ESSENTIAL OILS, PHYTOCHEMICALS AND BIOACTIVITY STUDIES OF Curcuma aeruginosa Aff. AND Kaempferia rotunda Linn.

NUR ADILLA CHE JAMALLUDDIN

UNIVERSITI TEKNOLOGI MALAYSIA

ESSENTIAL OILS, PHYTOCHEMICALS AND BIOACTIVITY STUDIES OF *Curcuma aeruginosa* Aff. AND *Kaempferia rotunda* Linn.

NUR ADILLA CHE JAMALLUDDIN

A thesis submitted in fulfillment of the requirements of the award of the degree of Master of Science (Chemistry)

> Faculty of Science Universiti Teknologi Malaysia

> > MAY 2014

To my beloved mother and father

ACKNOWLEDGEMENTS

First of all, thanks to Almighty Allah for His blessing that I can finish up this study with contribution from others. I would like to express my deepest appreciation to my supervisors, Associate Professor Dr Farediah Ahmad and my co-supervisor Prof. Dr. Hasnah Mohd Sirat for their guidance, motivation and precious advice during the completion of this thesis.

I would like to express my gratitude towards all the laboratory assistants who have helped me in running the GC, GC-MS, NMR and IR. Also not forget, the entire lecturers in Faculty of Science especially in the Chemistry Department. Their contributions in my thesis and lab work were very important for me to finish this study.

Sincerely, to all my laboratory mate and friends who had guide me in the laboratory work, gave me motivation and encouraged me during my hard time, a million of thanks. Thanks for all of your support.

Special gratitude to my beloved parents, brother and sister for giving me their love, support and encouragement.

ABSTRACT

The essential oils, phytochemicals and bioactivities of Curcuma aeruginosa Aff., known as temu hitam and *Kaempferia rotunda* Linn. or kunvit putih have been studied. Hydrodistillation of the fresh rhizomes of C. aeruginosa and K. rotunda gave 77.1% and 89.3% oils respectively. These oils were analyzed by GC and GC-MS. The chemical compositions were identified by comparison of the mass spectral data of Wiley Library and Kovats Indices with literature values. A total of 42 components were identified from C. aeruginosa with epicurzerenone (19.47%) as the major constituent followed by 1,8-cineole (15.89%), trans-\beta-farnesene (9.75%), βelemene (6.61%) and camphor (5.84%). K. rotunda essential oil was found to have 33 components with high concentration of benzyl benzoate (31.48%), bornyl acetate (5.56%), camphor (5.45%) and camphene (5.04%). Extractions by soxhlet apparatus were carried out on the dried samples to get the crude extracts. Fractionation and purification on the crude extracts of both species resulted in the isolation of sesquiterpenoids, cyclohexane oxide, esters and fatty acid. The structures of the isolated compounds were identified by spectroscopic techniques including IR and NMR (1D and 2D) spectroscopies and mass spectrometry. Seven sesquiterpenoids were isolated from C. aeruginosa and identified as curzerenone, furanodienone, germacrone, dehydrocurdione, curcumenone, curcumenol and curzeone. Four compounds were successfully isolated from K. rotunda and characterized as benzyl benzoate, crotepoxide, lignoceric acid and stigmasterol. The crude extracts, essential oils and several pure phytochemicals were screened for antibacterial and antityrosinase activities. Disc diffusion method followed by minimum inhibition concentration (MIC) and minimum bactericidal concentration (MBC) against Gram positive and Gram negative bacteria were used for antibacterial assay. All the essential oil, crude extracts and compounds of C. aeruginosa and K. rotunda exhibited weak to inactive activity against the entire tested microorganisms. Modified dopachrome method with L-DOPA as the substrate was chosen to screen the antityrosinase activity. Among all the tested samples, the essential oil of C. aeruginosa possessed the highest activity with 15.62% inhibition against the mushroom tyrosinase. This inhibition value was far lower compared to the standard kojic acid which showed 81.81% inhibition.

ABSTRAK

Kajian ke atas minyak pati, fitokimia dan bioaktiviti C. aeruginosa Aff., yang dikenal sebagai cekur hitam dan K. rotunda Linn. atau kunyit putih telah dijalankan. Penyulingan hidro terhadap rizom segar C. aeruginosa dan K. rotunda masingmasing telah memberikan 77.1% dan 89.3% minyak pati. Minyak ini dianalisis menggunakan GC dan GC-MS. Komposisi bahan kimia telah dikenalpasti melalui perbandingan data spektrum jisim daripada perpustakaan Wiley dan nilai Indeks Kovats daripada literatur. Sebanyak 42 komponen telah dikenalpasti daripada C. aeruginosa dengan epikurzerenon (19.47%) sebagai komponen utama diikuti oleh 1,8-sineol (15.89%), trans-β-farnesena (9.75%), β-elemena (6.61%) dan kamfor (5.84%). Minyak pati K. rotunda terdiri daripada 33 komponen dengan komponen utamanya benzil benzoat (31.48%), bornil asetat (5.56%), kamfor (5.45%) dan kamfena (5.04%). Pengestrakan soxhlet telah dijalankan ke atas rizom kering untuk mendapatkan ekstrak mentah. Pemeringkatan dan penulenan ke atas ekstrak mentah bagi kedua-dua spesies telah mengasingkan sebatian seskuiterpenoid, sikloheksana oksida, ester dan asid lemak. Struktur semua sebatian telah dikenalpasti menggunakan teknik spektroskopi termasuk spektroskopi IR dan NMR (1D dan 2D) serta spektrometri jisim. Tujuh sebatian seskuiterpenoid telah diasingkan daripada C. aeruginosa dan dikenalpasti sebagai kurzerenon, furanodienon, germakron, dehidrokurdion, kurkumenon, kurkumenol dan kurzeon. Empat sebatian telah berjaya diasingkan daripada K. rotunda dikenalpasti sebagai benzil benzoat, krotepoksida, asid lignoserik dan stigmasterol. Ekstrak mentah, minyak pati dan beberapa fitokimia tulen telah disaring untuk aktiviti antibakteria dan antitirosinase. Kaedah penyebaran cakera diikuti dengan kepekatan perencatan minimum (MIC) dan kepekatan bakterisida minimum (MBC) terhadap bakteria Gram positif dan Gram negatif telah digunakan untuk saringan antibakteria. Kesemua sampel minyak pati, ekstrak mentah dan fitokimia C. aeruginosa dan K. rotunda menunjukkan keaktifan yang lemah sehingga tidak aktif terhadap seluruh mikroorganisma yang diuji. Kaedah pengubahsuaian dopakrom dengan L-DOPA sebagai substrat dipilih sebagai kaedah untuk menguji aktiviti antitirosinase. Antara semua sampel yang diuji, minyak pati C. aeruginosa memiliki aktiviti tertinggi dengan 15.62% perencatan berbanding tirosinase cendawan. Nilai ini adalah jauh lebih rendah daripada piawai asid kojik dengan 81.81% perencatan.

