

**SYNTHESIS OF THE ANTI-OXIDANT EUGENOL BENZOATE USING  
*Candida rugosa* LIPASE IMMOBILIZED ONTO MULTI WALLED  
CARBON NANOTUBES**

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This dissertation is dedicated to my parents, my husband and my lovely daughter for their endless support and encouragement.

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## ABSTRACT

Multi walled carbon nanotube (MWCNTs) having high surface area was used for the immobilization of lipase from *Candida rugosa* (CRL) by physical adsorption. The ability of the MWCNT immobilized CRL was compared against the free CRL in the esterification of benzoic acid with eugenol to produce the antioxidant eugenol benzoate, a potent lipoxygenase inhibitor. In this study, the effect of four parameters; reaction time, temperature, molecular sieves and enzyme loading were evaluated. The maximum yield of ester of reactions catalyzed by immobilized CRL and free lipase were found to be 31.62 and 27.2%, respectively. The study revealed that the optimal conditions for the highest yield of ester attained for both free CRL and immobilized CRL were similar. The attained optimum conditions were 5 mg/mL enzyme, 0.5:1.5 molar ratio of benzoic acid/eugenol dissolved in 38.5 mL chloroform, 500 mg of molecular sieves with 50 mL total volume of reaction mixture, stirred under 200 rpm at 40°C for 4 h. It was observed the MWCNT immobilized CRL afforded slightly higher yield of ester in all parameters investigated as compared to the free CRL. The outcome suggests that the MWCNT immobilized CRL could be used to synthesize eugenol benzoate. Furthermore, treatment of the esterification process is now easier due to simpler methods of separation of the end product and the biocatalyst could be recycled.

## ABSTRAK

Tiub nano berdinding lapis (MWCNT) mempunyai luas permukaan yang besar telah digunakan untuk immobilisasi lapis daripada *Candida rugosa* (CRL) secara penjerapan fizikal. Kebolehan CRL yang terimmobilisasi atas tiub nano berdinding lapis telah dibandingkan dengan CRL bebas di dalam pemangkinan pengesteran asid benzoik dengan eugenol untuk menghasilkan antioksidan eugenol benzoat, suatu perencat lipoksigenase. Di dalam kajian ini kesan empat parameter; masa tindakbalas, suhu, ayak molekul dan muatan enzim telah dikaji. Hasil maksimum ester untuk tindakbalas yang dimungkinkan oleh CRL terimmobilisasi dan CRL bebas didapati masing-masing adalah 31.62 dan 27.2%. Kajian mendapati keadaan optimum untuk pembentukan ester yang paling banyak yang dicapai oleh kedua-dua lapis bebas dan terimmobilisasi adalah serupa. Keadaan optimum yang tercapai ialah 5 mg/mL enzim, 0.5:1.5 nisbah molar asid benzoik/eugenol yang dilarutkan dalam 38.5 mL klorofom, 500 mg ayak molekul dengan jumlah isipadu campuran tindakbalas 50 mL, dikacau pada 200 rpm pada 40°C selama 4 jam. Didapati CRL yang terimmobilisasi ke atas MWCNT membentuk hasil ester yang sedikit tinggi di dalam kesemua parameter yang dikaji berbanding CRL bebas. Hasil kajian menunjukkan bahawa CRL yang terimmobilisasi ke atas MWCNT boleh digunakan untuk sintesis eugenol benzoat. Manakala, proses pengesteran kini lebih mudah hasil dari kaedah pemisahan produk akhir yang lebih mudah dan mungkin biologi yang boleh dikitar semula.

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**LIST OF ABBREVIATIONS**

AChE	-	Acetylcholinesterase
Ca-Alg	-	Calcium alginate
CALB	-	<i>Candida antarctica</i> lipase B
CRL	-	<i>Candida rugosa</i> lipase
CRL-MWCNT	-	<i>Candida rugosa</i> lipase immobilized onto multi-walled carbon nanotubes
CPW	-	Carboxypeptidase II
CNT	-	Carbon nanotubes
CCVD	-	Catalytic chemical vapour decomposition
CVD	-	Chemical vapour deposition
CMC	-	Critical micellular concentration
DLH	-	Dienlacton hydrolase
EDX	-	Energy Dispersive X-ray Spectroscopy
EC	-	Enzyme Commission
FESEM	-	Field Emission Scanning Electron Microscope
FT-IR	-	Fourier Transform Infrared
MWCNT-COOH	-	Functionalized multi-walled carbon nanotubes
GLP	-	<i>Geotrichum Candidum</i> lipase
GA	-	Glutaraldehyde
HAL	-	Haloalkane dehalogenase
HMDI	-	Hexamethylene diisocyanate
PI	-	Isoelectric point
MWNT	-	Multi-walled carbon nanotubes
NSAIDs	-	Nonsteroidal Anti-Inflammatory drugs
PEI	-	Polyethylenimine
PUFA	-	Polyunsaturated fatty acids
SEM	-	Scanning Electron Microscope

- SWNT - Single walled carbon nanotubes
- STDEV - Standard deviation
- TBME - Tert-Butyl methyl ether
- TEM - Transmission Electron Microscopy

**LIST OF SYMBOLS**

Å	-	Angstrom
Cm	-	Centimeter
°C	-	Degree Celcius
G	-	Gram
g/cm <sup>3</sup>	-	Gram per cubic centimeter
h	-	Hour
L	-	Liter
Mg	-	Microgram
Mg	-	Milligram
mg/mL	-	Milligram per milliliter
mL	-	Milliliter
Mm	-	Millimeter
mM	-	Millimolar
Nm	-	Nanometer
N	-	Normality
%	-	Percent
psig	-	Pounds per square inch gauge
rpm	-	Unit of agitation speed
U/mg	-	Unit per milligram
v/v	-	Volume per volume

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## CHAPTER 1

### INTRODUCTION

#### 1.1 Background of the study

Enzymes are protein compounds that have wide range of compositions along with structural complexities. The employment of enzyme as catalysts in biotransformation has acquired an escalating interest in the past several years due to their excessive catalytic activities alongside with special unique substrate specification in organic synthesis. The two key properties of enzymes are usually satisfactory to minimize energy and process consumption cost (Palmer, 1985; Perez *et al.*, 2007). Hydrolase are enzymes that acquire great attention because of their high possible commercial application. Lipases are one of several hydrolase enzymes which may have increased resistance to deactivation in non-aqueous mass media. Due to excessive stableness, they can be utilized regularly for many reactions with no significant loss of their activity (Stamatis *et al.*, 1993).

Reactions that are catalyzed by enzymes are usually considered more productive and efficient when compared to inorganic catalyzed reactions, due to ultimate decrease in the reactions activation energy which is why enzyme-catalyzed reactions are carried out in typically mild conditions such as; lower temperature, low atmospheric pressure and physiological pH values in comparison to the inorganic catalyst. The lipase-catalyzed reactions are mostly in different medium which include biphasic organic-aqueous, reversed micelles, organic and supercritical CO<sub>2</sub> system

(Martinek *et al.*, 1981; Steytler *et al.*, 1991; Yang *et al.*, 1997). Because many hydrophobic substrates are insoluble in aqueous media, a new method of utilizing enzyme in organic media have been proposed which leads to increase in use of lipase enzyme mainly because numerous hydrophobic substrates usually are soluble within this media (Okahata and Mori, 1997).

Esterification reactions are generally omnipresent inside fine compounds, more advanced along with petrochemical industries. Besides hydrolysis involving triglyceride, lipases can also be employed for esterification between fatty acid and alcohol. Numerous researches have been carried out in the facet of controlling normal water activity as a way to promote the reaction towards synthesis direction. The product isolation and excess substrate concentrations have been evaluated and removed by different method and the products are obtained in high yield (Bloomer *et al.*, 1992; Yadav *et al.*, 2012).

