

DECOLORIZATION AND BIODEGRADATION OF MORDANT ORANGE-1 BY  
NEWLY ISOLATED *TRICHODERMA HARZIANUM* RY 36 AND  
*ACREMONIUM SPINOSUM* RY 42

RUBIYATNO

UNIVERSITI TEKNOLOGI MALAYSIA

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RUBIYATNO

A thesis submitted in fulfillment of the  
requirements for the award of the degree of  
Master of Engineering (Environment)

Faculty of Civil Engineering  
Universiti Teknologi Malaysia

MAY 2014

*To my beloved mother, father and sister*

*Thanks for all your support, love and care...*

## ACKNOWLEDGEMENTS

First and foremost to ALLAH S.W.T for the blessings and grace, I manage to complete the thesis entitled “Decolorization and Biodegradation of Mordant Orange-1 by Newly Isolated *Trichoderma harzianum* RY 36 and *Acremonium spinosum* RY 42”.

I would like to especially thanks my supervisor, Dr. Tony Hadibarata M.Sc., for his numerous supports, encouragement, valuable suggestions and great concern to my work. A sincere thanks and appreciation also goes to Professors, Staff Members and my colleagues in the Faculty of Civil Engineering, Institute of Environmental and Water Resources Management (IPASA), UTM.

A special thanks to My Lab teammates, Mr. Meor Mohd Fikri, Mr. Musa Mutah, Mr. Ameer Badr Khudhair, Ms. Liyana Amalina Adnan, Mr. Teh Zee Chuang, Ms. Norul Hudai, Ms, Nur raisha, Ms. Noor Atikah, Ms. Mimi, Ms. Shakila, Ms. Nurasikin Mr. Mohd Hairul, Mr. Iezaat Emer Post-doctoral fellow Dr. P. Sathiskumar M.Sc., and Dr. Risky Ayu Kristanty M.Sc., for their kind willingness to share precious knowledge, information, and support for accomplish this thesis.

Finally, I wish to express my acknowledgment to the government of East Kalimantan (Indonesia) for their support and scholarship and also to Universiti Teknologi Malaysia (Malaysia) and University of Yamanashi (Japan) for providing sufficient and adequate materials, equipments and good laboratory environment in completing this research.

## ABSTRACT

The synthetic dyes are dangerous for human being and aquatic life when it pollute of water resources. Textile processing manufacture is the largest sector and initiator of fluid sewage in the shape of pollutants containing synthetic dyes. Several wastewater treatment technologies are used to treat of these pollutants including conventional and advances treatment such as Physico-chemical, electrochemical, membrane separation, and reverse osmosis. Nevertheless, all these technologies are high cost operating, need of huge space, limited flexibility and generate by-products. The microbial field which is fungal based bioremediation gives promising treatment for decolorize and degrade the synthetic dye in wastewater from textile industry. It was found to be an environmental friendly, low-cost operation and effective compared to conventional and advances treatments. In this present study, the fungal strains from soil and decayed wood isolated from Universiti Teknologi Malaysia (UTM) Campus and some region in Johor Bahru forest were screened and selected for its ability to decolorize the azo dye, Mordant Orange-1 (MO-1). Two isolates RY 36 and RY 42 showed its ability for decolorization of MO-1 dye, among fifty fungal strains collected. The degradation experiments were conducted in both of the solid and liquid medium amended with 50 ppm of MO-1 dye. The efficient degraders, RY 36 and RY 42 were identified using 18S rRNA sequence analysis and morphology characterization. From the results obtained, these fungi belong to the group of *Trichoderma harzianum* RY 36 and *Acremonium spinosum* RY 42, respectively. Further, the effect of various environmental factors parameters such as carbon and nitrogen sources, surfactant (Tween 80), aromatic compounds and pH on the dye decolorization by *Trichoderma harzianum* RY 36 and *Acremonium spinosum* RY 42 in the liquid medium was assessed. *Trichoderma harzianum* RY 36 showed efficient decolorization with addition of glucose (84.16%), ammonium nitrate (79.41%), tween 80 0.1 mL (27.68%), salicylic acid (84.73%) pH 3 (89.42%) and maximum biomass production of 6840 mg/L was achieved in the presence of yeast extract. Meanwhile, *Acremonium spinosum* RY 42 showed efficient decolorization of MO-1 with addition of glucose (86.6%), ammonium nitrate (70.21%), Tween 80 0.1 mL (12.77%), salicylic acid (84.68%) pH 3 (89.6%) and maximum biomass production of 7850 mg/L was achieved in the presence of Tween 80 (1.5 mL). Further, the degradation products of MO-1 by both of the isolates were identified using Thin Layer Chromatography (TLC) and Gas Chromatography Mass Spectrophotometer (GC-MS). The analytical results showed that maleic acid and Isophthalic acid were formed during the degradation of MO-1 by *Trichoderma harzianum* RY 36. In the case of *Acremonium spinosum* RY 42, salicylic acid and benzoic acid were identified as metabolic products during the degradation of MO-1 dye.

## ABSTRAK

Pewarna sintetik adalah berbahaya kepada manusia dan kehidupan akuatik apabila ia mencemari sumber air. Proses pembuatan tekstil merupakan sektor yang besar dan penyumbang utama air sisa kumbahan yang mengandungi pewarna sintetik. Beberapa teknologi rawatan air sisa yang digunakan untuk merawat pencemar tersebut termasuk kaedah konvensional dan rawatan termaju seperti fiziko-kimia, elektrokimia, pemisahan membran dan osmosis berbalik. Namun, semua teknologi tersebut memerlukan kos operasi yang tinggi, kawasan yang luas, tidak fleksibel dan mewujudkan hasil sampingan. Bidang microbial yang menggunakan kulat sebagai asas permuliharaan-bio memberikan jaminan rawatan untuk penyingkiran dan penguraian pewarna sintetik dari air sisa industri tekstil. Ia juga mesra alam sekitar, kos operasi yang rendah dan berkesan berbanding dengan kaedah konvensional dan rawatan termaju. Dalam kajian ini, *strain* kulat yang telah di dapati dari tanah dan kayu lapuk di kampus Universiti Teknologi Malaysia (UTM) dan sebahagiannya dari hutan di Johor Bahru, telah di saring dan di pilih bagi kebolehan untuk menyingkirkan pewarna Azo, *Mordant Orange-1* (MO-1). Dua pengasing seperti RY 36 dan RY 42 menunjukkan keberkesanan dalam penyingkiran pewarna MO-1, dikalangan lima puluh strain kulat yang dikumpul. Kajian penguraian telah dijalankan dalam dua bentuk keadaan iaitu pepejal dan cecair yang telah diubah dengan menggunakan 50 ppm pewarna MO-1. Keberkesanan pengurai dan jenis kulat RY 36 dan RY 42 telah dikenalpasti berdasarkan analisis turutan 18S rRNA dan ciri-ciri bentuk permukaan dimana masing-masing adalah dari jenis *Trichoderma harzianum* RY 36 dan *Acremonium spinosum* RY 42. Seterusnya, kesan-kesan kepelbagaian parameter faktor persekitaran seperti karbon, sumber nitrogen, bahan permukaan (Tween 80), sebatian aromatic dan pH terhadap penyingkiran pewarna oleh *Trichoderma harzianum* RY 36 dan *Acremonium spinosum* RY 42 pada keadaan cecair telah diuji. *Trichoderma harzianum* RY 36 menunjukkan keberkesanan penyingkiran warna dengan penambahan glukosa (84.16%), ammonium nitrate (79.41%), Tween 80 0.1 mL (27.68%), asid salicylic (84.73%), pH 3 (89.42%) dan penghasilan biomas yang maksimum sebanyak 6840 mg/L telah diperolehi dengan kehadiran ekstrak yis. Manakala, *Acremonium spinosum* RY 42 menunjukkan keberkesanan penguraian MO-1 dengan penambahan glukosa (86.6%), ammonium nitrate (70.21%), Tween 80 0.1 mL (12.77%), asid salicylic (84.68%) pH 3 (89.6%) dan penghasilan biomas yang maksimum sebanyak 7850 mg/L telah diperolehi dengan kehadiran Tween 80 (1.5 mL). Seterusnya, penguraian produk dari MO-1 oleh kedua-dua pengasing telah ditentukan menggunakan *Thin Layer Chromatography* (TLC) dan *Gas Chromatography Mass Spectrophotometer* (GC-MS). Hasil kajian analitik menunjukkan asid maleic dan asid isophthalic telah terbentuk semasa penguraian MO-1 oleh *Trichoderma harzianum* RY 36. Manakala, bagi kajian *Acremonium spinosum* RY 42, asid salicylic dan asid benzoic dikenalpasti sebagai produk metabolik semasa penguraian pewarna MO-1.

## TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xii
	LIST OF FIGURES	xiv
	LIST OF SYMBOLS	xx
	LIST OF APPENDICES	xxiii
<b>1</b>	<b>INTRODUCTION</b>	
	1.1 Background of Study	1
	1.2 Problem of Statements	3
	1.3 Objective of Study	4
	1.4 Scope of Study	4
	1.5 Significant of Study	4
<b>2</b>	<b>LITERATURE REVIEW</b>	
	2.1 Introduction	6
	2.2 Synthetic Dyes	7
	2.2.1 Classification of Dyes	7
	2.2.2 Technique of Removal Textile Effluent	8
	2.3 Azo Dyes	11
	2.3.1 Toxicity Consideration of Azo Dye	12
	2.3.2 Mechanism of Azo Dye Biodegradation	13

	2.3.2.1	Redox Mediator	15
	2.3.3	Factor Affecting Azo Dye on Decolorization	16
	2.3.4	Mordant Orange-1 Type of Azo Dye	17
2.4		Fungi	17
	2.4.1	Enzyme	18
		2.4.1.1 Enzyme Nomenclature	19
		2.4.1.2 Enzyme Classification	19
		2.4.1.3 Enzyme of Fungi	20
	2.4.2	Fungi Biodegradation	22
	2.4.3	Fungi Productions Metabolites	22
2.5		18S rRNA Identification of Fungi	23
	2.5.1	Phylogeny	25
2.6		Analytical technique	27
	2.6.1	UV-Visible Spectrophotometer	28
	2.6.2	Chromatography	29
		2.6.2.1 Thin Layer Chromatography (TLC)	29
		2.6.2.2 Gas Chromatography-Mass Spectrometry (GC-MS)	30

### **3 MATERIAL AND METHOD**

3.1		Introduction	31
3.2		Experiment Design	32
3.3		Materials	33
	3.2.1	Chemical and Dye	33
3.4		Experimental Method	33
	3.4.1	Screening of Fungi	34
		3.4.1.1 Sampling of Fungi	34
		3.4.1.2 Isolated of Fungi in Solid Medium	34
		3.4.1.3 Re-culture of Fungi	35
	3.4.2	Identification of Selected Fungi	35
		3.4.2.1 PDA and PDB Preparation	35



3.4.2.2	Fungi Culture	35
3.4.2.3	DNA Isolation	35
3.4.2.4	Agarose Gel Electrophoresis	36
3.4.2.5	PCR Amplification and Agarose Gel Electrophoresis	36
3.4.2.6	PCR Purification	37
3.4.2.7	DNA Sequencing	38
3.4.2.8	Assembly of Full Length of 18S rRNA	38
3.4.2.9	Homology Search and Construction of Phylogenetic Tree by Using Basic Local Alignment Search Tool for Nucleotide (BLASTN)	38
3.4.2.10	Morphology Characterization of Fungi	38
3.4.3	Analytical Methods	39
3.4.3.1	Study of Effect Environmental Factors in Liquid Medium	39
3.4.3.2	Effect of Carbon Sources	39
3.4.3.3	Effect of Nitrogen Sources	40
3.4.3.4	Effect of Surfactant (Tween 80) Concentration	40
3.4.3.5	Effect of Aromatic Compounds	40
3.4.3.6	Effect of pH	40
3.4.4	Biomass Determination	41
3.4.5	Thin Layer Chromatography (TLC)	41
3.4.5.1	Spraying Detectors	41
3.4.6	Identification of Metabolite Products	42

<b>4</b>	<b>RESULTS AND DISCUSSION</b>	
4.1	Introduction	43
4.2	Screening and Isolated of Fungi from Nature on Solid Medium	43
4.3	Identification of Selected Fungi	48
4.3.1	The Phylogenetic Trees Analysis by 18S rRNA	48
4.3.2	Morphology of Characterization <i>Trichoderma harzianum</i> RY 36	50
4.3.3	Morphology of Characterization <i>Acremonium spinosum</i> RY 42	51
4.4	Study of Effect Environmental Factors in Liquid medium	52
4.4.1	Decolorization of MO-1 Dye by <i>Trichoderma harzianum</i> RY 36 in Liquid Medium	52
4.4.1.1	Effect of Carbon Sources	52
4.4.1.2	Effect of Nitrogen Sources	55
4.4.1.3	Effect of Surfactant (Tween 80) Concentration	57
4.4.1.4	Effect of Aromatic Compounds	60
4.4.1.5	Effect of pH	62
4.4.2	Decolorization of MO-1 Dye by <i>Acremonium spinosum</i> RY 42 in Liquid Medium	65
4.4.2.1	Effect of Carbon Sources	65
4.4.2.2	Effect of Nitrogen Sources	67
4.4.2.3	Effect of Surfactant (Tween 80)	69
4.4.2.4	Effect of Aromatic Compounds	72
4.4.2.5	Effect of pH	74

4.5	Identification of Metabolites	76
4.5.1	Identification of Metabolites by <i>Trichoderma harzianum</i> RY 36	76
4.5.2	Identification of Metabolites by <i>Acremonium spinosum</i> RY 42	84
<b>5</b>	<b>CONCLUSION AND RECOMMENDATION</b>	
5.1	Conclusions	91
5.2	Recommendation for Future Study	93
	<b>REFERENCES</b>	94
	<b>APPENDICES A-F</b>	116

**LIST OF TABLES**

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Dye structures according to their chromophores	8
2.2	Advantages and disadvantages of several physico-chemical and biological decolorization processes applied to textile effluents (Robinson <i>et al.</i> , 2001)	9
2.3	Decolorization and biodegradation of azo dyes by microorganisms	14
3.1	Properties of Mordant Orange-1 (MO-1)	33
3.2	Amount of components used in PCR reaction	36
3.3	Thermal cycle profile for PCR reaction of 18S rRNA	37
4.1	Screening of fungi having ability to decolor MO-1 on solid medium	44
4.2	TLC analysis of MO-1 dye degradation samples after purifications column chromatography by <i>Trichoderma harzianum</i> RY 36	78
4.3	Mass spectra analysis of the principal metabolites detected during the degradation of MO-1 dye by <i>Trichoderma harzianum</i> RY 36	79
4.4	Mass spectra analysis of the principal metabolites detected during the degradation of MO-1 dye by <i>Acremonium spinosum</i> RY 42	84

4.5	TLC analysis of MO-1 dye degradation samples after purifications column chromatography by <i>Acremonium spinosum</i> RY 42	87
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## LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Proposed mechanism of degradation of synthetic dye (A) Degradation of Reactive Orange 16 by White rot fungi <i>Irpex lacteus</i> , the compound in brackets was not detected (Svobodová <i>et al.</i> , 2007) (B) Remazol Red degradation by <i>Galactomyces geotricum</i> (Waghmode <i>et al.</i> , 2012)	15
2.2	The mechanism of proposed reduction of azo dyes by redox mediator (Keck <i>et al.</i> , 1997)	16
2.3	Structure chemical of Mordant Orange-1 (MO-1) type of azo dye	17
2.4	Morphology of fungi (Rittman and McCarty, 2001)	18
2.5	The cycle of catalytic peroxidases (Keck <i>et al.</i> , 1997)	21
2.6	The cycle of catalytic laccase (Keck <i>et al.</i> , 1997)	21
2.7	The component of ribosome in eukaryotic cell (Lee, 2008).	24
2.8	Ribosomal DNA in fungal identification (Cralile <i>et al.</i> , 2001).	25
2.9	The three main domains in universal phylogenetic tree (Rittmann and McCarty, 2001)	25
2.10	Detailed tree for eukaryotic shows the trunk branching to kingdoms (Rittmann and McCarty, 2001)	26
2.11	Detailed tree for the archaea domain (Rittmann and	26

	McCarty, 2001)	
3.1	Flow chart of decolorization and biodegradation of Mordant Orange-1 (MO-1) by fungi from nature	32
3.2	Diameter growth and decolorization fungi on solid medium	34
4.1	Decolorization of MO-1 dye on solid medium by RY 36 (A) control 0, 7 and 13 days incubation, (B) top views 0, 7 and 13 days incubation, (C) bottom views 0, 7 and 13 days incubation	45
4.2	Decolorization of MO-1 dye on solid medium by RY 42 (A) control 0, 7 and 12 days incubation, (B) top views 0, 7 and 12 days incubation, (C) bottom views 0, 7 and 12 days incubation	46
4.3	Phylogenetic analysis of 18S rRNA sequence of fungal isolate RY 36 and RY 42 species. Distance tree was constructed using neighbor-joining method of MEGA4. The sequences which were retrieved from National Center for Biotechnology Information database showed the phylogenetic relationships of <i>Trichoderma harzianum</i> RY 36 KC139308 and <i>Acremonium spinosum</i> RY42 HE608637.1	49
4.4	Morphological character of <i>Trichoderma harzianum</i> RY 36 Conodia, Conidiophores, Hyphae and Phialides (A) , Mycelia (B), Fungal growth on PDA (C)	50
4.5	Morphological character of <i>Acremonium spinosum</i> RY 42 Conodia, Conidiophores and Phialides (A), Mycelia (B), Fungal growth on PDA (C)	51
4.6	Effect of addition carbon sources in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid	53

	medium	
4.7	Effect of addition carbon sources on biomass productions in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	53
4.8	Effect of addition nitrogen sources in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	55
4.9	Effect of addition nitrogen sources on biomass productions in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	56
4.10	Effect of addition surfactant (Tween 80) concentration in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	58
4.11	Effect of addition surfactant (Tween 80) concentration on biomass productions in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	59
4.12	Schematic diagrams of the variation of surface tension, interfacial and contaminant solubility with surfactant concentration (Rosen, 1989)	60
4.13	Effect of addition aromatic compounds in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	61
4.14	Effect of addition aromatic compounds on biomass production in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	61
4.15	Effect of pH in decolorization of MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	63
4.16	Effect of pH on biomass production in decolorization of	63



	MO-1 dye by <i>Trichoderma harzianum</i> RY 36 in liquid medium	
4.17	Effect of addition carbon sources in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	65
4.18	Effect of addition carbon sources on biomass productions in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	66
4.19	Effect of addition nitrogen sources in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	67
4.20	Effect of addition nitrogen sources on biomass productions in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	68
4.21	Effect of addition surfactant (Tween 80) concentration in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	70
4.22	Effect of addition surfactant (Tween 80) concentration on biomass productions in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	71
4.23	Effect of addition aromatic compounds in decolorization of MO-1 by <i>Acremonium spinosum</i> RY 42 in liquid medium	73
4.24	Effect of addition aromatic compounds on biomass productions in decolorization of MO-1 by <i>Acremonium spinosum</i> RY 42 in liquid medium	73
4.25	Effect of pH in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	75

