

ANTI INFLAMMATORY AND ANTI OXIDANT PROPERTIES OF PURIFIED  
STILBENE AND CRUDE EXTRACTS OF PERLIS *Vitis vinifera* ON CULTURED  
HUVECs

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*In the memories of my late grandmother,*

*Tok Baayah binti Che Mat*

*May ALLAH rest her soul in eternal peace (1927- April 2013)*

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## ABSTRACT

Air-dried and particle form of stems and leaves of *Vitis vinifera* cultivated in Perlis, Malaysia were extracted using cold, back, methanolic and ethanolic extraction methods. The ethyl acetate stems extract by back extraction (STBE EA) possessed the highest total phenolic content ( $916.27 \pm 0.01$ ) by Folin-Ciocalteu assay and radical scavenging activity at  $IC_{50} = 18 \mu\text{g/mL}$  by DPPH assay. The extracts was further investigated by thin layer chromatography resulted in detection of resveratrol at  $R_f = 0.425$  in STBE EA using petroleum ether:ethyl acetate at ratio 2:3. Advanced screening method of phenolic profiling by LC-MS/MS has detected seven phenolic compounds which were identified through LC-MS/MS library and Mass Bank Software. The compounds are *trans*-resveratrol, *trans*-cinnamic acid,  $\epsilon$ -viniferin, salicylic and caffeic acid, catechin, and epigallocatechin. Purification of the STBE EA has resulted in the isolation of *trans*-stilbene which was identified as 3, 4', 5-trihydroxystilbene (*trans*-resveratrol) by Nuclear Magnetic Resonance. The STBE EA and *trans*-resveratrol exhibited protective effects on HUVECs against oxidative cell damage by hydrogen peroxide ( $IC_{50} = 205 \mu\text{M}$ ) with survival rate 17.69% ( $IC_{50} = 1 \mu\text{g/mL}$ ) and 40.80% ( $IC_{50} = 0.01 \mu\text{M}$ ) and induced nitric oxide 3.41 ( $IC_{50} = 0.001 \mu\text{g/mL}$ ) and 3.96 mm ( $IC_{50} = 0.001 \mu\text{M}$ ) respectively. STBE EA and *trans*-resveratrol possessed dual inhibition of COX-2 and 15-LOX  $85.07 \pm 1.20$  (COX-2) and 60% with  $IC_{50}$  at  $7.5 \mu\text{M}$  (15-LOX) respectively. Furthermore purified *trans*-resveratrol exhibited better inhibition at  $96.87 \pm 0.10\%$  for COX-2 and 79.57% for 15-LOX. This study demonstrated that STBE EA extracts and *trans*-resveratrol isolated from *Vitis vinifera* stems posses potent anti-oxidant and anti-inflammatory properties.

## ABSTRAK

Pengekstrakan batang dan daun *Vitis vinifera* yang ditanam di Perlis, Malaysia diekstrak menggunakan teknik rendaman sejuk, bersiri, metanol dan etanol. Keseluruhan 16 ekstrak menjalani penyaringan terhadap ciri-ciri anti-oksidasi. Ekstrak etil asetat batang (STBE EA) melalui kaedah pengekstrakan bersiri memiliki kandungan jumlah fenol tertinggi ( $916.27 \pm 0.01$ ) melalui kaedah Folin-Ciocalteu dan pemerangkapan radikal bebas pada  $IC_{50}$  18  $\mu\text{g/mL}$  melalui kaedah DPPH. Komposisi fenolik di dalam ekstrak terus dikaji dan menunjukkan kehadiran resveratrol di  $R_f = 0.425$  pada ekstrak STBE EA menggunakan petroleum eter:etil asetat pada nisbah 2:3. Kaedah saringan profil fenolik oleh LC-MS/MS berjaya mengesan tujuh sebatian fenolik yang dikenal pasti melalui pengumpulan data LC-MS/MS dan perisian Mass Bank. Kompaun-kompaun dikenali sebagai *trans*-resveratrol, asid *trans*-cinnamic,  $\epsilon$ -viniferin, asid salisilik, asid caffeic, catechin, dan epigallocatechin. Penulenan STBE EA telah menemui *trans*-stilbene yang telah dikenal pasti sebagai 3, 4', 5-trihydroxystilbene (*trans*-resveratrol) berdasarkan kaedah spektroskopi oleh Resonans Magnetik Nuklear. STBE EA dan *trans*-resveratrol masing-masing menunjukkan kesan perlindungan pada HUVECs terhadap kerosakan oksidatif sel oleh hidrogen peroksida ( $IC_{50} = 205 \mu\text{M}$ ) dengan peningkatan sel hidup 17.69% ( $IC_{50} = 1 \mu\text{g/mL}$ ) dan 40.80% ( $IC_{50} = 0.01 \mu\text{M}$ ) dan nitrik oksida teraruh pada kadar 3.41 ( $IC_{50} = 0.001 \mu\text{g/mL}$ ) dan 3.96 mm ( $IC_{50} = 0.001 \mu\text{M}$ ). STBE EA dan *trans*-resveratrol memiliki dwi-perencatan COX-2 dan 15-LOX pada  $85.07 \pm 1.20$  dan 60% dengan  $IC_{50}$  pada 7.5  $\mu\text{M}$  masing-masing. *Trans*-resveratrol tulen menunjukkan perencatan tertinggi pada  $96.87 \pm 0.10\%$  untuk COX-2 dan 79.57% untuk 15-LOX. Kajian ini menunjukkan bahawa ekstrak STBE EA dan *trans*-resveratrol yang diasingkan daripada batang *Vitis vinifera* memiliki sifat-sifat anti-oksidasi dan anti-radang.

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**LIST OF ABBREVIATIONS**

%	Percentage
°C	Degree celcius
µg	Microgram
µM	Micromolar
<sup>13</sup> C	13-Carbon
15-LOX	15-Lipoxygenase
<sup>1</sup> H	1-Hydrogen
5-LOX	5-Lipoxygenase
ANOVA	Analysis of variance
C	Carbon
CC	Column chromatography
CO <sub>2</sub>	Carbon dioxide
COX-1	Cyclooxygenase-1
COX-2	Cyclooxygenase-2
DMSO	Dimethyl sulfoxide
DPPH assay	2,2-diphenyl-1-picrylhydrazyl
EGM-2	Endothelial growth media-2
EIA	Enzyme immunoassay
ESI	Electrospray ionization
<i>et al.</i> ,	And others
<i>etc.</i>	<i>Et cetera</i>
EtOH	Ethanol
g	Gram



GAE	Gallic acid equivalent
GC	Gas chromatography
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
H <sub>2</sub> O <sub>2</sub>	Hydrogen peroxide
He	Helium
HUVECs	Human umbilical vein endothelial cells
Hz	Hertz
IC <sub>50</sub>	Inhibitory concentration at 50 %
L/min	Litre/minutes
LC	Liquid chromatography
LC-MS/MS	Liquid chromatography tandem mass spectrometry
LDL	Low density lipoprotein
LTs	Leucotrienes
LV80 ET	Leaves crude extracts of <i>Vitis vinifera</i> derives from 80% ethanol extraction techniques using ethanol to water
LV80 ME	Leaves crude extracts of <i>Vitis vinifera</i> derives from 80% methanol extraction techniques using methanol to water
LVBE EA	Leaves crude extracts of <i>Vitis vinifera</i> derives from back extraction techniques using methanol
LVBE ME	Leaves crude extracts of <i>Vitis vinifera</i> derives from back extraction techniques using methanol
LVBE PE	Leaves crude extracts of <i>Vitis vinifera</i> derives from back extraction techniques using methanol
LVCE EA	Leaves crude extracts of <i>Vitis vinifera</i> derives from cold extraction techniques using ethyl acetate
LVCE ME	Leaves crude extracts of <i>Vitis vinifera</i> derives from cold extraction techniques using methanol
LVCE PE	Leaves crude extracts of <i>Vitis vinifera</i> derives from cold extraction techniques using petroleum ether
LXs	Lipoxins

