

**PRODUCTION AND OPTIMIZATION OF INDOLE-3-ACETIC ACID BY
*RHODOPSEUDOMONAS PALUSTRIS***

LEONG PEI PEI

UNIVERSITI TEKNOLOGI MALAYSIA

PRODUCTION AND OPTIMIZATION OF INDOLE-3-ACTIC ACID BY
RHODOPSEUDOMONAS PALUSTRIS

LEONG PEI PEI

A thesis submitted in fulfillment of
the requirements for the award of the degree of
Master of Engineering (Bioprocess)

Faculty of Chemical Engineering
Universiti Teknologi Malaysia

JULY 2015

Specially dedicated to *Mom and Dad*

I really love both of you.

ACKNOWLEDGEMENS

Foremost, I would like to express my immense gratitude to my supervisor Ass Prof. Dr Lee Chew Tin for her continuously support this project. Thank you for her patient, motivation, knowledge and encouragement. I am also very grateful for the support provided by my co-supervisors Dr Chua Lee Suan and Professor Mohammad Roji for their continuous support and contributions to this thesis, especially to Dr Chua Lee Suan for help me to conduct the LC-MS. Moreover, a very big thank to lab member Ong Pei Ying, Cassendra Bong to provide me some guidance throughout the research. Nevertheless, I would also like to thank to all the lab technicians for allow me to use the lab and facility at the Institute of Bioproduct Development

ABSTRACT

Indole-3-acetic acid (IAA) is known to be an important phytohormone that helps to regulate plant growth and development. In this study, the optimum culture medium for the production of IAA by *Rhodopseudomonas palustris* in shake flask culture was studied. *Rhodopseudomonas palustris* is a purple non-sulfurbacteria which has been well recognised as one of the most metabolically versatile bacteria. The research was divided into three parts. First, a pre-screening process based on Taguchi Design was conducted to identify the significant factors that could affect the production of IAA. The pre-screening indicated that three parameters were found to be significant, which include the concentration of tryptophan, glucose and potassium nitrate. These parameters were selected and used to optimize the production IAA by Response Surface Methodology (RSM). Lastly, a kinetic study for the bacterial growth and IAA production was investigated. The optimal amount of IAA was obtained after incubation of 48 hours at 35 °C in the presence of 5 g L⁻¹ of tryptophan, 4.94 g L⁻¹ of glucose and 0.60 g L⁻¹ of KNO₃, as recommended by the RSM. Under this condition, the experimental yield of IAA production was 80.77± 2.13 µg mL⁻¹, which was in close agreement with the value predicted by the RSM model (77.64 µg mL⁻¹). This was the highest yield of IAA that was reported compared to the IAA yields obtained from the 20 experiments designed under the RSM. The IAA production depends on growth stage as most of the IAA was produced during the stationary growth phase of *Rhodopseudomonas palustris*. This study has successfully optimized the production of IAA by *Rhodopseudomonas palustris* by statistical approach and proved that *Rhodopseudomonas palustris* has the potential to be used as plant bioenhancer or biofertiliser for plant growth development.

ABSTRAK

Asid indola-3-asetik (IAA) merupakan fitohormon yang dapat menggalakan dan mengawal pertumbuhan dan perkembangan tumbuh-tumbuhan. Dalam kajian ini, medium kultur yang optimum bagi penghasilan IAA oleh *Rhodopseudomonas palustris* dalam kelalang gochang telah dikaji. *Rhodopseudomonas palustris* adalah bakteria ungu bukan sulfur yang telah mempunyai metabolisme yang amat versatil. Kajian ini dibahagikan kepada tiga bahagian. Pertama, proses pra-saringan telah dijalankan untuk mengenalpasti faktor-faktor penting yang menentukan penghasilan IAA. Daripada proses pra-saringan tersebut, tiga parameter telah dikenalpasti sebagai parameter yang signifikan, yakni kepekatan triptofan, glukosa dan kalium nitrat. Pada bahagian yang kedua, ketiga-tiga parameter ini telah dipilih dan digunakan untuk mengoptimumkan penghasilan IAA melalui pendekatan statistik, iaitu Metodologi Permukaan Respon (RSM). Akhirnya, pertumbuhan bakteria dan penghasilan IAA telah dikaji. Penghasilan optimum IAA dicapai selepas penderaman selama 48 jam pada suhu 35 °C dengan kehadiran 5 g L⁻¹ triptofan, 4.94 g L⁻¹ glukosa dan 0.60 g L⁻¹ kalium nitrat, seperti yang dicadangkan oleh kaedah RSM. Dalam keadaan ini, hasil eksperimen pengeluaran IAA adalah 80.77 ± 2.13 g mL⁻¹, nilai ini agak hampir dengan nilai yang diramalkan oleh model RSM (77.64 g mL⁻¹). Ini adalah nilai hasil IAA yang tertinggi berbanding dengan hasil IAA yang diperolehi daripada 20 ujikaji lain yang disarankan oleh kaedah RSM. Kadar penghasilan IAA juga didapati berkait rapat dengan pertumbuhan sel, dimana kebanyakan IAA telah dihasilkan semasa fasa pertumbuhan statik sel *Rhodopseudomonas palustris*. Kajian ini telah berjaya dioptimumkan pengeluaran IAA dan membuktikan bahawa *Rhodopseudomonas palustris* mempunyai potensi untuk digunakan sebagai bio-peringkat tumbuhan atau bio-baja untuk menumbuhkan tumbuh-tumbuhan.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	x
	LIST OF FIGURES	xii
	LIST OF APPENDIXS	xvi
	LIST OF ABBREVIATIONS	xiv
	LIST OF SYMBOLS	xvi
1	INTRODUCTION	1
	1.1 Research Background	1
	1.2 Problem Statement	4
	1.3 Research Objective	6
	1.4 Scopes of Study	6
2	LITERATURE REVIEW	
	2.1 Production of Plant Growth Promoting Compound by Plant Growth-Promoting Bacteria	7
	2.2 <i>Rhodopseumonas Palustris</i> as Model Microbe for Plant Growth Promoting Bacteria	7

