

BIODEGRADATION AND METABOLIC PATHWAY OF CRESOL RED  
DYE BY SELECTED FUNGI

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*To my beloved mother, father, brother and sisters*

*Thanks for all your support, love and care...*

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## ABSTRACT

Massive amount of dye concentrations has been released into the environment as wastewater. Cresol red dyes remains as one of the most difficult dyes to be degraded. In this study, 20 different types of white rot fungi were isolated from a decayed wood in UTM forest and labelled as M 01-M 20. These fungi were screened for their ability to degrade cresol red dye after 30 days of incubation. New metabolic pathways of degradation of cresol red dyes by fungi M06 and M15 were explored in this research. Both M06 and M15 were chosen based on their ability to decolorize 10 ppm cresol red dye on agar and liquid medium. These fungi were further tested for their ability to degrade cresol red dye in liquid medium under several conditions which is carbon and nitrogen sources, pH, salinity and effect of different concentration of surfactant (Tween 80). From decolorization, M06 showed efficient decolorization with addition of glucose (60.01%), ammonium sulphate (67.22%), Tween 80 0.1 mL (53.82%), 50 g/L of NaCl concentration (59.64%) and pH 3 (67.50%). Meanwhile, M15 showed efficient degradation of cresol red dye with addition of glucose (72.85%), ammonium nitrate (82.40%), Tween 80 0.1 mL (74.35%), 150 g/L of NaCl concentration (78.64%) and pH 3 (85.46%). These fungi were then identified using 18S rRNA sequence analysis method. From the phylogenetic tree, fungi M06 belongs to *Trichoderma harzianum* and fungi M15 belongs to *Absidia spinosa*. Identification of metabolites was performed by combining several series of experiment through the use of UV-vis Spectrophotometer, Thin Layer Chromatography, and Gas Chromatography Mass Spectrophotometer. The analytical results showed that 2-hydroxybenzoic acid, and 2,4-dihydroxybenzoic acid were formed during the degradation of cresol red dye by *Trichoderma harzianum* M06. In the case of *Absidia spinosa* M15, benzoic acid and benzeneacetic acid were identified as the metabolic products produced during the degradation of cresol red dye. From the study, it is found that M 06 and M 15 have the capability to degrade cresol red dye. For further investigation, several experiments can be carried out to enhance the use of the isolated fungi in textile wastewater treatment.

## ABSTRAK

Terdapat sejumlah besar pewarna yang telah disingkirkan ke alam sekitar sebagai air sisaan. Salah satunya adalah pewarna 'cresol red' dan ia adalah salah satu pewarna yang sukar untuk disingkirkan. Dalam kajian ini, 20 kulat yang berlainan jenis telah dipilih dari hutan yang terdapat di sekitar kampus UTM dilabel sebagai M01 - M20. Kulat-kulat ini telah disaring berdasarkan keupayaannya untuk menyingkirkan pewarna 'cresol red' selepas 30 hari pengeraman. Laluan metabolik degradasi untuk pewarna 'cresol red' oleh kulat M06 dan M15 telah dikaji dalam penyelidikan ini. Selepas itu, keupayaan kedua-dua kulat M06 dan M15 ini untuk meyingkirkan pewarna 'cresol red' diuji dalam keadaan pepejal dan cecair dengan kepekatan pewarna 'cresol red' sebanyak 10 ppm. Kulat ini seterusnya telah diuji keupayaannya untuk menyingkirkan pewarna 'cresol red' dalam keadaan cecair dibawah kepelbagaian keadaan seperti sumber karbon dan nitrogen, pH, kemasinan dan kesan kepekatan surfaktan (Tween 80) yang berbeza. M06 menunjukkan penyingkiran warna efisien dengan penambahan glukosa (60.01%), ammonium sulfat (67.22%), Tween 80 0.1 mL (53.82%), 50 g / L kepekatan NaCl (59.64%) dan pH 3 (67.50%). Sementara itu, M15 menunjukkan degradasi efisien pewarna 'cresol red' dengan penambahan glukosa (72,85 %), ammonium nitrat ( 82,40 % ), Tween 80 0.1 mL ( 74,35 % ), 150 g / L kepekatan NaCl ( 78,64 % ) dan pH 3 ( 85.46 % ). Kulat-kulat ini telah dikenalpasti menggunakan kaedah 18s rRNA. Berdasarkan pokok filogenetik, kulat M06 adalah dari jenis *Trichoderma harzianum* dan kulat M15 adalah dari jenis *Absidia spinosa*. Seterusnya pengenalpastian produk metabolit telah dijalankan menggunakan UV-vis Spectrophotometer, Thin Layer Chromatography, dan Gas Chromatography Mass Spectrophotometer. Dari hasil analisis, dua produk metabolit iaitu 2, asid dihidrozybenzoik acid, dan 2-4 asid dihidroxybenzoik telah terhasil semasa penyingkiran pewarna 'cresol red' oleh *Trichoderma harzianum* M06. Untuk kulat M15, produk metabolit dijumpai adalah asid benzoic dan asid benzeneasetik semasa degradasi pewarna cresol red. Berdasarkan kejadian ini, didapati bahawa M 06 dan M 15 mempunyai keupayaan untuk menyingkirkan pewarna 'cresol red'. Untuk siasatan lanjut, beberapa eksperimen boleh dijalankan untuk meningkatkan keupayaan penggunaan kulat terpencil dalam rawatan air sisa tekstil.

