

EVALUATION OF *Acalypha indica* EXTRACTS FOR  
ANTIOXIDANT AND ANTIBACTERIAL ACTIVITIES

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**DEDICATION**

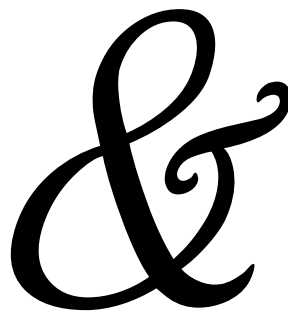
To

**ADABI**

*As the main sponsor of UTM-ADABI Cat Food project*

**MINISTRY OF HIGHER EDUCATION MALAYSIA and MARA**

*for providing scholarship and financial support for my study*



**FAMILY and FRIENDS**

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## ABSTRACT

For centuries, medicinal plants are being used as remedy for various ailments throughout the globe. The study was conducted emphasizing on the antibacterial and antioxidant activities of several *Acalypha indica* extracts. The plant was divided to leaves and stem, whole plant and roots and extracted with hexane, methanol and ethanol by successive method. Antioxidant activity was measured by 2,2-diphenyl-1-picrylhydrazyl (DPPH) Radical Scavenging assay and found to be highest in the ethanolic root extract with IC<sub>50</sub> of 206 µg/ml. The antibacterial activity screening of different extracts was conducted by using disc diffusion, minimum inhibition concentration (MIC) and minimum bactericidal concentration (MBC) against *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Enterococcus faecalis*. Hexane extracts from leaves and stem, whole plant and roots showed promising results against *Enterococcus faecalis* with high inhibition zone at 10 to 12 mm as compared to standard antibiotics, 6 to 10 mm. All extracts showed antibacterial activity with minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) values in the range from 60 to 15 mg/ml. This study concludes that *A. indica* explicit antioxidant and antibacterial activities may be potential for pharmaceuticals, cosmeceuticals, nutraceuticals, medical and food industry.

## ABSTRAK

Selama berabad, herba telah digunakan sebagai penawar pelbagai penyakit di seluruh dunia. Kajian ini dijalankan dengan menekankan aktiviti antioksidan dan antimikrob oleh beberapa ekstrak *Acalypha indica*. Tumbuhan tersebut dibahagikan kepada beberapa bahagian iaitu batang dan daun, keseluruhan pokok dan akar. Aktiviti antioksidan diukur menggunakan kaedah pemerangkapan radikal bebas 2,2-diphenyl-1-picrylhydrazyl (DPPH) dan menunjukkan ekstrak akar menggunakan ethanol mempunyai aktiviti antioksidan tertinggi dengan  $IC_{50}$  206 ug/ml. Penilaian aktiviti antimikrobial oleh pelbagai ekstrak dijalankan dengan cakera resapan, kepekatan perencatan minimum (MIC) dan kepekatan bakterisidal minimum (MBC) merencat pertumbuhan *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa* dan *Enterococcus faecalis*. Ekstrak menggunakan pelarut heksana menunjukkan hasil yang memberansangkan menentang *Enterococcus faecalis* dengan zon perencatan yang tinggi (10 hingga 12 mm) berbanding antibiotik standard (6 hingga 10 mm). Semua ekstrak menunjukkan nilai kepekatan perencatan minimum (MIC) dan kepekatan bakterisidal minimum (MBC) sekitar 60 hingga 15 mg/ml. Kesimpulannya, *A. indica* mempamerkan potensi aktiviti antioksidan dan anti-mikrob dan sesuai untuk penggunaan dalam industri farmaseutikal, nutraseutikal, kosmeseutikal, perubatan dan makanan.

## TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	x
	LIST OF FIGURES	xi
	LIST OF ABBREVIATIONS	xii
<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
	1.1 General Introduction	1
	1.2 Problem Statement	2
	1.3 Research Objectives	3
	1.4 Research Scope	3
	1.5 Significance of Research	4

<b>2</b>	<b>LITERATURE REVIEW</b>	<b>5</b>
2.1	Natural Product Approach in Drug Discovery	5
2.1.1	An Outlook on Plant Derived Drugs	6
2.1.2	Natural Product Trending	7
2.2	<i>Acalypha indica</i> , herb in grasp	8
2.2.1	Origin, Taxonomy and Morphology	8
2.2.2	Traditional Uses	10
2.2.3	Bioactive Compounds and its Applications	11
2.2.4	Potential use of <i>A. indica</i> Extracts in Industry	20
2.3	The Ground Knowledge of Extraction Methods	21
2.3.1	Synergism between Extraction Solvent and Bioactive Compounds	23
2.4	The science behind antioxidants from herbs	24
2.4.1	Natural Antioxidants: An Overview	25
2.5	Exploring Antimicrobial Properties of Natural Products	28
2.5.1	Targeted Microbes	29
<b>3</b>	<b>METHODS</b>	<b>33</b>
3.1	Materials	33
3.1.1	Samples	33
3.1.2	Chemicals and Reagents	34
3.1.3	Equipments	34
3.2	Extraction	34
3.3	Analysis of Extracts Bioactivities	35
3.3.1	Preparation of Extract	35
3.3.2	Screening of Antioxidant Activity	35
3.3.3	Antibacterial Activity Test	36
3.4	Statistical Analysis	39
<b>4</b>	<b>RESULT AND DISCUSSION</b>	<b>40</b>
4.1	<i>A. indica</i> Natural Products Extraction Yield and Efficiency	40