2.3	Auxin	12
2.4	IAA and IAA-Producing Bacteria	14
2.5	Biosynthesis Pathway of IAA	14
2.5.1	Tryptophan-dependent Pathway	16
2.5.2	Tryptophan-independent Pathway	18
2.6	The Environmental Factors That Affect the Biosynthesis of IAA in Bacteria	19
2.6.1	Effect of Tryptophan	19
2.6.2	Effect of Carbon Sources	20
2.6.3	Effect of pH	21
2.6.4	Effect of Nitrogen Source	22
2.6.5	Selection of Parameters	23
2.7	Optimization of IAA by DOE	24
2.7.1	Pre-Screening Process by DOE- Taguchi Design	27
2.7.2	Optimization of IAA by DOE-Response Surface Methodology (RSM)	30
2.8	Optimization of IAA by DOE-Response Surface Methodology (RSM)	35
2.9	Methods for IAA Analysis	37
2.9.1	Salkowski Assay	40
3	METHODOLOGY	42
3.1	Summary	42
3.2	Chemicals	44
3.3	<i>Rhodopseudomonas Palustris</i> and Its Growth Condition	44
3.4	Selection of culture medium for IAA production	45
3.5	Analytical Determination	47
3.5.1	Bacterial Cells Concentration	47
3.5.2	Analyses of IAA	48
3.5.2.1	Salkowski Assay	48
3.6	Pre-Screening of IAA Production by DOE-Taguchi Design	49

3.6.1	Selection of Parameters	52
3.6.2	ANOVA Analysis	53
3.7	Optimization for the Production of IAA by DOE - RSM	53
3.7.1	ANOVA Analysis	55
4	RESULTS AND DISCUSSION	56
4.1	Summary	56
4.2	Selection of the Culture Medium for IAA Production	57
4.3	Pre-Screening for the Significant Factors of IAA Production by Taguchi Design	60
4.3.1	Effect of Carbon Source on Production of IAA	65
4.3.2	Effect of Tryptophan on IAA Production	67
4.3.3	Effect of Nitrogen Source on IAA Production	70
4.3.4	Effect of pH On IAA Production	71
4.3.5	Selection of Significant Factors for IAA Production by Taguchi Design	72
4.4	Optimization of IAA Production by Response Surface Methodology (RSM)	74
4.4.1	Model Validation	83
4.5	IAA Production Profile by <i>Rhodopseudomonas palustris</i> and its IAA production under optimal condition.	85
4.6	Overall Discussion for the Production of IAA by <i>R. Palustris</i> As Compared To Previous Studies	88
5	CONCLUSION	90
	REFERENCES	92
	Appendices A-C	101-110

LIST OF TABLES

TABLE NO.	TITLE	PAGE
1.1	Application of auxin-like substances in agriculture (Hofrichter, 2010)	3
2.1	Characteristic and growth substrates of <i>R. palustris</i> (Guerrero, 2001)	10
2.2	IAA produced by microorganisms and their effect on plant morphology and development	14
2.3	The optimization method for production of IAA indifferent bacterial strain	25
2.4	Characteristic and formulas for type of S/N ratio in DOE-Taguchi Design	32
3.1	Nutrition contents in three different mediums	46
3.2	Different factors and their levels in the experiment for IAA production by Taguchi Design.	49
3.3	Level and Actual Value for Taguchi Design derived from DOE 8.0.6	50
3.4	Fixed Parameters for Taguchi Design	51
3.5	Coded values of variables used in the central composite design of RSM	53

4.1	Production of IAA in four different mediums	57
4.2	Experimental results for the production of IAA for the 16 experiments as designed by the DOE-Taguchi Method	61
4.3	Analysis of Variance (ANOVA) for the experimental results obtained for the experimental design by the DOE- Taguchi Design	62
4.4	Optimum conditions and their contribution	73
4.5	Central composite design matrix design with the experimental values of IAA (<i>Y</i>) produced by <i>R. palustris</i>	75
4.6	Regression analysis on the RSM for the production of IAA by <i>R. palustris</i>	77
4.7	Model coefficient estimated by student's t-test	78
4.8	Comparison of IAA yield against previous studies	89