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**LIST OF ABBREVIATIONS**

%	-	Percent
$\lambda$ max	-	Lambda Maximum
-C=O	-	Carbonyl
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	-	Ammonium Sulfate
°C	-	Celsius
°C/min	-	Celsius per Minute
C <sub>6</sub> H <sub>12</sub> O <sub>6</sub>	-	Sugar
C <sub>7</sub> H <sub>6</sub> O <sub>2</sub>	-	Benzoic Acid
C <sub>6</sub> H <sub>4</sub> (OH) <sub>2</sub>	-	Catechol
C <sub>7</sub> H <sub>6</sub> O <sub>3</sub>	-	Salicylic Acid
C	-	Carbon
CH <sub>3</sub>	-	Methyl
C.I.	-	Color Index
Cl	-	Chloride
Cl <sub>2</sub>	-	Chlorine
cm	-	Centimeter
COOH	-	Carboxyl
Cu <sup>+</sup>	-	Copper Ion
Cu <sup>2+</sup>	-	Copper (II) Ion
DNA	-	Deoxyribonucleic Acid
E' <sub>0</sub>	-	Redox Potential
e <sup>-</sup>	-	Electron
Fe <sup>3+</sup>	-	Iron (III) Ion
g/L	-	Gram per Liter
g/mol	-	Gram per Mole

GC	-	Gas Chromatography
GC-MS	-	Gas Chromatography-Mass Spectrometry
H <sub>2</sub> O <sub>2</sub>	-	Hydrogen Peroxide
H <sub>2</sub> O	-	Water
H <sup>+</sup>	-	Hydrogen Ion
HCL	-	<i>Hydrochloric Acid</i>
kPa	-	<i>Kilopascal</i>
Lac	-	<i>Laccase</i>
LMEs	-	Lignin-Modifying Enzymes
LiP	-	Lignin Peroxides
M <sup>+</sup>	-	Molecular Ion (peak)
M	-	Molar
MAE	-	Malt Extract Agar
Min	-	Minutes
mL	-	Milliliter
mg/L	-	Milligram Per Liter
mM	-	<i>Millimolar</i>
mm	-	<i>Milliliter</i>
MnP	-	Manganese Peroxides
Mn <sup>2+</sup>	-	<i>Manganese (II) Ion</i>
Mn <sup>3+</sup>	-	<i>Manganese (III) Ion</i>
MnO <sub>4</sub>	-	<i>Permanganate</i>
m/z	-	<i>Mass Spectrum (Mass-to-Charge Ratio)</i>
N	-	<i>Nitrogen</i>
NaOH	-	Sodium Hydroxide
NH <sub>2</sub>	-	Amines
NH <sub>4</sub> NO <sub>3</sub>	-	Ammonium Nitrate
NO	-	Nitro
NO <sub>2</sub>	-	Amino
Nm	-	Nanometer
O <sub>2</sub>	-	Oxygen
O <sub>3</sub>	-	Ozone

OH	-	Hydroxyl
OH <sup>·</sup>	-	Hydroxyl Radical
PAH	-	Polycyclic Aromatic Hydrocarbons
PCR	-	Polymerase Chain Reaction
pH	-	Power Hydrogen
ppm	-	Part per Million
$R_f$	-	Retention Factor
rpm	-	Rotor per Minute
rRNA	-	Ribosomal Ribonucleic Acid
S	-	Sulfur
Si	-	Silica
SO <sub>3</sub>	-	Sulfur Trioxide
TLC	-	Thin Layer Chromatography
TMS	-	Trimethylsilylation
$t_R$	-	Retention Time
UV	-	Ultraviolet
UV-Vis	-	Ultraviolet-Visible
v/v	-	Volume per Volume
w/v	-	Weight per Volume
μL	-	Microliter
μg	-	Microgram

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of Study

Synthetic dyes are widely used in many areas. It has been used in various branches of textile industry (Gupta *et al.*, 1992; Sokolowska-Gajda *et al.*, 1996), of the leather tanning industry (Tuñay *et al.*, 1999; Kabadasil *et al.*, 1999) in paper production (Ivanov *et al.*, 1996), in food technology (Bhat and Mathur, 1998; Slampova *et al.*, 2001), in agricultural research (Cook and Linden, 1997; Kross *et al.*, 1996), in light-harvesting arrays (Wagner and Lindsey, 1996), in photoelectrochemical cells (Wrobel *et al.*, 2001), and in hair colorings (Scarpi *et al.*, 1998). Moreover, synthetic dyes have been used for the control of the efficacy of sewage (Morgan-Sagastume *et al.*, 1997) and wastewater treatment (Hsu and Chiang, 1997; Orhon *et al.*, 1999), for the determination of specific surface area of activated sludge (Sorensen and Wakeman, 1996) for ground water tracing (Field *et al.*, 1995). The metabolic pathway during the degradation of cresol red dye is also been investigated through different series of steps.

Due to rapid growth of industrialization and urbanization, many chemicals including dyes are produced and used in day-to-day life. Dyes are synthetic and aromatic molecular structural compounds. According to their dissociation in an aqueous solution, dyes can be classified as acid, direct reactive dyes (anionic), basic dyes (cationic) and disperse dyes (nonionic) (Sathiya *et al.*, 2007). Dye solutions held together by physical adsorption through metals and salts using covalent bonds. Synthetic dyes have been used increasingly in textile and dyeing industries because



of their ease and cost effectiveness in synthesis, firmness and variety in color, compared to that of natural dyes

The exact amount of dyes produced in the world is not known. It is estimated to be over 10,000 tons per year. Exact data on the quantity of dyes discharged in the environment are also not available. It is assumed that a loss of 1–2% in production and 1–10% loss in use are a fair estimate. For reactive dyes, the loss can be about 4% (Forgacs *et al.*, 2004). Due to large-scale production and extensive application, synthetic dyes can cause considerable environmental pollution and are serious health-risk factors. Although, the growing impact of environmental protection on industrial development promotes the development of ecofriendly technologies (Desphande, 2001), reduced consumption of freshwater and lower output of wastewater (Knittel and Schollmeyer, 1996; Petek and Glavic, 1996), the release of important amounts of synthetic dyes to the environment causes public concern, legislation problems and are a serious challenge to environmental scientists.

