

IDENTIFICATION OF PROTEIN FROM *EURYCOMA LONGIFOLIA*

ROOT EXTRACT

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IDENTIFICATION OF PROTEIN FROM *EURYCOMA LONGIFOLIA*
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requirements for the award of the degree of
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Specially dedicated to my beloved umi, abah, siblings, supervisor and caring friend

Thank you for your support

“Jika mahu cemerlang, mesti sanggup menempuh semua halangan.
Halangan terbesar ialah MINDA kita sendiri, yang hanya sukakan
sesuatu yang selesa dan mudah.”

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ABSTRACT

This study was carried out to identify plant protein from the root samples of *Eurycoma longifolia* harvested from Pahang and Perak, Malaysia. Protein usually presents in small quantity which is only 0.001% in plant, therefore it is critical to determine the extraction method for high yield and good quality of protein for the subsequent process of protein identification spectrometrically. Four extraction methods, namely water, Triton X-100 (non-ionic detergent), phenol-SDS precipitation and TCA-acetone precipitation were investigated for plant protein extraction. The yield of protein extracted from the plant samples was determined using Bradford assay. Both water extracts (water and Triton X-100) methods contained slightly higher protein content (0.2-0.53 mg protein/ml crude protein) than the extracts of precipitation (phenol and TCA) methods (0.12-0.29 mg protein/ml crude protein). The water extraction method also produced the highest resolution of 15% polyacrylamide gel with six and five protein bands for *E. longifolia* Pahang and Perak, respectively. However, the number of protein bands decreased from five to three for the extraction method of Triton X-100, phenol-SDS and TCA-acetone, respectively. After trypsin digestion of the protein bands, the presence of protein was analyzed by liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS). The mass spectra matched with the databases showed that the addition of Triton X-100 could assist the extraction of mitochondrial protein, namely superoxide dismutase. Proteins that involved in energy metabolism such as phosphoenolpyruvate carboxykinase and plant protein inhibitor were also detected in the phenol-SDS buffer extraction because SDS acted as ionic detergent for cell lysis in this method. In line with previous studies, TCA-acetone did not exhibit clear gel image and subsequently less peptides were detected spectrometrically. In the present study, the addition of detergent (Triton X-100 and SDS) could enhance plant protein extraction from *E. longifolia* root from 46.7 to 72.7% w/w, but the use of TCA did not improve protein precipitation effectively.

ABSTRAK

Kajian ini dijalankan untuk mengenal pasti protein tumbuhan daripada sampel akar *Eurycoma longifolia* yang dituai dari Pahang dan Perak, Malaysia. Protein biasanya dalam kuantiti yang sedikit dalam tumbuhan, maka adalah penting untuk menentukan kaedah pengekstrakan bagi mendapatkan kandungan protein yang tinggi dan berkualiti supaya proses pengenalpastian protein dapat dilakukan secara spektrometrik. Empat kaedah pengekstrakan, iaitu air, Triton X-100 (detergen bukan ionik), pemendakan phenol-SDS dan pemendakan TCA-aseton telah dikaji untuk mengekstrak protein tumbuhan. Hasil protein daripada sampel tumbuhan telah ditentukan dengan menggunakan ujian Bradford. Hasilnya didapati pengekstrakan menggunakan air dapat mengekstrak hasil protein yang lebih tinggi (0.2-0.53 mg protein/ml protein mentah) daripada kedua-dua kaedah pemendakan (0.12-0.29 mg protein/ml protein mentah). Kaedah pengekstrakan air juga menghasilkan 15% poliakrilamida gel beresolusi tinggi dengan 6 dan 5 jalur protein bagi *E. longifolia* Pahang dan Perak masing-masing. Walau bagaimanapun, bilangan jalur protein menurun daripada 5 hingga 3 jalur bagi kaedah pengekstrakan Triton X-100, phenol-SDS dan TCA-aseton masing-masing. Selepas dicerna oleh trypsin, kehadiran protein dianalisis dengan menggunakan kaedah kromatografi cecair diintegrasikan dengan jisim spektrometri sejajar (LC-MS/MS). Spektra jisim dipadankan dengan pangkalan data dan didapati bahawa penambahan Triton X-100 membantu pengekstrakan protein mitokondria iaitu superosida dismutase. Protein terlibat dalam metabolisme tenaga seperti fosfenolpiruvate karboxykinase dan pemangkin protein dikesan dalam kaedah pemendakan phenol-SDS kerana SDS bertindak sebagai detergen ionik yang boleh memecahkan sel tumbuhan. Sebagaimana kajian sebelumnya, TCA-aseton tidak menunjukkan imej gel jelas dan kurang peptida dapat dikesan oleh spektrometer. Dalam kajian ini, penambahan detergen (Triton X-100 dan SDS) boleh meningkatkan proses pengekstrakan protein daripada tumbuhan sebanyak 46.7 hingga 72.7% w/w, tetapi penggunaan TCA tidak dapat memberikan hasil pemendakan protein secara berkesan.

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LIST OF ABBREVIATIONS

μl	-	Microliter
2DE	-	Two-Dimensional Electrophoresis
ACN	-	acetonitrile
AIDS	-	acquired immunodeficiency syndrome
APS	-	ammonium persulfate
ATP	-	Adenosine triphosphate
BCA	-	Bicinchoninic acid assay
BLAST	-	Basic Local Alignment Search Tool
BPB	-	bromophenol blue
BSA	-	bovine serum albumin
CBB	-	Coomassie Brilliant Blue
cm	-	centimeter
DNA	-	Deoxyribonucleic acid
DTT	-	dithiothreitol
EDTA	-	Ethylenediaminetetraacetic acid
ELISA	-	enzyme-linked immunosorbent assay
EPF	-	Epidermal Patterning Factor
FDA	-	US Food and Drug Administration
FRIM	-	Forest Research Institute Malaysia
FTIR	-	Fourier transform infrared spectroscopy
GC-MS	-	Gas chromatography-mass spectrometry
HCl	-	hydrogen chloride