The use of nanobiocatalyst is a rapid growing research area which refers to the use of enzymes that have been immobilized on nanomaterials. Among the used nanobiocatalyst, the carbon-based nanomaterial such as; carbon nanotubes (CNTs) and graphene have greatly attracted scientific interest (Kim *et al.*, 2008). The use of CNTs was first initiated by Endo in the late 19<sup>th</sup> century with a carbon tubular structure (also called carbon filaments in the sixties) that has a diameter of less than 100 nm (Cisseli, 2007). The multi-walled carbon nanotube which consist of two or more concentric cylindrical shells of graphene sheets that are arranged around a central hollow was the first CNT to be observed *via* Transmission Electron Microscopy (TEM) analysis and is currently the most attractive nanomaterial (Ajayan *et al.*, 2006).

*Candida rugosa* lipase (CRL) is one of the widely used industrial lipase that are mostly utilized in various reaction which include hydrolysis and esterification reactions due to its wide substrate specificity and also in pharmaceutical synthesis due to its stereoselectivity and regioselectivity (Takac and Bakkal, 2007). The immobilization of enzyme is a common strategy to obtain a conventional

heterogeneous catalyst onto biological catalyst with desirable features such as; enzyme immobilization on CNTs which other specific groups (e.g. organic, polymeric and biological molecules) to be functionalized to the CNTs which further makes it possible for the enzymes to specifically and selectively bind onto the CNTs. The key step in bionanofabrication is the immobilization of CRL on MWCNTs due to its high surface area, mechanical, thermal and electrical properties and biocompatibility (Kuchibhatla *et al.*, 2007).

In this study, the immobilization of a cheap and widely available enzyme (CRL) onto the MWCNTs would be carried out *via* adsorption and would further be used as catalyst in the esterification reaction for the synthesis of eugenol benzoate using benzoic acid as acyl donor, eugenol as substrate and chloroform as solvent. The frequent use of substituted esters (both aliphatic and aromatic) as plasticizers, solvents, flavors and precursors to a range of pharmaceuticals have been reported elsewhere (Yadav *et al.*, 2012).

## **1.2 Problem statement of the study**

Eugenol esters are chemically synthesized as potential inhibitors of lipoxygenase. The benefits of these compounds can be more enhance with the improvement of an efficient process for their synthesis under mild conditions. The use of lipases in non-aqueous medium has been described by Stamatis *et al.* (2001) for the preparation of phenolics acid ester. Besides, the esterification yield obtained using free lipases was very low. Other disadvantages include; less stable and low activity in organic solvent cannot withstand high temperature and pressure and requires the use of large amount of starting materials, these results in high waste generation in the industry. Hence, immobilization of the enzyme would drastically reduce the high use of starting materials which are quite expensive and environmental unfriendly.

The challenge of using biological element as catalyst is quite increasing due to their expensiveness, complexity as well as low stability. As such, a simple alternative method is required to reduce the above shortcomings without altering scientific interests. Therefore, enzyme immobilization onto a solid support (MWCNTs) is an alternative route because of its successful recycling property and providing mechanical stability to the enzyme three-dimensional structure. The use of enzyme immobilization is greatly important for the synthesis of eugenol benzoate. As such due to its ease of handling, faster purification, thermal and pH stability, high selectivity and substrate specificity, low energy and high yield of product.

### **1.3 Objectives of the study**

The objectives of the study are:

1. To immobilize CRL onto MWCNTs using physical adsorption method.
2. To determine best reaction conditions for the synthesis of eugenol benzoate.
3. To compare the free CRL and CRL-MWCNT to catalyze the synthesis of eugenol benzoate.

### **1.4 Scope of the study**

This study would report a greener alternative to the synthesis of valuable commercial products (eugenol benzoate) by the use of CRL immobilized onto MWCNT. To study how the molecules interacts with the enzyme and how often they affect the enzyme structure and how the enzyme is arranged on the MWCNTs. The reaction conditions to be optimized include; incubation time, temperature, presence of desiccant (molecular sieves) and effect of enzyme loading.

## 1.5 Significance of the study

CNTs are new nano-size carbon material and they have many potential applications to be exploited which include its use in the polymer composites and bioscience application that generated a remarkable starting point in the field of material science studies. Enzyme immobilization on CNTs presents some advantages over the bulk solid materials such as high surface area which can lead to higher enzyme loading, nanoscale dispersion and ease of surface functionalization (Kim *et al.*, 2008).

In general, the use of enzymes economically and efficiently in aqueous as well as in non-aqueous solvents allows the modification of their activity, selectivity and operational stability. Several applications reveals the use of lipase in an immobilized state due to its ease of separation, stability, reusability, better control of reaction and more favorable economical factors (Nasratun *et al.*, 2009). The study will determine whether the CRL immobilized onto MWCNTs by physical adsorption, is a viable option for the synthesis of eugenol benzoate by esterification reactions.

## REFERENCES

- Abdul Rahman, M. B., Jumbri, K., Mohd Ali Hanafiah, N. A., Abdulmalek, E., Tejo, B. A., Basri, M. and Salleh, A. B. (2012). Enzymatic esterification of fatty acid esters by tetraethylammonium amino acid ionic liquids-coated *Candida rugosa* lipase. *Journal of Molecular Catalysis B: Enzymatic*, 79, 61-65.
- Ajayan, P. M., Schadler, L. S. and Braun, P. V. (2006). *Nanocomposite science and technology*. (1<sup>st</sup> ed.). Germany: Wiley-VCH Verlag GmbH and Co. KGaA.
- Akoh, C.C. and Min, D.B (1998). Microbial lipases and enzymatic interesterification, In: *Food lipids: Chemistry. Nutrition and Biotechnology*. MarcelDeccer, New York, pp 641-698.
- Akoh, C. C., Lee, G. and Shaw, J. (2004). Protein engineering and applications of *Candida rugosa* lipase isoforms. *Lipids*. **39**(6), 513-526.
- Alberghina, L., Schmid, R. and Verger, R. (Eds.) (1991). *Lipases: Structure, Mechanism, and Genetic Engineering: Contributions to the CEC-GBF International Workshop, September 13 to 15, 1990, Braunschweig, Germany* (Vol. 16). VCH.
- Anazawa, K., Shimotani, K., Manabe, C., Watanabe, H. and Shimizu, M. (2002). High purity carbon nanotubes synthesis method by an arc discharging in magnetic field. *Applied Physics Letters*. **81**(4), 739-741.
- Ansari, S. A. and Husain, Q. (2012). Potential applications of enzymes immobilized on/in nano materials: a review. *Biotechnology Advances*. **30**(3), 512-523.
- Atsumi, T., Fujisawa, S., Satoh, K., Sakagami, H., Iwakura, I., Ueha, T. and Yokoe, I. (2000). Cytotoxicity and radical intensity of eugenol, isoeugenol or related dimmers. *Anticancer Research*. **20**(4), 2519-2524.
- Awasthi, P. K., Dixit, S. C., Dixit, N. and Sinha, A. K. (2008). Eugenol derivatives as future potential drugs. *Journal Pharmaceutical Research*. **1**(2), 215-220.

