

**ISOLATION AND CHARACTERIZATION OF PHOTOSYNTHETIC
BACTERIA FROM AQUACULTURE WASTEWATER
FOR HYDROGEN PRODUCTION**

KARZAN MOHAMMED KHALID

UNIVERSITI TEKNOLOGI MALAYSIA

ISOLATION AND CHARACTERIZATION OF PHOTOSYNTHETIC BACTERIA
FROM AQUACULTURE WASTEWATER
FOR HYDROGEN PRODUCTION

KARZAN MOHAMMED KHALID

A dissertation submitted in partial fulfilment of the
requirements for the award of the degree of
Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering
Universiti Teknologi Malaysia

JUNE 2014

Dedicated to my beloved parents, siblings and beloved Haifa

ACKNOWLEDGEMENT

I am grateful to Dr. Mohd Firdaus Bin Abdul Wahab, whose supervision was effective, he has guided, hinted and motivated me throughout the work. I wish to thank and appreciate him for his patience and all his efforts.

I would like to acknowledge and extend my heartfelt gratitude to Universiti Teknologi Malaysia (UTM) in general, and to the Faculty of Biosciences and Medical Engineering (FBME) in special, for giving me the chance to do this work under their management, and for providing the required items, devices and materials to accomplish such project.

I would like also to express my sincere gratitude to my lab mates, lab assistants and fellow graduate students for their encouragement, advice, and friendship.

ABSTRACT

Hydrogen is a clean energy substitute to fossil fuels, also environmentally friendly and does not lead to global warming. Photosynthetic bacteria produce hydrogen from organic compounds by an anaerobic light-dependent electron transfer process. Two hydrogen-producing photosynthetic bacteria have been isolated from aquaculture wastewater enriched with succinate (electron donor) under facultative condition. The isolated bacteria were identified as Gram negative and rod shaped bacteria. During the photohydrogen production process, hydrogen gas is produced at 30°C, initial pH of 7, and light intensity provided using white fluorescent lamp. The two isolated bacteria were then screened for hydrogen production using acetate and starch as substrate. Hydrogen production was measured using residual gas analyzer. These strains have the ability to utilize starch better than acetate as carbon sources for hydrogen production. The two strains, designated as AQ1 and AQ4, were identified to be *Pseudomonas* sp. via 16S rRNA characterization. The sequences have been deposited to GenBank with the accession number KJ854409 for *Pseudomonas* sp. AQ1 and KJ854410 for *Pseudomonas* sp. AQ4. In conclusion, the two isolates (AQ1 and AQ4) have been characterized as hydrogen producer in photohydrogen production process using starch and acetate as substrate. They have the potential to be used in simultaneous wastewater treatment and clean energy generation.

ABSTRAK

Hidrogen adalah pengganti tenaga bersih untuk bahan api fosil, mesra alam sekitar serta tidak menyumbang kepada pemanasan global. Bakteria fotosintetik menghasilkan hidrogen daripada sebatian organik oleh proses pemindahan elektron bergantung kepada cahaya dalam keadaan anaerobik. Dua bakteria penghasil hidrogen telah diasingkan daripada air sisa akuakultur yang diperkaya dengan suksinat (penderma elektron) di bawah keadaan fakultatif. Bakteria yang terpencil telah dikenal pasti sebagai Gram negatif dan berbentuk rod. Semasa proses foto-penghasilan hidrogen, gas hidrogen telah dihasilkan pada 30°C, pH 7, dibawah lampu pendarfluor putih. Bakteria terpencil itu kemudiannya telah digunakan untuk penghasilan hidrogen menggunakan asetat dan kanji sebagai substrat. penghasilan hidrogen ini diukur dengan menggunakan *Penganalisis Gas Sisa*. Strain ini mempunyai keupayaan untuk menggunakan asetat dan kanji sebagai sumber karbon bagi pengeluaran hidrogen. Strain AQ1 dan AQ4, telah dikenal pasti sebagai *Pseudomonas* sp. melalui pencirian 16S rRNA, dan telah didepositkan ke GenBank dengan *accession number* KJ854409 and KJ854410. Kesimpulannya, strain ini (AQ1 dan AQ4) telah dikenal pasti sebagai penghasil hidrogen di dalam proses penghasilan foto-hidrogen menggunakan kanji dan asetat sebagai substrat. Strain-strain ini mempunyai potensi untuk digunakan di dalam proses rawatan air sisa dan penghasilan tenaga bersih secara serentak.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	x
	LIST OF FIGURES	xi
	LIST OF APPENDICES	xii
1	INTRODUCTION	1
	1.1 Background	1
	1.2 Problem Statement	2
	1.3 The Objectives of the Study	3
	1.4 Scope of Study	3
	1.5 Significance of Study	4
2	LITERATURE REVIEW	5
	2.1 Hydrogen as Energy Source	5
	2.2 Hydrogen Production Processes	6
	2.2.1 Biological Hydrogen Production	7
	2.3 Photosynthetic Microorganisms	7
	2.3.1 Photosynthetic Bacteria (PSB)	9