HIV	-	human immunodeficiency virus
HPLC	-	High-Performance Liquid Chromatography
IAA	-	iodoacetamide
KCl	-	potassium chloride
kD	-	kilo Dalton
LC-MS/MS	-	Liquid Chromatography Coupled with Tandem Mass
MALDI-TOF	-	Matrix-assisted laser desorption/ionization - time-of-flight
mg	-	Miligram
ml	-	Mililiter
mM	-	Mili Molar
Mn	-	Manganese
MS	-	Mass Spectrometry
NCBI	-	National Center for Biotechnology Information
nm	-	nanometer
PCA	-	Principal component analysis
PMF	-	Peptide Mass Fingerprinting
pmol	-	picomoles
PMSF	-	phenylmethylsulfonyl fluoride
RALF	-	Rapid Alkalinization Factor; 49 amino acid residues
RIA	-	Radioimmunoassay
rpm	-	round per minute
SDS-PAGE	-	Sulphate Polyacrylamide Gel Electrophoresis
SELDI-MS	-	Surface Enhance Laser Desorption Ionization Mass Spectrometer
SPME	-	Solid Phase Microextraction
SRM	-	selected reaction monitoring
TCA	-	Trichloroacetic acid
TDIF	-	Tracheary Element Differentiation Inhibitory Factor
TEMED	-	tetramethylethylenediamine
TMV	-	Tobacco Mosaic Virus
TPD1	-	Tapetum Determinant 1
UV-Vis	-	Ultraviolet-visible

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CHAPTER 1

RESEARCH BACKGROUND

1.1 Introduction

Eurycoma longifolia or commonly known as Tongkat Ali in Malaysia is a tropical herbal plant found in several parts of South East Asia countries. It is also known as Payung Ali, Penawar Pahit, Setunjang Bumi, Bedara Pahit, Tongkat Baginda, Pokok Syurga, Tongkat Ali Hitam, Pokok Jelas and Jelaih. There are four common species of Tongkat Ali, namely *Eurycoma longifolia*, *Eurycoma apiculata*, *Polyathiabullata* and *Goniothalamus* sp. (Aziz *et al.*, 2003). Among these species, *Eurycoma longifolia* is commonly used in traditional medication for various diseases. Traditionally, *E. longifolia* is used for its aphrodisiac, anti-pyretic and anti-malarial effects as well as a general tonic (Kuo *et al.*, 2003). A decoction of its long woody tap root is taken orally. The benefits of the roots of *E. longifolia* include restoring energy and vitality, enhancing blood flow and functioning as herbal ingredient for women after child birth (Ismail *et al.*, 1999). This plant bears fruit after two and a half years of cultivation, while the root is generally taken to be processed after 4 years of cultivation (Athimulam *et al.*, 2006).

To our knowledge, plant protein from *E. longifolia* is seldom studied by researchers. It is believed that besides small metabolites, plant protein from the plant could be the bioactive constituent that contributed to the reported pharmacological properties (Chua *et al.*, 2011). The detection of bioactive peptide (4.3 kD) which had been reported to have aphrodisiac activity from *E. longifolia* (Asiah *et al.*, 2007) has encouraged more intensive research on protein or peptide in the plant. In the following year, another two plant proteins from *E. longifolia* with the molecular weight of 7.5 and 6.0 kD had been identified by Farouk *et al.* (2008). They reported the proteins showed antibacterial activity against human pathogenic bacteria. Antimicrobial peptides are ribosomally synthesised as natural antibiotics by almost all living organisms including bacteria, animals and plants. The plant still contains many proteins to be discovered, particularly for pharmacological applications.

1.2 Problem Statement

The hard cell walls of the plant have been reported to be the limitation factor for plant protein extraction and separation (Granier, 1988). Therefore, plant protein identification is a challenge since plant protein usually presents in small quantity. Plant proteins need to be extracted efficiently without degradation to ensure maximal yield of protein extraction. This study investigated the method of protein extraction that can give the highest number of proteins and the highest protein content. Usually, detergent is used to break the cell wall for proteins extraction. However, the type of detergent chosen must contribute to the lowest protein loss during extraction.

Recent technical advancement in sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) and mass spectrometry (MS) have made the effort of protein separation and identification to be faster and higher in accuracy and sensitivity. There are limited studies on proteins from *E. longifolia* from gel electrophoresis to LC-MS/MS technique. This is mainly due to the minute amount of plant protein. Fortunately, this technique only requires small amount of sample for analysis. Somehow, extra precaution in sampling handling such as protein alkylation and reduction must take into consideration before mass spectrometric analysis.

There are still many types of proteins need to be explored, mainly for pharmacological application. For instance, phytohormones which are used in intercellular responses for plant growth and development might be useful for drug development. Phytohormones involve in plant defense mechanism in response to wound signal transduction by pests (Lindsey *et al.*, 2002). Plant protein could also be used as a carrier such as plant non-specific lipid-transfer proteins in drug delivery (Cheng *et al.*, 2004). However, the number of plant protein in the database is lesser than protein from bacteria and animals (Kim *et al.* and Park *et al.*, 2002). Only 0.001% of the total 961,019 number of total protein sequence belongs to plant protein (National Center for Biotechnology Information (NCBI) database, assessed on 26th April 2014).

1.3 Objective

The objective of this study was to identify plant proteins from *Eurycoma longifolia* (Tongkat Ali) root extracts collected from Pahang and Perak in Malaysia.

1.4 Scope

In order to achieve the objective of this study, the scopes were:

1. To investigate the protein extraction methods for *E. longifolia* roots using four different extraction methods, namely water, Triton X-100 (non-ionic detergent), phenol-SDS buffer and TCA-acetone precipitation methods.
2. To fingerprint plant proteins extracted from *E. longifolia* roots using one dimensional gel electrophoresis and mass spectrometric approaches.

