# HEAVY METAL TOLERANCE OF *LYSINIBACILLUS FUSIFORMIS* ZB2 ISOLATED FROM TEXTILE EFFLUENT

WONG SIEW CHIAO

UNIVERSITI TEKNOLOGI MALAYSIA

## HEAVY METAL TOLERANCE OF *LYSINIBACILLUS FUSIFORMIS* ZB2 ISOLATED FROM TEXTILE EFFLUENT

WONG SIEW CHIAO

A dissertation submitted in partial fulfilment of the requirement for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

AUGUST 2015

A special dedication to my beloved family and friends who showered me abundantly with their love and continuous support

#### ACKNOWLEDGEMENTS

All praises to the Almighty Jesus Christ, the merciful for the strength and blessings throughout this study.

I express my sincere thanks from the bottom of my heart to my respected supervisor Assoc. Prof. Dr. Zaharah Binti Ibrahim for all her guidance, encouragement and active support throughout the course of this project. I also owe special debt of gratitude to Dr. Bay Hui Han for her patience, valuable suggestions and timely interventions at crucial junctures during the course of this project.

I am very thankful to all the lecturers, staffs and lab assistants in the Faculty of Biosciences and Medical Engineering for all their kindness and assistance in supplying materials required for this study. I also indebted to Universiti Teknologi Malaysia (UTM) for giving me the opportunity to pursue my Master study here.

I also wish to express my sincere appreciation to my fellow postgraduates, Asmaa, Ang Siow Kuang, Neoh Chin Hong, Lam Chi Yong, Mohd Fahmi and Muhd Hanif for their assistance throughout this study. I express my special thanks to my fellow coursemates especially Ong Leng Hui and Liew Kok Jun for their guidance, critics and friendship. I am very thankful to my beloved parents, family members and friends for their continuous prayers, motivations and love. This project has been enriched by the generosity and assistance of many people, without them, this dissertation would not be presented here.

### ABSTRACT

Heavy metal pollution has always been considered as one of the major threats to the environment and human health since these metals can accumulate in the food chain, inactivate cellular enzymes and may cause cancer related diseases. Conventional physiochemical methods do not provide economical treatment for the removal of heavy metals from heavy metal polluted environment. An effective and economical alternative method that has been widely reported is microbial bioremediation. In this study, the minimal inhibitory concentration (MIC) of Lysinibacillus fusiformis ZB2 for selected heavy metals, namely, cadmium (Cd), zinc (Zn), lead (Pb) and chromium (Cr(VI)) were determined. This bacteria was isolated previously from the textile effluent. It was grown in low phosphate medium (LPM) with glucose and tryptone as its carbon and nitrogen source respectively. L. fusiformis ZB2 reached its exponential growth within 48 hours of incubation in the LPM. The MIC of the bacteria for Cd, Zn, Pb and Cr(VI) were determined in solid and liquid media. The MIC obtained was relatively higher when using the liquid media. The MIC for Cd, Zn, Pb and Cr(VI) were 25, 75, 150, and 3500 ppm, respectively as compared to using the solid media MIC for Cd, Zn, Pb and Cr(VI) were 10, 75, 250, 3000 ppm, respectively. The order of toxicity of heavy metals towards Lysinibacillus fusiformis ZB2 was Cd>Zn>Pb>Cr(VI). The bacteria was found to be tolerant towards Zn, Pb and Cr(VI) with maximum tolerance towards Cr(VI).

### ABSTRAK

Pencemaran logam berat sentiasa dianggap sebagai satu daripada ancaman utama terhadap alam sekitar dan kesihatan manusia kerana logam ini boleh berkumpul dalam rantaian makanan, menyahakif enzim sel dan boleh dikaitkan dengan penyakit kanser. Kaedah penyingkiran logam berat dari persekitaran yang tercemar dengan logam berat menggunakan kaedah fizik kimia konvensional adalah tidak ekonomik. Satu kaedah alternatif yang berkesan dan ekonomik yang telah dilaporkan secara meluas adalah bioremediasi mikrob. Dalam kajian ini, minimal inhibitory concentration (MIC) daripada Lysinibacillus fusiformis ZB2 untuk logam berat terpilih, iaitu, kadmium (Cd), zink (Zn), plumbum (Pb) dan kromium (Cr(VI)) telah ditentukan. Bakteria ini telah diasingkan sebelum ini dari efluen tekstil. Ia dikultur dalam low phoshate medium (LPM) dengan glukosa dan tripton sebagai sumber karbon dan nitrogen masingmasing. L. fusiformis ZB2 mencapai pertumbuhan fasa eksponen dalam tempoh 48 jam pengeraman dalam LPM. MIC bakteria untuk Cd, Zn, Pb dan Cr(VI) ditentukan dalam media pepejal dan cecair. MIC yang diperolehi adalah lebih tinggi apabila menggunakan medium cecair. MIC untuk Cd, Zn, Pb dan Cr(VI) dalam medium cecair adalah 25, 75, 150, dan 3500 ppm masing-masing berbanding dengan medium pepejal MIC untuk Cd, Zn, Pb dan Cr(VI) adalah 10, 75, 250, 3000 ppm masing-masing. Urutan kesan ketoksikan logam berat terhadap Lysinibacillus fusiformis ZB2 adalah Cd> Zn> Pb> Cr(VI). Bakteria ini didapati toleran terhadap Zn, Pb dan Cr(VI) dan mempunyai toleransi maksimum terhadap Cr(VI).

### TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	Х
	LIST OF FIGURES	xi
	LIST OF ABBREVIATIONS	xiii
	LIST OF SYMBOLS	XV
	LIST OF APPENDICES	xvi
1	INTRODUCTION	1
	1.1 Background of Study	1
	1.2 Problem Statement	3
	1.3 Research Objectives	5
	1.4 Research Significance	5
	1.5 Scope of Research	6
2	LITERATURE REVIEW	7
	2.1 Heavy Metal Pollution	7
	2.2 Heavy Metal	8