$m/z$	Mass to charge ratio
MeOH	Methanol
mg	Milligram
MHz	Mega hertz
min	Minute
mL	Milliliter
mM	Millimole
MS <sup>2</sup>	Mass spectrometry/mass spectrometry
MTT assay	3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
N <sub>2</sub>	Nitrogen
NaOH	Sodium Hydroxide
nm	Nanometer
NMR	Nucleus magnetic resonance
NO	Nitric oxide
OH	Hydroxyl group
ROS	Reactive oxygen species
SFC	Supercritical fluid chromatography
SiO <sub>2</sub>	Silica gel
SPSS	Statistical analysis system
ST80 ET	Stem crude extracts of <i>Vitis vinifera</i> derives from 80% ethanol extraction techniques using ethanol to water
ST80 ME	Stem crude extracts of <i>Vitis vinifera</i> derives from 80% methanol extraction techniques using methanol to water
STBE EA	Stem crude extracts of <i>Vitis vinifera</i> derives from back extraction techniques using ethyl acetate
STBE ME	Stem crude extracts of <i>Vitis vinifera</i> derives from back extraction techniques using methanol
STBE PE	Stem crude extracts of <i>Vitis vinifera</i> derives from back extraction techniques using petroleum ether
STCE EA	Stem crude extracts of <i>Vitis vinifera</i> derives from cold extraction techniques using ethyl acetate
STCE ME	Stem crude extracts of <i>Vitis vinifera</i> derives from cold

	extraction techniques using methanol
STCE PE	Stem crude extracts of <i>Vitis vinifera</i> derives from cold extraction techniques using petroleum ether
TPC	Total phenolic content
USDA	United states department of agriculture
UV	Ultra Violet light
v/v	Volume over volume
VLC	Vacuum liquid chromatography
w/w	Weight over weight
WHO	World Health Organisation

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## CHAPTER 1

### INTRODUCTION

#### 1.1. Background of Research

Consumption of plant derived medicine has experienced rapid growing interest and is increasing extensively in both traditional and modern medicinal purpose. Since Sumerian civilisation dated 400 years before the Common Era, the therapeutic use of plants has been discovered. In western medicine, it has been reported that Dioscorides has successfully produced *De Materia Medica*, a compilation of information on more than 600 species of plants with medicinal value (Borzelleca *et al.*, 2008) while Hippocrates has been recorded using approximately 4000 different plant species for medical purpose. Natural products play a major role in the ancient traditional medicine system.

In decades, the uses of natural derived medicine were still in practice by Chinese, Ayurveda, and Egyptian (Sarker *et al.*, 2006). According to the statistic data by World Health Organization (WHO), herbal plant have been used for healthcare requirements by more than 80% of the world population, generally in developing countries (Farnsworth *et al.*, 1985; Canter *et al.*, 2005). After extensive research over 50 years, the National Institutes of Health have recently suggested that a diet rich in fresh foods derives from plant may reduces the risk of diseases including inflammation and cardiovascular diseases. Indeed, the right foods may act as medicines which increase the strength of immune system.

Cragg *et al.* (2002) has reported that, several numbers of modern drugs have been developed from medicinal plants due to their rich sources of bioactive compounds over the last century. Natural compounds such as phenolic acids, stilbenes, flavonoids, isoflavonoids, and lignans present diversely in plants may be responsible for their health benefits.

Based on previous studies, Vitaceae family is one of dicotyledonous flowering plants that produce many bioactive compounds in their biological activity. *V. vinifera* or commonly known as grapevine which belong to Vitaceae family are native to Mediterranean region, and Central Europe. It is liana flaky bark, with alternate leaves, palmately lobed and broad. The fruit is a berry known as grape. Grape was originally used in winemaking and the leaves were widely used in Indian Ayurvedic medicinal purposed. Since 1980 from its first trial to be grown in Malaysia, grape has experienced rapid growing interest in Malaysia's agricultural sector due to its high demand of local grown grape fruits. The sector itself has grown up to thousand hectares in 2010. After pruning season, large quantity of stems and leaves would remain on the field producing between 0.62 to 2.03 kg per plant. It is estimated that average pruning waste yielded approximately 5 tonnes per hectare of plant per year.

Ignat *et al.* (2011) proposed that phenolic compounds including flavonoids and stilbenes were found abundantly in agro-industrial residues compare to fruits and fresh vegetables itself. Phenolic compound comprise a large group of secondary metabolites obtain from phenylalanine and widely distributed throughout the plant. Phenolic compounds plays a pivotal role as imparting colours in plants' parts (leaves and fruits), appealing or repelling insects, and plant self-protection against harmful attack or injuries and ultraviolet radiation.

Chemically, phenolic compounds comprising an aromatic ring bearing one or more hydroxyl groups, including their derivatives. More than 8000 phenolic compounds have been identified in plants which predominantly substituted derivatives of hydrobenzoic and hydrocinnamic acids (Robbins, 2003; Wrolstad, 2005). These derivatives differ in patterns of hydroxylation process of their aromatic

rings (Harborne, 1994). The phenolic composition and its biosynthesis in plant are highly influence by viticulture and various biotic factors such as temperature, sunlight, rainfall, pathogenesis and soil nutrient content.

Downey *et al.*, (2006) observing the reduction in grape berry colour in hot climate. An additional climate impact on phenolic compounds is the positive relationship between sunlight exposure and increased flavonol accumulation. Mori *et al.*, (2005) reported that anthocyanin levels in Cabernet Sauvignon grapes are higher when day temperatures are constant at 20°C compared to 30°C. Therefore, increasing in anthocyanin content is highly related with grapes grown at higher altitudes. However, this association is complicated by the effect of diurnal differences in temperature with lower night temperature result in greater accumulation of anthocyanins.

Increasing in epidemiological studies and data collections suggests that high intake of fresh plant-based food provides an extensive number of health benefits against degenerative disease. Due to diverse phenolic composition in the plant, it possessed antioxidant activity which contributes to their protective effects against cardiovascular disease and cancer (Howitz *et al.*, 2003; Cho *et al.*, 2006; Valenzano *et al.*, 2006). Several specific plant phenolic compounds derives from plant extracts such as resveratrol and quercetin have been reported to exhibit anti-oxidant and anti-inflammatory via *in vitro* or *in vivo* (Pearson, 2008).

Therefore, in assessing the potential value of *V.vinifera* residues, it is pertinent to determine the phenolic compounds composition of its crude extracts and its anti-oxidant and anti-inflammatory properties. It has been shown that anti-oxidant and anti-inflammatory property varies with types of phenolic compounds present in the plant. Furthermore, the combination between phenolic compounds or together with other phytochemicals in extracts may react by different mechanisms, often interact synergistically or inhibitorily (Fuhrman *et al.*, 2000; Graversen *et al.*, 2008).



Inflammation is a basic fundamental pathological mechanism which underlies an array of disease (Tedgui, 2001). The inflammatory reaction may occur by interactions between inflammatory cells such as neutrophils, lymphocytes, and monocytes, and vascular cells - endothelial cells and smooth muscle cells. The nature of inflammatory reaction can potentially be stimulated by multiple cytokines and growth factors which are present at sites of inflammation. Endothelial cells and smooth muscle cells must integrate the signals produced by these multiple factors to effectively regulate the immunoinflammatory response during the expression of adhesion molecules, cytokines, chemokines, matrix metalloproteinases, and growth factors.

Research in vascular biology has progressed astonishingly in last decade, resulting in a better understanding of the vascular cell responses to inflammatory stimuli and identification of major intracellular inflammatory signalling pathways, particularly nuclear factor- $\kappa$ B (NF- $\kappa$ B) system. NF- $\kappa$ B may be found in all animal cell types and entailed in cellular response to stimuli stress, cytokines, free radicals, and oxidized Low Density Lipoprotein (LDL).

This study is designed to screen and build phenolic compound profiling, isolating targeted stilbene and investigate the anti-oxidant and anti-inflammatory effect of this tropical isolated stilbene on Human Umbilical Vein Endothelial Cell (HUVEC) by COX-2, 15-LOX and Nitric Oxide (NO) assay.

## **1.2. Problem statement**

Phenolic compounds and its biosynthesis were present in plant tissue as phytoalexins which is induced by biotic and abiotic stresses. Hence, the amount of phenolic compounds in grape is based on the variety of grapevine and influenced by viticultural (greenhouse, *etc.*) and environmental factors such as light, environment temperature, altitude, soil nutritional status, rain, pathogenesis, and other various development process (Downey *et al.*, 2006). Thus, the attention of this research is

focused on the stilbenes content from grapes of tropical countries such as Malaysia which has a hot, humid tropical climate throughout the year.

### **1.3. Objectives of study**

The objectives of this research are:

1. To evaluate total phenolic content and anti-oxidant activities of plant crude extracts by Folin-Ciocalteu and DPPH assay.
2. To develop LC-MS/MS library and rapid screening of phenolic compounds in selected crude extract of *V.vinifera*.
3. To isolate, purify, identify, and quantify targeted compound from selected active crude extract by Vacuum Liquid and Column Chromatography, and NMR Spectro-analysis.
4. To evaluate oxidative damage by hydrogen peroxide and protective effect of purified stilbene and selected crude extract on HUVECs
5. To evaluate the anti-inflammatory effect of purified stilbene and selected crude extract by production of Nitric Oxide and dual inhibition of COX-2 and 15-LOX.