REFERENCES

- Abdul, R. A. S., Sim, Y. M. M., Md Shakaff, A.Y., Ahmad, M.N., Dahari, Z., Ismail, Z. and Hitan, M.S. (2004). A microcontroller-based taste sensing system for the verification of *Eurycoma longifolia*. *Sens Actuators B*, 101:191-198.
- Aebersold, R. and Mann, M. (2003). Mass spectrometry-based proteomics. *Nature*, 422, 198 – 207.
- Ang, H. H., and Ngai, T. H. (2001). Aphrodisiac evaluation in non-copulator male rats after chronic administration of *Eurycoma longifolia* Jack. *Fundam Clin Pharmacol*,15:265–8.
- Ang, H. H., and Sim, M. K. (1997). *Eurycoma longifolia* Jack enhances libido in sexually experienced male rats. *J Exp Anim Sci*,46:287–90.
- Ang, H. H., Hitotsuyanagi, Y., and Fukaya, H. (2002a). Quassinoids from *Eurycoma longifolia*. *Phytochemistry*, 59. 833-837.
- Ang, H. H., Lee, K. L. (2002b). Effect of *Eurycoma longifolia* Jack on orientation activities in middle-aged male rats. *Fundam Clinic Pharmacol*,16: 479–83.
- Ang, H. H., Lee, K. L., and Kiyoshi, M. (2004). Sexual arousal in sexually sluggish old male rats after oral administration of *Eurycoma longifolia* Jack. *J Basic Clin Physiol Pharmacol*,15:303–9.
- Ang, H.H., Chan, K.L., and Mak, J.W. (1995). Effect of 7-day daily replacement of culture medium containing *Eurycoma longifolia* Jack constituents on the Malaysian Plasmodium falciparum isolates. *J Ethnopharmacol*, 49:171–5.
- Asiah, O., Nurhanan, M.Y. and Mohd Ilham, A. (2007). Determination of bioactive peptide (4.3kDa) as an aphrodisiac marker in six Malaysian Plants. *Journal of Tropical Forest Science*, 19/1. 61-63.
- Athimulam, A., Kumaresan, S., FOO, D. C. Y., Sarmidi, M. R. and Aziz, R. A. (2006). Modelling and Optimization of *Eurycoma longifolia* Water Extract Production. *Food and Bioproducts Processing*, 84(C2): 139–149.

- Aziz, R. A., Sarmidi, M. R., Kumaresan, S., Taher, Z. M., and Foo, D. C. Y. (2003). Phytochemical processing: the next emerging field in chemical engineering-aspects and opportunities. *Jurnal Kejuruteraan Kimia Malaysia*, 3: 45–60.
- Bajuk, B. P., Zdovc, I., Smrekar, V., Križaj, I., Leonardi A., and Drobni M. (2011). Dermatophyte Trichophyton mentagrophytes Produces Cysteine Protease Inhibitor. *Acta Chim. Slov*, 33, 58, 33–40.
- Barney, L. B., Messina, L., Vidal, A., Peric, M., and Nascimento, O. R. (1998). Precision Relative Aggregation Number Determinations of SDS Micelles Using a Spin Probe. A Model of Micelle Surface Hydration. *J. Phys. Chem. B*, 102 (50): 10347–10358.
- Berg, J. M., Tymoczko, J. L., and Stryer, L. (2002). *Biochemistry*. (5th edition). New York: W H Freeman; Section 4.1, The Purification of Proteins Is an Essential First Step in Understanding Their Function. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK22410/>.
- Bollag, D. M., Rozycki, M. D., and Edelstein, S. J. (1996). Protein Methods. *Second Edition. A John Wiley & Sons, Inc., Publication*, 12, 1-172.
- Bona, E., Marsano, F., Cavaletto, M., and Berta, G. (2007). Proteomic characterization of copper stress response in *Cannabis sativa* roots. *Proteomics*. 7, 1121 – 1130.
- Bradford, M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* 72, 248-254.
- Canovas, F. M., Dumas-Gaudot, E., Recorbet, G., Jorin, J., Mock, H. P., and Rossignol, M. (2004). Plant proteome analysis. *Proteomics*. 4, 285 – 298.
- Carpentier, S. C., Witters, E., Laukens, K., Deckers, P., Swennen, R., and Panis, B. (2005). Preparation of protein extracts from recalcitrant plant tissues: an evaluation of different methods for two-dimensional gel electrophoresis analysis. *Proteomics* 5, 2497-2507.
- Carpita, N. C, and Gibeaut, D. M. (1993). Structural models of primary cell walls in flowering plants: consistency of molecular structure with the physical properties of the walls during growth. *Plant Journal*, 3:1-30.
- Carvalho, A. O., and Gomes, V. M. (2009). Plant defensins—respects for the biological functions and biotechnological properties. *Peptides* 30, 1007–1020.

- Chan, K. L., Choo, C. Y., Abdullah, N. R., and Ismail, Z. (2004). Antiplasmodial studies of *Eurycoma longifolia* Jack using the lactate dehydrogenase assay of *Plasmodium falciparum*. *Journal of Ethnopharmacology*, 92: 223-227.
- Chan, K. L., Iitaka, Y., Noguchi, H., Sugiyama, H., Saito, I., and Sankawa, U. (1992). 6[α]-Hydroxyeurycomalactone, a quassinoid from *Eurycoma longifolia*. *Phytochemistry*, 31(12), 4295–4298.
- Chan, K. L., Lee, S. P., Sam, T. W., Tan, S. C., Noguchi, H., and Sankawa, U. (1991). 13 β , 18-Dihydroeurycomanol, a quassinoid from *Eurycoma longifolia*. *Phytochemistry*, 30(9), 3138–3141.
- Chan, K. L., Lee, S., Sam, T. W., and Han, B. H. (1989). A quassinoid glycoside from the roots of *Eurycoma longifolia*. *Phytochemistry*, 28:2857–9.
- Chan, K. L., Low, B. S., The, C. H., and Das, P. K. (2009). The effect of *Eurycoma longifolia* on sperm quality of male rats. *Nat Prod Commun*, 4: 1331–6.
- Cheng, C. S., Chen, M. N., Liu, Y. J., Huang, L. Y., Lin, K. F., and Lyu, P. C. (2004). Evaluation of plant non-specific lipid-transfer proteins for potential applications in drug delivery. *Enzyme and Microbial Technology*, 35: 532-539.
- Choo, C. Y., and Chan, K. L. (2002). High performance liquid in chromatography analysis of canthinone alkaloids from *Eurycoma longifolia*. *Planta Med*, 68:382-384.
- Christer, E., and Monica, N. (2003). Protein Extraction From Frozen Brain Samples. *Karolinska Institutet, Cancer Center Karolinska, Department of Oncology - Pathology*, R8:05, 171 76 Stockholm, Sweden
- Chua, L. S., Amin, N. A. M., Neo, J. C. H., Lee, T. H., Lee, C. T., Sarmidi, M. R., and Aziz, R. A. (2011). LC–MS/MS based metabolites of *Eurycoma longifolia* (Tongkat Ali) in Malaysia (Perak and Pahang). *Journal of Chromatography B*, 879 3909– 3919.
- Chua, L. S., Nurulaini, A. R., Bustanur, R. and Chew, T. L. (2012). Plant proteins, minerals and trace elements of *Eurycoma longifolia* (Tongkat Ali). *Natural Product Research*, 1–5.
- Covey, P. A., Subbaiah, C. C., Parsons, R. L., Pearce, G., Lay, F. T., and Anderson, M. A. (2010). A pollen-specific RALF from tomato that regulates pollen tube elongation. *Plant Physiol*, 153: 703–715.

