

REMOVAL OF SULFAMETHOXAZOLE AND CEPHALEXIN FROM WATER
BY CATALYTIC OZONATION PROCESS

JAVAID AKHTAR

UNIVERSITI TEKNOLOGI MALAYSIA

REMOVAL OF SULFAMETHOXAZOLE AND CEPHALEXIN FROM WATER
BY CATALYTIC OZONATION PROCESS

JAVAID AKHTAR

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (Chemical Engineering)

Faculty of Chemical Engineering
Universiti Teknologi Malaysia

OCTOBER 2011

Specially dedicated to my beloved mother and father

ACKNOWLEDGEMENT

Alhamdulillah, Praise to Allah, first, I would like to express my sincere, and deep appreciation to my supervisor, Prof. Dr Nor Aishah Saidina Amin for her advice, mentoring, guidance and support in my research. I hereby acknowledge her valuable contribution to my educational achievements and quality assurance. She always encouraged me in difficult times and helped me to surpass through challenges during three-year tenure. I hereby also acknowledge valuable support from Prof. Madya Dr. Zulkafi Buntat from Faculty of Electrical Engineering to solve my experimental issues related to ozone measurement. Finally, I would like to thank Prof. Madya Dr. Azmi Aris for his co-operation during analysis of my experimental samples.

I would like to thank all CREG members for their support and friendship over these years. In particular, to Fauzi, Maryam, Zaki, Mahdir, Linda, and Yani are greatly acknowledged for their helpful discussions and suggestions. I also acknowledge the technical support from Dr. Muhammad Khurram Zahoor from faculty of petroleum and renewable energy engineering. I wish them all the success in their future endeavors.

I would like to thank all laboratory technicians in particular Mr. Latfi, Siti Zalita and laboratory staff form FKA, for their assistance and cooperation throughout the research work to all the administration personnel in the Faculty of Chemical Engineering, Universiti Teknologi Malaysia. I would like to thank especially to Siti Zalita from Makmal Bioprocess for her support to run HPLC analysis of my samples. Her support enabled me to complete my research in time. Lastly, thanks to everyone that I have previously mentioned and to everyone who I may have unintentionally not recognized.

ABSTRACT

This study describes the removal of sulfamethoxazole and cephalexin by catalytic ozonation process in two types of reactors i) batch stirred type and ii) water circulation type. The first step was to screen a suitable catalyst during ozonation of sulfamethoxazole in a batch type reactor. It was observed that loading of $\text{Fe}_2\text{O}_3/\text{CeO}_2$ did not suppress the adsorption capacity of PAC and that adsorption process was by physisorption for $\text{Fe}_2\text{O}_3/\text{CeO}_2$ loaded PAC or PAC. Moreover, the loading of $\text{Fe}_2\text{O}_3/\text{CeO}_2$ synergized the effectiveness of powdered activated carbon (PAC), for removal of sulfamethoxazole during catalytic ozonation. Complete removal of sulfamethoxazole was observed using $\text{Fe}_2\text{O}_3/\text{CeO}_2$ loaded PAC catalyst within 5 min of ozonation on batch reactor. Further screening of catalyst suggested granular activated carbon (GAC) was a better catalyst compared to CeO_2 , MnO_2 , and $\text{MnO}_2\text{-CeO}_2$ metal oxides. In the presence of GAC as catalyst, approximately 90 % of cephalexin was removed in 5 min during batch ozonation process. GAC assisted ozonation of two antibiotics was conducted in a newly developed circulating reactors. Circulating batch reactor removed > 98 % of sulfamethoxazole and > 80% of COD using GAC as catalyst in 15 min duration. Similarly, 80-100% of cephalexin was removed using circulation batch reactor. Biodegradability was increased to more than 90% and 98% for cephalexin and sulfamethoxazole antibiotics respectively using circulating batch ozonation. Finally, a separate study was performed for solid phase regeneration of GAC to emulate the effectiveness of in-situ regeneration during ozonation process. In situ ozonation regenerated GAC efficiently. BET analysis, TPD- N_2 and TGA profiles of regenerated GAC resembled more of virgin GAC and differed from saturated GAC sample.

ABSTRAK

Kajian ini menerangkan penyingkiran sulfamethoxazole dan sefaleksin di dalam proses ozonisasi pemangkin di dalam dua jenis reaktor, iaitu (i) berkelompok teraduk dan (ii) edaran air. Langkah pertama adalah memilih mangkin yang sesuai semasa ozonisasi sulfametoksazol dalam reaktor berkelompok teraduk. Pemerhatian menunjukkan bahawa pemuatan $\text{Fe}_2\text{O}_3/\text{CeO}_2$ tidak menyekat keupayaan penjerapan serbuk karbon teraktivasi (PAC) dan proses penjerapan adalah berupa *physorption* untuk $\text{Fe}_2\text{O}_3/\text{CeO}_2$ dimuatkan PAC atau PAC sendiri. Tambahan pula, pemuatan $\text{Fe}_2\text{O}_3/\text{CeO}_2$ mensinergikan keberkesanan PAC, untuk penyingkiran sulfamethoxazole semasa ozonisasi sebagai pemangkin. Penyingkiran sulfamethoxazole yang lengkap telah diperhatikan apabila menggunakan mangkin $\text{Fe}_2\text{O}_3/\text{CeO}_2$ dimuatkan PAC dalam masa 5 minit ozonisasi pada reaktor kelompok. Pemeriksaan lanjut pemangkin mencadangkan karbon berbutiran diaktifkan (GAC) sebagai pemangkin yang lebih baik berbanding untuk CeO_2 , MnO_2 , dan oksida logam $\text{MnO}_2\text{-CeO}_2$. Dengan kehadiran GAC sebagai pemangkin, kira-kira 90% cephalexin dikeluarkan dalam 5 minit semasa proses ozonisasi kumpulan. GAC ozonisasi dibantu dua antibiotik telah dijalankan dalam reaktor berputar yang baru dibangunkan. Reaktor kelompok berputar mengeluarkan > 98% sulfamethoxazole dan > 80% COD menggunakan GAC sebagai pemangkin dalam tempoh 15 min. Begitu juga, 80-100% cephalexin telah disingkirkan menggunakan reaktor kelompok berputar. Biodegradasi telah meningkat kepada lebih daripada 90% dan 98% bagi antibiotik cephalexin dan sulfamethoxazole, masing-masing menggunakan kumpulan ozonisasi berputar. Akhir sekali, satu kajian berasingan telah dilaksanakan untuk penjaan semula fasa pepejal GAC untuk mengikuti keberkesanan penjaan semula in-situ semasa proses ozonisasi. Ozonisasi in-situ menjana semula GAC dengan cekap. Analisis BET, TPD- N_2 dan profil TGA untuk GAC yang dijana semula didapati menyerupai GAC asal dan berbeza dari sampel GAC tepu.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	TITLE	i
	DECLARATION	ii
	DEDICATIONS	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xv
	LIST OF FIGURES	xvi
	ABBREVIATIONS	xxiii
	LIST OF APPENDICES	xxvi
1	INTRODUCTION	1
	1.1 Pharmaceuticals as Water Pollutant	1
	1.2 Removal of Pharmaceuticals at Point Source	2
	1.3 Problem of Statement	6
	1.4 Research Objectives	8
	1.5 Scope of Research	8
2	LITERATURE REVIEW	9
	2.1 Introduction	9
	2.2 Sulfamethoxazole	12
	2.3 Cephalexin	13