4.26	Effect of pH on biomass productions in decolorization of MO-1 dye by <i>Acremonium spinosum</i> RY 42 in liquid medium	76
4.27	(A) TLC profile of extracted sample of MO-1 metabolites before separation with column chromatography, sprayed with bromocresol green and compared with six standard compounds (SA: Salicylic Acid, BA: Benzoic Acid, CA: Catechol, MA: Maleic Acid, IPA: Isophthalic Acid, and 2.4 HBA: 2.4 Hydroxybenzoic Acid)  (B) TLC profile of 5 fractions after separation with column chromatography, sprayed with bromocresol green. Fraction C (II) and E (I) showed identical $R_f$ value compared with two standard compounds maleic acid (MA) and isophthalic acid (IPA), respectively	78
4.28	Mass spectra of metabolite I identified as maleic acid (A) and metabolite II identified as isophthalic acid (B) by <i>Trichoderma harzianum</i> RY 36	80
4.29	Proposed pathways of MO-1 dye by <i>Trichoderma harzianum</i> RY 36, the intermediate compounds in the bracket were not identified in the culture extract	82
4.30	Mass spectra of metabolite I identified as benzoic acid (A) and metabolite II identified as salicylic acid (B) by <i>Acremonium spinosum</i> RY 42	85
4.31	(A) TLC profile of extracted sample of MO-1 metabolites before separation with column chromatography, sprayed with bromocresol green and compared with six standard compounds (SA: Salicylic Acid, BA: Benzoic Acid, CA: Catechol, MA: Maleic Acid, PA:	86

Phthalic Acid, and 2.4 HBA: 2.4 Hydroxybenzoic Acid)

(B) TLC profile of 5 fractions after separation with column chromatography, sprayed with bromocresol green. Fraction A (I) and B (II) showed identical  $R_f$  value compared with two standard compounds salicylic acid (SA) and benzoic acid (BA), respectively

4.32 Proposed pathways of MO-1 by *Acremonium spinosum* RY 42, the intermediate compounds in the bracket were not identified in the culture extract 89

## LIST OF SYMBOLS

%	-	Percent
—	-	Path length of radiation $b$ ; $l$ , $d$
$\lambda$ max	-	Lambda Maximum
-N=N-	-	Mono Azo Dye
-C=O	-	Carbonyl
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	-	Ammonium Sulfate
$\frac{A}{bc}$	-	Absorptivity $a$ , extinction coefficient $k$
$\frac{A}{bc}$	-	Molar absorptivity $\epsilon$ , Molar extinction coefficient
°C	-	Celsius
°C/min	-	Celsius per Minute
C <sub>6</sub> H <sub>12</sub> O <sub>6</sub>	-	Sugar
C <sub>7</sub> H <sub>6</sub> O <sub>2</sub>	-	Benzoic Acid
C <sub>6</sub> H <sub>4</sub> (OH) <sub>2</sub>	-	Catechol
C <sub>7</sub> H <sub>6</sub> O <sub>3</sub>	-	Salicylic Acid
C	-	Carbon
CH <sub>3</sub>	-	Methyl
C.I.	-	Color Index
Cl	-	Chloride
Cl <sub>2</sub>	-	Chlorine
cm	-	Centimeter
COOH	-	Carboxyl
Cu <sup>+</sup>	-	Copper Ion
Cu <sup>2+</sup>	-	Copper (II) Ion
DNA	-	Deoxyribonucleic Acid
E <sub>0</sub> '	-	Redox Potential
e <sup>-</sup>	-	Electron

Fe <sup>3+</sup>	-	Iron (III) Ion
g/L	-	Gram per Liter
g/mol	-	Gram per Mole
GC	-	Gas Chromatography
GC-MS	-	Gas Chromatography-Mass Spectrometry
H <sub>2</sub> O <sub>2</sub>	-	Hydrogen Peroxide
H <sub>2</sub> O	-	Water
H <sup>+</sup>	-	Hydrogen Ion
HCL	-	Hydrochloric Acid
kPa	-	Kilopascal
Lac	-	Laccase
LMEs	-	Lignin-Modifying Enzymes
LiP	-	Lignin Peroxides
$\log \frac{P_0}{P}$	-	Absorbance A, Optical density; extinction <i>E</i>
M <sup>+</sup>	-	Molecular Ion (peak)
M	-	Molar
MAE	-	Malt Extract Agar
Min	-	Minutes
mL	-	Milliliter
mg/L	-	Milligram Per Liter
mM	-	Millimolar
mm	-	Milliliter
MnP	-	Manganese Peroxides
Mn <sup>2+</sup>	-	Manganese (II) Ion
Mn <sup>3+</sup>	-	Manganese (III) Ion
MnO <sub>4</sub>	-	Permanganate
MO-1	-	Mordant Orange-1
m/z	-	Mass Spectrum ( <i>Mass-to-Charge Ratio</i> )
N	-	Nitrogen
NaOH	-	Sodium Hydroxide
NH <sub>2</sub>	-	Amines
NH <sub>4</sub> NO <sub>3</sub>	-	Ammonium Nitrate
NO	-	Nitro

NO <sub>2</sub>	-	Amino
Nm	-	Nanometer
O <sub>2</sub>	-	Oxygen
O <sub>3</sub>	-	Ozone
OH	-	Hydroxyl
OH <sup>•</sup>	-	Hydroxyl Radical
PAH	-	Polycyclic Aromatic Hydrocarbons
PCR	-	Polymerase Chain Reaction
PDA	-	Potato Dextrose Agar
PDB	-	Potato Dextrose Broth
pH	-	Power Hydrogen
		Energy of radian (in ergs) impinging on a 1-cm <sup>2</sup>
$P, P_0$	-	area of detector per second; radiation intensity $I, I_0$
$\frac{P_0}{P}$	-	Transmittance $T$
ppm	-	Part per Million
$R_f$	-	Retention Factor
rpm	-	Rotor per Minute
rRNA	-	Ribosomal Ribonucleic Acid
S	-	Sulfur
Si	-	Silica
SO <sub>3</sub>	-	Sulfur Trioxide
TLC	-	Thin Layer Chromatography
TMS	-	Trimethylsilylation
$t_R$	-	Retention Time
UV	-	Ultraviolet
UV-Vis	-	Ultraviolet-Visible
v/v	-	Volume per Volume
w/v	-	Weight per Volume
μL	-	Microliter
μg	-	Microgram

**LIST OF APPENDICES**

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
A	Calibration Curve of Mordant Orange-1 Standard Solution	116
B	Identification and Characterization Morphology of Selected Fungi	117
C	Screening and Isolation on Solid Medium	120
D	Study of Effect Environmental Factors	123
E	Thin Layer Chromatography (TLC) Analysis	126
F	Gas Chromatography-Mass Spectra (GC-MS) Analysis	130

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of Study

The synthetic dyes are dangerous for human being and aquatic life when they pollute water resources. Developing countries are faced with serious water pollution. The industries discharge 100 tons per day of wastewater effluent into river bodies (Allen *et al.*, 2004). The industrial processes including the pulp and paper, textile, chemical and petrochemical industries are generally utilizing dyes as colorants (Noroozi and Sorial, 2013). In excess of ten thousand variant synthetic dyes have been surpassed  $7 \times 10^5$  metric tons per annum in worldwide consumption and including manufactures (Deveci *et al.*, 2004). The presence of toxic dyes in water sources has stimulated much attention in recent decades because of their potential to cause environmental problems. Moreover, they lead to undesirable effects in the color, odor and taste of waters (Attia *et al.*, 2003).

Textile processing manufacture is the largest sector and is an initiator of fluid sewage in the shape of pollutants. Moreover, up to 1,000 tons per year are released in the form of wastewater industry based on report study (Ozmen *et al.*, 2007). The process it is predicted that 10%–20% of synthetic dyes will be lost in remaining fluids via partial washing and finishing operations (Deveci *et al.*, 2004). Besides, 1 kg of cloth produced generating approximately 40–65 L of textile effluent (Mezohegyi *et al.*, 2007). Dye treatment for effluent are presently able to reduce just partially of the dyes lost in wastewater streams. The international, national articles and magazines showed the pollution of rivers, agricultural lands and cases due to drinking water contamination by the effluent, coming out of the textile



industries. It has been prompted that find a new effective way is needed for the treatment of dye contaminated effluent to protect the ecosystem.

Several wastewater treatments are used including adsorption, physical-chemical treatment including coagulation, flocculation and filtration, advance treatment such as electrochemical, membrane separation, and reverse osmosis (Arulkumar *et al.*, 2011; Edward, 2005). Several literatures described that most of problem with physical-chemical treatment lies on the high cost operating with low efficiency, in addition to the need of huge space and limited flexibility. This is in contrast with another wastewater techniques and the treatment of the wastewater generated. Moreover, alternative by using adsorption process for decolorization of dyes is efficient, high competitive and simple (Sathishkumar *et al.*, 2011; Zheng *et al.*, 2009). Other innovative natural materials used in the production of carbonaceous solids include products from lignocelluloses substances e.g. palm shell, wood, sawdust, and also from vegetable waste substances e.g. fruit stones, nutshells, etc. (Adinata *et al.*, 2007; Alam *et al.*, 2009; Baccara *et al.*, 2009; Foo and Hameed, 2011; Srinivasakannan and Abu Bakar, 2004; Ucar *et al.*, 2009; Ould-Idriss *et al.*, 2011; Thio *et al.*, 2009; Yorgun *et al.*, 2009). The results from these lignocelluloses materials and wastes of vegetable origin reveal that their adsorption capacity was not so effective (Sarier, 2007). Furthermore, all technologies have advantages and disadvantages; these would be essential issues in search of any technology that are low-cost, easily obtained and environmentally friendly.

