SCREENING OF CELLULOLYTIC AND XYLANOLYTIC FUNGI FOR ENZYMES COCKTAIL STUDIES OF *GANODERMA BONINENSE* INHIBITION

ROHAYA BINTI MOHD NOOR

UNIVERSITI TEKNOLOGI MALAYSIA

SCREENING OF CELLULOLYTIC AND XYLANOLYTIC FUNGI FOR ENZYMES COCKTAIL STUDIES OF *GANODERMA BONINENSE* INHIBITION

ROHAYA BINTI MOHD NOOR

A dissertation submitted in partial fulfilment of the requirements for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

MARCH 2014

Specially dedicated to my supportive families and friends Thank you for all love and concern

ACKNOWLEDGEMENT

Alhamdulillah, first of all I would like to thank God, without his bless I,impossible for me to completed my masters dissertation. I also would like to express my indefinite gratitude to my main supervisor, Associate Prof Dr Madihah Bt Md Salleh for her encouragement, patience, guidance and strong motivation.

My sincere thanks are firstly to all the members of Biorefinery Technology Research Laboratory, Faculty Biosciences and Medical Engineering, Universiti Teknologi Malaysia, Skudai, Johor for creating a healthy working environment in the lab, which has been unique in its own way. For this my thanks are due to Noratiqah Binti Kamsani, Reza Soleimani, Kak Rachmawaty, Kak Huszalina Hussin, Ang Siow Kuang, Ahmad Fawwaz Mohd Raji and Shankar A/L Ramanathan for their unvaluable help, moral support, encouragement, advices, critism and sharing their knowledge with me.

My thank you also dedicated to all Staffs of Faculty Biosciences and Medical Engineering, UTM for their helps to provided me everything I need during my work.I am also obliged to express my appreciation towards my beloved parents and siblings for their enduring patience and financial supports. Lastly to all my friends and coursemate, who endure together with me, a heartfelt gratitude to all of you.

ABSTRACT

Ganoderma boninense is a basidiomycetes white rot fungus which causes basal stem rot (BSR), one major disease in oil palm plantation in Malaysia. The objective of the present study was to screen the most potential fungus that has a capability to produce crude xylanase and cellulase to be used as a biological control for Ganoderma boninense. Twenty four different strains of fungi were obtained from the Biorefinery Technology Research Laboratory of Faculty Biosciences and Medical Engineering, Universiti Teknologi Malaysia, Johor. They were screened for their potential to degrade cellulose and hemicelluloses. The screening of the fungi were based on the diameter of holozones on red color of congo red and the formation of yellow-opaque area on the carboxymethlycellulose(CMC) and xylan agar, respectively. Among these strains, five potential fungi which showed the maximum enzyme activities for endoglucanase (CMCase), filter paper degrading enzyme (FPase), beta-glucosidase, xylanase and reducing sugar were selected to produce enzyme cocktail on untreated oil palm trunk will be further study through its performance in solid state fermentation. The potential enzyme producers from fungi were CT2, Pycnoporus cinnabarinus, EFB1, EFB2 and TG6 that produced 304.04 U/g of CMCase, 13.25 U/g of FPase, 83.15 U/g for β -glucosidase, 523.10 U/g of xylanase and 164.04 U/g of reducing sugar respectively. Cellulase and xylanase cocktail produced from these five fungi were mixed and applied on the Ganoderma boninense at ratio 1:1:1:1:1 were made up to 0.025mL in the culture plates. However, the inhibition of Ganoderma boninense when treated with cellulase and xylanase cocktail was undetected in this study.

ABSTRAK

Ganoderma adalah merupakan basidiomykota kulat reput putih yang menyebabkan penyakit yang serius kepada kelapa sawit di Malaysia iaitu reput pangkal batang. Objektif di dalam kajian ini adalah melakukan penyaringan ke atas kulat yang berpotensi untuk menghasilkan enzim mentah sellulase dan xylanase yang akan digunakan sebagai kawalan biologi ke atas Ganoderma boninense. Sebanyak dua puluh empat strain kulat yang diambil daripada Makmal Penyelidikan Biorefineri, Fakulti Biosains dan Kejuruteraan Perubatan, Universiti Teknologi Malaysia, Johor. Kesemua strain ini dilakukan proses penyaringan berdasarkan keupayaan untuk menghuraikan sellulosa dan hemisellulosa. Penyaringan kulat tersebut adalah berdasarkan diameter holozon berwarna merah yang terbentuk daripada congo merah dan legap kuning pada karboksimetil sellulosa (CMC) dan agar xylan masing-masing. Lima jenis kulat yang berpotensi menghasilkan aktiviti endoglukanase (CMCase), enzim pengurai kertas turas (FPase), beta glukosidase, xylanase dan gula penurun yang maksimum telah dipilih untuk menghasilkan koktel enzim ke atas batang kelapa sawit yang tidak dirawat melalui proses fermentasi keadaan pepejal. Pengeluar enzim daripada kulat yang dikenalpasti berpotensi adalah CT2, Pycnoporus cinnabarinus, EFB1, EFB2 dan TG6 yang masing-masing menghasilkan 304.04 U/g CMCase, 13.25 U/g FPase, 83.15 U/g β-glukosidase, 523.10 U/g xylanase dan 164.04 U/g gula penurun. Koktel sellulase dan xylanase ini dicampurkan dan digunakan untuk merencatkan pertumbuhan Ganoderma boninense pada nisbah 1:1:1:1:1 menjadikan jumlah di dalam piring petri sebanyak 0.25ml. Walau bagaimanapun, perencatan Ganoderma boninense apabila dirawat dengan menggunakan koktel sellulase dan xylanase tidak berjaya di dalam kajian ini.