2.4	Gamma Proteobacteria	10
2.5	Hydrogen Production by PSB	10
2.5.1	Biohydrogen Production using Purple Non-Sulfur Bacteria	12
2.6	Industrial Application of PSB	12
2.7	Aquaculture Wastewater	13
2.8	Growth Condition for Hydrogen Production	13
2.8.1	Light intensity	13
2.8.2	Temperature and pH	14
2.8.3	Use of Suitable Microorganisms in Hydrogen Production Processes	15
2.8.4	Carbon Sources	15
3	METHODOLOGY	17
3.1	Experimental Design	17
3.2	Media Preparation	17
3.2.1	Succinate Media	18
3.2.2	Nutrient Agar (NA)	18
3.2.3	Acetate Medium	19
3.2.4	Starch medium	19
3.3	Isolation of Bacteria	20
3.4	Screening Pigment of PSB	21
3.5	Gram Staining	21
3.5.1	Preparation of Smear	21
3.5.2	Staining	22
3.6	Starch Utilization Test	22
3.7	Hydrogen production by PSB	23
3.7.1	Bacterial Solution (Pre-Culture)	23
3.7.2	Bacterial Growth and Hydrogen Generation	23
3.7.3	Gas Collection and RGA Technique	24
3.8	Identification of PSB using 16S rRNA	24
3.8.1	Genomic DNA Extraction (DNA	24

	isolation)	
	3.8.2 Colony PCR	24
	3.8.3 Agarose Gel Electrophoresis	25
	3.8.4 Polymerase Chain Reaction	25
	3.8.5 Purification of PCR Product	26
	3.8.6 DNA Sequencing and Analysis	26
	3.9 Work Flow of Experiment	27
4	RESULTS AND DISCUSSION	28
	4.1 Isolation of Bacteria	28
	4.2 Characterization of Isolated Single Colonies	29
	4.2.1 Identification of Pigment	29
	4.2.2 Gram Staining	31
	4.2.3 Starch Utilization Test	31
	4.3 Photofermentation Process (Photohydrogen Production)	32
	4.4 16S rRNA Analysis	34
	4.4.1 Genomic DNA Extraction	34
	4.4.2 Polymerase Chain Reaction (PCR)	35
	4.4.3 NCBI BLAST	35
	4.4.4 Phylogenetic Analysis	36
5	CONCLUSION AND FUTURE WORK	38
	5.1 Conclusion	38
	5.2 Future Work	39
	REFERENCES	40
	Appendices	47-53

LIST OF TABLES

TABLE NO	TITLE	PAGE
2.1	Different bacterial groups contain different types of bacteriochlorophyll	8
3.1	Composition of succinate media	18
3.2	Composition of acetate medium	19
3.3	Composition of starch medium	20
4.1	Colony and cellular characterization of AQ1 and AQ4 bacteria	31

LIST OF FIGURES

FIGURE NO	TITLE	PAGE
3.1	Flow of the experimental work performed in this research	27
4.1	Bacterial growth in succinate medium (pH 7), after 2-3 weeks incubation at 30°C in the presence of light.	28
4.2	UV-vis spectrum for AQ1.	30
4.3	UV-vis spectrum for AQ4.	30
4.4	Starch utilization test.	32
4.5	Production of H ₂ by AQ1 and AQ4 in starch medium. Line indicates the point when the valve is open, allowing gas flow into the RGA for measurement	33
4.6	Production of H ₂ by AQ1 and AQ4 in acetate medium. Line indicates the point when the valve is open, allowing gas to flow into the RGA for measurement.	34
4.7	PCR products of AQ1 and AQ4.	35
4.8	Phylogenetic tree for AQ1 strain. Neighbour-joining method was used to construct the tree.	36
4.8	Phylogenetic tree for AQ4 strain. Neighbour-joining method was used to construct the tree.	37