Microbial decolorization and biodegradation is greatly considered as an alternative for removal of textile dye effluent. Biological decolorization of dyes using fungi and bacteria has also been evaluated (Hadibarata *et al.*, 2011a; Maas and Chaudhari, 2005; Rodriguez-Couto *et al.*, 2003; Pearce *et al.*, 2003) to decolorized of azo, heterocyclic, polymeric dyes and triphenylmethane by white-rot fungus, *Polyporus* and *Phanerochaete chrysosporium* (Glen and Gold, 1983; Rodriguez *et al.*, 1999; Hadibarata *et al.*, 2011b; Pazarlioglu *et al.*, 2005). These fungi materials have been known to have good biodegradable capacity (Hadibarata *et al.*, 2011).

The microbial field which is fungal based bioremediation gives promising treatment on decolorization and degradation of polluting synthetic dyestuff. The

fungi strain that have capability to decolorize azo dye, type of Mordant Orange-1 (MO-1) were screened, isolated and identified. The forest is the best places for collecting fungi. The diversity of forest give fungi different capability to decolorize and degrade Mordant Orange-1 (MO-1). In addition, screening stage is the most important part to search new varieties of fungi strains in order to find the best degrader of Mordant Orange-1 (MO-1). A selected fungal strain was identified based on the 18S rRNA sequencing and microscope method to know the taxonomy and morphology characterization. Study of the effect of environmental factors (carbon, nitrogen sources, surfactant (Tween 80), aromatic compounds and pH) on decolorization was performed in the liquid medium and analyzed by UV-Visible Spectrophotometer. Finally, the metabolite product of MO-1 was analyzed in Gas Chromatography-Mass Spectrometry (GC-MS).

## 1.2 Problem of Statements

In Malaysia textile industry is mainly essential export sector besides plantation, oil and natural sources. This sector has been the seventh largest contributor to total earnings from manufactured exports, due to its high market either from inside and outside Malaysia. In Malaysia, the textile industry is concentrated mainly in the states of Johor, Perak, Penang and Selangor (Pang and Abdullah, 2013). Based on report from Department of Environment of Malaysia 2010, the textile industry produced approximately 743.99 metric tons per year of wastewater released by into river bodies and one of the substances were azo dyes. These compounds are one of the most difficult to treat. The majority of them are carcinogenic and toxic to living ecosystem. In fact, 1 ppm concentration might less rather than another chemical establish in wastewater, it can be visible instead at low concentrations (Godlewska *et al.*, 2009; Medvedev *et al.*, 1988).

### **1.3 Objectives of Study**

The present study was carried out to investigate the applicability of the fungi for the removal of mordant orange-1. The objectives of the study are:

1. To screen, isolate and identify the fungal strain isolated from nature for decolorization of an azo dye, Mordant Orange-1 (MO-1).
2. To investigate the effect of several environmental factors on decolorization of Mordant Orange-1 (MO-1).
3. To determine and identify the metabolite pathway of Mordant Orange-1 (MO-1) degradation.

### **1.4 Scope of Study**

The study utilized the filamentous fungi to decolorize Mordant Orange-1 (MO-1) in a solid and liquid medium in 15 and 30 days incubation time. The certain effect of environmental factors in decolorization such as carbon sources, nitrogen sources, effect of surfactant (Tween 80), aromatic compounds and initial pH value were investigated in the liquid medium. Identification of selected fungi was performed by using 18S rRNA sequence and microscope method. Metabolite pathway was determined by identify some products yield in degradation using some instruments such as TLC, UV-Visible Spectrophotometer, and GC-MS.

### **1.5 Significance of Study**

The significance of this research is to provide an alternative method for the decolorization and biodegradation of an azo dye Mordant Orange-1 (MO-1) by utilizing a new promising fungi from nature. Furthermore, the application of fungi for decolorization and degradation of synthetic dye is expected to be more

environmental friendly to human life, and contribute an effective method to solve problem in the treatment of wastewater from textile industry.

## REFERENCES

- Adinata, D., Daud, W. M. A. W. and Aroua, M. K. (2007). Preparation and characterization of activated carbon from palm shell by chemical activation with  $K_2CO_3$ . *Bioresource Technology*. 98: 145–149.
- Adosinda, M., Martins, M., Ferreira, I. C., Santos, I. M., Queiroz, M. J. and Lima, N. (2001). Biodegradation of bioaccessible textile azo dyes by *Phanerochaete crysoporium*. *Journal of Biotechnology*. 89: 91–98.
- Adosinda, M., Martins, M., Lima, N., Silvestre, A. and Queiroz, M. (2003). Comparative studies of fungal degradation of single or mixed bioaccessible reactive azo dyes. *Chemosphere*. 52: 967.
- Aguilera, L. M., Griffiths, R. P. and Caldwell B. A. (1993). Nitrogen in ectomycorrhizal mat and non-mat soils of different-age Douglas-fir forests. *Soil Biology and Biochemistry*. 25: 1015-1019.
- Alam, Md. Z., Ameen, E. S., Muyibi, S., Kabbashi, A. and Nassereldeen, A. (2009). The factors affecting the performance of activated carbon prepared from oil palm empty fruit bunches for adsorption of phenol. *Journal of Chemical Engineering*, 155: 191–198.
- Alexander, M. (1999). Biodegradation and bioremediation. New York: Academic press.
- Allen, S. J., McKay, G. and Porter, J. F. (2004). Adsorption isotherm models for basic dye adsorption by peat in single and binary component systems. *Journal of Colloid and Interface Science*. 280: 322–333.
- Ansari, A. A. and Thakur, B. D. (2000). Extraction, characterisation and application of a natural dye: The eco-friendly textile colorant. *Colourage*. 47: 15-16.

- Arulkumar, M., Sathishkumar, P. and Palvannan, T. (2011). Optimization of Orange G dye adsorption by activated carbon of *Thespesia populnea* pods using response surface methodology. *Journal of Hazardous Materials*. 186: 827–834.
- Arora, D. K. and Khachatourians, G. G. (2003). *Applied Mycology and Biotechnology*. Netherlands: Elsevier Science B.V.
- Asther, M., Corrieu, G., Drapron, R. and Odier, E. (1987). Effect of Tween 80 and oleic acid on ligninase production by *Phanerochaete chrysosporium* INA-12. *Enzyme and Microbial Technology*. 9: 245–249.
- Attia, A. A., Girgis, B. S. and Khedr, S. A. (2003). Capacity of activated carbon derived from pistachio shells by  $H_3PO_4$  in the removal of dyes and phenolics. *Journal of Chemical Technology and Biotechnology*. 78, 611–619.
- Baccar, R., Bouzida, J., Fekib, M. and Montiela, A. (2009). Preparation of activated carbon from Tunisian olive-waste cakes and its application for adsorption of heavy metal ions. *Journal of Hazardous Materials*. 162: 1522–1529.
- Badis, A., Ferradji, F. Z. Boucherit, A., Fodil, D. and Boutoumi, H. (2010). Removal of natural humic acids by decolorizing actinomycetes isolated from different soils (Algeria) for application in water purification. *Desalination*. 259: 216-222.
- Balan, D. S. L. and Monteiro, R. T. R. (2001). Decolorization of textile indigo dye by ligninolytic fungi. *Journal of Biotechnology*. 89: 141–145.
- Banat, I. M., Nigam, P., Singh, D. and Marchant, R. (1996). Microbial decolorization of textile dye-containing effluents: a review. *Bioresource Technology*. 58: 217–227.
- Barr, D. A. and Aust, S. D. (1994). Mechanisms white rot use to degrade pollutants. *Environmental Science and Technology*. 28: 79A–87A.

- Belsare, D. K. and Prasad, D. Y. (1988). Decolourization of effluent from the bagasse based pulp mills by white rot fungus. *Schizophyllum commune*. *Applied Microbiology and Biotechnology*. 28: 301-304.
- Bentley, R. and Bennett, J. W. (1988). In *Physiology of Industrial Fungi* (D.F. Berry, ed.). Oxford: Blackwell Scientific Publications.
- Blánquez, P., Casas, N., Font, X., Gabarrell, X., Sarrà, M., Caminal, G. and Vicent, T. (2004). Mechanism of textile metal dye biotransformation by *Trametes versicolor*. *Water Research*. 38: 2166–2172.
- Boer, C. G., Obici, L., Marques de Souza, C. G. and Paralta, R. M. (2004). Decolorization of synthetic dyes by solid state cultures of *Lentinula (Lentinus) edodes* producing manganese peroxidase as the main ligninolytic enzyme. *Bioresources Technology*. 94: 107–112.
- Boonchan, S., Britz, M. L. and Stanley, G. A. (2000). Degradation of high-molecular-weight polycyclic aromatic hydrocarbons by defined fungal-bacterial cocultures. *Applied and Environmental Microbiology*. 3: 1007-1019.
- Bras, R., Ferrá, M., Isabel, M., Pinheiro, H. M. and Goncalves, I. C. (2001). Batch tests for assessing decolorization of azo dyes by methanogenic and mixed cultures. *Journal of Biotechnology*. 89: 155-162.
- Bu'lock, J. D. (1980). In *The Biosynthesis of Mycotoxins, a Study in Secondary Metabolism* (P. S. Steyn, ed.). New York: Academic Press.
- Bumpus, J. A. and Tatarko, M. (1994). Biodegradation of 2,4,6-trinitrotoluene by *Phanerochaete chrysosporium*: identification of initial degradation products and the discovery of a TNT metabolite that inhibits lignin peroxidase. *Current Microbiology*. 28: 185-190.
- Call, H. P. and Mucke, I. (1997). Mini review: history, overview and applications of mediated ligninolytic systems, especially laccase-mediator-systems (Lignozym-Process). *Journal of Biotechnology*. 53: 163–202.

