OIL AND GREASE TREATMENT USING INDIGENOUS BACTERIAL BIOFILM SYSTEM

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Specially dedicated to my beloved

Family, Lecturers and Friends

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ABSTRACT

The Malaysian palm oil industry grows rapidly as the premier agriculturebased industry over the last two decades. However, the production of palm oil also resulted in the generation of large quantities of polluted wastewater commonly referred to as palm oil mill effluent (POME) as well as lignocellulosic biomass, especially from fronds, trunks, empty fruit bunch, palm kernel and mesocarp fibre. This study evaluates on the potential application of an immobilized bacterial-based system to reduce oil and grease (O&G) contamination in POME. From the 21 bacterial isolates, two isolates (i.e. isolate X7 and X10) were chosen for subsequent studies based on its high cell surface hydrophobicity, opaque halo formation and high culture turbidity. The bacterial isolates were evaluated for O&G degradation ability by varying contact time, initial POME concentration and initial pH. The isolated X10 showed the ability to completely degrade O&G from POME after 6 days of incubation at 200 rpm, 30°C and pH 7 in the batch study. In the continuous flowthrough column study, oil palm frond (OPF) - immobilized isolate X7 showed complete O&G degradation and 92.59% of chemical oxygen demand (COD) degradation from 100% (v/v) POME used after 8 days of contact time. Scanning Electron Microscope examination showed the presence of diverse morphology of microorganisms indicating the formation of mature biofilm after 200 days. One of the bacterial species isolated from the biofilm was identified as Bacillus cereus in whichmatch up to 99% of the query sequence. The finding of this study indicates the potential of using indigenous aerobic bacterial isolates as an alternative solution to degrade O&G in POME wastewaters.

ABSTRAK

Industri minyak sawit Malaysia berkembang dengan pesat sebagai industri berasaskan pertanian yang terulung sejak dua dekad yang lalu. Walau bagaimanapun, pemprosesan minyak kelapa sawit juga menyebabkan penghasilan kuantiti air sisa tercemar yang banyak, seringkali dirujuk sebagai buangan air sisa kilang minyak sawit (POME) atau dikenali sebagai biojisim lignoselulosik,khususnya pelepah, batang, buah tandan kosong, isirong sawit dan gentian mesokarp. Kajian ini memberi tumpuan terhadap penilaian potensi sistem bakteria tersekat gerak untuk mengurangkan pencemaran minyak dan gris (O&G) dalam POME. Berdasarkan 21 pencilan bakteria awal, dua pencilan bakteria (iaitu pencilan X7 dan X10) telah dipilih untuk kajian selanjutnya berdasarkan nilai yang tinggi bagi permukaan sel, formasi halo legap dan nilai kekeruhan kultur. Pencilan bakteria telah dinilai bagi keupayaan menguraikan O&G dengan mempelbagaikan masa tindakbalas, kepekatan asal POME dan pH asal. Pencilan X10 menunjukkan keupayaan untuk mendegradasi secara lengkap O&G dari POME selepas tempoh pengeraman selama 6 hari pada 200 rpm, 30°C dan pH 7 menerusi kajian kelompok. Bagi kajian menggunakan aliran turus yang berterusan, pencilan X7 yang tersekat gerak pada pelepah kelapa sawit (OPF) menunjukkan keupayaan untuk mendegradasi secara lengkap O&G dan degradasi keperluan oksigen kimia (COD) pada tahap 92.59% daripada 100% (v/v) POME yang digunakan selepas 8 hari masa tindakbalas. Analisis mikroskop elektron pengimbas menunjukkan kehadiran morfologi pelbagai jenis mikroorganisma yang menandakan pembentukan biofilem matang selepas 200 hari. Salah satu daripada spesis bakteria yang diasingkan daripada biofilem telah dikenalpasti sebagai Bacillus cereus dengan nilai padanan setinggi 99%. Hasil daripada kajian ini menunjukkan potensi penggunaan pencilan bakteria indigenus aerobik sebagai penyelesaian alternatif untuk menguraikan O&G dalam air sisa buangan POME.