4.2	Efficiency of <i>A. indica</i> Extracts as Natural Antioxidants	41
4.3	Potential Aspect of <i>A. indica</i> in Microbial Growth Inhibition	45
<b>5</b>	<b>CONCLUSION AND FUTURE RECOMMENDATIONS</b>	<b>53</b>
5.1	Conclusion	53
5.2	Future Recommendations	54
	<b>REFERENCES</b>	<b>55</b>
	<b>APPENDICES</b>	<b>66</b>



**LIST OF TABLES**

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
<b>2.1</b>	Summary of various <i>A. indica</i> traditional uses.	10
<b>2.2</b>	Preliminary phytochemical constituents of <i>A. indica</i> various extracts from previous works	12
<b>2.3</b>	List of bioactive compounds found in <i>A. indica</i> and their structure.	12
<b>2.4</b>	Classification of <i>E. coli</i>	30
<b>2.5</b>	Classification of <i>S. aureus</i>	31
<b>2.6</b>	Classification of <i>P. aeruginosa</i>	32
<b>2.7</b>	Classification of <i>E. faecalis</i>	33
<b>4.1</b>	Extraction yield (%) per solvent used for extraction of different parts of <i>A. indica</i>	42
<b>4.2</b>	Concentration at IC <sub>50</sub> of <i>A. indica</i> extracts and ascorbic acid as standard	46

<b>4.3</b>	Diameter of inhibition zones (mm) of hexane extracts against four test organisms	50
<b>4.4</b>	Diameter of inhibition zones (mm) of methanol extracts against four test organisms	51
<b>4.5</b>	Diameter of inhibition zones (mm) of ethanol extracts against four test organisms	52
<b>4.6</b>	MIC values of plant extracts against four test organisms	53
<b>4.7</b>	MBC values of plant extracts against four test organisms	54

**LIST OF FIGURES**

<b>FIGURE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
<b>2.1</b>	<i>Acalypha indica</i>	9
<b>2.2</b>	Soxhlet Extractor unit	23
<b>2.3</b>	The basic chemical structure of flavonoids	27
<b>4.1</b>	Scavenging effect of hexane extracts from different parts of <i>A. indica</i> and ascorbic acid on DPPH radical.	44
<b>4.2</b>	Scavenging effect of methanol extracts from different parts of <i>A. indica</i> and ascorbic acid on DPPH radical.	44
<b>4.3</b>	Scavenging effect of ethanol extracts from different parts of <i>A. indica</i> and ascorbic acid on DPPH radical.	45

**LIST OF ABBREVIATIONS**

<b>B.C.</b>	- Before Centuries
<b>DMSO</b>	- Dimethylsulfoxide
<b>DPPH</b>	- 2,2-diphenyl-1-picrylhydrazyl
<b>g</b>	- Gram
<b>h</b>	- Hour
<b>IC<sub>50</sub></b>	- 50% inhibitory concentration
<b>nm</b>	- nanometre
<b>m</b>	- Metre
<b>MBC</b>	- Minimum Bactericidal Concentration
<b>mg</b>	- Milligram
<b>MIC</b>	- Minimum Inhibitory Concentration
<b>mL</b>	- Millilitre
<b>mm</b>	- Millimetre
<b>NA</b>	- Nutrient agar
<b>NB</b>	- Nutrient broth
<b>rpm</b>	- Round per minute
<b>%</b>	- Percentage
<b>µg</b>	- Microgram
<b>µL</b>	- Microliter
<b>°C</b>	- Degree Celcius

## CHAPTER 1

### INTRODUCTION

#### 1.1 General Introduction

For thousand years, medicinal plants have been used as traditional remedies to treat numerous human illnesses in many different parts of the world. In the developing world, particularly in rural areas, herbal remedies continue to be an essential source of medicine. Scientifically, medicinal plants have been demonstrated as a numerous source of biologically active compounds, majority of them have already been formulated into beneficial therapeutic substances.