- Camarero, S., Ibarra, D., Martinez, M. J. and Martinez, A. T. (2005). Lignin-derived compounds as efficient laccase mediators for decolorization of different types of recalcitrant dyes. *Applied and Environmental Microbiology*. 71: 1775–1784.
- Cralile, M. J., Sarah, C. W. and Graham. (2001). *The Fungi*. London: Academic Press a Harcourt Science and Technology Company.
- Cervantes, F. J. (2002). Quinones as Electron Acceptors and Redox Mediators for the Anaerobic Biotransformation of Priority Pollutants. *Agrotechnology and Food Science, Sub-department of Environmental Technology*. Wageningen University, Netherland.
- Chen, K., Wu, J., Liou, D. and Hwang, S. J. (2003). Decolorization of the textile dyes by newly isolated bacterial strains. *Journal of Biotechnology*. 101: 57–68.
- Chen, B. Y., Lin, K. W., Wang, Y. M. and Yen, C. Y. (2009). Revealing Interactive Toxicity of Aromatic Amines to Azo Dye Decolorizer *Aeromonas hydrophila*. *Journal Hazardous Material*. 166: 187-194.
- Chung, K. T. and Cerniglia, C. E. (1992). Mutagenic of azo dyes: Structure-activity relationship. *Mutation Research*. 277: 201-220.
- Chivukula, M. and Renganathan, V. (1995). Phenolic azo dye oxidation by laccase from *Pyricularia oryzae*. *Applied and Environmental Microbiology*. 61: 4374-4377.
- Ciullini, I., Tilli, S., Scozzafava, A. and Briganti, F. (2008). Fungal laccase, cellobiose dehydrogenase, and chemical mediators: combined actions for the decolorization of different classes of textile dyes. *Bioresource Technology*. 99: 7003–7010.
- Cohen, R. Hadar, Y. and Yarden, O. (2001). Transcript and activity levels of different *Pleurotus ostreatus* peroxidases are differentially affected by Mn<sup>2+</sup>. *Environmental Microbiology*. 3: 312-322.



- Cole, D. W. (1981). Nitrogen uptake and translocation by forest ecosystems. In *Terrestrial Nitrogen Cycles* (ed. F. E. Clark & T. Rosswall). *Ecological Bulletins (Stockholm)*. 33: 219-232.
- Conneely, A., Smyth, W. F. and McMullan, G. (1999). Metabolism of phthiocyanine textile dye remazol turquoise blue by *Phanerochaete chrysosporium*. *FEMS Microbiology Letters*. 179: 333-337.
- Cooke, R. C. and Whipps, J. M. (1993). *Ecophysiology of fungi*. Oxford: Blackwell Scientific Publication.
- Craliell, C. M., Barclay, S. J., Naidoo, N., Buckley, C. A., Mulholland, D. A. and Senior, E. (1995). Microbial decolorization of a reactive azo dye under anaerobic conditions. *Water SA*. 21: 61–69.
- Cripps, C., Bumpus, J. A. and Aust, S. D. (1990). Biodegradation of azo and heterocyclic dyes by *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*. 56: 1114–8.
- Crawford, N. M and Arst, Jr. H. N. (1993). The molecular genetics of nitrate assimilation in fungi and plants. *Annual Review of Genetics*. 27: 115-146.
- Deveci, T., Unyayar , A. and Mazmanci, M. A. (2004). Production of Remazol Brilliant Blue R decolorizing oxygenase from the culture filtrates of *Funalia trogii* ATCC 20080. *Journal of Molecular Catalysis B. Enzymatic*. 30: 25–32.
- Department of Environment (DOE). (2010). Environmental Quality (Industrial Effluents) Regulations, 2009. Official Website of Department of Environment, Ministry of Natural Resources and Environment, Malaysia. <http://www.doe.gov.my/en/content/environmental-quality-act-1974> (Accessed April 2012).
- Dong, J. L., Zhang, Y. W., Zhang, R. H., Huang, W. Z. and Zhang, Y. Z. (2005). Influence of culture condition on laccase production and isozyme patterns in the white-rot fungus *Trametes gallica*. *Journal of Basic Microbiology*. 45: 190-198.

- dos Santos, A. B. (2005). Reductive decolourization of dyes by thermophilic anaerobic granular sludge. Sub-department of Environmental Technology. Wageningen University, Netherland.
- Dunnivant, F. M., Jardine, P. M., Taylor, D. L. and McCarthy, J. F. (1992). Cotransport of cadmium and hexachlorobipheyl by dissolved organic carbon through columns containing aquifer material. *International Journal of Environmental Science and Technology*. 26: 360–368.
- Edward, E. B. (2005). Water Treatment Plant Design 4<sup>th</sup> Edition. USA: McGraw-Hill Inc.
- El-Sheekh, M. M., Gharieb, M. M. and Abou-El-Souod, G. W. (2009). Biodegradation of dyes by some green algae and cyanobacteria. *International Biodeterioration and Biodegradation*. 63: 699-704.
- Ferreira, V. S., Magalhaes, D. B., Kling, S. H., Da Silva, J. G. and Bon E. P. S. (2000). N-demethylation of Methylene Blue by lignin peroxidase from *Phanerochaete chrysosporium*. *Applied Biochemistry and Biotechnology*. 84–86: 255–265.
- Fewson, C. A. (1988). Biodegradation of xenobiotics and other persistent compounds: causes of recalcitrant. *Trends in Biotechnology*. 6: 148-153.
- Finlay, R. D., Frostegard, A. and Sonnerfeld, A-M. (1992). Utilization of organic and inorganic nitrogen sources by ectomycorrhizal fungi in pure culture and in symbiosis with *Pinus contoria* Dougl. ex Loud. *New Phytogist*. 120: 105-115.
- Foo, K. Y. and Hameed, B. H. (2010). Fixed-bed adsorption of reactive azo dye onto granular activated carbon prepared from waste. *Journal of Hazardous Materials*. 175: 298–303.
- Foo, K. Y. and Hameed, B. H. (2011). Preparation and characterization of activated carbon from sunflower seed oil residue via microwave assisted K<sub>2</sub>CO<sub>3</sub> activation. *Bioresource Technology*. 102: 9794–9799.

- Fredrickson, J. K. Kostandaristhes, H. M., Li, S. W. Plymale, A. E. and Daly, M. J. (2000). Reduction of Fe(II), Cr (VI), U (VI) and Tc (VII) by *Deinococcus rediodurans* R1. *Applied and Environmental Microbiology*. 66: 2006-2011.
- Fu, Y., and Viraraghavan, T. (2001). Fungal decolorization of dye wastewaters: a review. *Bioresource Technology*. 79: 251–262.
- Fujita, K., Asami, Y. and Murata, E. (2002). Effects of thalidomide, cytochrome P-450 and TNF-alpha on angiogenesis in a three-dimensional collagen gel-culture. *Okajimas Folia Anatomica Japonica*.79: 101–106.
- Ganesh, R., Boardman, G. D. and Michelsen, D. (1994). Fate of azo dyes in sludges. *Water Research*. 28: 1367-1376.
- Godlewska, E. Z., Przystaś, W. and Sota E. G. (2009). Decolourization of triphenylmethane dyes and ecotoxicity of their end Products. Poland: Gliwice.
- Glen, J. and Gold, M. H., (1983). Decolourization of several polymeric dyes by the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*. 45: 1741–1747.
- Graciela, M. L. Ruiz-Aguilar., Jose, M., Ferná'ndez-Sa'nchez., Refugio, Rodrí'guez-Va'zquez., He'ctor. and Poggi-Varaldo. (2002). Degradation by white-rot fungi of high concentrations of PCB extracted from a contaminated soil. *Advances in Environmental Research*. 6: 559–568.
- Griffiths, J. (1984). Developments in the light absorption properties of dyes-color and photochemical reactions. In: Griffiths, J. (Ed.), *Developments in the Chemistry and Technology of Organic Dyes*. Oxford: *Society of Chemistry Industry*.
- Hadibarata, T., Tachibana, S. and Itoh, K. (2007). Biodegradation of Phenanthrene Fungi Screened from Nature. *Pakistan Journal of Biological Sciences*. 10: 2535-2543.

