ANTI-MICROBIAL AND ANTI-INFLAMMATORY ACTIVITIES OF CRUDE AND ACIDIFIED FRACTION OF SELECTED MALAYSIAN HONEY SAMPLES

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ABSTRACT

Honey has been reported to have anti-microbial and anti-inflammatory properties, due to the presence of polyphenol and hydrogen peroxide, respectively. To our knowledge, the detail compounds that contribute to the activities are still unknown. Therefore, this study was focused on anti-microbial and the antiinflammation activities of selected honey samples including Tualang, Acacia and Gelam honey from Malaysia. First, the honey samples were fractionated by acidified water (pH 2) in a solid phase extractor using C18 column. Then, the honey fraction was assayed for anti-microbial and anti-inflammatory activities using well diffusion method and enzymatic cyclooxygenase (COX-1 and -2) assay, respectively. For antimicrobial study, four different microbial strains consisted of gram positive (Staphylococcus aureus) and gram negative (Salmonella typimurium) bacteria, yeast (Candida albican), as well as fungi (Fusarium oxysporum) were tested on the honey samples. The result showed that only Salmonella typimurium and Candida albican were inhibited by crude and acidified fraction of Tualang, Acacia and Gelam honey, but no inhibition was observed for Staphylococcus aureus and Fusarium oxysporum. The minimum inhibitory concentration (MICs) of the crude honey samples (50.0% w/v) and their fractions (12.5-25.0% w/v) against Salmonella typimurium were about two to three times higher than the MICs of Manuka honey and its fraction (6.3-12.5% w/v). However, the MICs of honey samples (3.1-6.3% w/v for crude honey samples and 0.8-3.1% w/v for their acidified fractions) against Candida albican appeared to be better, or at least comparable to Manuka honey samples (25.0% w/v for crude honey samples and 3.1% w/v for their acidified fractions). On the other hand, the anti-inflammatory activity of honey samples at 50% of inhibition (IC₅₀) was found to be 0.7-1.7 mg/ μ L for COX-1 and 0.4-1.3 mg/ μ L for COX-2. Fractionation did not improve the anti-inflammatory activity of honey samples because their IC₅₀ values were increased to 320.0-1080.0 mg/L for COX-1 and 280.0-400.0 mg/L for COX-2. In term of selectivity ratio of COX-1/COX-2, the crude honey samples of Gelam (ratio=3.3) and its fraction (ratio=2.7) appeared to be the most selective COX-2 inhibitor, which was about two times higher than the selectivity of Manuka honey. The anti-microbial and anti-inflammatory properties of honey could be due to the presence of quercetin, chlorogenic acid, acacetin, apigenin-7-o-glucoside, myricetin and coumaryl quinic acid. These phenolic acids and flavonoids were detected in the honey fraction.

ABSTRAK

Madu telah dilaporkan mempunyai sifat anti-mikrob dan anti-inflamatori masing-masing disebabkan oleh kehadiran polifenol dan hidrogen peroksida. Dalam pengetahuan kami, sebatian yang menyumbang kepada aktiviti tersebut masih tidak diketahui. Oleh itu, kajian ini memberi tumpuan kepada aktiviti anti-mikrob dan antiinflamatori sampel madu terpilih dari Malaysia termasuk Tualang, Acacia dan Gelam. Sampel madu dipecahkan kandungannya menggunakan air berasid (pH 2) di dalam pemecah fasa pepejal kolum C18. Pecahan madu telah dinilai untuk kehadiran aktiviti anti-mikrob dan anti-inflamatori yang dilakukan ke atas sampel pecahan madu menggunakan kaedah masing-masing iaitu telaga resapan dan cerakin enzimatik siklooksigenase (COX-1 dan COX-2). Empat jenis strain mikrob yang berbeza terdiri daripada gram positif (Staphylococcus aureus), gram negatif (Salmonella typimurium), yis (Candida albican) dan juga kulat (Fusarium oxysporum) telah diuji ke atas sampel madu. Keputusan menunjukkan bahawa, hanya Salmonella typimurium dan Candida albican telah direncatkan oleh madu mentah dan pecahan berasid daripada sampel Tualang, Acacia dan Gelam, tetapi tiada perencatan diperhatikan bagi Staphylococcus aureus dan Fusarium oxysporum. Kepekatan perencatan minimum (MICs) sampel madu mentah (50.0% w/ v) dan pecahannya (12.5-25.0% w/v) terhadap Salmonella typimurium adalah kira-kira dua hingga tiga kali lebih tinggi daripada MICs daripada madu Manuka dan pecahannya (6.3-12.5% w/v). Walau bagaimanapun, MICs sampel madu (3.1-6.3% w/v bagi sampel madu mentah dan 0.8-3.1% w/v bagi sampel pecahan berasid) terhadap Candida albican adalah lebih baik, atau sekurang-kurangnya setanding dengan sampel madu Manuka (25.0% w/v bagi sampel madu mentah dan 3.1% w/v bagi sampel pecahan berasid). Sebaliknya, aktiviti anti-inflamasi terhadap sampel madu di 50% perencatan (IC₅₀) didapati 0.7-1.7 mg/µL bagi COX-1 dan 0.4-1.3 mg/µL bagi COX-2. Pemecahan tidak meningkatkan aktiviti anti-inflamasi sampel madu kerana nilai IC₅₀ telah meningkat kepada 320.0-1080.0 mg/L bagi COX-1 dan 280.0-400.0 mg/L bagi COX-2. Dari segi nisbah selektif COX-1/COX-2, sampel madu mentah dari Gelam (nisbah=3.3) dan pecahannya (nisbah=2.7) adalah merupakan perencat COX-2 yang paling selektif, di mana kira-kira dua kali lebih tinggi berbanding keselektifan madu Manuka. Ciri-ciri anti-mikrob dan anti-inflamasi madu mungkin disebabkan oleh kehadiran kuersetin, asid klorogenik, akasetin, apigenin-7-oglukosida, mirisetin dan asid kuinik kumaril. Asid fenolik dan flavonoid tersebut telah dikesan di dalam sampel pecahan madu.

