# BIODEGRADATION OF FRESH PALM OIL MILL EFFLUENT AND FUNGAL BIOMASS PROTEIN RECOVERY BY ASPERGILLUS NIGER AND TRICHODERMA VIRENS

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A thesis submitted in fulfilment of the requirements for the award of the degree of Master of Engineering (Bioprocess)

Faculty of Chemical Engineering Universiti Teknologi Malaysia

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To my beloved father and mother;

Jalaludin bin Hj Md Yusof and Sadaah Muhammad

Brother and sisters;

For their love, support and best wishes

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#### **ABSTRACT**

This study was conducted to evaluate the potential of filamentous fungi for fresh palm oil mill effluent (POME) biodegradation. Two strains of filamentous fungi namely Aspergillus niger and Trichoderma virens were utilized and their performance in fresh POME biodegradation were studied and compared. The results of preliminary study showed that A. niger demonstrated high capacity in enhancing the biodegradation process of fresh POME in terms of reduction of total suspended solids (TSS) (340 mg/l), turbidity (110 NTU) and chemical oxygen demand (COD) (3200 mg/l) than *T. virens* (TSS: 720 mg/l; turbidity: 224 NTU; COD: 3520 mg/l). Additionally, considerable amount of biosolids enriched with fungal biomass corresponding to higher protein yield was achieved in fungal treated fresh POME as greater amount was obtained in fresh POME treated by A. niger (13.9 g/l/3.09 g/l) as compared to T. virens (9.95 g/l/2.33 g/l). Effects of inoculum size (%), agitation rate (rpm) and temperature (°C) on the biodegradation process of the fresh POME by both fungal strains were examined using response surface methodology (RSM). Fresh POME treated by A. niger was more predictable compared to T. virens as high reduction of TSS and specific resistance to filtration (SRF) with greater recovery of biosolids enriched with fungal biomass (50.40 g/l) and protein (12.9 g/l) were achieved at optimal operating conditions. Monod kinetic model was utilized to define the biodegradation kinetic of fresh POME by both strains. High maximum specific growth rate  $(\mu_{max})$  (0.81 h<sup>-1</sup>) revealed in fresh POME treated by A. niger thus signify that A. niger has the fastest growth rate and thus would be most effective in reducing the pollutant levels of fresh POME compared to T. virens (0.009 h<sup>-1</sup>).

#### **ABSTRAK**

Kajian ini dijalankan untuk menilai potensi kulat filamen untuk biodegradasi efluen kilang kelapa sawit (POME) segar. Dua jenis kulat filamen iaitu Aspergillus niger dan Trichoderma virens telah digunakan dan prestasi mereka dalam biodegradasi POME segar telah dikaji dan dibandingkan. Hasil kajian awal menunjukkan bahawa A. niger menunjukkan kapasiti yang tinggi dalam meningkatkan proses biodegradasi POME segar dari segi pengurangan jumlah pepejal terampai (TSS) (340 mg/l), kekeruhan (110 NTU) dan permintaan oksigen kimia (COD) (3200 mg/l) berbanding kadar penyingkiran daripada T. virens (TSS: 720 mg/l; kekeruhan: 224 NTU; COD: 3520 mg/l). Selain itu, sejumlah besar biopepejal yang kaya dengan biomas kulat sepadan dengan hasil protein yang lebih tinggi telah dicapai dalam POME segar dirawat dengan kulat dengan jumlah yang lebih besar telah diperolehi dalam POME segar dirawat oleh A. niger (13.9 g/l/3.09 g/l) berbanding T. virens (9.95 g/l/2.33 g/l). Kesan saiz inokulum (%), kadar campuran (rpm) dan suhu (°C) pada proses biodegradasi POME segar menggunakan kedua-dua jenis kulat telah diperiksa menggunakan kaedah permukaan sambutan (RSM). POME segar dirawat oleh A. niger adalah lebih mudah diramalkan berbanding T. virens dengan pengurangan TSS dan rintangan khusus untuk penapisan (SRF) adalah lebih tinggi dan juga penghasilan biopepejal yang kaya dengan biomas kulat (50.40 g/l) dan protein (12.9 g/l) telah dicapai pada keadaan yang optima. Model kinetik Monod telah digunakan untuk menentukan kinetik biodegradasi daripada POME segar oleh kedua-dua jenis kulat. Kadar pertumbuhan khusus maksimum ( $\mu_{max}$ ) (0.81 h<sup>-1</sup>) yang tinggi yang ditunjukkan oleh POME segar dirawat oleh A. niger menunjukkan bahawa A. niger mempunyai kadar pertumbuhan paling pesat dan oleh itu berkesan dalam mengurangkan tahap pencemar akibat daripada POME jika dibandingkan dengan T. virens (0.009 h<sup>-1</sup>).

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### LIST OF ABBREVIATIONS

BOD - Biochemical Oxygen Demand

BSA - Bovine serum albumin

CCD - Central composite design

COD - Chemical Oxygen Demand

CPO - Crude Palm Oil

CuSO<sub>4</sub>.5H<sub>2</sub>O - Copper (II) sulphate pentahydrate

DOE - Department of Environmental

EFB - Empty Fruit Bunch

FBP - Fungal Biomass Protein

FFB - Fresh Fruit Bunch

NaCO<sub>3</sub> - Sodium carbonate

NaOH - Sodium hydroxide

POME - Palm Oil Mill Effluent

POMS - Palm Oil Mill Sludges

PPNJ - Persatuan Perladangan Negeri Johor

RSM - Response surface methodology

SRF - Specific Resistance to Filtration

TSS - Total Suspended Solid

VSS - Volatile Suspended Solid

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## LIST OF SYMBOLS

<sup>o</sup>C - Degree Celcius

μm - micrometer

g - gram L - Liter

m<sup>3</sup> - Cubic meter

mg - miligram mL - mililitre

v/v - volume per volume

w/w - weight per weight

 $\mu$  - growth rate constant

S - substrate concentration

Ks - substrate saturation constant

 $\mu_{max}$  - maximum growth rate

X - biomass

R<sup>2</sup> - determination of coefficient

 $\beta_i$  - linear coeffcient

 $\beta_{ii}$  - quadratic coefficient

 $\beta_{ij}$  - interaction coefficient

 $Y_{x/s}$  - yield coeffcient

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#### **CHAPTER 1**

### INTRODUCTION

## 1.1 Research background

The oil palm fruit processing not only produce crude palm oil as a major product but also generates large quantities of liquid wastes which results in unmanageable environmental pollution. Moreover, the industrial processes can also lead to negative environmental impacts which can considerably cause climate change, loss of natural resources, air and water pollution and extinction of species. Besides, palm oil mill effluent (POME) is the highest single polluter of Malaysian rivers. An estimated of 30 million tons of POME are produced annually from more than 300 palm oil mills in Malaysia. POME discharge and the problems associated with it are already alarming and dangerously keep growing. Pollution problems caused by POME relate mainly to its oxygen depleting effects. It was found that the oxygen depleting potential of POME is 100 times greater than that of domestic sewage (Singh *et al.*, 2010).

Currently, the most common method adopted by most palm oil processing-mills is biological treatment also identified as ponding system. The ponding system requires large area of land for their series of ponds so that the desired characteristics for the discharge are achieved (Hassan *et al.*, 2004). In addition, vast amount of corrosive and odorous biogas containing methane, carbon dioxide, and a very small amount of hydrogen sulphide are generated which can adversely affect the environment (Rupani *et al.*, 2010). Moreover, this commonly used conventional

method is unable to meet the guidelines set by the Department of Environmental (DOE) especially in terms of COD and TSS concentration.

Previous researches had found that filamentous fungi have been utilized to treat wastewater ingredients such as metals, inorganic nutrients and organic compounds (Karim and Kamil., 1989; Pechsuth *et al.*, 2001., Alam *et al.*, 2003; Santos and Linardi, 2004; Rodríguez-Couto, 2009) Therefore, implementation of filamentous fungi for POME treatment could help ease the maintenance, monitoring and land requirement required by the conventional ponding system. Since typical ponding system requires longer hydraulic retention time where it involves higher operating costs, hence, by applying filamentous fungi to treat POME it is believed that the retention time required for treatment can be considerably reduced. Moreover, as compared to the conventional treatment method, it may offer better quality final effluent, less usage of chemicals, low energy consumption and low maintenance and operating costs since filamentous fungi can be easily obtained in nature.