TABLE OF CONTENTS

CHAPTER	TITLE		PAGE
	DECI	LARATION	ii
	DEDI	CATION	iii
	ACK	NOWLEDGEMENTS	iv
	ABST	TRACT	V
	ABST	TRAK	vi
	TABI	LE OF CONTENTS	vii
	LIST	OF TABLES	xi
	LIST	OF FIGURES	xii
	LIST	OF SCHEMES	xiii
	LIST	OF ABBREVIATIONS	xiv
	LIST	OF APPENDICES	xvi
1	INTR	ODUCTION	
1	1.1	Background of the Problem	1
	1.2	<i>Curcuma</i> Species: Botany, Distribution and	3
		Usage	-
	1.3	<i>Kaempferia</i> Species: Botany, Distribution and	5
		Usage	
	1.4	Statement of Problem	6
	1.5	Objectives of Study	7
	1.6	Scope of the Study	7
2	LITE	RATURE REVIEW	
	2.1	Essential oil of Curcuma Species	8
	2.2	Essential oil of Kaempferia Species	11

2.3	Phytochemical Studies of Curcuma	13
	Species	
2.4	Phytochemical Studies of Kaempferia	15
	Species	
2.5	Bioactivity Studies on Curcuma Species	22
2.6	Bioactivity Studies on Kaempferia Species	24

3 ESSENTIAL OILS OF *CURCUMA AERUGINOSA* Aff. AND *KAEMPFERIA ROTUNDA* Linn.

3.1	Chemical Compositions of the Essential Oil of	26
	Curcuma aeruginosa	
3.2	Chemical Compositions of the Essential Oil of	29
	Kaempferia rotunda	

4 PHYTOCHEMICAL AND BIOACTIVITY STUDIES OF *CURCUMA AERUGINOSA* Aff. AND *KAEMPFERIA ROTUNDA* Linn.

4.1	Phytochemical Study of the Rhizomes of		32
	Curcu	ma aeruginosa Aff.	
	4.1.1	Curzerenone (1)	32
	4.1.2	Furanodienone (37)	35
	4.1.3	Curcumenone (108)	37
	4.1.4	Curcumenol (6)	38
	4.1.5	Germacrone (41)	40
	4.1.6	Curzeone (109)	42
	4.1.7	Dehydrocurdione (110)	44
4.2	Phytoe	chemical Study of the Rhizomes of	46
	Kaem	pferia rotunda Linn.	
	4.2.1	Benzyl Benzoate (43)	47
	4.2.2	Crotepoxide (51)	48
	4.2.3	Lignoceric acid (111)	50

	4.2.4	Stigmasterol (112)	52
4.3	Bioact	ivity Studies on <i>Curcuma aeruginosa</i> Aff.	54
	and <i>Ka</i>	aempferia rotunda Linn.	
	4.3.1	Antimicrobial Activity	54
	4.3.2	Antityrosinase Activity	57
EXPE	RIMEN	TAL	
5.1	Genera	al Experimental Procedures	59
5.2	Plant N	Materials	60
5.3	Solven	nts and Chemicals	61
5.4	Essent	ial Oils Extraction and Analysis of Fresh	61
	Rhizor	mes of <i>Curcuma aeruginosa</i> and	
	Kaemp	oferia rotunda	
5.5	Extrac	tion and Isolation of Curcuma aeruginosa	62
	5.5.1	Curzerenone (1)	63
	5.5.2	Furanodienone (37)	63
	5.1.3	Curcumenone (108)	64
	5.1.4	Curcumenol (6)	64
	5.1.5	Germacrone (41)	65
	5.1.6	Curzeone (109)	65
	5.1.7	Dehydrocurdione (110)	66
5.6	Extrac	tion and Isolation of Kaempferia rotunda	66
	5.2.1	Benzyl Benzoate (43)	66
	5.2.2	Crotepoxide (51)	67
	5.2.3	Lignoceric acid (111)	67
	5.2.4	Stigmasterol (112)	68
5.7	Bioact	ivity Studies	68
	5.7.1	Antimicrobial Activity	69
		5.7.1.1 Tested Samples and	69
		Microorganisms	

ix

Appendic	es		90-164
REFERE	NCES		76
6	CONCLUSIO	ON AND RECOMMENDATION	74
		5.7.3.2 Tyrosinase Inhibition Assay	72
		5.7.3.1 Chemicals and Instruments	72
	5.7.2	Antityrosinase Activity	72
		Concentration (MBC)	
		5.7.1.5 Minimum Bactericidal	71
		Concentration (MIC)	
		5.7.1.4 Minimum Inhibition	71
		5.7.1.3 Disc Diffusion Method	69
		5.7.1.2 Samples Preparation	69

LIST OF TABLES

TABLE NO.

TITLE

2.1	Isolated Compounds from Kampferia Species and Its	24
	Biological Properties	
3.1	Essential Oil Compositions of Curcuma aeruginosa	28
3.2	Essential Oil Compositions of Kaempferia rotunda	30
4.1	1 H, 13 C NMR and COSY Data of Compound (1)	35
4.2	¹ H, ¹³ C NMR, COSY and HMBC Data of Compound	40
	(6)	
4.3	¹ H, ¹³ C NMR, COSY and HMBC Data of Compound	44
	(109)	
4.4	¹ H, ¹³ C NMR, COSY and HMBC Data of Compound	46
	(110)	
4.5	Antimicrobial Activity of the Essential Oils, Crude	55
	Extracts and Pure Compounds from C. aeruginosa and	
	K. rotunda	
4.6	Results of MIC and MBC of the Essential Oils	56
4.7	Tyrosinase Inhibition Activity of the Essential Oils,	58
	Crude Extract and Several Pure Compounds of C.	
	aeruginosa and K. rotunda	

LIST OF FIGURES

FIGURES NO.