2.4	Occurrence of Sulfamethoxazole and Cephalexin in Water	14
2.5	Removal Options	15
2.5.1	Adsorptive Detoxification	16
2.5.1.1	Physical Adsorptions	16
2.5.1.2	Interactive Sorption	17
2.5.1.3	Functional Group Interactions	18
2.5.1.4	Dissociative Adsorption	20
2.5.2	Effect of Parameters on Adsorption	22
2.5.2.1	pH of Solution	22
2.5.2.2	Liquid phase concentration of PhCs	25
2.5.2.3	Ionic strength	26
2.5.3	Adsorptive Ozonation	27
2.5.4	Ozone as Oxidant	28
2.5.5	Mechanism for Adsorptive Ozonation	29
2.5.6	Ozonation of Sulfamethoxazole	33
2.5.7	Isothermal Equilibrium Models	35
2.5.7.1	Langmuir Model	35
2.5.7.2	Freundlich Model	35
2.5.7.3	Error Analysis	35
2.5.8	Theory of Adsorption Kinetic Models	36
2.5.8.1	Pseudo First Order Model	36
2.5.8.2	Pseudo Second Order Model	37
2.5.8.3	Intra-particle Diffusion Model	37
2.5.9	Theory of RSM	37
2.5.9.1	Statistical Model Fitting and Analysis	39
3	RESEARCH METHODOLOGY	40
3.1	Materials	40
3.2	Catalyst Preparation	40

3.3	General Research Methodology	42
3.4	Reactors Types used in this Study	44
3.4.1	Batch Ozonation Reactor	44
3.4.2	Circulating Reactor	45
3.4.3	Preparation of Antibiotic Solution	46
3.4.4	Sample Preparation	46
3.5	Experimental Procedure	47
3.5.1	Batch Adsorption Studies	47
3.5.2	Ozonation Experiments	48
3.6	Catalyst Characterization	49
3.6.1	X-Ray Diffraction (XRD)	50
3.6.2	BET Surface Area	51
3.6.3	Thermogravimetric analysis (TGA)	52
3.6.4	Temperature programmed desorption analysis	53
3.7	Analytical	53
3.7.1	Measurement of Dissolved Ozone Concentration	53
3.7.2	Ozone Utilization Efficiency	55
3.7.3	HPLC Analysis	55
3.7.4	Solid Phase Extraction (SPE)	56
3.7.5	GC-MS Analysis	57
3.7.6	TOC Analysis	58
3.7.7	COD Analysis	58
3.7.8	BOD Analysis	59
4	CHARACTERIZATION OF CATALYSTS	60
4.1	Catalyst Characterization	60
4.2	XRD Analysis	60
4.3	BET Surface Area	63
4.4	Regeneration of Granular Activated Carbon	65
4.5	Summary	67

5	BATCH OZONATION STUDIES	68
5.1	Introduction	68
5.2	Batch Ozonation of Sulfamethoxazole using Fe ₂ O ₃ /CeO ₂ Loaded Activated Carbon	68
5.2.1	Effect of Adsorbent Dosage	68
5.2.2	SMX Adsorption Kinetics	70
5.2.3	Intraparticle Diffusion Model	73
5.2.4	Isothermal Adsorption of SMX	75
5.2.5	Thermodynamic Parameters of Adsorption	77
5.2.6	Ozonation of SMX	78
5.2.7	Comparison among Catalyst Types for Removal Mechanism of SMX	82
5.3	Effect of Operating Conditions on Catalytic Ozonation of Sulfamethoxazole	84
5.3.1	Effect of Catalyst Types	84
5.3.2	Effect of Concentration of SMX	86
5.3.3	Effect of pH of Solution	88
5.3.4	Ozone Utilization Efficiency	91
5.3.5	Effect of Water Matrix Types	93
5.4	Effect of Operating Conditions on Removal of Cephalexin in Batch Reactor	95
5.4.1	Effect of GAC Dosage on Adsorption of Cephalexin	95
5.4.2	Effect of pH of Solution on Removal of Cephalexin	96
5.4.3	Effect of CEX Concentration on Removal of Cephalexin	97
5.4.4	Effect of GAC Dosage on Removal of Cephalexin	98
5.4.5	Biodegradability of Cephalexin (BOD/COD)	99

5.4.6	Effect of GAC dosage on COD removal	101
5.4.7	Effect of CEX Concentration on COD removal	102
5.4.8	Effect of pH of Solution on COD removal	103
5.4.9	GC-MS Analysis for Degradation Products of Cephalexin	104
5.5	Optimization Studies for Catalytic Ozonation of Cephalexin Antibiotic in a Batch Reactor	105
5.5.1	Response Surface Optimization for CEX Removal	105
5.5.2	Model Development	106
5.5.3	Surface Graphs and Contours	108
5.5.4	Response Surface Optimization for COD Removal	109
5.5.5	Effect of parameters on COD removal	111
5.6	Summary	112
6	REMOVAL OF SULFAMETHOXAZOLE AND CEPHALEXIN IN CIRCULATING REACTOR	114
6.1	Introduction	114
6.2	Effect of Operating Conditions for Catalytic Ozonation of Sulfamethoxazole	114
6.2.1	Effect of Circulation Flow Rate on SMX Removal	114
6.2.2	Effect of Concentration on SMX Removal	115
6.2.3	Effect of O ₃ dosage on SMX Removal	117
6.2.4	Effect of Circulation Rate on COD removal	118
6.2.5	Effect of GAC Dosage on COD removal	119
6.2.6	Effect of O ₃ Dosage on COD Removal	120

6.2.7	Effect of Ozonation time on COD Removal	121
6.2.8	Biodegradability	122
6.3	Optimization Studies for Catalytic Ozonation of Sulfamethoxazole	124
6.3.1	Empirical Model for SMX Removal	125
6.3.2	Surface Graphs	126
6.3.3	Four-parameter Optimization for COD Removal during Ozonation of SMX	128
6.3.4	Surface Graph	129
6.4	Effect of Operating Conditions for Removal of Cephalexin Antibiotic in a Circulating Reactor	131
6.4.1	Effect of O ₃ Dosage	131
6.4.2	Effect of GAC Dosage	131
6.4.3	Effect of Initial Concentration	133
6.4.4	Effect of Time Duration on COD Removal	133
6.4.5	Effect of Circulation Flow Rate on COD Removal	135
6.4.6	Effect of O ₃ Dosage	135
6.4.7	Biodegradability of Cephalexin Solution	136
6.5	Four-parameter Optimization for Removal of Cephalexin by Catalytic Ozonation in a Circulation Reactor	138
6.5.1	Model for CEX Removal	138
6.5.2	Surface Graphs	141
6.5.3	Model Equation for COD Removal in Four Parameter Optimization of CEX	142
6.5.4	Surface Graph	143
6.6	Assessment of Solid Phase Regeneration of GAC using O ₃ as Oxidant	145
6.7	Comparison for Batch and Circulating Reactors	147

6.8	Summary	150
7	CONCLUSIONS AND RECOMMENDATIONS	151
7.1	Conclusion	151
7.2	Contribution	152
7.3	Recommendations	153
	REFERENCES	154
	Appendices A-E	172-198