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LIST OF SYMBOLS

°C - Degree Celsius

% - Percentage

μg - Microgram

μL - Microliter

g - gram

d - Day

hr - Hour

kpa - kilopascal

kg - kilogram

K - Kelvin

rpm - revolutions per minute

ppm - parts per million

m - meter

mg - milligram

mM - millimolar

M - Molar

min - Minute

mL - Milliliter

mp - melting point

v/v - volume/volume

LIST OF ABBREVIATIONS

ADMI - American Dye Manufacturing Institute

HRT - Hydraulic retention time

NA - Nutrient AgarNB - Nutrient BrothLB - Luria Bertani

FT-IR - Fourier Transform Infrared

OPF - Oil Palm Frond

SEM - Scanning Electron Microscopy

COD - Chemical Oxygen Demand
BOD - Biological Oxygen Demand

TSS - Total Suspended Solid

VSS - Volatile Suspended Solid

TOC - Total Organic Carbon

POME - Palm Oil Mill Effluent

O&G - Oil and Grease

OD - Optical Density

BATH - Bacterial Adherence to Hydrocarbon

E₂₄ - Emulsification Index

CFU - Colony Forming Unit

EPS - Exopolymeric substances

PCR - Polymerase Chain Reaction

DNA - Deoxyribonucleic acid

rRNA - Ribosomal Ribonucleic Acid

FASTA - DNA and protein sequence alignment

BLAST - Basic Local Alignment Search Tool

NCBI - National Centre for Biotechnology Information

IUPAC - International Union of Pure and Applied Chemistry

BJH - Barrett-Joyner-Halenda

SDS - Sodium Dodecyl Sulfate

PBS Phosphate Buffer Saline

NaCl - Sodium Chloride

MeOH - methanol

HCl - Hydrochloric acid

H₂SO₄ - Sulphuric acid

NaOH - Sodium hydroxide

CaCl - Calcium chloride

 CH_2Cl_2 - Dichloromethane

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CHAPTER 1

INTRODUCTION

1.1 Background of the Study

The Malaysian palm oil industry has grown rapidly over the past few years to become the world's second largest producer of palm oil, accounting for 46% of the world exports and 37% of world palm oil production in 2011 (according to the Malaysian Palm Oil Council, MPOC), (2011). The year 2011 was both remarkable and significant for the Malaysian palm oil industry which recorded its highest ever earnings of RM 80.4 billion, surpassing the RM 65.2 billion mark recorded in 2008 by 23.3% (MPOC, 2011; Wu *et al.*, 2010). The palm oil industry also provides a source of livelihood to rural families in government land schemes and private small holders, as well as employment opportunities to agricultural workers in estates (Wu *et al.*, 2010).

Malaysia has implemented a wet process for palm oil milling. Even though the process is effective, it is not suitable to be applied in large scale productions (Prasertsan and Prasertsan, 1996). This process involves sterilizing, stripping and threshing of bunches to free the palm fruit, in which huge amounts of water and steam is required for washing and sterilizing. Other operational processes in the palm oil mills produce waste load in the form of gaseous emissions from boilers and incinerators, solid wastes materials and by-products such as empty fruit bunch, potash ash, palm kernel, fiber, shells and liquid waste. During the process, more than 50% of the water will be discharged to the environment as palm oil mill effluent

(POME), while the rest are lost either as steam in the boilers blow-down, washing water or leakage (Ahmad *et al.*, 2003a).

POME has always been regarded as the most significant pollutant from palm oil milling activities (Poh and Chong, 2009). POME is a viscous, brownish liquid containing about 95–96% water, 0.6–0.7% oil and 4–5% total solids that includes 2–4% total suspended solids (TSS) respectively (Salihu *et al.*, 2011). It is acidic (pH 4–5), hot (80–90°C), non-toxic (as no chemicals added during oil extraction), high organic content (COD 15,000–100,000 mg/L, BOD 10,250–43,750 mg/L) and contains appreciable amounts of plant nutrients such as phosphorus, potassium, calcium, magnesium, zinc and sodium (Najafpour *et al.*, 2005). It was estimated that for every tonne of crude palm oil produced, about 5–7.5 tonnes of water is required with more than 50% (about 2.5–3.5 tonne) ends up as POME (Norli *et al.*, 2006). It was then estimated that more than 40 million tonnes of POME could be generated from 372 mills throughout Malaysia (Yacob *et al.*, 2006). Most of the POME produced originates from different sources such as sterilizer condensate, hydrocyclone waste and clarifier sludge in the form of liquid effluent from the large amounts of steam or hot water used (Norli *et al.*, 2006).