*Acalypha indica* belongs to *Euphorbiaceae*, a large family of flowering plants. Majority of the species is distributed in the Indo-Malayan region and tropical America (Charles *et al.*, 2007) is an annual herb, about 80 cm high, a wild plant and commonly found in waste places or fields. It is locally known as “kucing galak” or “rumput lislis”, “kuppaimeni” in India and “t’ie han tsai” in China (Kirtikar & Basu, 1975). For a long time, *A. indica* has been used as traditional medicines of various countries and they are also reported to possess diuretic, purgative and anthelmintic properties, and also being used for bronchitis, asthma, pneumonia, scabies and other cutaneous diseases (Kirtikar & Basu, 1999).

Solvents selection have vital role in absorbing various bioactive compounds from plants. Polar solvent able to dissolve hydrophilic compounds, semi polar solvent able to dissolve both hydrophilic and lipophilic compounds while non-polar solvent absorbs lipophilic compounds. The usage of different solvents increase the possibilities to dissolve various different compounds (Charmi *et al.*, 2011) which in turn will affect the bioactivities such as antioxidant and antibacterial activities.

This experiment was conducted to evaluate the antioxidant activity of *A. indica* extracts using three different solvent by successive method. Next, the leaves and stem, roots and whole plant extract were screened on antioxidant and antibacterial properties to support the conventional therapeutic claim and to provide base line data for the scientific communities to conduct further study.

## **1.2 Problem Statement**

For decades, *A. indica* has been used as traditional remedies to treat various ailments such as skin diseases. However, there are still limited publications on the plants' biological activities such as antioxidant and antibacterial activity of *A. indica*, which might contribute to its various medicinal properties.

Nowadays, synthetic antioxidants such as butylated hydroxyanisole (BHA) and butylated hydroxytoluene (BHT) are often used in the food industries to prevent or inhibit the oxidative deterioration in foods. However, there are a few health and safety concerns in use of synthetic antioxidants due to their potential toxicity and carcinogenic (Izreen & Noriham, 2011). Thus, there is increasing importance to search the natural antioxidants from plants as an alternative to synthetic antioxidant. Besides, the knowledge on the potential of antioxidant properties of *A. indica* are still not well understood. Therefore, it is worthwhile to study the antioxidant activity of *A. indica* based on its medicinal benefits.

There have been an increase in bacterial resistance to antibacterial agents which in turn cause difficulties to treat infectious diseases (Prashanth, Asha & Amit, 2001). Thus, there is a need to search for new antibacterial agent specifically from plants as an alternative to synthetic antibiotics. This prompted the evaluation of *A. indica* as source of potential natural antibacterial agent by testing against human pathogens such as *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Enterococcus faecalis*.

### **1.3 Research Objectives**

- 1.3.1 To extract the leaves and stem, roots and whole plant of *A. indica* using various solvents by successive method.
- 1.3.2 To evaluate the antioxidant activity of the crude extracts.
- 1.3.3 To screen the antibacterial activities of the crude extracts.

### **1.4 Research Scope**

This research focuses on the study of the *A. indica* various extracts, its antioxidant activities and antibacterial activities. The fresh *A. indica* was extracted by using soxhlet extractor with hexane, methanol and ethanol. The biological activities such as antioxidant and antibacterial will be carried out on the crude extracts. 2, 2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity were conducted to assess the antioxidant activity. The antibacterial activity were evaluated by disc diffusion method, minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC).

## 1.5 Significance of Research

Even though the research of antibacterial agents towards important bacterial pathogens have increased throughout the world, it is well known that the number of new antibacterial agents being brought to the market has undergone a steady decline in the past several decades. Since antiquity, human has used plants to treat common infectious diseases and some of these traditional medicines are still included as part of the habitual treatment of various diseases (Rios & Recio, 2005). The antibacterial compounds from plants may inhibit bacterial growth by different mechanisms than antibiotics and may have a significant clinical value in treatment of resistant microbial strains (Eloff, 1998). This study contributing the overview of *A. indica* on antibacterial activities against common pathogenic bacteria.

Worldwide, there has been growing trend and interest in plants' natural antioxidants as natural additives in food and cosmetics. Plants are one of the most important targets to search for natural antioxidants as it is known safer compared to synthetic antioxidants (Yanislieva *et al.*, 2006). This study will contribute towards the growing database of knowledge on herbal medicines and help to advocate the safe and effective use of *A. indica* as traditional herbal remedies.