LIST OF APPENDICES

APPEENDIX NO	TITLE	PAGE
1	Mixed culture contains different single colonies on NA	47
2	AQ1 single colonies	47
3	AQ2 single colonies	48
4	AQ3 single colonies	48
5	AQ4 single colonies	49
6	AQ5 single colonies	49
7	Pigment of isolated PSB (AQ1 and AQ4)	50
8	Gas collection from starch medium (Tedlar bags connected to serum bottles)	50
9	Gas collection from acetate medium (Tedlar bags connected to serum bottles)	51
10	Extracted genomic DNA of AQ4	51
11	16S rRNA partial sequence of AQ1, (1091 bp)	52
12	16S rRNA partial sequence of AQ4, (1016 bp)	52
13	BLAST result for AQ1 strain	53
14	BLAST result for AQ4 strain.	53

CHAPTER 1

INTRODUCTION

1.1 Background

Hydrogen is a cleaner fuel that can potentially substitute fossil fuels (Barbosa *et al.*, 2001; and Koku *et al.*, 2003). Hydrogen is known as an energy carrier that does not cause pollution, environmentally friendly and does not lead to global warming (Melis, 2006). Furthermore, hydrogen can be used for electricity generation (Yu; and Takahashi, 2007). The sun energy can be used by photosynthetic bacteria (PSB) for producing hydrogen. At the same time, a small amount of organic acids will be consumed (Melis, 2005; Barbosa *et al.*, 2001).

There are several parameters such as light intensity, substrate concentration, and initial pH affecting hydrogen production (Koku *et al.*, 2003). Photo hydrogen production is a well-developed technology, thus PSB uses recycled organic materials and does not need fossil fuels as input (Kondo *et al.*, 2002; Kondo *et al.*, 2006). Now the technology is rapidly developing to improve hydrogen production and efficiency of conversion (Kondo *et al.*, 2002).

There are varieties of biological ways to produce hydrogen from either water, organic acids or from biomass (Melis, 2006). Many prokaryotes with different groups have the ability to produce hydrogen. This is especially true for bacteria that belong to photosynthetic bacteria family. They are characterized by the ability to produce hydrogen from organic substrates with light dependency (Sunita, and Mitra, 1993). PSB has two different pathways of energy metabolism and the ability to live in both dark-anaerobic and light-anaerobic conditions (Lu *et al.*, 2011).

Both PSB and anaerobic bacteria have the ability to provide a photohydrogen production system by using residual carbohydrates as substrates (Barbosa *et al.*, 2001). Hyvolution is a hybrid system of dark and photofermentation bacteria, which lead to higher yield of hydrogen. In this integrated project, biomass is used by dark fermentation (thermophilic bacteria) for hydrogen production, then the products of dark fermentation is utilized for photofermentation (by photosynthetic purple non-sulfur bacteria) (Boran *et al.*, 2010; Gebicki *et al.*, 2010; and Wu *et al.*, 2010). Ordinarily obligate and facultative anaerobic bacteria such as *Clostridium*, *Escherichia* and *Enterobacter* have the ability to produce high rate of hydrogen from glucose, but the produced hydrogen will be limited due to the production of organic materials. These bacteria can also be co-cultured with photosynthetic bacteria to gain a higher yield of hydrogen from glucose (Asada *et al.*, 2006; and Asada, 2008).

1.2 Problem Statement

The production and utilization of fossil fuels causing are negative impacts on the environment. Combustion of fossil fuels leads to the emission of greenhouse gases into the atmosphere. In response, hydrogen has emerged as an attractive substitute to the conventional fuels, as the combustion product is only water. Various sources for feasible hydrogen production are being researched, and production from biological sources is of high potential. Hydrogen production by

using photosynthetic bacteria is being actively studied. They are known as the most powerful hydrogen producers, as the use of the photobiological hydrogen evolution could provide a very simple and efficient system of solar energy conversion. So, PSB with efficient hydrogen production need to be identified, from various sources.

1.3 The Objectives of the Study

1. To isolate and characterize photosynthetic bacteria (PSB) from aquaculture wastewater.
2. To screen the isolated PSB for hydrogen production.
3. To identify the photosynthetic hydrogen producer by using 16S rRNA characterization.