Triphenylmethane (TPM) dyes are used extensively in textile industries for dyeing nylon, polyacrylonitrile modified nylon, wool, silk, and cotton. Dyes are recalcitrant molecules which are difficult to degrade biologically. From the treatment point of view, the degradation of dyes has received considerable attention. Conventional wastewater treatment processes are suitable for stabilization of nonxenobiotic compounds whereas these processes do not work well with the xenobiotic compounds. Environmental regulations in most of the countries now have made it mandatory to decolorize the dye wastewater prior to discharge. Increasing concerns about color in the effluents are leading to the worldwide efforts to develop more effective color removal processes (Azmi *et al.*, 1998).

Cresol red dyes belong to the group of triphenylmethane dye. These dyes have been released to the environment as synthetic dyes (Gill *et al.*, 2012). TPM dyes has been extensively used in cloth industry as dyeing silk, polyacrylonitrile customize nylon, coat, nylon, and cotton. They also are being used in the biological area as biological stains and marker. Leather and paper industries are also the major consumers of the TPM dyes. These groups of dyes are also used for coloring plastics,

varnish, gasoline, fats, oil, and waxes. Different types of triphenylmethane dyes are also used in cosmetic and food industries (Azmi *et al.*, 1998).

The majority of dyes are extremely poisonous and mutagenic. Artificial sources and complex aromatic configuration enable them to be more resistant to biodegradation activities (Godlewska *et al.*, 2009). Some of TPM dyes have been found in the soil due to improper waste disposal (Gill *et al.*, 2012). Therefore, innovative treatment technologies are needed to be investigated. Decolorization of dye wastewater by fungal metabolic activities is the subject of many studies. Fungi from the Basidiomycetes group, known as white rot fungi are a heterogeneous group of microorganisms but have a common capability to degrade lignin, as well as, other wood components (Kirk and Farrell, 1987). The white rot fungi are by far the most efficient ligninolytic microorganisms. They are able to degrade various recalcitrant pollutants including dyes (Sathiya *et al.*, 2007).

Extensive studies on biodegradation of synthetic dyes by ligninolytic fungi has been done with *Phanerochaete chrysosporium* (Paszczyński and Crawford 1995). White rot fungus are capable to discard dyes from industrial effluents. This fungus has been investigated for their ability to degrade resistant organic pollutants such as polyaromatic hydrocarbons, chlorophenols and polychlorinated biphenyl. The decolorization of phenol red, methylene blue, coomassive blue, and dextran blue has been applied to determine ligninolytic activity (Sathiya *et al.*, 2007).

The purpose of this study is to identify the isolated fungi using 18s rRNA method to know the identity of the targeted fungi by using bioinformatics tools. The isolated fungi are screened to determine their ability to degrade and decolorize cresol red dyes. The metabolic pathway during the degradation of cresol red dye is also being investigated through different series of steps.

## 1.2 Problem Statement

In Malaysia, textile industry sectors has been the seventh largest contributor to total earnings from manufactured exports, due to its high demanding market either from inside and outside Malaysia (Pang and Abdullah, 2013). Resulting from this, there are massive amount of dye concentrations that has been discharged to the environment as wastewaters. About 743.99 metric tons per year of waste has been released by the textile industry according to a report from Department of Environment of Malaysia (Environmental Quality Report, 2007). Approximately, 40-65 L of textile sewage is produced per kg of fabric formed by the textile industry. For the time being, there is no effective procedure to treat these waste water materials (Azmi *et al.*, 1998). In much worse scenario, the wastewater materials were discharged into the environment at high concentrations by the industries. Dyes may affect photosynthetic activity in water because they prevent light access through the water surface thus causing shortage of oxygen and subsequently decrease the survival rate of aquatic organism in the environment. Most of the dyes produced by textiles industries are greatly toxic and mutagenic.

The release of dyes into wastewaters by various industries poses serious environmental problems due to various dyes' persistent and recalcitrant nature. In Malaysia, a lot of industries including the textile industries also increased the water pollution level. From this industry, the water pollution has become worst especially in colour point of view. The effluent from the textile industry has a high level of colour concentration. Because of this, the best water treatment process is needed to encounter this pollution problem so that the best result will be achieved.

Presently, most of the processes used for the treatment of dye wastewater are chemical processes such as adsorption, coagulation, ion exchange and reduction. These processes are costly, produce large amount of sludge, and less efficient. Since then, biological processes including microorganisms application are getting extensive attention because they are cost effective, environmental friendly, and produce small quantities of sludge (Azmi *et al.*, 1998). In this study, fungi isolated from the environment were investigated for their potential in treating cresol red dye.

### **1.3 Research Objectives**

Several objectives are constructed as listed below. These objectives serve as guidelines in completing this study.

1. To screen, isolate, and identify fungi from nature that have the ability to degrade Cresol Red dyes.
2. To investigate the effect of different parameters to the degradation of Cresol Red dyes.
3. To investigate the metabolic pathway for cresol red dyes

### **1.4 Scope of the Study**

In this study, the degradation of cresol red dye by the selected fungi was investigated by measuring its decolorization percentage during the incubation period. After that, the capabilities of the isolated fungi to degrade cresol red dyes were further tested under different conditions with alteration of carbon sources, nitrogen sources, effect of surfactant (Tween 80), salinity and different initial pH. After that, the fungi were identified using 18s rRNA methods. Subsequently, the metabolic pathway employed by the fungi in degrading cresol red dye was also investigated using series of extraction methods. These methods were tested using UV-Spectrophotometre, TLC, Column Chromatography and GC-MS.

### **1.5 Significance of the Study**

The significance of this study is to provide an alternative method in term of decolorization and degradation of cresol red dyes wastewater using fungi which was isolated from the forest. In addition, biological process is more effective and more environmental friendly in the degradation of the synthetic dyes and can become an effective method in solving the problem of textile wastewater treatment.