2.3	Ph	ysico-ch	nemical Methods for Removal of Heavy	10
	Μ	etals		
	2.4	Efflue	nt Quality Discharge Limit Regulated by	12
		Legis	lation	
	2.5	Bioren	nediation of Heavy Metal	14
	2.6	Heavy	Metal Resistance Mechanism in Bacteria	16
	2.7	Lysini	bacillus	19
	2.8	Zinc		23
	2.9	Cadmi	um	25
	2.10	Lead		27
	2.11	Chron	nium	29
	MAT	FERIAL	S AND METHODS	31
	3.1	Materi	als	31
	3.2	Source	e of Microorganism	31
	3.3	Media	Preparation	32
		3.3.1	Nutrient Agar	32
		3.3.2	Nutrient Broth	32
		3.3.3	Low Phosphate Medium 1 (LPM1)	32
		3.3.4	Low Phosphate Medium 2 (LPM2)	33
		3.3.5	Low Phosphate Medium 3 (LPM3)	33
		3.3.6	Low Phosphate Medium 4 (LPM4)	34
		3.3.7	Low Phosphate Agar (LPA)	34
		3.3.8	Low Phosphate Agar (LPA) with	35
			Heavy Metal	
	3.4	Stock S	olution and Reagent Preparation	35
		3.4.1	Tryptone and Glucose Stock Solution	35
		3.4.2	Heavy Metal Stock Solutions	36
	3.5	Method	s	36
		3.5.1	Bacterial Culture and Inoculum	36
			Preparation	
		3.5.2	Carbon and Nitrogen Source in Low	37
			Phosphate Medium (LPM)	

viii

		3.5.3	Determination of MIC to Selected	38
			Heavy Metals with Solid Medium	
		3.5.4	Determination of MIC to Selected	38
			Heavy Metals in Liquid Medium	
	3.6	Statist	ical Analysis	39
	3.7	Flow	Chart of the Methodology	40
4	RES	ULTS A	AND DISCUSSION	41
	4.1	Growt	h of Lysinibacillus fusiformis ZB2 in Low	41
		Phospł	nate Medium (LPM) with different	
		combin	nations of C and N source	
	4.2	Deterr	nination of Lysinibacillus fusiformis ZB2	44
		Minim	al Inhibitory Concentration (MIC) of	
		Heavy	Metals with Low Phosphate Agar (LPA)	
	4.3	Deterr	nination of Lysinibacillus fusiformis ZB2	46
		Minim	al Inhibitory Concentration (MIC) of	
		Heavy	Metals with Low Phosphate Liquid	
		Mediu	m	
5	CON	NCLUSI	ONS	54
	5.1	Concl	usions	54
	5.2	Future	Work	55

REFERENCES	56
Appendices	70

### LIST OF TABLES

TITLE

TABLE NO.

2.1	Conventional metal removal technologies.	11
2.2	Acceptable conditions for discharge of industrial	13
	effluent or mixed effluent of Standards A and B.	
2.3	The MCL standards for most hazardous heavy metals.	14
2.4	Bacterial heavy metal resistance systems and	17
	mechanisms.	
2.5	Protein families important for heavy-metal transport.	18
2.6	Genes involved in heavy metals resistance that were	22
	identified in draft genome of Lysinibacillus fusiformis	
	ZB2.	
2.7	Zn <sup>2+</sup> bacterial transporter: names and mechanisms.	25
4.1	MIC of Lysinibacillus fusiformis ZB2 to different heavy	45
	metals and concentrations tested in LPA.	
4.2	MIC of Lysinibacillus fusiformis ZB2 in different heavy	51
	metals.	

PAGE

### LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	A modified periodic table showing commonly	9
	encountered regulated heavy metals, metalloids and	
	unregulated light metals.	
2.2	Graphical map of the genome of Lysinibacillus	21
	fusiformis ZB2.	
2.3	Protein families involved in bacterial heavy-metal	24
	metabolism.	
2.4	Protein families involved in bacterial heavy-metal	27
	metabolism.	
2.5	Selected mechanisms of cell protection against lead	28
	toxicity.	
2.6	Plausible mechanisms of enzymatic Cr <sup>6+</sup> reduction	30
	under aerobic and anaerobic conditions.	
3.1	The overall flow chart of the methodology.	40
4.1	Indirect growth measurement of Lysinibacillus	42
	fusiformis ZB2 in low phosphate media (LPM) with	
	different combinations of carbon and nitrogen source.	
4.2	Example of positive (a) and negative (b) growth of <i>L</i> .	44
	fusiformis ZB2 tested on different heavy metals with	
	different concentration on low phosphate agar (LPA).	
4.3	Percentage of Cd(II) tolerance of Lysinibacillus	47
	fusiformis ZB2 in low phosphate medium (LPM)	
	incubated at 37 $^{\circ}$ C for 48 hours.	

4.4	Percentage of Zn(II) tolerance of Lysinibacillus	48
	fusiformis ZB2 in low phosphate medium (LPM)	
	incubated at 37 °C for 48 hours.	
4.5	Percentage of Pb(II) tolerance of Lysinibacillus	49
	fusiformis ZB2 in low phosphate medium (LPM)	
	incubated at 37 °C for 48 hours.	
4.6	Percentage of Cr(VI) tolerance of Lysinibacillus	50
	fusiformis ZB2 in low phosphate medium (LPM)	
	incubated at 37 $^{\circ}$ C for 48 hours.	

### LIST OF ABBREVIATIONS

ADMI	-	American Dye Manufacturers Institute
CaCl <sub>2</sub>	-	Calcium chloride
Cd(II)	-	Cadmium (2 <sup>+</sup> ) ion
$Cd(NO_3)_2 \cdot 4H_2O$	-	Cadmium nitrate tetrahydrate
COD	-	Chemical Oxygen Demand
Cr(VI)	-	Chromium $(6^+)$ ion
HCl	-	Hydrochloric acid
$K_2Cr_2O_7$	-	Potassium dichromate
KCl	-	Potassium chloride
LPA	-	Low phosphate agar
LPM	-	Low phosphate medium
MIC	-	Minimal inhibitory concentration
Na <sub>2</sub> SO <sub>4</sub>	-	Sodium sulphate
NaCl	-	Sodium chloride
NADH	-	Nicotiamide adenine dinucleotide (reduced)
NADPH	-	Nicotiamide adenine dinucleotide phosphate (reduced)
NH4Cl	-	Ammonium chloride

OD <sub>600nm</sub>	-	Optical density at 600nm
Pb(II)	-	Lead $(2^+)$ ion
Pb(NO <sub>3</sub> ) <sub>2</sub>	-	Lead (II) nitrate
rpm	-	Revolutions per minute
Tris	-	2-Amino-2-(hydroxymethyl)-1,3-propanediol
Zn(II)	-	Zinc(2 <sup>+</sup> ) ion
ZnSO <sub>4</sub>	-	Zinc sulfate