In Malaysia, over 85% of the mills use the conventional anaerobic ponding system as the main treatment system for POME. The system usually based on the suspended growth of activated sludge with relatively low operation cost or something called as facultative pond. Other processes employed include aerobic and anaerobic digestions, physicochemical treatment and membrane filtration (Wu *et al.*, 2010). In Malaysia, all discharged effluents must adhere to the regulations as outlined in the Environmental Quality Act, 1974. Anaerobic process or biological treatment has considerable advantages over other processes such as less energy demands, minimum sludge formation and production of methane due to efficient break down of organic substances by anaerobic bacteria (Rincón et al., 2006).

1.2 Problem Statement

Oil and grease (O&G) is one of the major organic-rich compounds that directly contribute to severe environmental pollution as well as affecting biological wastewater treatment processes where the high-organic load may lead to system shut-down due various number of reason such as toxicity to the microbial population in the activated sludge, incomplete degradation process and huge volumes of sludge being generated. O&G would form layers on water surface that may prevent the dissolution of oxygen and reduce the oxygen transfer rates into the aqueous environment (Prasad and Manjunath, 2011). Being the preferred method for the treatment of POME in Malaysian palm oil mills, the ponding system has received a lot of attention from researchers notably in Malaysia. However, amongst notable disadvantages in the ponding system includes the requirement for large land area (to accommodate a series of ponds and volume of wastewater) and long hydraulic retention time (HRT), up to 20 days for each pond. Therefore, there is need to have alternative treatment system that requires small land area and able to shorten the HRT. This approach is expected to substantially reduce the operating cost and also minimalize the impact to the environment.

1.3 Objectives of Study

- i. To isolate, identify and characterize O&G degrading bacterial species from POME in batch and immobilized.
- ii. To evaluate the feasibility of using the indigenous O&G-degrading bacteria in an immobilized continuous flow O&G removal system.
- iii. To optimize indigenous O&G-bacteria degradation ability in batch and column study.

1.4 Scope of Study

This study focuses on the degradation of O&G in the mixing anaerobic pond from POME using biological treatment system and oil palm frond biomass as a support material. POME was characterized for biological and chemical parameters where the isolated indigenous O&G-degrading bacteria were determined for its O&G-degradation properties in POME. Oil palm frond was used as support material for the immobilization of bacteria for biofilm development. Factors that may affect optimum O&G degrading performance of the bacteria such as pH of the system, HRT, flowrate, influent and effluent COD concentrations and O&G contents were evaluated.

1.5 Significance of the Study

High concentration of O&G could cause severe environmental pollution and conflicts in water treatment process. Therefore, the estimated outcome of this study is initial indication on the feasibility of using an indigenous O&G-bacteria as alternative biological treatment system for the treatment of POME prior to entering the anaerobic pond (primary treatment system). It is expected that the retention time for the anaerobic system will be reduced significantly, hence reducing the overall treatment period for POME. Therefore, this alternative biological treatment may be suggested or implemented in the palm oil mills for its O&G degradation capability.

REFERENCES

- Abdullah, N., Ujang, Z. and Yahya, A. (2011). Aerobic Granular Sludge Formation for High Strength Agro-Based Wastewater Treatment. *Bioresource Technology*. 102, 6778-6781.
- Abdullah, A.Z., Salamatinia, B. and Kamaruddin, A.H. (2009). Application of Response Surface Methodology for The Optimization of NaOH Treatment on Oil Palm Frond Towards Improvement in The Sorption of Heavy Metals. *Desalination*. 244, 227-238.
- Abouseoud, M., Maachi, R. and Amrane, A. (2007). Biosurfactant Production from Olive Oil by *Pseudomonas fluorescens*. Communicating Current Research and Educational Topics and Trends in Applied Microbiology A. Méndez-Vilas (Ed.).
- Affandi, I.E., Suratman, N.H., Abdullah, S., Ahmad, W.A., and Zakaria, Z.A. (2014). Degradation of Oil and Grease from High- Strength Industrial Effluents using Locally Isolated Aerobic Biosurfactant- Producing Bacteria. *International Biodeterioration & Biodegradation*. doi.org/10.1016/j.ibiod.2014.04.009.
- Agensi Inovasi Malaysia (2011). National Biomass Strategy 2020: New wealth creation for Malaysia's palm oil industry.
- Ahmad, A. L., Ismail, S. and Bhatia, S. (2003a). Water Recycling from Palm Oil Mill Effluent (POME) using Membrane Technology. *Desalination*. 157, 87-95.
- Ahmad, A. L., Sethupathi, S. and Hameed, B.H. (2003b). Chitosan: A Biopolymer as Residue Oil Sorbent in Palm Oil Mill Effluent (POME). *International Conference on Chemical and Bioprocess Engineering*, Kota Kinabalu. 44-49.
- Ahmad, A. L., Sumathi, S. and Hameed, B. H. (2004). Chitosan: A Natural Biopolymer for The Adsorption of Residue Oil from Oily Wastewater. *Adsorption Science & Technology*. 22, 75-88.
- Ahmad, A. L., Ismail, S. and Bhatia, S. (2005a). Membrane Treatment for Palm Oil Mill Effluent: Effect of Transmembrane Pressure and Crossflow Velocity. *Desalination*. 179, 245-255.