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Occurrence of commonly detected pharmaceuticals in different water sources	10
2.2	Standard for COD and BOD in effluents of different industry in Malaysian waters [62]	11
2.3	Experimental design for three independent variables	38
3.1	Materials used in the study	41
3.2	Process conditions for analysis of sulfamethoxazole on HPLC using Synergi hydro C-18 column	56
3.3	Process conditions for analysis of cephalexin on HPLC using Synergi hydro C-18 column	56
3.4	Operating conditions for GC-MS analysis	57
4.1	BET surface area of PAC and MOPAC catalysts	63
4.2	BET surface area of VGAC, SGAC, and RGAC samples	66
5.1	Kinetic model for adsorption of SMX on PAC and MOPAC	72
5.2	Freundlich and Langmuir isotherms for adsorption of SMX on PAC and MOPAC	75
5.3	Thermodynamic parameters for adsorption of SMX on PAC and MOPAC	78
5.4	Increase in biodegradability of CEX solution during ozonation	101
5.5	Complete experimental design of uncoded values and experimental response variables	106
5.6	ANOVA table for removal of CEX from solution	108
5.7	Table ANOVA table for removal of COD from solution	110

6.1	Increase in biodegradability of CEX solution during ozonation	123
6.2	Experimental design for four-parameter optimization of sulfamethoxazole during GAC catalyzed ozonation	124
6.3	ANOVA table for SMX removal during four-parameter optimization	126
6.4	ANOVA table for COD removal during four-parameter optimization	128
6.5	Increase in biodegradability of CEX solution during ozonation	138
6.6	Four-parameter experimental design for removal of CEX from solution and experimental response variables	139
6.7	ANOVA table for removal of CEX from solution in four-parameter optimization	140
6.8	ANOVA table for removal of COD during four-parameter optimization of CEX	143
D.1	Peak area for initial and depleted samples of SMX during effect of O ₃ dosage on ozonation of SMX	192

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
1.1	Pathways for pharmaceutical compounds in the aquatic environment	2
2.1	Sulfamethoxazole (a) structural formula (b) pH speciation	13
2.2	Cephalexin antibiotic	14
2.3	Effect of pH of solution on adsorption of different antibiotics	23
2.4	Effect of pH of solution on adsorption of antibiotic	24
2.5	Mechanism for removal of pollutant compound from water by ozone and hydroxyl radical reactions in the presence of activated carbon surface	31
3.1	Procedure for preparation of metal oxides and impregnated metal oxide catalysts	42
3.2	General research methodology (a) batch ozonation studies (b) Ozonation in circulating reactor	43
3.3	Batch ozonation set up; reactor and accessories	44
3.4	Circulating ozonation set up; reactor and accessories	45
3.5	BET surface area graph for calculation of W_m	51
4.1	XRD analysis of mix MOPAC, PAC, and Fe_2O_3/CeO_2 samples	61
4.2	XRD analysis (a) CeO_2 , (b) MnO_2 , (c) MnO_2-CeO_2 samples	62
4.3	BET analysis for PAC, MOPAC, and GAC catalyst samples	64

4.4	Pore size distributions of PAC, MOPAC and GAC samples	64
4.5	Single point BET surface areas for VGAC, RGAC, and SGAC samples	65
4.6	TGA analyses for VGAC, RGAC, and SGAC samples	66
4.7	TPD-N ₂ analyses for VGAC, RGAC, and SGAC sample	67
5.1	Effect of adsorbent dosage on SMX removal	69
5.2	Change in pH of solution during adsorption process	70
5.3	Amount of TOC adsorbed on individual catalysts as a function of time	71
5.4	Intraparticle diffusion model for SMX removal	74
5.5	Equilibrium isotherms for adsorption of SMX	76
5.6	Effect of catalyst type and pH of solution on removal of SMX from solution	79
5.7	Catalytic ozonation of SMX: % TOC removal during ozonation of SMX solution as a function of initial concentration	80
5.8	Ozone utilization curve and percentage η_{O_3} during catalytic ozonation of SMX	82
5.9	Effect of MOPAC on products of ozonation	83
5.10	Decomposition byproducts of SMX ozonation (a) PAC catalyst (b) No catalyst	83
5.11	% removal SMX and COD (SMX _i 150-160 mg/L, pH 7) (a) Adsorption, 60 min (b) Catalytic ozonation, 20 min	85
5.12	Adsorption on GAC (●) COD _i 200 mg/L and GAC/O ₃ ozonation (■) COD _i 340 mg/L; (▲) COD _i 250 mg/L; (◆) COD _i 150 mg/L; pH = 5.	87
5.13	% removal SMX and COD (SMX _i 150-160 mg/L, pH 7) (a) Adsorption, 60 min (b) Catalytic ozonation, 20 min	87

5.14	Removal of COD by GAC/O ₃ , COD _i (290 mg/L) (a) Effect of initial pH of solution (b) change in pH of solution during GAC/O ₃ ozonation	90
5.15	Variations in dissolved ozone concentration at different pH values Conditions same as in Figure 5.14a	91
5.16	Amount of ozone consumed during ozonation under different pH of solution. Operating conditions same as in Figure 5.14a.	92
5.17	Effect of water matrix on removal of SMX and COD Operating conditions: pH _i = 4, SMX _i 200 mg/L	93
5.18	Effect of water matrix on removal of SMX and COD Operating conditions: pH _i = 4 (b) COD _i (290-310 mg/L)	94
5.19	Effect of GAC dosage on adsorption of CEX Conditions: Time 1 hour; Temperature 26 ± 1 °C, CEX concentration, 200 mg/L	96
5.20	Effect of pH of solution on removal of CEX concentration Conditions: CEX concentration 200 mg/L, O ₃ dosage 21 mg/L, GAC dosage 4 g/L, Temp. 26 ± 1 °C.	97
5.21	Effect of initial concentration of CEX on removal of CEX. Conditions: GAC dosage 4 g/L, O ₃ dosage 21 mg/L, Temperature 26 ± 1°C, pH 7-7.5	98
5.22	Effect of initial concentration of CEX on removal of CEX Conditions: GAC dosage 4 g/L, O ₃ dosage 21 mg/L, Temperature 26 ± 1°C, pH 7-7.5	99
5.23	Increase in biodegradability of CEX solution during ozonation	100
5.24	Effect of GAC dosage on removal of CEX and COD during ozonation Condition: CEX conc. 200 mg/L, pH 7- 7.5, O ₃ dosage 21 mg/L, Time CEX 5 min, COD 15 min, Temperature 26 ± 1°C	102

5.25	Effect of CEX concentration on removal of CEX and COD during ozonation Condition: GAC dosage 3 g/L, pH 7-7.5, O ₃ dosage 21 mg/L, Time CEX 5 min, COD 15 min, Temperature 26 ± 1°C	103
5.26	Effect of pH of solution on removal of CEX and COD during ozonation Conditions CEX conc. 200 mg/L, GAC dosage 3 g/L, O ₃ dosage 21 mg/L, Time CEX 5 min, COD 15 min, Temperature 26 ± 1°C	105
5.27	Decomposition byproducts of CEX ozonation in presence of GAC	105
5.28	Effect of ozone dosage and CEX conc. on removal of CEX	109
5.29	Surface graph for removal of COD as a function of CEX conc. and O ₃ dosage	111
6.1	Effect of circulation rate on removal of SMX from solution Conditions: SMX conc. 100 mg/L, GAC dosage 4 g/L, pH 7-7.5, O ₃ dosage 21 mg/L	116
6.2	Effect of SMX concentration on removal of SMX from solution Conditions GAC dosage 4 g/L, pH 7-7.5, O ₃ dosage 21 mg/L, Circulation rate 8 L/min, Sample volume 1100 mL.	116
6.3	Effect of O ₃ dosage on removal of SMX from solution conditions: SMX conc. 200 mg/L GAC dosage 4 g/L, pH 7-7.5, Circulation rate 8 L/min	117
6.4	Effect of circulation rates on removal of SMX from solution: SMX conc. 200 mg/L GAC dosage 4 g/L, pH 7-7.5, O ₃ dosage 21 L/min, and sample volume 1100 mL, Ozonation time COD 15 min, SMX 5 min	119
6.5	Effect of GAC dosage on removal of SMX from solution Conditions: SMX conc. 200 mg/L GAC dosage 4 g/L, pH 7-7.5, Circulation rate 8 L/min, Sample volume 1100 mL, ozonation time COD 15 min, SMX 5 min	120