### LIST OF SYMBOLS

%	-	percent
C	-	degree Celsius
μm	-	micrometre
g	-	gram
g/L	-	gram per litre
h	-	hour
kPa	-	kilopascal
L	-	Litre
mg/L	-	milligram per litre
mL	-	millilitre
mM	-	millimolar
nm	-	nanometer
ppm	-	parts per million
v/v	-	volume per volume
w/v	-	weight per volume

### LIST OF APPENDICES

APPENDIX	TITLE	PAGE
А	Preparation of working concentration from stock	67
	solution of heavy metal	

### **CHAPTER 1**

### INTRODUCTION

### 1.1 Background of Study

Heavy metals are elements with atomic weight ranging from 63.5 to 200.6 and have specific gravity more than 5.0 (Srivastava and Majumder, 2008). Toxic metals such as mercury (Hg), chromium (Cr), lead (Pb), zinc (Zn), copper (Cu), nickel (Ni), cadmium (Cd), arsenic (As), cobalt (Co) and tin (Sn); precious metals, such as palladium (Pd), platinum (Pt), silver (Ag), gold (Au) and ruthenium (Ru); and radionuclides such as uranium (U), radium (Ra) and americium (Am). These three types of heavy metals are rising concerns because of their negative impact towards the environment and human health (Wang and Chen, 2006).

Heavy metal pollution is known as a critical environmental problem as a result of the metals toxic effect and their accumulation throughout the food chain can cause severe problems to the ecology and human health (Malik, 2004). Wastewater containing heavy metals are discharged directly and indirectly into the environment particularly in developing countries (Fu and Wang, 2011). Coal, natural gas, paper and chlor-alkali, metal plating, mining, fertilizer, tanneries, batteries and pesticides industries are known to be the source for heavy metal pollution (Matlock *et al.*, 2002; Fu and Wang, 2011). Growing interests among researchers in bioremediation of heavy metals by microorganisms in recent years are possibly due to its potential application in industry and scientific novelty of the microorganism (Singh *et al.*, 2013). Metal ions can be readily adsorbed and accumulated by bacteria, algae and fungi (Abbott *et al.*, 2005; Volesky and Holan, 1995). In the study for the heavy metal tolerance, the medium composition affects interactions between metal ions and microbes in terms of bioavailability due to accumulation and precipitation of metal ions (Kumar *et al.*, 2013). As a result of low carbon source and negligible phosphate, low phosphate medium (LPM) provide more reliable results as compared to complex medium also provide conditions that is more similar to those found in the environmental sample compared to that of rich medium (Karelova *et al.*, 2011).

Heavy metals are often used in textile processes (Rybicki *et al.*, 2004) and eventually found in the textile wastewater in the form of free ionic metals or complex metals (Hill *et al.*, 1993). Heavy metals such as lead (Pb), cadmium (Cd), chromium (Cr), zinc (Zn), copper (Cu) and iron (Fe) are present in the textile dye of the textile effluent (Halimoon and Goh, 2010; Siddiqui *et al.*, 2011). In previous study by Kee *et al.* (2015), *Lysinibacillus fusiformis* strain ZB2 together with *Bacillus pumilus* strain ZK1, *Bacillus cereus* strain ZK2, *Brevibacillus panacihumi* strain ZB1 was used to treat real textile wastewater successfully for decolourization purpose and *L. fusiformis* ZB2 was identified as the dominant species in the mature granules.

Bacterial strains are potential candidates for simultaneous removal of metals from wastes as they have high tolerance to different metals (Malik, 2004). Since heavy metal pollution is becoming one of the major threats to the environment and possess many health hazards, bioremediation potential of bacterial isolates should be assessed by preliminary study in terms of their resistance level and the minimal inhibitory concentrations (MICs). This is fundamental in order to check for the tolerance of the bacterial strains towards different heavy metals to develop suitable heavy metal waste remediation.

#### **1.2 Problem Statement**

Heavy metal tolerance test of bacteria in complex medium results in higher tolerance behavior by the bacteria. This is because the metals chelate with constituents of the complex medium resulting in metal precipitation and non-uniform availability of metals in the medium. Low phosphate medium (LPM) is preferred over complex medium as the metal's precipitation is reduced due to negligible phosphate and low carbon source, thus, more metal ions are available to the bacteria (Kumar *et al.*, 2013). However, the suitability of LPM as *Lysinibacillus fusiformis* ZB2 growth medium is still yet to be tested.

In the last few decades, the river water and sediments are receiving increasing concentration of heavy metals. Although industrialization has long been accepted as a hallmark of civilization, it is undeniable that industrial discharges have been causing negative impacts to the environment. Mining, milling, surface finishing industries are the main sources of heavy metal pollution that discharge a variety of toxic metals such as Cd, Co, Cu, Ni, Pb and Zn into the environment. The toxics and heavy metals in the industrial effluents are often discharged into the river which might be a source of drinking water for another town downstream (Moore, 1990; Ewan and Pamphlett, 1996). According to Barakat (2011), heavy metals are hazardous to human health as they may inhibit growth and development, cause organ damage and cancer, damage to nervous system and in extreme cases, death.

Furthermore, there is an increasing demand to shift to cleaner production methods in different industries and develop environmental friendly, economical and efficient treatment technique for metal contaminated effluent as many industries have adhered to more stringent environmental regulations (Malik, 2004; Fu and Wang, 2011). A variety of methods are employed for the removal of heavy metal ions such as chemical precipitation, ion-exchange, adsorption, membrane filtration, electrochemical treatment technologies, etc. (Fu and Wang, 2011). However, these methods have their disadvantages as they are expensive, not environmental friendly and involve complex processes (Karman *et al.*, 2015). Also, remediation of heavy metal contaminated sites are unfeasible as heavy metals may disperse both horizontally and vertically as they migrate (Nikolaidis *et al.*, 1999).

Since heavy metals are the environmental priority pollutants and cause one of the most serious environmental problems, removal of these toxic heavy metals is essential in order to protect the people and environment (Fu and Wang, 2011). In the past few decades, microbial mediated detoxification technologies are still being valued over physicochemical ones. Biological remediation is receiving more attention as they are more economic and have long lasting nature (Ali *et al.*, 2009). Interactions of microbes and metals have significant environmental implications and monitoring as the microbes have adapted to resist the presence of metals or utilizes the metals for their growth. One of the useful environmental implications is the use of bacteria to clean up metal polluted areas (Nithya *et al.*, 2011).