- Daisuke, T., Shenkui, L., and Tetsuo, T. (2011). A rapid chemical method for lysing *Arabidopsis* cells for protein analysis. *Plant Methods*, 7:22 doi:10.1186/1746-4811-7-22.
- Darise, M., Kohda, H., Mizutani, K., and Tanaka, O. (1982). Eurycomanone and eurycomanol, quassinoids from the roots of *Eurycoma longifolia*. *Phytochemistry*, 21:2091-2093.
- Dominique, M., and Alain, A. (1995). Application to plant proteins of gel electrophoretic methods. *Journal of Chromatography A*, 698: 263-279.
- Doustjalali, S. R., Marzalina, M., and Nor Datiakma, M. A. (2005). A Gel-Based Proteomic Kit to Screen the Quality of Water-soluble Root Extracts of *Eurycoma longifolia*. *Journal of Tropical Forest Science*, 17(3): 479—480.
- Eichel, J., Gonzhez, J. C., Hotze, M., Matthews R. G., and Schroder, J. (1995). Vitamin-B12-independent methionine synthase from a higher plant (*Catharanthus roseus*) Molecular characterization, regulation, heterologous expression, and enzyme properties. *Eur. J. Biochem*, 230, 1053-1058.
- Farouk, A. E. A., Mohd Nawawi, M. N., and Hassan, S. (2008). Antibacterial peptides from *Eurycoma longifolia* (Tongkat Ali) and *Labisia pumila* (Kacip Fatimah) leaves in Malaysia. *Scientia Bruneiana*, 9: 55-64.
- Farouk, A. E., and Benafri, A. (2007). Antibacterial activity of *Eurycoma longifolia* Jack. A Malaysian medicinal plant. *Saudi Med J*, 28:1422—4.
- Gills, J. J., LoPiccolo, J., Tsurutani, J., Shoemaker, R. H., Best, C. J. M., Abu-Asab, M., Borojerdi, J., Warfel, N. A., Gardner, E. R., Danish, M., Hollander, M. C., Kawabata, S., Tsokos, M., Figg, W. D., Steeg, P.S., and Dennis, P. A. (2007). Nelfinavir, A Lead HIV Protease Inhibitor, Is a Broad-Spectrum, Anticancer Agent that Induces Endoplasmic Reticulum Stress, Autophagy, and Apoptosis In vitro and In vivo. *Clinical Cancer Research* 13, (17): 5183—94.
- Gimlette, J. D., and Thomson, H.W. (1977). *A dictionary of Malayan medicine*. Kuala Lumpur: Oxford University Press, 183 pp.
- Goh, S. H., Chuah, C. H., Mok, J. S. L., and Soepadmo, E. (1995). Malaysian Medicinal Plants for the Treatment of Cardiovascular Disease. *Pelanduk Publication*, Kuala Lumpur, Malaysia.
- Goreja, W. G. (2004). *Tongkat Ali: The Tree that Cures a Hundred Diseases*. vol. 2, Amazing Herb Press, New York, NY, USA.

- Gorg, A., Weiss, W., and Dunn, M. J. (2004). Current two-dimensional electrophoresis technology for proteomics. *Proteomics*, 4, 3665 – 3685.
- Granier, F. (1988). Extraction of plant proteins for two-dimensional electrophoresis. *Electrophoresis*, 9:712.
- Gygi, S. P., Corthals, G. L., Zhang, Y., Rochon, Y., and Aebersold, R. (2000). Evaluation of two-dimensional gel electrophoresis-based proteome analysis technology. *Proc. Natl. Acad. Sci. USA*, 97, 9390 – 9395.
- Hara, K., Kajita, R., Torii, K. U., Bergmann, D. C., and Kakimoto, T. (2007). The secretory peptide gene EPF1 enforces the stomatal one-cell-spacing rule. *Genes Dev*, 21: 1720–1725.
- Hara, K., Yokoo, T., Kajita, R., Onishi, T., Yahata, S., and Peterson, K. M. (2009). Epidermal cell density is autoregulated via a secretory peptide, Epidermal Patterning Factor 2 in *Arabidopsis* leaves. *Plant Cell Physiol*, 50: 1019–1031.
- Harlow, E., and Lane, D. (1999). *Immunoglobulins; Immunochemistry; Laboratory manuals*. Cold Spring Harbor Laboratory Press (Cold Spring Harbor, N.Y.). Book (ISBN 0879695447)
- Harrison, P.A. and Black, C.C. (1982). Two-dimensional Electrophoretic Mapping of Proteins of Bundle Sheath and Mesophyll Cells of the C4 Grass *Digitaria sanguinalis* (L.) Scop. (Crabgrass). *Plant Physiol*. 70:1359.
- Higashiyama, T. (2010). Peptide signaling in pollen–pistil interactions. *Plant Cell Physiol*, 51: 177–189.
- Hirakawa, Y., Kondo, Y., and Fukuda, H. (2010a). TDIF peptide signaling regulates vascular stem cell proliferation via the WOX4 homeobox gene in *Arabidopsis*. *Plant Cell*, 22: 2618–2629.
- Hirakawa, Y., Kondo, Y., and Fukuda, H. (2010b). Establishment and maintenance of vascular cell communities through local signaling. *Curr. Opin. Plant Biol*.
- Hout, S., Chea, A., Bunb, S. S., Elias, R., Gasquet, M., and Timon, D. P. (2006). Screening of selected indigenous plants of Cambodia for antiplasmodial activity. *J Ethnopharmacol*, 107:12–8.
- Hunt, L., and Gray, J. E. (2009). The signaling peptide EPF2 controls asymmetric cell divisions during stomatal development. *Curr. Biol*, 19: 864–869.
- Huntington, J. A. and Stein, P. E. (2001). Structure and properties of ovalbumin. *Journal of Chromatography*, B756, 189-198.