## REFERENCES

- Abedin, R. M. A. (2009). Decolorization and biodegradation of crystal violet and malachite green by *Fusarium solani* (Martius) Saccardo. A comparative study on biosorption of dyes by the dead fungal biomass. *Am J Bot.* 2, 01–15.
- Adosinda, M., Martins, M., Ferreira, I. C., Santos, I. M., Queiroz, M. J. and Lima, N. (2001). Biodegradation of bioaccessible textile azo dyes by *Phanerochaete crysoporium*. *Journal of Biotechnology.* 89: 91–98.
- Aguilera, L. M., Griffiths, R. P., Caldwell B. A. (1993). Nitrogen in ectomycorrhizal mat and non-mat soils of different-age Douglas-fir forests. *Soil BiolBiochem.* 25, 1015-1019.
- Alalewi, A., Jiang C. (2012) Bacterial Influence on Textile Wastewater Decolorization. *J of Env Protection.* 3, 889-901.
- Ali, H. (2010). Biodegradation of Synthetic Dyes—A Review. *Water Air Soil Pollut.* 213, 251–273.
- Ali, H., Ahmad, W., Haq, T. (2009). Decolorization and degradation of malachite green by *Aspergillus flavus* and *Alternaria solani*. *Afr J Biotechnol.* 8, 1574–1576.

- Annur, M. S. M., Adnan, S., Vikineswary, S., Chisti, Y. (2009). Kinetics and energetics of azo dye decolorization by *Pycnoporus sanguineus*. *Water Air Soil Pollut.* 202, 179–188.
- Anliker, R. (1979) Ecotoxicological Assessment of Dyes with particular reference to ETAD's Activities. *Journal of the Society of Dyers and Colourists.* 95, 317–326.
- Asther, M., Corrieu, G., Drapron, R. and Odier, E. (1987). Effect of Tween 80 and oleic acid on ligninase production by *Phanerochaete chrysosporium* INA-12. *Enzyme and Microbial Technology.* 9, 245–249.
- Azmi, W., Sani, R. K., and Banerjee, U. C. (1998). Biodegradation of triphenylmethane dyes. *Enzyme Microbial Technol.* 22, 185-191.
- Belsare, D. K. and Prasad, D. Y. (1988). Decolourization of effluent from the bagasse based pulp mills by white rot fungus. *Schizophyllum commune*. *Applied Microbiology and Biotechnology.* 28:301-304.
- Benny, G. L., Humber, R. A., and Morton, J. B. (2001). Zygomycota: zygomycetes. In: *The Mycota. Part A. J SystEvol.* 7, 113-146.
- Bhat R,V., Mathur P. (1998). Changing scenario of food colours in India. *Curr Sci.* 74:198–202.
- Blánquez, P., Casas, N., Font, X., Gabarrell, X., Sarrà, M., Caminal, G. and Vicent, T. (2004). Mechanism of textile metal dye biotransformation by *Trametes versicolor*. *Water Res.* 38, 2166–2172
- Bu'lock, J. D. (1980). In *The Biosynthesis of Mycotoxins, a Study in Secondary Metabolism* (P. S. Steyn, ed.). Academic Press, New York.

- Bruce E. Rittmann, Perry L. McCarty Environmental biotechnology: principles and applications, McGraw-Hill, 2001 - Business & Economics- 754 pages
- Carlile, M. J., Sarah, C. W. and Graham. (2001). The Fungi. London: Academic Press a Harcourt Science and Technology Company.
- Carmen,Z. and Daniela, S. (2012). Textile Organic Dyes – Characteristics, Polluting Effects and Separation/Elimination Procedures from Industrial Effluents – A Critical Overview, Organic Pollutants Ten Years After the Stockholm Convention - Environmental and Analytical Update, Dr. Tomasz Puzyn (Ed.)
- Cheremisinoff, N.P., (2002). Handbook of Water and Wastewater Treatment Technologies.Butterworth-Heinemann, Boston.
- Chen, K. C., Wu, J. Y., Liou, D. J., & Huang, S. C. J. (2003). Decolorization of the textile dyes by newly isolated bacterial strains. *J. of Biotechnol.* 101, 57-68.
- Cohen, R. Hadar, Y. and Yarden, O. (2001). Transcript and activity levels of different *Pleurotostreatus* peroxidases are differentially affected by Mn<sup>2+</sup>. *Environmental Microbiology*.3: 312-322.
- Cook S, M, F., Linden D, R. (1997) Use of rhodamine WT to facilitate dilution andanalysis of atrazine samples in short-term transport studies. *J Environ Qual.* 26:1438– 41.
- Cooke, R. C. and Whipps, J. M. (1993).*Ecophysiology of fungi*.Blackwell Scientific Publication, Oxford, UK.

Daiyo, D., Guo, J., Zeng, G., & Sun, G. (2008). Decolorization of anthraquinone, triphenylmethane and azo dyes by a new isolated *Bacillus cereus* strain DC11q. *IntBiodeteriorationBiodegrad.* 62, 263-269.

Desphande SD. Ecofriendly dyeing of synthetic fibres. *Ind J Fibre Text Res* 2001;26:136–42.

Department of Environment.Putrajaya, “Environmental Quality Report”, (2007),

E. Bruce .Rittmann and Perry,”Priciples and Applications,” Environmental Biotechnology USA: McGraw Hill,2001.

Field M, S., Wilhelm R, G., Quinlan J, F., Aley T, J. (1995). An assessment of the potential adverse properties of fluorescent tracer dyes used for groundwater tracing. *Environ Monit Assess* 38,75 – 97.

Finlay, R. D., Frostegard, A. and Sonnerfeld, A-M.(1992). Utilization of organic and inorganic nitrogen sources by ectomycorrhizal fungi in pure culture and in symbiosis with *Pinuscontoria*Dougl.ex Loud. *New Phytogist.*120, 105-115.

Forgacs,a E., Cserhati, T., & Oros, G. (2004). Removal of synthetic dyes from wastewaters: a review*Environ Int.* 30, 953-971.