Thus, in this study, the low phosphate medium (LPM) with different combinations of carbon and nitrogen source was used to grow *Lysinibacillus fusiformis* ZB2 in order to test the suitability of this medium as the growth medium for the bacteria and subsequent heavy metal testing. Furthermore, this bacteria was used to screen for its heavy metal tolerance and determine its minimal inhibitory concentration (MIC) to selected heavy metals which are Cd, Zn, Pb and Cr(VI) since these heavy metals are present mostly in the industrial effluents such as textile effluents.

#### **1.3 Research Objectives**

There are three objectives for this study:

- i. To determine suitable carbon and nitrogen co-substrate for growth of *Lysinibacillus fusiformis* ZB2 in low phosphate medium (LPM)
- ii. To screen resistance of *L. fusiformis* ZB2 to selected heavy metals
- iii. To determine minimal inhibitory concentration (MIC) of *L. fusiformis*ZB2 to selected heavy metals

### 1.4 Research Significance

Heavy metals are used in many industries and this causes a lot of concerns as they are not biodegradable and can remain the environment for a long period of time. It causes harm to both environment and human health. Conventional physicochemical methods are employed for the removal of the metals from heavy metal polluted sites, however, it has several disadvantages making remediation of heavy metal pollution a challenging task to achieve. With the emergence of biotechnology, microorganisms such as bacteria, algae and fungi are studied and are found to be capable of tolerating with heavy metals. In order to explore more potential microbe candidates for the remediation of heavy metal and overcome the limitation of physicochemical methods, more bacteria can be studied for their heavy metal tolerance. Therefore, in this study *Lysinibacillus fusiformis* ZB2 isolated previously from textile effluent was used to test for its heavy metal tolerance.

#### **1.5** Scope of Research

This study was mainly focused on heavy metal tolerance or resistance of Lysinibacillus fusiformis ZB2 in four selected heavy metals, namely, cadmium (Cd), zinc (Zn), lead (Pb), and chromium (Cr(VI)). Cd, Zn, Pb, and Cr(VI) are usually present in many industrial effluent and cause hazardous effect to the environment and Therefore, these heavy metals were chosen for this study. L. living organisms. fusiformis ZB2 was previously isolated from the textile effluent. The bacteria was grown in low phosphate medium (LPM) with optimisation of carbon and nitrogen source and the exponential growth phase in the best LPM was determined. The heavy metal resistance of L. fusiformis ZB2 was tested by spot inoculation on low phosphate agar (LPA) and by growing the bacteria in LPM supplemented with different concentrations of heavy metals. In general, the heavy metal resistance was determined in terms of minimal inhibitory concentration (MIC), which is the concentration of heavy metal that inhibit the bacteria growth in LPA and LPM. The MIC and percentage of tolerance were investigated by evaluating growth in LPA and measurement of bacteria concentration at OD<sub>600</sub> nm when the bacteria was grown in LPM incorporated with different concentrations of heavy metals.

#### REFERENCES

- Abbott, A. P., Capper, G., Davies, D. L., Rasheed, R. K., and Shikotra, P. (2005). Selective extraction of metals from mixed oxide matrixes using choline-based ionic liquids. *Inorganic Chemistry*. 44: 6497-6499.
- Ackerley, D. F., Gonzalez, C. F., Park, C. H., Blake, R., Keyhan, M., and Matin, A. (2004). Chromate-reducing properties of soluble flavoproteins from *Pseudomonas putida* and *Escherichia coli*. Applied and Environmental Microbiology. 20: 873-882.
- Ahluwalia, S. S., and Goyal, D. (2007). Microbial and plant derived biomass for removal of heavy metals from wastewater. *Bioresource Technology*. 98: 2243-2257.
- Alex, R. (2012). *Bioremediation of hydrocarbons by Lysinibacillus fusiformis BTTS10*.PhD Thesis. Cochin University of Science and Technology, Cochin, India.
- Alex, R., and Bhat, S. G. (2012). Isolation and characterization of a solvent tolerant alkaliphilic marine bacteria. *International Journal of Scientific and Engineering Research.* 3: 2229-5518.
- Ali, N., Hameed, A., and Ahmed, S. (2009). Physicochemical characterization and bioremediation perspective of textile effluent, dyes and metals by indigenous bacteria. *Journal of Hazardous Materials*. 164: 322–328.
- Apell, H. J. (2004). How do P-type ATPases transport ions? *Bioelectrochemistry*. 63: 149–156.
- Arguello, J. M. (2003). Identification of ion-selectivity determinants in heavy-metal transport P1B-type ATPases. *Journal of Membrane Biology*. 195: 93–108.

- Azlina, W. A., Shafinaz, S., and Zakaria, Z. A. (2009). Mechanisms of bacterial detoxification of Cr (VI) from industrial wastewater in the presence of industrial effluent as potential energy source (Final Report). Universiti Teknologi Malaysia, Skudai, Malaysia.
- Babel, S., and Kurniawan, T. A. (2003). Low-cost adsorbents for heavy metals uptake from contaminated water: a review. *Journal of Hazardous Materials*. 97: 219– 243.
- Barakat, M. A. (2011). New trends in removing heavy metals from industrial wastewater. *Arabian Journal of Chemistry*. 4: 361–377.
- Baruthio, F. (1992). Toxic effects of chromium and its compounds. *Biological Trace Element Research.* 32: 145-153.
- Bay, H. H., Lim, C. K., Kee, T. C., Ware, I., Chan, G. F., Shahir, S., and Ibrahim, Z. (2014). Decolourisation of Acid Orange 7 recalcitrant auto-oxidation coloured by-products using an acclimatised mixed bacterial culture. *Environmental Science and Pollution Research*. 21: 3891-3906.
- Beard, S. J., Hashim, R., Membrillo-Hernandez, J., Hughes, M. N., and Poole, R. K. (1997). Zinc(II) tolerance in *Escherichia coli* K-12: evidence that the *zntA* gene (o732) encodes a cation transport ATPase. *Molecular Microbiology*. 25: 883-891.
- Bhimani, H. D. (2011). Bacterial degradation of azo dyes and its derivatives. DoctorPhilosophy. Saurashtra University, Gujarat, India.
- Blencowe, D. K., and Morby, A. P. (2003). Zn(II) metabolism in prokaryotes. FEMS Microbiology Reviews. 27: 291–311.
- Blindauer, C. A., Harrison, M. D., Robinson, A. K., Parkinson, J. A., Bowness, P. W., Sadler, P. J., and Robinson, N. J. (2002). Multiple bacteria encode metallothioneins and SmtA-like zinc fingers. *Molecular Microbiology*. 45: 1421–1432.
- Bruins, M. R., Kapil, S., and Oehme, F. W. (2000). Microbial resistance to metals in the environment. *Ecotoxicology and Environmental Safety*. 45: 198–207.
- Cavet, J. S., Borrelly, G. P., and Robinson, N. J. (2003). Zn, Cu and Co in cyanobacteria:

selective control of metal availability. *FEMS Microbiology Reviews*. 27: 165–181.