TABLE OF CONTENT

CHAPTER

1

TITLE

PAGE

SUPERVISOR DECLARATION	
TITLE	i
AUTHOR DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGEMENTS	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	xi
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	XV
LIST OF APPENDICES	xvii
INTRODUCTION	1

1.1	General introduction	1
1.2	Objectives of study	4
1.3	Problem statement	5
1.4	Significant of study	5

2 LITERATURE REVIEW

2.1	Introduction to Ganoderma boninense	6
2.2	Enzyme defence related enzymes towards fungal	8
	pathogen	
2.3	Oil palm trunk	10
2.4	Cellulase	10
2.5	Xylanase	11
2.6	Mechanism of cellulase reaction in cellulose	12
	hydrolysis process	
2.7	Mechanism of xylanase reaction in xylan	13
	hydrolysis process	
2.8	Microorganims producing cellulase and xylanase	15
2.9	Application of cellulase	19
2.10	Application of xylanase	20
2.11	Composition of lignocellulosic material	
	2.11.1 cellulose	21
	2.11.2 Hemicellulose	22
	2.11.3 Lignin	23
2.12	Lignocellulosic biomass as a resource for	24
	cellulose	
2.13	Lignocellulose hydrolysis	25
	2.13.1 Acid hydrolysis	25
	2.13.2 Enzymatic hydrolysis	26
2.14	Solid state fermentation	28
2.15	Enzyme cocktail	30

3

4

METHODS AND MATERIALS

3.1	Micro	organism	32
3.2	Experi	mental design	33
3.3	Scree	ning of xylanolytic and celluloytic activities	34
	3.3.1	carboxymethylcellulose agar method	34
	3.3.2	xylan agar method	34
3.4	Inocul	um preparation	35
3.4	Growt	h medium (PDA)	36
3.5	Produc	ction medium	36
3.6	Crude	cellulase and xylanase extraction	37
3.7	Analy	tical method	
	3.7.1	Determination of endoglucanase	37
	3.7.2	Determination of exoglucanase activity	37
	3.7.3	Determination of β -glucosidase activity	38
	3.7.4	Determination of xylanase activity	38
	3.7.5	Determination of protein content	38
	3.7.6	Determination of reducing sugar assay	39
3.8	Ganoa	lerma boninense culture	39
3.9	Applic	cation of crude cellulase and xylanase	39
	cockta	il against Ganoderma boninense	
	3.9.1	Antifungal activity bioassay	39
RESU	LTS A	ND DISCUSSION	
4.1	Qualit	ative screening of cellulolytic and	41
	xyland	olytic fungi	

4.2 Quantitative screening of cellulolytic and 44 xylanolytic fungi

APPENDIC	CES A -	Н	85-104
REFEREN	CES		74
	5.2	Suggestion for Future Works	73
	5.1	Conclusion	72
5	CON	ICLUSION	
		Cocktail on Ganoderma inhibition	
	4.3	Application of crude cellulase and xylanase	68

LIST OF TABLES

TABLE NO	TITLE	PAGE
	IIIDD	Inde

2.1	Types of cellulase and its functions	11
2.2	Types of xylanase and its functions	12
2.3	Microorganisms producing cellulase on various	17
	substrate in solid state fermentation	
2.4	Microorganisms producing xylanase on various	18
	substrates in solid state fermentation	
2.5	Application of cellulase	19
2.6	Application of xylanase	20
4.1	Qualitative analysis of cellulase and xylanase	43
	producing fungi	
4.2a	Quantitative analysis of cellulase of 24 strains	46
	of fungi	
4.2b	Quantitative analysis of xylanase and reducing	48
	sugars of 24 strains of fungi	
4.3	Highest cellulase, xylanase and reducing	50
	sugars in five selected fungi	
4.4	Cellulase and xylanase activities of five selected	51
	fungi and their specific activities	

4.5a	Comparison of cellulase by various fungi	52
	under solid state fermentation	
4.5b	Comparison of cellulase by various fungi	53
	Under solid state fermentation	
4.5c	Comparison of xylanase and reducing sugars	54
	by various fungi under solid state fermentation	
4.6	Qualitative analysis of fungus CT2	59
4.7	Qualitative analysis of fungus Pycnoporus	61
	cinnabarinus	
4.8	Qualitative analysis of fungus EFB1	63
4.9	Qualitatitve analysis of fungus EFB2	65
4.10	Qualitative analysis of fungus TG6	6

LIST OF FIGURES

FIGURE NO

TITLE

PAGE

1.1	The Ganoderma boninense basidiomata	3
2.1	Ganoderma boninense fruiting bodies found at the bottom	7
	of sick oil-palm stem	
2.2	The cellulose biodegradation into glucose	13
2.3	Structure of xylan and the sites of its attack by	14
	xylanolytic enzyme	
2.4	Structural formula of cellulase	22
2.5	Structure of three lignin precursors	23
2.6	Structure of lignocelluloses materials	24
4.1	Time course of cellulase, xylanase and reducing sugars	58
	by fungus CT2	
4.2	Time course of cellulase, xylanase and reducing sugars	60
	by fungus Pycnoporus cinnabarinus	
4.3	Time course of cellulase, xylanase and reducing sugars	62
	by fungus EFB1	
4.4	Time course of cellulase, xylanase and reducing sugars	64
	by fungus EFB2	
4.5	Time course of cellulase, xylanase and reducing sugars by	66

fungus TG6

4.6	Culture of Ganoderma boninense after 10 days	68
4.7	Antifungal activity bioassay	68
4.8	Interaction between Ganoderma boninense and cellulase	69
	and xylanase cocktail	