Treatment of POME does generate greater amount of biosolids/ sludge thus proper management of the biosolids is vital in order to prevent loss of effective treatment capacity as well as to alleviate public health risks due to unsafe disposal. Since POME is non toxic therefore reuse of the biosolids for variety of applications are considered safe and would also be beneficial in terms of economic. Moreover, POME is rich with high biodegradable content of various suspended components of complex compounds such as proteins, fats, starch, cellulose, hemicelluloses, free organic acids and minor organic and mineral elements making them ideal substrates for production of fungal biomass protein. The fungal biomass protein could be converted into marketable products such as human and animal feed and may compensate the operating costs of the treatment process. Fortunately, the fungal biomass contains substantial quantities of nutrients such as carbohydrates, lipids, minerals and proteins thus making them also advantageous to be used as aquaculture feed ingredients. Furthermore, due to its high value proteins and fibres, fungal biomass could possibly be applied in the food sectors as human and animal foodstuffs replacing traditional plant or animal sources (Moore and Chiu, 2001).

In addition, kinetic studies play a major role in providing useful information either chemical or biological reactions progression occurred in a system (Jin *et al.*, 2002). In this research, the study of biodegradation kinetic is important since it is helpful in describing how the filamentous fungi work and predicting its behavior in a system. Moreover, the kinetic coefficients which will be obtained in this study will be able to illustrate the biodegradation mechanism occurred. Besides, the kinetics study can be used to describe the relationship between theoretical states of variables and explains the biodegradation system performance (Pazaouki *et al.*, 2008).

#### 1.2 Problem statement

Filamentous fungi have already been applied to treat either domestic wastewater (Alam et al., 2003; Mannan et al., 2005) or industrial wastewater (Karim and Kamil., 1989; Pechsuth et al., 2001; Garcia et al., 2000). Nonetheless, application of filamentous fungi especially fungal genera of Aspergillus and Trichoderma on fresh POME biodegradation does receive lack of attention with less information published in the literatures. It is believed that the filamentous fungi exhibit strong capability to treat wastewater with very high organic loadings. Furthermore, the current method of ponding system for treatment of POME does have some drawbacks such as large of space requirement, long retention time of process treatment and also generates vast amount of odorous gas which may adversely affect the environment. Moreover, existing alternatives techniques for POME treatment based on physical and chemical treatments (Ahmad et al., 2005; Laohaprapanon et al., 2005; Agustin et al., 2008) involve high operating cost and expensive equipment. Realizing to this, an eco-friendly and low cost technique to treat fresh POME is crucial thus performing an investigation to study the effectiveness of the filamentous fungi on fresh POME biodegradation would be beneficial.

Both mixed and pure fungal cultures have been used successfully in the biodegradation of wastewater ingredients (Jianlong *et al.*, 2001; Khalaf, 2008; Singh and Dikshit, 2010; Tsekova *et al.*, 2010). The degradation of wastewater components has been investigated previously and it was concluded that most fungal species are excellent metabolizer (Okoro *et al.*, 2008). As mentioned before, there are few researches on POME treatment by filamentous fungi thus establishement the role of filamentous fungi in POME degradation by exposing POME to pure fungal cultures is important. It is crucial to determine the pontential of pure culture in application for POME degradation before implementing the investigation using mixed cultures. In this study, the ability of two pure fungal cultures, *Aspergillus niger* and *Trichoderma virens* to degrade fresh POME are demonstrated.

Management of biosolids generated during POME treatment process plays an important role in preventing loss of effective treatment capacity due to sludge accumulation. Because POME is non toxic and contains large amounts of organic matters, macro and micro-nutrients therefore safe handling and use of the biosolids/ fungal biomass protein generated for human and animal consumption would be beneficial. Additionally, it is believed that filamentous fungi exhibit high potential ability to convert the ingredients present in the wastewater into valuable end products of fungal biomass protein which may enrich with appreciable amounts of protein and amino acids that can be used as a protein-rich animal feed. Moreover, the biomass produced by filamentous fungi can be used as such without any further processing because it provides carbohydrates, lipids, minerals, vitamins and proteins (Nitayavardhana and Khanal, 2010). Furthermore, it was reported that bioconversion of wastewater sludge by fungi into value added products such as organic acids, industrial enzymes, and, biopolymers have been completed with successful results therefore may able to compensate the operating costs of the treatment process (Barnab'e et al., 2009; More et al., 2010).

Nevertheless, to the best of our knowledge, there is lack of information on aerobic biodegradation of fresh POME by filamentous fungi as well as the kinetics data compared to the anaerobic digestion. Therefore, the kinetics study of the aerobic treatment of POME by filamentous fungi should be looked into and explored by

determination of kinetic coefficients so that the biodegradation process can be understood. The knowledge of kinetic coefficients is necessary and useful for biological design control process as well as the optimization of the operational conditions (Nakhla *et al.*, 2006). Moreover, by conducting kinetics study the relation between growth rate and substrate utilization could be estimated and would be beneficial for mass balance evaluation in a wastewater treatment plant (Firozjaee *et al.*, 2011).

## 1.3 Objectives of the study

The main objective of the study is to compare the performance of two species of filamentous fungi namely *Aspergillus niger* and *Trichoderma virens* in fresh palm oil mill effluent (POME) biodegradation and fungal biomass protein recovery.

## 1.4 Scopes of the study

- Evaluation of fresh POME degradation including total suspended solids (TSS), turbidity, specific resistance to filtration (SRF) and chemical oxygen demand (COD) removal rate.
- 2. Assessment of fungal biomass and protein recovery during treatment process
- 3. Optimization of biodegradation and fungal biomass protein recovery. Parameters chosen to be optimize are inoculums size (5-20%), agitation rate (100-150 rpm) and temperature (30-36°C)
- 4. Determination of biodegradation kinetic coefficients ( $\mu_{max}$ ,  $K_s$ ,  $Y_{x/s}$ ) based on Monod's model.

#### REFERENCES

- Abdallah, R. and Taha, S. (2012). Biosorption of methylene blue from aqueous solution by nonviable *Aspergillus fumigatus*. *Chemical Engineering Journal*, 195–196: 69–76
- Abd El- Rahim, W. M., El-Ardy, O.A.A.M., and Mohammad, F. H.A. (2009). The effect of pH on bioremediation potential for the removal of direct violet textile dye by *Aspergillus niger*. *Desalination*, 249: 1206-1211
- Adebayo, G.B., Otunola, B. and Ajao, T.A. (2010). Assessment and biological treatment of effluent from textile industry. African Journal of Biotechnology, 9(49): 8365-8368
- Agamuthu, P., Tan, E. L. and Shaiful, A.A.A. (1986). Effect of Aeration and Soil Inoculum on the Composition of Palm Oil Mill Effluent (POME). *Agricultural Wastes*. 15:121-132
- Agustin, M. B., Sengpracha., W. P., and Phutdhawong, W. (2008). Electrogulation of Palm Oil Mill Effluent. *Int. J. Environ. Res. Public Health.* 5(3): 177-180
- Ahmad A.L., Ismail, S. and Bhatia, S. (2005).Ultrafiltration behavior in the treatment of agro-industry effluent: pilot scale studies. *Chem Eng Sci* .60: 5385–94.
- Ahmad, A. L., Chan, C. Y., Abd Shukor, S. R. and Mashitah M. D. (2008). Optimization of oil and carotenes recoveries from palm oil mil effluent using response surface methodology. *J Chem Technol Biotechnol*, 84:1063-1069
- Ahmad, A.L., Chong, M.F. and Bhatia, S. (2008). Population balance model (PBM) for flocculation process: simulation and experimental studies of palm oil mill effluent (POME) pretreatment. *Chemical Engineering Journal*, 140: 86-10