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Four growth modes of <i>Rhodopseudomonas palustris</i> (Larimer <i>et al.</i> , 2004)	9
2.2	Structure of Indole-3-acetic acid (C ₁₀ H ₉ NO ₂)	12
2.3	Model of phototropism in shoot (Kimura and Kagawa, 2006).	13
2.4	Biosynthesis of IAA (Zhao, 2010)	17
2.5	Layout of two-by-two factorial design on the left, whereas OFAT experiment is shown at the upper right.	28
2.6	Flow diagram for the optimization of IAA production	29
2.7	Standard procedures for Taguchi Design	31
2.8	Array Selectors for Taguchi Design.	34
2.9	Three-dimensional surface and the corresponding contour plot, where x_i and x_j are parameter 1 and parameter 2 respectively.	36
2.10	Bacterial growth curve	38
2.11	Salkowski reagent reacted with IAAs to form a pink colour chelates.	41
2.12	Simplified diagram and function of instrument of LC-MS	43
3.1	Flow sheet of the experimental work	43
4.1	The profile of <i>R. palustris</i> cell concentration, g L ⁻¹ (bar) and IAA concentration, µg mL ⁻¹ (line) achievable in four culture mediums. All mediums were supplemented with 1mg mL ⁻¹ of tryptophan.	58
4.2	Percentage of contribution of all four factors (tryptophan, carbon source, nitrogen source and pH) for the production of IAA production.	64
4.3	Effect of type of carbon source for the production of IAA by <i>R. palustris</i> .	66

4.4	Effect of amount of tryptophan (g L^{-1}) on IAA production ($\mu\text{g mL}^{-1}$)	69
4.5	Effect of nitrogen source on production of IAA by <i>R. palustris</i>	70
4.6	Effect of pH on IAA production	72
4.7	Two- and three-dimensional contour plots of the combined interaction effect of tryptophan (g L^{-1}) and glucose (g L^{-1}) for the production of IAA by <i>R. palustris</i> .	79
4.8	Two- and three-dimensional contour plots of the combined interaction effect of tryptophan (g L^{-1}) and KNO_3 (g L^{-1}) for the production of IAA by <i>R. palustris</i>	80
4.9	Two- and three-dimensional contour plots of the combined interaction effect of amount of glucose (g L^{-1}) and KNO_3 (g L^{-1}) for the production of IAA by <i>R. palustris</i> .	81
4.10	Comparison of actual production of IAA versus the predicted IAA production by <i>R. palustris</i>	84
4.11	Normal probability of Internally studentized residuals	84
4.12	Grow curve for <i>R. palustris</i> and its IAA production in the <i>Rhodopseudomonas</i> medium supplemented with 5 g L^{-1} of tryptophan, 4.94 g L^{-1} of glucose and 0.60 g L^{-1} of KNO_3	87

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Calibration Data	109
B	Raw Data	113
C	Calculation Formula	117

LIST OF ABBREVIATIONS

3D	-	Three-Dimensional
ACC	-	1-Aminocyclopropane-1-Carboxylate
ALA	-	5-Aminolevulonic Acid
ANOVA	-	Analysis Of Variance
AO1	-	Aldehyde Protein
AR	-	Analytical Reagent
C18	-	Column 18
CCD	-	Central Composite Design
DOE	-	Design Of Experiment
EM	-	Effective Microorganism
Eq	-	Equation
<i>GC-MS</i>	-	Gas Chromatography-Mass Spectrometry
H ₂	-	Hydrogen Gas
H ₂ O	-	Water
<i>HPLC</i>	-	High-Performance Liquid Chromatography
IAA	-	Indole-3-Acetic Acid
IAAld	-	Indole -3-Acetaldehyde
IAM	-	Indole-3-Acetamide
IAN	-	Indole -3-Acetonitrile
IAOx	-	Indole-3-Acetaldoxime

IPA	-	Indole-3-Pyruvic Acid
KNO ₃	-	Potassium Nitrate
LC-MS	-	Liquid Chromatography-Mass Spectrometry
m/z	-	Mass-To-Charged
MRM	-	Multiple Reaction Monitoring
NaNO ₃	-	Nitrate
OD	-	Optical Density
OFAT	-	One-Factor-At-A-Time
PGPB	-	Plant Growth-Promoting Bacteria
<i>p</i> -values	-	Probability Value
<i>R. palustris</i>	-	<i>Rhodopseudomonas palustris</i>
RNA	-	Ribonucleic Acid
<i>RSM</i>	-	Response Surface Methodology
S/N	-	Signal-To-Noise
<i>sp</i>	-	Species
sur1	-	Supperroot
TAM	-	Tryptamine
Tryp	-	Tryptophan
UV	-	Ultra Violet
YUCCA	-	Favin Monooxygenase-Like Protein

LIST OF SYMBOLS

a.u.	-	Arbitrary units
%	-	Percentage
v/v	-	Volume per volume
min	-	Minute
v	-	Voltage
SS	-	Sum of square
R^2	-	Regression coefficient
df	-	Degree of freedom
h	-	Hour
$^{\circ}\text{C}$	-	Degree Celsius
rpm	-	Rotation per minute
g L^{-1}	-	Gram per liter

CHAPTER 1

INTRODUCTION

a. 1.1 Research Background

One of the major challenges for the twenty first century will be to create an environmental and sustainable crop production. In the current agricultural practices, improper using of the chemical fertiliser and pesticide have lead to a long list of environmental and health problems (Gunnell *et al.*, 2007; Leach and Mumford, 2008). Moreover, new emerging and treatening plant disease continue to challenge the plant biosecurity and health worldwide (Miller *et al.*, 2009). Altogether was caused increasing the demand of using ecologically compatible strategies in the agricultural sectors, for example using the beneficial bacterial to increase the crop productivity. Plant growth-promoting bacteria (PGPB), is a group of beneficial bacteria which can offer a diverse functions for plant growth and at the same time fullfill a promising solution for an environmentally friendly and sustainable agriculture. The PGPB able to enhance the plant growth by increasing the nutrient availability, release of phytohormone for phytostimulation, plant strengthening and biocontrolling.