### **1.4. Scope of study**

In this research, stems and leaves of *V.vinifera* were extracted using four different extraction methods. The total phenolic content and anti-oxidant activities were evaluated by Folin-Ciocalteu and DPPH assay. Rapid screening in phenolic compounds profiling were analysed by simple thin layer chromatography technique and LC-MS/MS spectra. The selected crude extract by screening of its potential antioxidant properties and phenolic compounds profile was further undergo partitioning by using Vacuum Liquid Chromatography technique, purifies using column chromatography and was assessed by using  $^1\text{H}$  and  $^{13}\text{C}$  of NMR spectral

data. NMR was used to evaluate the purities of compound isolated and so as to identify the structure of isolated compound. The anti-inflammatory effect of purified compound and selected crude were evaluated on Human Umbilical Vein Endothelial Cell (HUVEC) via COX-2 inhibition assay, 15-LOX colometric inhibition assay and nitric oxide production by Griess assay.

### **1.5. Significant of Study**

Nature has been a fundamental source of therapeutic agents for centuries which led to the developments of modern drugs typically based on their used in ancient traditional medicine. However, these modern derives drugs such as nonsteroidal anti-inflammatory drugs (NSAIDs) have its negative side-effects on human. Thus, the results obtain from this study will be used as further guidance in research work in findings new bioactive compounds led to more effective natural anti-inflammatory medicine with less side effects on human consumption. This study will contribute mainly in the science and clinical chemistry.

## REFERENCES

- Aggarwal, B. B., Shishodia, S., Sandur, S. K., Pandey, M. K. and Sethi, G. (2006). Inflammation and cancer: how hot is the link? *Journal of Biochemistry and Pharmacology*. 72: 1605-1621.
- Alderton W. K., Cooper, C. E., and Knowles R. G. (2001). Nitric oxide synthases: structure, function and inhibition. *Biochem J*. 357: 593-615.
- Alonso-Garcia, A., Cancho-Grande, B., Simal and Gandara, J. (2004). Development of a rapid method based on solid-phase extraction and liquid chromatography with ultraviolet absorbance detection for the determination of polyphenols in alcohol-free beers. *Journal of Chromatography A*. 1054: 175–180.
- Altıok, E., Baycin, D., Bayraktar, O., Ulku S. (in press). Isolation of polyphenols from the extracts of olive leaves (*Olea europaea* L.) by adsorption on silk fibroin. *Separation and Purification Technology*.
- Amelia, F., Galih N. A., Arini M., Alia N. F., Sisca U., and Mimiek M. (2012). Extraction and Stability Test of Anthocyanin from Buni Fruits (*Antidesma Bunius* L) as an Alternative Natural and Safe Food Colorants. *J.Food Pharm.Sci*. 1: 49-53.
- Amr, A., Al-Tamimi, E. (2007). Stability of the crude extracts of *Ranunculus asiaticus* anthocyanins and their use as food colourants. *International Journal of Food Science Technology*. 42: 985–991.

- Andreasen, M. F., Christensen, L. P., Meyer, A. S. and Hansen, A. (2000). Content of phenolic acids and ferulic acid dehydrodimers in 17 rye (*Secale cereale* L.) varieties. *Agricultural and Food Chemistry*. 48: 2837 - 2842.
- Arichi, H., Kimura, Y., Okuda, H., Baba, K., Kozawa, M. and Arichi, S. (1982). Effects of stilbene components of the roots of *Polygonum cuspidatum* Sieb. et Zucc. on lipid metabolism. *Chem Pharm Bull, Tokyo*. 30: 1766-1770.
- Arnold C., Schnitzler, A., Douard, A., Peter, R., Gillet, F. (2005). Is there a future for wild grapevine (*Vitis vinifera* subsp. *silvestris*) in the Rhine Valley? *Biodiversity and Conservation*. 14: 1507–1523.
- Arnold, C., Gillet, F., Gobat, J. M. (1998). Situation de la vigne sauvage *Vitis vinifera* subsp. *silvestris* en Europe. *Vitis*. 37: 159–170.
- Artaud-Wild, S. M., Connor, S. L., Sexton, G. (1993). Differences in coronary mortality can be explained by differences in cholesterol and saturated fat intakes in 40 countries but not in France and Finland. A paradox. *Circulation*. 88:2771–2779.
- Aruoma, O. I. (2002). Methodological considerations for characterizing potential antioxidant actions of bioactive components in plant foods. *Mutation Research*. 523 - 524: 9 - 20.
- Augustin, H. G., Kozian, D. H. and Johnson, R. C. (1994). Differentiation of endothelial cells: analysis of the constitutive and activated endothelial cell phenotypes. *BioEssays*. 16: 901-906.
- Awika, J. M., Rooney, L. W., & Waniska, R. D. (2005). Anthocyanins from black sorghum and their antioxidant properties. *Food Chemistry*. 90: 293–301.

- Baderschneider, B., & Winterhalter, P. (2000). Isolation and characterization of novel stilbene derivatives from Riesling wine. *Journal of Agriculture and Food Chemistry*. 48: 2681–2686.
- Balas, A., and Popa, V. I. (2007). Bioactive aromatic compounds – The influences of natural aromatic compounds on the development of *Lycopersicon esculentum* plantlets. *BioResources*. 2(3): 363–370.
- Balasundram, N., Sundram, K., and Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. *Food Chemistry*. 99: 191–203.
- Bate-Smith, E. C. (1973). Haemanalysis of tannins: the concept of relative astringency. *Phytochemistry*. 12: 907 - 910.
- Bleve, M., Ciurlia, L., Erroi, E., Lionetto, G., Longoc, L., Rescioa, L. (2008). An innovative method for the purification of anthocyanins from grape skin extracts by using liquid and sub-critical carbon dioxide. *Separation and Purification Technology*. 64: 192–197.
- Bombardelli, E., Morazzonni, P. (1995). *Vitis vinifera* L. *Fitoterapia*. 66: 291-317.
- Borzelleca, Joseph, F.; Lane, Richard, W., (2008). The Art, the Science, and the Seduction of Toxicology: an Evolutionary Development. Eds. Hayes,
- Bourhis, M., Theodore, N., Weber, J. F., Vercauteren, J. (1996). Isolation and identification of (Z)- and (E)- $\epsilon$ -viniferins from stalks of *Vitis vinifera*. XVIII Journee Internationale Groupe Polyphenols, Bordeaux. 1: 43-44.
- Bucic'-Kojic' , A., Planinic' , M., Tomas, S., Bilic' , M., and Velic, D. (2007). Study of solid– liquid extraction kinetics of total polyphenols from grape seeds. *Journal of Food Engineering*. 81: 236–242.

- Burda, S. and Oleszek, W. (2001). Antioxidant and antiradical activities of flavonoids. *Journal of Agricultural and Food Chemistry*. 49: 2774 - 2779.
- Burns, J., Yokota, T., Ashihara, H., Lean, M. E., Crozier, A. (2002). Plant foods and herbal sources of resveratrol. *Journal of Agriculture and Food Chemistry*. 50(11): 3337-40.
- Cadenas, S. and Barja, G. (1999). Resveratrol, melatonin, vitamin E, and PBN protect against renal oxidative DNA damage induced by the kidney carcinogen KBrO<sub>3</sub>. *Free Radical Biological Medicine*. 26: 1531-1537.
- Canter, P.H., Thomas, H., Ernst, E. (2005). Bringing medicinal plants into cultivation: opportunities and challenges for biotechnology. *Trends Biotechnol.* 23: 180-185.
- Caridi, D., Trenerry, V. C., Rochfort, S., Duong, S., Laughler, D., and Jones, R. (2007). Profiling and quantifying quercetin glucosides in onion (*Allium cepa* L.) varieties using capillary zone electrophoresis and high performance liquid chromatography. *Food Chemistry*. 105: 691–699.
- Castañeda-Ovando, A., de Lourdes Pacheco-Hernández, Ma, Elena, Ma, Páez-Hernández, J. A., RodríguezGalán-Vidal, J. A., & Galán-Vidal, C. A. (2009). Chemical studies of anthocyanins: A review. *Food Chemistry*. 113: 859–871.
- Chen, R. S., Wu, P. L., Chiou, Y.Y. (2002). Peanut Roots as a Source of Resveratrol. *Journal of Agriculture and Food Chemistry*. 50(6): 1665-1667
- Cho, M., Howard, L., Prior, R., Clark, J. (2004). ‘Flavonoid glycosides and antioxidant capacity of various blackberry, blueberry and grape genotypes determined by high-performance liquid chromatography/mass spectrometry. *Journal of Science and Food Agriculture*. 84: 1771-1782.
- Clifford, M. N. (1999). Chlorogenic acids and other cinnamates: nature, occurrence and dietary burden. *Science of Food and Agriculture*. 79: 362 - 372.