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LIST OF ABREVIATIONS

А	-	Acacia
AA	-	Arachidonic Acid
APCI	-	Atmospheric Pressure Chemical Ionisation
ATCC	-	American Type Culture Collection
ATP	-	Adinose trisphosphate
CA	-	California
COX	-	Cyclooxygenase
COX-1	-	Cyclooxygenase-1
COX-2	-	Cyclooxygenase-2
DNA	-	Deoxyribonucleic acid
ELISA	-	Enzyme-Linked Immunosorbant Assay
ESI	-	Electrospray ionization
FAMA	-	Federal Agricultural Marketing Authority
G	-	Gelam
GC	-	Gas Chromatography
GOx	-	Glucose Oxidase Enzyme
H_2O_2	-	Hydrogen peroxide
HCL	-	Hydrochloric acid
HMF	-	Hydroxymethylfurfural
HPLC	-	High-Performance Liquid Chromatography
IC ₅₀		Maximal Inhibitory Concentration at 50%
IL1β	-	Interleukin B
IL-6	-	And Interleukin-6
iNOS	-	Inducible Nitric Oxide Synthase
LC-MS/MS	-	Liquid Chromatography Tandem Mass
		Spectrometry

LOX	-	Lipoxygenase
М	-	Manuka
MA	-	Massachusets
MBC	-	Minimum Bactericidal Concentration
MIC	-	Minimum Inhibitory Concentration
MM6	-	Macro Mac 6
MO	-	Missouri
MRM	-	Multiple Reaction Monitoring
MRSA	-	Methicillin-Resistant Staphylococcus Aureus
MSA	-	Methicillin-Sensitive Staphylococcus Aureus
NA	-	Not available
ND	-	Not detected
NO	-	Nitric oxide
NSAID	-	Non-Steroidal Anti-Inflammatory Drugs
PAMPs	-	Pathogen-Associated Molecular Pattern
PDA	-	Potato Dextrose Agar
PGD	-	ProstagIndin
PGD ₂	-	Prostaglandin D ₂
PLA ₂	-	Phospholipase A ₂
ppm	-	Parts per million
PRRs	-	Pattern Recognition Receptors
RNS	-	Reactive Nitrogen Species
ROS	-	Reactive Oxygen Species
SDA	-	Sabaraud Dextrose Agar
sp	-	Species
SPE	-	Solid Phase Extraction
Т	-	Tualang
TLC	-	Thin Layer Chromatography
TNF-α	-	Tumour Necrosis Factor-A
TRIS-HCL	-	Trisbase Hydrochloric Acid
UMF	-	Unique Manuka Factor
UPLC	-	Ultra Performance Liquid Chromatography
USA	-	United State Of America
VCE	-	Vancomycin-Resistant Entrococcus

LIST OF SYMBOLS

Amu/s	-	Atomic mass unit per second
cm	-	Centimeter
eV	-	Electron volt
g	-	Gram
kg	-	Kilogram
kV	-	Kilovolt
Μ	-	Molarity
m/z	-	Mass to charge ratio
mg	-	Miligram
mg/µL	-	Milligram per microlitre
mg/kg	-	Milligram per kilogram
mL	-	Mililitre
mL/min	-	Mililitre per minute
mm	-	Milimeter
mM	-	Milimolar
MΩ-cm	-	Megaohm-centimeter
Ν	-	Normality
ppm	-	Parts per million
psi	-	Pound per square inch
V	-	Volt
v/v	-	Volume per volume
w/v	-	Weight per volume
μg	-	Microgram
μL	-	Microlitre

μL/min	-	Microlitre per minute
μm	-	Micrometer
μΜ	-	Micromolar
-	-	Minus
+	-	Plus
°C	-	Degree Celcius
%	-	Percentage

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CHAPTER 1

INTRODUCTION

1.1 Introduction to Research Background

This chapter contains the research background, problem statement, significance of study, objectives and scopes of this work under the section of 1.2 to 1.5. General information about honey and its composition, as well as its applications are also discussed in this chapter.

Honey is a thick, liquid form of natural product consisted of carbohydrates, free amino acids, vitamins, trace elements, phenolic compounds, organic acids, proteins and enzymes (Kassim *et al.*, 2010 and Ferreres, 1993). Mostly, honey majority consists of saturated sugars such as fructose (38.0%) and glucose (31.0%) (Gheldof *et al.*, 2002). Honey contains approximately 17.7% of water, 0.1% of total acidity and 0.2% of ashes (Nagai *et al.*, 2006). However, honey compositions are different depending on its environmental and climate condition, as well as processes that it undergoes during processing (Gheldof *et al.*, 2002)

Honey exhibits antioxidant, anti-bacterial, antiviral, anti-microbial and antiinflammatory activities (Martos *et al.*, 2008). Honey is also being widely used as traditional medicine. Honey is also consumed to provide gastric protection against gastric lesions (Caravaca *et al.*, 2006). It also helps to treat certain illnesses such as athma, cough, skin cancer and to promote wound healing from infection and burns (Cooper *et al.*, 1999; Molan, 1999; Fox, 2002; Molan, 2006).