- Ahmad, A. L., Bhatia, S., Ibrahim, N. and Sumathi, S. (2005b). Adsorption of Residue Oil from Palm Oil Mill Effluent using Rubber Powder. *Brazilian Journal of Chemical Engineering*. 22, 371-379.
- Ahmad, A. L., Sumathi, S. and Hameed, B. H. (2005c). Adsorption of Residual Oil from Palm Oil Mill Effluent using Powder and Flake Chitosan: Equilibrium and Kinetic Studies. *Water Research*. 39, 2483-2494.
- Ahmad, A.L., Sumathi, S., Hameed, B.H., (2005d). Residual oil and suspended solid removal using natural adsorbents chitosan, bentonite and activated carbon: a comparative study. *Journal of Chemical Engineering*. 108, 179-185.
- Ahmad, A. L., Sumathi, S. And Hameed, B. H. (2006). Coagulation of Residue Oil and Suspended Solid in Palm Oil Mill Effluent by Chitosan, Alum and PAC. *Chemical Engineering Journal*. 118, 99-105.
- APHA (2005). Standard Methods for the Examination of Water & Wastewater. (21st Ed.). Washington, DC.: American Public Health Association.
- Babu, I. S. and Rao, G. H (2007). Optimization of Process Parameters for The Production of Lipase in Submerged Fermentation By *Yarrowia lipolytica*NCIM 3589. *Research Journal of Microbiology*. 2, 88-93.
- Baharuddin, A. S., Razak, M. N. A., Hock, L. S., Ahmad, M. N., Aziz, S. A,Rahman, N. A. A. (2010). Isolation and Characterization of Thermophilic Cellulase-Producing Bacteria from Empty Fruit Bunches-Palm Oil Mill Effluent Compost. *American Journal of Applied Sciences*. 7(1), 56-62.
- Bahiyah, N. (2007). Degradation of Palm Oil Mill Secondary Effluent (POMSE) Using Biostructure. Master of Engineering (Civil Environmental Management), Faculty of Civil Engineering, Universiti Teknologi Malaysia.
- Basiron, Y. and Chan, K. W. (2004). The Oil Palm and Its Sustainability. *Journal of Oil Palm Research*. 16(1), 1-10.
- Bussamara, R., Fuentefria, A. M., de Oliveira, E., Broetto, L., Simcikova, M., Valente, P., Schrank, A. and Vainstein, M. H. (2010). Isolation of a Lipase Secreting Yeast for Enzyme Production in a Pilot-Plant Scale Batch Fermentation. *Bioresource Technology*. 101, 268-275.
- Bhumibhamon, O., Koprasertsak, A. and Funthong, S. (2002). Biotreatment of High Fat and Oil Wastewater by Lipase Producing Microorganisms. *Kasetsart Journal* (*Nature Science*.). 35, 261-267.