- Chen, C., and Wang, J. (2007). Response of *Saccharomyces cerevisiae* to lead ion stress. *Applied Microbiology and Biotechnology*. 74: 683–687.
- Chen, X. C., Wang, Y. P., Lin, Q., Shi, J. Y., Wu, W. X. and Chen, Y. X. (2005). Biosorption of copper (II) and zinc(II) from aqueous solution by *Pseudomonas putida* CZ1. *Colloids and Surfaces B: Biointerfaces*. 46: 101–107.
- Cheung, K. H., and Gu, J. D. (2007). Mechanism of hexavalent chromium detoxification by microorganisms and bioremediation application potential: A review. *International Biodeterioration and Biodegradation*. 59: 8–15.
- Coleman, J. E. (1998). Zinc enzymes. *Current Opinion in Chemical Biology*. 2: 222-234.
- Davis, T. A., Volesky, B., and Muccib, A. (2003). A review of the biochemistry of heavy metal biosorption by brown algae. *Water Research*. 37: 4311–4330.
- De Rore, H., Top, E., Houwen, F., Mergeay, M., and Verstraete, W. (1994). Evolution of heavy metal resistant transconjugants in a soil environment with a concomitant selective pressure. *FEMS Microbiology Ecology*. 14: 263-273.
- Dintilhac, A., Alloing, G., Granadel, C., and Claverys, J. P. (1997). Competence and virulence of *Streptococcus pneumoniae*: Adc and PsaA mutants exhibit a requirement for Zn and Mn resulting from inactivation of putative ABC metal permeases. *Molecular Microbiology*. 25: 727-739.
- Dongfeng, Z., Weilin, W., Yunbo, Z., Qiyou, L., Haibin, Y., and Chaocheng, Z. (2011). Study on isolation, identification of a petroleum hydrocarbon degrading bacterium *Bacillus fusiformis sp.* and influence of environmental factors on degradation efficiency. *China Petroleum Processing and Petrochemical Technology.* 13: 74-82.
- Dopson, M., Baker-Austin, C., Koppineedi, P. R., and Bond, P. L. (2003). Growth in sulfidic mineral environments: metal resistance mechanisms in acidophilic micro-organisms. *Microbiology*. 149: 1959–1970.

- Durve, A., Naphade, S., Bhot, M., Varghese, J., and Chandra, N. (2012). Characterisation of metal and xenobiotic resistance in bacteria isolated from textile effluent. *Advances in Applied Science Research*. 3: 2801-2806.
- Elangovan, R., Abhipsa, S., Rohit, B., Philip, L., and Chandraraj, K. (2006). Reduction of Cr(VI) by a *Bacillus* sp. *Biotechnology Letters*. 28:247–253.

Environmental Quality (Industrial Effluent) Regulations. (2009).

- Ewan, K. B., and Pamphlett, R. (1996). Increased inorganic mercury in spinal motor neurons following chelating agents. *Neurotoxicology*. 17: 343–349.
- Fagan, M. J., and Saier, M. H. J. (1994). P-type ATPase of eukaryotes and bacteria: sequence comparisons and construction of phylogenetic trees. *Journal of Molecular Evolution*. 38: 57-99.
- Fath, M. J., and Kolter, R. (1993). ABC-transporters: the bacterial exporters. *Microbiology Reviews*. 57: 995-1017.
- Ferchichi, M., Crabbe, E., Hintz, W., Gil, G. H., and Almadidy, A. (2005). Influence of culture parameters on biological hydrogen production by *Clostridium saccharoperbutylacetonicum* ATCC 27021. World Journal of Microbiology and *Biotechnology*. 21: 855-862.
- Fu, F., and Wang, Q. (2011). Removal of heavy metal ions from wastewaters: A review. Journal of Environmental Management. 92: 407-418.
- Gadd, G. M. (1988). Accumulation of metals by microorganisms and algae, In Rehm,
  H. J. (Ed.) *Biotechnology-special microbial processes, Vol. 6B* (pp. 401-433).
  Weinheim: VCH Verlagsgellschaft.
- Garbisu, C., and Alkorta, I. (1997). Bioremediation: principles and future. Journal of Clean Technology, Environmental Toxicology and Occupational Medicine. 6: 351–366.
- Garbisu, C., and Alkorta, I. (2003). Basic concepts on heavy metal soil bioremediation. *The European Journal of Mineral Processing and Environmental Protection*. 3: 58–66.
- Goyer, R. A., and Clarkson, T. W. (2001). Chapter 23 Toxic effects of metals (pp. 533-

573). New York: McGraw-Hill.

- Grass, G., Franke, S., Taudte, N., Nies, D. H., Kucharski, L. M., Maguire, M. E., and Rensing, C. (2005). The metal permease ZupT from *Escherichia coli* is a transporter with a broad substrate spectrum. *Journal of Bacteriology*. 187: 1604– 1611.
- Halder, S. (2014). Bioremediation of heavy metals through fresh water microalgae: a review. *Scholars Academic Journal of Biosciences*. 2: 825-830.
- Halimoon, N. and Goh, R. S. Y. (2010). Removal of heavy metals from textile wastewater using zeolite. *EnironmentAsia*. 3: 124-130.
- Hasnain, S., Yasmin, S., and Yasmin, A. (1993). The effects of lead resistant pseudomonads on the growth of *Triticum aestivum* seedlings under lead stress. *Environmental Pollution*. 81: 179–184.
- Hassan, M. E. T., van der Lelie, D., Springael, D., Romling, U., Ahmed, N., and Mergeay, M. (1999). Identification of a gene cluster, *czr*, involved in cadmium and zinc resistance in *Pseudomonas aeruginosa*. *Gene*. 238: 417–425.
- He, M., Li, X., Liu, H., Miller, S. J., Wang, G., and Rensing, C. (2011). Characterization and genomic analysis of a highly chromate resistant and reducing bacterial strain *Lysinibacillus fusiformis* ZC1. *Journal of Hazardous Materials*. 185: 682-688.
- Hill, W. E., Perkins, S., and Sandlin, G. S. (1993). Removal and speciation of transition metal ions textile dyeing wastewaters. *Textile Chemist and Colorist.* 25: 26–27.
- Hoostal, M. J., Bidart-Bouzat, M. G., and Bouzat, J. L. (2008). Local adaptation of microbial communities to heavy metal stress in polluted sediments of Lake Erie. *FEMS Microbiology Ecology*. 65: 156-168.
- Jain, S., and Bhatt, A. (2014). Molecular and in situ characterization of cadmiumresistant diversified extremophilic strains of *Pseudomonas* for their bioremediation potential. *Biotechnology*. 4: 297–304.
- Jansen, E., Michels, M., Til, M., and Doelman, P. (1994). Effects of heavy metals in soil on microbial diversity and activity as shown by the sensitivity-resistance

index, an ecologically relevant parameter. *Biology and Fertility of Soils*. 17: 177-184.