- Hurkman, W. J., and Tanaka, C. K. (1986). Solubilization of plant membrane proteins for analysis by two-dimensional gel electrophoresis. *Plant Physiology*, 81: 802-806.
- Husen, R., Hawariah, A., Pihie, L., and Nallappan, M. (2004). Screening for antihyperglycaemic activity in several local herbs of Malaysia. *J Ethnopharmacol*, 95:205–8.
- Isaacson, T., Damasceno, C. M. B., Saravanan, R. S., He, Y. H., Catal, C., Saladi, M., and Rose, J. K. C. (2006). Sample extraction techniques for enhance proteomics analysis of plant tissues. *Nature Protocol*, 1, 769 – 774.
- Ismail, Z., Ismail, N. and Lassa, J. (1999). *Malaysian Herbal Monograph*. (Malaysian Monograph Committee, Kuala Lumpur).
- Ito, Y., Nakanomyo, I., Motose, H., Iwamoto, K., Sawa, S., and Dohmae, N. (2006). Dodeca-CLE peptides as suppressors of plant stem cell differentiation. *Science*, 313: 842–845.
- Itokawa, H., Kishi, E., Morita, H., and Takeya, K. (1992). Cytotoxic quassinoids and tirucallane-type triterpenes from the wood of *Eurycoma longifolia*. *Chemical and Pharmaceutical Bulletin*, 40:1053.
- Itokawa, H., Kishi, E., Morita, H., Takeya, K., and Iitaka, Y. (1991). Eurylene, a new squalene-type triterpene from *Eurycoma longifolia*. *Tetrahedron Letters*, 32(15), 1803–1804.
- Jaquinod, M., Villiers, F., Kieffer-Jaquinod, S., Hugouvieux, V., Bruley, C., Garin, J., and Bourguignon, J. (2007). A proteomics dissection of *Arabidopsis thaliana* vacuoles isolated from cell culture. *Mol. Cell Proteomics*, 6, 394 – 412.
- Jefferson, R. A., Tony, A. K., and Michael, W. B. (1987). GUS fusions: β -glucuronidase as a sensitive and versatile gene fusion marker in higher plants. *The EMBO Journal*. vol.6 no. 13 pp.3901 -3907.
- Jun, J.H., Fiume, E. and Fletcher, J.C. (2008). The CLE family of plant polypeptide signaling molecules. *Cell. Mol. Life Sci*, 65: 743–755.
- Kardono, L. B. S., Angerhofer, C. K., Tsauri, S., Padmawinata, K., Pezzuto, L. M., and Kinghorn, A. D. J. (1991a). Studies on Indonesian medicinal plants. IV Cytotoxic and antimalarial constituents of the roots of *Eurycoma longifolia*. *Journal of Natural Products*, 54(5), 1360–1367.

- Kardono, L. B. S., Angerhofer, C. K., Tsauri, S., Padmawinata, K., Pezzuto, L. M., and Kinghorn, A. D. J. (1991b). Cytotoxic and antimalarial constituents of the roots of *Eurycoma longifolia*. *Journal of Natural Products*, 54:1360–7.
- Khairi, N., Aisha, A. F.A., and Ismail, Z.. (2013). Extraction, Preliminary Chemical Characterization, and Antioxidant Properties of Polysaccharides From *Eurycoma Longifolia* Jack. *Bentham Open*, DOI: 10.2174/2210289201304010181.
- Kim, S. I., Kim, S. J., Nam, M. H., and Kim, S. (2002). Proteome analysis of aniline-induced proteins in *Acinetobacter lwoffii* K24. *Current Microbiol.*, 44, 61–66.
- Kondo, T., Kajita, R., Miyazaki, A., Hokoyama, M., and Nakamura-Miura, T. (2010). Stomatal density is controlled by a mesophyll-derived signaling molecule. *Plant Cell Physiol*, 51: 1–8.
- Kondo, Y., Hirakawa, Y., Kieber, J.J., and Fukuda, H. (2011). CLE peptides can negatively regulate protoxylem vessel formation via cytokinin signaling. *Plant Cell Physiol*, 52: 37–48.
- Kuo, P. C., Damu, A. G., Lee, K. H. and Wu, T. S. (2003). Characterization of the water soluble fraction from the root extract of *Eurycoma longifolia*. *Pharmaceutical Society of Republic of China, Taipei, Taiwan, Province de Chine*, vol. 55, pp. 257-265.
- Kuo, P. C., Damu, A. G., Lee, K. H. and Wu, T. S. (2004). Cytotoxic and antimalarial constituents from the roots of *Eurycoma longifolia*. *Bioorganic & Medicinal Chemistry*, 12: 537-544.
- Kuo, P. C., Shi, L. S., Damu, A. G., Su, C. R., Huang, C. H., and Ke, C. H. (2003). Cytotoxic and antimalarial β -carboline alkaloids from the roots of *Eurycoma longifolia*. *J Nat Prod*, 66:1324–7.
- Kuo, P. C., Su, C. R., Damu, A. G., and Wu, T. S. (2004). Eurycomalin A, a new dimeric dihydrobenzofuran from *Eurycoma longifolia*. *Heterocycles*, 63(9), 2123–2129.
- Kuramata, M., Masuya, S., Takahashi, Y., Kitagawa, E., Inoue, C., Ishikawa, S. (2009). Novel cysteine-rich peptides from *Digitaria ciliaris* and *Oryza sativa* enhance tolerance to cadmium by limiting its cellular accumulation. *Plant Cell Physiol*, 50: 106–117.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227, 680 – 685.