Honey is a good anti-microbial agent. It was reported that honey could kill various classes of microbes, namely, gram positive and gram negative bacteria, fungi and yeast (Efem *et al.*, 1992; Nasser *et al.*, 2003; Halco'n and Milkus, 2004; Omoya and Akharaiyi, 2010). Honey was also reported to inhibit antibiotic resistant microbes such as Meticillin Resistant *Stapylococcus aureus* (MRSA) and Vancomycin Resistant *Enterococcus* (VRE) (Allen *et al.*, 2000). There are several factors that contribute to this anti-microbial effect such as low pH value, osmolarity effect and H_2O_2 content of honey. It is believed that there are compounds that give its anti-microbial property, particularly flavonoids and phenolic acids (Havsteen, 1983).

Many studies showed honey could reduce inflammation and treat inflammatory related disease (Subhramanyam, 1998). Honey could heal inflammation when applied directly on wound, and accelerating wound healing (Subhramanyam, 1998; Molan, 2006). The anti-inflammatroy property of honey is closely related to its flavonoid and phenolic content. For example, galangin and chrysin (flavonoid) were likely to inhibit enzyme that executed inflammation (Raso *et al.*, 2001; Kim *et al.*, 2002). Flavonoids in honey could expel free radicals that might contribute to inflammation (Garcia-Lafuente *et al.*, 2009).

1.2 Research Problem Statement

Honey has known for its numerous applications since ancient time. Honey is a potential source of anti-microbial and anti-inflammatory agents. However, scientific information regarding anti-microbial and anti-inflammatory properties of Malaysian honey is very limited. Although, there were many studies done on antimicrobial activity of honey, little research was based on the use of Malaysian honey (Mulu *et al.*, 2004; Al-Jabri *et al.*, 2003). Therefore, it is important to collect data regarding anti-microbial and anti-inflammatory properties of local honey samples.

Many researches have been done to investigate the group of compounds that responsible for anti-microbial and anti-inflammatory effects of honey (Russel *et al.*, 1990). However, the reported compounds that responsible for these effects were varied according to the honey origin. Hence, it is essential to determine the compound in local honey samples that contribute to these biological activities.

Cyclooxygenase (COX) assay has been used to test for anti-inflammatory drugs. However, little information regarding COX assay on honey samples. This is because honey is a complex mixture of compounds. Somehow, this COX assay could be used to have a rapid screening on semi-purified honey fraction for bioactive compound identification.

1.3 Significance of Study

Honey is a natural food that has been consumed since ancient time. It contains about 181 substances (Caravaca *et al.*, 2006). Honey has also been used as remedies for health promotion. This indicates that honey has many benefits yet to be explored. In recent years, modern societies have become more conscious about natural treatment for disease fighting and the production of honey related products is increasing. Although some people view the idea as somewhat primitive or ignorant, many remedies are the result of empirical observation since thousands of years.

Data obtained from this research can provide information regarding antimicrobial and anti-inflammatory effect of Malaysian honey. This study also determines the groups of compounds that exhibit anti-microbial and antiinflammatory effect. The information obtained is essential for the development of new antibiotic and anti-inflammatory drug.

The findings of this study can prove that Malaysian honey is comparable to imported Manuka honey in terms of its ability to prevent microbial infections and inflammation. This can help to boost the quality value of Malaysian honey, thus leading to the increment of Malaysia economy through import and export activity. The increase of Malaysia economy can provide more job opportunities to Malaysian.

1.4 Objectives of Study

The objective of this study was to identify bioactive compounds from Tualang, Acacia and Gelam honey samples for anti-microbial and anti-inflammatory activities based on well diffusion technique and COX assay, respectively.

1.5 Scope of Study

The scopes of this research included:

- (i) To fingerprint the acidified fractions collected from honey samples.
- (ii) To investigate anti-microbial activity of crude and acidified fraction of honey samples by using well diffusion technique.
- (iii) To investigate anti-inflammatory activity of crude and acidified fraction of honey samples by using cyclooxygenase assay.

REFERENCES

- Adock, D. (1912). The Effect of Catalase on Inhibine and Peroxide Values of Various Honeys. *Journal of Apicultural Research*. Vol. 1, 38–40.
- Allen, K. L., Molan, P.C. and Reid, G.M., (1991). A Survey of the Antibacterial Activity of Some New Zealand Honeys. *Journal of Pharmacy and Pharmacology*. Vol. 43, 817-822.
- Allen, K. L., Hutchinson, G. and Molan, P.C., (2000). The Potential for Using Honey to Treat Wounds Infected with MRSA and VRE. First Worldwide Healing Congress. 10-13 September Melbourne Australia.
- Al-Jabri, A. A., Nzeako, B., Mahrooqi, Z Al., Naqdy, A. Al. and Nsanze, H. (2003). In vitro Antibacterial Activity of Omani and African honey. *Journal of Biomedical Science*. Vol. 60, 1-4.
- Aljadi, A.M., and Kamaruddin, M.Y. (2004). Evaluation of Phenolic Content and Antioxidant Capacities of Two Malaysian Floral Honeys. *Food Chemistry*. Vol. 85, 513-518.
- Al-Mamary, M., Al-Meeri, A. and Al-Habori, M. (2002). Antioxidant Activities and Total Phenolics of Different Types of Honey. *Nutrition Research*. Vol. 22, 1041-1047.
- Alzahrani, H.A., Alsabehi, R., Boukraa, L., Abdellah, F., Bellik, Y. and Bakhotmah,
 B.A. (2012). Antibacterial and Antioxidant Potency of Floral Honeys from
 Different Botanical and Geographical Origins. *Molecules*. Vol. 17, 10540-10549.
- Altoparlak, U., Aktas, F., Selebi, D., Ozkurt, Z. and Akcay, M., (2005). Prevalence of Metallo-b-lactamase Among *Pseudomonas aeruginosa* and *Actinobacter baumanii* Isolated from Burn Wounds and In-vitro Activities of Antibiotic Combinations Against these Isolates. *Burns*. Vol. 31, 707–710.