- Coleman, J. W. (2001) Nitric oxide in immunity and inflammation. *Int. immunopharmacology*. 1: 1397-1406.
- Cooks R. G., Caprioli R. M. (2000). Special feature on electrospray ionization. *J Mass Spectrom*. 35:761.
- Corrales, M., Fernández García, A., Butz, P., and Tauscher, B. (2009). Extraction of anthocyanins from grape skins assisted by high hydrostatic pressure. *Journal of Food Engineering*. 90: 415–421.
- Cragg, G. M. and Newman, D. J. (2002). Chemical diversity: a function of biodiversity. *Trends Pharmacology Science*. 23: 404–405.
- Cragg, G. M., Newmann, D. J., and Snader, K. M. (1997). Natural products in drug discovery and development. *Natural Product*. 60: 52–60.
- Creasy, L. L., Coffee M. (1988). Phytoalexin production potential of grape berries. *American Social and Horticulture Sciences*. 113: 230-234.
- Crespan M. (2004). Evidence on the evolution of polymorphism of microsatellite markers in varieties of *Vitis vinifera* L. *Theoretical and Applied Genetics*. 108: 231–237.
- Cuvelier, M.-E., Richard, H. and Berst, C. (1992). Comparison of the antioxidative activity of some phenolic acids: structure-activity relationship. *Bioscience Biotechnology and Biochemistry*. 56: 324 - 325.
- Darley-USmar, V. and Halliwell, B. (1996). Blood radicals. Reactive nitrogen species, reactive oxygen species, transition metal ions, and the vascular system. *Pharmaceutical Research*. 13: 649 - 662.



- Das, S., Alagappan, V. K., Bagchi, D., Sharma, H. S., Maulik, N. and Das, D. K. (2005). Coordinated induction of iNOS-VEGF-KDR-eNOS after resveratrol consumption: a potential mechanism for resveratrol preconditioning of the heart. *Vascular Pharmacology*. 42: 281-289.
- Dasgupta, N. and De, B. (2004). Antioxidant activity of *Piper betle* L. leaf extract in vitro. *Food Chem*. 88: 219-224.
- De Caterina, R., Libby, P., Peng, H. B., Thannikal, V. J., Rajavashisth, T. B., Gimbrone, M. A., Jr., Shin, W. S., Liao, J. K. (1995). Nitric oxide decreases cytokine-induced endothelial activation. *Journal of Clinical Investigation*. 96: 60-8.
- De Heer, M. I., Korth, H. G. and Mulder, P. (1999). Polymethoxy phenols in solution: O-H bond dissociation enthalpies, structures, and hydrogen bonding. *Journal of Organic Chemistry*. 64: 6969 - 6975.
- Decendit, A., Waffo-Teguo, P., Richard, T., Krisa, S., Vercauteren, J., Monti, J.P., *et al.* (2002). Galloylated catechins and stilbene diglucosides in *Vitis vinifera* cell suspension cultures. *Journal of Phytochemistry*. 60: 795–798.
- Delcour, J. A., Vandenberghe, M. M., Corten, P. F. and Dondeyne, P. C. (1984). Flavor thresholds of polyphenolics in water. *American Journal of Enology and Viticulture*. 35: 134 - 139.
- Diouf, P. N., Stevanovic, T., and Cloutier, A. (2009). Study on chemical composition, antioxidant and anti-inflammatory activities of hot water extract from *Picea mariana* bark and its proanthocyanidin-rich fractions. *Food Chemistry*. 113: 897–902.
- DiSilvestro, R. A. (2001). Flavonoids as Antioxidants. Eds. Wildman, R. E. C. *Handbook of Nutraceuticals and Functional Foods*. CRC Press, New York, NY. 127 - 142.

- Dixon, R. A. (2001). Natural products and plant disease resistance. *Nature* . 411:843–847.
- Douillet-Breuil, A. C.; Jeandet, P.; Adrian, M.; Bessis, R.; 1999: Changes in the phytoalexin content of various *Vitis* spp. in response to UV-C elicitation. *Journal of Agriculture and Food Chemistry*. 47: 4456-4461.
- Downey, M. O., Dokoozlian, N. K., Krstic, M.P. (2006). Cultural practice and environmental impacts on the flavonoid composition of grapes and wine: a review of recent research. *American Journal of Enology and Viticulture*. 57 :257–268.
- Downey, M.O., Mazza, M., Krstic, M.P. (2007). Development of a stable extract for anthocyanins and flavonols from grape skin. *American Journal of Enology and Viticulture*. 58: 358–364.
- Duke, J. (1981). *Phytochemical Database*, USDA-ARS-NGRL. Eds. Tiwari, B., Gowen, A., and McKenna B. *Pulse Foods: Processing, Quality and Nutraceutical Applications*. Elsevier, Amsterdam. 437
- Duke, J. A. (1983). *Handbook of Energy Crops*. Unpublished.
- Escarpa, A., Morales, M. D., and Gonzalez, M. C. (2002). Analytical performance of commercially available and unavailable phenolic compounds using real samples by high-performance liquid chromatography-diode array detection. *Analytica Chimica Acta*. 460: 61–72.
- Farnsworth, N. R. (1990). The role of ethnopharmacology in drug development in Bioactive Compounds from Plants. Eds. Chadwick, D. J. and Marsh, J., John Wiley and Sons, New York. 2-21.
- Farnsworth, N. R., Akerele, O., Bingel, A. S., Soejarto, D. D. and Guo, Z. (1985). Medicinal plants in therapy. *Bull. WHO*. 63: 965-981

- Folin, O., Ciocalteu, V. (1927): Tyrosine and tryptophan determination on proteins. *J. Biol. Chem.* 73: 627-649.
- Frankel, E. N., Waterhouse, A. L. and Teissedre, P. L. (1993). Inhibition of oxidation of human low-density protein by phenolic substances in red wine. *Lancet.* 341: 1103 - 1104.
- Fred-Jayesimi, Adediwura, A., Oredipe, A. (2013). Anti-depressant activities of the methanol extracts, petroleum ether, and ethyl acetate fractions of *Morus mesozygia* stem bark. *Journal of Pharmacology and Pharmacy.* 4: 100-103
- Friedman, M., and Jürgens, H. (2000). Effect of pH on the Stability of Plant Phenolic Compounds. *J. Agric. Food Chem.* 4(6): 2101–2110.
- Fu, J. Y., Masferrer, J. L., Seibert, K., Raz, A., Needleman, P. (1990). The induction and suppression of prostaglandin H<sub>2</sub> synthase (cyclooxygenase) in human monocytes. *J Biol Chem.* 1990;265:16737–40.
- Fuhrman, B., Volcova, N., Rosenblat, M. and Aviram, M. (2000). Lycopene synergistically inhibits LDL oxidation in combination with vitamin E, glabridin, rosmarinic acid, carnosic acid, or garlic. *Antioxidants and Redox Signalling.* 2: 491-505.
- Fukumoto, L. R. and Mazza, G. (2000). Assessing antioxidant and prooxidant activities of phenolic compounds. *Journal of Agricultural and Food Chemistry.* 48: 3597 - 3604.
- Furst, D. E. (1999). Pharmacology and efficacy of cyclooxygenase (COX) inhibitors. *Am J Med.* 107S:18–26S.
- Gao, H., Shupe, T. F., Eberhardt, T., Hse, C.Y. (2006). Antioxidant activity of extracts from the wood and bark of Port-Orford cedar. *J. Wood Sci.*