- Hadibarata, T. and Tachibana, S. (2009). Bioremediation of Phenanthrene, Chrysene, and Benzo[a]pyrene by Fungi Screened from Nature. *ITB Journal of Science*. 41: 88-97.
- Hadibarata, T., Tacibana, S. and Askari, M. (2011a). Identification of metabolites from phenanthrene oxidation by phenoloxidases and dioxygenases of *polyporus* sp. S133. *Journal of Microbiology and Biotechnology*. 21: 299-304.
- Hadibarata, T., Yusoff, A. R. M. and Kristanti, R. A. (2011b). Decolorization and metabolism of antraquinone-type dye by laccase of white-rot fungi *polyporus* sp. S133. *Water Air and Soil Pollution*. DOI:10.1007/s11270-011-0914-6.
- Hadibarata, T., Yusoff, A. R. M., Salmiati, Hidayat, T. and Kristanti, R. A. (2012). Decolorization of azo, triphelylmethane and antraquinone dyes by laccase of a newly isolated *Armillaria* sp. F022. *Water Air and Soil Pollution*. DOI:10.1007/s11270-011-0922-6.
- Hadibarata, T. and Kristanti, R. A. (2012). Effect of Environmental Factors in the Decolorization of Remazol Brilliant Blue R by *polyporus* sp. S133. *Journal of the Chemical Society*. 57: 1095-1098.
- Hadibarata, T., Adnan. L. A., Yusoff, A. R. M., Yuniarto, A., Rubiyatno., Zubir, M. M. F. A., Khudhair, A. B., Teh, Z. C. (2013). Microbial Decolorization of an Azo Dye Reactive Black 5 Using White-Rot Fungus *Pleurotus eryngii* F032. *Water Air and Soil Pollution*. DOI 10.1007/s11270-013-1595-0
- Hall, G. G. (2008). *Phylogenetic Trees Made Easy*. USA: Sinauer Associates.
- Hatvani, N. and Mecs, I. (2002). Effect of the nutrient composition on dye decolorisation and extracellular enzyme production by *Lentinus edodes* on solid medium. *Enzyme and Microbial Technology*. 30: 381–386.
- Haug, W., Schmidt, A. and Nortemann. (1991). Mineralization of the sulfonated azo dye mordant yellow 3 by 6-aminonaphthalene-2-sulfonate degrading bacterial consortium. *Applied and Environmental Microbiology*. 57: 3144-3149.

- Hawksworth, D. L., Kirk, T. M., Sutton, B. C., and Pegler, D. N. (1995). *Ainsworth and Biesby's Dictionary of Fungi*. UK: CAB International.
- Hawksworth, D. L. (2001). The magnitude of fungal diversity: the 1.5 million species estimate revisited. *Mycological Research*. 105: 1422–1432.
- Hawksworth, D. L. (2002). Mycological Research News. *Mycological Research*. 106: 1-3.
- Heinflig, A., Martinez, M. J., Martinez, A. T., Bergbauer, M. and Szewzyk, U. (1998). Transformation of industrial dyes by manganese peroxidases from *Bjerkandera adusta* and *Pleurotus eryngii* in a manganese-independent reaction. *Applied and Environmental Microbiology*. 64: 2788–2793.
- Higson, F. K. (1991). Degradation of xenobiotics by white rot fungi. *Reviews of Environmental Contamination and Toxicology*. 122: 111–152.
- Hofrichter, M., Bublitz, F. and Fritsche, W. (1997). Fungal attack on coal. II. solubilization of low-rank coal filamentous fungi. *Fuel Processing Technology*. 52: 55–64.
- Howard, R., Abotsi, L., Rensburg, E-J. V. E. and Howard, S, L. (2003). Lignocellulose biotechnology: issues of bioconversion and enzyme production. *African Journal of Biotechnology*. 2: 602-619.
- Idris, A. B., Eltayeb, M. A. H., Potgieter-Vermaak, S. S., Grieken, R. V. and Potgieter, J. H. (2007). Assessment of heavy metal pollutant in Sudanese Harbours Along the Red Sea Coast. *Microbiology and Chemistry journal*. 87: 104-112.
- Ishibashi, Y., Cervantes, C. and Silver, S. (1990). Chromium reduction in *Pseudomonas putida*. *Applied and Environmental Microbiology*. 56: 2268-2270.

- Jadhav, U. U., Dawkar, V. V., Tamboli, D. P. and Govindwar, S. P. (2009). Purification and characterization of veratryl alcohol oxidase from *Comomonas* sp. UVS and its role in decolorization of textile dyes. *Biotechnology and Bioprocess Engineering*. 14: 369-376.
- Jadhav, J. P., Kalyani, D. C., Phugare, S. S. and Govindwar, S. P. (2010). Evaluation of the efficacy of a bacterial consortium for the removal of color, reduction of heavy metals, and toxicity from textile dye effluent. *Bioresource Technology*. 101: 165-173.
- Jennings, D. (1995). *Physiology of Fungal Nutrition*. New York: Cambridge University Press.
- Juhasz, T., Szengyel, Z., Reczey, K., Siika-Aho, M. and Viikari, L. (2005). Characterization of cellulases and hemicellulases produced by *Trichoderma reesei* on various carbon sources. *Process Biochemistry*. 40: 3519-3525.
- Kahraman, S. S., and Gurdal, I. H. (2002). Effect of synthetic and natural culture media on laccase production by white rot fungi. *Bioresource Technology*. 82: 215-7.
- Kalme, S., Ghodake, G. and Govindwar, S. (2007). Red HE7B degradation using desulfonation by *Pseudomonas desmolyticum* NCIM 2112. *International Biodeterioration and Biodegradation*. 60: 327-333.
- Kalme, S., Jadhav, S., Jadhav, M. and Govindwar, S. (2009). Textile dye degrading laccase from *Pseudomonas desmolyticum* NCIM 2112. *Enzyme and Microbial Technology*. 44: 65-71.
- Kalyani, D. C., Patil, P. S. Jadhav, J. P. and Govindwar, S. P. (2009). Biodegradation of reactive textile dye red BLI by an isolated bacterium *Pseudomonas* sp. SUK1. *Bioresource Technology*. 99: 4635-4641.
- Kariminiaae-Hamedani, H. R., Sakurai, A. and Sakakibara, M. (2007). Decolorization of synthetic dyes by a new manganese peroxidase-producing white rot fungus. *Dyes and Pigments*. 72: 157-162.

- Karigar, C. S. and Rao, S. S. (2011). Role of Microbial Enzymes in the Bioremediation of Pollutants: A Review. *Enzyme Research*. doi:10.4061/2011/805187
- Karthikeyan, K., Nanthakumar, K., Shanthi, K. and Lakshmanaperumalsamy, P. (2010). Response surface methodology for optimization of culture conditions for dye decolorization by a fungus *Aspergillus niger* HM11 isolated from dye affected soil. *Iranian Journal of Microbiology*. 2: 213–222.
- Keck, A., Klein, J., Kudlich, M., Stolz, A., Knackmuss, H. J. and Mattes, R. (1997). Reduction of azo dyes by redox mediators originating in the naphthalenesulfonic acid degradation pathway of *Sphingomonas sp.* strain BN6. *Applied Environmental Microbiology*. 63: 3684–3690.
- Kersten, P. J., Tien, M., Kalyanaraman, B. and Kirk, T. K. (1985). The ligninase from *Phanerochaete chrysosporium* generates cation radicals from methoxybenzenes. *Journal of Biological Chemistry*. 260: 2609–2612.
- Kirk, T. K. and Farrell, R. L. (1987). Enzymatic combustion. The microbial degradation lignin. *Annual Review Microbiology*. 41: 465-505.
- Kirk, T. K., Schultz, E., Connors, W. J., Lorenz, L. F. and Zeikus, J. G. (1978). Influence of culture parameters on lignin metabolism by *Phanerochaete chrysosporium*. *Archives of Microbiology*. 117: 277– 85.
- Kotterman, M. J. J., Rietberg, H. J., Hage, A. and Field, J. A. (1998). Polycyclic aromatic hydrocarbon oxidation by the white-rot fungus *Bjerkandera sp.* strain BOS55 in the presence of nonionic surfactants. *Biotechnology and Bioengineering*. 57: 220–227.
- Kuhad, R. C., Singh, A. and Eriksson, K-E. L. (1997). Microorganisms and enzymes involved in the degradation of plant fiber cell walls. In: *Biotechnology in the pulp and paper industry. Advances in Biochemical Engineering Biotechnology*. 57: 97–105.

- Kuhn, S. P. and Pfister, R. M. (1990). Accumulation of cadmium by immobilized *zoolea ramigea* 115. *Indian Journal of Microbiology*. 6: 123-128.
- Kulla, H. G., Klausener, F., Meyer, U., Luedeke, B. and Leisinger, T. (1983). Evolution of new bacterial enzyme activities during adsorption to azo dyes. *Archives of Microbiology*. 135: 1-7.
- Kwon, O. U., Ogino, K. and Ishikawa, H. (1991). The Longest 18S Ribosomal RNA Ever Known. *European Journal of Biochemistry*. 202: 827-833.
- Lacina, C., Germain, G. and Spiros, A. N. (2003). Utilization of fungi for biotreatment of raw wastewater. *African Biotechnology*. 2: 620-630.
- Lee, R. E. (2008). Phycology. USA : Publish by Cambridge University Press.
- Laha, S. and Luthy, R. G. (1992). Inhibition of phenanthrene mineralization by nonionic surfactants in soil water systems. *International Journal of Environmental Science and Technology*. 25: 1920–1930.
- Leatham, G. F. and Kirk, T. K. (1983). Regulation of ligninolytic activity by nutrient N in white-rot basidiomycetes. *FEMS Microbiology Letters*. 16: 65–67.
- Lehninger, A. L., Nelson, D. L. and Cox, M. M. (2004). Lehninger's Principles of Biochemistry. USA: New York. W.H. Freeman.
- Levin, L., Papinutti, L. and Forchiassin, F. (2004). Evaluation of Argentinean white rot fungi for their ability to produce lignin-modifying enzymes and decolorize industrial dyes. *Bioresources Technology*. 94: 169–176.
- Lilly, V. M. and Barnett, H. L. (1951). Physiology of the fungi. New York: McGraw-Hill Book Co.
- Lovely, D. R., Fraga, J. L., BluntHarris, E. L., Hayes, L. A., Philips, E. J. P. and Coates, J. D. (1998). Humic substances as a mediator for microbially catalyzed metal reduction. *Acta Hydrochemistry Hydrobiology*. 26: 152-157.