Phytohormones are low molecular weight signal molecules that are naturally occurring and capable to influence plant growth and development in low concentration. Among the five major groups of phytohormones, indole-3-acetic acid (IAA) has been well recognized for pivotal functions in nearly every aspect related to plant growth and architecture. For examples, cell division, cell elongation, apical dominance, adventitious and lateral roots initiation, and cell and vascular differentiation (Chen *et al.*, 2009).

Biosynthesis of IAA can be found in the plants and microorganisms. The biosynthesis process occurs through several pathways, such as indole-3-pyruvic acid (IPA) pathway, tryptamine (TAM) pathway, indole-3-acetamide pathway (IAM) and indole-3-acetaldoxime (IAOx) pathways. The biosynthesis of IAA in the microorganism could be very comprehensive because the level of the IAA biosynthesis can be altered by various environmental and genetic factors. The environmental factors include the presence of tryptophan, carbon source, nitrogen source, pH, temperature. The production of IAA by *Az. Braisilense* and the expression of key gene *ipdC* have been found to be decreased during the reduction of growth rate, carbon limitation and under acidic condition (Ositadinma Ona *et al.*, 2005; Vande Broek *et al.*, 2005). In term of genetic factor, it was found that the biosynthesis of IAA can be affected by the gene location, mode of gene expression and presence of transcriptional regulators across the microorganism (Spaepen *et al.*, 2007).

Basically, auxin-type plant regulators are the oldest compounds that used in the agricultural sectors. After IAA was identified, it was chemically synthesized into the industry (Hofrichter, 2010). Currently a number of synthetic auxin-like substances such as indole-butyric acid (IBA), 2,4dichlorophenoxyacetic acid (2,4-D), and naphthalene acid (NAA) were found to be have similar effects to IAA on plant growth development (Hofrichter, 2010). The applications of these auxin-like substances in the agricultural sector were shown in Table 1.1.

Table 1.1: Application of auxin-like substances in agriculture (Hofrichter, 2010)

Auxin-like compounds	Application
Indole-3-acetic acid	Cell enlarger, disease controller, anti-transpirant fruit ripening inhibitor
indole-butyric acid (IBA)	Stimulation of root development in the propagation of stem cuttings
2,4 dichlorophenoxyacetic acid (2,4-D)	As herbicide: stimulates uncontrolled growth in broadleaf weeds in grasses
naphthalene acid (NAA)	Reduction of excessive fruit set to avoid development of many small fruits
4-Chlorophenoxyacetic acid (4-CPA)	Increase of fruit set in tomato and other solanaceous plants
Indole-butyric acid (IBA) 2,4-D together with gibberellic acid (GA ₃)	Prolongation of the pre-harvest and post-harvest life of navel oranges
2,4-Dichlorophenoxyacetic acid (2,4-D) naphthalene-acetic acid (NAA)	Delay of fruit abscission and senescence of the fruit button in grapefruit, prevention of fruit drop of apple, pear and lemon

In order to meet the demand from the market, a statistical approach was used to optimize the production of IAA. The Taguchi Design and Response Surface Methodology (RSM) are powerful statistical tools used for identifying the significant factors and optimization of the production. Both techniques have significant advantages compared to the conventional methods. For examples, they require less labour and time compared to other approaches. These methods have successfully been applied for the optimization of media and culture conditions in many cultivation processes for the production of primary and secondary metabolites, for instance enzymes, amino acids, ethanol and flavouring compounds.

b. 1.2 Problem Statement

Auxin is an important plant hormone that could influence the physiological processes of plant growth by modulating their development events, such as embryogenesis, root initiation, apical dominance, gravitropism and phototropism. Currently, chemical auxins are the most common phytohormone used in the market such as indole-3-acetic acid (IAA), 2,4-dichlorophenoxyacetic acid, 2,4,5-trichlorophenoxy acetic acid, α -naphthalene acetic acid, 2-methoxy-3,6-dichlorobenzoic acid and 4-amino-3,5,6-trichloropicolinic acid for promoting plant growth. However excessive use of these chemical or synthetic auxins are not sustainable for soil and environment as the chemical auxin could increase heavy metal in the soil, nutrient imbalance and soil acidification by changing the aggregation degree of the potassium nitrate (KNO_3) and nitrate ($NaNO_3$) in the soil (Savci, 2012). In addition, public are increasing concern on the quality of foods and health, as well as on their nutritional properties which stimulate the agriculture trend to a more sustainable and environmental friendly approach. In this context, application of soil microorganisms with beneficial activities on plant growth represents an attractive alternative approach as compared to the conventional agriculture that uses chemical or synthetic fertilizer (Choudhary *et al.*, 2011; Miransari, 2011; Verma *et al.*, 2011).

The release of tryptophan in the root exudates may results in its conversion into IAA by PGPB. In the previous study found that PGPB from different genera (Alcali gene faecalis, Enterobacter, Azospirillum, Klebsiella) and fungi have shown to enhance plant growth by synthesis of IAA (Reineke *et al.* 2008; Torres-Rubio *et al.*, 2008). The *Streptomyceslydicus* WYEC108 and *Streptomycesgriseoviridis* K61 are used commercially for IAA production under the trade of Mycostop(Khamna *et al.* 2010).