## REFERENCES

- Abou Elkair. E, Fadda. H and Abu Mohsen (2010). Antibacterial Activity and Phytochemical Analysis of Some Medicinal Plants from Gaza Strip-Palestine. *Journal of Al Azhar University-Gaza*, Vol. 12, 45-54.
- Abu-Shanab. B, Adwan. G, Abu-Safiya. D, Jarrar. N and Adwan. K (2004). Antibacterial Activities of Some Plant Extracts Utilized in Popular Medicine in Palestine. *Turkish Journal of Biology* Vol. 28, 99-102.
- Agarwal, S. S. and Paridhavi, M. (2005). Clinically Useful Herbal Drugs, Ahuja Publishing House, 281-282.
- Amarowicz, R., Pegg, R. B., Rahimi-Moghaddam, P., Barl, B., & Weil, J. A. (2004). Free-radical Scavenging Capacity and Antioxidant Activity of Selected Plant Species from the Canadian Prairies. *Food Chemistry*, 84(4), 551-562.
- Balandrin, M.F., Klocke, J.A., Wurtele, E.J, and Bollinger, W.H. (1985). Natural Plant Chemicals: Sources of Industrial and Medicinal Materials. *Science New Series*, 228(4704), 1154-1160.
- Ben-Erik Vanwyk, Micheal Wink. (2009) Medicinal Plant of the World. Published by Briz Publication, South Africa, Edition I, 403.
- Bernhoft, A., (2010). A Brief Review on Bioactive Compounds in Plants. In: Bioactive Compounds in Plants - Benefits and Risks for Man and Animals. Oslo: The Norwegian Academy of Science and Letters.
- Bennett R. N., Wallsgrove R. M. (1994). Secondary Metabolites in Plant Defense Mechanisms. *Phytotherapy Research*. 127: 617-633
- Blokhina, O., Virolainen, E., & Fagerstedt, K. V. (2003). Antioxidants, Oxidative Damage and Oxygen Deprivation Stress: A Review. *Annals of botany*, 91(2), 179-194.
- Bravo, L. (1998). Polyphenols: Chemistry, Dietary Sources, Metabolism, and Nutritional Significance. *Nutrition reviews*, 56(11), 317-333.

- Butler, M.S., 2004. The role of natural product chemistry in drug discovery. *Journal of Natural Products* 67, 2141-2153.
- Charles, C. D., Maribeth, L., Daniel, L. N., Kenneth, J. W., David, A. B. (2007) Floral Gigantism in Rafflesiaceae. Science Express, USA.
- Charmi, P.S., Vachhani, U.D., Trivedi, M.N., Joshi, V.J., and Santani, D.D. (2011). Evaluation of Antibacterial Potential of *Terminalia chebula* against Pathogenic Organisms of Lacrimal system. *Pharmacology online* 1:506-509.
- Cheremisinoff NP. 2003. Industrial solvents Handbook. Second Edition. Marcel Dekker, Inc. New York.
- Chopra, R. N., Nayar, S. L., Chpora. I. C. (2006) Glossary of Indian Medicinal Plants, National Institute of Science Communication and Information Resources, 54.
- Cowan, M. M. (1999). Plant Products as Antimicrobial Agents. *Clinical Microbiology Reviews*, 12(4), 564–582.
- Cragg, G. M., Grothaus, P. G., & Newman, D. J. (2012). Natural Products in Drug Discovery: Recent Advances. Plant Bioactives and Drug Discovery: Principles, Practice, and Perspectives. 4th ed., Hoboken: Wiley, 1-42.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15(10), 7313-7352.
- Decker, E. A., Warner, K., Richards, M. P., & Shahidi, F. (2005). Measuring Antioxidant Effectiveness in Food. *Journal of agricultural and food chemistry*, 53(10), 4303-4310.
- Dixon, R. A., Dey, P. M., & Lamb, C. J. (1983). Phytoalexins: enzymology and molecular biology. *Advances in enzymology and related areas of molecular biology*, 55, 1-136.
- Edwards, R., Tiller, S. A., & Parry, A. D. (1997). The Effect of Plant Age and Nodulation on the Isoflavonoid Content of Red Clover (*Trifolium pratense*). *Journal of plant physiology*, 150(5), 603-610.