Forss, J., & Welander, U. (2009). Decolorization of reactive azo dyes with microorganisms growing on soft wood chips. *IntBiodeteriorationBiodegrad.* 63, 752-758.

Gaucher, G. M.(1969). An Introduction to Chromatography. *J. Chem. Educ*Robinson. 46, 729-733.



- Gill, PK., Arora, DS., and Chander, M. (2002). Biodecolourization of azo and triphenylmethane dyes by *Dichomitussqualens* and *Phlebiaspp.*. *J. Ind. Microbiol. Biotechnol.* 28, 201-203.
- Godlewska, E. Z., Przysaś, W., and Sota, E. G. (2009). Decolourization Of Triphenylmethane Dyes And Ecotoxicity Of Their End Products. *Environ. Prot. Eng.* 35, 44-100.
- Godlewska, E. Z., Przysaś, W., and Sota, E. G. (2012). Biological Removal of Azo and Triphenylmethane Dyes and Toxicity of Process By-Products. *Water Air Soil Pollut.* 223, 1581-1592.
- Gregory, P. Dyes and dyes intermediates. In: *Encyclopedia of Chemical Technology* Vol. 8 (Kroschwitz, J. I., Ed.). John Wiley & Sons. New York. 1993. 544-545
- Grondona, I., M. R. Hermosa, M. Tejada, M. D. Gomis, P. F. Mateos, P. Bridge, E. Monte, and I. Garcí'a-Acha. 1997. Physiological and biochemical characterization of *Trichoderma harzianum*, a biological control agent against soilborne fungal plant pathogens. *Appl. Environ. Microbiol.* 63,3189-3198.
- Gupta G, S., Shukla S, P., Prasad G., Singh V, N. (1992). China clay as an adsorbent for dye house wastewaters. *Environ Technol.* 13,925– 36.
- Hadibarata, T., and Kristanti, R. A. (2012). Fate and cometabolic degradation of benzo[a]pyrene by white-rot fungus *Armillaria* sp. F022. *Bioresour. Technol.* 107, 314-318.

- Hadibarata, T., Yosoff, A. R. M. and Kristanti, R. A. (2011b). Decolorization and metabolism of anthraquinone-type dye by laccase of white-rot fungi *polyporus* sp. S133. *Water Air Soil Pollut.* DOI:10.1007/s11270-011-0914-6.
- Hadibarata, T., Yosoff, A. R. M., Salmiati, Hidayat, T. and Kristanti, R. A. (2011). Decolorization of azo, triphenylmethane and anthraquinone dyes by laccase of a newly isolated *Armillaria* sp. F022. *Water Air Soil Pollut.* DOI:10.1007/s11270-011-0922-6.
- Hai, F. I., Yamamoto, K., Nakajima, F., & Fukushi, K. (2008). Removal of structurally different dyes in submerged membrane fungi reactor-biosorption/PAC-adsorption, membrane retention and biodegradation. *J. Membr. Sci.* 325, 395–403.
- Hall, G. G. (2008). *Phylogenetic Trees Made Easy*. USA: Sunderland, MA: Sinauer Associates.
- Haugland, R. A., and J. L. Heckman. (1998). Identification of putative sequence-specific PCR primers for detection of the toxigenic fungal species *Stachybotrys chartarum*. *Mol. Cell. Probes.* 12, 387-396.
- Hawksworth, D. L., Kirk, T. M., Sutton, B. C., and Pegler, D. N. (1995). *Ainsworth and Biesby's Dictionary of Fungi*. UK: CAB International
- Heinfling, A., Bergbauer, M., and Szewzyk, U. (1997). Biodegradation of azo and phthalocyanine dyes by *Trametes versicolor* and *Bjerkandera adusta*. *Appl. Microbiol. Biotechnol.* 48, 261-266.
- Higson, F. K. (1991). Degradation of xenobiotics by white rot fungi. *Reviews of Environmental Contamination and Toxicology.* 122, 111–152

- Howard, R., Abotsi, L., Rensburg, E-J. V. E. and Howard, S, L. (2003). Lignocellulose biotechnology: issues of bioconversion and enzyme production. *African Journal of Biotechnology*.2: 602-619.
- Hsu T, C., Chiang C, S. (1997). Activated sludge treatment of dispersed dye factory wastewater. *J Environ Sci Health Part A, Environ Sci Eng Toxic Hazard Substance Control A* 32,1921– 32.
- Husseiny, S. M. (2008). Biodegradation of the reactive and direct dyes using Egyptian isolates. *J. Appl. Sci. Res*4, 599–606.
- Hofrichter, M., Bublitz, F. and Fritsche, W. (1997). Fungal attack on coal. II. solubilization of low-rank coal filamentous fungi. *Fuel Processing Technology*. 52, 55–64.
- Ivanov K., Gruber E., Schempp W., Kirov D, (1996). Possibilities of using zeolite as filler and carrier for dyestuffs in paper. *Das Papier* 50,456– 60 [in German].
- Juhasz, T., Szengyel, Z., Reczey, K., Siika-Aho, M. and Viikari, L. (2005). Characterization of cellulases and hemicellulases produced by *Trichoderma reesei* on various carbon sources. *Process Biochemistry*.40: 3519-3525.
- Jadhav, U. U., Dawkar, V. V., Tamboli, D. P. and Govindwar, S. P. (2009). Purification and characterization of veratryl alcohol oxidase from *Comomonas* sp. UVS and its role in decolorization of textile dyes. *Biotechnology and Bioprocess Engineering*. 14: 369-376.
- Jadhav, J. P., Kalyani, D. C., Phugare, S. S. and Govindwar, S. P. (2010). Evaluation of the efficacy of a bacterial consortium for the removal of color, reduction

of heavy metals, and toxicity from textile dye effluent. *Bioresource Technology*. 101: 165-173.

Jennings, D. (1995). *Physiology of Fungal Nutrition*. Cambridge University Press, New York.