Rhodopseudomonas palustris, a purple nonsulfur photosynthetic bacteria with extraordinary diversity of enzymes, can be considered as one of the promising PGPB for natural soil enhancer compared to other microorganisms. It is not only able to fix carbon dioxide from atmosphere into biomass, but also fixing the nitrogen gas to form ammonia source for plants (Larimer *et al.*, 2004). Nevertheless, there are evidences from the genetic traits showing the capability of *R. palustris* to synthesis IAA. These features enable *R. palustris* to exhibit potential to enhance plant productivity by increasing the nitrogen availability, release of functional phytohormones and even detoxifying contaminated soil from overuse of chemical fertilizers and pesticides (Chen *et al.*, 2007; Elder and Kelly, 1994; Kim *et al.*, 2004; Liu *et al.*, 2011). To date, the use of *R. palustris* for agriculture application is seldom been reported, particularly for the production of phytohormone such as the IAA. Therefore, in this study, the *R. palustris* strain NRRL-B4276 was chosen for validation as a candidate strain to produce IAA via submerged fermentation

The level of IAA biosynthesis by bacteria can be influenced by a number of factors, such as bacteria strain, concentration of tryptophan, nutrient availability, pH, temperature and time of incubation (Apine and Jadhav, 2011; Chaiharn and Lumyong, 2011). Thus, it is necessary to consider which type of factors would affect the production of IAA in the *R. palustris*. In the present study, four key factors: the amount of tryptophan, types of carbon and nitrogen sources and pH was chosen by the Taguchi Design under the pre-screening process. Subsequently, only the significant factors were chosen and proceeded for the optimisation study by RSM analysis.

c. 1.3 Research Objective

The objective of this study is to optimize the key parameters for the production of IAA by *Rhodopseudomonas palustris* under submerged fermentation using statistical analysis approach.

d. 1.4 Scopes of Study

The scopes of the study are to:

- a) determine the significant factors that affect the production of IAA by *R. palustris* using Taguchi Design as pre-screening step;
- b) optimize the parameters of IAA production by *R. palustris* using RSM analysis

REFERENCES

- Apine, O. A., Jadhav, J. P. (2011). *Optimization of medium forⁱ indole-3-acetic acid production using Pantoea agglomerans strain PVM*. J Appl Microbiol, 110(5), 1235-1244.ⁱⁱ
- Baudoin, E., Lerner, A., Mirza, M. S., El Zembrany, H., Prigent-Combaret, C., Jurkevich, E., et al. (2010). *Effects of Azospirillum brasilense with genetically modified auxin biosynthesis gene ipdC upon the diversity of the indigenous microbiota of the wheat rhizosphere*. Res Microbiol, 161(3), 219-226.
- Bennett, K. D. (1998). *The power of movement in plants*. Trends Ecol Evol, 13(9), 339-340.
- Berg, G. (2009). *Plant-microbe interactions promoting plant growth and health: perspectives for controlled use of microorganisms in agriculture*. Applied Microbiology and Biotechnology, 84(1), 11-18.
- Brandl, M. T., and Lindow, S. E. (1996). *Cloning and characterization of a locus encoding an indolepyruvate decarboxylase involved in indole-3-acetic acid synthesis in Erwinia herbicola*. Appl Environ Microbiol, 62(11), 4121-4128.
- Carreno-Lopez, R., Campos-Reales, N., Elmerich, C., and Baca, B. E. (2000). *Physiological evidence for differently regulated tryptophan-dependent pathways for indole-3-acetic acid synthesis in Azospirillum brasilense*. Mol Gen Genet, 264(4), 521-530.

- Carrillo, A. E., Li, C. Y., and Bashan, Y. (2002). *Increased acidification in the rhizosphere of cactus seedlings induced by Azospirillum brasilense*. *Naturwissenschaften*, 89(9), 428-432
- Chaiharn, M., and Lumyong, S. (2011). *Screening and optimization of indole-3-acetic acid production and phosphate solubilization from rhizobacteria aimed at improving plant growth*. *Curr Microbiol*, 62(1), 173-181.
- Chakrabarti, J., Chatterjee, S., Ghosh, S., Chatterjee, N. C., and Dutta, S. (2010). *Synergism of VAM and Rhizobium on production and metabolism of IAA in roots and root nodules of Vigna mungo*. *Curr Microbiol*, 61(3), 203-209.
- Chen, Zhang, B., Hicks, L. M., Wang, S., and Jez, J. M. (2009). *A liquid chromatography-tandem mass spectrometry-based assay for indole-3-acetic acid-amido synthetase*. *Anal Biochem*, 390(2), 149-154.
- Chen, C.-Y., Lu, W.-B., Wu, J.-F., and Chang, J.-S. (2007). *Enhancing phototrophic hydrogen production of Rhodospseudomonas palustris via statistical experimental design*. *International Journal of Hydrogen Energy*, 32(8), 940-949.
- Chen, Q., Zhang, B., Hicks, L. M., Wang, S., and Jez, J. M. (2009). *A liquid chromatography-tandem mass spectrometry-based assay for indole-3-acetic acid-amido synthetase*. *Anal Biochem*, 390(2), 149-154.
- Chen, Q., Zhang, B., Hicks, L. M., Wang, S., and Jez, J. M. (2009). *A liquid chromatography-tandem mass spectrometry-based assay for indole-3-acetic acid-amido synthetase*. *Analytical Biochemistry*, 390(2), 149-154.
- Choorit, W., Saikur, A., Chodok, P., Prasertsan, P., and Kantachote, D. (2011). *Production of biomass and extracellular 5-aminolevulinic acid by Rhodospseudomonas palustris KG31 under light and dark conditions using volatile fatty acid*. *Journal of Bioscience and Bioengineering*, In Press, Corrected Proof.