LIST OF ABREVIATIONS

et. al	and friends
BSR	Basal stem rot
CMC	Carboxymethyl cellulose
CMCase	Carboxymethyl cellulose
FPase	Filter paper culture enzyme
SSF	Solid state fermentation
SmF	Submerged fermentation
Н	Hour
μ	Micro
μ nm	Micro Nanometer
nm	Nanometer
nm °C	Nanometer Degree celcius
nm °C PNPG	Nanometer Degree celcius p-nitrophenyl β-D-glucoside
nm °C PNPG Rpm	Nanometer Degree celcius p-nitrophenyl β-D-glucoside Rotation per minute

m	Slope
L	Liter
mL	Mililiter
cm	Centimetre
g	Gram
NaOH	Sodium hydroxide
NaCl	Sodium chloride
PDA	Potato dextrose agar
OD	Optical density
BSA	Bovine serum albumin
v/v	Volume per volume
w/v	Weight per volume
μL	Micro liter

LIST OF APPENDICES

APPENDIX

TITLE PAGE

A	Spore counting using haemocytometer	85
В	Preparation of Dinitrosalicyclic acid	
	(DNS) reagent	87
С	Determination of CMCase activity using	88
	DNS method	
D	Determination of FPase activity using	91
	DNS method	
E	Determination of β -glucosidase activity using	94
	DNS method	
F	Determination of Xylanase activity	97
G	Determination of Protein content using	100
	Lowry method	
Н	Determination of reducing sugar using	103
	DNS method	

CHAPTER 1

INTRODUCTION

1.1 General Introduction

Malaysia has been the second largest oil palm plantation areas after Indonesia (Chong *et al.*, 2012) and one of the largest contribution for an economic crops in our industrial. The amount of biomass produced reached 225 million tonnes per annum with 4.5 million ha of oil palm (Mohamed and Alimon, 2012). The rate estimation of palm oil production in our country is more than twelve million tonnes (Najafpour et al., 2007). The important oil palm producing countries reported by (Paterson et al., 2013) are Indonesia, Malaysia, Nigeria, Democratic Republic of Congo, the Ivory Coast, Brazil, Colombia, Costa Rica and Ecuador.

In palm oil industry, the main residues are created from two major sources of processing which is from plantations or field residues and from palm oil milling. Oil palm trunks (OPT) and oil palm fronds (OPF) were categorized as field residues while empty fruit bunch (EFB), palm kernel cake (PKC), palm oil mill effluent (POME), palm press fibre (PPF) and shell are created from palm oil milling process (Zahari *et al.*, 2012). Approximately 5 million tonnes of agricultural wastes were generated from the industry (Ibrahim, 2006). Therefore, all the co-products produced

from the palm oil industry need to be processed further to prevent the land pollution as well as to reduce a detrimental impact by convert it into value added product by using microorganism degradation.

There are many products had been produced from the agricultural waste. For example, fiber can be used to make pulp, paper and particle boards. Empty fruit bunch can be used to produce energy and the fertilizer while the oil palm trunk can be made as a furniture, particle board and also biofuels (Ng *et al.* 2011).

One of the most important diseases attacking the oil palm in Malaysia is a *Ganoderma* white rot. This disease caused by one of the pathogen which is called *Ganoderma boninense*. Azadeh *et al.* (2010) stated that cultural, chemical and clean clearing strategies only focused on the way to minimize the incidence of BSR in replanting, prolonging, the productive life of infected palms and delaying the duration of *Ganoderma* infection. The other method to reduce the chemical products application in agriculture is by using the biological control from the microorganism.

Ganoderma white rot disease may be controlled by the enzyme inhibitors. *Ganoderma* white rot can damage many living trees and wood which were used for manufacturing industry. *Ganoderma* can infected rubber trees and tea which developed red root rot disease in both of these plants (Zakaria *et al.*, 2009). This fungus also may attack the part of oil palm and seriously infects undamaged trees. It can infects palms as young as 12-24 months after planting and serious infects on palms of 4-5 years age, usually in replanted areas (Bivi *et al.*, 2010). Research by Abdullah *et al.* (2012) mentioned that our industry would lose about RM80 million each year if 2.5 percent of the total acreage of oil palm plantation in Malaysia is suffered by this disease. *Ganoderma* species can cause a major disease in palm oil industry. One of the major parts in the disease process is a lignin biodegradation which produce carbon dioxide and water molecule. Once lignin was degraded, fungus obtained energy from the cellulose (Paterson, 2007). This fungus species are grown in wood and classified as wood decaying fungi. It degrades cellulose, hemicellulose and lignin. *Ganoderma* is known as polypores because they possessed tiny pores underside their cap where the reproductive spores were hiding. The caps of the spores will always change when the age of that species were changed where it spongy hardening to a shiny when fresh while smooth woody structure when it was matured (Figure 1.1). The colour of the caps also always changed which were ranges from brown to yellowish (Wei *et al.*, 2003). The surface of the pore was cream in colour while the spores were brown colour.