- Bakkali, F., Averbeck, S., Averbeck, D. and Idaomar, M. (2008). Biological effects of essential oils-a review. *Food and Chemical Toxicology*. **46**(2), 446-475.
- Balcao V.M., Pavia A.L. and Malcata F.X. (1996). Bioreactors with immobilized lipases: state of the art. *Enzyme and Microbial Technology*, 18, 392-416.
- Ballesteros, A., Bornscheuer, U., Capewell, A., Combes, D., Condoret, J. S., Koenig, K. and Xenakis, A. (1995). Enzymes in non-conventional phases. *Biocatalysis and Biotransformation*. **13**(1), 1-42.
- Barabino, R. C., Gray, D. N. and Keyes, M. H. (1978). Coupled reactions of immobilized enzymes and immobilized substrates: Clinical application as exemplified by amylase assay. *Clinical Chemistry*. **24**(8), 1393-1398.
- Barceloux, D. G. (2008). *Medical toxicology of natural substances: foods, fungi, medicinal herbs, plants, and venomous animals*. John Wiley and Sons.
- Barros, M., Fleuri, L. F. and Macedo, G. A. (2010). Seed lipases: sources, applications and properties-a review. *Brazilian Journal of Chemical Engineering*. **27**(1), 15-29.
- Basri, M., Ampon, K., Yunus, W. M., Razak, C. N. A. Salleh, A. B. (1994). Immobilization of hydrophobic lipase derivatives on to organic polymer beads. *Journal of Chemical Technology and Biotechnology*. **59**(1), 37-44.
- Bebarta, B., Jhansi, M. J., Kotasthane, P. and Sunkireddy, Y. R. (2013). Medium chain and behenic acid incorporated structured lipids from sal, mango and kokum fats by lipase acidolysis. *Food Chemistry*. **136**(2), 889-894.
- Benjamin, S. and Pandey, A. (1998). *Candida rugosa* lipases: Molecular biology and versatility in biotechnology. *Yeast*. **14**(12), 1069-1087.
- Bertrand, F., Basketter, D. A., Roberts, D. W. and Lepoittevin, J. P. (1997). Skin sensitization to eugenol and isoeugenol in mice: possible metabolic pathways involving ortho-quinone and quinone methide intermediates. *Chemical Research in Toxicology*. **10**(3), 335-343.
- Bhardwaj, K., Raju, A. and Rajasekharan, R. (2001). Identification, purification, and characterization of a thermally stable lipase from rice bran. A new member of the (phospho) lipase family. *Plant Physiology*. **127**(4), 1728-1738.
- Blanco, R. M., Calvete, J. J. and Guisan, J. M. (1989). Immobilization-stabilization of enzymes; variables that control the intensity of the trypsin (amine)-agarose (aldehyde) multipoint attachment. *Enzyme and Microbial Technology*. **11**(6), 353-359.

- Bloomer, S., Adlercreutz, P. and Mattiasson, B. (1992). Facile synthesis of fatty acid esters in high yields. *Enzyme and Microbial Technology*. **14**(7), 546-552.
- Brocca, S., Persson, M., Wehtje, E., Adlercreutz, P., Alberghina, L. and Lotti, M. (2000). Mutants provide evidence of the importance of glycosidic chains in the activation of lipase 1 from *Candida rugosa*. *Protein Science*. **9**(5), 985-990.
- Burghard, M., Duesberg, G., Philipp, G., Muster, J. and Roth, S. (1998). Controlled adsorption of carbon nanotubes on chemically modified electrode arrays. *Advanced Materials*. **10**(8), 584-588.
- Cañete-Rosales, P., Ortega, V., Álvarez-Lueje, A., Bollo, S., González, M., Ansón, A. and Martínez, M. T. (2012). Influence of size and oxidative treatments of multi-walled carbon nanotubes on their electrocatalytic properties. *Electrochimica Acta*, **62**, 163-171.
- Cang-Rong, J. T. and Pastorin, G. (2009). The influence of carbon nanotubes on enzyme activity and structure: Investigation of different immobilization procedures through enzyme kinetics and circular dichroism studies. *Nanotechnology*. **20**(25).
- Cao, L. (2005). Immobilised enzymes: Science or art? *Current Opinion in Chemical Biology*, **9**(2), 217-226.
- Carriere, F., Thirstrup, K., Hjorth, S. and Boel, E. (1994). Cloning of the classical guinea pig pancreatic lipase and comparison with the lipase related protein 2. *FEBS Letters*. **338**(1), 63-68.
- Carta, G., Gainer, J. L. and Gibson, M. E. (1992). Synthesis of esters using a nylon immobilized lipase in batch and continuous reactors. *Enzyme and Microbial Technology*. **14**(11), 904-910.
- Chaibakhsh, N., Basri, M., Mohamed Anuar, S. H., Abdul Rahman, M. B. and Rezayee, M. (2012). Optimization of enzymatic synthesis of eugenol ester using statistical approaches. *Biocatalysis and Agricultural Biotechnology*. **1**(3), 226-231.
- Che, G., Lakshmi, B. B., Fisher, E. R. and Martin, C. R. (1998). Carbon nanotubule membranes for electrochemical energy storage and production. *Nature*. **393**(6683), 346-349.



- Chen, J., Hamon, M. A., Hu, H., Chen, Y., Rao, A. M., Eklund, P. C. and Haddon, R. C. (1998). Solution properties of single-walled carbon nanotubes. *Science*. **282**(5386), 95-98.
- Chen, R. J., Bangsaruntip, S., Drouvalakis, K. A., Wong Shi Kam, N., Shim, M., Li, Y. and Dai, H. (2003). Noncovalent functionalization of carbon nanotubes for highly specific electronic biosensors. *Proceedings of the National Academy of Sciences of the United States of America*. **100**(9), 4984-4989.
- Chen, Y., Zhu, C. and Wang, T. (2006). The enhanced ethanol sensing properties of multi-walled carbon nanotubes/SnO<sub>2</sub> core/shell nanostructures. *Nanotechnology*. **17**(12), 3012-3017.
- Chiang, C., Hsiau, L. and Lee, W. (2004). Immobilization of cell-associated enzymes by entrapment in polymethacrylamide beads. *Biotechnology Techniques*. **11**(2), 121-125.
- Chiang, Y. C., Lin, W. H. and Chang, Y. C. (2011). The influence of treatment duration on multi-walled carbon nanotubes functionalized by H<sub>2</sub>SO<sub>4</sub>/HNO<sub>3</sub> oxidation. *Applied Surface Science*. **257**(6), 2401-2410.
- Chiou, S. H. and Wu, W. T. (2004). Immobilization of *Candida rugosa* lipase on chitosan with activation of the hydroxyl groups. *Biomaterials*. **25**(2), 197-204.
- Ciselli, P. (2007). *The potential of carbon nanotubes in polymer composites*. PhD thesis, Eindhoven University of Technology.
- Collins, P. G., Bradley, K., Ishigami, M. and Zettl, A. (2000). Extreme oxygen sensitivity of electronic properties of carbon nanotubes. *Science*. **287**(5459), 1801-1804.
- Corrias, M., Caussat, B., Ayrat, A., Durand, J., Kihn, Y., Kalck, P. and Serp, P. (2003). Carbon nanotubes produced by fluidized bed catalytic CVD: First approach of the process. *Chemical Engineering Science*. **58**(19), 4475-4482.
- Cygler, M. and Schrag, J. D. (1999). Structure and conformational flexibility of *Candida rugosa* lipase. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*. **1441**(2), 205-214.
- Cygler, M., Schrag, J. D., Sussman, J. L., Harel, M., Silman, I., Gentry, M. K. and Doctor, B. P. (1993). Relationship between sequence conservation and three dimensional structure in a large family of esterase, lipases, and related proteins. *Protein Science*. **2**(3), 366-382.

- Daenen, M., de Fouw, R.D., Hamers, B., Janssen, P.G.A., Schouteden, K. and Veld, M.A.J. (2003). *The Wondrous World of Carbon Nanotubes*. Unpublished, Denmark/Eindhoven University of Technology.
- Dandavate, V. and Madamwar, D. (2007). Novel approach for the synthesis of ethyl isovalerate using surfactant coated *Candida rugosa* lipase immobilized in microemulsion based organogels. *Enzyme and Microbial Technology*. **41**(3), 265-270.
- Dandavate, V., Keharia, H. and Madamwar, D. (2009). Ethyl isovalerate synthesis using *Candida rugosa* lipase immobilized on silica nanoparticles prepared in nonionic reverse micelles. *Process Biochemistry*. **44**(3), 349-352.
- Dave, R. and Madamwar, D. (2008). *Candida rugosa* lipase immobilized in Triton-X100 microemulsion based organogels (MBGs) for ester synthesis. *Process Biochemistry*. **43**(1), 70-75.
- Dawar, P., Bhagavan Raju, M. and Ramakrishna, R. A. (2011). One-pot esterification and Ritter reaction: chemo- and regioselectivity from tert-butyl methyl ether. *Tetrahedron Letters*. **52**(33), 4262-4265.
- De La Casa, R. M., Sánchez-Montero, J. M. and Sinisterra, J. V. (1996). Water adsorption isotherm as a tool to predict the pre equilibrium water amount in preparative esterification. *Biotechnology Letters*. **18**(1), 13-18.
- Domínguez De María, P., Sánchez-Montero, J. M., Sinisterra, J. V. and Alcántara, A. R. (2006). Understanding *Candida rugosa* lipases: An overview. *Biotechnology Advances*. **24**(2), 180-196.
- Dussán, K. J., Cardona, C. A. and Giraldo, O. (2012). Immobilization and characterization of the *Candida rugosa* lipase enzyme on magnetic particles. *Revista Mexicana de Física S*. **58**(2), 47-51.
- Dyal, A., Loos, K., Noto, M., Chang, S. W., Spagnoli, C., Shafi, K. V. and Gross, R. A. (2003). Activity of *Candida rugosa* lipase immobilized on  $\gamma$ -Fe<sub>2</sub>O<sub>3</sub> magnetic nanoparticles. *Journal of the American Chemical Society*. **125**(7), 1684-1685.
- Ebbesen, T. W. and Ajayan, P. M. (1992). Large-scale synthesis of carbon nanotubes. *Nature*. **358**(6383), 220-222.