- Choudhary, D. K., Sharma, K. P., and Gaur, R. K. (2011). *Biotechnological perspectives of microbes in agro-ecosystems*. *Biotechnol Lett*, 33(10), 1905-1910.
- Condra, L. *Design of experiments Value-added Management with Design of Experiments*. Netherlands:Springer .14-54; 1995
- Cooke, T. J., Poli, D., Sztein, A. E., and Cohen, J. D. (2002). *Evolutionary patterns in auxin action*. *Plant Mol Biol*, 49(3-4), 319-338.
- Datta, C., and Basu, P. S. (2000). *Indole acetic acid production by a Rhizobium species from root nodules of a leguminous shrub, Cajanus cajan*. *Microbiological Research*, 155(2), 123-127.
- Dimkpa, C. O., Svatos, A., Dabrowska, P., Schmidt, A., Boland, W., and Kothe, E. (2008). *Involvement of siderophores in the reduction of metal-induced inhibition of auxin synthesis in Streptomyces spp.* *Chemosphere*, 74(1), 19-25.
- Dimkpa, C. O., Zeng, J., McLean, J. E., Britt, D. W., Zhan, J., and Anderson, A. J. (2012). *Production of indole-3-acetic acid via the indole-3-acetamide pathway in the plant-beneficial bacterium Pseudomonas chlororaphis O6 is inhibited by ZnO nanoparticles but enhanced by CuO nanoparticles*. *Appl Environ Microbiol*, 78(5), 1404-1410.
- Elder, D. J., and Kelly, D. J. (1994). *The bacterial degradation of benzoic acid and benzenoid compounds under anaerobic conditions: unifying trends and new perspectives*. *FEMS Microbiol Rev*, 13(4), 441-468.
- Fibach-Paldi, S., Burdman, S., and Okon, Y. (2012). *Key physiological properties contributing to rhizosphere adaptation and plant growth promotion abilities of Azospirillum brasilense*. *FEMS Microbiol Lett*, 326(2), 99-108.

- Ghosh, S., and Basu, P. S. (2006). *Production and metabolism of indole acetic acid in roots and root nodules of Phaseolus mungo*. Microbiological Research, 161(4), 362-366.
- Gibon, Y., and Rolin, D. Aspects of Experimental Design for Plant Metabolomics Experiments and Guidelines for Growth of Plant Material. In: Hardy & R. D. Hall (ed.), *Plant Metabolomics*. NorthWest Washington:Humana Press. 860:13-30; 2012
- Guerrero, R. (2001). *Bergey's manuals and the classification of prokaryotes*. Int Microbiol, 4(2), 103-109.
- Gunnell, D., Eddleston, M., Phillips, M., and Konradsen, F. (2007). *The global distribution of fatal pesticide self-poisoning: Systematic review*. BMC Public Health, 7(1), 357.
- Gutierrez, C. K., Matsui, G. Y., Lincoln, D. E., and Lovell, C. R. (2009). *Production of the phytohormone indole-3-acetic acid by estuarine species of the genus Vibrio*. Appl Environ Microbiol, 75(8), 2253-2258.
- Higa, T., and Parr, J. F. (1994). *Beneficial and effective microorganisms for a sustainable agriculture and environment* (Vol. 1): International Nature Farming Research Center Atami,, Japan.
- Hotta, Y., Tanaka, T., Takaoka, H., Takeuchi, Y., and Konnai, M. (1997). *Promotive effects of 5-aminolevulinic acid on the yield of several crops*. Plant Growth Regulation, 22(2), 109-114.
- Idris, E. E., Iglesias, D. J., Talon, M., and Borriss, R. (2007). *Tryptophan-dependent production of indole-3-acetic acid (IAA) affects level of plant growth promotion by Bacillus amyloliquefaciens FZB42*. Mol Plant Microbe Interact, 20(6), 619-626.
- Iordache, O. (2012). *Design of Experiments Self-Evolvable Systems*. BerlinHeidelberg: Springer.217-238

- Jurków, D., and Stiernstedt, J. (2014). *Investigation of High Temperature Co-fired Ceramics sintering conditions using Taguchi Design of the experiment*. *Ceramics International*, 40(7, Part B), 10447-10455.
- Kamilova, F., Kravchenko, L. V., Shaposhnikov, A. I., Azarova, T., Makarova, N., and Lugtenberg, B. (2006). *Organic acids, sugars, and L-tryptophane in exudates of vegetables growing on stonewool and their effects on activities of rhizosphere bacteria*. *Mol Plant Microbe Interact*, 19(3), 250-256.
- Kantachote, D., Torpee, S., and Umsakul, K. (2005). *The potential use of anoxygenic phototrophic bacteria for treating latex rubber sheet wastewater*. *Electronic Journal of Biotechnology*, 8, 0-0.
- Khamna, S., Yokota, A., Peberdy, J.F. and Lumyong, S. (2010). *Indole-3-acetic acid production by Streptomyces sp. isolated from some Thai medicinal plant rhizosphere soils*. *EurAsia J Biosci* 4, 23–32.
- Khare, E., and Arora, N. K. (2010). *Effect of indole-3-acetic acid (IAA) produced by Pseudomonas aeruginosa in suppression of charcoal rot disease of chickpea*. *Curr Microbiol*, 61(1), 64-68.
- Kim, M. K., Choi, K. M., Yin, C. R., Lee, K. Y., Im, W. T., Lim, J. H., et al. (2004). *Odorous swine wastewater treatment by purple non-sulfur bacteria, Rhodopseudomonas palustris, isolated from eutrophicated ponds*. *Biotechnol Lett*, 26(10), 819-822.
- Kimura, M., and Kagawa, T. (2006). *Phototropin and light-signaling in phototropism*. *Curr Opin Plant Biol*, 9(5), 503-508.
- Koh, R. H., and Song, H. G. (2007). *Effects of application of Rhodopseudomonas sp. on seed germination and growth of tomato under axenic conditions*. *J Microbiol Biotechnol*, 17(11), 1805-1810.