TITLE

PAGE

5.1	Rhizomes of Curcuma aeruginosa	60
5.2	Rhizomes of Kaempferia rotunda	61
5.3	The Arrangement of Discs in the Agar Plate	70
5.4	The Arrangement in the 96-well Flat Bottom Plastic	73
	Microtiter Plate (Greinich)	

LIST OF SCHEMES

SCHEMES	NO.
----------------	-----

TITLE

PAGE

4.1	Fragmentation Pattern of Curzerenone (1)	34
4.2	Fragmentation Pattern of Germacrone (41)	42
4.3	Fragmentation Pattern of Benzyl Benzoate (27)	48
4.4	Fragmentation Pattern of Crotepoxide (44)	50
4.5	Fragmentation Pattern of Stigmasterol (112)	53

LIST OF ABBREVIATIONS

α	Alpha
β	Beta
br	Broad
CC	Column Chromatography
COSY	Correlation Spectroscopy
¹³ C	Carbon-13
CDCl ₃	Deuterated chloroform
CHCl ₃	Chloroform
cm ⁻¹	Per centimeter
δ	Chemical shift
d	Doublet
dd	Doublet of doublets
DEPT	Distortionless Enhancement by Polarization Transfer
DMSO	Dimethyl sulphoxide
DPPH	2,2-Diphenyl-1-picrylhydrazyl
EIMS	Electron Impact Mass Spectrometry
Et ₂ O	Diethyl ether
GC	Gas Chromatography
GC-MS	Gas Chromatography-Mass Spectrometry
$^{1}\mathrm{H}$	Proton
HMBC	Heteronuclear Multiple Bond Correlation
HMQC	Heteronuclear Multiple Quantum Coherence
Hz	Hertz
IR	Infrared
J	Coupling constant
KI	Kovats Indices
L	Liter

m	Multiplet
\mathbf{M}^+	Molecular ion
m.p.	Melting point
MBC	Minimum Bactericidal Concentration
MIC	Minimum Inhibitory Concentration
MS	Mass Spectrometry
<i>m/z</i> ,	Mass to charge ion
$MgSO_4$	Magnesium sulphate
NA	Nutrient agar
NB	Nutrient broth
NMR	Nuclear Magnetic Resonance
ppm	Parts per million
\mathbf{R}_{f}	Retention factor
S	Singlet
SiO ₂	Silica gel
t	Triplet
TLC	Thin Layer Chromatography
VLC	Vacuum Liquid Chromatography

LIST OF APPENDICES

APPENDIX

TITLE

PAGE

1	GC Chromatogram of Curcuma aeruginosa Rhizome Oil	90
2	GC Chromatogram of Kaempferia rotunda Rhizome Oil	91
3	IR Spectrum of Curzerenone (1)	92
4	¹ H NMR Spectrum of Curzerenone (1)	93
5	¹ H - ¹ H COSY Spectrum of Curzerenone (1)	94
6	¹³ C NMR Spectrum of Curzerenone (1)	95
7	DEPT Spectra of Curzerenone (1)	96
8	EIMS Spectrum of Curzerenone (1)	97
9	HMQC Spectrum of Curzerenone (1)	98
10	HMBC Spectrum of Curzerenone (1)	99
11	IR Spectrum of Furanodienone (37)	100
12	¹ H NMR Spectrum of Furanodienone (37)	101
13	¹³ C NMR Spectrum of Furanodienone (37)	102
14	DEPT Spectra of Furanodienone (37)	103
15	HMQC Spectrum of Furanodienone (37)	104
16	HMBC Spectrum of Furanodienone (37)	105
17	IR Spectrum of Curcumenone (108)	106
18	¹ H NMR Spectrum of Curcumenone (108)	107
19	¹ H - ¹ H COSY Spectrum of Curcumenone (108)	108
20	¹³ C NMR Spectrum of Curcumenone (108)	109
21	DEPT Spectra of Curcumenone (108)	110
22	HMQC Spectrum of Curcumenone (108)	111
23	HMBC Spectrum of Curcumenone (108)	112
24	IR Spectrum of Curcumenol (6)	113
25	¹³ C NMR Spectrum of Curcumenol (6)	114

26	DEPT Spectra of Curcumenol (6)	115
27	¹ H NMR Spectrum of Curcumenol (6)	116
28	HMBC Spectrum of Curcumenol (6)	117
29	IR Spectrum of Germacrone (41)	118
30	¹³ C NMR Spectrum of Germacrone (41)	119
31	DEPT Spectra of Germacrone (41)	120
32	¹ H NMR Spectrum of Germacrone (41)	121
33	¹ H - ¹ H COSY Spectrum of Germacrone (41)	122
34	HMQC Spectrum of Germacrone (41)	123
35	HMBC Spectrum of Germacrone (41)	124
36	EIMS Spectrum of Germacrone (41)	125
37	IR Spectrum of Curzeone (109)	126
38	¹ H NMR Spectrum of Curzeone (109)	127
39	¹ H - ¹ H COSY Spectrum of Curzeone (109)	128
40	¹³ C NMR Spectrum of Curzeone (109)	129
41	DEPT Spectra of Curzeone (109)	130
42	HMBC Spectrum of Curzeone (109)	131
43	HMQC Spectrum of Curzeone (109)	132
44	IR Spectrum of Dehydrocurdione (110)	133
45	¹ H NMR Spectrum of Dehydrocurdione (110)	134
46	¹ H - ¹ H COSY Spectrum of Dehydrocurdione (110)	135
47	¹³ C NMR Spectrum of Dehydrocurdione (110)	136
48	DEPT Spectra of Dehydrocurdione (110)	137
49	HMQC Spectrum of Dehydrocurdione (110)	138
50	HMBC Spectrum of Dehydrocurdione (110)	139
51	IR Spectrum of Benzyl Benzoate (43)	140
52	¹ H NMR Spectrum of Benzyl Benzoate (43)	141
53	¹ H - ¹ H COSY Spectrum of Benzyl Benzoate (43)	142
54	¹³ C NMR Spectrum of Benzyl Benzoate (43)	143
55	DEPT Spectra of Benzyl Benzoate (43)	144
56	HMQC Spectrum of Benzyl Benzoate (43)	145
57	HMBC Spectrum of Benzyl Benzoate (43)	146
58	EIMS Spectrum of Benzyl Benzoate (43)	147