- Al-Waili, N.S. (2004). Investigating the Antimicrobial Activity of Natural Honey and its Effect on the Pathogenic Bacterial Infections of Surgical Wounds and Conjuctiva. *Journal of Medicinal Food*. Vol.7. 210-222.
- Allwood, J.W. and Goodacre, R. (2009). An Introduction to Liquid Chromatography-Mass Spectrometry Instrumentation Applied in Plant Metabolomic Analyses. *Phytochemical Analysis*. Vol. 21, 33-47.
- Anyanwu, C.U. (2012). Investigation of In-vitro Antifungal Activity of Honey. Journal of Medicinal Plants Research. Vol. 6, 3512-3516.
- Atrott, J. and Henle, T. (2009). Methylglyoxal in Manuka Honey- Correlation with Antibacterial Properties. *Czech Journal of Food Sciences*. Vol. 27, 163-165.
- Aween, M.M., Hassan, Z., Faujani, N.H., Emdakim, M.M. and Muhialdin, B.J. (2014). Potency of Honey as Antibacterial Agent Against Multiple Antibiotic Resistant Pathogens Evaluated by Different Methods. *American Journal of Applied Sciences*. Vol 11. 1773-1783.
- Aysan, E., Ayar, E. and Aren, A. (2002). The Role of Intra-peritoneal Honey Administration in Preventing Post-operative Peritoneal Adhesions. *European Journal of Obstetrics and Gynaecology and Reproductive Biology*. Vol. 104, 152-155.
- Barros, L., Duenas, M., Pinela, J., Carvalho, A.M., Buelga, C.S. and Ferreire,
 I.C.F.R. (2012). Characterization and Quantification of Phenolic Compounds in
 Four Tomato (*Lycopersicon Esculentum* L.) Farmer Varieties in Northeastern
 Portugal Homegardens. *Plant Food for Human Nutrition*. Vol. 67, 229-234.
- Basualdo, C., Sgroy, V., Finola, M.S. and Marioli, M. (2007). Comparison of the Antibacterial Activity of Honey from Different Provence Against Bacteria Usually Isolated from Skin Wounds. *Veterinary Microbiology*. Vol. 124, 375-381.
- Berg, A.J.J.V.D., Worm, E.V.D., Halkes, S.B.A., Hoekstra, M.J. and Beukelman, C.J. (2008). An In vitro Examination of the Antioxidant and Anti-inflammatory Properties of Buckwheat Honey. *Journal of Wound Care*. Vol. 17, 172-178.
- Bilsel, Y., Bugra, D. and Yamaner, S. (2002). Could Honey Have a Place in Colitis Therapy? Effects of honey, Prednisolone, and Disulfiram on Inflammation, Nitric Oxide, and Free Radical Formation. *Digestive Surgery*. Vol. 19, 306-12.

- Bogdanov, S. (2009). Honey Composition . S. Bogdanov. Bee hexagon. Botting, R.M. (2006). Inhibitors of Cyclooxygenases: Mechanisms, Selectivity and Uses. *Journal of Philosophy and Pharmacology*. Vol 5. 113-124.
- Bonnie Alvey, R.N. and Beck, D. E. (2008). Peristomal Dermatology. Clinical Colon Rectal Surgery. Vol. 21, 41-44.
- Bramer, S.E.V. (1998). An Introduction to Mass Spectrometry. Department of Chemistry, Widener University. Chester PA.
- Brose, S.A., Thuen, B.T. and Golovko, M.Y. (2011). LC/MS/MS Method for Analysis of E2 Series Prostaglandins and Isoprostanes. *Journal of Lipid Research*. Vol.52, 850-859.
- Candiracci, M., Citterio, B. and Piatti, E. (2012). Anti-fungal Activity of Honey Flavonoid Extract against *Candida albican*. *Food chemistry*. Vol., 131, 493-499.
- Cao, H., Yu, R., Tao, Y., Nikolic, D., van Breemen, R.B. (2011). Measurement of Cyclooxygenase Inhibition Using Liquid Chromatography–tandem Mass Spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*. Vol. 54, 230-235.
- Cao, H., Xiao, L., Park, G.Y., Wang, X., Azim, A.C., Christman, J. W. and Breeman, R.B.v. (2007). An Improved LC-MS/MS Method for the Quantification of Prostaglandins E2 and D2 Production in Biological Fluids. *Analytical Biochemistry*. Vol, 372, 41-51.
- Caravaca, A.M.G., Romero, M.G., Roman, D.A., Carretero and A.S., Guitierrez, A.F. (2006). Advances in the Analysis of Phenolic Compounds in Products Derived from Bees. *Journal of Pharmaceutical and Biomedical Analysis*. Vol. 41, 1220-1234.
- Chen, Y.C., Shen, S.C., Lee, W.R., Hou, W.C., Yang, L.L. and Lee, T.J.F. (2001). Inhibition of Nitric Oxide Synthase Inhibitors and Lipopolysaccharide Induced Inducible NOS and Cyclooxygenase-2 Gene Expression by Rutin, Quercetin, and Quercetin Pentaacetate in RAW 264.7 Macrophages. *Journal of Cellular Biochemistry*. Vol. 82, 537-548.
- Chua, L.S., Rahaman, N.L.A., Sarmidi, M.R. and Aziz, R. (2012). Multi-Elemental Composition and Physical Properties of Honey Samples from Malaysia. *Food Chemistry*. Vol. 135, 880-887.