- Jarosławiecka, A., and Piotrowska-Seget, Z. (2014). Lead resistance in microorganism. *Microbiology*. 160: 12–25.
- Kalil, M. S., Alshiyab, H. S., and Yusoff, W. M. W. (2008). Effect of nitrogen source and carbon to nitrogen ratio on hydrogen production using *C. acetobutylicum*. *American Journal of Biochemistry and Biotechnology*. 4: 393-401.
- Karelova, E., Harichova, J., Stojnev, T., Pangallo, D., and Ferianc, P. (2011). The isolation of heavy-metal resistant culturable bacteria and resistace determinants from a heavy-metal-contaminated site. *Biologia*. 66: 18-26.
- Karman, S. B., Diah, S. Z. M., and Gebeshuber, I. C. (2015). Raw materials synthesis from heavy metal industry effluents with bioremediation and phytomining: A biomimetic resource management approach. *Advances in Materials Science and Engineering*. 2015: 1-21.
- Kee, T. C., Bay, H. H., Lim, C. K., Muda, K., and Ibrahim, Z. (2015). Development of bio-granules using selected mixed culture of decolorizing bacteria for the treatment of textile wastewater. *Desalination and Water Treatment*. 54(1):132-139.
- Khezami, L., and Capart, R. (2005). Removal of chromium(VI) from aqueous solution by activated carbons: kinetic and equilibrium studies. *Journal of Hazardous Material*. 123: 223-231.
- Khudairy, A. A., Alrokayan, S. A. H., and Al-Minsed, F. A. (2001). Cadmium toxicity and cell stress response. *Pakistan Journal of Biological Sciences*. 4: 1046–1049.
- Kim, B. N., Joo, Y. C., Kim, Y. S., Kim, K. R., and Oh, D. K. (2012). Production of 10-hydroxystearic acid from oleic acid and olive oil hydrolyzate by an oleate hydratase from *Lysinibacillus fusiformis*. *Applied Microbiology and Biotechnology*. 95: 929-37.
- Kolenbrander, P. E., Andersen, R. N., Baker, R. A., and Jenkinson, H. F. (1998). The adhesion-associated sca operon in *Streptococcus gordonii* encodes an inducible

high-affinity ABC transporter for Mn<sup>2+</sup> uptake. *Journal of Bacteriology*. 180: 290-295.

- Krznaric, E., Verbruggen, N., Wevers, J. H. L., Carleer, R., Vangronsveld, J., and Colpaert, J. V. (2009). Cd-tolerant *Suillus luteus*: a fungal insurance for pines exposed to Cd. *Environmental Pollution*. 157: 1581-1588.
- Kumar, R., Nongkhlaw, M., Acharya, C., and Joshi, S. R. (2013). Growth media composition and heavy metal tolerance behaviour of bacteria characterized from the sub-surface soil of uranium rich ore bearing site of Domiasiat in Meghalaya. *Indian Journal of Biotechnology*. 12: 115-119.
- Lee, S. W., Glickmann, E., and Cooksey, D. A. (2001). Chromosomal locus for cadmium resistance in *Pseudomonas Putida* consisting of a cadmiumtransporting ATPase and a merr family response regulator. *Applied and Environmental Microbiology*. 67: 1437–1444.
- Liang, B., Lu, P., Li, H., Li, R., Li, S., and Huang, X. (2009). Biodegradation of fomesafen by strain *Lysinibacillus* sp. ZB-1 isolated from soil. *Chemosphere*. 77: 1614–1619.
- Liu, H., Song, Y., Chen, F., Zheng, S., and Wang, G. (2013). Lysinibacillus manganicus sp. nov., isolated from manganese mining soil. International Journal of Systematic and Evolutionary Microbiology. 63: 3568-3573.
- Lopez-Maury, L., Garcia-Dominguez, M., Florencio, F. J., and Reyes, J. C. (2002). A two component signal transduction system involved in nickel sensing in the *Cyanobacterium synechocystis* sp. PCC 6803. *Molecular Microbiology*. 43: 247–56.
- Lu, D., Boyd, B., and Lingwood, C. A. (1997). Identification of the key protein for zinc uptake in *Haemophilus influenzae*. *Journal of Biological Chemistry*. 272: 29033-29038.
- Lu, W. B., Shi, J. J., Wang, C. H., and Chang, J. S. (2006). Biosorption of lead, copper and cadmium by an indigenous isolate *Enterobacter* sp. J1 possessing high heavy metal resistance. *Journal of Hazardous Material*. 134: 80-86.

- Lucious, S., Reddy, E. S., Anuradha, V., Vijaya, P. P., Ali, A. S., Yogananth, N., Rajan, R., and Parveen, P. K. (2013). Heavy metal tolerance and antibiotic sensitivity of bacterial strains isolated from tannery effluent. *Asian Journal of Experimental Biological Sciences*. 4: 597-606.
- Malik, A. (2004). Metal bioremediation through growing cells. *Environment International*. 30: 261–278.
- Mani, D., and Kumar, C. (2014). Biotechnological advances in bioremediation of heavy metals contaminated ecosystems: an overview with special reference to phytoremediation. *International Journal of Environmental Science and Technology*. 11: 843–872.
- Martinez, R. J., Wang, Y., Raimondo, M. A., Coombs, J. M., Barkay, T., and Sobecky, P.A. (2006). Horizontal gene transfer of PIB-type ATPases among bacteria isolated from radionuclide- and metal-contaminated subsurface soils. *Applied Environmental Microbiology*. 72: 3111-3118.
- Matlock, M. M., Henke, K. R., and Atwood, D. A. (2002). Effectiveness of commercial reagents for heavy metal removal from water with new insights for future chelate designs. *Journal of Hazardous Materials*. 92: 129-142.
- Mclean, J., and Beveridge, T. J. (2000). Chromate reduction by a pseudomonad isolated from a site contaminated with chromated copper arsenate. *Applied and Environmental Microbiology*. 67: 1076–1084.
- Melnick, R. L., Suárez, C., Bailey, B. A., and Backman, P.A. (2011). Isolation of endophytic endospore-forming bacteria from *Theobroma cacao* as potential biological control agents of cacao diseases. *Biological Control.* 57: 236-245.
- Mergeay, M., Nies, D., Schlegel, H. G., Gerits, J., Charles, P., and van Gijsegem, F. (1985). Alcaligenes eutrophus CH34 is a facultative chemolithotroph with plasmid-bound resistance to heavy metals. Journal of Bacteriology. 162: 328-334.
- Miwa, H., Ahmed, I., Yokota, A., and Fujiwara, T. (2009). *Lysinibacillus* parviboronicapiens sp. nov., a low-boron-containing bacterium isolated from