6.6	Effect of O ₃ dosage on removal of SMX from solution Conditions: SMX conc. 200 mg/L GAC dosage 4 g/L, pH 7-7.5, Circulation rate 8 L/min, Sample volume 1100 mL.	121
6.7	Amount of COD removed as a function of time. Conditions: SMX conc. 200 mg/L GAC dosage 4 g/L, pH 7-7.5, Circulation rate 8 L/min, Sample volume 1100 mL.	122
6.8	Increase in biodegradability as a function of time. Conditions: SMX conc. 200 mg/L GAC dosage 4 g/L, pH 7-7.5, Circulation rate 8 L/min, Sample volume 1100 mL.	123
6.9	Surface graphs for removal of SMX during ozonation	129
6.10	Surface graphs for removal of COD during ozonation of SMX solution	130
6.11	Effect of O ₃ dosage on removal of CEX from solution	131
6.12	Effect of GAC dosage on removal of CEX from solution	132
6.13	Effect of CEX concentration on removal of CEX from solution	133
6.14	Removal of COD during ozonation of CEX	134
6.15	Removal of COD during ozonation of CEX as function of circulation flow rate	135
6.16	Removal of COD during ozonation of CEX as function of O ₃ dosage	136
6.17	Increase in biodegradability of CEX solution as a function of time	137
6.18	Surface graph for removal of CEX from solution.	141
6.19	Surface graphs for removal of COD during ozonation of CEX on circulation reactor	144
6.20	Saturation curve for adsorption of CEX and COD onto VGAC.	145
6.21	Saturation curve for adsorption of CEX and COD onto VGAC and RGAC. Initial CEX = 300 mg/L, gentle stirring, 26 ± 1°C.	146

6.22	Amount of COD adsorbed at equilibrium on RGAC and VGAC.	146
6.23	Amount of CEX adsorbed at equilibrium conditions on RGAC and VGAC	147
6.24	Change in biodegradability and COD values of CEX solution using two reactors. CEX concentration 200 mg/L, pH 7-7.5, Time 30 min, Initial COD 190, O ₃ dosage 21 mg/L, Volume of reactor; 200 mL (stirred batch), 1100 mL (circulating batch)	149
A.1	Batch type ozonation reactor used in this study	172
A.2	Circulating type reactor developed in CREG laboratory for catalytic ozonation of selected pharmaceuticals	173
C.1	Removal of SMX and secondary products during ozonation of SMX solution in the presence of GAC. SMX solution was prepared in deionized water. (Conditions: pH = 4, SMX _i = 200 ppm, O ₃ dosage = 50 mg/L).	178
C.2	Removal of cephalexin and secondary products during ozonation of cephalexin solution in the presence of GAC. Cephalexin solution was prepared in deionized water. Operating conditions: pH = 4, SMX _i = 200 ppm, O ₃ dosage = 50 mg/L	179
C.3	Representative curve for GC-MS analysis of SMX in the presence of MOPAC catalyst. Samples were drawn according to procedure given in section 3.5.2.	180
C.4	Disinfection by-products during ozonation of sulfamethoxazole in the presence of MOPAC catalyst. (a) Sulfanilamide (b) 6-Aminobenzoxazole (c) Propylmaleamic acid (d) 2-Acetylthiazole (e) 2-Propylthiazole (f) Sulfathiazole (g) Sulfonyl phenyl aminol (h) 5-methyl Thiazole	183

C.5	Representative curve for GC-MS analysis of CEX in the presence of granular activated carbon. Samples were drawn according to procedure given in section 3.5.2.	183
C.6	Disinfection by-products during ozonation of Cephalexin in the presence of granular activate carbon catalyst. (a) dimethyl furyl pridine (b) 1-phenyl propanediole, (c) isonitrosoacetophenone (d) Benzenactic acid, methylester (e) Pyroazole, 5-amin 3-methyl phenyl	185
D.1	Van't Hoff plot for calculation of thermodynamic parameters during adsorption of sulfamethoxazole on MOPAC and PAC Results are given in Table 5.3.	186
D.2	Langmuir adsorption isotherms for SMX onto PAC (see section 5.1.7)	187
D.3	Freundlich adsorption isotherms for SMX onto MOPAC (see section 5.1.7)	187
D.4	Freundlich adsorption isotherms for PAC (see section 5.1.7)	188
D.5	Freundlich adsorption isotherms for PAC (see section 5.1.7)	188
D.6	Ozone consumption efficiency and total amount of O ₃ consumed during ozonation of SMX in the presence of two catalysts	190
E.1	Comparison of experimental and predicted values	192
E.2	Surface contours for CEX removal. Effect of ozone dosage and pH	193
E.3	Comparison of experimental and predicted values for COD removal	193
E.4	Contour plot for removal of COD removal in batch ozonation	194
E.5	Comparison of experimental and predicted response for SMX removal	194

E.6	Contour plot for removal of SMX during ozonation Effect of circulation flow rate and GAC dosage	195
E.7	Comparison for experimental and predicted values for COD removal	195
E.8	Contour plots for removal of COD during ozonation of SMX solution	196
E.9	Experimental vs. predicted response for removal of CEX in four-parameter optimization	196
E.10	Contour plot for removal of CEX from solution during four-parameter optimization of CEX	197
E.11	Experimental values vs. predicted response for COD removal in ozonation	197
E.12	Contour plot for removal of COD during ozonation of CEX in circulating reactor	198

LIST OF ABBREVIATIONS

Al ₂ O ₃	-	Aluminum dioxide
ANOVA	-	Analysis of Variance
AOPS	-	Advanced oxidation process
BOD	-	Biological oxygen demand
CEX	-	Cephalexin
COD	-	Chemical oxygen demand
CTNs	-	Carbon nanotubes
DBPs	-	Disinfection byproducts
GAC	-	Granular activated carbon
GC-MS	-	Gas chromatography mass spectroscopy
HPLC	-	High performance liquid chromatography
MOPAC	-	Metal oxide impregnated powdered activated carbon
MPS _{BET}	-	Multipoint surface area
MPSD	-	Marquardt's percent standard deviation
MWNTs	-	Multiwalled nanotubes
OH	-	Hydroxyl radicals
O ₃	-	Ozone
O ₂	-	Oxygen
PAC	-	Powdered activated carbon
PCAC	-	Petroleum coke based activated carbon
PhCs	-	Pharmaceutical compounds
RGAC	-	Regenerated activated carbon
RSM	-	Response surface methodology
SiO ₂	-	Silicon dioxide
SGAC	-	Saturated activated carbon
SMX	-	Sulfamethoxazole
SOGs	-	Surface active group

SPE	-	Solid phase extraction
SPS_{BET}	-	Single point surface area measured at $P/P_0 = 0.02535$
SSE	-	sum of error squares
SWNTs	-	Single wall nanotubes
TBAM	-	Tetrabutylammonium montmorillonite
TiO_2	-	Titanium dioxide
TOC	-	Total organic contents
VGAC	-	Virgin granular activated carbon
V_{mes}	-	Mesoporous volume
V_{micro}	-	Microporous volume
V_{Total}	-	Total volume
WWTPs	-	Wastewater treatment plant