- Leibfried, A., To, J. P., Busch, W., Stehling, S., Kehle, A., and Demar, M. (2005). WUSCHEL controls meristem function by direct regulation of cytokinin-inducible response regulators. *Nature*, 438: 1172–1175.
- Leila, F., Muhammad, I., Rafael, F. P. L., Hervé, C., and Elisabeth, J. (2006). Evaluation of cell wall preparations for proteomics: a new procedure for purifying cell walls from *Arabidopsis hypocotyls*. *Plant Methods*, 2:10 doi:10.1186/1746-4811-2-10.
- Liang, X. Q., Luo, M., Holbrook, C. C. and Guo, B. Z. (2006). Storage protein profiles in Spanish and runner market type peanuts and potential markers. *BMC Plant Biol*, 6, 24 – 32.
- Lin, L. C., Peng, C. Y., Wang, H. S., Lee, K. W., and Wang, P. S. (2001). Reinvestigation of the chemical constituents of *Eurycoma longifolia*. *Chinese Pharmaceutical Journal*, 53(2), 97–106.
- Lindsey, K., Casson, S., and Chilley, P. (2002). Peptides: new signaling molecules in plants. *Trends in Plant Science*, 7: 78-83.
- Magdeldin, S. (2012). *Gel Electrophoresis – Principles and Basics*. InTech Janeza Trdine 9, 978-953-51-0458-2.
- Malik, N. A. and Bradford, J. M. J. (2005). *Food Agric. Environ.*, 3, 246 –248.
- Manfred, R. (2012). LC-MS/MS for protein and peptide quantification in clinical chemistry. *Elsevier B.V. Journal of Chromatography B*, 59-67.
- Mann, M., and Pandey, A. (2001). Use of mass spectrometry-derived data to annotate nucleotide and protein sequence databases. *Trends Biochem Sci*, 26: 54–61.
- Mary, J.. (2013). Detergents: Triton X-100, Tween-20, and More. *Princeton*, 3:163.
- Maserti, B. E., Della Croce, C. M., Luro, F., Morillon, R., Cini, M., and Caltavuturo, L. (2007). A general method for the extraction of citrus leaf proteins and separation by 2D electrophoresis: a follow up. *Journal of Chromatography B*, 849, 351–356.
- Mathesius, U., Keijzers, G., Natera, S. H., Weinman, J., Djordjevic, M., and Rolfe, B. (2001). Establishment of a root proteome reference map for the model legume *Medicago truncatula* using the expressed sequence tag database for peptide mass fingerprinting. *Proteomics*, 1, 1424 – 40.
- Matsubayashi, Y. (2011). Post-translational modifications in secreted peptide hormones in plants. *Plant Cell Physiol*, 52: 5–13.

- Matsubayashi, Y., and Sakagami, Y. (1996). Phytosulfokine, sulfated peptides that induce the proliferation of single mesophyll cells of *Asparagus officinalis* L. *Proc. Natl Acad. Sci. USA*, 93: 7623–7627.
- Matsuzaki, Y., Ogawa-Ohnishi, M., Mori, A., and Matsubayashi, Y. (2010). Secreted peptide signals required for maintenance of root stem cell niche in *Arabidopsis*. *Science*, 329: 1065–1067.
- Mohamad, M., Ali, M. W., and Ahmad, A. (2010). Modelling Extraction of Major Phytochemical Components from *Eurycoma longifolia*. *Journal of Applied Sciences*, 10(21): 2572-2577.
- Mohd, R. M. A. R., Sow, A., Noor, R. A., Mohd Ilham, A., and Zakiah, I. (2007). *Eurycoma longifolia* extract-artemisinin combination: parasitemia suppression of Plasmodium yoelii-infected mice. *Trop Biomed*, 24: 111–8.
- Montchamp, J. C., Piehler, L. T., and Frost, J. W. (1992). Diastereoselection and in vivo inhibition of 3-dehydroquinate synthase. *J. Am. Chem. Soc.*, 114 (12), pp 4453–4459 DOI: 10.1021/ja00038a002.
- Morita, H., Kishi, E., Takeya, K., and Itokawa, H. (1992). Biphenylneolignans from wood of *Eurycoma longifolia*. *Phytochemistry*, 31(11), 3993–3995.
- Morita, H., Kishi, E., Takeya, K., Itokawa, H., and Iitaka, Y. (1993). Highly oxygenated quassinoids from *Eurycoma longifolia*. *Phytochemistry*, 33(3), 691–696.
- Morita, H., Kishi, E., Takeya, K., Itokawa, H., and Iitaka, Y. (1993). Squalene derivatives from *Eurycoma longifolia*. *Phytochemistry*, 34(3), 765–771.
- Morita, H., Kishi, E., Takeya, K., Itokawa, H., and Tanaka, O. (1990). New quassinoids from the roots of *Eurycoma longifolia*. *Chemistry Letters*, 19, 749–752.
- Mukerjee, P., and Mysels, K. J. (1971). Critical Micelle Concentration of Aqueous Surfactant Systems. *US. Government Printing Office, Washington, D.C. NSRDS-NBS*, 36.
- Nazrun, A. S., Firdaus, M., Tajul, A. A. S., Norliza, M., Norazlina, M., and Ima, N. S. (2011). The anti-osteoporotic effect of *Eurycoma longifolia* in aged orchidectomised rat model. *The Aging Male*, vol. 14, no. 3, pp. 150–154.
- Newman, D. J., Cragg, G. M., and Snader, K. M. (2003). Natural products as sources of new drugs over the period. *J Nat Prod*, 66(7):1022-1037.