- Gimbrone, M. A. J., Cotran, R. S., Folkman, J. (1974). Human Vascular Endothelial Cells in Culture, Growth, and DNA Synthesis. *Journal of Cell Biology*. 60(3): 673-684.
- Golce, N., Keaney, J. J., Vita, J. A. (1998). Endotheliopathies: clinical manifestations of endothelial dysfunction. Eds: Loscalzo, J., Shafer, A. In: *Thrombosis and hemorrhage*. 2<sup>nd</sup> Edition. Baltimore: Williams and Wilkins.
- Gomez-Caravaca, A. M., Segura-Carretero, M. G' A., Gomez-Romero, A. D. (2006). Advances in the analysis of phenolic compounds in products derived from bees. *Journal of Pharmaceutical and Biomedical Analysis*. 41: 1220–1234.
- Gordon, M. H. (1990). The Mechanism of Antioxidant Action In Vitro. Eds. Hudson, B. J. F. *Food Antioxidants*. Elsevier Applied Science, London. 1 – 18.
- Graversen, H.B., Becker, E.M., Skibsted, L.H. and Andersen, M.L. 2008. Antioxidant synergism between fruit juice and alpha-tocopherol. A comparison between high phenolic black chokeberry (*Aronia melanocarpa*) and high ascorbic blackcurrant (*Ribes nigrum*). *European Food Research and Technology*. 226 (4): 737-743.
- Guyot, S., Marnet, N., Laraba, D., Sanoner, P., Drilleau, J.F. (1998). Reversed-phase HPLC following thiolysis for quantitative estimation and characterization of the four main classes of phenolic compounds in different tissue zones of a French cider apple variety (*Malus domestica* Var. Kermerrien). *Journal of Agricultural and Food Chemistry*. 46: 1698–1705.
- Hafizah, A. H., Zaiton, Z., Zulkhairi, A., Ilham, M. A., Nordin, M. M., Anita, A., Zaleha, A. M. (2010). *Piper sarmentosum* as an antioxidant on oxidative stress in human umbilical vein endothelial cells induced by hydrogen peroxide. *Biomed & Biotechnol*. 11(5): 357-365

- Halliwell, B., Murcia, M. A., Chirico, S. and Aruoma, O. I. (1995). Free radicals and antioxidants in food and in vivo: what they do and how they work. *Critical Reviews in Food Science and Nutrition*. 35: 7 - 20.
- Harborne, J. B. (1967). *Comparative Biochemistry of the Flavonoids*. Academic Press. New York, NY.
- Harborne, J. B. (1984). *Phytochemical methods: a guide to modern techniques of plant analysis*. Chapman and Hall. 1-288.
- Harborne, J. B. (1994). *Flavonoids*. Chapman and Hall, London and New York.
- Harborne, J. B. and Williams, C. A. (2000). Advances in flavonoid research since 1992. *Phytochemistry*. 55: 481 - 504.
- Hart, J. H. (1981). Role of phytoestrogens in decay and disease resistance. *Annual Review Phytopathology*. 19: 437-458.
- Hattori, R., Otani, H., Maulik, N. and Das, D. K. (2002). Pharmacological preconditioning with resveratrol: role of nitric oxide. *American Journal in Physiological Heart Circulation Physiology*. 282: H1988-1995.
- Health. CRC Press, Boca Raton, FL. 3-20.
- Heitzer, T., Schlinzig, T., Krohn, K. (2001). Endothelial dysfunction, oxidative disease. *Circulation*. 104: 2673-2678.
- Hensley, K., Mou, S., Pye, Q. N., Dixon, R. A., Summner, L. W. and Floyd, R. A. (2004). Chemical versus pharmacological actions of nutraceutical phytochemicals: Antioxidant and anti-inflammatory modalities. *Current Topics in Nutraceutical Research*. 2: 13 - 26.
- Hermann, K. M., Weever, L. M. (1999) The Shikimate Pathway. *Annual Review Plant Physiology Plant Molecular Biology*. 50: 473-503.

- Herrmann K. (1989). Occurrence and content of hydroxycinnamic and hydroxybenzoic acid compounds in food. *Crit Rev Food Sci Nutr.* 28: 315–347.
- Hobbs, A. J. (1997). Soluble guanylate cyclase: the forgotten sibling. *Trends pharmacol sci.* 18:484-491.
- Hooper, L., and Cassidy, A. (2006). A review of the health care potential of bioactive compounds. *Journal of the Science of Food and Agriculture*, 86, 1805–1813.
- Howitz K. T., Bitterman K. J., Cohen H.Y., Lamming D. W., Lavu S., and Wood J. G. (2003). Small molecule activators of sirtuins extended *Saccharomyces cerevisiae* lifespan. *Nature.* 425:191–6.
- Hu, C. and Kitts, D. D. (2001). Evaluation of antioxidant activity of epigallocatechin gallate in biphasic model systems in vitro. *Molecular and Cellular Biochemistry.* 218: 147 - 155.
- Huang, C. A. and Zayas, J. F. (1991). Phenolic acid contribution to taste characteristics of corn germ protein flour products. *Journal of Food Science.* 56: 1308 - 1312.
- Huang, M. T., Ho, C. T. and Lee, C. Y. (1992). Phenolic Compounds in Food and Their Effects on Health II. ACS Symposium Series 507. American Chemical Society, Washington, DC.
- Hughes, M. N. (1999). Relationships between nitric oxide, nitroxyl ion, nitrosonium cation and peroxyxynitrite. *Biochim biophys acta.* 1411: 263-272.
- Ignat, I., Volf, I., Popa V. (2011). A critical review of methods for characterisation of polyphenolic compounds in fruits and vegetables. *Food Chemistry.* 126: 1821-1835.

- Ihsan, S., Kamarazaman, Z. A., Rasadah, M. A., Abdah, M. A., Khairunnuur, F. A., Daryl, J. A., Kamal, N. H., Shahidan, M. A., Zamree, M. S., and Khairul, K. A. K. (2012). Protective effects of *Tinospora crispa* extracts on H<sub>2</sub>O<sub>2</sub>-induced oxidative stress and TNF- $\alpha$ -induced inflammation on human umbilical vein endothelial cells (HUVECs). *Journal of Medicinal Plants Research*. 6(15): 3013-3021.
- Imamura, G., Bertelli, A. A., Bertelli, A., Otani, H., Maulik, N. and Das, D. K. (2002). Pharmacological preconditioning with resveratrol: an insight with iNOS knockout mice. *American Journal in Physiological Heart Circulation Physiology*. 282: H1996-2003.
- Jaffe, E. A., Nachman, R. L., Becker, C. G., Minick, C. R. (1973). Culture of Human Endothelial Cells Derived from Umbilical Veins. Identification By Morphologic And Immunologic Criteria. *The Journal of Clinical Investigation*. 52: 2745 - 2756
- Jang, M., Cai, L., Udeani, G. O., Slowing, K. V., Thomas, C. F., Beecher, C.W., Fong, H. H., Farnsworth, N. R., Kinghorn, A. D., Mehta, R. G., Moon, R. C., and Pezzuto, J. M. (1997). Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science*. 275: 218-220.
- Jang, M.; Cai, L.; Udeani, G. O.; Slowing, K. U.; Thomas, C. F.; Beecher, C. W.; Fong, H. H. S.; Farnsworth, N. R.; Kinghorn, A. D.; Mehta, R. G.; Moon, R. C.; Pezzuto, J. M. (1997). Cancer chemoprotective activity of resveratrol, a natural product derived from grapes. *Journal of Science*. 275: 218-220.
- Jeandet, P., Bessis, R., Maume, B. F., Meunier, P., Peyron, D., Trollat, P. (1995). Effect of enological practices on the resveratrol isomers content of wine. *Agriculture and Food Chemistry*. 43: 316- 319.

- Jeandet, P., Breuil, A. C., Adrian, M., Weston, L. A., Debord, S., Meunier, P. (1997). HPLC analysis of grapevine phytoalexins coupling photodiode array detection and fluorometry. *Analytical Chemistry*. 69(24): 5172–5177.
- Jeandet, P., Douillt-Breuil, A.C., Bessis, R., Debord, S., Sbaghi, M., Adrian, M. (2002). Phytoalexins from the Vitaceae: Biosynthesis, phytoalexin gene expression in transgenic plants, antifungal activity, and metabolism. *Journal of Agriculture and Food Chemistry*. 50: 2731–2741.
- Jean-Denis, J. B.; Pezet, R.; Tabacchi, R. (2006). Rapid analysis of stilbenes and derivatives from downy mildew-infected grapevine leaves by liquid chromatography-atmospheric pressure photoionisation mass spectrometry. *Journal of Chromatography*. 1112: 263-268.
- Jianping, S., Liang, F., Bin, Y., Li, P., Duan, C. (2007). Screening Non-colored Phenolics in Red Wines using Liquid Chromatography/Ultraviolet and Mass Spectrometry/Mass Spectrometry Libraries. *Molecules*. 12: 679-693.
- Jouzeau JY, Terlain B, Abid A, Nedelec E, Netter P. Cyclo-oxygenase isoenzymes. How recent findings affect thinking about nonsteroidal anti-inflammatory drugs. *Drugs*. 53: 563–82.
- Kamal-Eldin, A. and Appelqvist, L. A. (1995). The effects of extraction methods on sesame oil stability. *J. Am. Oil. Chem. Soc.* 72(8): 967–969.
- Kapasakalidis, P. G., Rastall, R. A., and Gordon, M. H. (2006). Extraction of polyphenols from processed black currant (*Ribes nigrum* L.) residues. *Journal of Agricultural and Food Chemistry*. 54: 4016–4021.
- Knekt, P., Kumpulainen, J., Jarvinen, R., Rissanen, H., Heliovaara, M., Reunanen, A., Hakulinen, T. and Aromaa, A. (2002). Flavonoid intake and risk of chronic diseases. *The American Journal of Clinical Nutrition*. 76: 560 - 568.