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Reactor types used in this study	171
B	SPE extraction protocols	173
C	Brief overview of HPLC and GC-MS obtained results	177
D	Graphs and calculations	185
E	Graphs for statistical optimization	181

CHAPTER 1

INTRODUCTION

1.1 Pharmaceuticals as Water Pollutant

Advancements in personal care sector injected numerous varieties of pharmaceuticals in modern day health facilities. Although medications served as life saving drugs both for human and animals, their indirect addition to ecosystem has raised many questions to the environment protection [1]. Medicines are stable structures chemically to prolong medication time within the body, which sense non-degradability of such items [2]. Persistence of pharmaceuticals in industrial and municipal water streams is one of environmental hazards polluting ecosystem. Clotrimazole, Mefenamic, diclofenac, erythromycin, colifibric acid [3], Ibuprofen [4], sulfamethoxazole [5] are examples of pharmaceuticals frequently detected in municipal and waste water treatment plant effluents. Researchers have raised concerns about the transportation of pharmaceutically polluted water resources as drinking water supplies or long-term implications to the aquatic life. Although direct effect of pharmaceutical polluted water is less susceptible since concentration of pharmaceuticals in water, streams far lower than prescribed dosages level. Pharmaceuticals are design to interact with biological matter in living organisms and in their physico-chemical behavior. Many of the pharmaceuticals are lipophilic to ease their passage through cell membranes and are reactive to specific types of metabolic interactions only; otherwise remain persistent in the body cells. In a way these pharmaceuticals easily bioaccumulate within the body and induce the harmful effects of terrestrial or aquatic organisms. Figure 1.1 illustrates the exposure, fate, and long-term effects of medical compounds on aquatic organisms. Pharmaceuticals

undergo biodegradation into metabolites during the fate of such substances in the environment. Occurrence of pharmaceutical active compounds and metabolites in the environment depends upon their resistance to the biodegradability. However, presence of these pharmaceutically active substances in ground water, surface, or ocean water shows their persistence for longer time duration and mobile nature.

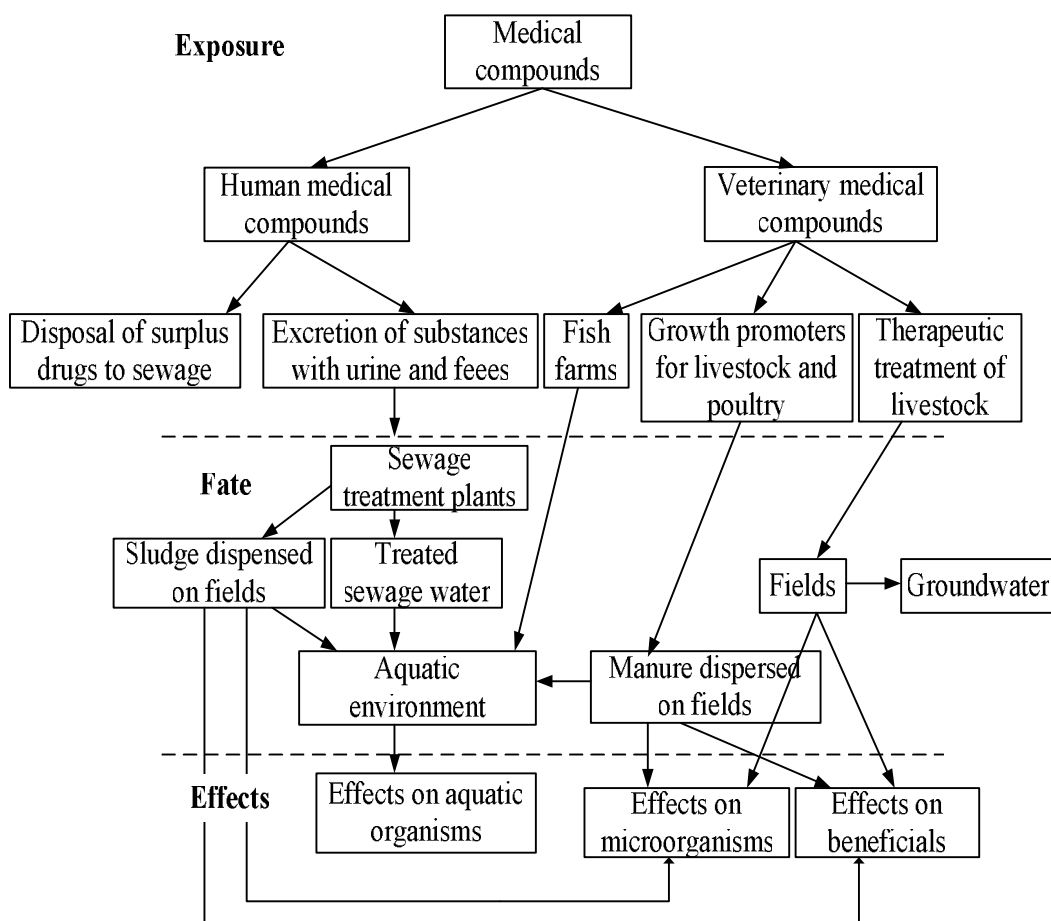


Figure 1.1 Pathways for pharmaceutical compounds in aquatic environment [6]

1.2 Removal of Pharmaceuticals at Point Source

Major sources for induction of in the aquatic environment are urban wastewater, hospitals, pharmaceutical manufacturing facilities, and treatment plants. Proper treatment of these substances at the exit of their source points may reduce the significant volume of pharmaceuticals in the aquatic environment. Treatment at the

exit point seems one viable option if we are to save our water supplies from such pollutants.

Several methods have been adopted in water treatment ranging from conventional filtration [7], biological treatments [8], coagulation [8] to activated carbon [9], electrochemical and advanced oxidation processes [10-11]. These processes differ in treatment capability, operational cost, selectivity and removal efficiency. Biological methods like biofilters, activated sludge are quite effective for biodegradable pollutants. Physical techniques like adsorption, coagulations flocculation, and precipitations are suitable to remove insoluble suspended particles. Activated carbon can effectively remove dissolved organic contaminations. Reverse osmosis, micro, and nano filtrations are other methods for selective removal of micro pollutants. Other than these, advanced oxidation processes such as ozonation, UV, $\text{H}_2\text{O}_2/\text{O}_3$, UV/ O_3 , chlorination are capable of oxidizing soluble, insoluble organic and inorganic contaminants [5]. However, it is true most of the organic and inorganic toxins are removable through water treatment techniques, none of techniques is solely appropriate to handle all types of contaminations. Biological methods cannot grasp synthetic and inorganic pollutions; coagulations and flocculation are inefficient to dissolved micro pollutants; membranes are costly, clog able, and unable to treat macro pollutants; production of DBPs in advanced oxidation processes question usefulness of such operations. Presence of pharmaceuticals in wastewater treatment plants (WWTPs) effluents and water streams also confirms inefficiency of traditional techniques like coagulations, flocculation, and sedimentations [3, 5, 12]. Though it is true, the most WWTPs are equipped to handle various types of contaminations by integration of techniques in series. Bar screening, preliminary clarification, trickling filter, active sludge and UV treatment scheme is an example of such integrations applied in Howdon water treatment works [3]. It is believed that inclusion of ozonation or advanced oxidation processes within this integration may reduce soluble contaminations. Some researchers have reported removal of soluble pollutants using ozone and ozone-assisted oxidations [13-16]. Thus, advanced oxidation processes may be capable of reducing pharmaceuticals and synthetic dyes in wastewater streams.