- Breed, R. S. and Murray, E. G. (1984). Bergey's Manual of Determinative Bacteriology [M], The Williams and Wilkins Company, Loudan.
- Divakaran, R. and Pillai, V. N. S. (2001). Flocculation of Kaolinite Suspensions in Water by Chitosan. *Water Research*. 35, 3904-3908.
- Campere, A.K., Hayes, J.T., Sturman, P.J., Jones, W.L. & Cunninghan, A.B. (1993) Effect of Motility and Absorption Rate Coefficient on Transport of Bacteria through Saturated Porous Media. *Applied and Environmental Microbiology*. 59, 3455–3462.
- Cao, S. G., Yong, H., Ma, L. and Guo, S. C. (1996). Enzymatic Properties by The Immobilization Method. *Applied Biochemistry and Biotechnology*. 59, 7–14.
- Chan, Y.J., Chong, M.F. and Law, C.L. (2012). An integrated Anaerobic-Aerobic Bioreactor (IAAB) for The Treatment of Palm Oil Mill Effluent (POME): Start-up and Steady State Performance. *Process Biochemistry*. 47, 485-495.
- Chen, S. Y., Wei, Y. H. and Chang, J. S. (2007). Repeated pH-stat fed-batch fermentation for rhamnolipid production with indigenous *Pseudomonas* aeruginosa S2. Applied Microbiology and Biotechnology. 76 (1), 67-74.
- Comte, S., Guibaud, G. and Baudu, M. (2006). Biosorption properties of extracellular polymeric substances (EPS) resulting from activated sludge according to their type: soluble or bound. *Process Biochemistry*. 41, 815–823.
- Cooper, D. G. and Goldenberg, B. G. (1984). Surface-Active Agents from Two Bacillus Species. Applied and Environmental Microbiology. 53(2), 224-229.
- Davranov, K. (1994) Microbial lipases in biotechnology, (Review) *Applied Biochemistry and Microbiology*. 30, 427 432.
- Donlan, R. M. and Costerton, J. W. (2002). Biofilms: Survival Mechanisms of Clinically Relevant Microorganisms. Clinical Microbiology Reviews. 167-193.
- El-Masry, M. H., El-Bestawy, E. and El-Adl, N. I. (2004). Bioremediation of vegetable oil and grease from polluted wastewater using a sand biofilm system. *World Journal of Microbiology and Biotechnology*. 20, 551-557.
- Eboigbodin, K. E. (2008). *Biophysical Investigation of Bacterial Aggregation*. Doctor of Philosophy, University of Sheffield, UK.
- Farrell, M. J. and Finkel, S. E. (2003). The Growth Advantage in Stationary-Phase Phenotype Conferred by *rpoS* Mutations Is Dependent on the pH and Nutrient Environment. *Journal of Bacteriology*. 7004- 7052.

- Flemming, H. (1995) Sorption sites in biofilms. *Water Science and Technology*. 32(8), 27—33.
- Fritsche, W. and Hofrichter, M. (2008). Aerobic Degradation by Microorganisms. In: *Biotechnology: Environmental Processes I.* Germany: Wiley-VCH Verlag GmbH.
- Habib, M. A. B., Yusoff, F. M., Phang, S.M., Ang, K.J. and Mohamed, S. (1997). Nutritional values of chironomid large grown in palm oil mill effluent and algal culture. *Aquaculture*. 158, 95-105.
- Hassan, M. A., Yacob, S. and Shirai Y. (2004). Treatment of Palm Oil Mill Wastewaters. In Wang, L.K., Hung, Y., Lo, H.H. and Yapijakis, C. (Ed.). *Handbook of Industrial and Hazardous Wastes Treatment* (pp. 719 736). New York: Marcel Dekker, Inc.
- Hasan, F., Shah, A. A. and Hameed, A. (2006). Industrial applications of microbial lipases. *Enzyme and Microbial Technology*. 39, 235-251.
- Houdt, R. V. and Michiels, C. W. (2005). Role of bacterial cell surface structures in *Escherichia coli* biofilm formation. Research in Microbiology. 156, 626-633.
- Iwara, A.I., Ewa, E. E., Ogundele, F. O., Adeyemi, J. A. and Out, C. A. (2011).
 Ameliorating Effects of Palm Oil Mill Effluent on the Physical and Chemical properties of Soil in Ugep, Cross River State, South-Southern Nigeria.
 International Journal of Applied Science and Technology. 1, 5.
- Kosaric, N. (1992). Biosurfactants in industry. *Pure and Applied Chemistry*. 64(11), 1731-1737.
- Lai, L. W. and Idris, A. (2013). Disruption of Oil Palm Trunks and Fronds by Microwave-Alkali Pretreatment. *BioResources*. 8, 2.
- Lam, M. K., Lee, K. T., (2011). Renewable and sustainable bioenergies production from palm oil mill effluent (POME): win–win strategies toward better environmental protection. *Biotechnology Advances*. 29, 124–141.
- Lam, M. K., Tan, K. T., Lee, K. T., Mohamed, A. R. (2009). Malaysian palm oil: Surviving the food versus fuel dispute for a sustainable future. *Renewable Sustainable Energy Reviews*.13, 1456–64.
- Lan, W. U., Gang, G. E. and Jinbao, W. A. N. (2009). Biodegradation of oil wastewater by free and immobilized *Yarrowia lipolytica* W29. *Journal of Environmental* Sciences. 21, 237-242.