- Endo, M., Hayashi, T., Kim, Y. A., Terrones, M. and Dresselhaus, M. S. (2004). Applications of carbon nanotubes in the twenty-first century. *Philosophical Transactions of the Royal Society of London. Series A: Mathematical, Physical and Engineering Sciences*. **362**(1823), 2223-2238.
- Fadiloglu, S. and Söylemez, Z. (1998). Olive oil hydrolysis by celite-immobilized *Candida rugosa* lipase. *Journal of Agricultural and Food Chemistry*. **46**(9), 3411-3414.
- Feng, W. and Ji, P. (2011). Enzymes immobilized on carbon nanotubes. *Biotechnology Advances*. **29**(6), 889-895
- Ferrer, P., Montesinos, J. L., Valero, F. and Solà, C. (2001). Production of native and recombinant lipases by *Candida rugosa*. *Applied Biochemistry and Biotechnology-Part A Enzyme Engineering and Biotechnology*. **95**(3), 221-255.
- Fessenden, R. J. and Fessenden, J. S. (1990). *Fundamentals of organic chemistry*. (4<sup>th</sup> ed.) Harper and Row. California: Cole Publishing Co.
- Flahaut, E., Bacsa, R., Peigney, A. and Laurent, C. (2003). Gram-scale CCVD synthesis of double-walled carbon nanotubes. *Chemical Communications*. **9**(12), 1442-1443.
- Flood, M. T. and Kondo, M. (2001). Safety evaluation of lipase produced from *Candida-rugosa*: Summary of toxicological data. *Regulatory Toxicology and Pharmacology*. **33**(2), 157-164.
- Foresti, M. L., Pedernera, M., Bucalá, V. and Ferreira, M. L. (2007). Multiple effects of water on solvent-free enzymatic esterifications. *Enzyme and Microbial Technology*. **41**(1-2), 62-70.
- Freitas, L., Paula, A. V., dos Santos, J. C., Zanin, G. M. and de Castro, H. F. (2010). Enzymatic synthesis of monoglycerides by esterification reaction using penicillium camembertii lipase immobilized on epoxy SiO<sub>2</sub>-PVA composite. *Journal of Molecular Catalysis B: Enzymatic*. **65**(1-4), 87-90.
- Gao, S., Wang, Y., Diao, X., Luo, G. and Dai, Y. (2010). Effect of pore diameter and cross-linking method on the immobilization efficiency of *Candida rugosa* lipase in SBA 15. *Bioresource Technology*. **101**(11), 3830-3837.

- Gao, Y. and Kyratzis, I. (2008). Covalent immobilization of proteins on carbon nanotubes using the cross-linker 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide A critical assessment. *Bioconjugate Chemistry*. **19**(10), 1945-1950.
- Gill, I. (2001). Bio-doped nanocomposite polymers: Sol-gel bioencapsulates. *Chemistry of Materials*. **13**(10), 3404-3421.
- Goldoni, A., Larciprete, R., Petaccia, L. and Lizzit, S. (2003). Single-wall carbon nanotube interaction with gases: Sample contaminants and environmental monitoring. *Journal of the American Chemical Society*. **125**(37), 11329-11333.
- Gómez, J. M., Romero, M. D. and Fernández, T. M. (2005). Immobilization of  $\beta$ -glucosidase on carbon nanotubes. *Catalysis Letters*. **101**(3-4), 275-278.
- Goyanes, S., Rubiolo, G. R., Salazar, A., Jimeno, A., Corcuera, M. A. and Mondragon, I. (2007). Carboxylation treatment of multi walled carbon nanotubes monitored by infrared and ultraviolet spectroscopies and scanning probe microscopy. *Diamond and Related Materials*. **16**(2), 412-417.
- Graber, M., Bousquet-Dubouch, M. P., Sousa, N., Lamare, S. and Legoy, M. D. (2003). Water plays a different role on activation thermodynamic parameters of alcoholysis reaction catalyzed by lipase in gaseous and organic media. *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics*. **1645**(1), 56-62.
- Grochulski, P., Li, Y., Schrag, J. D. and Cygler, M. (1994). Two conformational states of *Candida rugosa* lipase. *Protein Science*. **3**(1), 82-91.
- Grochulski, P., Li, Y., Schrag, J. D., Bouthillier, F., Smith, P., Harrison, D. and Cygler, M. (1993). Insights into interfacial activation from an open structure of *Candida-rugosa* lipase. *Journal of Biological Chemistry*. **268**(17), 12843-12847.
- Guisan, J.M., 2006. *Immobilization of Enzymes and Cells*. (2<sup>nd</sup> ed.). Totowa: Humana Press Inc.
- Guiseley, K. B. (1989). Chemical and physical properties of algal polysaccharides used for cell immobilization. *Enzyme and Microbial Technology*. **11**(11), 706-716.

- Guncheva, M., Tashev, E., Zhiryakova, D., Tosheva, T. and Tzokova, N. (2011). Immobilization of lipase from *Candida rugosa* on novel phosphorous-containing polyurethanes: Application in wax ester synthesis. *Process Biochemistry*. **46**(4), 923-930.
- Guo, T., Nikolaev, P., Thess, A., Colbert, D. T. and Smalley, R. E. (1995). Catalytic growth of single-walled nanotubes by laser vaporization. *Chemical Physics Letters*. **243**(1-2), 49-54.
- Hasan, F., Shah, A. A. and Hameed, A. (2006). Industrial applications of microbial lipases. *Enzyme and Microbial Technology*. **39**(2), 235-251.
- Hilding, J., Grulke, E. A., George Zhang, Z. and Lockwood, F. (2003). Dispersion of carbon nanotubes in liquids. *Journal of Dispersion Science and Technology*. **24**(1), 1-41.
- Hinds, B. J., Chopra, N., Rantell, T., Andrews, R., Gavalas, V. and Bachas, L. G. (2004). Aligned multiwalled carbon nanotube membranes. *Science*. **303**(5654), 62-65.
- Honda, T., Miyazaki, M., Nakamura, H. and Maeda, H. (2006). Facile preparation of an enzyme-immobilized microreactor using a cross-linking enzyme membrane on a micro channel surface. *Advanced Synthesis and Catalysis*. **348**(15), 2163-2171.
- Horchani, H., Ben Salem, N., Zarai, Z., Sayari, A., Gargouri, Y. and Chaâbouni, M. (2010). Enzymatic synthesis of eugenol benzoate by immobilized *Staphylococcus aureus* lipase: Optimization using response surface methodology and determination of antioxidant activity. *Bioresource Technology*. **101**(8), 2809-2817.
- Hou, H., Ge, J. J., Zeng, J., Li, Q., Reneker, D. H., Greiner, A. and Cheng, S. Z. D. (2005). Electrospun polyacrylonitrile nanofibers containing a high concentration of well-aligned multi walled carbon nanotubes. *Chemistry of Materials*. **17**(5), 967-973.
- Huang, W., Taylor, S., Fu, K., Lin, Y., Zhang, D., Hanks, T. W. and Sun, Y. (2002). Attaching proteins to carbon nanotubes via diimide-activated amidation. *Nano Letters*. **2**(4), 311-314.