xviii

59	IR Spectrum of Crotepoxide (51)	148
60	¹ H NMR Spectrum of Crotepoxide (51)	149
61	¹ H - ¹ H COSY Spectrum of Crotepoxide (51)	150
62	¹³ C NMR Spectrum of Crotepoxide (51)	151
63	DEPT Spectra of Crotepoxide (51)	152
64	HMQC Spectrum of Crotepoxide (51)	153
65	HMBC Spectrum of Crotepoxide (51)	154
66	EIMS Spectrum of Crotepoxide (51)	155
67	IR Spectrum of Lignoceric acid (111)	156
68	¹³ C NMR Spectrum of Lignoceric acid (111)	157
69	DEPT Spectra of Lignoceric acid (111)	158
70	¹ H NMR Spectrum of Lignoceric acid (111)	159
71	IR Spectrum of Stigmasterol (112)	160
74	EIMS Spectrum of Stigmasterol (112)	161
73	¹ H NMR Spectrum of Stigmasterol (112)	162
74	¹³ C NMR Spectrum of Stigmasterol (112)	163

CHAPTER 1

INTRODUCTION

1.1 Background of the Problem

Malaysia is a unique country with high biological diversity. There are diverse of plants in a variety of forest which can be found all over Malaysia such as plants of Zingiberaceae, Leguminoceae and Annonaceae. About 12 to 18% of trees, shrubs and herbs from more than 7,000 species of angiosperms and 600 species of ferns in Malaysia are reported to have medicinal values [1]. As a result of chemical studies from isolation of active compounds from various plants, many biologically active plant-derived compounds were discovered and being used in industry according to their medicinal properties [2].

Zingiberaceae which have been reported to be useful as food, traditional medicine, spice, condiment, dye and flavour is one of the largest monocotyledonous family of the order Zinziberales. Approximately 160 species from 18 genera are found, mostly grow naturally in damp, shaded parts of the lowland or hill slopes, as scattered plants or thickets in Peninsular Malaysia [3]. About 23 genera and 200 species of Zingiberaceae have been recognized in Malaysia. Among the common species found are *Curcuma, Zingiber, Alpinia, Achasma, Amonum, Costus, Boesenbergia, Kaempferia, Languas, Elettaria, Globba, Hedychium,* and *Nicolaia* [4].

One of the genus of Zingiberaceae family is *Curcuma* which consists of about 40 species out of 1500 species in Zingiberaceae. Most of the *Curcuma* species are

growing in mountainous area of the world [5]. *Curcuma* is a monocotyledon plant with a fleshy complex rhizome and the roots bearing ellipsoid tubers and usually has more or less erect leaf-blades. *Curcuma* species are distributed in India, Southeast Asia and Northern Australia. *Curcuma* species such as *C. longa*, *C. mangga*, *C. aeruginosa*, *C. xanthorrhiza*, *C. zedoaria*, *C. viridiflora*, *C. phaeocaulis*, *C. aromatica*, *C. domestica*, and *C. angustifolia* are well-known as village plants in Malaysia [6].

Beside *Curcuma* species, *Kaempferia* is also one of the genus in Zingiberaceae family. The name *Kaempferia* is taken from a German botanist name, Engelbert Kaempfer (1651-1716). The possible range of distribution for this genus is in Africa and Asia because *Kaempferia* can adapts to life in seasonal climates and more open places. *Kaempferia* is known as small herbs with tuberous root and short rhizomes. The flowers of the genus appear to consist of four lobes, surrounded by three thin and narrow corolla lobes [6].

Both *Curcuma* and *Kaempferia* have their own medicinal values. Some of the medicinal properties reported were antioxidant, antibacterial, anti-inflammatory and anti-tyrosinase. Many antibacterial products are sold today especially for cleaning and hand washing, while antioxidants are widely used in dietary supplement in order to help maintaining human health [7].

An essential oil is natural oil with a distinctive scent secreted by the glands of certain aromatic plants. It is a very complex natural mixtures which includes two groups of components from distinct biosynthetically origin. The main group consists of terpenes and terpenoids while the second group is composed of aromatics and aliphatic constituents of low molecular weight. Essential oils are extracted from the plants by hydrodistillation, steam distillation, extraction with cold neutral fats or solvents such as alcohol or pressing. They are used in preservation of food, antimicrobial, analgesic, anti-inflammatory, and local anesthetic remedies because of their fragrance, antiseptic and medicinal properties [8].

1.2 *Curcuma* Species: Botany, Distribution and Usage

The genus *Curcuma* is naturally distributed throughout south and Southeast Asia with the highest diversity in India and Thailand. This genus comprises of about 70 species of rhizomatous herbs. The genus *Curcuma* is economically important due to its pharmacological effects. Rhizomes of *Curcuma* are useful in traditional medicine in Asia. Several of the *Curcuma* species are reported to possess anti-tumor and anti-virus activities [9].

Previous study on *Curcuma* species showed that the phytochemicals of this species consisted of monoterpenoids, sesquiterpenoids and diarylheptanoids. *C. xanthorrhiza, C. zedoaria, C. longa, C. domestica* and *C. aromatica* are the species found in Malaysia and Indonesia. These species are widely investigated by researchers in the area of natural products and food chemistry due to their wider application in traditional medicine [6].

C. aeruginosa is a native tropical plant of Southeast Asia, including Malaysia, Myanmar, Cambodia, Vietnam, Indonesia and Thailand. It is a perennial plant with leafy shoots of 45–60 cm high and roots or tubers are oblong. This plant is distinguished by red corolla lobes and ferruginous or greenish-blue rhizome. In Malaysia, it is known locally as *temu hitam* [6]. Fresh rhizomes of this species emit the aroma of ginger-like and are mildly aromatic. The rhizomes have been used for many years in traditional medicine especially for gastrointestinal remedies treatment of diarrhea, colic and for postpartum care, uterine involution, treatment of uterine pain and uterine inflammation [10].

C. zedoaria is considered to be native in North-East of India. It is known as zedoary and grows in tropical and subtropical wet forest regions. This plant has fragrant and bears yellow flowers with red and green bracts also has long leaf shoots which can reach 1 meter in height. The underground stem section is large and tuberous with numerous branches. *C. zedoaria* or locally known as *'temu kuning'* is used as a medicine in India and Malaysia. This *Curcuma* species is used as remedies

in the countries such as the provinces of Madras, Bengal and also Indonesia because of digestive, diuretic, emollient and antiscurbotic properties [6].