soil. International Journal of Systematic and Evolutionary Microbiology. 59: 1427-1432.

- Mohan, N., and Naveena, L. (2015). Isolation and determination of efficacy of acephate degrading bacteria from agricultural soil. *Journal of Environmental Science, Toxicology and Food Technology*. 9: 2319-2402.
- Moore, J. W. (1990). Inorganic contaminants of surface water residuals and monitoring priorities. (pp. 178-210). New York: Springer-Verlag.
- Naseem, R., and Tahir, S.S. (2001). Removal of Pb(II) from aqueous solution by using bentonite as an adsorbent. *Water Research*. 35: 3982-3986.
- Nies, D. H. (1995). The cobalt, zinc, and cadmium efflux system CzcBAc from *Alcaligenes eutrophus* functions as a cation-proton-antiporter in *Escherichia coli*. *Journal of Bacteriology*. 177: 2707-2712.
- Nies, D. H. (1999). Microbial heavy-metal resistance. Applied Microbiology and Biotechnology. 51: 730-750.
- Nies, D. H. (2003). Efflux-mediated heavy metal resistance in prokaryotes. FEMS Microbiology Reviews. 27: 313–339.
- Nies, D. H. and Silver S. (1995). Ion efflux systems involved in bacterial metal resistances. *Journal of Industrial. Microbiology*. 14: 186–199.
- Nies, D. H., and Silver, S. (1989). Plasmid-determined inducible efflux is responsible for resistance to cadmium, zinc, and cobalt in *Alcaligenes eutrophus*. *Journal of Bacteriology*. 171: 896-900.
- Nies, D. H., and Silver, S. (1995). Ion efflux systems involved in bacterial metal resistances. *Journal of Industrial Microbiology*. 14: 186-199.
- Nies, D. H., Nies, A., Chu, L., and Silver, S. (1989). Expression and nucleotide sequence of a plasmid-determined divalent cation efflux system from *Alcaligenes eutrophus*. *Proceedings of the National Academy of Sciences of the United States of America*. 86: 7351–7355.
- Nies, D., Mergeay, M., Friedrich, B., and Schlegel, H. G. (1987). Cloning of plasmid genes encoding resistance to cadmium, zinc and cobalt in *Alcaligenes eutrophus*

CH34. Journal of Bacteriology. 169: 4865-4868.

- Nikaido, H. (1996). Multiple efflux pumps of gram-negative bacteria. *Journal of Bacteriology*. 178: 5853-5859.
- Nikolaidis, N. P., Hellerich, L. A., and Lackovic, J. A. (1999). Methodology for sitespecific, mobility based cleanup standards for heavy metals in glaciated soils. *Environmental Science and Technology*. 33: 2910–2916.
- Nithya, C., Gnanalakshmi, B., and Pandian, S. K. (2011). Assessment and characterization of heavy metal resistance in Palk Bay sediment bacteria. *Marine Environmental Research*. 71: 283-294.
- Oyaro, N., Juddy, O., Murago, E. N. M., and Gitonga, E. (2007). The contents of Pb, Cu, Zn and Cd in meat in Nairobi, Kenya. *Food, Agriculture and Environment* (*JFAE*). 5: 119-121.
- Parkouda, C. (2010). Microorganisms associated with maari, a baobab seed fermented product. *International Journal of Food Microbiology*. 142: 292-301.
- Patzer, S. I., and Hantke, K. (1998). The ZnuABC high-affinity zinc uptake system and its regulator Zur in *Escherichia coli*. *Molecular Microbiology*. 28: 1199-1210.
- Paulsen, I. T., and Saier, M. H. Jr. (1997). A novel family of ubiquitous heavy metal ion transport proteins. *Journal of Membrane Biology*. 156: 99-103.
- Paulsen, I. T., Brown, M. H., and Skurray, R. D. (1996). Proton-dependent multidrug efflux systems. *Microbiology Reviews*. 60: 575-608.
- Paulsen, I. T., Sliwinski, M. K., Nelissen, B., Goffeau, A., Saier, M. H. (1998). Unified inventory of established and putative transporters encoded within the complete genome of *Saccharomyces cerevisiae*. *FEBS Letters*. 430: 116-125.
- Poopal, A. C., and Laxman, R. S. (2008). Hexavalent chromate reduction by immobilized *Streptomyces griseus*. *Biotechnology Letters*. 30: 1005–1010.
- Prabha, M. S., Divakar, K., Priya, J. D. A., Selvam, G. P., Balasubramanian, N., and Gautam, P. (2015). Statistical analysis of production of protease and esterase by a newly isolated *Lysinibacillus fusiformis* AU01: purification and application of

protease in sub-culturing cell lines. Annals of Microbiology. 65: 33-46.