- Knowles, R. G. and Moncada, S. (1994) Nitric oxide synthases in mammals. 298:249-258.
- Kopp, P. (1998). Resveratrol, a phytoestrogen found in red wine. A possible explanation for the conundrum of the 'French paradox'? *Eur J Endocrinol.* 138: 619-620.
- Koskimaki, J. J., Hokkanen, J., Jaakola, L., Suorsa, M., Tolonen, A., Sampo, M., Pirttila, A.M., Hohtola, A. (2009). Flavonoid biosynthesis and degradation play a role in early defence responses of bilberry (*Vaccinium myrtillus*) against biotic stress. *European Journal of Plant Pathol.* 125: 629–640.
- Kothari, V., Seshadri, S. (2010). Antioxidant activity of seed extracts of *Annona squamosa* and *Carica papaya*. *Nutrition and Food Science.* 40(4):403-408.
- Kubes, P. and McCafferty, D. M. (2000) Nitric oxide and intestinal inflammation. *America j med.* 109: 150-158.
- Kuh, D., Ben-Shlomo, Y. (1997). A life course approach to chronic disease epidemiology. Oxford University Press, NY.
- Langcake, P., Mccarthy, W.V. (1979). The relationship of resveratrol production to infection of grapevine leaves by *Botrytis cinerea*. *Vitis.* 18: 244-253.
- Langcake, P., Pryce, R. J. (1976). The production of resveratrol by *Vitis vinifera* and other members of the Vitaceae as a response to infection or injury. *Physiology and Plant Pathology.* 9: 77-86.
- Lapornik, B., Prosek, M., & Golc, Wondra. A. (2005). Comparison of extracts prepared from plant by-products. *Food Engineering.* 71: 214–222.
- Lardos, A., Kreuter, M. H. (2000). Red vine leaf. Eds. Kreuter, M. H. *Phytopharmacology and Phytochemistry Products.* Flachsmann AG. Zurich. 1-7.

- Le-Marchand, L., Murphy, S. P., Hankin, J. H., Wilkens, L. R. and Kolonel, L. N. (2000). Intake of flavonoids and lung cancer. *Journal of the National Cancer Institute*. 92: 154 - 160.
- Levadoux, L. D. (1956). Wild and cultivated populations of *Vitis vinifera* L. *Annales de l'Amelioration des Plantes*. 6: 59–118.
- Lewis, L. J., Hoak, J. C., Maca, R.D., Fry, G. L. (1973). Replication of Human Endothelial Cells in Culture. *Journal of Science*. 181: 453-454
- Loredana, L.; Torre, G. L. L.; Saitta, M.; Vilasi, F.; Pellicanó, T.; Dugo G. (2006). Direct determination of phenolic compounds in Sicilian wines by liquid chromatography with PAD and MS detection. *Food Chem*. 94: 640-650.
- Lorenz, P., Roychowdhury, S., Engelmann, M., Wolf, G., Horn, T.F.W. (2003). Oxyresveratrol and resveratrol are potent antioxidants and free radical scavengers: Effect on nitrosative and oxidative stress derived from microglial cells. *Journal of Nitric Oxide*. 9:64–76.
- Loub, W. D., Farnsworth, N. R., Soejarto, D. D., and Quinn, M. L. (1985). NAPRALERT: computer handling of natural product research data. *Chem. Inf. Comput. Sci*. 25: 99–103.
- Lucas, K. A., Pitari, G. M., Kazerounian, S., Ruiz-Stewart, I., Park, J., Schulz, S., Chepenik, K. P. and Waldman, S. A. (2000). Guanylyl cyclases and signaling by cyclic GMP. *Pharmacol rev*. 52: 375-414.
- Luthria, D. L., and Pastor-Corrales, M. A. (2006). Phenolic acids content of 15 dry edible bean (*Phaseolus vulgaris* L.) varieties. *Journal of Food Composition and Analysis*. 19: 205–211.
- Macheix, J. J., Fleuriet, A. Billot, J. (1990). *Fruit Phenolics*. CRC Press, Boca Raton, FL.

- Macheix, J., Fleuriet, A. and Billot, J. (1990). Fruits phenolics. *CRC Press*. 24–31, 295–342.
- Manach, C., Scalbert, A., Morand, C., Jimenez, L. (2004). Polyphenols: Food sources and bioavailability. *American Journal of Clinical Nutrition*. 79: 727–747.
- Marletta, M. A., Hurshman, A. R. and Rusche K. M. (1998). Catalysis by nitric oxide synthase. 2 :656-663.
- Martel-Pelletier, J., Lajeunesse, D., Reboul, P. (2002). Therapeutic role of dual inhibitors of 5-LOX and COX, selective and non-selective non-steroidal anti-inflammatory drugs. *Ann rheum*. 62: 501–509.
- Martin, A. R., Villegas, I., La Casa, C. and de la Lastra, C. A. (2004). Resveratrol, a polyphenol found in grapes, suppresses oxidative damage and stimulates apoptosis during early colonic inflammation in rats. *Biochemical Pharmacology*. 67: 1399 - 1410.
- Marwan, A. G. and Nagel, C. W. (1982). Separation and purification of hydroxycinnamic acid derivatives in cranberries. *Journal of Food Science*. 47: 585 - 590.
- Mattila, P., and Kumpulainen, J. J. (2002). Determination of free and total phenolic acids in plant derived foods by HPLC and diode array detection. *Journal of Agricultural and Food Chemistry*. 48: 3660–3667.
- Mauro, M. C., Toutaina, S., Pinck, W. L., Ottenc, L., Coutos-Theve- notd, P., Deloiree, A., Barbierd, P. (1999). High efficiency regeneration of grapevine plants transformed with the GFLV coat protein gene. *Plant Science*. 112: 97–106.

- McDaniel, R., Ebert-Khosla, S., Hopwood, D. A. and Khosla, C. (1995). Rational design of aromatic polyketide natural products by recombinant assembly of enzymatic subunits. *Nature*. 375: 549 - 554.
- McKenna, D. J., Jones, K., Hughesm K. (1998). Efficacy, safety, and use of ginkgo biloba in clinical and preclinical applications. *Altern Ther Health Med*. 5:70-86, 88-90.
- Melzoch K., Hanzlíková I., Filip V., Buckiová D., Šmidrkal J. (2001). Resveratrol in parts of vine and wine originating from Bohemian and Moravian vineyard regions.
- Merken, H. M., and Beecher, G. R. (2000). Measurement of food flavonoids by high-performance liquid chromatography: A review. *Journal of Agricultural and Food Chemistry*. 48: 577–599.
- Monagas, M.; Suárze, R.; Gómez-Cordovés, C.; Bartolomé, B. (2005). Simultaneous determination of nonanthocyanins in phenolic compounds in wines by HPLC-DAD/ESI-MS. *Am. J. Enol. Vitic*. 56 (2): 139-147.
- Moncada S, Higgs A. (1993). The L-arginine-nitric oxide pathway. *N Engl J Med*. 329: 2002–2012.
- Mori, A., Nishino, C., Enoki, N. and Tawata, S. (1987). Antibacterial activity and mode of action of plant flavonoids against *Proteus vulgaris* and *Staphylococcus aureus*. *Phytochemistry*. 26(8): 2231–2234
- Mori, K., H., Saito, N., Goto-Yamamoto, M., Kitayama, S., Kobayashi, S., Sugaya, H., Gemma, and Hashizume, K. (2005). Effects of abscisic acid treatment and night temperatures on anthocyanin composition in Pinot noir grapes. *Vitis*. 44: 161-165.

- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assay. *J. Immunol. Methods*. 65 (1-2): 55-63.
- Murias, M., Handler, N., Erker, T., Pleban, K., Ecker, G., Saiko, P. (2004). Resveratrol analogue as selective cyclooxygenase-2 inhibitors: synthesis and structure-activity relationship. *Bioorganic Medicine Chemistry*. 12(21): 5571-5578.
- Nathan, C. (1997). Inducible nitric oxide synthase: what difference does it make? *100*:2417-2423.
- Nature Review Drug Discovery. (2006)
- Nichenametla, S. N., Taruscio, T. G., Barney, D. L. and Exon, J. H. (2006). A review of the effects and mechanisms of polyphenolics in cancer. *Critical Reviews in Food Science and Nutrition*. 46: 161 - 183.
- Noguchi, N. and Niki, E. (1998). Chemistry of Active Oxygen Species and Antioxidants. Eds: Papas, A. M. Antioxidant Status, Diet, Nutrition, and
- Norbaek R., Kondo T. (1999) Further anthocyanins from flowers of *Crocus antalyensis* (Iridaceae). *Journal of Phytochemistry*. 50: 325–328.
- Parr. A. J., Bolwell, G. P. (2000). Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying the phenol content or profile. *Agriculture and Food Chemistry*. 80: 985-1012.
- Pearson T. A., Mensah G. A., Alexander R. W. (2003). Markers of inflammation and cardiovascular disease: application to clinical and public health practice. A statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. *Circulation*.107: 499–511.

- Pezet, R., Gindro, K., Viret, O., and Spring, J. L. (2004). Glycosylation and oxidative dimerization of resveratrol are respectively associated to sensitivity and resistance of grapevine cultivars to downy mildew. *Physiological and Molecular Plant Pathology*. 65: 297–303.
- Pezet, R., Perret, C., Bernard, J., Jean-Denis, J. B., Tabacchi, R, Gindro, K. (2003).  $\epsilon$ -viniferin, a resveratrol dehydrodimer: One of the major stilbenes synthesized by stressed grapevine leaves. *Journal of Agriculture and Food Chemistry*. 51: 5488–5492.
- Pezet, R., Pont, V. (1988). Demonstration of pterostilbene in clusters of *Vitis vinifera*. *Plant Physiology and Biochemistry*. 26: 603–7.
- Pietta, P., Simonetti, P., and Mauri, P. (1998). Antioxidant activity of selected medicinal plants. *Agricultural and Food Chemistry*. 46: 4487–4490.
- Pietta, P.-G. (2000). Flavonoids as antioxidants. *Journal of Natural Products*. 63: 1035 - 1042.
- Pinelo, M., Del Fabbro, P., Manzocco, L., Nunez, M. J., & Nicoli, M. C. (2005). Optimization of continuous phenol extraction from *Vitis vinifera* byproducts. *Food Chemistry*. 92: 109–117.
- Popkin, B. M., Keyou, G., Fengying, Z., Guo, X., Haijiang, M., Zohoori, N. (1993). The nutrition transition in China: a cross-sectional analysis. *European Journal of Clinical Nutrition*. 47: 333-346.
- Proctor, A. and Bowen, D. J. (1996). Ambient-temperature extraction of rice bran oil with hexane and isopro- panol, *J. Am. Oil. Chem. Soc.* 73(6): 811–813.
- Püssa, T., Floren, J., Kuldkepp, P., Raal, A. (2006). Survey of grapevine *Vitis vinifera* stem polyphenols by liquid chromatography-diode array detection-tandem mass spectrometry. *Journal of Agricultural and Food Chemistry*. 54:7488-7494.

- Ramawat, K. G., Dass, S., Mathur, M. (2009). The chemical diversity of bioactive molecules and therapeutic potential of medicinal plants. Eds. Ramawat, K. G. *Herbal drugs: ethnobotanical to modern perspective*. Springer, Heidelberg.
- Renaud, S., de Lorgeril, M. (1992). Wine, alcohol, platelets, and the French paradox for coronary heart disease. *Lancet*. 339:1523–1526.
- Richard, J. L., Cambien, F., Ducimetie`re, P. (1981). Epidemiologic characteristics of coronary disease in France. *Nouv Presse Med*. 10:1111–1114.
- Robbins, R. J. (2003). Phenolic acids in foods: An overview of analytical methodology. *Agricultural and Food Chemistry*. 51: 2866 - 2887.
- Roberts, P. J., Morgan, K., Miller, R., Hunter, J. O and Middleton, S. J. (2001). Neuronal COX-2 expression in human myenteric plexus in active inflammatory bowel disease. *Gut*. 48:468-472.
- Roberts, P. J., Riley, G. P., Morgan, K., Miller, R., Hunter, J. O. and Middleton S. J. (2001). The physiological expression of inducible nitric oxide synthase (iNOS) in the human colon. *J Clin Pathol*. 297: 54-293.
- Roggero, J. P. (1997). Wine phenolics analysis via direct HPLC injection. *American Laboratory News*. January: 12D-12G.
- Romero-Pérez, A. I., Lamuela-Raventós, R. M., Andrés- Lacueva, C., de la Torre-Boronat, M. C. (2001). Method for the quantitative extraction of resveratrol and piceid isomers in grape berry skins. Effect of powdery mildew on the stilbene content. *Journal of Agricultural*.

- Ross, K. A., Beta, T., and Arntfield, S. D. (2009). A comparative study on the phenolic acids identified and quantified in dry beans using HPLC as affected by different extraction and hydrolysis methods. *Food Chemistry*. 113: 336–344.
- Ross, R. (1999) Atherosclerosis—an inflammatory disease. *N Engl J Med* 340:115–126.
- Rossetto, M., McNally, J., Henry, R. J. (2002). Evaluating the potential of SSR flanking regions for examining relationships in Vitaceae. *Theoretical and Applied Genetics*. 104: 61–66.
- Russell, W. R., Scobbie L., Labat, A., Duncan, G. J. Duthie, G. G., (2008). Phenolic acid content of fruits commonly consumed and locally produced in Scotland. *Food Chemistry*.
- Sakihama, Y., Cohen, M. F., Grace, S. C. and Yamasaka, H. (2002). Plant phenolic antioxidant and prooxidant activities: Phenolics-induced oxidative damage mediated by metals in plants. *Toxicology*. 177: 67 - 80.
- Salie, F., Eagles, P. F. K., Leng, H. J. M. (1996). Preliminary antimicrobial screening of four South African Asteraceae species. *Journal of Ethnopharmacology*. 52: 27–33.
- Salvemini, D., Misko, T. P., Masferrer, J. L., Seibert, K., Currie, M. G. and Needleman, P. (1993) Nitric oxide activates cyclooxygenase enzymes. *Proc natl acad sci USA*. 90: 7240-7244.
- Sana, T. R., Waddell, K., Fischer, S. M. (2008). A sample extraction and chromatographic strategy for increasing LC/MS detection coverage of the erythrocyte metabolome. *Chromatography B*. 871(2): 314-321.



- Sanoner, P., Guyot, S., Marnet, N., Molle, D., Drilleau, J. F. (1999). Polyphenol profiles of French cider apple varieties (*Malus domestica* sp.). *Journal of Agricultural and Food Chemistry*. 47: 4847–4853.
- Santamaria, A. R., Antonacci, D., Caruso, G., Cavaliere, C., Gubbiotti, R., Lagan, A., Valletta, A., Pasqua G. (2010). Stilbene production in cell cultures of *Vitis vinifera* L. cvs Red Globe and Michele Palieri elicited by methyl jasmonate, *Natural Product Research: Formerly Natural Product Letters*. 24(15): 1488-1498.
- Santos-Buelga, C., Scalbert, A. (2000). Proanthocyanidins and tannin-like compounds: nature, occurrence, dietary intake and effects on nutrition and health. *Journal of Food Science and Agriculture* (in press).
- Sarker, S. D., Latif, Z. and Gray, A. I. (2006). *Natural Product: History Perspective in Natural Product Isolation*. Humana Press, New Jersey. 2: 4-5.
- Scalbert, A. and Williamson, G. (2000). Dietary intake and bioavailability of polyphenols. *Nutrition*. 130: 2073S - 2085S.
- Schachinger V., Britten, M. B., Zeiher A. M. (2000). Prognostic impact of coronary disease. *Circulation*. 101: 1899 -1906.
- Schuler, P. (1990). Natural Antioxidants Exploited Commerically. In: *Food Antioxidants*. Eds. Hudson, B. J. F. Elsevier Applied Science, New York, N. Y.
- Sefc, K. M., Steinkellner, H., Lefort, F., (2003). Evaluation of the genetic contribution of local wild vines to European grapevine cultivars. *American Journal of Enology and Viticulture*. 54: 15–21.
- Shahidi, F. (2000). Antioxidants in food and food antioxidants. *Nahrung*. 44: 158 - 163.