- Eloff, J. N. (1998). Which Extractant Should Be Used For The Screening And Isolation Of Antimicrobial Components From Plants. *Journal of Ethnopharmacology*, 60(1), 1-8.
- Erhola, M., Kellokumpu-Lehtinen, P., Metsä-Ketelä, T., Alanko, K., & Nieminen, M. M. (1996). Effects of Anthracyclin-Based Chemotherapy on Total Plasma Antioxidant Capacity in Small Cell Lung Cancer Patients. *Free Radical Biology and Medicine*, 21(3), 383-390.
- Faure, H., Coudray, C., Mousseau, M., Ducros, V., Douki, T., Bianchini, F., & Favier, A. (1996). 5-Hydroxymethyluracil Excretion, Plasma TBARS and Plasma Antioxidant Vitamins in Adriamycin-Treated Patients. *Free Radical Biology and Medicine*, 20(7), 979-983.
- Fawole, A.M. (2009). *Pharmacology and phytochemistry of South African traditional medicinal plants used as antimicrobials*. MSc thesis. University of KwaZulu Natal, Pietermaritzburg.
- Ferruzzi, M.G., Bohm, V., Courtney, P.D., & Schwartz, S.J. (2002). Antioxidant And Antimutagenic Activity of Dietary Chlorophyll Derivatives Determined by Radical Scavenging And Bacterial Reverse-mutagenesis Assays. *Journal of Food Science*. 67(7), 2589-2595.
- Ganesan, A., (2008). The Impact of Natural Products upon Modern Drug Discovery. *Current Opinion in Chemical Biology* 12, 306-317.
- Gordon M. H. (2001). Measuring Antioxidant Activity. In: Pokorný J, Yanishlieva N, Gordon M, editors. Antioxidants in food: practical application. Cambridge England: Woodhead Publishing Limited. P73-84.
- Green, R. J. (2004). Antioxidant Activity of Peanut Plant Tissues. Master's Thesis. NC State University.
- Hall C. 2001. Sources of Natural Antioxidants: Oilseeds, Nuts, Cereals, Legumes, Animal Products and Microbial Sources. In: Pokorný J, Yanishlieva N, Gordon

M, editors. Antioxidants in food: practical applications. Cambridge England: Woodhead Publishing Limited. P159-209.

Harborne, J. B. (2013). *The flavonoids: advances in research since 1980*. Springer.

Hsu, B., Coupar, I. M., & Ng, K. (2006). Antioxidant Activity of Hot Water Extract from the Fruit of the Doum Palm, *Hyphaene thebaica*. *Food Chemistry*, 98(2), 317-328.

Houghton, P.J., and Raman, A. (1998). Laboratory Handbook for the Fractionation of Natural Extracts 1st Edition. New York, NY: Chapman & Hall.

Hussain, A. Z., & Ignatius, A. (2010). GC-MS Analysis and Antimicrobial Activity of *Acalypha indica* Linn. *Asian Journal of chemistry*, 22(5), 3591.

Izreen, I., & Noriham, A. (2011). Evaluation of the Antioxidant Potential of Some Malaysian Herbal Aqueous Extracts As Compared With Synthetic Antioxidants and Ascorbic Acid in Cakes. *International Food Research Journal*, 18, 583-587.

Jacobs, F. (1985). *Breakthrough: The True Story of Penicillin*. Dodd Mead.

Jagatheeswari, D., & Deepa, J. (2013). *Acalypha indica* L-an Important Medicinal Plant: a Review of Its Traditional Uses, and Pharmacological Properties. *International Journal of Research in Botany*, 3(2009), 19–22.

Jones, W. P., & Kinghorn, A. D. (2005). Extraction of plant secondary metabolites. In *Natural products isolation* (pp. 323-351). Humana Press.

Joshi, B., Lekhak, S., Sharma A., (2009). Antibacterial Property of Different Medicinal Plants: *Ocimum sanctum*, *Cinnamomum zeylanicum*, *Xanthoxylum armatum* and *Origanum majorana*. *Kathmandu University Journal of Science, Engineering and Technology*, 5(1): 143 – 150.

Kalia, K., Sharma, K., Singh, H. P., & Singh, B. (2008). Effects of Extraction Methods on Phenolic Contents and Antioxidant Activity in Aerial Parts of *Potentilla*

atrosanguinea Lodd. and Quantification of Its Phenolic Constituents by RP-HPLC. *Journal of agricultural and food chemistry*, 56(21), 10129-10134.

Karakoca, K., Ozusaglam, M. A., Cakmak, Y. S., & Erkul, S. K. (2013). Antioxidative, Antimicrobial and Cytotoxic Properties of *Isatis floribunda boiss. ex bornm.* Extracts. *Excli Journal*, 12, 150-167.

Karou D., Savadogo A., Canini A., Yameogo S., Montesano C., Simpore J., Collizi V. and Traore A. (2005). Antibacterial Activity of Alkaloids from *Sida acuta*. *African Journal of Biotechnology*. 4 (12): 1452-57.

Kinghorn, A. D., and Manuel F. B. (1993). Human medicinal agents from plants. American Chemical Society (ACS).

Kirtikar, K.R. and Basu B.D. (2006) Indian Medicinal Plants, International book distributors, 2856-2860.

Kirtikar, K. R. and Basu, B. D. (1999). Indian Medicinal Plant. Volume III, International Book Distributors, Dehradun, India, 2262-2263.

Kirtikar, K. R. and Basu, B. D. (1975). Indian Medical Plants. Volume II. Second Edition. Jayyed Press, New Delhi, 30-45.