Kaal, J. E. E., A. J. Field and W. T. Joyce (1995). Increasing ligninolytic enzyme activities in several white-rot Basidiomycetes by nitrogen-sufficient media. *Bioresour. Technol.* 53, 133-139.

Kabadasil I., Tu'nay O., Orhon D. (1999). Wastewater control and management in a leather tanning district. *Water Sci Technol.* 1,261– 7.

Kariminiaae-Hamedani, H. R., Sakurai, A. and Sakakibara, M. (2007). Decolorization of synthetic dyes by a new manganese peroxidase-producing white rot fungus. *Dyes and Pigments*. 72: 157–162.

Kaushik, P., & Malik, A. (2009). Fungal dye decolorization: Recent advances and future potential. *Environment International*. 35, 127-141.

Kirk, T. K. and Farrell, R. L. (1987). Enzymatic combustion. The microbial degradation lignin. *Annual Review Microbiology*. 41: 465-505.

Knittel D, Schollmeyer E. (1996). Prevention of water pollution in dyeing processes of synthetic textiles. *Eur Water Pollut Control* 6,6– 10.

Kopka, J., Fernie, A. F., Weckwerth, W., Gibon, Y., and Stitt, M. (2004) Metabolite profiling in Plant Biology: Platforms and Destinations. *Genome Biol.* 6,109-117.

Kothandaraman, U., Aboo, K. M., and Sastry, C. (1976). Characteristics of wastes from a textile mill. *Indian J. Environ. Health*. 18,99-112.

- Kotterman, M. J. J., Rietberg, H. J., Hage, A. and Field, J. A. (1998). Polycyclic aromatic hydrocarbon oxidation by the white-rot fungus *Bjerkandera* sp. strain BOS55 in the presence of nonionic surfactants. *Biotechnology and Bioengineering*. 57, 220–227.
- Kristina S, I., Charles B, K,. (2014). Recurrent furunculosis – challenges and management: a review. *Clinical, Cosmetic and Investigational Dermatology*. 7 59–64
- Kross B, C., Nicholson H, F., Ogilvie LK. (1996) Methods development study for measuring pesticide exposure to golf course workers using video imaging techniques. *Appl Occup Environ Hyg*. 11,1346–51.
- Kuhad, R. C., Sood, N., Tripathi, K. K., Singh, A., & Ward, O. P. (2004). Developments in microbial methods for the treatment of dye effluents. *Adv Appl Microbiol*. 56,185-213.
- Levin, L., Papinutti, L. and Forchiassin, F. (2004). Evaluation of Argentinean white rot fungi for their ability to produce lignin-modifying enzymes and decolorize industrial dyes. *Bioresources Technology*. 94: 169–176.
- Lee, J.-W., Choi, S.-P., Thiruvengkatachari, R., Shim, W.-G., Moon, H.. (2006). Evaluation of the performance of adsorption and coagulation processes for the maximum removal of reactive dyes. *Dyes Pigments* 69, 196-203.
- Machado, K. M. G., Matheus, D.R., and Bononi, V. L. R.(2005). Ligninolytic enzymes production and remazol brilliant blue R decolorization by tropical brazilian basidiomycetes fungi. *Braz. J. Microbiol*. 36,246-252.

- Maihafizah., (2004), "Microbiol Physiology of biofilm and its application in colour and heavy metals removal from textile wastewater". BSc Thesis, University Technology Malaysia.
- Mishra, A., Bajpai, M., (2006). The flocculation performance of *Tamarindus mucilagein* relation to removal of vat and direct dyes. *Bioresour. Technol.* 97, 1055-1059.
- Mishra, A., Bajpai, M., Pandey, S..(2006). Removal of dyes by biodegradable flocculants:a lab scale investigation. *Sep. Sci. Technol.* 41, 583-593.
- Morgan-Sagastume, J, M., Jimenez, B, Noyola, A. (1997) Tracer studies in a laboratory and pilot scale UASB reactor. *Environ Technol.* 18,817– 26.
- Mueller, A., Duechting, P., and Weiler, E. W. (2002) A multiplex GC-MS/MS technique for the sensitive and quantitative single-run analysis of acidic phytohormones and related compounds, and its application to *Arabidopsis thaliana*. *Plant*, 216,44-56.
- Nigam, P., Banat, I. M., Singh, D. and Marchant, R. (1996).Microbial process for the decolorization of textile effluent containing azo, diazo and reactive dyes.*Process Biochemistry.* 31: 435–442.
- Novotny, C., Rawal, B., Bhatt, M., Milind, P., Sasek, V., and Molitoris, H.P.(2001). Capacity of *Irpex lacteus* and *Pleurotus ostreatus* for decolorization of chemically different dyes. *J Biotech.* 89, 113-122.
- Novotny, C., Svobodova, K., Erbanova, P., Cajthaml, T., Kasinath, A., Lang, E., et al. (2004a). Ligninolytic fungi in bioremediation: extracellular enzyme production and degradation rate. *Soil Biol Biochem.* 36,1545–1551.

- Novotny, C., Svobodova, K., Kasinath, A., & Erbanova, P. (2004b). Biodegradation of synthetic dyes by *Irpex lacteus* under various growth conditions. *IntBiodeteriorationBiodegrad.* 54, 215–223.
- Nozaki, K., Beh, C. H., Mizuno, M., Isobe, T., Shiroishi, M., Kanda, T., et al. (2008). Screening and investigation of dye decolorization activities of basidiomycetes. *J. Biosci. Bioeng.* 105, 69-72.
- Nyanhongo, G., Gomes, J., Gübitz, G., Zvauya, R., Read, J. and Steiner, W. (2002). Production of laccase by a newly isolated strain of *Trametesmodesta*. *Bioresource Technology.* 84: 259-263.
- O'Donnell, K.L. 1979. *Zygomycetes in culture*. Palfrey Contributions in Botany.No. 2.Department of Botany, University of Georgia, Athens, Georgia.257 p.
- Orhon, D, So"zen S, Go"rgu"n E, Cokgo" r E, Artan N. (1999). Technological aspects of wastewater management in coastal tourist areas. *Water Sci Technol* 8,177– 84.
- Pandey, A., Singh, P., & Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *IntBiodeteriorationBiodegrad.* 59, 73-84.
- Pang, Y. L. and Abdullah, A. Z. (2013). Current Status of Textile Wastewater Management and Research Progress in Malaysia: A Review. *Clean Soil Air Water.* doi: 10.1002/clen.201000318.
- Paszczynski A, Crawford RL (1995). Potential for bioremediation of xenobiotic compounds by the white rot fungus *Phanerochaete chrysosporium*. *Biotechnol. prog.* 11, 368-379.