- Kravchenko, L. V., Azarova, T. S., Makarova, N. M., and Tikhonovich, I. A. (2004). *The effect of tryptophan of plant root metabolites on the phyto stimulating activity of rhizobacteria*. *Mikrobiologiya*, 73(2), 195-198.
- Larimer, F. W., Chain, P., Hauser, L., Lamerdin, J., Malfatti, S., Do, L., et al. (2003). *Complete genome sequence of the metabolically versatile photosynthetic bacterium Rhodopseudomonas palustris*. *Nature biotechnology*, 22(1), 55-61.
- Larimer, F. W., Chain, P., Hauser, L., Lamerdin, J., Malfatti, S., Do, L., et al. (2004). *Complete genome sequence of the metabolically versatile photosynthetic bacterium Rhodopseudomonas palustris*. *Nat Biotechnol*, 22(1), 55-61.
- Leach, A. W., and Mumford, J. D. (2008). *Pesticide Environmental Accounting: A method for assessing the external costs of individual pesticide applications*. *Environmental Pollution*, 151(1), 139-147.
- Lee, K.-H., Koh, R.-H., and Song, H.-G. (2008). *Enhancement of growth and yield of tomato by Rhodopseudomonas sp. under greenhouse conditions*. *The Journal of Microbiology*, 46(6), 641-646.
- Lehmann, T., Hoffmann, M., Hentrich, M., and Pollmann, S. (2010). *Indole-3-acetamide-dependent auxin biosynthesis: a widely distributed way of indole-3-acetic acid production?* *Eur J Cell Biol*, 89(12), 895-905.
- Leljak-Levanic, D., Jezic, M., Cesar, V., Ludwig-Muller, J., Lepedus, H., Mladinic, M., et al. (2010). *Biochemical and epigenetic changes in phytoplasma-recovered periwinkle after indole-3-butyric acid treatment*. *J Appl Microbiol*, 109(6), 2069-2078.
- Leveau, J. H., and Lindow, S. E. (2005). *Utilization of the plant hormone indole-3-acetic acid for growth by Pseudomonas putida strain 1290*. *Appl Environ Microbiol*, 71(5), 2365-2371.

- Liu, S., Zhang, F., Chen, J., and Sun, G. (2011). *Arsenic removal from contaminated soil via biovolatilization by genetically engineered bacteria under laboratory conditions*. J Environ Sci (China), 23(9), 1544-1550.
- Marschner, H. *Effect of Internal and External Factors on Root Growth and Development*. In: H. Marschner (Ed.), *Mineral Nutrition of Higher Plants (Second Edition)*. London: Academic Press. 508-536; 1995
- Marschner, H., Kirkby, E. A., and Cakmak, I. (1996). *Effect of mineral nutritional status on shoot-root partitioning of photoassimilates and cycling of mineral nutrients*. J Exp Bot, 47, 1255-1263.
- Merzaeva, O. V., and Shirokikh, I. G. (2010). *Production of auxins by the endophytic bacteria of winter rye*. Prikl Biokhim Mikrobiol, 46(1), 51-57.
- Miller, S. A., Beed, F. D., and Harmon, C. L. (2009). *Plant disease diagnostic capabilities and networks*. Annu Rev Phytopathol, 47, 15-38.
- Miransari, M. (2011). *Soil microbes and plant fertilization*. Appl Microbiol Biotechnol, 92(5), 875-885.
- Mujahid, M., Sasikala, C., and Ramana Ch, V. (2010). *Aniline-induced tryptophan production and identification of indole derivatives from three purple bacteria*. Curr Microbiol, 61(4), 285-290.
- Muller, A., and Weiler, E. W. (2000). *Indolic constituents and indole-3-acetic acid biosynthesis in the wild-type and a tryptophan auxotroph mutant of Arabidopsis thaliana*. Planta, 211(6), 855-863.
- Nadeem, S., Naveed, M., Zahir, Z., and Asghar, H. (2013). *Plant-Microbe Interactions for Sustainable Agriculture: Fundamentals and Recent Advances*. In N. K. Arora (Ed.), *Plant Microbe Symbiosis: Fundamentals and Advances* (pp. 51-103): Springer India.
- Normanly, J. (2010). *Approaching cellular and molecular resolution of auxin biosynthesis and metabolism*. Cold Spring Harb Perspect Biol, 2(1), 15-94.