1.4 Scope of Study

In this study PSB were isolated from aquaculture wastewater. Isolation was performed in several steps, such as dilution, shake, and plate cultivation. Furthermore, the colonies and cells of isolated PSB were characterized and the pigment was screened. PSB were then screened for the ability to produce hydrogen. Identification of hydrogen producer was done using 16S rRNA technique.

1.5 Significance of Study

This study contributed to the knowledge on the production hydrogen from biological sources, especially via photosynthetic method. Hydrogen is a clean energy source and can be used as feedstock for some industries. Therefore, demand on hydrogen production has increased considerably in recent years. This PSB isolated in this study has the potential to be applied in photosynthetic bioreactors, such as for waste treatment, with usable energy generation.

REFERENCES

- Adessi, A., Philippis, R. D. (2014). Photobioreactor Design and Illumination Systems for H₂ Production with Anoxygenic Photosynthetic Bacteria. *International Journal of Hydrogen Energy*, 39, 3127-3141.
- Androga, D. D., Sevinç, P., Koku, H., Yücel, M., Gündüz, U., and Eroglu, I. (2014). Optimization of Temperature and Light Intensity for Improved Photofermentative Hydrogen Production using *Rhodobacter capsulatus* DSM 1710. *International Journal of Hydrogen Energy*, 39, 2472-2480.
- Asada, Y., Ohsawa, M., Naga, Y., Ishimi, K., Fulkatsu, M., Hideno, A., Wakayama, T., and Miyake, J. (2008). Re-evaluation of Hydrogen Productivity from Acetate by Some Photosynthetic Bacteria. *International Journal of Hydrogen Energy*, 33(19), 5147-5150.
- Asada, Y., Takumoto, M., Aihara, Y., Oku, M., Ishimi, K., Wakayama, T., Miyake, T., Tomyama, M., and Kohno, H. (2006). Hydrogen Production by Co-Cultures of *Lactobacillus* and a Photosynthetic Bacterium, *Rhodobacter sphaeroids* RV. *International Journal of Hydrogen Energy*, 31(11), 1509-1513..
- Bai, H., Zhang, Z., Yang, G., and Li, B., (2008). Bioremediation of Cadmium by Growing *Rhodobacter sphaeroides*: Kinetic Characteristic and Mechanism Studies. *Bioresource Technology*, 99(16), 7716–7722.
- Balat, M. (2009). Production of Hydrogen via Biological Processes. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 31(20), 1802-1812.
- Barbosa, M. J., Rocha, J. M. S., Tramper, J., and Wijffels, R. H. (2001). Acetate as a Carbon Source for Hydrogen Production by Photosynthetic Bacteria. *Journal of Biotechnology*, 85(1), 25-33.

- Basak, N., and Das, D. (2007). The Prospect of Purple Non-Sulfur (PNS) Photosynthetic Bacteria for Hydrogen Production: The Present State of the Art. *World Journal of Microbiology and Biotechnology*, 23(1), 31-42.
- Bhise., A.D. (2005). Biomimetic Approach for Synthesizing Artificial Light-Harvesting System Using Self- Assembled. *Institute of Nanotechnology*, 5-7.
- Boboescu, I. Z., Gherman, V. D., Mirel, I., Pap, B., Tengölics, R., Rákhely, G., and Maróti, G. (2014). Simultaneous Biohydrogen Production and Wastewater Treatment Based On the Selective Enrichment of the Fermentation Ecosystem. *International Journal of Hydrogen Energy*, 39(3), 1502-1510.
- Boran, E., Ozgur, E. Van der Burg, J., Yucel, M., Gunduz, V., and Eroglu, I. Biological Hydrogen Production by *Rhodobacter capsulatus* in Solar Tubular Photobioreactor. *Journal of Clean Production*, 18(2010), 529-535.
- Bryant, D. A., Costas, A. M. G., Maresca, J. A., Chew, A. G. M., Klatt, C. G., Bateson, M. M., and Ward, D. M. (2007). *Candidatus chloracidobacterium Thermophilum*: An Aerobic Phototrophic Acidobacterium. *Science*, 317(5837), 523-526.
- Cai, J., and Wang, G. (2014). Photo-biological Hydrogen Production by an acid Tolerant Mutant of *Rhodovulum sulfidophilum* P5 Generated by Transposon Mutagenesis. *Bioresource Technology*, 154, 254-259.
- Chang, J. S., Lee, K. S., and Lin, P. J. (2002). Bio-hydrogen Production with Fixed-Bed Bioreactor. *Hydrogen Energy*, 27, 1167–1176.
- Euzéby, J.P. (1997). "List of Bacterial Names with Standing in Nomenclature: a Folder Available on the Internet". *International Journal of Systematic Bacteriology*, 47(2): 590–592.
- Franchi, E., Tosi, C., Scolla, G., Della Penna, G., Rodriguez, F., and Pedroni, P. M. (2004). Metabolically Engineered *Rhodobacter sphaeroides* RV Strains for Improved Biohydrogen Photoproduction Combined with Disposal of Food Wastes. *Marine Biotechnology*, 6(6), 552-565.
- Gadhamshtetty, V., Sukumaran, A., and Nirmalakhandan, N. (2010). Photoparameters in Photofermentative Biohydrogen Production. *Critical Reviews in Environmental Science and Technology*, 41(1), 1-51.