- Ahmad, A. L., Sumathi, S. and Hameed, B. H. (2005). Adsorption of residue oil from palm oil mill effluent using powder and flake chitosan: Equilibrium and kinetic studies. *Water Research* .39:2483–2494
- Ahmad, A.L., Ismail, S. and Bhatia, S. (2003). Water recycling from Palm oil mill Effluent (POME) using Membrane Technology. *Desalination*. 157:87-95
- Ahmad, F., Jameel, A. T., Kamarudin, M. H. and Mel, M. (2011). Study of growth kinetic and modeling of ethanol production by *Saccharomyces cerevisae*. *African Journal of Biotechnology*. 16(81): 18842-18846
- Akthar M. N. and Mohan, P. M. (1995). Bioremediation of toxic metal ions from polluted lake waters and industrial effluents by fungal biosorbent. *Curr. Sci.* 69: 1028-1030
- Akhtar, K., Khalid, A.M., Akhtar, M.W. and Ghauri, M.A. (2009). Removal and recovery of uranium from aqueous solutions by Ca-alginate. *Bioresource Technology*, 100:4551-4558
- Akhtar, Y and Ghaffar, A. (1986). Removal of NH<sub>3</sub>-N from domestic wastewater by fungi. *Biotechnology Letters*, 8 (8): 601-604
- Alam, M. Z., Fakhru'l-Razi, A. and Molla, A.H. (2003) Biosolids Accumulation and Biodegradation of Domestic Wastewater Treatment Plant Sludge by Developed Liquid State Bioconversion Process using a Batch Fermenter. *Water Research*, **37** (15): 3569–3578.
- Alam, M.Z. and Fakhru'l-Razi, A. (2003). Enhanced settleability and dewaterability of fungal treated domestic wastewater sludge by liquid state bioconversion process. *Water Research*. 37:1118–1124
- Alam, M.Z., Fakhru'l-Razi, A. and Molla, A.H. (2003). Biosolids accumulation and biodegradation of domestic wastewater treatment plant sludge by developed liquid state bioconversion process using a batch fermenter. *Water Research*. 37: 3569–3578
- Alam, M.Z., Fakhru'l-Razi, A. and Molla, A.H. (2003a). Optimization of liquid state bioconversion process for microbial treatment of domestic wastewater sludge. *Journal of Environmental Engineering Science*, 2: 299-306

- Alam, M.Z., Fakhru'l-Razi, A., Molla, A.H. and Roychoudhury, P.K., (2001). Treatment of wastewater sludge by liquid state bioconversion process. *Journal of Environmental Science Health*, 36 (7): 1237–1243.
- Amini, M., Younesi, H. and Bahramifar, N. (2009). Biosorption of nickel (II) from aqueous solution by Aspergillus niger: Response surface methodology and isotherm study. *Chemosphere*, 75:1483-1491
- Ammary, B.Y. (2004). Nutrients requirements in biological industrial wastewater treatment. *African Journal of Biotechnology*, 3 (4): 236-238
- Anand, P., Isar, J., Saran, S. and Saxena, R. K. (2006). Bioaccumulation of copper by *Trichoderma viride*. *Bioresource Technology*, 97: 1018-1020
- Anastasi, A., Spina, F., Romagnolo, A., Tigini, V., Prigione, V. and Varese, G.C. (2012). Integrated fungal biomass and activated sludge treatment for textile wastewaters bioremediation. *Bioresource Technology*, 123: 106–111
- Ardestani, F. (2011). Investigation of the Nutrient Uptake and Cell Growth Kinetics with Monod and Moser Models for *Penicillium brevicompactum* ATCC 16024 in Batch Bioreactor. *Iranica Journal of Energy & Environment*, 2 (2):117-121
- Asgher, M., Jamil, F. and Iqbal, H.M. N. (2012). Bioremediation Potential of Mixed White Rot Culture of *Pleurotus Ostreatus* IBL-02 and *Coriolus Versicolor* IBL-04 for Textile Industry Wastewater. *J Bioremed Biodegrad*, S1:007. doi:10.4172/2155-6199.S1-007
- Auterská, P. and Nová, K. (2006). Sucessful solution for high nitrogen content wastewater treatment from rendering plants. *Water Science & Technology*, 54 (10): 23-30
- Baharuddin, A. S., Hock, L.S., Yusof, M.Z.M., Rahman, N.A.A., Shah, U.K.M., Hassan, M.A., Wakisaka, M., Sakai, K. and Shirai, Y. (2010). Effects of palm oil mill effluent (POME) anaerobic sludge from 500 m<sup>3</sup> of closed anaerobic methane digested tank on pressed-shredded empty fruit bunch (EFB) composting process. *African Journal of Biotechnology*, 9 (16): 2427-2436

- Barnab'e, S., Brar, S.K., Tyagi, R.D., Beauchesne, I. and Surampalli, R.Y. (2009). Pre-treatment and bioconversion of wastewater sludge to value-added products-fate of endocrine disrupting compounds. *Science of the Total Environment*, 407: 1471-1488
- Bezerra, M.A., Santelli, R.E., Oliveira, E.P. Villar, L.S. and Escaleira, L.A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76:965-977
- Bharati, T., Kulkarni, J.H., Krishnaraj, P.U. and Alagawadi A.R. (2007). Effect of Different Carbon Sources on the Biomass of *Metarhizium anisopliae* (Ma2)\*. *Karnataka J. Agric. Sci.*, 20 (2): 310-311
- Bhatia, S., Othman, Z. and Ahmad, A.L. (2007). Pretreatment of palm oil mill effluent using *Moringa Oleifera* seeds as natural coagulant. Journal of Hazardous Material. 145: 120-126
- Borja, R., Banks, C.J. and Sinchez, E. (1996). Anaerobic treatment of palm oil mill effluent in a two-stage up-flow anaerobic sludge blanket (UASB) system. Journal of Biotechnology, 45: 125-135
- Borja-Padilla, R. and Banks, C.J. (1993). Thermophilic semi-continuous anaerobic treatment of palm oil mill effluent. *Biotechnology Letters*, 15:761-766
- Cai, W., Li, J. and Zhang, Z. (2007). The characteristics and mechanisms of phenol biodegradation by *Fusarium sp. Journal of Hazardous Materials*, 148: 38-42
- Cail, R.G. and Barford, J.P. (1985). Mesophilic semi-continuous anaerobic digestion of palm oil mill effluent. *Biomass*, 7: 287-295
- Cameron, M.D., Timofeevski, S., and Aust, S.D. (2000). Enzymology of *Phanerochaete chrysosporium* with respect to the degradation of recalcitrant compounds and xenobiotics. *Applied Microbiology and Biotechnology*, 54 (6):751–758.
- Carlsen., M., Nielsen, J. and Villadsen, J. (1995). Growth and α-amylase production by Aspergillus Oryzae during continous cultivation. *Journal of Biotechnology*, 45:81-93

- Carman, P.C. (1938). Fundamental principals of industrial filtration (A Critical Review of Present Knowledge). *Trans Inst Chem Eng*, 16:168
- Chakroun, H., Mechichi, T., Martinez, M. J., Dhouib, A. and Sayadi, S. (2010). Purification and characterization of a novel laccase from the ascomycete *Trichoderma atroviride*: Application on bioremediation of phenolic compounds. *Process Biochemistry*, 45: 507–513
- Chin, K.K., Lee, S.W. and Mohammad, H.H. (1996). A study of palm oil mill effluent treatment using a pond system. *Water Science & Technology*, 34:119-123
- Christov, L.P., van Driessel, B. and du Plessis, C.A. (1999). Fungal biomass from *Rhizomucor pusillus* as adsorbent of chromophores from a bleach plant effluent. *Process Biochemistry*, 35: 91–95
- Chow, M.C. (1991). Palm oil mill effluent analysis. Kuala Lumpur: Palm Oil Research Institute of Malaysia, p.11-8
- Cooney, M. J. and McDonald, K. A. (1993). The Monod Constant for Growth of *Candida utilis* on ammonium in continuous and batch cultures. *Biotechnol. Prog*, 9: 93-96
- Correia, R.T.P., McCue, P. Magalhães, M.M.A., Mac^edo, G.R. and Shetty, K. (2004). Production of phenolic antioxidants by the solid-state bioconversion of pineapple waste mixed with soy flour using *Rhizopus oligosporus Process Biochemistry*, 39:2167–2172
- Coulibaly, L and Agathos, S. N. (2007). Effects of *Aspergillus niger* inoculum concentration upon the kinetics of starchy wastewater pretreatment in a tanks-inseries bioreactor under transitory conditions. *Brazilian Journal of Chemical Engineering*, 24(04): 499-507
- Coulibaly, L., Gourene, G. and Agathos, S. N. (2003). Utilization of fungi for biotreatment of raw wastewaters. *African Journal of Biotechnology*, 2 (12): 620-630