C. xanthorrhiza which is a member of the Zingiberaceae grows in Southeast Asia. This plant is sometimes referred to as Javanese Turmeric or *temu lawak*. They can grow up to 8 feet tall and the flower is yellow in color. It has an aromatic, pungent odor and a bitter taste. The large leaves stem from the root and the large rhizome of this plant contains herbal qualities. The blend of the juice from the rhizomes of *C. xanthorrhiza* is used to treat indigestion and rheumatism or applied to the body after childbirth [3, 6].

C. domestica is a native plant of South Asia and is cultivated extensively throughout the warmer parts of the world. Its rhizomes are oblong, ovate, pyriform and often short-branched. The rhizome is widely used in powder form as food additive to impart flavor and a yellow color. It is very popular in Asian medicine for the treatment of coryza, hepatic disorders and rheumatism [11].

C. mangga (locally known amongst Malays as '*temu pauh*') cultivated in Java and Malaysia. It is a plant of the ginger family which is closely related to turmeric. The structure of the rhizomes is very similar to ginger but it has a raw mango taste and always used for the preparation of food, supplements and traditional medicine. Its aromatic leaves are useful for flavouring steamed and baked fish while its rhizomes are always used in making pickles in several parts of India. The fresh tips of young rhizomes and shoots are sometimes consumed with rice. The rhizomes are useful to treat stomach, chest pains, fever and also used in postpartum care, specifically to aid healing of wounds [6, 12, 13].

1.3 *Kaempferia* Species: Botany, Distribution and Usage

Kaempferia is one of the important genus in Zingiberaceae family which can be found in the Southeast Asia. Plants of the genus are small and herbaceous with short, fleshy or slender rhizomes and one to a few leaves [14]. The *Kaempferia* species can be found in very damp, shaded areas and usually close to streams or in boggy conditions [15].

Several *Kaempferia* species are used to be cultivated in villages for food, spice and folk medicine. Other than that the rhizomes part of *Kaempferia* species has been proven by many researchers to display health benefit properties. The extracts of rhizomes from some *Kaempferia* species can exhibit anti-inflammatory and HIV-1 protease inhibitory activity [16].

K. rotunda Linn. is a stemless and rhizomatous herb. The leaves are erect, oblong, up to 30 cm long, while the flowers are large, light purple and appear before leaves, from radical spike [6]. The leaves and rhizomes of this species are used to treat sprain and stomachic aches by the locals of Northeast India. The paste of the ground rhizomes was mixed with other herbs and applied to the sprains and covered with a bandage while the leaves paste is used as body lotion. This usage of *K. rotunda* is popular in Assam, Manipur and Mizoram area [17].

K. galanga Linn. is said to be native in India and widely cultivated throughout Southeast Asia. This species is known as *Chandramula* by the Assamese live in Northeast India. It is a common village plant in Malaysia known as *Chekur*. *K. galanga* is used both for medicine and to flavor food [6]. The rhizomes of *K. galanga* L. were externally used, taken orally and sometimes used together with the rhizomes of *Z. montanum* and *Z. officinale* as local medicine. The leaves and rhizomes were useful in treating indigestion, cold, pectoral and abdominal pains, headache, carminative and toothache and menstrual pain. They are also effective for dandruff or scabs on the head, antidiarrheal and against poisoning when there is blood vomiting [18-19].

In China, *K. galanga* L. is used to spice food and important ingredient for medicinal industry. The rhizomes of this plant are used traditionally to treat symptoms ranging from hypertension, pectoral and abdominal pains, headache, toothache, rheumatism, dyspepsia, coughs and inflammatory tumor. It also has been used as fragrance to help restlessness, stress, anxiety and depression. While in Japan, it has been used as one of the main ingredients in a scent bag which is recognized as improving sleep or minimizing stressful situations, due to this species possesses a strong characteristic balsamic odor [20].

K. pandurata (*Syn. B. pandurata*) cultivated in some tropical countries including Myanmar, Indonesia, Malaysia, and Thailand is a perennial herb of the Zingiberaceae family. In Indonesia, Malaysia, and Thailand, this species is a popular folk medicine for the treatment of diseases such as ulcer, dry mouth, stomach discomfort, leucorrhea, and dysentery. The fresh rhizomes are used as a flavoring agent in Thai cuisine due to its characteristic aroma [21].

K. parviflora Wall., which is locally known in Thai as *Kra-chai-dam*, belongs to the family Zingiberaceae. *K. parviflora* species has a black rhizome and have been used for the treatment of colic disorder as well as peptic and duodenal ulcers. It is well-known as 'Thai ginseng' in Thailand [22].

1.4 Statement of Problem

The plants of *Curcuma* and *Kaempferia* from Zingiberaceae family have great interest due to their variety in essential oil compositions, phytochemicals and biological properties. The essential oils, phytochemicals and bioactivity studies which include antibacterial, antioxidant and anti-inflammatory of *Curcuma* and *Kaempferia* species are well explored. However, the anti-tyrosinase activity of the essential oils and phytochemicals from these species has not been investigated yet.

1.5 Objectives of Study

The objectives of this research are as follows:

- (i) To extract the essential oils from the fresh rhizomes of *C. aeruginosa* and *K. rotunda*.
- (ii) To determine the chemical compositions of the essential oils.
- (iii) To extract and purify phytochemicals from the dried rhizomes of *C*. *aeruginosa* and *K. rotunda*.
- (iv) To identify the structure of the pure phytochemicals.
- (v) To screen the antibacterial, antioxidant and anti-tyrosinase of the essential oils, crude extracts and pure phytochemicals.

1.6 Scope of the Study

The essential oils and phytochemicals from the rhizomes of *C. aeruginosa* and *K. rotunda* will be investigated. The essential oils will be extracted from the fresh rhizomes using hydrodistillation technique and the chemical compositions of the essential oils will be analyzed using GC and GC-MS. The crude extracts will be extracted from the dried rhizomes using soxhlet extractor. The extracts will be fractionated using vacuum liquid chromatography and column chromatography to get the pure isolated phytochemicals which will be elucidated structurally using spectroscopic techniques including 1D and 2D NMR, IR, MS and GC-MS. The essential oils, crudes and selected phytochemicals from both species will be screened for antibacterial using disc diffusion method followed by minimum inhibition concentration (MIC) and minimum bactericidal concentration (MBC). The modified dopachrome method with L-DOPA will be performed for anti-tyrosinase activity.