- Gebicki, J., modigell, M., Schumacher, M., Van der Burg, j., and Roebroeck, E. Comparison of Two Reactor Concepts for Anoxygenic H₂ Production by *Rhodobacter capsulatus*. *Journal of Clean Production*, 18(2010), 536-542.
- Ghirardi, M. L., Zhang, L., Lee, J.W., Flynn, T., Seibert, M., and Greenbaum, E. (2000). Microalgae: A Green Source of Renewable H₂. *Trends in Biotechnology*, 18(12), 506–511.
- Hallenbeck, P. C., Ghosh, D., Skonieczny, M. T., and Yargeau, V. (2009). Microbiological and Engineering Aspects of Biohydrogen Production. *Indian Journal of Microbiology*, 49(1), 48-59.
- Han, H., Liu, B., Yang, H., and Shen, J. (2012). Effect of Carbon Sources on the Photobiological Production of Hydrogen using *Rhodobacter sphaeroides* RV. *International Journal of Hydrogen Energy*, 37:12167-12174.
- Han, S. K., and Shin, H. S. (2003). Bio-hydrogen Production by Anaerobic Fermentation of Food Waste. *Hydrogen Energy* 29, 569–577.
- Handbook on the Biology of Bacteria, 3–37. Edited by M. Dworkin, S. Falkow, E. Rosenberg, K. H. Schleifer and E. Stackebrandt. New York: Springer.
- He, D., Bultel, Y., Magnin, JP., Roux, C., and Willison, JC. (2005). Hydrogen Photosynthesis by *Rhodobacter capsulatus* and Its Coupling to PEM Fuel Cell. *Journal of Power Sources*, 14(1), 19–23.
- Holt, J. G., Krieg, N. R., Sneath, P. H., Staley, J. T., and Williams, S. T. (1994). *Bergey's Manual of Determinative Bacteriology*. Baltimore, MD, 152.
- Inui, T., Anpo, M., Izui, K., Yanagida, S., and Yamaguchi, T. (Eds.). (1998). *Advances in Chemical Conversions for Mitigating Carbon Dioxide*. Elsevier.
- Kahveci, M. I. (2007). Biohydrogen Production via Dark Fermentation. MSc Thesis, Istanbul Technical University Institute of Science and Technology, Istanbul, Turkey.
- Kahyaoglu, M., Şahin, Ö., and Saka, C. (2012). Biohydrogen Production from Waste Substrates as a Clean Energy. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 34(12), 1084-1103.
- Kantachote, D., Torpee, S., and Umsakul, K. (2005). The Potential use of Anoxygenic Phototrophic Bacteria for Treating Latex Rubber Sheet Wastewater. *Electronic Journal of Biotechnology*, 8(3), 0-0.