Figure 1.1 The *Ganoderma boninense* basidiomycete (Abdullah *et al.*, 2012)

However, *Ganoderma* species were not listed among the group of edible mushrooms because of its physical appearance. Its fruiting bodies were always thick, corky and tough and do not have the fleshy texture characteristics of true edible fungi. Dry rot occurs when *Ganoderma* species attack the palm roots and spread to the stem of the palm. If the branch of palm is shedding, the wound surface will be attacked by airborne spore and this is how *Ganoderma* starts their lives. As the consequences, the transportation and absorption of nutrients by roots were disturbed. When the fungus degrade the_lignin, which is one of the component of wood, it will form a white cellulose and this is one of the sources for white rot fungi to get their energy.

Applications of enzymes and microorganism can be used for degradation of lignobiomass of palm oil to produce value added products such as polyoses, organic acid, biomaterials, biofuel, bioenergy and biolignin. The cost effective process developed for inhibition of *Ganoderma* species growth can be patented for the future. One of the effective ways to overcome this problem is through the application of green technology using microbial or enzymatic reaction. Enzymes production from the microbial reaction with the agrowaste was sustainable due to the renewable and ubiquitous nature of biomass and to be an excellent carbon source in microbial enzyme production (Bivi *et al.*, 2010).

1.2 Objectives of study

a) To screen the potential defense related enzymes producing fungi capable to inhibit *Ganoderma boninense*.

b) To apply crude cellulase and xylanase cocktail produce by fungi CT2, *Pycnoporus cinnabarinus*, EFB1, EFB2 and TG6 for *Ganoderma boninense* inhibition.

1.3 Problem of statement

Ganoderma spesies is one of the fungi which capable to attack palm oil. One of the major diseases in oil palm is Basal Stem Rot (BSR). *Ganoderma boninense* attacks the root system of oil palm from second and subsequent planting cycles. Oil palm has 25 to 30 years of economic life span but this disease can kill more than 80% of stands by the time they are half-way through a normal economic life. The infection of this disease resulted in crop loss up to 45% of the yield of oil palm (Bivi *et al.*, 2010). Basal stem root in oil palm can be controlled effectively by management practices in the early stage of disease development.

1.4 Significance of study

The normal conventional ways to solve *Ganoderma boninense* problems in oil palm industry are by using pesticide and burning which would cause disaster for the surrounding. These activities would cause massive air pollution or even release the toxic compound to soil, environment and give dangerous impact to the human's health. Green technology by using microbial or enzymatic reaction will become the replacement for the chemical pesticide in order to treat *Ganoderma boninense*. The cost-effective process developed for inhibition of *Ganoderma boninense* growth can be patented for the future. The treatment of *Ganoderma boninense* with cellulase and xylanase cocktail do not give bad influence the environment and therefore provides a glimmer of hope to environmentalist.

REFERENCES

- Abe, H., Murata, Y., Kubo, S., Watanabe, K., Tanaka, R., Sulaiman, O., Hashim, R., Ramle, S. F. M., Zhang, C. & Noshiro, S. (2013). Estimation of the ratio of vascular bundles to parenchyma tissue in oil palm trunks using NIR Spectroscopy. *BioResources*. 8(2).1573-1581.
- Abdel-Monem, O. A., El-Baz, A. F., Shetaia, Y. M. & El-Sabbagh, S. M. (2012). Production and application of thermostable cellulase-free xylanase by *Aspergillus fumigatus* from agricultural wastes. *Industrial Biotechnology*. 8. 152-161.
- Abdullah, A., Shakaff, A. M., Adom, A., Ahmad, M., Zakaria, A., Ghani, S., Samsudin, N., Saad, F., Kamarudin, L. & Hamid, N. (2012). P2. 1.7 Exploring MIP Sensor of Basal Stem Rot (BSR) disease in Palm Oil Plantation. *Tagungsband*. 1348-1351.
- Acida, O. D. H. (2010). Optimization of acid hydrolysis of bagasse from agave tequilana weber. *Revista Mexicana de Ingeniería Químic*.9. 91-97.
- Ahamed, A. & Vermette, P.(2008). Enhanced enzyme production from mixed cultures of *Trichoderma reese*i RUT-C30 and *Aspergillus niger* LMA grown as fed batch in a stirred tank bioreactor. *Biochemical Engineering Journal*.42. 41-46.
- Alam, M. Z., Mamun, A. A., Qudsieh, I. Y., Muyibi, S. A., Salleh, H. M. & Omar, N. M. (2009). Solid state bioconversion of oil palm empty fruit bunches for cellulase enzyme production using a rotary drum bioreactor. *Biochemical Engineering Journal*. 46. 61-64.
- Amin, M., Soom, R., Ahmad, I. & Lian, H. (2006). Carboxymethyl cellulose from palm oil empty fruit bunch-their properties and use as a film coating agent. *Jurnal Sains Kesihatan Malaysia*. 4. 53-62.
- Amir, I., Anwar, Z., ZAfar, Y., Hussain, I., Muhammad, A., Irshad, M. & Mehmood,S. (2011). Optimization of cellulase enzyme production from corn cobs using

Alternaria alternata by solid state fermentation. Journal of Cell andMolecularBiology. 9. 51-56.