- Norris, J. L., Porter, N. A. and Caprioli, R. M. (2003). Mass Spectrometry of Intracellular and Membrane Proteins Using Cleavable Detergents. *Anal Chem*, 6642-6647.
- Nurhanan, M. Y., Asiah, O., Rafedah, A., Mohd Ilham, A., Anee Suryani, S. and Mohd Radzi, A. (2004). Protein and chemical fingerprints of different plant parts of *E. longifolia*. *Poster presented at the 3rd MMBPP Symposium, 27–28 July 2004, Kuala Lumpur*.
- Nurhanan, M. Y., Hawariah, L. P. A., Ilham, A. M., and Shukri, M. A. M. (2005). Cytotoxic effects of the root extracts of *Eurycoma longifolia* Jack. *Phytother Res*, 19: 994–6.
- Ohyama, K., Ogawa, M., and Matsubayashi, Y. (2008). Identification of a biologically active, small, secreted peptide in *Arabidopsis* by in silico gene screening, followed by LC-MS-based structure analysis. *Plant J*, 55: 152–160.
- Ohyama, K., Shinohara, H., Ogawa-Ohnishi, M., and Matsubayashi, Y. (2009). A glycopeptide regulating stem cell fate in *Arabidopsis thaliana*. *Nat. Chem. Biol*, 5: 578–580.
- Okamoto, S., Ohnishi, E., Sato, S., Takahashi, H., Nakazono, M., and Tabata, S. (2009). Nod factor/nitrate-induced CLE genes that drive HAR1-mediated systemic regulation of nodulation. *Plant Cell Physiol*, 50: 67–77.
- Okuda, S., Tsutsui, H., Shiina, K., Sprunck, S., Takeuchi, H., Yu, R. (2009). Defensin-like polypeptide LUREs are pollen tube attractants secreted from synergid cells. *Nature*, 458: 357–361.
- Osakabe, Y., Arinaga, N., Umezawa, T., Katsura, S., Nagamachi, K., Tanaka, H., Ohiraki, H., Yamada, K., Seo, S. U., Abo, M., Yoshimura, E., Shinozaki, K., and Yamaguchi-Shinozaki, K. (2013). Osmotic stress responses and plant growth controlled by potassium transporters in *Arabidopsis*. *Plant Cell*, 25(2):609-24. doi: 10.1105/tpc.112.105700.
- Osman, A., Jordan, B., Lessard, P. A., Muhammad, N., Haron, M. R., Riffin, N. M., Sinskey, A. J., Rha, C. K., and Houseman, D.E. (2003). Genetic diversity of *Eurycoma longifolia* inferred from single nucleotide polymorphisms. *Plant Physiol*, 131:1294-1301.
- Park, K. S., Kim, H., Kim, N.G., and Cho, S. Y. (2002). Proteomic analysis and molecular characterization of tissue ferritin light chain in hepatocellular carcinoma. *Hepatology*, 35, 1459–1466.

- Peltier, J., Ytterberg, J., Liberles, D. A., Roepstorff, P., and Van, W. K. J. (2001). Identification of a 350 kDa ClpP protease complex with 10 different Clp isoforms in chloroplasts of *Arabidopsis thaliana*. *J Biol Chem*, 276: 16318–16327.
- Prive, G. G. (2007). Detergents for stabilization and crystallization of membrane proteins. *Elsevier Inc, Method* 41. 388-397.
- Raharjo, T. J., Widjaja, I., Roytrakul, S. and Verpoorteb, R. J. (2004). Comparative proteomics of *Cannabis sativa* plant tissues. *Biomol. Tech.*, 15, 97 – 106.
- Rao, N. M., Rao H. N. and Pattabiraman, T. N. (1983). Enzyme inhibitors from plants. Isolation and characterization of a protease inhibitor from arrow root (*Maranta arundinaceae*) tuber. *J. Biosci.*, Vol.5, Number 1, pp. 21—33.
- Rio, D. C., Manuel, A. J., Gregory J. H., and Timothy W. N. (2010). Purification of RNA by SDS Solubilization and Phenol Extraction. *Spring Harb Protoc.* doi:10.1101/pdb.prot5438.
- Rodney, F. B. (2012). *Biochemistry Laboratory: Modern Theory and Techniques, 2/E*. Hope College: Prentice Hall.
- Rodrigues, E. P., Torres, A. R., Batista, J. S. D. S., Huergo, L. And Hungria, M. (2012). A simple, economical and reproducible protein extraction protocol for proteomics studies of soybean roots. *Genetics and Molecular Biology*, 35, 1 9suppl0, 348-352.
- Rowley, A., Choudhary, J. S., Marzioch, M., Ward, M. A., Weir, M., Solari, R. C. and Blackstock, W. P. (2000). Applications of protein mass spectrometry in cell biology. *Elsevier*. Volume 20, Issue 4: Pages 383–397.
- Schmidt, A.C., Steier, S., & Otto, M. (2009). Evaluation of the arsenic binding capacity of plant proteins under conditions of protein extraction for gel electrophoretic analysis. *Talanta*, 77, 1830–1836.
- Scopes, R.K. (1982). Protein Purification. *Principles and Practice*. Springer-Verlag, New York, Pp. 185-193. 282 pages.
- Shafiqul Islam, A. K. M., Ismail, Z., Saad, B., Othman, A. R., Ahmad, M. N., and Md. Shakaff, A. Y. (2006). Correlation studies between electronic nose response and headspace volatiles of *Eurycoma longifolia* extracts. *Sens Actuators B*, 120:245–51.
- Shen, S. H., Jing, Y. X. and Kuang, T. Y. (2003). Proteomics approach to identify wound-response related proteins from rice leaf sheath. *Proteomics*, 3, 527 – 535.