- Maas, R. and Chaudhari, S. (2005). Sorption and biological decolorization of azo dye reactive red 2 in semi continuous anaerobic reactors. *Process Biochemistry*. 40: 699–705.
- Malaysian Textile Manufacturers Association. (2008). Statistic of Malaysian import and export. [http://www.fashion-asia.com/page.cfm?name=Statistic%20\(Import%20and%20Export\)](http://www.fashion-asia.com/page.cfm?name=Statistic%20(Import%20and%20Export)) (Accessed 25. 04. 2012).
- Margesin, R. (2000). Potential of cold-adapted microorganisms for bioremediation of oil-polluted Alpine soils. *International Biodeterioration and Biodegradation*, 46: 3-10.
- Mansur, M., Arias, M. E., Copa-Patino, J. L., Flardh, M. and Gonzalez, A. E. (2003). The white-rot fungus *Pleurotus ostreatus* secretes laccase isozymes with different substrate specificities. *Mycologia*. 95: 1013-1020.
- McMullan, G., Meehan, C., Conneely, A., Kirby, N., Robinson, T. and Nigam, P. (2001). Microbial decolourisation and degradation of textile dyes. *Applied Microbiology and Biotechnology*. 56: 81–87.
- Mechichi, T., Mhiri, N. and Sayadi, S. (2006). Remazol Brilliant Blue R decolorization by the laccase from *Trametes trogii*. *Chemosphere*. 64: 998-1005.
- Medvedev, Z. A., Crowne, H. M. and Medvedeva, M. N. (1988). Age related variations of hepatocarcinogenic effect of azo dye (30-MDAB) as linked to the level of hepatocyte polyploidization. *Mechanisms of Ageing and Development*. 46: 159–174.
- Mezohegyi, G. Kolodkin, A. Castro, U. I., Bengoa, C., Stuber, F., Font, J. and Fabregat, A. (2007). Effective Anaerobic Decolorization of Azo Dye Acid Orange 7 in Continuous Upflow Packed-Bed Reactor Using Biological Activated Carbon System. *Industrial and Engineering Chemistry Research*. 46: 6788-6792.

- Milojkovic-Opsenica, D. M., Lazarevic, K., Ivackovic, V. and Tesic, Z. L. (2003). Reversed-phase thin-layer chromatography of some foodstuff dyes. *Journal of Planar Chromatography-Modern TLC*. 16: 276-279,
- Moir, D., Masson, S. and Chu, I. (2001). Structure-activity relationship study on the bioreduction of azo dyes by *Clostridium paraputrificum*. *Environmental of toxicology in chemistry*. 20: 479-488.
- Moldes, D. and Sanromán, M. A. (2006). Amelioration of the ability to decolorize dyes by laccase: relationship between redoxmediators and laccase isoenzymes in *Trametes versicolor*. *World Journal of Microbiology and Biotechnology*. 22: 1197-1204.
- Moss, M. O. (1984). In *The Ecology and Physiology of the Fungal Mycelium* (D. H. Jennings and A. D. M. Rayner, ed.). Cambridge, UK: Cambridge University Press.
- Murugesan, K., Yang, I. H., Kim, Y. M., Jeon, J. R. and Chang, Y. S. (2009). Enhanced transformation of malachite green by laccase of *Ganoderma lucidum* in the presence of natural phenolic compounds. *Applied Microbiology and Biotechnology*. 82: 341-350.
- Nigam, P., Banat, I. M., Singh, D. and Marchant, R. (1996). Microbial process for the decolorization of textile effluent containing azo, diazo and reactive dyes. *Process Biochemistry*. 31: 435-442.
- Noroozil, B. and Sorial, G. A. (2013). Applicable models for multi-component adsorption of dyes: A review. *Journal of Environmental Sciences*. 25: 419-429.
- Nyanhongo, G., Gomes, J., Gübitz, G., Zvauya, R., Read, J. and Steiner, W. (2002). Production of laccase by a newly isolated strain of *Trametes modesta*. *Bioresource Technology*. 84: 259-263.
- O'Neill, C., Hawkes, F. R., Hawkes, D. L., Lourenco, N. D., Pinheiro, H. M. and Delee, W. (1999). Color in textile effluent sources, measurement, discharge,

- consents and stimulation: a review. *Journal of Chemical Technology and Biotechnology*. 74: 1009-1018.
- Ould-Idriss, A., Stitou, M., Cuerda-Correa, E. M., Fernández-González, C., Macías-García, A., Alexandre-Franco, M. F. and Gómez-Serrano, V. (2011). Preparation of activated carbons from olive-tree wood revisited I. Chemical activation with H<sub>3</sub>PO<sub>4</sub>. *Fuel Processing Technology*. 92: 261–265.
- Ozmen, E. Y., Erdemir, S., Yilmaz, M. and Bahadir, M. (2007). Removal of carcinogenic direct azo dyes from aqueous solution using calix[n]arene derivatives, *Clean Soil Air Water*. 35: 612–619.
- Palmieri, G., Cennamo, G. and Sannia, G. (2005). Remazol Brilliant Blue R decolourization by the fungus *Pleurotus ostreatus* and its oxidative enzymatic system. *Enzyme and Microbial Technology*. 36: 17–24.
- Pandey, G., Paul, D. and Jain, R. K. (2005). Conceptualizing “suicidal genetically engineered microorganisms” for bioremediation applications. *Biochemical and Biophysical Research Communications*. 327: 637-639.
- Pandey, A. Singh, P. and Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *International Biodeterioration and Biodegradation*. 59: 73-84.
- Pang, Y. L. and Abdullah, A. Z. (2013). Current Status of Textile Wastewater Management and Research Progress in Malaysia: A Review. *Clean Soil Air Water*. doi: 10.1002/clen.201000318.
- Paszczynski, A., Pasti-Grigsby, M. B., Goszczynski, S., Crawford, R. L. and Crawford, D. L. (1992). Mineralization of sulfonated azo dyes and sulfanilic acid by *Phanerochaete chrysosporium* and *Streptomyces chromofuscus*. *Applied and Environmental Microbiology*. 58: 3598–3604.
- Pazarlioglu, N. K., Urek, R. O. and Ergun, F. (2005). Biodecolourization of Direct Blue 15 by immobilized *Phanerochaete chrysosporium*. *Process Biochemistry*. 40: 1923–1929.

- Pearce, C. I., Lloyd, J. R. and Guthrie, J. T. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments*. 58: 179–96.
- Pereira, L., Coelho, A. V., Viegas, C. A., Correia dos Santos, M. M., Robalo, M. P. and Martins, L.O. (2009). Enzymatic biotransformation of the azo dye sudan orange G with bacterial CotA-laccase. *Journal of Biotechnology*. 139: 68-77.
- Poole, C. F. and Poole, S. K. (1992). Gas chromatography. In *Chromatography*, 5<sup>th</sup> edition, fundamentals and applications of chromatography and related differential migration methods. New York: E. Heftmann Ed. Elsevier.
- Providenti, M. A., Lee, H. and Trevors, J. T. (1993). Selected factors limiting the microbial degradation of recalcitrant compounds. *Journal of Industrial Microbiology*. 12: 379–395.
- Ratner, B. D., Hoffman, A. S., Schoen, F. J. M. D. and Lemons, J. E. (2004). *Biomaterials Science*. USA: Elsevier Academic Press.
- Reichstein, T. F. (1992). *Chromatography*, 5<sup>th</sup> edition, fundamentals and applications of chromatography and related differential migration methods. New York: E. Heftmann Ed. Elsevier.
- Reinhammar, B. (1984). Laccase. In: Lontie R (ed) *Copper proteins and copper enzymes*. Boca Raton: CRC.
- Rieger, P-G., Meier, H-M., Gerle, M., Vogt, U., Groth, T. and Knackmuss, H-J. (2002). Xenobiotics in the environment: present and future strategies to obviate the problem of biological persistence. *Journal of Biotechnology*. 94: 101–123.
- Rittmann, B. E. and McCarty, P. L. (2001). *Environmental Biotechnology: Principles and Applications*. USA: McGraw Hill Inc.
- Robinson, T., McMullan, G., Marchant, R. and Nigam, P. (2001). Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. *Bioresource Technology*. 77: 247–55.

- Rodriguez, E., Pickard, M. A. and Vazquez-Duhalt, R. (1999). Industrial dye decolourization by laccases from ligninolytic fungi. *Current Microbiology*. 38: 27–32.
- Rodriguez-Couto, S., Moldes, D., Liébanas, A. and Sanromán, A. (2003). Investigation of several bioreactor configurations for laccase production by *Trametes versicolor* operating in solid-state conditions. *Biochemical Engineering Journal*. 15: 21–26.
- Rogalski, J. and Leonowicz, A. (1992). *Phlebia radiata* laccase forms induced by veratric acid, and xyloidine in relation to lignin peroxidase, and manganese-dependent peroxidase. *Acta Biotechnologica*. 12: 213–221.
- Rosen, M. J. (1989). *Surfactants and Interfacial Phenomena*. New York: Wiley.
- Rouse, J. D., Sabatini, D. A., Suflita, J. M. and Harwell, J. H. (1994). Influence of surfactant on microbial degradation of organic compounds. *Critical Reviews in Environmental Science and Technology*. 24: 325–370.
- Ruijter, G. J., Kubicek, C. P. and Visser, J. (2002). Production of Organic Acids by Fungi. In *The Mycota X. Industrial Applications* (H.D.Osiewacz, ed). Berlin Heidelberg: Springer-Verlag. pp. 213-230.
- Sam, M. and Yesilada, O. (2001). Decolorization of Orange II dye by white-rot fungi. *Folia Microbiologica*. 46: 143-145.
- Salami, M. and Vandamme, A. M. (2003). *The Phylogenetic Hand Book: A Practical Approach to DNA And Protein Phylogeny*. UK: Cambridge University publisher.
- Sarier, N. (2007). Specific features of sorption of azo dyes on fly ash. *Russian Chemical Bulletin international Edition*. 56: 566–569.
- Sarioglu, M., Bali, U. and Bisgin, T. (2007). The removal of C.I. basic red 46 in a mixed methanogenic anaerobic culture. *Dyes and Pigments*. 74: 223–229.