Advanced oxidation processes (AOPs) have been employed for removing pharmaceuticals active compounds [17-19]; dyes and dyestuff [14, 20-21]; bacterial disinfection [22-23]; pesticides degradation [24-25] and soil decontaminations [26-27]. AOPs rely on production of hydroxyl radicals (OH) through chemical, photochemical and photo catalytic energy that is capable of converting organics into dehydrogenated products [28]. Conventional oxidants within AOPs category include ozone, H₂O₂, chlorine, chlorine dioxide, Hydroxyl ions [29]. These are called aqueous phase oxidants, which attach almost all types of organic and inorganic contaminations. Oxidation potential is one criterion to judge pollutants removal efficiency in such treatments like ozone OH (2.86), O (2.42), O₃ (2.07), H₂O₂ (1.78), Cl (1.36), ClO₂ (1.27). Performance of individual process is also dependent upon generation of hydroxyl ion (OH[•]) which is the most powerful oxidant of this group. For this reason, ozone and H₂O₂ are preferable due their ability to oxidize contaminations directly and through OH ion generation [30]. Due to this ability, ozone has emerged as one major pollutant oxidizer for microorganism's inactivation, metals and suspended solids oxidation, dyes and pigments discoloration, dissolved organic matter and humic acids oxidation, micro pollutants removal. Whilst chlorine and its derivatives are enough to disinfect bacteria present in water their ability to generate lethal chlorinated organic compounds by reacting organic species has limited their role as disinfectants [31]. Electrochemical, Fenton, Photo-Fenton [32], TiO₂/UV [33] are names of AOPs oxidation processes in which induce energy is utilized to generate radicals and ions. Fenton reagents and TiO₂ mediums generate radicals by absorbing near-UV radiations within 300-400 nm range. Electrochemical oxidation involves anodic reactions at high voltage electrodes thus breaking water molecule into hydroxyl radical (OH). In literature, Pt, PbO₂, doped PbO₂, doped SnO₂ have been employed dominantly as anode. Ion generation reaction in equation 2.1 [28].



AOPs are suitable to waste water treatments containing chemically stable, lethal, and/or non-biodegradable pollutants. AOPs have property to degrade any type of contaminations indiscriminately without producing any toxic intermediates at

room temperatures [20]. AOPs effluents are biodegradable due radical's ability to replace chlorines attached to ring structures of organic compounds. Rate constant of organic molecules destruction remains in order of 10^6 - 10^9 $M^{-1} S^{-1}$, thus minimizing process residence times [29]. AOPs have certain advantages over conventional water treatment methods. AOPs are not refractory to wide varieties of feed contaminations and disinfection byproducts are not usually produced which simplifies operations. AOPs are better than bioremediation and chemical coagulations because later produce sludge waste materials and operate selectively on specific types of pollutants. Post processing is costly in membrane processes due to choking problem while AOPs completely mineralize organic matter and avoid any further processing of organic materials. Carbon catalyst poisoning is the major drawback in activated carbon absorption whereas no such problems are associated with AOPs (Spartan water treatment). However, high capital and operating cost of AOPs is a major drawback when compared to biological treatments and chemical coagulations. Literature usually recommends integration of different oxidants for treatment process like O_3/UV , O_3/H_2O_2 , Photo/Fenton, TiO_2/UV [30, 34] mainly due their inability to produce high concentrations of hydroxyl ions (OH) individually. One of the commonly used advanced oxidant (ozone) is highly energy intensive consumes high voltages in order of 4-20 kV. As ozone is degradable to simpler oxygen at room temperatures, high concentrations of ozone need continuous ozone generation. Other AOPs, Fenton/ H_2O_2 systems produce considerable amounts of iron sludge wastes [35]. Electrochemical processes usually involve costly electrodes.

Irrespective of the practical limitations, advanced oxidation processes continued their penetration in water and wastewater treatments. Ozone has emerged as one of the popular oxidant in recent times [13, 15, 17, 19, 21, 36-38]. Probably this is because i) ozone is easily soluble in water (0.57 g/L 20 °C), ii) ozone decomposes readily into hydroxyl ion (OH), iii) oxidation potential is high (2.87 V) [38]. Major pollutants divisions which have been tested for ozone dosages are i) metals and inorganic substances removal ii) Oxidation of suspended and dissolved organic matter iii) bacterial and viral disinfection iv) Discoloration and v) detoxification of harmful chemicals [13, 39]. Camel and Bermond, [39] divided existing literature on ozonation in three dosage levels pre-oxidation, intermediate

ozonation and final disinfection. Ozone is added at pre-oxidation stage to remove colorants and odors, inorganic and suspended materials; to increase coagulations-decantation. Micro pollutants and DBPs are generally removed in second stage dosage, which also enhances biodegradability of organic matter. Final disinfection stage is capable of removing all types of microorganisms, micro pollutants and reducing DBPs [39]. Number of citations notified effects of ozonation on pharmaceuticals degradation from wastewater streams [17, 36, 40-43]. Thus, ozonation processes are widely accepted techniques in removal of micro pollutants like pharmaceuticals from water streams.

1.3 Problem of Statement

Organic compounds such as pharmaceuticals, active personal care products (PPCPs), industrial and household chemicals are potential threat to human health and aquatic ecosystem. These organic chemical collectively called micropollutants involve endocrine disrupting effects and chronic effects on long-term exposure [44]. Some of the pharmaceuticals have shown ineffectiveness to advance treatment technologies such as membrane separation, activated carbon adsorption, ultraviolet radiations, and ozonation [45]. Pharmaceutical compounds are even more likely in effluents of conventional treatment plants. Therefore, it seems necessary to investigate on modern technologies to treat these new types of pollutants in water resources. Moreover, due to low concentration of these micropollutants, conventional treatments based on physical or biological treatments fail to eliminate these compounds from water properly. It may be helpful to investigate on modern treatment methods for treatment of micropollutants.

Ozonation is one attractive option to degrade pharmaceuticals at the exit of point source. Simple procedure can be the reaction of dissolved ozone with pharmaceutical compound. Pharmaceuticals are relatively active species due to the presence of different functional groups that are designed to interact with metabolism. Therefore, it is presumable that main pharmaceutical compound can degrade in short exposure to dissolved ozone. However, degree of mineralization might be low.

Simple ozonation also may not be effective in achieving high ozone mass transfer efficiency. Coupling of simple ozonation with a suitable adsorptive catalyst such as activated carbon might perform effective role in removing pharmaceutical compounds from water. Activated carbon acts as an adsorbent and catalyst during the process. Activated carbon can absorb sufficient amount of pharmaceuticals on its surface in origin and oxidized byproducts form due to its porous structure and non-selective nature. At the same time, activated carbon can decompose the dissolved ozone into oxidants such as OH/O radicals. Decomposition of dissolved ozone also induces the transfer of ozone mass from gas to liquid.

In general, sufficient amount of ozone pass through the reactor column in unutilized form during ozonation process. That might be due to many reasons such as excess amount of ozone in the feed gas, incapability of system to dissolve gas phase ozone into the solution or inefficient reasons between pharmaceutical and ozone. Addition of catalyst as activated carbon may help the better utilization of input ozone gas. Various studies highlighted such an issue where outgoing gas retains sufficient quantity of ozone gas which either need to trap in solutions or to destroy [22, 29]. Extended post processing of gas adds capital and operation cost of ozonation processes besides wasting costly O_3 into atmosphere. Proper utilization of generated O_3 is challenging in ozonation processes that may be solved by utilizing proper absorber design, catalyzed ozonation, and ozone diffusers.