- D'Annibale, A., Casa, R., Pieruccetti, F., Ricci, M. and Marabottini, R. (2004). *Lentinula edodes* removes phenols from olive-mill wastewater: impact on durum wheat (Triticum durum Desf.) germinability. *Chemosphere*, 54: 887–894
- D'Annibale, A., Quaratino, D., Federici, F. and Fenice, M. (2006). Effect of agitation and aeration on the reduction of pollutant load of olive mill wastewater by the white-rot fungus *Panus tigrinus*. *Biochemical Engineering Journal*, 29: 243–249
- D'Annibale. A., Crestini, C., Vinciguerra, V. and Sermanni, G.G. (1998). The biodegradation of recalcitrant effluents from an olive mill by a white-rot fungus. *Journal of Biotechnology*, 61: 209–218
- Dubey, S., Singh, A. and Banerjee, U.C. (2011). Response surface methodology of nitrilase production by recombinant *Escherichia coli*. *Brazillian Journal of Microbiology*, 42: 1085-1092
- Dursun, A.Y., Uslu, G., Cuci, Y. and Aksu, Z. (2003). Bioaccumulation of copper (II), lead (II) and chromium (VI) by growing *Aspergillus niger*. *Process Biochemistry*, 38: 1647-1651
- Ellouze, M., Aloui, F. and Sayadi, S. (2008). Detoxification of Tunisian landfill leachates by selected fungi. *Journal of Hazardous Materials*, 150: 642–648
- Elson, M. K., Schisler, D. A. and Jackson, M. A. (1998). Carbon-to-nitrogen ratio, carbon concentration, and amino acid composition of growth media influence conidiation of *Helminthosporium solani*. *Mycologia*, 90(3): 406–413.
- Ergűl, F.E., Sargin, S., Öngen, G. and Sukan, F.V. (2009). Dephenolisation of olive mill wastewater using adapted *Trametes versicolor*, *International Biodeterioration & Biodegradation*, 63: 1–6
- Fadil, K., Chahlaoui, A., Quahbi, A., Zaid, A. and Borja R. (2003). Aerobic biodegradation and detoxification of wastewaters from the olive oil industry. *International Biodeterioration and Biodegradation*, 51:37-41
- Fadzilah, K. and Mashitah, M. D. (2010). Cellulase production in palm oil mill effluent: Effect of aeration and agitation. *Journal of Applied Sciences*, 10 (24): 3307-3312

- Fakhru'l-Razi, A. and Molla, A. H. (2007). Enhancement of bioseparation and dewaterability of domestic wastewater sludge by fungal treated dewatered sludge. *Journal of Hazardous Material*, 147: 350-356
- Feijoo, G. and Lema, J. M. (1995). Treatment of forest industry effluents with toxic and recalcitrant compounds by the white rot fungi. *Afnidad*, 52:171-180
- Feng, X., Deng, J., Lei, H., Bai, T., Fan, Q. and Li, Z. (2009). Dewaterability of waste activated sludge with ultrasound conditioning. *Bioresource Technology*, 100: 1074–1081
- Field, J. A., de Jong, E., Feijoo Costa, G. and de Bont J. A. M. (1993). Screening for ligninolytic fungi applicable to the degradation of xenobiotics. *TiBtech*,14: 44-49.
- Firozjaee, T.T, Najafpour, G.D., Khavarpour, M., Bakshi, Z., Pishgar, R. and Mousavi, N. (2011). Phenol Biodegradation Kinetics in an Anaerobic Batch Reactor. *Iranica Journal of Energy & Environment*, 2 (1): 68-73
- Fosdick, L.E., Bohlmann, J.A., Trinkle, J.R. and Ray, B.L. *Glucosamine and Method of Making Glucosamine from Microbial Biomass*. U.S. Patent, 20110319358, 2011
- Freitas, A.C., Ferreira, F., Costa, A.M., Pereira, R., Antunes, S.C., Gonçalves, F., Rocha-Santos, T.A.P., Diniz, M.S. (2009). Biological treatment of the effluent from a bleached kraft pulp mill using basidiomycete and zygomycete fungi. *Science of the Total Environment*, 407: 3282–3289
- Friedrich, J., Cemerman, A. and Perdih, A. (1996). Comparison of different of cellulolytic fungi or bioconversion of apple distillery waste. *Applied Microbiology and Biotechnology*. 24: 243-247
- Fu, Y. and Viraraghavan, T. (2002). Removal of Congo Red from an aqueous solution by fungus *Aspergillus niger*. *Advances in Environmental Research*, 7: 239-247

- Fujita, M., Era, A., Ike, M., Soda, S., Miyata, N. and Hirao, T. (2000). Decolorization of heat-treatment liquor of waste sludge by a bioreactor using polyurethane foam-immobilized white rot fungi equipped with an ultramembrane filtration unit. *J. Biosc. Bioeng*, 90: 387-394.
- Garcia Garcia, I., Jimenez Pena, P.R., Bonilla Venceslada, J.L., Martin martin, A., Martin Santos, M.A. and Ramos Gomez, E. (2000). Removal of phenol compounds from olive mill wastewater using *Phanerochaete chrysosporium*, *Aspergillus niger*, *Aspergillus terreus* and *Geotrichum candidum*. Process Biochemistry, 35: 751–758
- Gauthier, H., Yargeau, V. and Cooper, D.G. (2010). Biodegradation of pharmaceuticals by *Rhodococcus rhodochrous* and *Aspergillus niger* by cometabolism. *Science of the Total Environment*, 408: 1701–1706
- Ghafari, S., Aziz, H. A., Isa, M. H., and Zinatizadeh A. A. (2009). Application of response surface methodology (RSM) to optimize coagulation—flocculation treatment of leachate using poly-aluminum chloride (PAC) and alum. *Journal of Hazardous Materials*, 163:650-656
- Hageskal, G., Lima, N. and Skaar, I. (2009). The study of fungi in drinking water. *Mycological research*. 113:165-172
- Hamdi, M. and Ellouz R. (1992). Use of *Aspergillus niger* to improve filtration of olive mill waste waters. *J Chem Tech Biotechnol*, 53:195-200
- Hamman, A., Brust, D. and Osiewacz, H. D. (2008). Apoptosis pathways in fungal growth, development and ageing. *Trends in Microbiology*, 16 (6): 276-283
- Han, Y.W., Lee, J.S. and Anderson, A.W. (1975). Chemical composition and digestibility of rye grass straw. *Journal of Agricultural and Food Chemistry*, 23:928-931
- Habib, M.A.B., Yusoff, F.M., Phang, S.M., Ang, K.J. and Mohamed, S. (1997).
  Nutritional values of chironomid larvae grown in palm oil mill effluent and algal culture. *Aquaculture*, 158:95-105