- Li. Ji., Luo. M., Zhao. C. J., Li. C. Y., Wang. W., Zu. Y. G. and Fu. Y. J. (2013). Oil removal from water with yellow horn shell residues treated by ionic liquid. *Bioresource Technology*. 128, 673-678.
- Lin, T. C., Young, C. C., Ho, M. J., Yeh, M. S., Chou, C. L., Wei, Y. H. and Chang, J. S. (2005). Characterization of floating activity of indigenous diesel-assimilating bacterial isolates. *Journal of Bioscience and Bioengineering*. 99, 4656-472.
- Lindsay, D., Brozel, V. S. and Holy, A. V. (2002). Differential efficacy of a chlorine dioxide-containing sanitizer against single species and binary biofilms of a dairy-associated Bacillus cereus and a *Pseudomonas fluorescens* isolate. *Journal of Applied Microbiology*. 92, 352-361.
- Lopez, E. R., Hernandez, J. C., Dendooven, L., Rangel, P. and Thalasso, P. (2003).Characterization of five agricultural by-products as potential biofilters carriers.Bioresource Technology. 88, 259-263.
- Malaysia (2009). Environmental Quality Act 1974-Environmental Quality (Industrial Effluent) Regulation 2009. P.U. (A) 434.
- Malaysian Palm Oil Council (2011). Annual Report- The Natural Colour of Health. Malaysia: Malaysian Palm Oil Council.
- Matsumiya, Y., Wakita, D., Kimura, A., Sanpa, S. and Kubo, M. (2007). Isolation and Characterization of a Lipid-Degrading Bacterium and Its Application to Lipid-Containing Wastewater Treatment. *Journal of Bioscience and Bioengineering*. 103(4), 325-330.
- MPOB., (2008). Malaysian Palm Oil Board. A summary on the performance of the Malaysian oil palm industry. http:// econ.mpob.gov.my/economy/Performance-130109.htm.
- Metcalf, R. G., James, M. J., Mantzioris, E., & Cleland, L. G. (2003). A practical approach to increasing intakes of n-3 polyunsaturated fatty acids: Use of novel foods enriched with n-3 fats. *European Journal of Clinical Nutrition*. 57(12), 1605-1612.
- Najafpour, G. D., Zinatizadeh, A. A. L., Mohamed, A. R., Isa, M H. and Nasrollahzadeh, H. (2005). High-Rate Anaerobic Digestion of Palm Oil Mill Effluent in An Upflow Anaerobic Sludge-Fixed Film Bioreactor. *Process Biochemistry*, 41, 370-379.

- Noudeh, G. D., Noodeh, A. D., Moshafi, M. H., Behravan, E., Afzadi, M. A. and Sodagar M. (2010). Investigation of cellular hydrophobicity and surface activity effects of biosynthesed biosurfactant from broth media of PTCC1561. *African Journal of Microbiology Research*. 4(17), 1814-1822.
- Nwuche, C. O. and Ogbonna, J. C. (2011). Isolation of Lipase Producing Fungi from Palm Oil Mill Effluent (POME) Dump Sites at Nsukka. *Brazillian Archives of Biology and Technology*. 54(1), 113-116.
- Norli, I., Ho, C. M. and Ling, Y. L. (2006). Anaerobic Digestion of Mixed ChemicalPulping and Palm Oil Mill Effluent in Suspended Growth Anaerobic Digester. *International Conference on Green and Sustainable Innovation*. Chiang Mai, Thailand.
- Norsuhada binti Abdul Karim (2012). Chromate Reductase Activitiy in Whole Cells and crude Cell Free Extract of Acinetobacter haemolyticus. Master of Science, Universiti Teknologi Malaysia, Skudai.
- Nur Haizarisha binti Suratman (2014). *Electrical and Electronic Waste Characterization and Biodegradation of Oil and Grease by Isolated Indigenous Bacteria*. Master of Science, Universiti Teknologi Malaysia, Skudai.
- Nur Zulaikha binti Yusof (2010). *Isolation and Application of Violet Pigment Extracted from Chromobacterium violaceum*. Master of Science, Universiti Teknologi Malaysia, Skudai.
- Oswal, N., Sarma, P. M., Zinjarde, S. S. and Pant, A. (2002). Palm oil mill effluent treatment by a tropical marine yeast. *Bioresource Technology*. 85, 35-37.
- Pantzaris, T. P. and Ahmad, M. J. (2001). Properties and Utilization of Palm Kernel Oil. Palm Oil Developments. 35: 11-23.
- Larkin, P. (2011). Infrared and Raman Spectroscopy, Principles and Spectral Interpretation. USA: Elsevier.
- Prasad, M. P. and Manjunath, K. (2011). Comparative study on biodegradation of lipid-rich wastewater using lipase producing bacterial species. *Indian Journal of Biotechnology*. 10, 121-124.
- Prasertsan, S. and Prasertsan, P. (1996). Biomass residues from palm oil mills in Thailand: An Overview on Quality and Potential Usage. *Biomass Bioenergy*. 11, 387-395.

