (2003). Microbial α-amylases: a biotechnological perspective. *Process Biochemistry*, 38(11), 1599-1616.
- Hanefeld, M. and Schaper, F. (2007). The role of alpha-glucosidase inhibitors (acarbose) *Pharmacotherapy of Diabetes: New Developments* (pp. 143-152): Springer.
- Heacock, P. M., Hertzler, S. R., Williams, J. A. and Wolf, B. W. (2005). Effects of a medical food containing an herbal α-glucosidase inhibitor on postprandial glycemia and insulinemia in healthy adults. *Journal of the American Dietetic Association*, 105(1), 65-71.
- Hui, A. L., Sevenhuysen, G., Harvey, D. and Salamon, E. (2014). Food Choice Decision-Making by Women with Gestational Diabetes. *Canadian Journal of Diabetes*, 38(1), 26-31.
- Hussain, F., Sheikh, M. A., Arif, M., Arshad, A. and Jamil, A. (2008). Nonenzymatic glycation in diabetic complications. *Saudi medical journal*, 29(2), 303-304.

- Hussin, A. H. (2001). Adverse effects of herbs and drug-herbal interactions. Malaysian Journal of Pharmacy, 1(2), 39-44.
- Jadhav, R. and Puchchakayala, G. (2012). Hypoglycemic and antidiabetic activity of flavonoids: boswellic acid, ellagic acid, quercetin, rutin on streptozotocinnicotinamide induced type 2 diabetic rats. *International Journal of Pharmacy & Pharmaceutical Sciences*, 4(2).
- Jain, S. and Saraf, S. (2010). Type 2 diabetes mellitus—its global prevalence and therapeutic strategies. *Diabetes & Metabolic Syndrome: Clinical Research & Reviews*, 4(1), 48-56.
- Jones, K., Sim, L., Mohan, S., Kumarasamy, J., Liu, H., Avery, S., et al. (2011). Mapping the intestinal alpha-glucogenic enzyme specificities of starch digesting maltase-glucoamylase and sucrase-isomaltase. *Bioorganic & medicinal chemistry*, 19(13), 3929-3934.
- Kamboj, V. (2000). Herbal medicine. Current science-bangalore, 78(1), 35-38.
- Kee, K. T., Koh, M., Oong, L. X. and Ng, K. (2013). Screening culinary herbs for antioxidant and α-glucosidase inhibitory activities. *International Journal of Food Science & Technology*, 48(9), 1884-1891.
- Kim, J.-S., Kwon, C.-S. and Son, K. H. (2000). Inhibition of alpha-glucosidase and amylase by luteolin, a flavonoid. *Bioscience, biotechnology, and biochemistry*, 64(11), 2458-2461.
- Krall, L. P., Beaser, R. S. and Center, J. D. (1989). Joslin Diabetes Manual: Lea & Febiger.
- Kumar, A., Ilavarasan, R., Jayachandran, T., Deecaraman, M., Kumar, R. M., Aravindan, P., et al. (2008). Anti-inflammatory activity of Syzygium cumini seed. *African Journal of Biotechnology*, 7(8).

- Lebovitz, H. (1998). Postprandial hyperglycaemic state: importance and consequences. *Diabetes research and clinical practice*, 40, S27-28.
- Lefèbvre, P. and Silink, M. (2006). Diabetes fights for recognition. *The Lancet*, 368(9548), 1625-1626.
- Lenhard, J. M. and Gottschalk, W. K. (2002). Preclinical developments in type 2 diabetes. Advanced drug delivery reviews, 54(9), 1199-1212.
- Lieberman, M. and Marks, A. D. (2009). *Marks' basic medical biochemistry: a clinical approach*: Lippincott Williams & Wilkins.
- Lodish, H., Berk, A., Zipursky, S. L., Matsudaira, P., Baltimore, D. and Darnell, J. (2000). Transport across Epithelia.
- Loh, S. P. and Hadira, O. (2011). In vitro inhibitory potential of selected Malaysian plants against key enzymes involved in hyperglycemia and hypertension. *Mal J Nutr*, 17(1), 77-86.
- Malviya, N., Jain, S. and Malviya, S. (2010). Antidiabetic potential of medicinal plants. *Acta Pol Pharm*, 67(2), 113-118.
- Martini, F., Ober, W. C., Garrison, C. W. and Welch, K. (1998). Fundamentals of anatomy and physiology (Vol. 4): Prentice Hall New Jersey.
- Ménan, H., Banzouzi, J.-T., Hocquette, A., Pélissier, Y., Blache, Y., Koné, M., et al. (2006). Antiplasmodial activity and cytotoxicity of plants used in West African traditional medicine for the treatment of malaria. *Journal of ethnopharmacology*, 105(1), 131-136.
- Mkhombo, M. (2010). The effects of clausena anisata (WILLD) hook (RUTACEAE) leaf extracts on selected diabetic related carbohydrate metabolizing enzymes. University of Limpopo.