- Nunkaew, T., Kantachote, D., Nitoda, T., and Kanzaki, H. (2012). *The use of rice straw broth as an appropriate medium to isolate purple nonsulfur bacteria from paddy fields.*
- Oda, Y., Samanta, S. K., Rey, F. E., Wu, L., Liu, X., Yan, T., et al. (2005). *Functional genomic analysis of three nitrogenase isozymes in the photosynthetic bacterium Rhodospirillum rubrum.* J Bacteriol, 187(22), 7784-7794.
- Ona, O., Van Impe, J., Prinsen, E., and Vanderleyden, J. (2005). *Growth and indole-3-acetic acid biosynthesis of Azospirillum brasilense Sp245 is environmentally controlled.* FEMS Microbiology Letters, 246(1), 125-132.
- Perley, J. E., and Stowe, B. B. (1966). *The production of tryptamine from tryptophan by Bacillus cereus (KVT).* Biochem J, 100(1), 169-174.
- Podile, A., and Kishore, G. K. (2006). *Plant growth-promoting rhizobacteria.* In S. Gnanamanickam (Ed.), *Plant-Associated Bacteria* (pp. 195-230). Netherlands: Springer.
- Reineke, G., Heinze, B., Schirawski, J., Buettner, H., Kahmann, R. and Bassei, C.W. (2008). *Indole-3-acetic acid (IAA) biosynthesis in the smut fungus Ustilago maydis and its relevance for increased IAA levels in infected tissue and host tumor formation.* Molecular Plant Pathology, 9, 39–355.
- San-Francisco, S., Houdusse, F., Zamarreño, A. M., Garnica, M., Casanova, E., and García-Mina, J. M. (2005). *Effects of IAA and IAA precursors on the development, mineral nutrition, IAA content and free polyamine content of pepper plants cultivated in hydroponic conditions.* Scientia Horticulturae, 106(1), 38-52.
- Savci, S. (2012). *An agricultural pollutant: chemical fertilizer.* Int J. of Environ Sci and Dev, 3(1), 77-80.

- Seo, M., Akaba, S., Oritani, T., Delarue, M., Bellini, C., Caboche, M., et al. (1998). *Higher activity of an aldehyde oxidase in the auxin-overproducing superroot1 mutant of Arabidopsis thaliana*. Plant Physiol, 116(2), 687-693.
- Sheng, X., He, L., Wang, Q., Ye, H., and Jiang, C. (2008). *Effects of inoculation of biosurfactant-producing Bacillus sp. J119 on plant growth and cadmium uptake in a cadmium-amended soil*. J Hazard Mater, 155(1-2), 17-22.
- Shokri, D., and Emtiazi, G. (2010). *Indole-3-acetic acid (IAA) production in symbiotic and non-symbiotic nitrogen-fixing bacteria and its optimization by Taguchi design*. Curr Microbiol, 61(3), 217-225.
- Spaepen, S., Vanderleyden, J., and Remans, R. (2007). *Indole-3-acetic acid in microbial and microorganism-plant signaling*. FEMS Microbiol Rev, 31(4), 425-448.
- Sugawara, S., Hishiyama, S., Jikumaru, Y., Hanada, A., Nishimura, T., Koshiba, T., et al. (2009). *Biochemical analyses of indole-3-acetaldoxime-dependent auxin biosynthesis in Arabidopsis*. Proc Natl Acad Sci U S A, 106(13), 5430-5435.
- Thanawan Kantha, C. C., Duangporn Kantachote, Suchada Sukrong and Amorntip Muangprom. (2010). *Selection of photosynthetic bacteria producing 5-aminolevulinic acid from soil of organic saline paddy fields from the Northeast region of Thailand*. African Journal of Microbiology Research, 4(17), 1848-1855.
- Torres-Rubio, M.G., Valencia-Plata, S.A., Bernal-Castillo, J. and Martinez-Nieto, P. (2000). *Isolation of enterobacteria, Azotobacter sp. and Pseudomonas sp. producers of indole-3-acetic acid and siderophores, from Colombian rice rhizosphere*. Rev Latinoam Microbiol 42, 171-176.

- Van Puyvelde, S., Cloots, L., Engelen, K., Das, F., Marchal, K., Vanderleyden, J., et al. (2011). *Transcriptome analysis of the rhizosphere bacterium Azospirillum brasilense reveals an extensive auxin response*. *Microb Ecol*, 61(4), 723-728.
- Vande Broek, A., Gysegom, P., Ona, O., Hendrickx, N., Prinsen, E., Van Impe, J., et al. (2005). *Transcriptional analysis of the Azospirillum brasilense indole-3-pyruvate decarboxylase gene and identification of a cis-acting sequence involved in auxin responsive expression*. *Mol Plant Microbe Interact*, 18(4), 311-323.
- promotion potential of siderophore producing endophytic *Streptomyces* from *Azadirachta indica* A. Juss. *J Basic Microbiol*, 51(5), 550-556.
- Yaxley, J. R., Ross, J. J., Sherriff, L. J., and Reid, J. B. (2001). *Gibberellin biosynthesis mutations and root development in pea*. *Plant physiology*, 125(2), 627-633.
- Yurekli, F., Geckil, H., and Topcuoglu, F. (2003). *The synthesis of indole-3-acetic acid by the industrially important white-rot fungus Lentinus sajor-caju under different culture conditions*. *Mycol Res*, 107(3), 305-309.
- Zhao, Yⁱⁱⁱ. (2010). *Auxin biosynthesis and its role in plant development*. *Annu Rev Plant Biol*, 61, 49-6
- Verma, V. C., Singh, S. K., and Prakash, S. (2011). *Bio-control and plant growth*