- Anis, M., Siti Nadrah, A., Kamarudin, H., Astimar, A. & Mohd Basri, W. (2011).
 Isolation and functional properties of hemicelluloses from oil palm trunks.
 Journal of Oil Palm Research. 23. 1178-1184.
- Bansal, N., Tewari, R., Gupta, J. K., Soni, R. & Soni, S. K. (2011). A novel strain of *Aspergillus niger* producing a cocktail of hydrolytic depolymerising enzymes for the production of second generation biofuels. *BioResources*. 6. 552-569.
- Bakar, N. K. A., Zanirun, Z., Abd-Aziz, S., Ghazali, F. M. & Hassan, M. A. (2012). Production of fermentable sugars from oil palm empty fruit bunch using crude cellulase cocktails with *Trichoderma asperellum* UPM1 and *Aspergillus fumigatus* UPM2 for bioethanol production. *BioResources*. 7. 3627-3639.
- Brienzo, M., Arantes, V. & Milagres, A. M. (2008). Enzymology of the thermophilic ascomycetous fungus *Thermoascus aurantiacus*. *Fungal Biology Reviews*. 22. 120-130.
- Brijwani, K., Oberoi, H. S. & Vadlani, P. V. (2010). Production of a cellulolytic enzyme system in mixed-culture solid-state fermentation of soybean hulls supplemented with wheat bran. *Process Biochemistry*, 45, 120-128.
- Bivi, M. R., Farhana, M., Khairulmazmi, A. & Idris, A.(2010). Control of *Ganoderma boninense:* A causal agent of basal stem rot disease in oil palm with endophyte bacteria in vitro. *International Journal of Agriculture and Biology*. 12. 833-839.
- Bahrin, E. K., Seng, P. Y. & Abd-Aziz, S. (2011). Effect of oil palm empty fruit bunch particle size on cellulase production by *Botryosphaeria* sp. under solid state fermentation. *Australian Journal of Basic and Applied Science*. 5(3). 276-280.
- Chai Chu Chia. Enhanced Production of Lignin Peroxidase and Manganese Peroxidase by Phanerochaete chrysosporium in a Submerged Culture

Fermentation and Their Application in Decolorisation of Dyes. Masters of Science, Universiti Sains Malaysia, Penang.(2002).

- Chang, P., Tsai, W.-S., Tsai, C.-L. & Tseng, M.-J. (2004). Cloning and characterization of two thermostable xylanases from an alkaliphilic *Bacillus firmus. Biochemical and Biophysical Research Communications*. 319. 1017-1025.
- Chen, C.-L., Qi, W. & Wang, J.-Y. (2012). Microbial cocktail for bioconversion of green waste to reducing sugars. *Journal of Bioscience and Bioengineering*.115(1).82-85.
- Chong, K., Markus, A. & Rossall, S. (2012). The susceptibility of different varieties of oil palm seedlings to *Ganoderma boninense* infection. *Pakistan Journal of Botany.* 44. 2001-2004.
- Dahot, M. U. & Noomrio, M. H. (1996). Microbial production of cellulases by Aspergillus fumigatus using wheat straw as a carbon source. Journal of Islamic Academy of Sciences. 9. 119-124.
- Das, A., Paul, T., Halder, S. K., Jana, A., Maity, C., Das Mohapatra, P. K., Pati, B.
 R. & Mondal, K. C. (2012). Production of cellulolytic enzymes by *Aspergillus fumigatus* ABK9 in wheat bran-rice straw mixed substrate and use of cocktail enzymes for deinking of waste office paper pulp. *Bioresource technology*.128.290-296.
- De Ascensao, A. R. & Dubery, I. A.(2000). Panama disease: cell wall reinforcement in banana roots in response to elicitors from *Fusarium oxysporum* f. sp. cubense race four. *Phytopathology*. 90.1173-1180.
- Dhillon, G. S., OberoI, H. S., Kaur, S., Bansal, S. & Brar, S. K. (2011). Valueaddition of agricultural wastes for augmented cellulase and xylanase production through solid-state tray fermentation employing mixed-culture of fungi. *Industrial Crops and Products*. 34. 1160-1167.
- Duenas, R., Tengerdy, R. & Gutierrez-Correa, M. (1995). Cellulase production by mixed fungi in solid-substrate fermentation of bagasse. World Journal of Microbiology and Biotechnology .11. 333-337.