- Plou, F. J., Ferrer, M., Nuero, O. M., Calvo, M. V., Alcalde, M., Reyes, F. and Ballesteros, A. (1998). Analysis of Tween 80 as an esterase/lipase substrate for lipolytic activity assay. *Biotechnology Techniques*. 12(3), 183-186.
- Poh, P. E., Chong, M. F. (2009). Development of anaerobic digestion methods for palm oil mill effluent (POME) treatment. *Bioresource Technology*. 100, 1-9.
- Poku, K. (2002). Small Scale Palm Oil processing in Africa. Agriculture and Consumer Protection. FAO. *Agricultural Services Bull*.148, 62.
- Rajeb, A. B., Kallali, H., Aissa, N. B., Bouzaiene, O., Jellali, S., Jedidi, N. and Hassen, A. (2009). Soil microbial growth and biofilm expansion assessment under wastewater infiltration percolation treatment process: column experiments. *Desalination*. 246, 514-525.
- Rincón, B., Raposo, F. Domínguez, J. R., Millán, F., Jiménez, A. M., Martín, A. and Borja, R. (2006). Kinetic models of an anaerobic bioreactor for restoring wastewater generated by industrial chickpea protein production. *International Biodeterioration & Biodegradation*, 57: 114-120.
- Rosenberg, E., and Ron, E. Z. (1999). High and low-molecular-mass microbial surfactants. *Applied Microbiology and Biotechnology*. 52(2), 154-162.
- Rosenberg, E. (2006) Hydrocarbon-Oxidizing Bacteria. *Prokaryotes*. 2,564-577.
- Rupani, P. F., Singh, R. P., Ibrahim, M. H. and Esa, N. (2010). Review of Current Palm Oil Mill Effluent (POME) Treatment Methods: Vermicompositing as a Sustainable Practice. *World Applied Sciences Journal*. 11(1), 70-81.
- Saeed, H. M., Zaghloul, T. I., Khalil, A. I. and Abdelbaeth, M. T. (2005). Purification and characterization of two extracellular lipases from *Pseudomonas aeruginosa* Ps–x. *Polish Journal of Microbiology*. 54, 233 -240.
- Saka, C. (2012). BET, TG-DTG, FT-IR, iodine number analysis and preparation of activated carbon from acorn shell by chemical activation ZnCl₂. *Journal of Analytical and Applied Pyrolysis*. 95, 21-24.
- Salamatinia, B., Kamaruddin, A.H. and Abdullah, A.Z. (2008). Modeling of the continuous copper and zinc removal by sorption onto sodium hydroxide-modified oil palm frond in a fixed-bed column. *Chemical Engineering Journal*. 145, 259-266.
- Salihu, A. and Alam, M. Z. (2012). Palm oil mill effluent: a waste or a raw material? Journal of Applied Sciences Research. 8(1), 466-473.