- Park, C., Lee, M., Lee, B., Kim, S. W., Chase, H. A., Lee, J., et al. (2007). Biodegradation and biosorption for decolorization of synthetic dyes by *Funalia trogii*. *BiochemEng J.* 36, 59-65.
- Petek J, Glavic P. (1996) An integral approach to waste minimization in process industries. *Resour Conserv Recycl* 17,169–88.
- Pointing, S. B., & Vrijmoed, L. L. P. (2000). Decolorization of azo and triphenylmethane dyes by *Pycnoporus sanguineus* producing laccase as the sole phenoloxidase. *World JMicrobBiot.* 16, 317-318.
- Ramalho, P. A. (2005). Degradation of dyes with microorganisms-Studies with ascomycete yeasts. PhD thesis, Biology Department, University of Minho
- Ramsay, J. A., & Nguyen, T. (2002). Decoloration of textile dyes by *Trametes versicolor* and its effect on dye toxicity. *Biotechnol.Lett.* 24, 1757–1761.
- Randerath, K., “Thin Layer Chromatography” (2<sup>nd</sup> Ed), Academic Press, New York, 1966.
- Reddy, C. A. (1995). The potential of white rot fungi in the treatment of pollutants. *Curr.Opin.Cell.Biol.* 6, 320-328.
- Reife. A.(1993). Dyes; environmental chemistry. In: *Encyclopedia of Chemical Technology* Vol. 8 (Kroschwitz, J. I. Ed.). John Wiley & Sons, New York., 754
- Ribes, J. A., Vanover-Sams C. L., and Baker D. J.(2000). Zygomycetes in human disease. *ClinMicrobiol Rev.* 13, 236–301.



- Ritz, N., Ammann R. A., Casaulta A. C, Gugger, M., Jatton, K., Schmid R. A., Aebi, C.(2005). Failure of voriconazole to cure disseminated zygomycosis in an immunocompromised child. *Eur J Pediatr.* 164, 231–235
- Robinson, T., McMullan, G., Marchant, R., & Nigam, P. (2001). Remediation of dyes in textile effluent: A critical review on current treatment technologies with a proposed alternative. *Bioresour.Technol.* 77, 247-255.
- Rogalski, J. and Leonowicz, A. (1992).Phlebiaradiatalaccase forms induced by veratric acid, and xylidine in relation to lignin peroxidase, and manganese-dependent peroxidase. *ActaBiotechnologica.* 12, 213–221.
- Rosen, M. J. (1989). Surfactants and Interfacial Phenomena.Wiley, NewYork.
- Ryan, D., Shellie, R., Tranchida, P., Casilli, A.,Mondello, L., and Marriott, P. (2004). Analysis of roasted coffee bean volatiles by using comprehensive two-dimensional gas Chromatography-time-of-flight mass spectrometry. *J Chromatogr.* 1054, 57–65.
- Salami, M. and Vandamme, A. M. (2003). The Phylogenetic Hand Book: A Practical Approach to DNA And Protein Phylogeny. UK; Cambridge University publisher.
- Sarnthima, R., Khammuang, S., & Svasti, J. (2009). Extracellular ligninolytic enzymes by *Lentinus polychrous* under solid-state fermentation of potential agro-industrial wastes and their effectiveness in decolorization of synthetic dyes. *Biotechnol. Bioprocess Eng.* 14, 513–522.

- Scarpi, C., Ninci, F., Centini, M., Anselmi, C. (1998) High-performance liquid chromatography determination of direct and temporary dyes in natural hair colourings. *J Chromatogr, A* 796,319– 25.
- Selvam, K., Swaminathan, K., & Chae, K. S. (2003). Decolorization of azo dyes and a dye industry effluent by a white rot fungus *Thelephora* sp. *Bioresour.Technol.* 88, 115-119.
- Shah, V., and Nerud, F. (2002). Lignin degrading system of white-rot fungi and its exploitation for dye decolorization. Review. *Can J Microbiol.* 48, 857-870.
- Shedbalkar, U., Dhanve, R., & Jadhav, J. (2008). Biodegradation of triphenylmethane dye Cotton blue by *Penicillium ochrochloron* MTCC 517. *J. Hazard. Mater.* 157, 472–479.
- Sinha, A. E., Hope, J. L., Prazen, B. J., Nilsson, E. J., Jack, R. M., and Synovec, R. E. (2004b). Algorithm for locating analytes of interest based on mass spectral similarity in GC×GC-TOF-MS data: analysis of metabolites in human infant urine. *J Chromatogr.* 1058,209–215.
- Sinha, A. E., Fraga, C. G., Prazen, B. J., and Synovec, R. E (2004a). Trilinear chemometric analysis of twodimensional comprehensive gas chromatography-time-of-flight mass spectrometry data. *J Chromatogr.* 1027,269–277.
- Sinha, A. E., Prazen, B. J., and Synovec, R. E. (2004c). Trends in chemometric analysis of comprehensive two-dimensional separations. *Anal Bioanal Chem* 378:1948–1951
- Slampova, A., Smela, D., Vondrackova, A., Jancarova, .I, Kuban, V. (2001). Determination of synthetic colorants in foodstuffs. *Chem Listy.* 95, 163– 8.