- Kantachote, D.; Salwa, T.; Kamontam, U., (2005). The Potential Use of Anoxygenic Photosynthetic Bacteria for Treating Latex Rubber Sheet Wastewater. *Electronic Journal of Biotechnology*, 8(3), 314-323.
- Kapdan, I. K., and Kargi, F. (2006). Bio-hydrogen Production from Waste Materials. *Enzyme and Microbial Technology*, 38(5), 569-582.
- Kerstens, K., De Vos, P., Gillis, M., Swings, J., Vandamme, P., and Stackebrandt, E. (2006). Introduction to the Proteobacteria. *The Prokaryotes: Proteobacteria: Alpha and Beta Subclasses*, 5, 3-37.
- Khatipove, E., Miyake, M., Miyake, J., and Asada, Y. (1998). Accumulation of Poly-B-Hydroxybutyrate by *Rhodobacter Sphaeroides* on Various Carbon and Nitrogen Substrates. *Federation of European Microbiological Societies Microbiology Letters*, 162, 39-45.
- Kim, D. H., and Kim, M. S. (2011). Hydrogenases for Biological Hydrogen Production. *Bioresource Technology*, 102(18), 8423-8431.
- Koku, H., Eroglu, I., Gunduz, U., Yucel, M., and Turker, L. (2003). Kinetics of Biological Hydrogen Production by the Photosynthetic Bacterium *Rhodobacter sphaeroides* O U 001. *International Journal of Hydrogen Energy*, 28(4), 381-388.
- Kondo, T., Arakawa, M., Wakayama, T., and Miyake, J. (2002). Hydrogen Production by Combining Two Types of Photosynthetic Bacteria with Different Characteristics. *International Journal of Hydrogen Energy*, 27(11), 1303-1308.
- Kondo, T., Arakawa, M., Wakayama, T., and Miyake, J. (2006). Efficient Hydrogen Production using a Multi-Photobioreactor and a Photosynthetic Bacterium Mutant with Reduced Pigment. *International Journal of Hydrogen Energy*, 31, 1522-1526.
- Kosamu, I. B. M.; Obst, M. (2009). The Influence of Picocyanobacterial Photosynthesis on Calcite Precipitation. *International Journal of Environmental Science and Technology*, 6(4), 557-562.
- Li, D., and Chen, H. (2007). Biological Hydrogen Production from Steam-Exploded Straw by Simultaneous Saccharification and Fermentation. *International Journal of Hydrogen Energy*, 32(12), 1742-1748.
- Lo, S. C., Shih, S. H., Chang, J. J., Wang, C. Y., and Huang, C. C. (2012). Enhancement of Photoheterotrophic Biohydrogen Production at Elevated

- Temperatures by the Expression of a Thermophilic Clostridial Hydrogenase. *Applied Microbiology and Biotechnology*, 95(4), 969-977.
- Lu, H., Zhang, G., and Dong, S. (2011). Quantitative Study of PNSB Energy Metabolism in Degrading Pollutants under Weak Light-Micro Oxygen Condition. *Bioresource Technology*, 102(8), 4968-4973.
- Lu, H., Zhang, G., Wan, T., and Lu, Y. (2011). Influences of Light and Oxygen Conditions On Photosynthetic Bacteria Macromolecule Degradation: Different Metabolic Pathways. *Bioresource Technology*, 102(20), 9503-9508.
- Luo, W., Deng, X., Zeng, W., and Zheng, D. (2012). Treatment of Wastewater from Shrimp Farms using a Combination of Fish, Photosynthetic Bacteria, and Vegetation. *Desalination and Water Treatment*, 47(1-3), 221-227.
- Madigan M T, Martinko J M, Dunlap P V, Clark D P. (2009). Brock Biology of Microorganisms 12-th edition, 453-454.
- Madukasi, E.I., Chunhua, H. and Zhang. G. (2011). Isolation and Application of a Wild Strain Photosynthetic Bacterium to Environmental Waste Management. *International Journal of Environmental Science and Technology*, 8(3), 513-522.
- Madukasi, E.I., Dai, X., He, C., and Zhou, J., (2010). Potentials of Phototrophic Bacteria in Treating Pharmaceutical Wastewater. *International Journal of Environmental Science and Technology*, 7(1), 165–174.
- Masset, J., Calusinska, M., Hamilton, C., Hiligsmann, S., Joris, B., Wilmotte, A., and Thonart, P. (2012). Fermentative Hydrogen Production from Glucose and Starch using Pure Strains and Artificial Co-Cultures of *Clostridium* spp. *Biotechnology for Biofuels*, 5(1), 35.
- Melis, A. 2002. Green alga hydrogen production: Progress, Challenges and Prospects. *International Journal of Hydrogen Energy*, 27(11), 1217–1228.
- Melis, A., and Melnicki, M. R. (2006). Integrated Biological Hydrogen Production. *International Journal of Hydrogen Energy*, 31(11), 1563-1573.
- Nath, K., Kumar, A., and Das, D. (2005). Hydrogen Production by *Rhodobacter sphaeroides* Strain OU 001 using Spent Media of *Enterobacter cloacae* Strain DM11. *Applied Microbiology and Biotechnology*, 68(4), 533-541.