- Mohamed, E. A. H., Siddiqui, M. J. A., Ang, L. F., Sadikun, A., Chan, S. H., Tan, S. C., et al. (2012). Potent α-glucosidase and α-amylase inhibitory activities of standardized 50% ethanolic extracts and sinensetin from Orthosiphon stamineus Benth as anti-diabetic mechanism. *BMC complementary and alternative medicine*, 12(1), 176.
- Mohan, V. and Janardhanan, K. (1995). Chemical determination of nutritional and anti-nutritional properties in tribal pulses. *J. Food Sci. Technol*, 32.
- Mokshagundam, S. P. L. and Broadstone, V. L. (2007). Diabetes Mellitus *Primary Care in Obstetrics and Gynecology* (pp. 309-325): Springer.
- Mølgaard, P., Nielsen, S. B., Rasmussen, D. E., Drummond, R. B., Makaza, N. and Andreassen, J. (2001). Anthelmintic screening of Zimbabwean plants traditionally used against schistosomiasis. *Journal of ethnopharmacology*, 74(3), 257-264.
- Monago, C. and Alumanah, E. (2005). Antidiabetic effect of chloroform-methanol extract of Abrus precatorius linn seed in alloxan diabetic rabbit. J Appl Sci Environ Mgt, 9(1), 85-88.
- Neal, M. J. (2012). *Medical pharmacology at a glance* (Vol. 70): John Wiley & Sons.

Ogbonnia, S. and Anyakora, C. (2009). Chemistry and Biological Evaluation of Nigerian Plants with Anti-Diabetic Properties. Proceedings of the 2009 ACS symposium series, 185-207.

- Organization, W. H. (2006). International Diabetes Federation. Definition and diagnosis of diabetes mellitus and intermediate hyperglycemia: report of a WHO/IDF consultation. *World Health Organization*, 21.
- Ortiz-Andrade, R., Garcia-Jimenez, S., Castillo-Espana, P., Ramirez-Avila, G., Villalobos-Molina, R. and Estrada-Soto, S. (2007). α-Glucosidase inhibitory

activity of the methanolic extract from Tournefortia hartwegiana: An antihyperglycemic agent. *Journal of ethnopharmacology*, 109(1), 48-53.

- Pal, R. S., Ariharasivakumar, G., Girhepunje, K. and Upadhyay, A. (2009). In-vitro antioxidative activity of phenolic and flavonoids compounds extracted from seeds of Abrus precatorius. *International Journal of Pharmacy and Pharmaceutical Sciences*, 1(2), 136-140.
- Palanuvej, C., Hokputsa, S., Tunsaringkarn, T. and Ruangrungsi, N. (2009). In vitro glucose entrapment and alpha-glucosidase inhibition of mucilaginous substances from selected Thai medicinal plants. *Sci Pharm*, 77, 837-849.
- Park, M.-H., Ju, J.-W., Park, M. and Han, J. (2013). Daidzein inhibits carbohydrate digestive enzymes in vitro and alleviates postprandial hyperglycemia in diabetic mice. *European journal of pharmacology*, 712(1), 48-52.
- Peet, A. (2012). *Marks' Basic Medical Biochemistry*: Lippincott Williams & Wilkins.
- Pistia-Brueggeman, G. and Hollingsworth, R. I. (2001). A preparation and screening strategy for glycosidase inhibitors. *Tetrahedron*, 57(42), 8773-8778.
- Pistia-Brueggeman, G. and Hollingsworth, R. I. (2003). The use of the o-nitrophenyl group as a protecting/activating group for 2-acetamido-2-deoxyglucose. *Carbohydrate research*, 338(5), 455-458.
- Preedy, V. R. (2012). Dietary Sugars: Chemistry, Analysis, Function and Effects: Royal Society of Chemistry.
- Qatanani, M. and Lazar, M. A. (2007). Mechanisms of obesity-associated insulin resistance: many choices on the menu. *Genes & development*, 21(12), 1443-1455.