- Shahidi, F. and Naczk, M. (1995). Food Phenolics: Sources, Chemistry, Effects, Applications. *Technomic Publishing Company*. Lancaster, P. A.
- Shahidi, F. and Naczk, M. (2004). Phenolics in Food and Nutraceuticals. *CRC Press*. Boca Raton, FL.
- Sharififar, F., Dehghn-Nudeh, G., and Mirtajaldini, M. (2009). Major flavonoids with antioxidant activity from *Teucrium polium* L. *Food Chemistry*. 112: 885–888.
- Shimokawa H. (1999). Primary endothelial dysfunction: atherosclerosis. *J Mol Cell Cardiol*. 31: 23–37.
- Signorelli, P., and Ghidoni, R. (2005). Resveratrol as an anticancer nutrient: Molecular basis, open questions and promises. *Journal of Nutritional Biochemistry*. 16: 449–466.
- Singleton, V. L. and Rossi, J. A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*. 16: 144 - 158.
- Slinkard, K. and Singleton, V. L. (1977). Total phenol analysis: Automation and comparison with manual methods. *American Journal of Enology and Viticulture*. 28: 49 - 55.
- Soares, D. G., Andreazza, A. C. and Salvador, M. (2003). Sequestering ability of butylated hydroxytoluene, propyl gallate, resveratrol, and vitamins C and E against ABTS, DPPH, and hydroxyl free radicals in chemical and biological systems. *Journal of Agricultural and Food Chemistry*. 51: 1077 - 1080.
- Soares, J.R., Dins, T.C.P., Cunha, A.P., Ameida, L.M. (1997). Antioxidant activity of some extracts of *Thymus zygis*. *Free Radical Res*. 26: 469–478.

- Sobolev, V. S.; Cole, R. J. (1999). *trans*-Resveratrol content in commercial peanuts and peanut products. *Journal of Agriculture and Food Chemistry* 47: 1435-1439.
- Sánchez-Rabaneda, F.; Jáuregui, O.; Casals, I.; Andrés-Lacueva, C.; Izquierdo-Pulido, M. (2003). Liquid chromatographic/electrospray ionization tandem mass spectrometric study of the phenolic composition of cocoa (*Theobroma cacao*). *J. Mass Spectrom.* 38: 35-42.
- Sotheeswaran, S., Pasupathy, V. (1993). Distribution of resveratrol oligomers in plants. *Phytochemistry*. 32 (5): 1083-1092.
- Stalikas, C.D. (2007). Extraction, separation, and detection methods for phenolic acids and flavonoids. *J. Sep. Sci.* 30: 3268-3295.
- Stecher, G., Huck, C. W., Stoggl, W. M., Bonn, G. K. (2003). Phytoanalysis: a challenge in phytomics. *Trends in Analytical Chemistry*. 22(1).
- Strack D. (1997). Phenolic metabolism. In: Dey PM, Harborne JB, eds. *Plant Biochemistry*. London, UK: Academic Press. 387–416.
- Subbaramaiah, K. and Dannenberg, A. J.( 2003). Cyclooxygenase 2: a molecular target for cancer prevention and treatment. *Trends Pharmacology Science*. 24: 96-102.
- Subbaramaiah, K., Chung, W. J., Michaluart, P., Telang, N., Tanabe,T., Inoue, H., Jang, M., Pezzuto, J. M., Dannenberg, A. J., (1998). Resveratrol inhibits cyclooxygenase-2 transcription and activity in phorbol ester-treated human mammary epithelial cells. *Journal of Biological Chemistry*. 273: 21875–21882.
- Sumpio, B. E., Riley, J. T., Dardik, A. (2002). Cells in focus: endothelial cells. *The International Journal of Biochemistry and Cell Biology*. 34: 1508 - 1512

- Sun, Z. J., Pan, C. E., Liu, H. S., Wang, G. J. (2002). Anti-hepatoma activity of resveratrol *in vitro*. *World J Gastroenterol*. 8(1):79-81.
- Swain, T. and Hillis, W. E. (1959). Phenolic constituents of *Prunus domestica*. I. Quantitative analysis of phenolic constituents. *Journal of the Science of Food and Agriculture*. 10: 63 - 68.
- Szmitko P. E., Wang, C. H., Weisel, R. D., de Almeida, J. R., Anderson. T. J. (2003). New Markers of Inflammation and Endothelial Cell Activation : Part I. *Circulation*. 108: 1917-1923.
- Taylor, R. (1993). Non-communicable diseases in the tropics. *Medical Journal of Australia*. 159: 266-269.
- Tedgui, A., and Mallat, Z. (2001). Anti-Inflammatory Mechanisms in the Vascular. *Circulation Research*. 88: 877-887.
- This, P., Jung, A., Boccacci, P., (2004). Development of a common set of standard varieties and standardized method of scoring microsatellites markers for the analysis of grapevine genetic resources. *Theoretical and Applied Genetics*. 109: 1448–1458.
- Trela, B. and Waterhouse, A. (1996). Resveratrol: isomeric molar absorptivities and stability. *Agriculture and Food Chemistry*. 44: 1253- 1257.
- Valenzano, D. R., Terzibasi, E., Genade, T., Cattaneo, A., Domenici, L., and Cellerino, A. (2006). Resveratrol prolongs lifespan and retards the onset of age-related markers in a short-lived vertebrate. *Current Biology*. 16(3): 296–300.

- Vallverdú, A. Q., Olga, J., Alexander, M. R., Rosa, M. Ş., Lamuela-Raventós. (2011). Evaluation Of A Method Based On LC–ESI–MS/MS For The Characterization Of The Polyphenol Profile Of Organic And Conventional Tomatoes. 5th International Symposium on *Recent Advances in Food Analysis*, Czech Republic.
- Vastano B., Chen, Y., Zhu, N., Ho, C., Zhou, Z., Rosen, R. (2000). Isolation and identification of stilbenes in two varieties of *Polygonum cuspidatum*. *Journal of Agriculture and Food Chemistry*. 48: 253-256.
- Verma, S., Anderson, T.J. (2002). Fundamentals of endothelial function for the clinical cardiologist. *Circulation*. 105: 546–549.
- Wael E. H., and Marcel J. (2005). Isolation of Marine Natural Products. Eds. S. D. Sarker, Z. Latif, and A. I. Gray (2006), *Natural Products Isolation*. Second Edition. Humana Press Inc., Totowa.
- Wallace, A. *Principles and methods of toxicology* (5th ed.). Taylor & Francis Group. 13.
- Wang, Y., Zhao, H., Sheng, X., Gambino P. E., Costello, B., Bojanowski, K. (2002). Protective effect of Fructus lycii polysaccharides against time and hyperthermia-induced damage in cultured seminiferous epithelium. *J. Ethnopharmacol*. 82:169-175.
- Wang, Z. H., Hsu, C. C., and Yin, M. C. (2009). Antioxidative characteristics of aqueous and ethanol extracts of glossy privet fruit. *Food Chemistry*. 112: 914–918.
- Waterhouse, A. L. (2005). Determination of Total Phenolics. Eds. Wrolstad, R. E., Acree, T. E., Decker, E. A., Penner, M. H., Reid, D. S., Schwartz, S. J., Shoemaker, C. F., Smith, D. and Sporns, P. *Handbook of Food Analytical Chemistry: Pigments, Colorants, Flavors, Texture, and Bioactive Food Components*. John Wiley & Sons, Incorporated. Hoboken, NJ. 463 - 470.

- Wrolstad, R. E. (2005). Bioactive Food Components. Eds. Wrolstad, R. E., Acree, T. E., Decker, E. A., Penner, M. H., Reid, D. S., Schwartz, S. J., Shoemaker, C. F., Smith, D. and Sporns, P. *Handbook of Food Analytical Chemistry: Pigments, Colorants, Flavors, Texture, and Bioactive Food Components*. John Wiley & Sons, Incorporated. Hoboken, NJ. 459.
- Xu, H. X., Wan, M., Dong, H., But, P. P. H. and Foo, L. Y. (2000). Inhibitory activity of flavonoids and tannins against HIV-1 protease. *Biological Pharmacological Bulletin*. 23: 1072 - 1076.
- Yach, D., Puska, P. (2002). Globalisation, diets, and non communicable disease. World Health Organisation, Geneva.
- Yang, B., Karlsson, R. M., Oksman, P. H. and Kallio, H. P. (2001). Phytosterols in sea buckthorn (*Hippophae rhamnoides* L.) berries: Identification and effects of different origins and harvesting times. *Journal of Agricultural and Food Chemistry*. 49: 5620 - 5629.
- Yue, X., Zang, W., Deng, M. (2011). Hyper-production of <sup>13</sup>C-labeled trans-resveratrol in *Vitis vinifera* suspension cell culture by elicitation and *in situ* adsorption. *Journal of Biochemical Engineering*. 53: 292 – 296.
- Zycova, T. A., Zhu, F., Zhai, X., Ma, X. Y., Ermakova, S. P., Lee, K. W. (2008). Resveratrol directly targets COX-2 to inhibit carcinogenesis. *Molecular carcinogens*. 47(10): 797-805.