Koehn, F. E., & Carter, G. T. (2005). The Evolving Role of Natural Products in Drug Discovery. *Nature reviews Drug discovery*, 4(3), 206-220.

Kowti, R., Harsha, R., Ahmed, M., Hareesh, A., Thammanna Gowda, S., Dinesha, R., & Irfan Ali, M. (2010). Antimicrobial Activity of Ethanol Extract of Leaf and Flower of *Spathodea campanulata P. Beauv.* *Research Journal of Pharmaceutical, Biological and Chemical Science*, 3(1), 691-8.

Kumar. M, Agarwala. R, Deyb. K, Raib. V, Johnsonc. B (2009) Antimicrobial Activity of Aqueous Extract of *Terminalia chebula* Retz. On Gram positive and Gram negative Microorganisms. *International Journal of Current Pharmaceutical Research* Vol. 1 (1): 56-60.

Leeds, J.A., Schmitt, E.K. and Krastel, P. (2006). Recent Development in Antibacterial Drug Discovery: Microbe-Derived Natural Products - From Collection to Clinic. *Expert Opinion on Investigational Drugs*, 15(3), 211-226.

- Lucera, A., Costa, C., Conte, A., & Del Nobile, M. A. (2012). Food Applications of Natural Antimicrobial Compounds. *Frontiers in Microbiology*, 3, 287. doi:10.3389/fmicb.2012.00287
- Lupo, M. P. (2001). Antioxidants and Vitamins in Cosmetics. *Clinics in dermatology*, 19(4), 467-473.
- Mackie TJ, McCartney JE (1989). Microbial Infections. Medical Microbiology. 13th Edition Longman Group Limited, London.
- Mahon, C. R., & Manuselis, G. (1995). Enterobacteriaceae. *Textbook of Diagnostic Microbiology*, 2, 463-511.
- Mahugo Santana, C., Sosa Ferrera, Z., Esther Torres Padrón, M., & Juan Santana Rodríguez, J. (2009). Methodologies for the Extraction of Phenolic Compounds from Environmental Samples: New Approaches. *Molecules*, 14(1), 298-320.
- Mancini, I., Defant, A. and Guella, G. (2007). Recent Synthesis of Marine Natural Products with Antibacterial Activity. *Anti-infective agents in Medicinal Chemistry*, 6, 17-48.
- Martin, G.J. (1995). *Ethnobotany: A people and plants conservation manual*. United Kingdom. Chapman and Hall. pp. 68-69.
- Mdlolo, C.M. (2009). Phytochemical Analysis and Selected Biological Activity of *Phyllanthus parvulus* Sond. var. *gariensis*. MSc thesis. University of Zululand. South Africa.
- Miller, K. W., Reo, N. V., Uiterkamp, A. S., Stengle, D. P., Stengle, T. R., & Williamson, K. L. (1981). Xenon NMR: Chemical Shifts of a General Anesthetic in Common Solvents, Proteins, and Membranes. *Proceedings of the National Academy of Sciences*, 78(8), 4946-4949.
- Mollik, M. A. H., Hossan, M. S., Paul, A. K., Taufiq-Ur-Rahman, M., Jahan, R., & Rahmatullah, M. (2010). A Comparative Analysis of Medicinal Plants Used By Folk Medicinal Healers in Three Districts of Bangladesh and Inquiry as To Mode of Selection of Medicinal Plants. *Ethnobotany Research and Applications*, 8, 195-218.

- Nadkarni, K. M. (2009) Indian Materia Medica. Volume 1. Bombay Popular Prakashan, 285-286.
- Moreno, S., Scheyer, T., Romano, C. S., & Vojnov, A. A. (2006). Antioxidant And Antimicrobial Activities Of Rosemary Extracts Linked To Their Polyphenol Composition. *Free radical research*, 40(2), 223-231.
- Naczk, M., & Shahidi, F. (2006). Phenolics in Cereals, Fruits and Vegetables: Occurrence, Extraction and Analysis. *Journal of pharmaceutical and biomedical analysis*, 41(5), 1523-1542.
- Nicolaou, K.C., Chen, J.S., Edmonds, D.J. and Estrada, A.A. (2009). Recent Advances in the Chemistry and Biology of Naturally Occurring Antibiotics. *Angewandte Chemie International Edition English*, 48(4), 660-719.
- Nijveldt, R. J., Van Nood, E., Van Hoorn, D. E., Boelens, P. G., Van Norren, K., & Van Leeuwen, P. A. (2001). Flavonoids: A Review of Probable Mechanisms of Action and Potential Applications. *The American journal of clinical nutrition*, 74(4), 418-425.
- Palmer, J., Flint, S., & Brooks, J. (2007). Bacterial Cell Attachment, the Beginning of a Biofilm. *Journal of industrial microbiology & biotechnology*, 34(9), 577-588.
- Papo, N., & Shai, Y. (2005). A Molecular Mechanism for Lipopolysaccharide Protection of Gram-Negative Bacteria from Antimicrobial Peptides. *Journal of Biological Chemistry*, 280(11), 10378-10387.
- Parish ME, Davidson PM. (1993). Methods For Evaluation: In Antimicrobials in foods, 2nd Ed. Marcel Dekker, Inc., New York, 597-615.
- Phillipson, J. D., & O'Neill, M. J. (1989). New Leads To The Treatment Of Protozoal Infections Based On Natural Product Molecules. *Acta Pharmaceutica Nordica*, 1(3), 131-144.
- Pokorný J. and Korczak J. (2001). Preparation of natural antioxidants. In: Pokorný J, Yanishlieva N, Gordon M, editors. Antioxidants in food: practical application. Cambridge England: Woodhead Publishing Limited. P311-41.