- Ni, M., Leung, D. Y., Leung, M. K., and Sumathy, K. (2006). An Overview of Hydrogen Production from Biomass. *Fuel Processing Technology*, 87(5), 461-472.
- Oh, Y. K., Park, M. S., Seol, E. H., Lee, S. J., & Park, S. (2003). Isolation of Hydrogen-Producing Bacteria from Granular Sludge of an Upflow Anaerobic Sludge Blanket Reactor. *Biotechnology and Bioprocess Engineering*, 8(1), 54-57.
- Otsuki, T., Uchiyama, S., Fujiki, K., Fukunaga, S., (1998). Hydrogen Production by a Floating-Type Photobioreactor. In: Zaborsky, O.R. (Ed.), *Biohydrogen*. Plenum Press, London, 369–374.
- Prasertsan, P., Jaturapnripat, M. and Siripatana, C. (1997). Utilization and Treatment of Tuna Condensate by Photosynthetic Bacteria. *Pure and Applied Chemistry*, 69(11), 2438-2445.
- Proctor, L. M. (1997). Nitrogen-fixing, Photosynthetic, Anaerobic Bacteria Associated With Pelagic Copepods. *Aquatic Microbial Ecology*, 12(2), 105-113.
- Sangeetha, R., and Karunanithi, T. (2010). Optimization of Growth of *Rhodobacter Sphaeroides* using Mixed Volatile Fatty Acids by Response Surface Methodology. *International Journal of Chemical Engineering*, 3, 115-118.
- Saxena, R. C., Adhikari, D. K., and Goyal, H. B. (2009). Biomass-based Energy Fuel through Biochemical Routes: A Review. *Renewable and Sustainable Energy Reviews*, 13(1), 167–178.
- Shi, X. Y., and Yu, H. Q. (2006). Conversion of Individual and Mixed Volatile Fatty Acids to Hydrogen by *Rhodospseudomonas capsulata*. *International Biodeterioration and Biodegradation*, 58(2), 82-88.
- Sunita, M., and Mitra, C.K. (1993). Photoproduction of Hydrogen by Photosynthetic Bacteria from Sewage and Wastewater. *Journal of Bioscience*, 18(1), 155-160.
- Sutcliffe, I. C. (2010). A Phylum Level Perspective on Bacterial Cell Envelope Architecture. *Trends in Microbiology*, 18(10), 464-470.
- Vijayaraghavan, K., and Soom, M. A. M. (2004). Trends in Biological Hydrogen Production—A Review. *International Journal of Hydrogen Energy*.

- Vogl, K., Tank, M., Orf, G. S., Blankenship, R. E., and Bryant, D. A. (2012). Bacteriochlorophyll f: Properties of Chlorosomes Containing the “Forbidden Chlorophyll”. *Frontiers in Microbiology*, 3: 1-12.
- Wakayama, T., and Miyake, J. (2001). Hydrogen from Biomass. In: Biohydrogen II: An Approach to Environmentally Acceptable Technology, Miyake, J., Matsunaga, T., and San Pietro, A. (Eds.). New York: Pergamon, 3–32.
- Wang, Y. Z., Liao, Q., Zhu, X., Li, J., and Lee, D. J. (2011). Effect of Culture Conditions on the Kinetics of Hydrogen Production by Photosynthetic Bacteria in Batch Culture. *International Journal of Hydrogen Energy*, 36(21), 14004-14013.
- Weinel, C., Nelson, K. E., and Tümmler, B. (2002). Global Features of the *Pseudomonas putida* KT2440 Genome Sequence. *Environmental Microbiology*, 4(12), 809-818.
- Winter, C. J. (2005). Into the Hydrogen Energy Economy—Milestones. *International Journal of Hydrogen Energy*, 30(7), 681-685.
- Wu, X., Wang, X., Yang, H., and Gue, L. (2010). A Comparison of Hydrogen Production among Three Photosynthetic Bacteria Strains. *International Journal of Hydrogen Energy*, 35(13), 7194-7199.
- Yu, J., and Takahashi, P. (2007). Biophotolysis-based Hydrogen Production by Cyanobacteria and Green Microalgae. *Communicating Current Research and Educational Topics and Trends in Applied Microbiology*, 1, 79-89.