REFERENCES

- Kumara, K., Dan, Y.M. and Tuan Marina, T.I. Economic Significance of Medicinal Plants in Peninsular Malaysia. Kuala Lumpur: Forestry Department, Peninsular Malaysia. 1998.
- Itokawa, H., Morris, N.S.L., Akiyama T. and Lee K.H. Plant-Derived Natural Product Research Aimed at New Drug Discovery. *J. Nat. Med.* 2008. 62: 263-280.
- Ruslay, S., Abas, F., Shaari, K., Zainal, Z., Maulidiani, Sirat, H., Ahmad Israf, D., Lajis, N.H. (2007). Characterization of the Components Present in the Active Fractions of Health Gingers (*Curcuma xanthorrhiza* and *Zingiber zerumbet*) by HPLC–DAD–ESIMS. *Food Chemistry*. 104: 1183–1191.
- 4. Singh, G., Singh, O.P., Maurya, S. (2002)). Chemical and Biocidal Investigations on Essential Oils of Some Indian Curcuma Species. *Progress in Crystal Growth and Characterization of Materials*. 75-81.
- Usman, L.A., Hamid, A.A., George, O.C., Ameen, O.M., Muhammad, N.O., Zubair, M.F., Lawal, A. (2009). Chemical Composition of Rhizome Essential Oil of *Curcuma longa* L. Growing in North Central Nigeria. *World J. Chemistry* 4 (2):178-181.

- Holttum, R.E. *The Zingiberaceae of the Malay Peninsular*. Singapore: S.N. Publisher. 1950. 65-73.
- Amirah, T.S., Samah, O.A., Taher, M., Susanti, D., Qaralleh, H. (2012). Antimicrobial activity and essential oils of *Curcuma aeruginosa*, *Curcuma mangga*, and *Zingiber cassumunar* from Malaysia. *Asian Pacific Journal Tropical Medicine*. 202-209.
- Bakkali, F., Averbeck, S., Averbeck, D., Idaomar, M. (2008). Biological Effects of Essential Oils – A review. *Food and Chemical Toxicology*. 46:446–475.
- Thaina, P., Tungcharoen, P., Wongnawa, M., Reanmongkol, W. (2009). Uterine Relaxant Effects of *Curcuma aeruginosa* Roxb. Rhizome Extracts. *J. Ethnopharmacology* 121:433–443.
- Sukari, M.A, Saad, S.M., Lajis, N., Rahman, M., Muse, R., Yusuf, U.K., Riyanto, S. (2007). Chemical Constituents and Bioactivity of *Curcuma* aeruginosa Roxb. Natural Products Sciences. 13(3):175-179.
- Qin, N.Y., Yang, F.Q., Wang, Y.T., Li, S.P. (2007). Quantitative Determination of Eight Components in Rhizome (*Jianghuang*) and Tuberous Root (*Yujin*) of *Curcuma longa* Using Pressurized Liquid Extraction and Gas Chromatography–Mass Spectrometry. J. Pharmaceutical and Biomedical Analysis. 43:486–492.
- 12. Liu, Y., Nair, G.M. (2012). *Curcuma longa* and *Curcuma mangga* leaves exhibit functional food property. *Food Chemistry*. 135: 634–640.

- Liu, Y., Nair, G.M. (2011). Labdane diterpenes in *Curcuma mangga* rhizomes inhibit lipid peroxidation, cyclooxygenase enzymes and human tumour cell proliferation. *Food Chemistry*. 124: 527–532.
- 14. Sirat, H.M., Jamil, S., Hussain, J. (1998). Essential Oil of *Curcuma aeruginosa* Roxb. From Malaysia. *J. Essent. Oil Res.* 10: 453-458.
- Vanijajivaa, O., Sirirugsab, and P., Suvachittanont, W. (2005). Confirmation of Relationships among *Boesenbergia* (Zingiberaceae) and Related Genera by RAPD. *Biochemical Systematics and Ecology*. 33:159–170.
- 16. Jing, L.J., Mohamed, M., Rahmat, A., and Abu Bakar., M.F. (2010). Phytochemicals, Antioxidant Properties and Anticancer Investigations of the Different Parts of Several Gingers Species (*Boesenbergia rotunda*, *Boesenbergia pulchella* var attenuata and Boesenbergia armeniaca). J. Med. Plants Research. 4(1):027-032.
- Tushara, Basaka, S., Sarma, G.C., Rangan, L. (2010). Ethnomedical Uses of Zingiberaceous Plants of Northeast India. *J. Ethnopharmacology*.
- Wan Ibrahim, W.I., Sidik, K., Kuppusamy, U.R. (2010). A High Antioxidant Level in Edible Plants is Associated with Genotoxic Properties. *Food Chemistry*. 122:1139–1144.
- Kanjanapothi, D., Panthong, A., Lertprasertsuke, N., Taesotikul, T., Rujjanawate, C., Kaewpinit, D., Sudthayakorn, R., Choochote, W., Chaithong, U., Jitpakdi, A., Pitasawat, B. (2004). Toxicity of Crude Rhizome Extract of *Kaempferia galanga* L. (Proh Hom). *J. Ethnopharmacology*. 90:359–365.

- Huang, L., Yagura, T., Chena, S. (2008). Sedative Activity of Hexane Extracts of *Kaempferia galanga* L. and Its Active Compounds. J. *Ethnopharmacology*. 120: 123–125.
- Win, N.N., Awale, S., Esumi, H., Tezuka, Y., and Kadota, S. (2007). Bioactive Secondary Metabolites from *Boesenbergia pandurata* of Myanmar and Their Preferential Cytotoxicity against Human Pancreatic Cancer PANC-1 Cell Line in Nutrient-Deprived Medium. *J. Nat. Prod.* 70 (10): 1582-1587.
- 22. Tewtrakul, S., Subhadhirasakul, S., Karalai C., Ponglimanont, C. and Cheenpracha, S. (2009). Anti-inflammatory Effects of Compounds from *Kaempferia parviflora* and *Boesenbergia pandurata*. *Food Chem.* 115: 534-538.
- Jarikasem, S., Thubthimthed, S., Chawananoreseth, K., and Suntorntanasat, T. (2005). Essential Oils from Three *Curcuma* Species Collected in Thailand. *Acta Hort* 675, 37-41.
- Mau, J.L., Eric, Y.C., Wang, N.P., Chen, C.C., Chang, C.H., Chyau, C.C. (2003). Composition and Antioxidant Activity of the Essential Oil from *Curcuma zedoaria*. *Food Chemistry*. 82:583-591.
- Jatoi, S.A., Kikuchi, A., Gilani, S.A., and Watanabe, K.N. (2007). Phytochemical, Pharmacological and Ethnobotanical Studies in Mango Ginger (*Curcuma amada* Roxb.; Zingiberaceae). *Phytother. Res.* 21: 507– 516.
- Abdul Wahab, I.R., Blagojevic, P.D., Radulovic, N.S., and Boylan, F. (2011).
 Volatiles of *Curcuma mangga* Val. & Zijp (Zingiberaceae) from Malaysia.
 Chemistry & Biodiversity. Vol. 8.