- Elliott. M. L. & Broschat. T. K. (2000). *Ganoderma* Butt Rot of Palms. *Plant Pathology Department*. pp- 54.
- El-Zawawy, W. K., Ibrahim, M. M., Abdel-Fattah, Y. R., Soliman, N. A. & Mahmoud, M. M. (2011). Acid and enzyme hydrolysis to convert pretreated lignocellulosic materials into glucose for ethanol production. *Carbohydrate Polymers*. 84. 865-871.
- Flood, J.,Cooper. R., Rees. R., Potter. U. &Hasan. Y. (2010). Some latest R&D on Ganoderma Diseases in oil palm. IOPRI/ MPOB Seminar: Advances In The Controlling of Devastating Disease of Oil Palm (Ganoderma). May 2010. South East Asia, Jogjakarta, Indonesia, 1-21.
- Guimarães, L. H. S., Peixoto-Nogueira, S. C., Michelin, M., Rizzatti, A. C. S., Sandrim, V. C., Zanoelo, F. F., Aquino, A. C. M., Junior, A. B. & Polizeli, M. D. L. (2006). Screening of filamentous fungi for production of enzymes of biotechnological interest. *Brazilian Journal of Microbiology*. 37. 474-480.
- Gutierrez-Correa, M. & Tengerdy, R. P.(1998). Xylanase production by fungal mixed culture solid substrate fermentation on sugar cane bagasse. *Biotechnology Letters*. 20.45-47.
- Gupta, V. K., Gaur, R., Yadava, S. K., & Darmwal, N. S. (2009). Optimization of xylanase production from free and immobilized cells of *Fusarium solani* F7. *BioResources*. 4(3). 932-945.
- Hu, H., Van Den Brink, J., Gruben, B., Wösten, H., Gu, J.-D. & DE Vries, R.(2011).
 Improved enzyme production by co-cultivation of *Aspergillus niger* and *Aspergillus oryzae* and with other fungi. *International Biodeterioration & Biodegradation*. 65. 248-252.
- Haltrich, D., Nidetzky, B., Kulbe, K. D., Steiner, W. & Župančič, S. (1996). Production of fungal xylanases. *Bioresource Technology*. 58. 137-161.
- Hamisan, A., Abd-Aziz, S., Kamaruddin, K., Shah, U. M., Shahab, N. & Hassan, M. (2009). Delignification of oil palm empty fruit bunch using chemical and microbial pretreatment methods. *International Journal Agricultural Resources.* 4. 250-256.

- Howell, C. (2003). Mechanisms employed by *Trichoderma* species in the biological control of plant diseases: the history and evolution of current concepts. *Plant disease*. 87. 4-10.
- Ibrahim, C. (2006). Xylanase production by a local isolate, *Trichoderma* spp. FETL c3-2 via solid state fermentation using agricultural wastes as substrates.
- Jecu, L. (2000). Solid state fermentation of agricultural wastes for endoglucanase production. *Industrial Crops and Products*. 11. 1-5.
- Jelani. A. R., Hitam. A., Khalid. R., Seman. I.A., Shuib. A.B., Aminulrashid & Ismail.F.(2004).Mechanical trunk injection for control of *Ganoderma*. *Malaysia Palm Oil Board Information Series* no 215. ISSN no.1511-7871.
- Kamble, R. D. & Jadhav, A. R.(2012). Isolation, purification, and characterization of xylanase produced by a new species of *Bacillus* in solid state fermentation. *International journal of microbiology*.2012.
- Kapilan, R. & Arasaratnam, V. (2011). Paddy husk as support for solid state fermentation to produce xylanase from *Bacillus pumilus*. *Rice Science*. 18. 36-45.
- Kapoor, M., Nair, L. M. & Kuhad, R. C.(2008). Cost-effective xylanase production from free and immobilized *Bacillus pumilus* strain MK001 and its application in saccharification of *Prosopis juliflora*. *Biochemical Engineering Journal*. 38. 88-97.
- Karthikeyan, M., Radhika, K., Mathiyazhagan, S., Bhaskaran, R., Samiyappan, R. & Velazhahan, R. (2006). Induction of phenolics and defense-related enzymes in coconut (*Cocos nucifera L.*) roots treated with biocontrol agents. *Brazilian Journal of Plant Physiology*. 18. 367-377.
- Kapilan, R., & Arasaratnam, V. (2011). Paddy Husk as Support for Solid State Fermentation to Produce Xylanase from *Bacillus pumilus*. *Rice Science*. 18(1). 36-45.
- Kavya, V. & Padmavathi, T. (2009). Optimization of growth conditions for xylanase production by Aspergillus niger in solid state fermentation. Polish Journal of Microbiology.58. 125-130.

- Kheng, P. P. & Omar, I. C. (2005). Xylanase production by a local fungal isolate, *Aspergillus niger* USM AI 1 via solid state fermentation using palm kernel cake (PKC) as substrate. *Songklanakarin Journal Sciences and Technology*. 27. 325-336.
- Kim Phin Chong. The role of phenolics in the interaction between oil palm and *Ganoderma bonisense* the causal agent of basal stem rot. Degree of Philosophy, University of Nottingham. (2010).
- Lakshmi, G. S., Rao, C. S., Rao, R. S., Hobbs, P. J. & Prakasham, R. S.(2009). Enhanced production of xylanase by a newly isolated *Aspergillus terreus* under solid state fermentation using palm industrial waste: A statistical optimization. *Biochemical Engineering Journal*. 48. 51-57.
- Larkin, R. P. & Fravel, D. R. (1998). Efficacy of various fungal and bacterial biocontrol organisms for control of Fusarium wilt of tomato. *Plant disease*. 82. 1022-1028.
- Lavania, M., Chauhan, P. S., Chauhan, S., Singh, H. B. & Nautiyal, C. S. (2006). Induction of plant defense enzymes and phenolics by treatment with plant growth–promoting rhizobacteria *Serratia marcescens* NBRI1213. *Current Microbiology*. 52. 363-368.
- Lin,Y.S.,Cheng,W.C.,Chen,S.H.& Yeh.A.I. (2012). Preparation of nano/submicron Ganoderma tsugae and its stability.Journal of Food and Drug Analysis. (20)4. 900-907.
- Lelong, C. C., Roger, J.-M., Brégand, S., Dubertret, F., Lanore, M., Sitorus, N. A., Raharjo, D. A. & Caliman, J.-P. (2010). Evaluation of oil-palm fungal disease infestation with canopy hyperspectral reflectance data. *Sensors*. 10. 734-747.
- Lo, T., C. (1998). General mechanisms of action of microbial biocontrol agents. Plant Pathology Buletin. 7.155-166
- Madi, L., Katan, T., Katan, J. & Henis, Y.(1997). Biological control of Sclerotium rolfsii and Verticillium dahliae by Talaromyces flavus is mediated by different mechanisms. Phytopathology. 87. 1054-1060.