- Hassan, M.A., Shirai, Y., Kusubayashi, N., Karim, M.I.A., Nakanishi, K. and Hashimoto, K. (1996). Effect of organic acid profiles during anaerobic treatment of palm oil mill effluent production of polyhyroxyalkanoates by *Rhodobacter spahaeroides*. *Journal of Fermentation and Bioengineering*, (82) 2: 151-156
- Hassan, M.A., yacob, S., Shirai, Y. and Hung, Y.T. (2004). Treatment of palm oil wastewaters. In: Lawrence K. Wang, Yung-Tse Hung, Howard H. Lo, Constantine Yapijakis. *Handbook of Industrial of hazardous Waste Treatment*. New York: Marcel Dekker, Inc. 719-735
- Henze, M., Harremoes, P., Jansen, J., Arvin, E. (1997). Wastewater Treatment, second edition, *Springer*
- Ho, C.C and Tan, Y.K (1983). Centrifugal fractionation studies on the particulates of the palm oil mill effluent. *Water Research*, 17: 613-618
- Ho, C.C., Tan, Y.K. and Wang, C.W. (1984). The distribution of chemical constituents between the soluble and the particulate fractions of palm oil mill effluent and its significance on its utilisation/ treatment. *Agricultural Wastes*, 11:61-71
- Ho, C.C. and Tan, Y.K. (1988). The treatment of anaerobically digested palm oil mill effluent by pressurized activated sludge. *Journal of Chemical Technology and Biotechnology*. 41:75-84
- Imai, Y., Suzuki, M., Masamoto, M. and Nagayao, K. (1993). Amylase production by *Aspergillus Oryzae* in new kind of fermenter with a rotary draft tube. *Journal of Fermentation Bioengineering*, 76(6): 459-464
- Jackson, A.M., Whipps, J.M. and Lynch J.M. (1991). Effects of temperature, pH and water potential on growth of four fungi with disease biocontrol potential. World Journal of Microbiology and Biotechnology, 7: 494-501
- Jamal, P., Idris, Z. M. and Alam, M.Z. (2011). Effects of physicochemical parameters on the production of phenolic acids from palm oil mill effluent under liquid-state fermentation by *Aspergillus niger* IBS-103ZA. *Food Chemistry*, 124: 1595–1602

- Jiang, T. and Ting, Y. (2004). Optimisation on bioleaching of incinerator fly ash by Aspergillus niger—use of central composite design. Enzyme and Microbial Technology, 35: 444-454
- Jianlong, W., Xinmin, Z., Decai, D. and Ding, Z. (2001). Bioadsorption of lead (II) from aqueous solution by fungal biomass of *Aspergillus niger*. *Journal of Biotechnology*, 87: 273–277
- Jin, B., van Leuween, H. J., Patel, B. and Yu, Q. (1998). Utilisation of starch processing wastewater for production of microbial biomass protein and fungal α-amylase by Aspergillus oryzae. Bioresource Technology, 66:201-206
- Jin, B., Yan, X.Q., Yu, Q. and van Leeuwen, J.H. (2002). A comprehensive pilot plant system for fungal biomass protein production and wastewater reclamation. Advances in Environmental Research, 6:179-189
- Justino, C.I., Duarte, K., Loureiro, F., Pereira, R., Antunes, S.C., Marques, S.M., Goncalves, F., Rocha-Santos, T.A.P. and Freitas, A.C. (2009). Toxicity and organic content characterization of olive oil mill wastewater undergoing a sequential treatment with fungi and photo-Fenton oxidation. *Journal of Hazardous Materials*, 172: 1560–1572
- Kacprzak, M., Neczaj, E. and Okoniewska, E. (2005). The comparative mycological analysis of wastewater and sewage sludge from selected wastewater treatment plants. *Desalination*, 185: 363-370
- Kapat, A., Jung, J.K., Park, S.Y. and Hong, H.K. (1998). Effects of agitation and aeration on the production of extracellular glucose oxidase from a recombinant *Saccharomyces cerevisiae*. *Bioprocess Engineering*, 18: 347 351
- Kapoor, A. and Viraraghavan, T. (1998). Biosorption of heavy metals on *Aspergillus niger*: Effect of pretreatment. *Bioresource Technology*, 63: 109-113
- Karim, M. I. A. and Hie, L. L. (1987) The Use of Coagulating and Polymeric Flocculating Agents in The Treatment of Palm Oil Mill Effluent (POME). *Biological Wastes*, 22(3): 209-218

- Karim M.I.A. and Kamil. A .Q . A (1989). Biological treatment of POME using Thricoderma viride. Biological waste. 27: 143-152
- Kerry, B. R., Kirkwood, I. A., de Leij, Barba, J., Leijdens, M. B. and Brookes, P.A. (1993). Growth and survival of *Verticillium chlamydosporium Goddard*, a parasite of nematodes in soil. *Biocontrol Sci Technol*, 3: 355–365.
- Khalaf, M. A. (2008). Biosorption of reactive dye from textile wastewater by non-viable biomass of *Aspergillus niger* and *Spirogyra sp. Bioresource Technology*, 99: 6631–6634.
- Khalid, R. and Wan Mustafa, W.A. (1992). External benefits of environmental regulation: Resource recovery and the utilisation of effluents. *The Environmentalist*, 12: 277-285
- Kumar, V., Wati, L., Nigam, P., Yadav, B. S., Singh, D., and Marchant, R. (1998).
  Decolorization and biodegradation of anaerobically digested surgarcane molasses spent wash effluent from biomethanation plants by white-rot fungi.
  Process Biochem, 33: 83-88.
- Kumar, R., Bhatia, D., Singh, R., Rani, S. and Bishnoi, N. (2011). Sorption of heavy metals from electroplating effluent using immobilized biomass *Trichoderma viride* in a continuous packed-bed column. *International Biodeteriation & Biodegradation*, 65: 1133-1139
- Kyanko, V., Canel, R., de la Osa, O. and Ludemann, V. (2010). Filamentous fungi unconventional protein ingredients with good hydration properties. *International Conference on Food Innovation*, October 25-29, 2010. Food Innova Universidad Politecnica de Valencia.1-4
- Lacina, C., Germain, G. and Spiros, A. N. (2003). Utilization of fungi for biotreatment of raw wastewaters. African Journal of Biotechnology, 2 (12): 620-630
- Lam, M.K & Lee, K.T. (2011). Renewable and sustainable bioenergies production from palm oil mill effluent (POME): Win–win strategies toward better environmental protection. *Biotechnology Advances*, 29:124-141

- Laohaprapanon, T., Prasertsan, P. and Kittikun, A. H. (2005). Physical and Chemical Separation of Oil and Suspended Solids from POME. *Asian J. Energy Environ*, 6 (1): 39-55.
- Laws of Malaysia, 1994. Environmental Quality Act 1974 and Regulations, fourth ed. MDC Sdn. Bhd, Malaysia.
- Lee, C.H. and Liu, J.C. (2000) Enhanced Sludge Dewatering by Dual Polyelectrolytes Conditioning. *Water Research*, 34 (18): 4430-4436
- Lee, S.Y., Choi, J-i. and Wong, H. H. (1999). Recent advances in polyhdroxyalkanoate production by bacterial fermentation: mini review. *International Journal of Biological Macromolecules*, 25:31-36
- Legal Research Board (2002). Environmental Quality Act 1974 (Act 127) & Subsidiary Legislation (As at 5<sup>th</sup> June 2002). International Law Book of Services.
- Li, Y., Cui, F., Liu, Z., Xu, Y. and Zhao, H. (2007). Improvement of xylanase production by *Penicillium oxalicum* ZH-30 using response surface methodology. *Enzyme and Microbial Technology*, 40: 1381-1388
- Liu, T., Hu, H., He, Z. and Ni, Y. (2011). Treatment of poplar alkaline peroxide mechanical pulping (APMP) effluent with *Aspergillus niger*. *Bioresource Technology*, 102: 7361–7365
- Liu, Y-g., Eng, G-m,. Li, X., Ong, Q., Zhou, M., Xu, W-h., and Huang, Y-e. (2006). Removal of cadmium and zinc ions from aqueous solution by living *Aspergillus niger*. *Trans. Nonferrous Met. Soc. China*, 16:681-686
- Lopez Errasquin, E. and Vasquez, C. (2003). Tolerance and uptake of heavy metals by *Trichoderma atroviride* isolated from sludge. *Chemosphere*, 50: 137-143
- Lowry, O. H., Rosebrough, N. J., Farr, A. L., Randall, R. J. (1951). Protein measurement with the folin phenol reagent. *Journal Biol. Chem*, 193: 265-275