- Cooper, R.A., Molan, P.C. and Harding K.G. (1999). Antibacterial Activity of Honey Against Strains of *Staphylococcus aureus* from Infected Wounds. *Journal of the Royal Society of Medicine*. Vol. 92, 283-285.
- Cooper, R., Halas, E. and Molan, P.C. (2002). The Efficacy of Honey in Inhibiting Strains of *Pseudomonas aeruginosa* from Infected Burns. *Journal of Burn Care* and Rehabilitation. Vol. 23, 366–370.
- Cushnie, T.P.T. and Lamb, A.J. (2005). Detection of Galangin-induced Cytoplasmic Membrane Damage in *Staphylococcus aureus* by Measuring Potassium Loss. *Journal of Ethno-pharmacology*. Vol. 101, 243-248.
- Das, A., Datta, S., Mukherjee, S., Bose, S., Ghosh, S., Dhar, P. (2012). Evaluation of Antioxidative, Antibacterial and Probiotic Growth Stimulatory Activities of *Sesamum indicum* honey Containing Phenolic Compounds and Lignans. *LWT-Food Science and Technology*. Vol. 2014, 1-7.
- Dimitrova, B., Gevrenova, R. and Anklam, E. (2007). Analysis of Phenolic Acids in Honey of Different Floral Origin by Solid-phase Extraction and Highperformance Liquid Chromatography. *Phytochemical Analysis*. Vol. 18, 24-32.
- Doner, L.W. (1977). The Sugars of Honey- A Review. *Journal of the Science of Food and Agriculture*. Vol. 28, 443-456.
- Edgar, J.A., Roeder, E. L. and Molyneux, R. J. (2002). Honey from Plants Containing Pyrrolizidine Alkaloids: A Potential Threat to Health. *Journal of Agriculture and Food Chemistry*. Vol. 50, 2719-2730.
- Efem, S.E.E., Udoh, K.T. and Iwara, C.I. (1992). The Antimicrobial Spectrum of Honey and its Clinical Significance. *Infection*. Vol.4, No.2.
- Estevinho, L., Pereire, A.P., Moreire, L., Dias, L.G., and Pereire, E. (2008). Antioxidant and Antimicrobial Effects of Phenolic Compounds Extracts of Northeast Portugal Honey. *Food Chem Toxicol*. Vol. 46, 374-3779.
- Ferreres, F., Viguera, C.G., Lorente, F.T. and Barberan, F.A.T. (1993). Hesperetin: A Marker of the Floral Origin of Citrus Honey. *Journal Food Science Agriculture*. Vol. 61, 121-123.
- Fox, C. (2002). Honey as a Dressing for Chronic Wounds. British Journal of Community Nursing. Vol.7. No.10.
- Freshney, R.I. *Culture of Animal Cells: A Manual of Basic Technique*. Fifth edition. New Jersey: John Wiley and Sons, 2005.

- Garcia-Lafuente, A., Guillamon, E., Villares, A., Rostagno, M.A. and Martinez, J.A. (2009). Flavanoids as Anti-Inflammatory Agents Implications in Cancer and Cardiovascular Disease. *Inflammation Research*. Vol. 58, 537-552.
- Gheldof, N., Wang, X.H. and Engeseth, N.J. (2002). Identification and Quantification of Antioxidant Components of Honeys from Various Floral Sources. *Journal of Agriculture and Food Chemistry*. Vol. 50, 5870-5877.
- Hadjmohamadi, M. R., Nazari, S. and Kamel, K. (2009). Determination of Flavonoid Markers in Honey with SPE and LC using Experimental Design. *Chromatographia*. Vol. 69, 1291-1297.
- Halco'n, L. and Milkus, K. (2004). Staphylococcus aureus and Wounds: A Review of Tea Tree Oil as a Promising Antimicrobial. American Journal of Infection Control. Vol. 32, 402–408.
- Havsteen, B. (1983). Flavonoids: Class of Natural Products of High Pharmacological Potency. *Biochemical Pharmacology*. Vol. 32, 1141–8.
- Hermosin, I., Chicon, R.M. and Cabezudo, M.D. (2003). Free Amino Acid Composition and Botanical Origin of Honey. *Food Chemistry*. Vol.84, 263-268.
- Hsueh, P. Chen, W. and Nad Luh, K. (2005). Relationship Between Anti-microbial Resistance in Gram-negative Bacteria Causing Nonsocomical Infections from 1991-2003 at a University Hospital in Taiwan. *International Journal of Antimicrobial Agents*. Vol. 26, 463-472.
- Idborg, H., Pawelzik, S.C., Manso, M.P., Bjork, L., Hamrin, J., Herlenius, E. and Jakobsson, P.J. (2014). Evaluation of Urinary Prostaglandin E2 Metabolite as a Biomarker in Infants with Fever due to Viral Infection. *Prostaglandin, Leukotrienes and Essential Fatty Acids*. Vol. 91, 269275.
- Imlay, J.A. (2013). The Molecular Mechanisms and Physiological Consequences of Oxidative Stress Lessons from A Model Bacterium. *Nature Reviews Microbiology*. Vol.11, 443-454.
- Karin, M., Lawrence, T. and Nizet, V. (2006). Innate Immunity Gone Awry Linking Microbial Infections to Chronic Inflammation and Cancer. *Cell*. Vol. 124, 823-835.
- Kassim, M., Achoui, M., Mustafa, M.R., Mohd, M.A. and Yusoff, K.M. (2010). Ellagic Acid, Phenolic Acids, and Flavonoids in Malaysian honey Extracts Demonstrate In-vitro Anti-inflammatory Activity. *Nutrition Research*. Vol. 30, 650-659.