- Marli, C. (2012). Cellulase determination: Modifications to make the filter paper assay easy, fast, practical and efficient. *Journal of Analytical & Bioanalytical Techniques*.1.125
- Mohamed, W. Z. & Alimon, A.R. (2012). Recent advances in the utilization of oil palm by-products as animal feed. International Conference On Livestock Production and Veterinary Technology (ICARD).01-04 October. Ciawi, Bogor, Indonesia, 211-219
- Mojsov, K. (2010). Application of solid-state fermentation for cellulase enzyme production using *Trichoderma viride*. *International Cross Industry Journal*.5(2). 108-110.
- Nair, S. G., Sindhu, R. & Shashidhar, S. (2008). Fungal xylanase production under solid state and submerged fermentation conditions. *African Journal of Microbiology Research*. 2. 82-86.
- Najafpour, G., Ideris, A., Salmanpour, S. & Norouzi, M.(2007). Acid hydrolysis of pretreated palm oil lignocellulosic wastes. *IJE Transactions*. 20. 147-156.
- Naher, L., Tan, S. G., Yusuf, U. K., Ho, C. L. & Siddiquee, S.(2012). Activities of chitinase enzymes in the oil palm (*Elaeis guineensis* Jacq.) in interactions with pathogenic and non-pathogenic fungi. *Plant Omics.* 5. 333.
- Norhamly bin Mohd Nor. Production of Xylanase enzyme from *Aspergillus niger* using sugarcane baggase; the effect of substrate concentration. Degree of Chemical Engineering, Kolej Universiti Kejuruteraan & Teknologi Malaysia. (2006).
- Ncube, T., Howard, R. L., Abotsi, E. K., Van Rensburg, E. L. J. & Ncube, I.(2012). Jatropha curcas seed cake as substrate for production of xylanase and cellulase by Aspergillus niger FGSCA733 in solid-state fermentation. Industrial Crops and Products. 37.118-123.
- Norazlina, I., Meenalosani, N. & Ku Halim, K. (2013). Production of Xylanase by *Trichoderma* sp. Via Solid State Culture Using Sugarcane Bagasse. *International Journal of Energy Science*. 3(2).99-105.

- Nurul Kartini, A., Zanirun, Z., Abd-Aziz, S., Ghazali, F. M. & Hassan, M. A. (2012). Production of fermentable sugars from oil palm empty fruit bunch using crude cellulase cocktails with *Trichoderma asperellum* UPM1 and *Aspergillus fumigatus* UPM2 for bioethanol production. *BioResources*. 7. 3627-3639.
- Ojumu, T. V., Solomon, B. O., Betiku, E., Layokun, S. K. & Amigun, B.(2003). Cellulase Production by Aspergillus flavus linn isolate NSPR 101 fermented in sawdust, bagasse and corncob. African Journal of Biotechnology. 2. 150-152.
- Onyeagoro, G. (2012). Influence of surface lignin concentration on fibre surface characteristics and tensile properties of oil palm fibre/urea-formaldehyde resin composite. *Academic Research International*. 3(1).491-498.
- Paterson, R. (2007). *Ganoderma* disease of oil palm—A white rot perspective necessary for integrated control. *Crop Protection*. 26.1369-1376.
- Paterson, R. R. M., Sariah, M. & Lima, N. (2013). How will climate change affect oil palm fungal diseases? *Crop Protection*. 46. 113-120
- PercivaL Zhang, Y.-H., Himmel, M. E. & Mielenz, J. R.(2006). Outlook for cellulase improvement: screening and selection strategies. *Biotechnology advances*. 24. 452-481.
- Rajmane, S. & Korekar, S. (2012). Cellulase enzyme production of post-harvest fungi under the influence of carbon and nitrogen sources. *Current Botany*. 3(2).13-15.
- Ramos, L., Zandoná Filho, A., Deschamps, F. & Saddler, J. (1999). The effect of *Trichoderma* cellulases on the fine structure of a bleached softwood kraft pulp. *Enzyme and Microbial Technology*. 24. 371-380.
- Reese, E. T. (1956). Enzymatic hydrolysis of cellulose. *Applied Microbiology*. 4. 39-45.
- SAliu, B. K. & Sani, A.(2012). Bioethanol potentials of corn cob hydrolysed using cellulases of Aspergillus niger and Penicillium decumbens. Experimental and Clinical Sciences International Journal.11.468-479.