- Lu, Y., Yan, L., Wang, Y., Zhou, S., Fu, J. and Zhang, J. (2009). Biodegradation of phenolic compounds from coking wastewater by immobilized white rot fungus *Phanerochaete chrysosporium*. *Journal of Hazardous Materials*, 165: 1091– 1097
- M. L. and Kargi, F. (2002). *Bioprocess Engineering* (2<sup>nd</sup> ed). Prentice Hall PTR: Prentice-Hall, Inc.
- Ma, A. N. (1995). A novel treatment for palm oil mill effluent. *Palm Oil Res Instit Malaysia (PORIM)*. 29:201–12.
- Magan, N. (2007). Fungi in extreme environments C.P. Kubicek and I. S. Druzhinina (Eds.) *Environmental and Microbial Relationships*. (pp. 85-103): Springer-Verlag Berlin Heidelberg
- Mannan, S., Fakhru'l-Razi, A. and Alam, M.A. (2005) Use of Fungi to Improve Bioconversion of Activated Sludge. *Water Research*, 39: 2935–2943
- Mason, R.L., Gunst, R.F., and Hess, J.L. (2003). *Statistical Design and Analysis of Experiments*. New Jersey: John Wiley & Sons, Inc.
- Mapari, S.AS., Meyer, A.S., Thrane, U. and Frisvad, J.C. (2009). Identification of potentially safe promising fungal cell factories for the production of polyketide natural food colorants using chemotaxanomic rationale. *Microbial Cell Factories*, 8,24: doi:10.1186/1475-2859-8-24
- Mausumi, M. (2008). Role of surface properties during biosorption of copper by pretreated *Aspergillus niger* biomass. *Colloids and Surfaces A: Physicochem. Eng. Aspects*, 329: 95–99
- Md. Din, M.F., Mohanadoss, P., Ujang, Z., van Loosdrecht, M., Yunus, S.M., Chelliapan, S., Zambare, V. and Olsson, G. (2012). Development of Bio-PORec system for polyhdroxyalkanoates (PHA) production and its storage in mixed cultures of palm oil mill effluent (POME). *Bioresource Technology*, 124: 208-216

- Mendoca, E., Martins, A. and Anselmo, A. M. (2004). Biodegradation of natural phenolic compounds as single and mixed substrates by *Fusarium flocciferum*. *Electronic Journal of Biotechnology*, 7(1): 31-37
- Ming, K. K. and Chandramohan, D. (2002). Malaysian palm oil industry at crossroad and its future direction. *Oil Palm Ind. Econ. J.* 2:10-15
- Miranda, M.P., Benito, G.G., Cristobal, N.S. and Nieto, C.H. (1996). Color elimination from molasses wastewater by *Aspergillus niger*. *Bioresource Technology*, 57: 229-235
- Mishra, B.K., Arora, A. and Lata. (2004). Optimization of a biological process for treating potato chips industry wastewater using a mixed culture of *Aspergillus foetidus* and *Aspergillus niger*. *Bioresource Technology*, 94: 9–12
- Mo, M., Xu, C., and Zhang, K. (2005). Effects of carbon and nitrogen sources, carbon-to-nitrogen ratio, and initial pH on the growth of nematophagous fungus *Pochonia chlamydosporia* in liquid culture. *Mycopathologia*. 159: 381–387
- Mohd Ali Hassan, Shahrakbah Yacob, Shirai, Y. and Hung, Y. T. (2004). Treatment of Palm Oil Wastewaters. In: Lawrence K. Wang, Yung- Tse Hung, Howard H.Lo, Constantine Yapijakis. *Handbook of Industrial and Hazardous Waste Treatment*. New York: Marcel Dekker, Inc. 719-735.
- Molla, A.H., Fakhru'l-Razi, A., Abd-Aziz, S., Hanafi, M.M., Roychoudhury, P.K. and Alam, M.Z. (2002). A potential resource for bioconversion of domestic wastewater sludge. *Bioresource Technology*, 85: 263–272
- Molla, A.H., Fakhru'l-Razi, A. and Alam, M.Z., (2004). Evaluation of solid-state bioconversion of domestic wastewater sludge as a promising environmental friendly disposal technique. *Water Research*, 38 (19), 4143–4152.
- Monod, J. (1949). The growth of bacterial cultures. Ann. Rev. Microbiol. 3: 371-394.
- Montgometry, D.C. (1997). *Design and Analysis of Experiments*. New York: John Wiley and Sons.

- Moore, D. & Chiu, S. W. (2001). Fungal products as food. Chapter 10 in *Bio-Exploitation of Filamentous Fungi* (ed. S. B. Pointing & K. D. Hyde), pp. 223-251. Fungal Diversity Press: Hong Kong
- More, T.T., Yan, S., Tyagi, R. D. and Surampalli, R.Y. (2010). Potential use of filamentous fungi for wastewater sludge treatment. *Bioresourse Technology*, 101: 7691-7700
- Muhrizal, S., Shamsuddin, J., Husni, M.H.A., Fauziah, I. (2006). Alleviation of aluminium toxicity in an acid sulphate soil using organic materials. *Communications in Soil Sciences and Plant Analysis*. 34: 2993-3011
- Mungasavalli, D. P., Viraraghavan, T., Jin, Y. (2007). Biosorption of chromium from aqueous solutions by pretreated *Aspergillus niger*: Batch and column studies. *Colloids and Surfaces A: Physicochem. Eng. Aspects*, 301: 214–223
- Murugesan, G.S., Sathishkumar, M. and Swaminathan, K. (2005). Supplementation of waste tea fungal biomass as a dietary ingredient for broiler chicks. *Bioresource Technology*, 96: 1743-1748
- Muthukumar, M., Sargunamani, D., Selvakumar, N. and Venkata Rao, J. (2004). Optimisation of ozone treatment for colour and COD removal of acid dye effluent using central composite design experiment. *Dyes and Pigments*, 63:127-134
- Najafpour, G.D., Zinatizadeh, A.A.L., Mohamed, A.R., Hasnain Isa, M. and Nasrollahzadeh, H. (2006). High-rate anaerobic digestion of palm oil mill effluent in an upflow anaerobic sludge-fixed film bioreactor. *Process Biochemistry*, 41: 370-379
- Nakhla, G., Liu, V. and Bassi, A. (2006). Kinetic modeling of aerobic biodegradation of high oil and grease rendering wastewater. *Bioresource Technology*. 97:131–139
- Nasseri, S., Mazaheri Assadi, M., Noori Sepehr, M., Rostami, Kh., Shariat, M. and Nadafi, K. (2002). Chromium Removal from Tanning Effluent using Biomass of *Aspergillus oryzae. Pakistan Journal of Biological Sciences*, 5 (10): 1056-1059

- Nilsson, I., Moller, A., Mattiasson, B., Rubindamayugi, M.S.T. and Welander, U. (2006). Decolorization of synthetic and real textile wastewater by the use of white-rot fungi. *Enzyme and Microbial Technology*, 38: 94-100
- Nitayavardhana S. and Khanal, S.K. (2010). Innovative biorefinery concept for sugar-based ethanol industries: Production of protein-rich fungal biomass on vinnase as an aquaculture feed ingredient. *Bioresource Technology*, 101: 9078-9085
- Nour, A.H., Nour, A. H., Vissaliny, A.P.S. and Rajaletchumy A.P.V. (2010). Kinetics Study of Sewage Sludge Treatment by an Aerobic Digestion. *Journal of Applied Sciences*, 10 (3):226-230
- Okoro, C. C. (2008). Biodegradation of hydrocarbons in untreated produce water using pure fungal cultures. *African Journal of Microbiology Research*, 2: 217-223
- Okpowasili, G.C. and Nweke, C. O. (2005). Microbial growth and substrate utilization. *African Journal of Biotechnology*, 5 (4): 305-317
- Osiewacz, H. D. (2000). The genetic basis of aging in the fungal aging model *Podospora anserina. Futura.* 15:237-241
- Oswal, N., Sarma, P.M., Zinjarde, S.S. and Pant, A. (2002). Palm oil mill effluent treatment by a tropical marine yeast. *Bioresource Technology*, 85:35-37
- Palma, C., Moreira, M.T., Mielgo, I., Feijoo, G. and Lema, J.M. (1999). Use of a fungal bioreactor as a post treatment step for continuous decolorization of dyes. *Water Science and Technology*, 40: 131-136
- Park, D., Yun, Y-S. and Park, J.M. (2005). Use of dead fungal biomass for the detoxification of hexavalent chromium: screening and kinetics. *Process Biochemistry*, 40: 2559–2565
- Park, D., Yun, Y-S., Jo, J.H. and Park, J.M. (2005). Mechanism of hexavalent chromium removal by dead fungal biomass of *Aspergillus niger*. *Water Research*, 39: 533–540