- Salihu, A., Alam, M. Z., Karim, M. I. A. and Salleh H. M. (2011). Suitability of using palm oil mill effluent as a medium for lipase production. *African Journal of Biotechnology*. 11(10), 2044-2052.
- Sawain, A., Taweepreda, W., Puetpaiboon, U. and Suksaroj, C. (2009). The Effect of pH on the Stability of Grease and Oil in Wastewater from Biodiesel Production Process. International Conference on Innovations in Agricultural, Food and Renewable Energy Productions of Mankind.
- Shavandi, M., Mohebali, G., Haddadi, A., Shakarami, H. and Nuhi, A. (2011). Emulsification potential of a newly isolated biosurfactant-producing bacterium, *Rhodococcus* sp. Strain TA6. *Colloids and Surfaces B: Biointerfaces*. 82, 477-482.
- Shavandi, M. A., Haddadian, Z., Ismail, M. H. S. and Abdullah, N. (2012). Continuous metal and residual oil removal from palm oil mill effluent using natural zeolite-packed column. *Journal of the Taiwan Institute of Chemical Engineers*. 43, 934-941.
- Sierra, G. (1957). Studies on bacterial esterases 1) Part III- Influence of the composition of the culture media on the production of all-esterase and lipase of pseudomonas aeruginosa.
- Sirisha, E., Rajasekar, N. and Narasu, M. L. (2010). Isolation and Optimization of Lipase Producing Bacteria from Oil Contaminated Soils. *Advances in Biological Research*. 4(5), 249-252.
- Sidik, S.M., Jalil, A.A., Triwahyono, S., Adam, S.H., Satar, M.A.H. and Hameed, B.H. (2012). Modified oil palm leaves adsorbent with enhanced hydrophobicity for crude oil removal. *Chemical Engineering Journal*. 203, 9-18.
- Sugimori, D., Nakamura, M., Tusbochi, N., Tusbochi, K., Obata, T., Uzura, K., and Miyazaki, K. (2002). Treatment of lipidcontaminated wastewater by a lipid-degrading microbial preparation. *Seibutsu-kogaku*, 80, 559–562.
- Sumathi, S., Chai, S. P. and Mohamed, A. R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable Sustain Energy Reviews*. 12, 2404-2421.
- Thong, S. O., Mamimin, C. and Prasertsan, P. (2011). Effect of temperature and initial pH on biohydrogen production from palm oil mill effluent: long-term evaluation and microbial community analysis. *Microbial Biotechnology*. 14(5).

- Tong, S. L. and Jaafar, A. B. (2004). Waste to energy: methane recovery from anaerobic digestion of palm oil mill effluent, ENERGY SMART.
- Tres, A., Ruiz-Samblas, C., Veer, G. V. D. and Ruth, S. M. V. (2013). Geographical provenance of palm oil by fatty acid and volatile compound fingerprinting techniques. *Food Chemistry*. 137, 142-150.
- Ginkel, C.G.V. (1989). Complete degradation of xenobiotics surfactants by consortium of aerobic micro-organisms. *Biodegradation*. 7(2), 151-164.
- Vijayaraghavan, K., Ahmad, D.and Aziz, M. E. A. (2007). Aerobic Treatment of Palm Oil Mill Effluent. *Journal of Environmental Management*. 82, 24-31.
- Wanrosli, W.D., Zainuddin, Z., Law, K.N. and Asro, R. (2007). Pulp from oil palm fronds by chemical processes. *Industrial Crops and Products*. 25, 89-94.
- Watnick, P. and Kolter, R. (2000). Biofilm: city of microbes. *Journal of Bacteriology*. 182, 2675–2679.
- Wiseman, A.(1995).Introduction to principles. In: Wiseman, A (ed.,) Handbook of enzyme biotechnology 3rd edition. Padstow, Cornwall, U.K: Ellis Horwood Ltd., T.J Press Ltd pp 3 -8.
- Wong, P. H., Nik Meriam Sulaiman, Nachiappan, M. and Varadaraj, B. (2002). Pretreatment and Membrane Ultrafiltration Using Treated Palm Oil Mill Effluent (POME). *Songklanakarin Journal of Science and Technology*. 24, 891 898.
- Wu, T. Y., Mohammad, A. W., Jahim, J. M. and Anuar, N. (2010). Pollution control technologies for the treatment of palm oil mill effluent (POME) through end-of-pipe processes. *Journal of Environmental and Management*. 91, 1467-1490.
- Wan Haslinda binti Wan Ahmad (2013). Hexavalent Chromium Reduction by Acinetobacter haemolyticus using Agricultural Waste. Doctor of Philosophy, Universiti Teknologi Malaysia, Skudai.
- Yacob, S., Shirai, Y., Hassan, M. A., Wakisaka, M. and Subash, S. (2006). Start-up Operation of Semi-Commercial Closed Anaerobic Digester for Palm Oil Mill Effluent Treatment. *Process Biochemistry*. 41, 962-964.
- Yacob, S., Shirai, Y. and Hassan, M. A. (2009). Treatment of Palm Oil Wastewaters. Waste treatment in the food processing industry, CRC Press.
- Yusoff, S. (2006). Renewable energy from palm oil e innovation on effective utilization of waste. *Journal of Cleaner Production*.14, 87-93.

Zainul Akmar Zakaria (2006). Development of Bacterial –Based Remediation System for The Removal of Chromium (VI) from Electroplating Industrial Effluent.

Doctor of Philosophy, Universiti Teknologi Malaysia, Skudai.