- Huang, W., Xia, Y. M., Gao, H., Fang, Y. J., Wang, Y. and Fang, Y. (2005). Enzymatic esterification between n-alcohol homologs and n-caprylic acid in non-aqueous medium under microwave irradiation. *Journal of Molecular Catalysis B: Enzymatic*. **35**(4), 113-116.
- Ibrahim, N. A., Guo, Z. and Xu, X. (2008). Enzymatic interesterification of palm stearin and coconut oil by a dual lipase system. *JAOCS, Journal of the American Oil Chemists' Society*. **85**(1), 37-45.
- Iijima, S. and Ichihashi, T. (1993). Single-shell carbon nanotubes of 1nm diameter. *Nature*. **363**(6430), 603-605.
- Iijima, S. (1991). Helical microtubules of graphitic carbon. *Nature*. **354**(6348), 56-58.
- Jia, H., Zhu, G. and Wang, P. (2003). Catalytic behaviors of enzymes attached to nanoparticles: The effect of particle mobility. *Biotechnology and Bioengineering*. **84**(4), 406-414.
- Jin, W. and Brennan, J. D. (2002). Properties and applications of proteins encapsulated within sol-gel derived materials. *Analytica Chimica Acta*. **461**(1), 1-36.
- Joshi, K. A., Prouza, M., Kum, M., Wang, J., Tang, J., Haddon, R. and Mulchandani, A. (2006). V-type nerve agent detection using a carbon nanotube-based amperometric enzyme electrode. *Analytical Chemistry*. **78**(1), 331-336.
- Joshi, K. A., Tang, J., Haddon, R., Wang, J., Chen, W. and Mulchandani, A. (2005). A disposable biosensor for organophosphorus nerve agents based on carbon nanotubes modified thick film strip electrode. *Electroanalysis*. **17**(1), 54-58.
- Journet, C., Maser, W. K., Bernier, P., Loiseau, A., De La Chapelle, M. L., Lefrant, D. L. S. and Fischer, J. E. (1997). Large-scale production of single-walled carbon nanotubes by the electric-arc technique. *Nature*. **388**(6644), 756-758.
- Kahveci, D. and Xu, X. (2011). Repeated hydrolysis process is effective for enrichment of omega 3 polyunsaturated fatty acids in salmon oil by *Candida rugosa* lipase. *Food Chemistry*. **129**(4), 1552-1558.
- Karajanagi, S. S., Vertegel, A. A., Kane, R. S. and Dordick, J. S. (2004). Structure and function of enzymes adsorbed onto single-walled carbon nanotubes. *Langmuir*. **20**(26), 11594- 11599.

- Katchalski-Katzir, E. and Kraemer, D. M. (2000). Eupergit<sup>®</sup>C, a carrier for immobilization of enzymes of industrial potential. *Journal of Molecular Catalysis B: Enzymatic*. **10**(1), 157-176
- Katchalski-Katzir, E. (1993). Immobilized enzymes-learning from past successes and failures. *Trends in Biotechnology*. **11**(11), 471-478.
- Keeling-Tucker, T. and Brennan, J. D. (2001). Fluorescent probes as reporters on the local structure and dynamics in sol-gel-derived nanocomposite materials. *Chemistry of Materials*. **13**(10), 3331-3350.
- Kim, B. J., Kang, B. K., Bahk, Y. Y., Yoo, K. H. and Lim, K. J. (2009). Immobilization of horseradish peroxidase on multi-walled carbon nanotubes and its enzymatic stability. *Current Applied Physics*. **9**(4), 263-265.
- Kim, J., Grate, J. W. and Wang, P. (2008). Nanobiocatalysis and its potential applications. *Trends in Biotechnology*. **26**(11), 639-646.
- Kim, J., Grate, J. W. and Wang, P. (2006). Nanostructures for enzyme stabilization. *Chemical Engineering Science*. **61**(3), 1017-1026.
- Kim, S. M. and Rhee, J. S. (1991). Production of medium-chain glycerides by immobilized lipase in a solvent-free system. *Journal of the American Oil Chemists' Society*. **68**(7), 499-503.
- Klotzbach, T. L., Watt, M., Ansari, Y. and Minter, S. D. (2008). Improving the microenvironment for enzyme immobilization at electrodes by hydrophobically modifying chitosan and nafion<sup>®</sup> polymers. *Journal of Membrane Science*. **311**(1-2), 81- 88.
- Knezevic, Z., Milosavic, N., Bezbradica, D., Jakovljevic, Z. and Prodanovic, R. (2006). Immobilization of lipase from *Candida rugosa* on eupergit<sup>®</sup>C supports by covalent attachment. *Biochemical Engineering Journal*. **30**(3), 269-278.
- Komatsu, T., Nagayama, K. and Imai, M. (2004). Interesterification activity of *Rhizopus delemar* lipase in phospholipid microemulsions. *Colloids and Surfaces B: Biointerfaces*. **38**(3), 175-178).
- Kónya, Z. and Kiricsi, I. (2004). *Catalytica production, purification, characterization and application of single and multi-walled carbon nanotubes*. Unpublished.

- Krishnan, A., Dujardin, E., Ebbesen, T. W., Yianilos, P. N. and Treacy, M. M. J. (1998). Young's modulus of single-walled nanotubes. *Physical Review B*. **58**(20), 14013.
- Kuchibhatla, S. V., Karakoti, A. S., Bera, D. and Seal, S. (2007). One dimensional nanostructured materials. *Progress in Materials Science*. **52**(5), 699-913.
- Kumakura, M. and Kaetsu, I. (2003). Immobilization of cellulase using porous polymer matrix. *Journal of Applied Polymer Science*. **29**(9), 2713-2718.
- Kumar, R., Mago, G., Balan, V. and Wyman, C. E. (2009). Physical and chemical characterizations of corn stover and poplar solids resulting from leading pretreatment technologies. *Bioresource Technology*. **100**(17), 3948-3962.
- Larroche, C. and Pandey, A. (2005). First International Congress on Bioprocesses in Food Industries (ICBF-2004). *LWT-Food Science and Technology*. **38**(6), 695.
- Laurent, N., Haddoub, R. and Flitsch, S. L. (2008). Enzyme catalysis on solid surfaces. *Trends in Biotechnology*. **26**(6), 328-337.
- Lee, G., Kim, J. and Lee, J. H. (2008). Development of magnetically separable polyaniline nanofibers for enzyme immobilization and recovery. *Enzyme and Microbial Technology*. **42**(6), 466-472.
- Legoy, M., Goldberg, M. and Barbotin, J. (1991). Lipase and water. In *Lipases: Structure, Mechanism, and Genetic Engineering: Contributions to the CEC-GBF International Workshop, September 13-15, 1990, Braunschweig, Germany* (Vol. 16, p. 183). VCH.
- Li, H. and Zhang, X. (2005). Characterization of thermostable lipase from thermophilic geobacillus sp. TW1. *Protein Expression and Purification*. **42**(1), 153-159.
- Li, W. Z., Xie, S. S., Qian, L. X., Chang, B. H., Zou, B. S., Zhou, W. Y. and Wang, G. (1996). Large-scale synthesis of aligned carbon nanotubes. *Science*. **274**(5293), 1701-1703.
- Li, Y., Zhang, X. B., Tao, X. Y., Xu, J. M., Huang, W. Z., Luo, J. H. and Geise, H. J. (2005). Mass production of high-quality multi-walled carbon nanotube bundles on a Ni/MO/MgO catalyst. *Carbon*. **43**(2), 295-301.