- Kassim, M., Achoui, M., Mansor, M. and Yusoff, K.M. (2010). The Inhibitory Effects of Gelam Honey and its Extracts on Nitric Oxide and Prostaglandin E2 in Inflammatory Tissues. *Fitoterapia*. Vol. 81, 1196-1201.
- Khalil, M.I., Mahaneem, M., Jamalullail, S.M.S., Alam, N. and Sulaiman, S.A. (2011). Evaluation of Radical Scavenging Activity and Colour Intensity of Nine Malaysian Honeys of Different Origin. *Journal of ApiProduct and ApiMedical Sciences*. Vol. 3, 4-11.
- Kim, E.J, Kwon, K.J., Park, J.Y., Lee, S.H., Moon, C.H. and Baik, E.J. (2002). Effect of Perixisome Proliferator-activated Receptor Against LPSinducedneuronal Death in Mixed Cortical Neurons: Associated with iNOS and COX-2. *Brain Research*. Vol. 941, 1-10.
- Kirnpal-Kaur, B.S., Tan, H.T., Boukraa, L. and Gan, S.H. (2011). Different Solid Phase Extraction Fractions of Tualang (*Koompassia excelsa*) Honey Demonstrate Diverse Antibacterial Properties Against Wound and Enteric Bacteria. *Journal of ApiProduct and ApiMedical Science*. Vol. 3, 59-65.
- Ko, H.H., Sao, L.T., Yu, K.L., Liu, C.T., Wang, J. P. and Lin, C.N. (2003). Structure-activity Relationship Studies on Chalcone Derivatives: The Potent Inhibition of Chemical Mediators Release. *Bioorganic and Medicinal Chemistry Letters*. Vol. 11, 105-111.
- Kumar, S.V., Satyanarayana, T., Mathew, A., Ganesh, B. and Venukumar, R. (2012).
 In-vivo Anti-inflammatory Activity of Methanolic Extract of *Cissus Pallida*.
 Journal of Chemical and Pharmaceutical Sciences. Vol. 5, 113-116.
- Kwakman, P.H.S. and Zaat, S.A.J (2012). Antibacterial Components of Honey. International Union of Biochemistry and Molecular Biology. Vol. 64, 48-55.
- Lin Jiang (2011). Comparison of Disk Diffusion, Agar Dilution and Broth Microdilution for Antimicrobial Susceptibility Testing of Five Chitosans. Master of Science, Lousiana State University and Agricultural and Mechanical College.
- Liyana-Pathirana, C.M. and Shahidi, F. (2005). Antioxidant Activity of Commercial Soft and Hard Wheat (Triticum aestivum L.) as Affected by Gastric pH conditions. *Journal of Agricultural and Food Chemistry*. Vol 53, 2433-2440.
- Makawi, S.Z.A., Gadkariem, E.A. and Ayoub, S.M.H. (2009). Determination of Antioxidant Flavanoids in Sudanese Honey Samples by Solid Phase Extraction and High Performance Chromatography. *E-journal of Chemistry*. Vol. 6, 429-437.

- Malika, N., Mohamed, F. and Chakib, E.A. (2004). Antimicrobial Activities of Natural Honey from Aromatic and Medicinal Plants on Antibiotic-resistant Strains of Bacteria. *International Journal of Agriculture and Biology*. Vol. 6, 289-293.
- Marcato, P.D., De Paula, L.B., Melo, P.S., Ferreira, I.R., Almeida, A.B.A, Torsoni, A.S. and Alves, O.L. (2015). In vivo Evaluation of Complex Biogenic Silver Nanoparticle and Enoxaparin in Wound Healing. *Journal of Nanomaterials*. Vol. 2015, 1-10.
- Martos, I., Ferreres, F. and Tomas-Barberian, F.A. (2000). Identification of Flavanoid Markers for the Botanical Origin of *Eucalyptus* Honey. *Journal of Agriculture and Food Chemistry*. Vol. 48, 1498-1502.
- Martos, M.V., Navajas, Y.R., Lopez´, J.F. and A´Lvarez, J.A.P. (2008). Functional Properties of Honey, Propolis and Royal Jelly. *Journal of Food Science*.Vol.73. No. 9.
- Mato, I., Huidobro, J.F., Simal-Lozano, J. and Sancho, M.T. (2003). Significance of Non-aromatic Organics Acids in Honey. *Journal of Food Protection*. Vol.12, 2371-2376.
- Matuschek, E., Brown, D.F.J. and Kahlmeter, G. (2013). Development of the EUCAST Disk Diffusion Antimicrobial Susceptibility Testing Method and its Implementation in Routine Microbiology Laboratories. *Clinical Micorbila Infection*. Vol. 20, 255-266.
- Mulu, A., Tessema, B. and Derbie, F. (2004). In Vitro Assessment of The Antimicrobial Potential of Honey on Common Human Pathogens. *Ethiopian Journal* of *Health Development*. Vol.18, 107-118.
- Mundo, M.A., Padilla-Zakour, O.I and Worobo, R.W. (2004). Growth Inhibition of Foodborne Pathogens and Food Spoilage Organism by Select Raw Honeys. *Journal of Food Microbiology*. Vol. 97, 1-8.
- Murnaghan, I. (2013, September 2). Using Animal for Testing: Pros Versus Cons. About Animal Testing, September 2, 2013, http://www.aboutanimaltesting.co.uk/using-animals-testing-pros-versuscons.html.
- Molan, P.C. (1999). The Role of Honey in the Management of Wounds. *Journal of Wound Care*. Vol. 8. No.8.