- Pazouki, M., Najafpour, G., and Raouf Hosseini, M. (2008) Kinetic models of cell growth, substrate utilization and bio-decolorization of distillery wastewater by *Aspergillus fumigatus* UB260. *African Journal of Biotechnology*, 7 (9):1369-1376
- Pechsuth, M., Prasertsan, P. and Ukita, M. (2001). Biopretreatment of palm oil mill effluent by thermotolerant polymer producing fungi. *Songklanakarin Journal of Science Technology*. 23: 772-777
- Petre, M., Peng, M. X. and Mao L. X. (2010). The Influence of Culture Conditions on Fungal Pellet Formation by Submerged Fermentation of Cordyceps Synensis. In Proceeding of Fifth International Conference of Mushroom Biology and Mushroom Products, 99-104
- Pokhrel, D. and Viraghavan, T. (2008). Arsenic removal from an aqueous solution by modified *A. niger* biomass: Batch kinetic and isotherm studies. *Journal of Hazardous Materials*, 150: 818–825
- Polak-Berecka, M., Wasko, A., Kordowska-Wiater, M., Targonski, Z. and Kubik-Komar, A. (2011). Application of response surface methodology to enhancement of biomass production by *Lactobacillus rhamnosus e/n. Brazillian Journal of Microbiology*, 42:1485-1494
- Pujari, V and Chandra, T. S. (2000). Statistical optimization of medium components fo enhanced riboflavin production by UV-mutant of *Eromothecium ashbyii*. *Process Bichemistry*, 36: 31-37
- Radha, K.V., Arunagiri, R. A., Murugesan, T. (2005). Decolorization studies of synthetic dyes using Phanerochaete chrysosporium and their kinetics. Process Biochemistry, 40: 3337-3345
- R. Borja, R., Banks, C. J., Martin, A. and Khalfaoui B. (1995) Water waste: an overall kinetic model for methane production and substrate utilization. *Bioprocess Engineering*, 13: 87-95
- Rao, J.R. and Viraraghavan, T. (2002). Biosorption of phenol from an aqueous solution by biomass. *Bioresource Technology*, 85:165–171

- Rashid, S.S., Alam, M.Z., Karim, M.I.A. and Salleh, M. H. (2009). Management of palm oil mill effluent through production of cellulases by filamentous fungi. *World J Mocrobiol Biotechnol.* 25: 2219-2226
- Razak, C.N.A., Salleh, A.B., Musani, R., Samad, M.Y. and Basri, M. (1997). Some characteristics of lipases from thermophilic fungi isolated from palm oil mill effluent. *Journal of Molecular Catalysis B: Enzymatic*, 3: 153-159
- Rigas, F., Dritsa, V., Marchant, R., Papadopulou, K., Avramides, E.J. and Hatzianestis, I. (2005). Biodegradation of lindane by *Pleurotus ostreatus* via central composite design. *Environmental International*, 31:191-196
- Robles , A., Lucas, R., de Ciefuegos, G. A., Galve, A. (2000) Biomass production and detoxification pf wastewaters from the olive oil industry by strains pf Penicillium isolated from wastewater disposal ponds. *Bioresource Technology*.74:217
- Rodríguez-Couto, S. (2009). Dye removal by immobilized fungi. *Biotechnology Advances*. 27:227-235
- Rodríguez-Couto, S. (2012). A promising inert support for laccase production and decolouration of textile wastewater by the white-rot fungus *Trametes* pubescesns. Journal of Hazardous Materials, 233–234: 158–162
- Rostami, Kh. And Joodaki, M.R. (2002). Some studies of cadmium adsorption using Aspergillus niger, Penicillium austurianum, employing an airlift fermenter. Chemical Engineering Journal, 89: 239–252
- Rupani, P.F., Singh, P.R., Ibrahim, M.H. and Esa, N. (2010). Review of Current Palm Oil Mill Effluent (POME) Treatment Methods: Vermicomposting as a Sustainable Practice. *World Applied Sciences Journal*, 11 (1): 70-81
- Saeeda, A., Iqbala, M. and Zafar, S. I. (2009). Immobilization of *Trichoderma viride* for enhanced methylene blue biosorption: Batch and column studies. *Journal of Hazardous Materials*, 168: 406-415
- Sadhasivam, S., Savitha, S. and Swaminathan, K. (2010). Deployment of *Trichoderma harzianum* WL1 laccase in pulp bleaching and paper industry effluent treatment. *Journal of Cleaner Production*, 18: 799-806

- Sadhasivam, S. Savitha, S. and Swaminathan, K. (2007). Exploitation of *Trichoderma harzianum* mycelial waste for the removal of rhodamine 6G from aqueous solution. *Journal of Environmental Management*, 85: 155-161
- Salihu, A., Alam, M.Z., AbdulKarim, M.I. and Salleh, H.M. (2011). Optimization of lipase production by Candida cylindracea in palm oil mill effluent based medium using statistical experimental design. *Journal of Molecular Catalysis B: Enzymatic*, 69: 66-73
- Santos, V.L. and Linardi, V.R. (2004). Biodegradation of phenol by a filamentous fungi isolated from industrial effluents-identification and degradation potential. *Process Biochemistry*. 39:1001-1006
- Saranraj, P., Sumathi, V., Reetha, D. and Stella, D. (2010). Fungal Decolourization of Direct Azo Dyes and Biodegradation of Textile Dye Effluent. *Journal of Ecobiotechnology*, 2/7: 12-16
- Sati, S. C. and Bisht. S. (2006). Utilization of various caron sources for the growth of waterborne conodial fungi. *Mycologia*, 98 (5): 678–681.
- Schmidt S. K, Simkins S. and Alexander, M. (1985). Models for the kinetics of biodegradation of organic compounds not supporting growth. *Appl. Environ. Microbiol*, 50: 323-331
- Sepehr, M. N., Nasseri, S., Zarrabi, M., Samarghandi, M. R. and Amrane, A. (2012). Removal of Cr (III) from tanning effluent by *Aspergillus niger* in airlift bioreactor. *Separation and Purification Technology*, 96: 256–262
- Setiadi, T., Husaini and Djajadiningrat, A. (1996) Palm Oil Mill Effluent Treatment by Anaerobic Baffled Reactors: Recycle Effects and Biokinetic Parameters. *Water Science Technology*, 34 (11): 59-66
- Shah, M.P., Patel, K.A., Nair, S.S. and Darji, A.M. (2013). Microbial degradation of Textile Dye (Remazol Black B) by Bacillus spp. ETL-2012. *Bioremediation & Biodegradation*, 4:2
- Shahvali, M., Assadi, M.M., Rostami, K. (2000). Effect of environmental parametrs on decolorization of textile wastewater using Phanerochaete chrysosporium. *Bioprocess Engineering*, 23:721-726

- Shang, H., Yang, Q. and Zhang, Y. (2011). Kinetic Models of Dichloroethylene biodegradation by two strains of aerobic bacteria. *Biomed Environ Sci*, 24(3):261-267
- Shayegan, J., Pazouki, M. and Afshari, A. (2005). Continuous decolorization of anaerobically digested distillery wastewater. *Process Biochemistry*, 40: 1323–1329
- Shuler, M.L. and Kargi, F. *Bioprocess Engineering*. 2<sup>nd</sup>. ed. Upper Saddle River, NJ 07458: Prentice-Hall. 2002
- Simkins, S. and Alexander, M. (1984). Models for mineralization kinetics with the variables of substrate concentration and population density. *Appl. Environ. Microbiol*, 47: 1299-1306.
- Singh, R.P., Hakimi Ibrahim, M., Esa, N., Iliyana, M.S. (2010). Composting of waste from palm oil mill: A Sustainable Waste Management Practice. *Rev Environ Sci Biotechnol*, 9:331-344
- Singh, S.S. and Dikshit, A.K. (2010). Optimization of the parameters for decolourization by *Aspergillus niger* of anaerobically digested distillery spentwash pretreated with polyaluminium chloride. *Journal of Hazardous Materials*, 176: 864–869
- Sladka, A. (1996). The most common fungi in biological treatment plants. *Hydrobiologia*.31:350-362
- Souza, J.V.B, da Silva, E.S., da Silva, F.T. and Paiva, T.C.B. (2005). Fungal treatment of a delignification effluent from a nitrocellulose industry. *Bioresource Technology*, 96: 1936–1942
- Spina, F., Anastasi, A., Prigione, V., Tigini, V. and Varese, G.C. (2012). Biological treatment of industrial wastewaters: a fungal approach, *Chemical Engineering Transactions*, 27:175-180 DOI: 10.3303/CET1227030
- Standard Methods (2005). Standard Methods for the Examination of Water and Wastewater. 21<sup>st</sup> edition. APHA, AWWA, WPCF, Washington, D.C., U.S.A.

- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of azo dyes. *Applied Mirobiology and Biotechnology*, 56:69-80
- Subramanian, B.S., Yan, S., Tyagi, R.D., Surampalli, R.Y., (2008). A new, pellet-forming fungal strain: its isolation, molecular identification, and performance for simultaneous sludge-solids reduction, flocculation, and dewatering. *Water Environment Research*, 80 (9), 840–852.
- Sujatha, P., Kalarani, V. and Naresh Kumar, B. (2012). Effective Biosorption of Ni (II) from Aqueous Solution using *Trichoderma viride*. *Journal of Chemistry*, http://dx.doi.org/10.1155/2013/716098
- Sulaiman, N.M. Nachiappan and Vadaraj, B. (2003). Ultrafiltration of treated Palm oil mill effluent (POME), *Proceedings of the International Environmental Management and Technology A Clean Environment towards Sustainable Development*, August 4-6, Putrajaya, Malaysia.
- Sumathi Sethupathi. Removal of residue oil from palm oil mill effluent (POME) using chitosan. Ms.c. Thesis. Universiti Sains Malaysia; 2004
- Taştan, B. E., Ertuğrul, S. and Dönmez, G. (2010). Effective bioremoval of reactive dye and heavy metals by Aspergillus versicolor. Bioresource Technology, 101: 870–876
- Tejocote-Pérez, M., Balderas-Hernández, P., Barrera-Díaz, C., Roa-Morales, G. and Bárcena, V. (2012). Effect of *Chytriomyces hyalinus* on Industrial Wastewater Pre-Treated with Electrocoagulations in a Continuous System. *Natural Resources*, 3: 118-125
- Ting, S. Y. A. and Choong, C. (2009). Bioaccumulation and biosorption efficacy of *Trichoderma* isolate SP2F1 in removing copper (Cu (II)) from aqueous solutions. *World Journal of Mirobiology & Biotechnology*, 25(8): 1431-1437
- Torrades, F., Saiz, S., Garcia-Hortal, J. A. (2011). Using central composite experimental design optimize the degradation of black liquor by Fenton reagent. *Desalination*, 268:97-102

- Tsekova, K., Todorova, D., Dencheva, V. and Ganeva, S. (2010). Biosorption of copper(II) and cadmium(II) from aqueous solutions by free and immobilized biomass of *Aspergillus niger*. *Bioresource Technology*, 101: 1727–1731
- Tung, T. Q., Miyata, N. and Iwahori, K. (2004). Growth of Aspergillus oryzae during Treatment of Cassava Starch Processing Wastewater with High Content of Suspended Solids. Journal of Bioscience and Bioengineering, 97 (5):329-335
- Ugoji, E.O., (1997) Anaerobic Digestion of Palm Oil Mill Effluent and Its Utilization as Fertilizer for Environmental Protection. *Renewable Energy*, 10(2-3): 291-294
- Vassilev, N., Fenice, M., Federici, F. and Azcon R. (1997). Olive mill waste water treatment by immobilized cells of *Aspergillus niger* and its enrichment with soluble phosphate. *Process Biochemistry*, 32 (7): 617-620
- Vijayaraghavan, K. and Ahmad, D. (2006) Biohydrogen Generation from Palm Oil Mill Effluent Using Anaerobic Contact Filter. *International Journal of Hydrogen Energy*, 31(10): 1284 1291
- Vijayaraghavan, K., Ahmad, D., Abdul Aziz, M.E. (2007). Aerobic treatment of palm oil mill effluent. *J. Environ. Manage*. 82:24-31.
- Wang, Y.X. and Lu, Z.X. (2005). Optimization of processing parameters for the mycelial growth and extracellular polysaccharide production by *Boletus spp*. 50328. *Process Biochemistry*, 40: 1043-1051
- Wu, J., Xiao, Y-Z. and Yu, H-Q. (2005). Degradation of lignin in pulp mill wastewaters by white-rot fungi on biofilm. *Bioresource Technology*, 96: 1357–1363
- Wu, T.Y., Mohammad, A.W., Md. Jahim, J. and Anuar, N. (2006). Investigations on protease production by a wild-type Aspergillus terreus strain using diluted retentate of pre-filtered palm oil mill effluent (POME) as substrate. *Enzyme and Microbiology*, 39: 1223-1229
- Wu, T. Y., Mohammad, A. W., Md. Jahim, J. and Anuar, N. (2009). Holistic approach to managing palm oil mill effluent (POME): Biotechnological advances in the sustainable reuse of POME. *Biotechnology Advances*, 27: 40–52

- Wu, T. Y., Mohammad, A. W., Md. Jahim, J. and Anuar, N. (2010). Pollution controltechnologies for the treatment of palm oil mill effluent (POME) through end-of-pipe processes. *Journal of Environmental Management*, 91:1467-1490
- Xiong, X-J., Meng, X-J. and Zheng, T-L. (2010). Biosorption of C.I. Direct Blue 199 from aqueous solution by nonviable *Aspergillus niger*. *Journal of Hazardous Materials*, 175: 241–246
- Xu, T-J. and Ting, Y-P. (2004). Optimisation on bioleaching of incinerator fly ash by *Aspergillus niger*-use of central composite design. Enzyme and Microbial Technology, 35:444-454
- Yin, X., You, Q. and Jiang, Z. (2011). Optimization of enzyme assisted extraction of polysaccharides from *Tricholoma matsutake* by response surface methodology. *Carbohydrate Polymers*. 86: 1358-1364
- Yesilada, O., Sik, S. and Sam, M. (1997). Biodegradation of olive oil mill wastewaterby Coriolus versicolor and Funalia Trogii: Effects of agitation, initial Cod concentration, inoculum size and immobilization. *World Journal of Microbiology and Biotechnology*, 14: 37-42
- Zhang, Z.Y., Jin, B., Bai, Z.H. Wang, X. Y. (2008). Prodiuction of fungal biomass protein using microfungi from winery wastewater treatment. *Bioresourse technology*, 99: 3871-3876
- Zhou, X., Xu, S. X. Liu, L. and Chen, J. (2007). Degradation of cyanide by *Trichoderma* mutants constructed by restriction enzyme enzyme mediated integration (REMI). *Bioresource Technology*, 98: 2958-2962
- Zhen, G., Lu, X., Zhao, Y. and Chai, X. (2012). Enhanced dewaterability of sewage sludge in the presence of Fe (II)-activated persulfate oxidation. Bioresource technology, 116: 259-265
- Zinatizadeh, A.A.L., Mohamed, A.R., Najafpour, G.D., Hasnain isa, M. and Nasrollahzadeh, H. (2006). Kinetic evaluation of palm oil mill effluent digestion in a high rate up-flow anaerobic sludge fixed film bioreactor. *Process Biochemistry*, 41: 1038-1046

- Zinatizadeh, A. A. L., Salamatinia, B., Zinatizadeh, S. L., Mohamed, A. R. and Hasnain Isa, M. (2007). Palm Oil Mill Effluent Digestion in an Up-Flow Anaerobic Sludge Fixed Film Bioreactor. *International Journal of Environmental Research*, 1(3): 264-271
- Zinatizadeh, A.A.L., Mohamed, A. R., Mashitah, M.D. Abdullah, A.Z. and Hasnain Isa, M. (2007). Optimisation of pre-treated palm oil mill effluent digestion in an up-flow anaerobic sludge fixed film bioreactor: A comparative study. *Biochemical Engineering Journal*, 35: 226-237