- Raja, C. E., and Omine, K. (2012): Characterization of boron resistant and accumulating bacteria Lysinibacillus fusiformis M1, Bacillus cereus M2, Bacillus cereus M3, Bacillus pumilus M4 isolated from former mining site, Hokkaido, Japan. Journal of Environmental Science and Health. 47: 1341-1349.
- Rathnayake, I. V. N., Megharaj, M., Krishnamurti, G. S. R., Bolan, N. S., and Naidu, R. (2013). Heavy metal toxicity to bacteria – Are the existing growth media accurate enough to determine heavy metal toxicity? *Chemosphere*. 90: 1195– 1200.
- Rensing, C., Mitra, B., and Rosen, B. P. (1997). The zntA gene of Escherichia coli encodes a Zn(II)-translocating P-type ATPase. Proceedings of the National Academy of Sciences of the United States of America. 24: 14326-14331.
- Roane, T. M., and Kellogg, S. T. (1996). Characterization of bacterial communities in heavy metal contaminated soils. *Canadian Journal of Microbiology*. 42: 593– 603.
- Robinson, N. J., Gupta, A., Fordham-Skelton, A. P., Croy, R. R. D., Whitton, B. A., and Huckle, J. W. (1990). Prokaryotic metallothionein gene characterization and expression: Chromosome crawling by ligation-mediated PCR. *Proceedings of the Royal Society B*. 242: 241–247.
- Rubio, J., Souza, M. L., and Smith, R.W. (2002). Overview of flotation as a wastewater treatment technique. *Minerals Engineering*. 15: 139-155.
- Rybicki, E., Swiech, T., Lesniewska, E., Albinska, J., Szynkowska, M. I., Paryjczak, T., and Sypniewski, S. (2004). Changes in hazardous substances in cotton after mechanical and chemical treatments of textiles. *Fibres and Textiles in Eastern Europe*. 12: 67–73.
- Saier M. H. J. (1994). Computer-aided analyses of transport protein sequences; gleaning evidence concerning function, structure, biogenesis, and evolution. *Microbiology Reviews*. 58: 71-93.

Saier, M. H., Tam, R., Reizer A., and Reizer, J. (1994). Two novel families of bacterial

membrane proteins concerned with nodulation, cell division and transport. *Molecular Microbiology*. 11: 841-847.

- Saluja, B., and Sharma, V. (2014). Cadmium resistance mechanism in acidophilic and alkalophilic bacterial isolates and their application in bioremediation of metal-contaminated soil. *Soil and Sediment Contamination: An International Journal*. 23: 1-17.
- Saratale, R. G., Gandhi, S. S., Purankar, M. V., Kurade, M. B., Govindwar, S. P., Oh, S. E., and Saratale, G. D. (2013). Decolorization and detoxification of sulfonated azo dye CI Remazol Red and textile effluent by isolated *Lysinibacillus* sp. RGS. *Journal of Bioscience and Bioengineering*. 115: 658-667.
- Sengupta, A. K. (1994). Principles of Heavy Metals Separation: An Introduction. Environmental Separation of Heavy Metals – Engineering Processes. USA: Lewis Publishers.
- Siddiqui, M. F., Wahid, Z. A., and Sakinah, M. (2011). Bioremediation and biofouling perspective of real batik effluent by indigenous bacteria. *International Journal* of Chemical and Environmental Engineering. 2: 302-308.

Silver, S. (1996). Bacterial resistances to toxic metal ions: a review. Gene. 179: 9–19.

- Silver, S., and Phung, L. T. (2005). A bacterial view of the periodic table: genes and proteins for toxic inorganic ions. *Journal of Industrial Microbiology and Biotechnology*. 32: 587–605.
- Silver, S., Nucifora, G., and Phung, L. T. (1993). Human Menkes X-chromosome disease and the staphylococcal cadmium-resistance ATPase: a remarkable similarity in protein sequences. *Molecular Microbiology*. 10: 7–12.
- Singh, Y., Ramteke, P. W., Tripathy, A., and Shukla, P. K. (2013). Isolation and characterization of *Bacillus* resistant to multiple heavy metals. *International Journal of Current Microbiology and Applied Sciences*. 2: 525-530.
- Srivastava, N. K., and Majumder, C. B. (2008). Novel biofiltration methods for the treatment of heavy metals from industrial wastewater. *Journal of Hazardous Material*. 151: 1-8.

- Stanbury, P. F., Whitaker, A., and Hall, S. J. (1994). Principles of Fermentation Technology, 2<sup>nd</sup> Edn. (pp: 350). Oxford: Butterworth-Heinemann.
- Stohs, S. J., and Bagchi, D. (1995). Oxidative mechanisms in the toxicity of metalions. *Free Radical Biology and Medicine*. 18: 321–336.
- Sun, F., and Shao, Z. (2007). Biosorption and bioaccumulation of lead by *Penicillium* sp. Psf-2 isolated from the deep sea sediment of the Pacific Ocean. *Extremophiles*. 11: 853–858.
- Thelwell, C., Robinson, N. J., and Turner-Cavet, J. S. (1998) An SmtB-like repressor from Synechocystis PCC 6803 regulates a zinc exporter. Proceedings of the National Academy of Sciences of the United States of America. 95: 10728-10733.
- Trajanovska, S., Britz, M. L., and Bhave, M. (1997). Detection of heavy metal ion resistance genes in gram-positive and gram-negative bacteria isolated from a lead-contaminated site. *Biodegradation*. 8: 113–124.
- Tsai, K. J., and Linet, A. L. (1993). Formation of a phosphorylated enzyme intermediate by the cadA Cd<sup>2+</sup>-ATPase. *Archives of Biochemistry and Biophysics*. 305: 267-270.
- Vallee, B. L., and Ulmer, D. D. (1972). Biochemical effects of mercury, cadmium and lead. Annual Review of Biochemistry. 41: 91–128.
- Volesky, B. (2001). Detoxification of metal-bearing effluents: biosorption for the next century. *Hydrometallurgy*. 59: 203–216.
- Volesky, B., and Holan, Z. R. (1995). Biosorption of heavy metals. *Biotechnology Progress*. 11: 235-250.
- Wang, J., and Chen, C. (2009). Biosorbents for heavy metals removal and their future. *Biotechnology Advances*. 27: 195–226.
- Wang, J., and Chen, C. (2006). Biosorption of heavy metals by Saccharomyces cerevisiae: A review. Biotechnology Advances. 24: 427–451.
- Wang, J., Fan, Y., and Yao, Z. (2010). Isolation of a *Lysinibacillus fusiformis* strain with tetrodotoxin-producing ability from puffer fish *Fugu obscurus* and the characterization of this strain. *Toxicon*. 56: 640-643.

- Weast, R. C. (1984). *CRC handbook of chemistry and physics*. (64<sup>th</sup> ed.) Florida: Boca Raton CRC.
- Yilmaz, E. I. (2003). Metal tolerance and biosorption capacity of *Bacillus circulans* strain EB1. *Research in Microbiology*. 154: 409–415.
- Zanardini, E., Andreoni, V., Borina, S., Cappitellia, F., Daffonchio, D., Talottaa, P., Sorlinia, C., Ranallib, G., Brunic, S., and Cariatic, F. (1997). Lead-resistant microorganisms from red stains of marble of the Certosa of Pavia, Italy and use of nucleic acid based techniques for their detection. *International Biodeterioration and Biodegradation*. 40: 171–182.