- Saranaik, S. and Kanekar, P. (1995). Bioremediation of color of methyl violet and phenol from a dye industry waste effluent using *Pseudomonas sp.* isolated from factory soil. *Journal of Applied Bacteriology*. 79: 459–469.
- Sathishkumar, P. Arulkumar, M. and Palvannan, T. (2011). Utilization of agro-industrial waste *Jatropha curcas* pods as an activated carbon for the adsorption of reactive dye Remazol Brilliant Blue R (RBBR). *Journal of Cleaner Production*. 22: 67-75.
- Shiau, B. J., Sabatini, D. A. and Harwell, J. H. (1995). Properties of food grade (edible) surfactants affecting subsurface remediation of chlorinated solvents. *International Journal of Environmental Science and Technology*. 29: 2929–2935.
- Singh, A., Manju., Rani, S. and Bishnoi, N. R. (2012). Malachite green dye decolorization on immobilized dead yeast cells employing sequential design of experiments. *Ecological Engineering*. 47: 291-296.
- Sirianuntapiboon, S., Phothisilangka, P. and Ohmomo, S. (2004). Decolorization of molasses wastewater by a strain No.BP103 of acetogenic bacteria. *Bioresource Technology*. 92: 31-39.
- Sherma, J. (2000). Planar chromatography. *Analytical Chemistry*. 72: 9-25.
- Skoog, D. A., Holler, E. J. and Nieman, T. A. (1999). Principles of instrumental analysis 5<sup>th</sup> edition. Philadelphia: Saunders collage publishing.
- Spadaro, J. T., Gold, M. H. and Renganathan, V. (1992). Degradation of azo dyes by the lignin degrading fungus *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*. 58: 2397–2401.
- Spadaro, J. T. and Renganathan, V. (1994). Peroxidase-catalyzed oxidation of azo dyes: mechanism of Disperse Yellow 3 degradation. *Archives of Biochemistry and Biophysics*. 312: 301-307.
- Spain, J. C. and Gibson, D. T. (1991). Pathway for biodegradation of p-nitrophenol in a *Moraxella sp.* *Applied and Environmental Microbiology*. 57: 812-819.

- Srinivasakannan, C. and Bakar, A. M. Z. (2004). Production of activated carbon from rubber wood sawdust. *Biomass and Bioenergy*. 27: 89–96.
- Steffen, K. T., Hofrichter, M. and Hatakka, A. (2000). Mineralisation of <sup>14</sup>C-labelled synthetic lignin and ligninolytic enzyme activities of litter-decomposing basidiomycetous fungi. *Applied Microbiology and Biotechnology*. 54: 819–825.
- Svobodová, K., Senholdt, M., Novotny, C. and Rehorek, A. (2007). Mechanism of Reactive Orange 16 degradation with the white rot fungus, *Irpex lacteus*. *Process Biochemistry*. 42: 1279–1284.
- Swamy, J. and Ramsay, J. A. (1999). The evaluation of white rot fungi in the decolouration of textile dyes. *Enzyme and Microbial Technology*. 24: 130–137.
- Tatarko, M. and Bumpus, J. A. (1998). Biodegradation of Congo Red by *Phanerochaete chrysosporium*. *Water Research*. 32: 1713-1717.
- Telke, A. A., Kalyani, D. C., Dawkar, V. V. and Govindwar, S. P. (2009). Influence of Organic and Inorganic Compounds on Oxidoreductive Decolorization of Sulfonated Azo Dye C.I.Reactive Orange 16. *Journal of Hazardous Materials*. 172: 298-309.
- Thibault, S. L., Anderson, M. and Frankenberger Jr. W. T. (1996). Influence of surfactants on pyrene desorption and degradation in soils. *Applied and Environmental Microbiology*. 62: 283–287.
- Thio, C. C., Mirna, M. M., Sunarso, J., Sudaryanto, Y. and Ismadji, S. (2009). Activated carbon from durian shell: Preparation and characterization. *Journal of the Taiwan Institute of Chemical Engineer*. 40: 457–462.
- Thurston, C. F. (1994). The structure and function of fungal laccase. *Microbiology*. 140: 19–26.
- Tien, M. and Kirk, T. K. (1983). Lignin-degrading enzyme from the Hymenomycete *Phanerochaete chrysosporium* Burds. *Science*. 221: 661–663.

- Turner, W. B. (1971). *Fungal Metabolites*. London: Academic Press.
- Turner, W. B. and Aldridge, D. C. (1983) *Fungal Metabolites 11*. London: Academic Press.
- Ucar, S., Erdem, M., Tay, T. and Karago, S. (2009). Preparation and characterization of activated carbon produced from pomegranate seeds by ZnCl<sub>2</sub> activation. *Applied Surface Science*. 225: 8890–8896.
- Umezawa, T. and Higuchi, T. (1987). Mechanism of aromatic ring cleavage of h-O-4 lignin substructure models by lignin peroxidase. *FEBS Letters*. 218: 255–260.
- Van der Zee, F. P. (2002). Anaerobic azo dye reduction. Doctoral Thesis, Wageningen University, Wageningen. Netherlands.
- Vasdev, K., Kuhad, R. C. and Saxena, R. K. (1995). Decolorization of triphenylmethane dyes by the bird's nest fungus by *Cyathus bulleri*. *Current Microbiology*. 30: 269-272.
- Volkering, F., Breure, A. M., Van Andel, J. G. and Rulkens, W. H. (1995). Influence of nonionic surfactants on bioavailability and biodegradation of polycyclic aromatic hydrocarbons. *Applied and Environmental Microbiology*. 61: 1699–1705.
- Waghmode, T. R., Kurade, M. B., Kabra, A. N. and Govindwar, S. P. (2012). Degradation of Remazol Red Dye by *Galactomyces geotrichum* MTCC 1360 leading to increased iron uptake in *Sorghum vulgare* and *Phaseolus mungo* from Soil. *Biotechnology Bioengineering*. 17: 117–126.
- Wallace, T. H. (2001). Biological treatment of synthetic dye water and an industrial textile wastewater containing azo dye compounds. Master thesis, Virginia Polytechnic Institute and State University.
- Wesenberg, D., Kyriakides, I. and Agathos, N. (2003). White-rot fungi and their enzymes for the treatment of industrial dye effluents. *Biotechnology Advances*. 22: 161–187.



- Willmott, N. J., Guthrie, J. T. and Nelson, G. (1998). The biotechnology approach to colour removal from textile effluent. *Journal of the Society of Dyers and Colourists*. 114: 38–41.
- Will, R., Ishikawa, Y. and Leder, A. (2000). Synthetic dyes, chemical economics handbook: synthetic dyes. Menlo Park (CA): SRI Chemical and Health Business Service.
- Yang, Q., Yediler, A., Yang, M. and Kettrup, A. (2005). Decolorization of an azo dye, Reactive Black 5 and MnP production by yeast isolate: *Debaryomyces polymorphus*. *Biochemical Engineering Journal*. 24: 249-253.
- Yang, J. G., Liu, X., Long, T., Yu, G., Peng, S. and Zheng, L. (2003). Influence of nonionic surfactant on the solubilization and biodegradation of phenanthrene. *Journal of Environmental Sciences*. 15: 859–862.
- Yemendzhiev, H., Alexieva, Z. and Krastanov, A. (2009). Decolorization of synthetic dye Reactive Blue 4 by mycelial culture of white-rot fungi *Trametes versicolor* 1. *Biotechnology and Biotechnological Equipment*. 23: 1337–9.
- Yorgun, S., Vural, N. and Demiral, H. (2009). Preparation of high-surface area activated carbons from Paulownia wood by ZnCl<sub>2</sub> activation. *Microporous and Mesoporous Materials*. 122: 189–194.
- Xin, B., Xia, Y., Zhang, Y., Aslam, H., Liu, C. and Chen, S. (2012). A feasible method for growing fungal pellets in a column reactor inoculated with mycelium fragments and their application for dye bioaccumulation from aqueous solution. *Bioresource Technology*. 105: 100–105.
- Zak, J. C. and Wildman, H. G. (2004). Fungi in stressful environments. In *Measuring and Monitoring Biological Diversity: Standard Methods for Fungi*, Mueller, G., Bills, G. (Eds.). Washington, DC: Smithsonian Institution Press.
- Zhang, F., Yediler, A., Liang, X. and Kettrup, A. (2004). Effects of dye additives on the ozonation process and oxidation by products: a comparative study using hydrolysed CI Reactive red 120. *Dyes and Pigments*. 60: 1–7.

- Zheng, L., Su, Y., Wang, L. and Jiang, Z. (2009). Utilization of industrial waste products as adsorbents for the removal of dyes. *Separation and Purification Technology*. 68: 244–24.
- Zhou, J. L. and Banks, C. J. (1991). The adsorption of humic acid fractions by fungal biomass. *Environmental Technology*. 12: 519–530.
- Zhou, W. and Zimmermann, W. (1993). Decolorization of industrial effluents containing reactive dyes by actinomycetes. *FEMS Microbiology Letters*. 107: 157–162.
- Zouari-Mechichi, H., Mechichi, T., Dhoui, A., Sayad, S., Martínez, A. T. and Martínez, M. J. (2006). Laccase purification and characterization from *Trametes trogii* isolated in Tunisia: Decolorization of textile dyes by the purified enzyme. *Enzyme and Microbial Technology*. 39: 141–148.