In this research we focused on the maximizing the ozone utilization during the ozonation process. Options that we tried include the usage of activated carbon as catalyst and adsorbent. Secondly, we proposed the circulating absorber column reactor with using venturi mixture. Two antibiotics were selected (sulfamethoxazole and cephalexin) as model compounds. These two are commonly prescribed medicines in daily healthcare activities across the world and are often detected in the urban water and in effluents of wastewater treatment plants. Secondly, these two belong to different class of antibiotics and represent major prescribed antibiotic classes. By using these two antibiotics, it is assumed, ozonation can be applied to other antibiotics or pharmaceuticals as well.

1.4 Research Objectives

Major objectives of the research are as follows

1. To study the degradation of two antibiotic compounds (sulfamethoxazole and cephalexin) during catalytic ozonation process.
2. To screen suitable catalyst for removal of antibiotics during ozonation,
3. To compare the performance of stirred batch reactor and circulating reactor for removal of two antibiotic compounds.

1.5 Scope of Research

1. Initial screening of catalyst is performed for degradation of sulfamethoxazole antibiotic. Initial screening is performed by comparing the performance of activated carbons, metal oxides and metal loaded activated carbon catalysts. The selected catalyst is investigated further to assess the effect of operating parameters and kinetics of sulfamethoxazole. Removal of cephalexin is investigated with screened catalyst only.
2. Dissolved ozone concentration is investigated to compare the ozone decomposition behavior of catalysts. Dissolved ozone concentration is measured in case of selected catalyst for both sulfamethoxazole and cephalexin. Some experiments are conducted to measure ozone utilization efficiency for both antibiotics.
3. Performance comparison of two reactors is investigated by degrading cephalexin and sulfamethoxazole antibiotics in circulating reactor and comparing the results with that of stirred batch reactor.
4. Analysis of the antibiotics is performed using high performance liquid chromatography (HPLC) to measure their concentration during experiments. Degree of mineralization is measured by TOC and COD analysis. While for cephalexin is analyzed by COD and biological oxygen demand (BOD) analysis. Secondary byproducts for two antibiotics are analyzed in gas chromatography mass spectroscopy (GC-MS).

REFERENCES

1. Peterson, J. W., Burkhart, R. S., Shaw, D. C., Schuiling, A. B., Haserodt, M. J. and Seymour, M. D. (2010). Experimental determination of ampicillin adsorption to nanometer-size Al_2O_3 in water. *Chemosphere*. 80: 1268-1273.
2. Poznyak, T., Bautista, G. L., Chaírez, I., Córdova, R. I. and Ríos, L. E. (2008). Decomposition of toxic pollutants in landfill leachate by ozone after coagulation treatment. *Journal of Hazardous Materials*. 152: 1108-1114.
3. Roberts, P. H. and Thomas, K. V. (2006). The occurrence of selected pharmaceuticals in wastewater effluent and surface waters of the lower tyne catchment. *Science of The Total Environment*. 356: 143-153.
4. Klavarioti, M., Mantzavinos, D. and Kassinos, D. (2009). Removal of residual pharmaceuticals from aqueous systems by advanced oxidation processes. *Environment International*. 35: 402-417.
5. Kim, S. D., Cho, J., Kim, I. S., Vanderford, B. J. and Snyder, S. A. (2007). Occurrence and removal of pharmaceuticals and endocrine disruptors in south korean surface, drinking, and waste waters. *Water Research*. 41: 1013-1021.
6. Halling-Sørensen, B., Nors Nielsen, S., Lanzky, P. F., Ingerslev, F., Holten Lützhøft, H. C. and Jørgensen, S. E. (1998). Occurrence, fate and effects of pharmaceutical substances in the environment- a review. *Chemosphere*. 36: 357-393.
7. Stackelberg, P. E., Gibs, J., Furlong, E. T., Meyer, M. T., Zaugg, S. D. and Lippincott, R. L. (2007). Efficiency of conventional drinking-water-treatment processes in removal of pharmaceuticals and other organic compounds. *Science of The Total Environment*. 377: 255-272.
8. Clara, M., Strenn, B., Gans, O., Martinez, E., Kreuzinger, N. and Kroiss, H. (2005). Removal of selected pharmaceuticals, fragrances and endocrine disrupting compounds in a membrane bioreactor and conventional wastewater treatment plants. *Water Research*. 39: 4797-4807.

9. Sánchez-Polo, M., Rivera-Utrilla, J. and von Gunten, U. (2006). Metal-doped carbon aerogels as catalysts during ozonation processes in aqueous solutions. *Water Research*. 40: 3375-3384.
10. Kusic, H., Koprivanac, N. and Bozic, A. L. (2006). Minimization of organic pollutant content in aqueous solution by means of aops: Uv- and ozone-based technologies. *Chemical Engineering Journal*. 123: 127-137.
11. Vogna, D., Marotta, R., Napolitano, A., Andreozzi, R. and d'Ischia, M. (2004). Advanced oxidation of the pharmaceutical drug diclofenac with uv/h₂O₂ and ozone. *Water Research*. 38: 414-422.
12. Han, G. H., Hur, H. G. and Kim, S. D. (2006). Ecotoxicological risk of pharmaceuticals from wastewater treatment plants in Korea: Occurrence and toxicity to *Daphnia magna*. *Environmental Toxicology and Chemistry*. 25: 265-271.
13. Khadhraoui, M., Trabelsi, H., Ksibi, M., Bouguerra, S. and Elleuch, B. (2009). Discoloration and detoxification of a Congo red dye solution by means of ozone treatment for a possible water reuse. *Journal of Hazardous Materials*. 161: 974-981.
14. Arslan, I. and Balcioglu, I. A. (1999). Degradation of commercial reactive dyestuffs by heterogeneous and homogeneous advanced oxidation processes: A comparative study. *Dyes and Pigments*. 43: 95-108.
15. Erol, F. and Özbelge, T. A. (2008). Catalytic ozonation with non-polar bonded alumina phases for treatment of aqueous dye solutions in a semi-batch reactor. *Chemical Engineering Journal*. 139: 272-283.
16. Rosal, R., Rodríguez, A., Perdigón-Melón, J. A., Mezcua, M., Hernando, M. D., Letón, P., García-Calvo, E., Agüera, A. and Fernández-Alba, A. R. (2008). Removal of pharmaceuticals and kinetics of mineralization by O₃/H₂O₂ in a biotreated municipal wastewater. *Water Research*. 42: 3719-3728.
17. Hua, W., Bennett, E. R. and Letcher, R. J. (2006). Ozone treatment and the depletion of detectable pharmaceuticals and atrazine herbicide in drinking water sourced from the upper Detroit River, Ontario, Canada. *Water Research*. 40: 2259-2266.