- Lima, F. V., Pyle, D. L. and Asenjo, J. A. (1995). Factors affecting the esterification of lauric acid using an immobilized biocatalyst: Enzyme characterization and studies in a well-mixed reactor. *Biotechnology and Bioengineering*. **46**(1), 69-79.
- Liu, Y. and Gao, L. (2005). A study of the electrical properties of carbon nanotube NiFe<sub>2</sub>O<sub>4</sub> composites: Effect of the surface treatment of the carbon nanotubes. *Carbon*. **43**(1), 47-52.
- Liu, Y. C., Chen, H. L., Lin, H. K., Liu, W. L., Chou, Y. W., Lo, S. C. and Tai, C. H. (2005). DNA condensation induced by nanoparticle-embedded dendrimer leading to pearl-chain nanowires. *Biomacromolecules*. **6**(6), 3481-3485.
- Liu, Y., Chen, D. and Yan, Y. (2013). Effect of ionic liquids, organic solvents and supercritical CO<sub>2</sub> pretreatment on the conformation and catalytic properties of *Candida rugosa* lipase. *Journal of Molecular Catalysis B: Enzymatic*, **90**, 123-127.
- Madoery, R. R. and Fidelio, G. D. (2001). A simple method to obtain a covalent immobilized phospholipase A2. *Bioorganic and Medicinal Chemistry Letters*. **11**(13), 1663-1664.
- Mansee, A. H., Chen, W. and Mulchandani, A. (2005). Detoxification of the organophosphate nerve agent coumaphos using organophosphorus hydrolase immobilized on cellulose materials. *Journal of Industrial Microbiology and Biotechnology*. **32**(11-12), 554-560.
- Malcata, F. X., Reyes, H. R., Garcia, H. S., Hill, C. G. and Amundson, C. H. (1992). Kinetics and mechanisms of reactions catalysed by immobilized lipases. *Enzyme and Microbial Technology*. **14**(6), 426-446.
- Martinek, K., Semenov, A. N. and Berezin, I. V. (1981). Enzymatic synthesis in biphasic aqueous-organic systems. I. Chemical equilibrium shift. *Biochimica et Biophysica Acta (BBA)-Enzymology*. **658**(1), 76-89.
- Mateo, C., Palomo, J. M., Fuentes, M., Betancor, L., Grazu, V., López-Gallego, F. and Guisán, J. M. (2006). Glyoxyl agarose: A fully inert and hydrophilic support for immobilization and high stabilization of proteins. *Enzyme and Microbial Technology*. **39**(2), 274-280.
- Matsuura, K., Saito, T., Okazaki, T., Ohshima, S., Yumura, M. and Iijima, S. (2006). Selectivity of water-soluble proteins in single-walled carbon nanotube dispersions. *Chemical Physics Letters*. **429**(4-6), 497-502.

- McNeill, G. P., Ackman, R. G. and Moore, S. R. (1996). Lipase-catalyzed enrichment of long-chain polyunsaturated fatty acids. *Journal of the American Oil Chemists' Society*. **73**(11), 1403-1407.
- Micheal, D.D., Noah, N. and John, M.A. (2000). Synthesis of single-walled carbon nanotubes in flames. *Journal of Physical Chemistry B*. **104**, 9615-9620.
- Montero, S., Blanco, A., Virto, M. D., Carlos Landeta, L., Agud, I., Solozabal, R. and Serra, J. L. (1993). Immobilization of *Candida rugosa* lipase and some properties of the immobilized enzyme. *Enzyme and Microbial Technology*. **15**(3), 239-247.
- Moore, C. M., Akers, N. L., Hill, A. D., Johnson, Z. C. and Minter, S. D. (2004). Improving the environment for immobilized dehydrogenase enzymes by modifying Nafion with tetraalkylammonium bromides. *Biomacromolecules*. **5**(4), 1241-1247.
- Moore, S. R. and McNeill, G. P. (1996). Production of triglycerides enriched in long chain n-3 polyunsaturated fatty acids from fish oil. *Journal of the American Oil Chemists' Society*. **73**(11), 1409-1414.
- Murty, V. R., Bhat, J. and Muniswaran, P. K. A. (2002). Hydrolysis of oils by using immobilized lipase enzyme: a review. *Biotechnology and Bioprocess Engineering*. **7**(2), 57-66.
- Nagayama, K., Matsu-ura, S. and Doi, T. (1997). Biocatalytic synthesis of fatty acid ester by lipase in microemulsion system. 4<sup>th</sup> Asia-Pacific. *Biochemical Engineering Conference*. 481-484.
- Nasratun, M., Said, H. A., Noraziah, A. and Alla, A. A. (2009). Immobilization of lipase from *Candida rugosa* on chitosan beads for transesterification reaction. *American Journal of Applied Sciences*. **6**(9), 1653.
- Nepal, D. and Geckeler, K. E. (2006). pH-Sensitive Dispersion and Debundling of Single-Walled Carbon Nanotubes: Lysozyme as a Tool. *Small*. **2**(3), 406-412.
- Nie, Y., Xu, Y., Mu, X. Q., Tang, Y., Jiang, J. and Sun, Z. H. (2005). High-yield conversion of (R)-2-octanol from the corresponding racemate by stereoinversion using *Candida rugosa*. *Biotechnology Letters*. **27**(1), 23-26.
- Niesz, K., Siska, A., Vesselényi, I., Hernadi, K., Méhn, D., Galbács, G. and Kiricsi, I. (2002). Mechanical and chemical breaking of multi walled carbon nanotubes. *Catalysis Today*. **76**(1), 3-10.

- Okada, T. and Morrissey, M. T. (2007). Production of n-3 polyunsaturated fatty acid concentrate from sardine oil by lipase-catalyzed hydrolysis. *Food Chemistry*. **103**(4), 1411-1419.
- Okahata, Y. and Mori, T. (1997). Lipid-coated enzymes as efficient catalysts in organic media. *Trends in Biotechnology*. **15**(2), 50-54.
- Olempska-Beer, Z. S., Merker, R. I., Ditto, M. D. and DiNovi, M. J. (2006). Food-processing enzymes from recombinant microorganisms-a review. *Regulatory Toxicology and Pharmacology*. **45**(2), 144-158.
- Ollis, D. L., Cheah, E., Cygler, M., Dijkstra, B., Frolow, F., Franken, S. M. and Goldman, A. (1992). The  $\alpha/\beta$  hydrolase fold. *Protein Engineering*. **5**(3), 197-211.
- Ozmen, E. Y., Sezgin, M. and Yilmaz, M. (2009). Synthesis and characterization of cyclodextrin-based polymers as a support for immobilization of *Candida rugosa* lipase. *Journal of Molecular Catalysis B: Enzymatic*. **57**(1-4), 109-114.
- Ozyilmaz, G. and Gezer, E. (2010). Production of aroma esters by immobilized *Candida rugosa* and porcine pancreatic lipase into calcium alginate gel. *Journal of Molecular Catalysis B: Enzymatic*. **64**(3-4), 140-145.
- Palmer, T. (1981). *Understanding enzymes*. (2<sup>nd</sup> ed.). England: Ellis Harwood.
- Pandey, A., Benjamin, S., Soccol, C. R., Nigam, P., Krieger, N. and Soccol, V. T. (1999). The realm of microbial lipases in biotechnology. *Biotechnology and Applied Biochemistry*. **29**(2), 119-131.
- Pavlidis, I. V., Vorhaben, T., Tsoufis, T., Rudolf, P., Bornscheuer, U. T., Gournis, D. and Stamatis, H. (2012). Development of effective nanobiocatalytic systems through the immobilization of hydrolases on functionalized carbon-based nanomaterials. *Bioresource Technology*, 115, 164-171.
- Perez, V. H., da Silva, G. S., Gomes, F. M. and de Castro, H. F. (2007). Influence of the functional activating agent on the biochemical and kinetic properties of *Candida-rugosa* lipase immobilized on chemically modified cellulignin. *Biochemical Engineering Journal*. **34**(1), 13-19.
- Pitcher Jr, W. H. (1980). Immobilized enzymes for food processing. CRC Press Inc.