- Siangming, K., Hunjiat, T. & Weichee, W. (2013). In vitro growth of *Ganoderma* boninense isolates on novel palm extract medium and virulence on oil palm (*Elaeis guineensis*) seedlings. *Malaysian Journal of Microbiology*. 9. 33-42.
- Saida, L., Oberoi H.S, & Narasu, M.L.(2013). Studies on cellulase production by solid state fermentation using sweet *sorghum baggase*. *Helix*.1. 261-266.
- Sánchez, C. (2009). Lignocellulosic residues: biodegradation and bioconversion by fungi. *Biotechnology advances*. 27. 185-194.
- Shahriarinour, M., Wahab .M. N. A., Mustafa. S., Mohamad. R & Ariff. A. B. (2011). Cellulase from palm waste. Bioresources. 6(1). 291-307.
- Shata, H. M.(2005). Extraction of milk-clotting enzyme produced by solid state fermentation of Aspergillus oryzae. Polish Journal of Microbiology. 54. 241-247.
- Sharaf,A.N.,Abdelkader,H.S.,El-Hadi,A.E.A.A.&Ahmed,D.S.(2008).Overexpression of rice chitinase gene: evaluation of chitinase ability as a bio-antifungal agent. *Arab Journal Biotechnology*. 12. 85-98.
- Shafique,S.,Bajwa,R.,and Shafique,S.(2007).Cellulase production potential of selected strains of Aspergilli. Pakistan Journal of Phytopathology.19(2).196-206.
- Singh, A., Singh, N. & Bishnoi, N. R.(2009). Production of cellulases by Aspergillus heteromorphus from wheat straw under submerged fermentation. *International Journal of Civil and Environmental Engineering*. 1. 23-26.
- Sitarz, A. K., Mikkelsen, J. D., Højrup, P. & Meyer, A. S. (2013). Identification of a laccase from *Ganoderma lucidum* CBS 229.93 having potential for enhancing cellulase catalyzed lignocellulose degradation. *Enzyme and Microbial Technology*. 53. 378-385.
- Sukumaran, R. K., Singhania, R. R. & Pandey, A.(2005). Microbial cellulases-Production, applications and challenges. *Journal of Scientific and Industrial Research*. 64. 832.
- Sun, Y. & Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource technology*. 83. 1-11.

- Sunna, A. & Antranikian, G. (1997). Xylanolytic enzymes from fungi and bacteria. *Critical reviews in biotechnology*. 17.39-67.
- Tabet, T. A. & Aziz, F. A. (2013). Cellulose microfibril angle in wood and its dynamic mechanical significance.Ven,T.V.D & Godbout,L. Cellulose-Fundamental Aspects (pp. 113-142).Sabah: InTechOpen
- Tengerdy, R. & Szakacs, G. (2003). Bioconversion of lignocellulose in solid substrate fermentation. *Biochemical Engineering Journal*. 13. 169-179.
- Thakker, J. N., PateL, S. & Dhandhukia, P. C.(2012). Induction of defense-related enzymes in banana plants: Effect of live and dead pathogenic strain of *Fusarium oxysporum* f. sp. *cubense*. *ISRN Biotechnology*. 2013.(601303).1-6.
- Umbrin, I., Abdul, M., Khalid, H., Khalid, N., Skakil, A. & Muhammad, N.(2011). Solid state fermentation of Vigna mungo for cellulase production by *Aspergillus niger. World Applied Sciences Journal.* 12. 1172-1178.
- Verma, A., Kumar, S. & Jain, P. (2011). Key pretreatment technologies on cellulosic ethanol production. *Journal of Sciences Resources*. 55. 57-63.
- Vinale, F., Sivasithamparam, K., Ghisalberti, E. L., Marra, R., Woo, S. L. & Lorito, M. (2008). *Trichoderma*-plant-pathogen interactions. *Soil Biology and Biochemistry*. 40 (2008).1-10.
- Virupakshi, S., Babu, K. G., Gaikwad, S. R. & Naik, G.(2005a). Production of a xylanolytic enzyme by a thermoalkaliphilic *Bacillus* sp. JB-99 in solid state f ermentation. *Process Biochemistry*. 40(2005).431-435.
- Virupakshi, S., Babu, K. G., Gaikwad, S. R. & Naik, G. R. (2005b). Production of a xylanolytic enzyme by a thermoalkaliphilic *Bacillus* sp.JB-99 in solid state f ermentation. *Process Biochemistry*. 40(2005).431-435.
- Wahid, M. Z. A., Salleh, M., Yusof, F., Karim, M. I. A. & Alam, M. Z. (2011).
 Factors affecting endoglucanase production by *Trichoderma reesei* RUT C-30 from solid state fermentation of oil palm empty fruit bunches using Plackett-Burman design. *African Journal of Biotechnology*. 10.9402-9409.

- Wei, Y., Van Houten, R. T., Borger, A. R., Eikelboom, D. H., & Fan, Y. (2003). Minimization of excess sludge production for biological wastewater treatment. *Water Research*. 37(18). 4453-4467.
- Yang, B., Dai, Z., Ding, S.-Y. & Wyman, C. E. (2011). Enzymatic hydrolysis of cellulosic biomass. *Biofuels*. 2. 421-450.
- Zhu, D., Wu, Q. & Wang, N. (2011). 3.02 Industrial Enzymes. *In:* Moo-Young, M. (ed.) *Comprehensive Biotechnology (Second Edition)*. Burlington: Academic Press.
- Zakaria, L., Ali, N., Salleh, B. & Zakaria, M. (2009). Molecular analysis of Ganoderma species from different hosts in peninsula Malaysia. Journal of Biological Sciences. 9. 12-20.
- Zahari, M. W., Alimon, A. & Wong, H. (2012). Utilization of oil palm co-products as feeds for livestock in Malaysia. Makkar, H.P.S.*Biofuel Co-products As Livestock Feed-Opportunities and Challenge*. (243-302). Rome: Food and Agriculture Organization Of The United Nations.