- Qujeq, D. and Babazadeh, A. (2013). The Entrapment Ability of Aqueous and Ethanolic Extract of Teucrium Polium: Glucose Diffusion into the External Solution. *International journal of molecular and cellular medicine*, 2(2), 93.
- Rai, M., Cordell, G. A., Martinez, J. L., Marinoff, M. and Rastrelli, L. (2012). *Medicinal plants: biodiversity and drugs*: CRC Press.
- Rajaram, N. and Janardhanan, K. (1992). The chemical composition and nutritional potential of the tribal pulse, Abrus precatorius L. *Plant foods for human nutrition*, 42(4), 285-290.
- Rege, A. and Chowdhary, A. S. (2014). Evaluation of Alpha-Amylase and Alpha-Glucosidase Inhibitory Activities of Ocimum sanctum Linn. *International Journal* of Pharmaceutical Sciences Review & Research, 25(1).
- Russell, W. and Duthie, G. (2011). Plant secondary metabolites and gut health: the case for phenolic acids. *Proceedings of the Nutrition Society*, 70(03), 389-396.
- Sarker, S. D., Latif, Z. and Gray, A. I. (2006). Natural products isolation.

Scully, T. (2012). Diabetes in numbers. *Nature*, 485(7398), S2-S3.

- Shaw, J., Sicree, R. and Zimmet, P. (2010). Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes research and clinical practice*, 87(1), 4-14.
- Sheehan, M. T. (2003). Current therapeutic options in type 2 diabetes mellitus: a practical approach. *Clinical medicine & research*, 1(3), 189-200.
- Shim, Y.-J., Doo, H.-K., Ahn, S.-Y., Kim, Y.-S., Seong, J.-K., Park, I.-S., et al. (2003). Inhibitory effect of aqueous extract from the gall of Rhus chinensis on alpha-glucosidase activity and postprandial blood glucose. *Journal of ethnopharmacology*, 85(2), 283-287.

- Sohn, S.-H., Lee, E.-Y., Lee, J.-H., Kim, Y., Shin, M., Hong, M., et al. (2009). Screening of herbal medicines for recovery of acetaminophen-induced nephrotoxicity. *Environmental toxicology and pharmacology*, 27(2), 225-230.
- Soro, R. Y., Diopoh, J. K., Willemot, R.-M. and Combes, D. (2007). Enzymatic synthesis of polyglucosylfructosides from sucrose alone by a novel α-glucosidase isolated from the digestive juice of Archachatina ventricosa (Achatinideae). *Enzyme and microbial technology*, 42(1), 44-51.
- Stumvoll, M., Goldstein, B. J. and van Haeften, T. W. (2005). Type 2 diabetes: principles of pathogenesis and therapy. *The Lancet*, 365(9467), 1333-1346.
- Subramanian, R., Asmawi, M. Z. and Sadikun, A. (2008). In vitro alpha-glucosidase and alpha-amylase enzyme inhibitory effects of Andrographis paniculata extract and andrographolide. *Acta Biochim Pol*, 55(2), 391-398.
- Tripathi, B. K. and Srivastava, A. K. (2006). Diabetes mellitus: complications and therapeutics. *Med Sci Monit*, 12(7), 130-147.
- Van Tilburg, J., Van Haeften, T. W., Pearson, P. and Wijmenga, C. (2001). Defining the genetic contribution of type 2 diabetes mellitus. *Journal of medical genetics*, 38(9), 569-578.
- von Reis, A. S. (1977). Exploring the herbarium. *Scientific American*, 236(5), 96-104.
- Wah, L. E. (2001). Lqsg Chemistry O Level 2e. *Pearson Education South Asia*, 2nd edition, 108.
- Wambebe, C. and Amosun, S. L. (1984). Some neuromuscular effects of the crude extracts of the leaves of abrus precatorius. *Journal of ethnopharmacology*, 11(1), 49-58.

West, I. (2000). Radicals and oxidative stress in diabetes. *Diabetic Medicine*, 17(3), 171-180.

WHO. (2013). World Diabetes Day 2013. WHO.

Zajac, J., Shrestha, A., Patel, P. and Poretsky, L. (2010). The main events in the history of diabetes mellitus *Principles of Diabetes Mellitus* (pp. 3-16): Springer.