- 27. Sirat, H.M., Jamil, S., Siew, L.W. (2005). The Rhizome Oil of *Kaempferia rotunda* Val. *J. Essential Oil Research*. 17:3, 306-307.
- Woerdenbag, H.J., Windono, T., Bos R., Riswan, S., and Quax, W.J. (2004).
 Composition of the Essential Oils of *Kaempferia rotunda* L. and *Kaempferia angustifolia* Roscoe Rhizomes from Indonesia. *Flav. Fragr. J.* 19: 145-148.
- Tewtrakul, S., Yuenyongsawad, S., Kummee, S., and Atsawajaruwan, L. (2005). Chemical Components and Biological Activities of Volatile Oil of *Kaempferia galanga* Linn. *Songklanakarin J. Sci. Technol.*27 (2): 503-507.
- Phan Minh, G., Phan Tong, S., Matsunami, K., Otsuka, H. (2007). New sesquiterpenoids from *Curcuma* aff. *aeruginosa* Roxb. *Heterocycles*. 74:977-981.
- Shiobara, Y., Asakawa, Y., Kodama, M., Yasuda, K., Takemoto, T. (1998). Curcumenone, Curcumanolide B, Three Sesquiterpenoids from *Curcuma zedoaria*, *Phytochemistry*, 24(11):2629-2633.
- Hwang, J.K., Shim, J.S., Pyun, Y.R. (2000). Antibacterial Activity of Xanthorrhizol from *Curcuma xanthorrhiza* Against Oral Pathogens. *Fitoterapia*. 71:321-323.
- Pancharoen, O., Tuntiwachwuthkul, P., and Taylor, W.C. (1996). Cyclohexane Diepoxides from *Kaempferia rotunda*. *Phytochemistry*. 43 (1): 305-308.
- 34. Win, N.N., Awale, S., Esumi, H., Tezuka, Y., and Kadota, S. (2008). Panduratins D-I, Novel Secondary Metabolites from Rhizomes of

Boesenbergia pandurata. Chem. Pharm. Bull. 56 (4): 491-496.

- Jaipetch T, Kanghae S, Pancheroen O. (1982). Constituents of *Boesenbergia* pandurata (syn. Kaempferia pandurata): Isolation, Crystal Structure and Synthesis of (±)-Boesenbergin A. Aust. J. Chem. 35: 351-361.
- Tewtrakul, S., Subhadhirasakul, S. and Kummee, S. (2008). Anti-allergic Activity of Compounds from *Kaempferia parviflora*. J. *Ethnopharmacol*.116:191-193.
- Tewtrakul, S. and Subhadhirasakul, S. (2008). Effects of Compounds from Kaempferia parviflora on Nitric Oxide, Prostaglandin E2 and Tumor Necrosis Factor-Alpha Productions in RAW264.7 Macrophage Cells. J. Ethnopharmacol. 120: 81-84.
- Pancharoen, O., Tuntiwachwuttikul, P. and Taylor, W.C. (1989).
 Cyclohexane Oxide Derivatives from *Kaempferia Angustifolia* and *Kaempferia* Species. *Phytochemistry*, Vol 28, No 4: 1143-1148.
- Reanmongkol, W., Subhadirasakul, S., Khaisombat, N., Fuengnawakit, P., Jantansila, S., Khamjun, A. (2006). Investigation the Antinoceptive, Antipyretic and Anti-Inflammatory Activities of *Curcuma aeruginosa* Roxb. Extracts in Experimental Animals. J. Sci. Technology. 28(5):999-1008.
- 40. Panji, C., Grimm, C., Wray, V. (1993). Insecticidal Contituents from our Species of the Zingiberaceae. *Phytochemistry*. 34:415-419.
- Wilson, B., Abraham, G., Manju, V.S., Mathew, M., Vimala, B., Sundaresan, S., Nambisan, B. (2005). Antimicrobial Activity of *Curcuma zedoaria* and *Curcuma malabrica* tubers. *J. Ethnopharmacology*. 99:147-151.

- Navarro D.F., de Souza, M.M., Neto, R.A., Golin, V., Niero, R., Yunes, R.A., Delle Monache, F., Cechinel Filho, V. (2002). Phytochemical Analysis and Analgesic Properties of *Curcuma zedoaria* Grown in Brazil. *Phytomedicine*. 9:427–432.
- 43. Hikino, H., Agatsuma, K., Takemoto, T. (1968). Furanodiene, a Precursor of Furan-Containing Sesquiterpenoids. *Tetrahedron Letters*. 8:931-933.
- Win, N.N., Awale, S., Esumi, H., Tezuka, Y., and Kadota, S. (2008).
 Panduratins D-I, Novel Secondary Metabolites from Rhizomes of *Boesenbergia pandurata. Chem. Pharm. Bull.* 56 (4): 491-496.
- 45. Yun, J.M., Kweon, M.H., Kwon, H.J., Hwang, J.K., and Mukhtar, H. (2006). Induction of Apoptosis and Cell Cycle Arrest by a Chalcone Panduratin A Isolated from *Kaempferia pandurata* in Androgen-independent Human Prostate Cancer Cells PC3 and DU145. *Carcinog.* 27 (7): 1454-1464.
- 46. Hwang, J.K., Chung, J.Y., Baek, N.I., Park, J.H. (2004). Isopanduratin A from *Kaempferia pandurata* as an Active Antibacterial Agent Against Cariogenic *Streptococcus mutans*. *Int. J. Antimicrob. Agents*. 23: 377-381.
- 47. Yenjai, C., Prasanphen, K., Daodee, S., Wongpanich, V. and Kittakoop, P. (2004). Bioactive Flavonoids from *Kaempferia parviflora*. *Fitoterapia*. 75: 89-92.
- Thongnest, S., Mahidol, C., Sutthivaiyakit, S., Ruchirawat, S. (2005).
 Oxygenated Pimarane Diterpenes from *Kaempferia marginata*. J. Nat. Prod. 68:1632-1636.