- Planeix, J. M., Coustel, N., Coq, B., Brotons, V., Kumbhar, P. S., Dutartre, R. and Ajayan, P. M. (1994). Application of carbon nanotubes as supports in heterogeneous catalysis. *Journal of the American Chemical Society*. **116**(17), 7935-7936.
- Prlainović, N. Z., Bezbradica, D. I., Knežević-Jugović, Z. D., Stevanović, S. I., Avramov Ivić, M. L., Uskoković, P. S. and Mijin, D. T. (2013). Adsorption of lipase from *Candida rugosa* on multi walled carbon nanotubes. *Journal of Industrial and Engineering Chemistry*. **19**(1), 279-285.
- Pryakhin, A. N., Chukhrai, E. S. and Poltorak, O. M. (1977). Glucose 6-phosphate dehydrogenase immobilized by adsorption on silica gel solid supports. *Vest Moskov University Ser 2 Khim*. **18**(1), 125.
- Rahman, R. N. Z. R. A., Baharum, S. N., Basri, M. and Salleh, A. B. (2005). High-yield purification of an organic solvent-tolerant lipase from pseudomonas sp. strain S5. *Analytical Biochemistry*. **341**(2), 267-274
- Rangheard, M. S., Langrand, G., Triantaphylides, C. and Baratti, J. (1989). Multi-competitive enzymatic reactions in organic media: a simple test for the determination of lipase fatty acid specificity. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*. **1004**(1), 20-28.
- Resasco, D. E., Alvarez, W. E., Pompeo, F., Balzano, L., Herrera, J. E., Kitiyanan, B. and Borgna, A. (2002). A scalable process for production of single-walled carbon nanotubes (SWNTs) by catalytic disproportionation of CO on a solid catalyst. *Journal of Nanoparticle Research*. **4**(1-2), 131-136.
- Sadeghian, H., Seyedi, S. M., Saberi, M. R., Arghiani, Z. and Riazzi, M. (2008). Design and synthesis of eugenol derivatives, as potent 15-lipoxygenase inhibitors. *Bioorganic and Medicinal Chemistry*. **16**(2), 890-901.
- Saito, S. (1997). Carbon nanotubes for next-generation electronics devices. *Science*. **278**(5335), 77-78.
- Salimon, J. and Salih, N. (2009). Improved low temperature properties of 2-ethylhexyl 9(10)-hydroxy-10(9)-acyloxystearate derivatives. *European Journal of Scientific Research*. **31**(4), 583-591.
- Sánchez, A., Ferrer, P., Serrano, A., Pernas, M. A., Valero, F., Rúa, M. L. and Solà, C. (1999). Characterization of the lipase and esterase multiple forms in an enzyme preparation from a *Candida rugosa* pilot-plant scale fed-batch fermentation. *Enzyme and Microbial Technology*. **25**(3), 214-223.

- Satoh, K., Sakagami, H., Yokoe, I., Kochi, M. and Fujisawa, S. (1998). Interaction between eugenol-related compounds and radicals. *Anticancer Research*. **18**(1), 425-428.
- Saxena, R. K., Sheoran, A., Giri, B. and Davidson, W. S. (2003). Purification strategies for microbial lipases. *Journal of Microbiological Methods*. **52**(1), 1-18.
- Scherer, R. P., Dallago, R. L., Penna, F. G., Bertella, F., de Oliveira, D., de Oliveira, J. V. and Pergher, S. B. C. (2012). Influence of process parameters on the immobilization of commercial porcine pancreatic lipase using three low-cost supports. *Biocatalysis and Agricultural Biotechnology*. **1**(4), 290-294.
- Schmid, R. D. and Verger, R. (1998). Lipases: interfacial enzymes with attractive applications. *Angewandte Chemie International Edition*. **37**(12), 1608-1633.
- Schmitt-Rozieres, M., Vanot, G., Deyris, V. and Comeau, L. C. (1999). Borago officinalis oil: Fatty acid fractionation by immobilized *Candida rugosa* lipase. *Journal of the American Oil Chemists' Society*. **76**(5), 557-562.
- Serp, P., Corrias, M. and Kalck, P. (2003). Carbon nanotubes and nanofibers in catalysis. *Applied Catalysis A: General*. **253**(2), 337-358.
- Shah, S., Solanki, K. and Gupta, M. N. (2007). Enhancement of lipase activity in non aqueous media upon immobilization on multi-walled carbon nanotubes. *Chemistry Central Journal*. **1**(30), 1-6.
- Sheldon, R. A. (2007). Enzyme immobilization: The quest for optimum performance. *Advanced Synthesis and Catalysis*. **349**(8-9), 1289-1307.
- Shi, Y., Jin, F., Wu, Y., Yan, F., Yu, X. and Quan, Y. (1997). Improvement of immobilized cells through permeabilizing and crosslinking. **13**(1), 111-113.
- Shim, M., Shi Kam, N. W., Chen, R. J., Li, Y. and Dai, H. (2002). Functionalization of carbon nanotubes for biocompatibility and biomolecular recognition. *Nano Letters*. **2**(4), 285-288
- Shimada, Y., Fukushima, N., Fujita, H., Honda, Y., Sugihara, A. and Tominaga, Y. (1998). Selective hydrolysis of borage oil with *Candida rugosa* lipase: two factors affecting the reaction. *Journal of the American Oil Chemists' Society*. **75**(11), 1581-1586.
- Shnyrov, V. L., Martínez, L. D., Roig, M. G., Lyubarev, A. E., Kurganov, B. I. and Villar, E. (1999). Irreversible thermal denaturation of lipase B from *Candida rugosa*. *Thermochimica Acta*. **325**(2), 143-149.

- Shu, C., Cai, J., Huang, L., Zhu, X. and Xu, Z. (2011). Biocatalytic production of ethyl butyrate from butyric acid with immobilized *Candida rugosa* lipase on cotton cloth. *Journal of Molecular Catalysis B: Enzymatic*. **72**(3), 139-144.
- Soo, E. L., Salleh, A. B., Basri, M., Rahman, R. A. and Kamaruddin, K. (2004). Response surface methodological study on lipase-catalyzed synthesis of amino acid surfactants. *Process Biochemistry*. **39**(11), 1511-1518.
- Spahn, C. and Minteer, S. D. (2008). Enzyme immobilization in biotechnology. *Recent patents on Engineering*. **2**(3), 195-200.
- Stamatis, H., Xenakis, A., Menge, U. and Kolisis, F. N. (1993). Kinetic study of lipase catalyzed esterification reactions in water-in-oil microemulsions. *Biotechnology and Bioengineering*. **42**(8), 931-937.
- Stamatis, H., Sereti, V. and Kolisis, F. N. (2001). Enzymatic synthesis of hydrophilic and hydrophobic derivatives of natural phenolic acids in organic media. *Journal of Molecular Catalysis B: Enzymatic*. **11**(4), 323-328.
- Steytler, D. C., Moulson, P. S. and Reynolds, J. (1991). Biotransformations in near critical carbon dioxide. *Enzyme and Microbial Technology*. **13**(3), 221-226.
- Subramanian, A., Kennel, S. J., Oden, P. I., Jacobson, K. B., Woodward, J. and Doktycz, M. J. (1999). Comparison of techniques for enzyme immobilization on silicon supports. *Enzyme and Microbial Technology*. **24**(1-2), 26-34.
- Suzuki, Y., Sugiyama, K. and Furuta, H. (1985). Eugenol-mediated superoxide generation and cytotoxicity in guinea pig neutrophils. *Japanese Journal of Pharmacology*. **39**(3), 381-386.
- Takac, S. and Bakkal, M. (2007). Impressive effect of immobilization conditions on the catalytic activity and enantioselectivity of *Candida rugosa* lipase toward S-Naproxen production. *Process Biochemistry*. **42**(6), 1021-1027.
- Tasis, D., Tagmatarchis, N., Bianco, A. and Prato, M. (2006). Chemistry of carbon nanotubes. *Chemical Reviews*. **106**(3), 1105-1136.
- Torres, S. and Castro, G. R. (2004). Non-aqueous biocatalysis in homogeneous solvent systems. *Food Technology and Biotechnology*. **42**(4), 271-277.
- Tosa, T., Sato, T., Mori, T., Yamamoto, K., Takata, I., Nishida, Y. and Chibata, I. (1979). Immobilization of enzymes and microbial cells using carrageenan as matrix. *Biotechnology and Bioengineering*. **21**(10), 1697-1709.