- Sokolowska-Gajda J, Freeman HS, Reife A. (1996). Synthetic dyes based on environmental considerations: 2. Iron complexed formazan dyes. *DyesPigm.* 30,1–20.
- Sherma, J. (2000). Planar chromatography. *Analytical Chemistry*. 72: 9-25
- Skoog, D. A., Holler, E. J. and Nieman, T. A. (1999). Principles of instrumental analysis 5<sup>th</sup> edition. Philadelphia: Saunders collage publishing.
- Somasiri W, Ruan W, Xiufen L, Jian C. (2006). Decolourization of textile wastewater containing acid dyes in UASB reactor system under mixed anaerobic granular sludge. *Electronic J Environ Agricul Food Chem.* 5(1), 1224–1234.
- Sorensen BL, Wakeman RJ. (1996). Filtration characterization and specific surface area measurement of activated sludge by rhodamine B adsorption. *Water Res.* 30, 115– 21.
- Steffen, K. T., Hofrichter, M. and Hatakka, A. (2000). Mineralisation of <sup>14</sup>C-labelled synthetic lignin and ligninolytic enzyme activities of litter-decomposing basidiomycetous fungi. *Applied Microbiology and Biotechnology.* 54, 819–825
- Stahl, E., “Thin Layer Chromatography ; A Laboratory Handbook,” Academic Press, New York, 1963.
- Stajic, M., L. Persky, D. Friesem, Y. Hadar, S. P. Wasser, E. Nevo and J. Vukojevic (2006). Effect of different carbon and nitrogen sources on laccase and peroxidases production by selected *Pleurotus* species. *Enzyme Microb Tech* 38, 65-73.

- Strainh., H., And Sherma, J, and Grandolfo, M., (1967). Analytical Chemistry. 39, 926
- Swamy, J., and Ramsay, J.A.(1999). The evaluation of white rot fungi in the decoloration of textile dyes. *Enzyme Microb Tech*, 24, 130-137.
- Thakur, D., Bora, T. C., Bordoloi, G. N. and Mazumdar, S. (2009). Influence of nutrition and culturing conditions for optimum growth and antimicrobial metabolite production by *Streptomyces* sp. 201. *J Med Mycol* 19,161-167
- Thirion-Delalande, C., Guillot, J., Jensen, H. E, Crespeau, F. L., Bernex, F.,(2005). Disseminated acute concomitant aspergillosis and mucormycosis in a pony. *Journal of Veterinary Medicine A*. 52, 121–124
- Tien, M. and Kirk, T. K. (1983). Lignin-degrading enzyme from the Hymenomycete *Phanerochaete chrysosporium* Burds. *Science*. 221: 661–663
- Tu'nay O, Kabdasli I, Ohron D, Cansever G. (1999) Use and minimalization of water in leather tanning processes. *Water Sci Technol* 1,237– 44.
- Turner, W. B. (1971). Fungal Metabolites. Academic Press, London.
- Turner, W. B. and Aldridge, D. C. (1983) Fungal Metabolites 11. Academic Press, London.
- Umezawa, T. and Higuchi, T. (1987). Mechanism of aromatic ring cleavage of h-O-4 lignin substructure models by lignin proxidase. *FEBS Letters*. 218: 255–260.
- Van Deursen, M. M., Beens, J., Janssen, H. G., Leclercq, P. A., and Cramers, C. A. (2000) Evaluation of time-offlight mass spectrometric detection for fast gas chromatography. *J Chromatogr*. 878, 205-213.

- Von Arx, J.A. (1984). On Mucoraceae s. str. and other families of the Mucorales. *Sydowia* 35:10-26.
- Wagner R,W, Lindsey JS. (1996). Boron-dipyrrromethane dyes for incorporation in synthetic multi-pigment light-harvesting arrays. *Pure Appl Chem* 68,1373–80.
- Wesenberg, D., Kyriakides, I., and Agathos, S.N. (2003). White rot fungi and their enzymes for the treatment of industrial dye effluents. *Biotechnology. Adv.* 22, 161-187.
- Wrobel D, Boguta A, Ion R,M. (2001). Mixtures of synthetic organic dyes in a photoelectronic cell. *J Photochem Photobiol, A Chem.* 13, 7 – 22.
- Wu, Z., Tsumura, Y., Blomquist G., Wang X, R.(2003). 18S rRNA gene variation among common airborne fungi, and development of specific oligonucleotide probes for the detection of fungal isolates. *Appl. Environ. Microbiol.* 9, 5389-5397/
- Yang, X. Q., Zhao, X. X., Liu, C. Y., Zheng, Y., & Qian, S. J. (2009). Decolorization of azo, triphenylmethane and anthraquinone dyes by a newly isolated *Trametes* sp. SQ01 and its laccase. *Process Biochem.* 44, 1185-1189.
- Yuzhu Fu., Viraraghavan, T. (2001)Fungal decolorization of dye wastewaters: a review. *Biores Techno*79,251–262.
- Youssef, A. S., El-Sherif, M. F., & El-Assar, S. A. (2008). Studies on the decolorization of Malachite green by the local isolate *Acremonium kiliense*. *Biotechnology*, 7, 213-223.

Zak, J. C. and Wildman, H. G. (2004). Fungi in stressful environments. In *Measuring and Monitoring Biological Diversity: Standard Methods for Fungi*, Mueller, G., Bills, G. (Eds.). Smithsonian Institution Press. Washington, DC.

Zouari-Mechichi, H., Mechichi, T., Dhoui, A., Sayad, S., Martínez, A. T., and Martínez, M. J. (2006). Laccase purification and characterization from *Trametes trogii* isolated in Tunisia: Decolorization of textile dyes by the purified enzyme. *Enzyme Microb Tech*, 39, 141-148.

Zycha, H., R. Siepmann, and G. Linnemann. 1969. *Mucorales a description of all genera and species of this fungal group*. Lehre, J. Cramer. 355 p