- Molan, P.C. (2002). Not All Honeys are the Same for Wound Healing. *Bulletin of European Tissue Repair Society*. Vol. 9, 5–6.
- Molan, P.C., Cooper, R. and White, R. Honey A Modern Wound Management Product. United Kingdom: Wound-UK Books. 2005.
- Molan, P.C., The Evidence Supporting the Use of Honey a Wound Dressing. (2006). *The International Journal of Lower Extremity Wounds*. Vol. 5, 40-54.
- Moniruzzaman, M., Sulaiman, S.A., Azlan, S.A.M. and Gan, S.W. (2013). Two-year Variation of Phenolics, Flavonoids and Antioxidant Contents in Acacia Honey. *Molecules*. Vol. 18, 14694-14710.
- Moniruzzaman, M., Sulaiman, S.A., Azlan, S.A.M. and Gan, S.W. (2013).
 Physiochemical and Antioxidant Properties of Malaysian Honey Produced by *Apis cerana, Apis dorsata* and *Apis mellifera. BMC Complementary and Alternative Medicine*. Vol. 13, 1-12.
- Moussa, A., Saad, A., Djebli, N.D., Meslem, A. and Benhalima, A.E.K. (2011). Antifungal Activity of Four Honeys of Different Types from Algeria against Pathogenic Yeast: *Candida albican* and *Rhodotorula sp. International Journal* of Microbiological Research. Vol. 2, 276-279.
- Nagai, T., Inoue, R., Kanamori, N., Suzuki, N. and Nagashima, T. (2006). Characterization of Honey from Different Floral Sources. Its Functional Properties and Effects of Honey Species on Storage of Meat. *Food chemistry*. Vol. 97, 256-262.
- Nasser, S., Mabrouk, A. and Maher, A. (2003). Colonization of Burn Wounds in Ain Shams University Burn Unit. *Burns*. Vol. 29, 229–233.
- Ng, W.J., Ken, K.W., Kumar, R.V., Gunasagaran, H., Chandramogan, V. and Lee, Y.Y. (2014). Invitro Screening of Malaysian Honey from Different Floral Sources for Antibacterial Activity on Human Pathogen Bacteria. *African Journal* of Traditional, Complementary Alternative Medicine. Vol. 11, 315-318.
- Noreen, Y., Ringbom, T., Perera, P., Danielson, H. and Bohlin, L. (1998).
 Development of a Radiochemical Cyclooxygenase-1 and -2 in Vitro Assay for Identification of Natural Products as Inhibitors of Prostaglandin Biosynthesis. *Journal of Natural Product*. Vol. 61, 2-7.
- Omoya, F.O. and Akharaiyi, F.C. (2010). A Pasture Honey Trial Potency on Some Selected Pathogenic Bacteria. *Journal of Natural Products*. Vol. 3, 5-11.

- Pan, M.H., Lai, C.S. and Ho, C.T. (2010). Anti-inflammatory Activity of Natural Dietary Flavonoids. *The Royal Society of Chemistry*. Vol.1, 15-31.
- Pan, M.H., Lai, C.S., Dushenkov, S. and Ho, C.T. (2009). Modulation of Inflammatory Genes by Natural Dietary Bioactive Compound. *Journal of Agricultural and Food Chemistry*. Vol. 57, 4467-4477.
- Paradkar, P.N, Blum, P.S, Berhow, M.A, Baumann, H and Kuo, S.M. (2004). Dietary Isoflavones Supress Endotoxin-induced Inflammatory Reaction in Liver and Intestine. *Cancer Letters*. Vol. 215, 21-28.
- Patil, S. and McCormick, B.A. (2014). Mucosal Inflammatory Response to Salmonella typhimurium Infection. Frontiers in Immunology. Vol. 5, 1-10.
- Patton, T., Barrett, J., Brennan, J. and Moran, N. (2006). Use of Spectrophotometric Bioassay for Determination of Microbial Sensitivity to Manuka Honey. *Journal* of Microbiological Methods. Vol. 64, 84-95.
- Perez, R.A., Iglesias, M.T., Pueyo, E., Gonzalrez, M. and De Lorenzo, C. (2007). Amino Acid Composition and Antioxidant Capacity of Spanish Honeys. *Journal* of Agricultural and Food Chemistry. Vol. 55, 360-365.
- Pichichero, E., Canuti, L. and Canini, A. (2009). Charactersation of the Phenolic and Flavonoid Fractions and Antioxidant Power of Italian honeys of Different Botanical Origin. *Journal of Science Food Agriculture*. Vol. 89, 609-616.
- Plazonic, A., Bucar, F., Males, Z., Mornar, A., Nigovic, B. and Kujundzic, N. (2009).
 Identification and Quantification of Flavonoids and Phenolic Acids in Burr
 Parsley (*Caucalis platycarpos* L.), Using High-Performance Liquid
 Chromatography with Diode Array Detection and Electrospray Ionization Mass
 Spectrometry. *Molecules*. Vol.14, 2466-2490.
- Prakash, A., Medhi, B., Avti, P. K., Saikia, U.N., Pandhi, P. And Khanduja, K. L. (2008). Effect of different Doses of Manuka Honey in Experimentally Induced Inflammatory Bowel Dsease in Rats. *Phytotherapy Research*. Vol, 22, 1511-1519.
- Pulcini, P., Francesco, A., and Norma, F. (2006). Fast SPE Extraction and LC-ESI-MS-MS Analysis of Flavonoids and Phenolic Acids in Honey. *Apiacta*. Vol.41, 21-27.
- Pyrzynska, K. and Biesaga, M. (2009). Analysis of Phenolic Acids and Flavonoids in Honey. *Trends in Analytical Chemistry*. Vol. 28, 893-902.