- Potier, P., Gueritte-Voegelein, F. and Guenard, D. (1996). The search and discovery of, two new antitumor drugs, Navelbine and Taxorete, modified natural products. In Hostettmann, K., Chinyanganya, F. and Maillard, F. (Ed). *Chemistry, Biological and Pharmacological properties of African Medicinal Plants: Proceeding of the first international IOCD-Symposium* Victoria Falls, Zimbabwe, February 25-28. University of Zimbabwe Publications, pp. 69-76.
- Prashanth, D., Asha, M. K., & Amit, A. (2001). Antibacterial Activity of *Punica granatum*. *Fitoterapia*, 72(2), 171-173.
- Pratt D. and Hudson B. J. F. (1990). Natural Antioxidants Not Exploited Commercially. In:Hudson B.J.F, editor. Food antioxidants. New York: Elsevier. P171-91.
- Preedy, V. R. (Ed.). (2012). Isoflavones: chemistry, analysis, function and effects (No. 5). Royal Society of Chemistry.
- Prior, R. L., Wu, X., & Schaich, K. (2005). Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of agricultural and food chemistry*, 53(10), 4290-4302.
- Radojević. I, Stanković. O, Topuzović. M, Čomić. L & Ostojić. A (2012). Great Horetail (*Equisetum telmateia* Ehrh.): Active Substances Content and Biological Effects. *Experimental and Clinical Sciences International Journal*, Vol. 11:59-67.
- Rafii, F., Sutherland, J. B., & Cerniglia, C. E. (2008). Effects of Treatment with Antimicrobial Agents on the Human Colonic Microflora. *Therapeutics and Clinical Risk Management*, 4(6), 1343–1358.
- Rastogi, R. P., Mehrotra, B. N. (2004). Compendium of Indian Medicinal plants. Volume 4. Central Drug Research Institute, Lucknow and National Institute of Science Communication and Information Resources, New Delhi, 155-156.
- Rios, J. L., & Recio, M. C. (2005). Medicinal plants and antimicrobial activity. *Journal of ethnopharmacology*, 100(1), 80-84.
- Robak, J., & Gryglewski, R. J. (1988). Flavonoids are scavengers of superoxide anions. *Biochemical pharmacology*, 37(5), 837-841.



- Ryan K. J., Ray C. G. (2004) Sherris Medical Microbiology: An Introduction to Infectious Diseases. 4th edition. McGraw Hill Publishers.
- Sahoo, R. R. (2012). Antioxidant & Antimicrobial Efficacy of *Ficus religiosa L.* & *Ficus benghalensis L.* Master's Thesis. National Institute of Technology Rourkela.
- Sanseera, D., Niwatananun, W., Liawruangrath, B., Liawruangrath, S., Baramée, A., Trisuwan, K., & Pyne, S. G. (2012). Antioxidant and Anticancer Activities from Aerial Parts of *Acalypha indica* Linn. *Chiang Mai University Journal of Natural Sciences*, 11 (2), 157-168.
- Sanseera Duangsuree, Niwatananun Wirat, Liawruangrath Boonsom, Liawruangrath Saisunee, Baramée Aphiwat., and Stephen G. Pyne. (2010). Antimicrobial activities of various medicinal plant extracts in Family Euphorbiaceae. p. 562. Proceeding of Pure and Applied Chemistry for Sustainable Development 2010, 21-23 January 2000. Sunee Grand Hotel and Convention Center, Ubon Ratchathani, Thailand.
- Sarkar, D., Sharma, A. and Talukder, G. (1996). Plant Extracts as Modulators of Genotoxic Effects. *Botanical Review*, 62(4), 275-300.
- Sarker, S. D., Latif, Z., & Gray, A. I. (2005). Natural products isolation (Vol. 20). Springer Science & Business Media.
- Saxena, G. O. L. D. Y., Kalra, S. S., & Gupta, N. (2011). Antimicrobial Activity Pattern of Certain Terpenoids. *Int J Pharma Bio Sci*, 2, 87-91.
- Schefflan L, Jacobs MB. (1953). The handbook of solvents. First Edition. Robert E. Krieger Publishing Co., New York.
- Schillaci, C., Nepravishta, R., & Bellomaria, A. (2014). Antioxidants in Food and Pharmaceutical Research. *Albanian Journal of Pharmaceutical Sciences*, 1(1), 9-15.
- Sekar, D. (2012). Screening of *Phyllanthus amarus*, *Acalypha indica* and *Datura metel* for its Antimicrobial Activity against Selected Pathogens. *International Journal of Pharmaceutical & Biological Archive*, 3(5).