- 49. Nugroho, W.B., Schwarz, B., Wray, V., Proksch, P. (1996). Insecticidal constituents from Rhizomes Of *Zingiber cassumunar* and *Kaempferia rotunda*. *Phytochemistry*. 41 (1): 129-132.
- Adams, R.P. (2007). Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry, 4th Ed.Schmale Road: Allured Publishing Corporation. 31-51.
- Asghari, G., Jalali, M. and Sadoughi, E. (2012). Antimicrobial Activity and Chemical Composition Essential Oil from the Seeds of *Artemisia aucheri* Boiss, *Jundishapur J. Nat. Pharm. Prod.* 2012;7(1):11-15.
- 52. Yau Sui Feng, (2009). Chemical Constituents and Bioactivities of Malaysian and Indonesian Kaempferia rotunda. Universiti Teknologi Malaysia: Tesis Sarjana Sains Kimia.
- Dekebo, A., Dagne, E., Sterner, O. (2002). Furanosesquiterpenes from Commiphora sphaerocarpa and Related Adulterants of True Myrrh. Fitoterapia. 73: 48-55.
- Shajarahtunnur binti Jamil, (1996). Komponen Sebatian Semula Jadi bagi Spesis Curcuma dan Boesenbergia (Zingiberaceae). Universiti Teknologi Malaysia: Tesis Sarjana Sains Kimia.
- Dekebo, A., Dagne E., Hansen, L.K., Gautun, Odd. R., Aasen, A.J. (2000). Crystal Structures of Two Furanosesquiterpenes from *Commiphora* sphaerocarpa. Tetrahedron Letters. 41: 9875 – 9878.

- Sukari, M.A., Wah, T.S., Saad, S. M., Rashid, N.Y., Rahmani, M., Lajis, N.H., Taufiq. (2010). Bioactive Sesquiterpenes from *Curcuma ochrorhiza* and *Curcuma heyneana*. *Nat. Prod. Research.* 24 (9) : 838-845.
- 57. Ohshiro, M., Kuroyanagi, M., Ueno, A. (1990). Structures of Sesquiterpenes from *Curcuma longa*. *Phytochemistry*. 29 (7) : 2201-2205.
- 58. Firman, K., Kinoshita, T., Itai, A., Sankawa, U. (1988). Terpenoids from *Curcuma heyneana*. *Phytochemistry*. 27 (12) : 3887-3891.
- Yang, F.Q., Li, S.P., Chen, Y., Lao, S.C., Wang, Y.T., Dong, T.T.X., Tsim, K.W.K. (2005). Identification and Quantitation of Eleven Sesquiterpenes in Three Species of *Curcuma* Rhizomes by Pressurized Liquid Extraction and Gas Chromatography–Mass Spectrometry. *Journal Pharmaceutical and Biomedical Analysis*. 39 : 552-558.
- Shiobara, Y., Asakawa, Y., Kodama, M., Takemoto, T. (1986). Zedoarol, 13-Hydroxygemacrone and Curzeone, Three Sesquiterpenoids from *Curcuma* zedoaria. Phytochemistry. 25 (6) : 1351-1353.
- Kodpinid, M., Sadavongvivad, C., Thebtaranonth, C. and Thebtaranonth Y. (1984). Benzyl Benzoates from the |Root of Uvaria purpurea. *Phytochemistry*. 23 (1): 199-200.
- Kim, S., and Yi, K.Y. Synthetic Applications of Di-2-pyridyl Thionocarbonate as a Dehydration, a Dehydrosulfuriation, and a Thiocarbonyl Transfer Reagent. Bull. (1987). *Korean Chem. Soc.* 8 (6): 466-470.

- Zhang, Y.H., Ruan, H.L., Hui, F.P., Wu, J.Z., Sun, H.D. and Fujita, T. (2004). Structural Elucidation of Fritillahupehin from Bulbs of *Fritillaria hupehensis* Hsiao et K.C. Hsia. *Journal Asian Natural Products Research*. 6:1, 29-34.
- 64. Kamboj, A., Saluja, A.K. (2011). Isolation of Stigmasterol and β-Sitosterol from Petroleum Ether Extract of Aerial Parts of Ageratum onyzoides (Asteraceae). International Journal Pharmacy and Pharmaceutical Sciences. 3 (1). 94-96.
- Chan, E.W.C., Lim, Y.Y., Wong, L.F., Lianto, F.S., Wong, S.K., Lim, K.K., Joe, C.E., Lim, T.Y. (2008). Antioxidant and Tyrosinase Inhibition Properties of Leaves and Rhizomes of Ginger Species. *Food Chemistry*. 109 : 477–483.
- 66. Narayanaswamy, N., Duraisamy, A. and Balakrishnan, K.P. (2011). Screening of Some Medicinal Plants for Their Antityrosinase and Antioxidant Activities. *International Journal PharmTech Research*. 3(2) : 1107-1112.
- Mbaveng, A.T., Ngameni, B., Kuete, V., Simo, I.K., Ambassa, P., Roy, R., Bezabih, M., Etoa, F. X. Ngadjui, B.T., Abegaz, B.M., Meyer, J.J.M., Lall, N. and Beng, V.P. (2008). Antimicrobial Activity of thr Crude Extracts and Five Flavanoids from the Twigs of Dorstenia barteri (Moracea). J. Ethnopharmacol. 116: 483-489.
- Mackeen, M.M., Ali, A.M., El-Sharkawy, S.H., Manap, M.Y., Salleh, K.M., Lajis, N.H., Kawazu, K. (1997). Antimicrobial and Cytotoxic Properties of Some Malaysian Traditional Vegetables. *Int. J. Pharmacog.* 35, 174-178.

 Aligians N, Kalpoutzakis E, Mitaku S, Chinou IB (2001). Composition and Antimicrobial Activity of the Essential Oil from *Origanum* Species. J. Agric Food Chem, 49. 4168-4170.