- Sheoran, I. S., Ross, A. R. S., Olson, D. J. H. and Sawhney, V. K. (2009). Compatibility of plant protein extraction methods with mass spectrometry for proteome analysis. *Plant Science*, 176: 99-104.
- Shevchenko, A., Wilm, M., Vorm, O., and Mann, M. (1996) Mass spectrometric sequencing of proteins from silver stained polyacrylamide gels. *Anal. Chem.* 68, 850-858.
- Somaieh, A. Z., Xinghua, G., Gholamhassan, A. and Ernst, L. (2011). Optimization and application of microwave-assisted acid hydrolysis for rapid quantification of protein oxidation markers using LC-MS. *Elsevier B.V. Talanta*, 85. 1835-1841.
- Sugano, S. S., Shimada, T., Imai, Y., Okawa, K., Tamai, A., and Mori, M. (2010). Stomagen positively regulates stomatal density in *Arabidopsis*. *Nature*, 463: 241–244.
- Tada, H., Yasuda, F., Otani, K., Doteuchi, M., Ishihara, Y., and Shiro, M. (1991). New antiulcer quassinoids from *Eurycoma longifolia*. *Eur J Med Chem*, 26:345–9.
- Takashi, Y. and Isobe, H. (2008). Bottom-up synthesis of finite models of helical (n,m)-single-wall carbon nanotubes. *Nature Communications*, Article number:492 doi:10.1038/ncomms1505.
- Takayama, S., Shimosato, H., Shiba, H., Funato, M., Che, F. S., and Watanabe, M. (2001). Direct ligand–receptor complex interaction controls Brassica self-incompatibility. *Nature*, 413: 534–538.
- Tambi, M. I. (2002). Glycoprotein water-soluble extract of *Eurycoma longifolia* Jack as a health supplement in management of Health aging in aged men. *Proceedings of the 3rd World Congress on the Aging Male*, B. Lunenfeld, Ed., p. 6.
- Tambi, M. I. (2005). Standardized water soluble extract of *Eurycoma longifolia* (LJ100) on men's health. *International Journal of Andrology*, vol. 28.
- Tatjana, P., Martin, W., and Ralf, O. (2000). Identification of low-density Triton X-100-insoluble plasma membrane microdomains in higher plants. *European Journal of Biochemistry*. Volume 267, Issue 24, pages 6989–6995.
- Terrie, R. (2013). Advanced Molecular Biology. Bio 480/580. 3028D Biosciences. Laboratory Information.
- Thoi, L. V. and Suong, N. N. (1970). Constituents of *Eurycoma longifolia* Jack. *J Org Chem*, 35:1104-1109.

- Turro, N. J. and Yekta, A. (1978). Luminescent Probes for Detergent Solutions. A Simple Procedure for Determination of the Mean Aggregation Number of Micelles. *J. Am. Chem. Soc.*, 100,5951.
- Vyetrogon, K., Tebbji, F., Olson, D. J. H., Ross, A. R. and Matton, D. P. (2007). A comparative proteome and phosphoproteome analysis of differentially regulated proteins during fertilization in the self-incompatible species *Solanum chacoense* Bitt. *Proteomics*, 7, 232 – 247.
- Wang, W., Scali, M., Vignani, R., Spadafora, A., Sensi, E., Mazzuca, S., and Cresti, M. (2003). Protein extraction for two-dimensional electrophoresis from olive leaf, a plant tissue containing high levels of interfering compounds. *Electrophoresis.*, 24, 2369 – 2375.
- Weber, K. and Osborn, M. (1975). *The Proteins*. (Edition 3). H. Neurath and R. L. Hills, eds. Academic Press, vol. 1, p. 179.
- Wessel, D. and Flugge, U. I. (1984). A method for the quantitative recovery of protein in dilute solution in the presence of detergents and lipids. *Analytical Biochemistry*, Volume 138, Issue 1, April 1984, Pages 141–143.
- Xiaoying, Y., Donald, J. J. J., Ramin, M. H., Zhen, X., Zhaojing, M., Robert, G. U., Haleem, J. I., Timothy, D. V. and Josip, B. (2009). Optimization of protein solubilization for the analysis of the CD14 human monocyte membrane proteome using LC-MS/MS. *ScienceDirect. Journal of Proteomics*, 73: 112-122.
- Xie, H., Pan, S., Liu, S., Ye, K. and Huo, K. (2007). A novel method of protein extraction from *perennial Bupleurum* root for 2-DE. *Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim*, 28,871-875.
- Xingshen, L., Tingting, B., Yunfeng, L., Xiaolei, R., and Huaping, L. (2013). Proteomic analysis of *Fusarium oxysporum* f. sp. *Cubense* tropical race 4-inoculated response to *Fusarium* wilts in the banana root cells. *Proteome Sci*, 11: 41.
- Yang, S. L., Xie, L. F., Mao, H. Z., Puah, C. S., Yang, W. C., and Jiang, L. (2003). TAPETUM DETERMINANT1 is required for cell specialization in the *Arabidopsis* anther. *Plant Cell*, 15: 2792–2804.
- Yeboah, N. A., Arahira, M., Nong, V. H., Zhang, D., Kadokura, K., Watanabe, A., and Fukazawa, C. (1998). A class III acidic endochitinase is specifically expressed in the developing seeds of soybean (*Glycine max* [L.]. *Plant Molecular Biology*. Volume 36, Issue 3, pp 407-415.

- Yusmazura, Z., Asmah, R., Azimahtol, H. L. P., Noor, R. A., and Peter, J. H. (2009). *Eurycomanone* induce apoptosis in *HepG2* cells via up-regulation of *p53*. *BioMed Central Ltd*, 9:16.
- Zarembinski, T. I., Hung, L.W., Dieckmann, H. J. M., Kim, K. K., Yokota, H., Kim, R. and Kim, S. H. (1998). Structure-based assignment of the biochemical function of a hypothetical protein: A test case of structural geno. *Proc. Natl. Acad. Sci. USA*, Vol. 95, pp. 15189–15193.
- Zarkadas, C. G., Gagnon, C., Poysa, V., Khanizadeh, S., Cober, E. R., Chang, V., and Gleddie, S. (2007). Protein quality and identification of the storage protein subunits of tofu and null soybean genotypes, using amino acid analysis, one- and two-dimensional gel electrophoresis, and tandem mass spectrometry. *Food Research International*, 40, 111–128.
- Zhou, D., Lobo, Y. A., Batista, I. F. C., Marques-Porto, R., Gustchina, A., Oliva, M. L. V. and Wlodawer, A. (2013). Crystal Structures of a Plant Trypsin Inhibitor from *Enterolobium contortisiliquum* (*EcTI*) and of Its Complex with Bovine Trypsin. *Volume 8 .Issue 4*, e62252.
- Zhu, J. M., Chen, S. X., Alvarez, S., Asirvatham, V. S., Schachtman, D. P., Wu, Y. J. and Sharp, R. E. (2006). Cell wall proteome in the maize primary root elongation zone. I. Extraction and identification of water-soluble and lightly ionically bound proteins. *Plant Physiol*, 140, 311 – 325.
- Zielinski, R. E. (1998). Calmodulin and Calmodulin-binding Proteins in Plants. *Annual Review of Plant Physiology and Plant Molecular Biology*, Vol. 49: 697-725 DOI: 10.1146/annurev.arplant.49.1.697.