- Shahid Chatha, S. A., Anwar, F., Manzoor, M., & Rehman Bajwa, J. U. (2006). Evaluation of the Antioxidant Activity Of Rice Bran Extracts Using Different Antioxidant Assays. *Grasas y aceites*, 57(3), 328-335.
- Shanmugapriya, R. (2012). Evaluation of Antioxidant Potential and Antibacterial Activity of *Acalypha indica* Linn. Using In Vitro Model. *Asian Journal of Biomedical and Pharmaceutical Sciences*, 1(1).
- Silva, G. L., Lee, I. S., & Kinghorn, A. D. (1998). Special Problems with the Extraction of Plants. In *Natural Products Isolation* (pp. 343-363). Humana Press.
- Singh, J. (2008). Maceration, percolation and infusion techniques for the extraction of medicinal and aromatic plants. *Extraction Technologies for Medicinal and Aromatic Plants*, 67.
- Sultana, B., Anwar, F., & Przybylski, R. (2007). Antioxidant Activity of Phenolic Components Present in Barks of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica*, and *Eugenia jambolana* Lam. trees. *Food Chemistry*, 104(3), 1106-1114
- Tiwari, P., Kumar, B., Kaur, M., Kaur, G., & Kaur, H. (2011). Phytochemical Screening and Extraction: A Review. *Internationale pharmaceutica scientia*, 1(1), 98-106.
- Tromp, J. (1983). Nutrient Reserves in Roots of Fruit Trees, In Particular Carbohydrates and Nitrogen. In *Tree root systems and their mycorrhizas* (pp. 401-413). Springer Netherlands.
- Vaishnava, M. M., Tripathi, A. K., Gupta, K. R. (1993). Constituents of *Cassia fistula* Roots. *Fitoterapia*. 64(1), 93.
- Wiegand, I., Hilpert, K., & Hancock, R. E. (2008). Agar and Broth Dilution Methods to Determine the Minimal Inhibitory Concentration (MIC) Of Antimicrobial Substances. *Nature protocols*, 3(2), 163-175.
- Wade Jr, L. G. (2003). Infrared Spectroscopy and Mass Spectrometry. *Organic Chemistry. 5th ed. Pearson Education: Upper Saddle River, NJ*, 490-538.

- Wuyang, H. (2008). Traditional Chinese medicinal plants and their endophytic fungi: Isolation, identification and bioassay. *PhD thesis*. University of Hong Kong, China, pp. 1-1
- Yanishlieva N. (2001). Inhibiting Oxidation. In: Pokorný J, Yanishlieva N, Gordon M, editors. *Antioxidants in Food: Practical Application*. Cambridge England: Woodhead Publishing Limited. P22-70.
- Yanishlieva, N. V., Marinova, E., & Pokorný, J. (2006). Natural Antioxidants from Herbs and Spices. *European Journal of Lipid Science and Technology*, 108(9), 776-793.
- Zhao, W. H., Hu, Z. Q., Okubo, S., Hara, Y., & Shimamura, T. (2001). Mechanism of Synergy Between epigallocatechin gallate and  $\beta$ -lactams against methicillin-resistant *Staphylococcus aureus*. *Antimicrobial Agents and Chemotherapy*, 45(6), 1737-1742.
- Zhou, M., Luo, H., Li, Z., Wu, F., Huang, C., Ding, Z., & Li, R. (2012). Recent Advances in Screening of Natural Products for Antimicrobial Agents. *Combinatorial chemistry & high throughput screening*, 15(4), 306-315.
- Zicker, S. C. (2005). Cognitive and Behavioral Assessment in Dogs and Pet Food Market Applications. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 29(3), 455-459.
- Ziegler J. & Facchini P. (2008). Alkaloid Biosynthesis: Metabolism and Trafficking. *Annual Review of Plant Biology*, 59, 735-69.