- Tres, M. V., Ferraz, H. C., Dallago, R. M., Di Luccio, M. and Oliveira, J. V. (2010). Characterization of polymeric membranes used in vegetable oil/organic solvents separation. *Journal of Membrane Science*. **362**(1), 495-500.
- Tsai, H. and Doong, R. (2007). Preparation and characterization of urease-encapsulated biosensors in poly(vinyl alcohol)-modified silica sol-gel materials. *Biosensors and Bioelectronics*. **23**(1), 66-73.
- Uppenberg, J., Patkar, S., Bergfors, T. and Jones, T. A. (1994). Crystallization and preliminary X-ray studies of lipase B from *Candida antarctica*. *Journal of Molecular Biology*. **235**(2), 790-792.
- Vaidya, B. K., Ingavle, G. C., Ponrathnam, S., Kulkarni, B. D. and Nene, S. N. (2008). Immobilization of *Candida rugosa* lipase on poly (allyl glycidyl ether-co-ethylene glycol dimethacrylate) macroporous polymer particles. *Bioresource Technology*. **99**(9), 3623-3629.
- Vikbjerg, A. F., Peng, L., Mu, H. and Xu, X. (2005). Continuous production of structured phospholipids in a packed bed reactor with lipase from thermomyces lanuginose. *JAACS, Journal of the American Oil Chemists' Society*. **82**(4), 237-242.
- Villeneuve, P. and Foglia, T. A. (1997). Lipase specificities: Potential application in lipid bioconversions. *INFORM-International News on Fats, Oils and Related Material*. **8**(6), 640-650.
- Villeneuve, P., Barēa, B., Sarrazin, P., Davrieux, F., Boulanger, R., Caro, Y. and Graille, J. (2003). Synthesis of pyroglutamic acid fatty esters through lipase-catalyzed esterification with medium chains alcohols. *Enzyme and Microbial Technology*. **33**(1), 79-84.
- Villeneuve, P., Muderhwa, J. M., Graille, J. and Haas, M. J. (2000). Customizing lipases for biocatalysis: a survey of chemical, physical and molecular biological approaches. *Journal of Molecular Catalysis B: enzymatic*. **9**(4), 113-148.
- Wahab, R. A., Basri, M., Rahman, M. B. A., Rahman, R. N. Z. R. A., Salleh, A. B. and Leow, T. C. (2012). Combination of oxyanion Gln114 mutation and medium engineering to influence the enantioselectivity of thermophilic lipase from geobacillus zalihae. *International Journal of Molecular Sciences*. **13**(9), 11666-11680.

- Wan, L. S., Ke, B. B. and Xu, Z. K. (2008). Electrospun nanofibrous membranes filled with carbon nanotubes for redox enzyme immobilization. *Enzyme and Microbial Technology*. **42**(4), 332-339.
- Wang, H., Chhowalla, M., Sano, N., Jia, S. and Amaratunga, G. A. J. (2004). Large-scale synthesis of single-walled carbon nanohorns by submerged arc. *Nanotechnology*. **15**(5), 546-550.
- Wang, J., Musameh, M. and Lin, Y. (2003). Solubilization of carbon nanotubes by nafion toward the preparation of amperometric biosensors. *Journal of the American Chemical Society*. **125**(9), 2408-2409.
- Wang, L., Wei, L., Chen, Y. and Jiang, R. (2010). Specific and reversible immobilization of NADH oxidase on functionalized carbon nanotubes. *Journal of Biotechnology*. **150**(1), 57-63.
- Wang, X. and Cao, J. (2001). Wrinkling limit in tube bending. *Journal of Engineering Materials and Technology*. **123**(4), 430-435.
- Wang, Y., Iqbal, Z. and Malhotra, S. V. (2005). Functionalization of carbon nanotubes with amines and enzymes. *Chemical Physics Letters*. **402**(1-3), 96-101.
- Wang, Y., Joshi, P. P., Hobbs, K. L., Johnson, M. B. and Schmidtke, D. W. (2006). Nanostructured biosensors built by layer-by-layer electrostatic assembly of enzyme-coated single-walled carbon nanotubes and redox polymers. *Langmuir*. **22**(23), 9776-9783.
- Won, K., Kim, S., Kim, K. J., Park, H. W. and Moon, S. J. (2005). Optimization of lipase entrapment in Ca-alginate gel beads. *Process Biochemistry*. **40**(6), 2149-2154.
- Woolley, P. and Petersen, S. B. (Eds.) (1994). *Lipases: their structure, biochemistry and application*. Cambridge: Cambridge University Press.
- Wu, J. C., Song, B. D., Xing, A. H., Hayashi, Y., Talukder, M. M. R. and Wang, S. C. (2002). Esterification reactions catalyzed by surfactant-coated *Candida rugosa* lipase in organic solvents. *Process Biochemistry*. **37**(11), 1229-1233.
- Wu, C., Zhou, G., Jiang, X., Ma, J., Zhang, H. and Song, H. (2012). Active biocatalysts based on *Candida rugosa* lipase immobilized in vesicular silica. *Process Biochemistry*. **47**(6), 953-959.



- Wu, X., Li, J., Li, X., Hsieh, C. L., Burgers, P. M. and Lieber, M. R. (1996). Processing of branched DNA intermediates by a complex of human FEN-1 and PCNA. *Nucleic Acids Research*. **24**(11), 2036-2043.
- Xu, H., Li, M. and He, B. (1995). Immobilization of *Candida cylindracea* lipase on methyl acrylate-divinyl benzene copolymer and its derivatives. *Enzyme and Microbial Technology*. **17**(3), 194-199.
- Yadav, G. D. and Yadav, A. R. (2012). Insight into esterification of eugenol to eugenol benzoate using a solid super acidic modified zirconia catalyst UDCaT-5. *Chemical Engineering Journal*, *192*, 146-155.
- Yang, K. and Xing, B. (2007). Desorption of polycyclic aromatic hydrocarbons from carbon nanomaterials in water. *Environmental Pollution*. **145**(2), 529-537.
- Yang, I.R., Xu, H.J., Yao, S.J., and Zhu, Z.Q. (1997). Kinetics of butyl acetate synthesis by lipase-catalyzed transesterification in hexane. 4<sup>th</sup> Asia-Pacific Biochemical engineering. Conference, 296-299.
- Ye, P., Xu, Z. K., Wu, J., Innocent, C. and Seta, P. (2006). Nanofibrous poly (acrylonitrile-co-maleic acid) membranes functionalized with gelatin and chitosan for lipase immobilization. *Biomaterials*. **27**(22), 4169-4176.
- Yeşiloğlu, Y. (2005). Utilization of bentonite as a support material for immobilization of *Candida rugosa* lipase. *Process Biochemistry*. **40**(6), 2155-2159.
- Yilmaz, E., Can, K., Sezgin, M. and Yilmaz, M. (2011). Immobilization of *Candida rugosa* lipase on glass beads for enantioselective hydrolysis of racemic Naproxen methyl ester. *Bioresource Technology*. **102**(2), 499-506.
- Yu, W. H., Fang, M., Tong, D. S., Shao, P., Xu, T. N. and Zhou, C. H. (2013). Immobilization of *Candida rugosa* lipase on hexagonal mesoporous silicas and selective esterification in nonaqueous medium. *Biochemical Engineering Journal*, *70*, 97-105.
- Zaidi, A., Gainer, J. L. and Carta, G. (1995). Fatty acid esterification using nylon-immobilized lipase. *Biotechnology and Bioengineering*. **48**(6), 601-605.
- Zaks, A. and Klivanov, A. M. (1985). Enzyme-catalyzed processes in organic solvents. *Proceedings of the National Academy of Sciences*. **82**(10), 3192-3196.

- Zhang, D. H., Yuwen, L. X., Li, C. and Li, Y. Q. (2012). Effect of poly (vinyl acetate acrylamide) microspheres properties and steric hindrance on the immobilization of *Candida rugosa* lipase. *Bioresource Technology*, 124, 233-236.
- Zhang, P. and Henthorn, D. B. (2010). Synthesis of PEGylated single wall carbon nanotubes by a photoinitiated graft from polymerization. *AIChE Journal*. **56**(6), 1610-1615.
- Zhang, S., Minus, M. L., Zhu, L., Wong, C. P. and Kumar, S. (2008). Polymer transcrystallinity induced by carbon nanotubes. *Polymer*. **49**(5), 1356-1364.
- Zhao, H., Holladay, J. E., Brown, H. and Zhang, Z. C. (2007). Metal chlorides in ionic liquid solvents convert sugars to 5-hydroxymethylfurfural. *Science*. **316**(5831), 1597-1600.