- Raso, G.M., Meli, R., Carlo G.D., Pacilio, M. and Carlo, R.D. (2001). Inhibition of Inducible Nitric Oxide Synthase and Cyclooxygenase-2 Expression by Flavanoids in Macrophage J774A.1. *Life Sciences*. Vol. 68, 921-991.
- Rijke, E.D., Out, P., Niessen, W.M.A., Ariese, F., Goojier, C. and Brinkman, U.A.T. (2006). Analytical Separation and Detection Methods for Flavonoids. *Journal of Chromatography A*. Vol.1112, 31-63.
- Rotelli, A.E, Guardis, T., Juarez, A.O, Rocha, N. E. D. L. and Pelzer, L.E. (2003). Comparative Study of Flavonoids in Experimental Models of Inflammation. *Pharmacological Research*. Vol. 48, 601-606.
- Russell, K., Molan, P., Wilkins, A., Holland, P., (1990). Identification of Some Antibacterial Constituents of New Zealand Manuka Honey. *Journal of Agriculture Food Chemistry*. Vol. 38, 10–13.
- Salam, H.H.A. and Raju, R. (2009). In-vivo Screening Methods of Antiinflammatory Drugs-An Approach to Herbal Drugs. B.Pharm Projects.
- Sanagi, M.M., Sulaiman, A. and Ibrahim, W.A.W. Principle of Chemical Analysis. Universiti Teknologi Malaysia. Department of Chemistry, Faculty of Science. 2007.
- Sherlock, O., Dolan, A., Athman, R., Power, A., Gethin, G., Cowman, S. and Humphreys, H. (2010). Comparison on the Antimicrobial Activity of Ulmo Honey from Chile and Manuka Honey Against Methicillin-resistant Staphylococcus aureus, Escherichia coli and Pseudomonas aeruginosa. Biomedical Central Complementary and Alternative Medicine. Vol.10, 1-5.
- Snow, M. and Manley-Harris, M. (2004). On the Nature of Non-peroxide Antibacterial Activity in New Zealand Manuka honey. *Food Chemistry*. Vol. 84, 145–147.
- Srisayam, M. and Chantawannakul, P. (2010). Antimicrobial and Antioxidant Properties of Honeys Produced by *Apis mellifera* in ThailanD. *Journal of Apiproduct and Apimedical Science*. Vol.2, 77-83.
- Stalikas, C. D. (2007). Extraction, Separation and Detection Methods for Phenolic Acids and Flavanoids. *Journal of Separation Science*. Vol. 30, 3268-3295.
- Subrahmanyam, M. A. (1998). Prospective Randomised Clinical and Histological Study of Superficial Burn Wound Healing with Honey and Silver Sulfadiazine. *Burns*. Vol. 24, 157-61.

- Subrahmanyam, M., Sahapure, A.G. and Nagane, N.S. (2001). Effects of Topical Application of Honey on Burn Wound Healing. *Ann Burns Fire Disasters*. Vol. 14, 143-5.
- Subrahmanyam, M., Shahapure, A.G. and Nagane, N.S. (2003). Free Radical Control-The Main Mechanism of the Action of Honey in Burns. Ann Burns Fire Disasters. Vol. 16, 135-138.
- Sun, J., Liang, F., Bin, Y., Li, P. and Duan, C. (2007). Screening Non-colored Phenolics in Red Wines Using Liquid Chromatography/ Ultraviolet and Mass Spectrometry/Mass Spectrometry Libraries. *Molecules*.Vol.12, 679-693.
- Tan, H.T., Rahman, R.A., Gan, S.H., Halim, A.S., Hassan, S.A., Sulaiman, S.A. and Kimpal,- Kaur, B.S. (2009). The Antibacterial Properties of Malaysian Tualang Honey Against Wound and Enteric Microorganisms In Comparison to Manuka Honey. *Biomedical Central complementary and Alternative Medicine*. Vol. 9.
- Tasleem, S. and Naqvi, S.B.S. (2014). Comparisson of Invitro Antimicrobial Activity of Different Crude Pakistani Honey Samples and Commercials Antibiotics Against Clinical Pathogens. *World Journal of Pharmaceutical Research*. Vol. 3, 88-104.
- Teixeira, É.W., Negri G., Meira, R.M.S.A., Message, D. and Salatino, A. (2005). Plant Origin of Green Propolis: Bee Behaviour, Plant anatomy and Chemistry. *Evidence-based Complementary and Alternative Medicine*. Vol. 2, 85-92.
- Tistaert, C., Dejaegher, B., and Heyden, Y. V. (2011). Chromatographic Separation Techniques and Data Handling Methods for Herbal Fingerprint : A Review. *Analytica Chimica Acta*. Vol. 690, 148-161.
- Tonks, A.J., Cooper, R.A., Jones, K.P., Blair, S., Parton, J. and Tonks, A. (2002). Honey Stimulates Inflammatory Cytokine Production from Monocytes. *Cytokine*. Vol. 21, 242-247.
- Tumin, N., Halim, N.A.A., Hahjahan, M., Noor Izani, N.J., Sattar, M.A., Khan, A.H. and Mohsin, S.S.J (2005). Antibacterial Activity of Local Malaysian Honey. *Malaysian Journal of Pharmaceutical Sciences*. Vol. 3, 1-10.
- Vallianou, N.G., Gounari, P., Skourtis, A., Panagos, J. and Kazazis, C. (2014). Honey and its Anti-inflammatory, Anti-bacterial and Anti-oxidant Properties. *General Medicine*. Vol. 2, 1-5.
- Vuong, C. and Otto, M. (2002). Staphylococcus epidermidis Infections. Microbes and Infection. Vol. 4, 481–489.

- Wahdan, H.A.L. (1998). Causes of the Antimicrobial Activity of Honey. *Infection*. Vol. 26. No.1.
- Wang, L., Tu, Y.C., Lian, T.W., Hung, J.T., Yen, J.H and Wu, M.J. (2006). Distinctive and Anti-inflammatory Effects of Flavonols. *Journal of Agricultural* and Food Chemistry. Vol. 54, 9798-9804.
- White, J.W., Subers, M.H. and Schepartz, A.L. (1962). The Identification of Inhibine. *American Bee Journal*. Vol. 102, 430–431.
- Zainol, M.I., Yussof, K.M. and Yusof, M.Y.M. (2013). The Antibacterial Activity of Selected Malaysian Honey. *Biomedical Central and Alternative Medicine*. Vol.13, 129.