18. Ternes, T. A., Meisenheimer, M., McDowell, D., Sacher, F., Brauch, H.-J., Haist-Gulde, B., Preuss, G., Wilme, U. and Zulei-Seibert, N. (2002). Removal of pharmaceuticals during drinking water treatment. *Environmental Science & Technology*. 36: 3855-3863.
19. Zwiener, C. and Frimmel, F. H. (2000). Oxidative treatment of pharmaceuticals in water. *Water Research*. 34: 1881-1885.
20. Faouzi, M., Cañizares, P., Gadri, A., Lobato, J., Nasr, B., Paz, R., Rodrigo, M. A. and Saez, C. (2006). Advanced oxidation processes for the treatment of wastes polluted with azoic dyes. *Electrochimica Acta*. 52: 325-331.
21. Ledakowicz, S., Solecka, M. and Zylla, R. (2001). Biodegradation, decolourisation and detoxification of textile wastewater enhanced by advanced oxidation processes. *Journal of Biotechnology*. 89: 175-184.
22. Tiwari, B. K., O'Donnell, C. P., Muthukumarappan, K. and Cullen, P. J. (2009). Anthocyanin and colour degradation in ozone treated blackberry juice. *Innovative Food Science & Emerging Technologies*. 10: 70-75.
23. Guzel-Seydim, Z. B., Greene, A. K. and Seydim, A. C. (2004). Use of ozone in the food industry. *Lebensmittel-Wissenschaft und-Technologie*. 37: 453-460.
24. Javier Benitez, F., Acero, J. L. and Real, F. J. (2002). Degradation of carbofuran by using ozone, uv radiation and advanced oxidation processes. *Journal of Hazardous Materials*. 89: 51-65.
25. Somich, C. J., Muldoon, M. T. and Kearney, P. C. (1990). On-site treatment of pesticide waste and rinsate using ozone and biologically active soil. *Environmental Science & Technology*. 24: 745-749.
26. Takayama, M., Ebihara, K., Stryczewska, H., Ikegami, T., Gyoutoku, Y., Kubo, K. and Tachibana, M. (2006). Ozone generation by dielectric barrier discharge for soil sterilization. *Thin Solid Films*. 506-507: 396-399.
27. Pierpoint, A. C., Hapeman, C. J. and Torrents, A. (2003). Ozone treatment of soil contaminated with aniline and trifluralin. *Chemosphere*. 50: 1025-1034.
28. Maezono, T., Tokumura, M., Sekine, M. and Kawase, Y. (2011). Hydroxyl radical concentration profile in photo-fenton oxidation process: Generation and consumption of hydroxyl radicals during the discoloration of azo-dye orange ii. *Chemosphere*. 82: 1422-1430.

29. Lafi, W. K. and Al-Qodah, Z. (2006). Combined advanced oxidation and biological treatment processes for the removal of pesticides from aqueous solutions. *Journal of Hazardous Materials*. 137: 489-497.
30. Finzgar, N. and Lestan, D. (2006). Heap leaching of pb and zn contaminated soil using ozone/uv treatment of edta extractants. *Chemosphere*. 63: 1736-1743.
31. Siddiqui, M. S., Amy, G. L. and Murphy, B. D. (1997). Ozone enhanced removal of natural organic matter from drinking water sources. *Water Research*. 31: 3098-3106.
32. Arslan-Alaton, I. and Dogruel, S. (2004). Pre-treatment of penicillin formulation effluent by advanced oxidation processes. *Journal of Hazardous Materials*. 112: 105-113.
33. Arslan, I., Akmehmet, B. I. and Tuhkanen, T. (2000). Advanced treatment of dyehouse effluents by fe(ii) and mn(ii)-catalyzed ozonation and the h₂O₂/O₃ process. *Water Science & Technology*. 42: 13-18.
34. Alaton, I. A., Balcioglu, I. A. and Bahnemann, D. W. (2002). Advanced oxidation of a reactive dyebath effluent: Comparison of O₃, H₂O₂/uv-c and TiO₂/uv-a processes. *Water Research*. 36: 1143-1154.
35. Badawy, M. I., Ghaly, M. Y. and Gad-Allah, T. A. (2006). Advanced oxidation processes for the removal of organophosphorus pesticides from wastewater. *Desalination*. 194: 166-175.
36. Huber, M. M., Canonica, S., Park, G.-Y. and von Gunten, U. (2003). Oxidation of pharmaceuticals during ozonation and advanced oxidation processes. *Environmental Science & Technology*. 37: 1016-1024.
37. Bougrier, C., Albasi, C., Delgenès, J. P. and Carrère, H. (2006). Effect of ultrasonic, thermal and ozone pre-treatments on waste activated sludge solubilisation and anaerobic biodegradability. *Chemical Engineering and Processing*. 45: 711-718.
38. Magara, Y., Itoh, M. and Morioka, T. (1995). Application of ozone to water treatment and power consumption of ozone generating systems. *Progress in Nuclear Energy*. 29: 175-182.

39. Camel, V. and Bermond, A. (1998). The use of ozone and associated oxidation processes in drinking water treatment. *Water Research*. 32: 3208-3222.
40. Vieno, N. M., Härkki, H., Tuhkanen, T. and Kronberg, L. (2007). Occurrence of pharmaceuticals in river water and their elimination in a pilot-scale drinking water treatment plant. *Environmental Science & Technology*. 41: 5077-5084.
41. McDowell, D. C., Huber, M. M., Wagner, M., von Gunten, U. and Ternes, T. A. (2005). Ozonation of carbamazepine in drinking water: Identification and kinetic study of major oxidation products. *Environmental Science & Technology*. 39: 8014-8022.
42. Huber, M. M., Göbel, A., Joss, A., Hermann, N., Löffler, D., McArdell, C. S., Ried, A., Siegrist, H., Ternes, T. A. and von Gunten, U. (2005). Oxidation of pharmaceuticals during ozonation of municipal wastewater effluents: A pilot study. *Environmental Science & Technology*. 39: 4290-4299.
43. Andreozzi, R., Raffaele, M. and Nicklas, P. (2003). Pharmaceuticals in stp effluents and their solar photodegradation in aquatic environment. *Chemosphere*. 50: 1319-1330.
44. Musloff, A., Leschik, S., Reinstorf, F., Strauch, G. and Schirmer, M. (2010). Micropollutant loads in the urban water cycle *Environmental Science Technology*. 44: 4877-4888.
45. Jones, O. A., Lester, J. N. and Voulvoulis, N. (2005). Pharmaceuticals: A threat to drinking water? *TRENDS in Biotechnology*. 23: 163-167.
46. Drillia, P., Dokianakis, S. N., Fountoulakis, M. S., Kornaros, M., Stamatelatou, K. and Lyberatos, G. (2005). On the occasional biodegradation of pharmaceuticals in the activated sludge process: The example of the antibiotic sulfamethoxazole. *Journal of Hazardous Materials*. 122: 259-265.
47. Daughton, C. G. and Ternes, T. A. (1999). Pharmaceuticals and personal care products in the environment: Agents of subtle change? *Environmental Health Perspectives*. 107: 907-938.
48. Kümmerer, K. (2001). Drugs in the environment: Emission of drugs, diagnostic aids and disinfectants into wastewater by hospitals in relation to other sources - a review. *Chemosphere*. 45: 957-969.

49. Kim, S. H., Shon, H. K. and Ngo, H. H. (2010). Adsorption characteristics of antibiotics trimethoprim on powdered and granular activated carbon. *Journal of Industrial and Engineering Chemistry*. 16: 344-349.
50. Kasprzyk-Hordern, B., Ziólek, M. and Nawrocki, J. (2003). Catalytic ozonation and methods of enhancing molecular ozone reactions in water treatment. *Applied Catalysis B: Environmental*. 46: 639-669.
51. Esplugas, S., Bila, D. M., Krause, L. G. T. and Dezotti, M. (2007). Ozonation and advanced oxidation technologies to remove endocrine disrupting chemicals (edcs) and pharmaceuticals and personal care products (ppcps) in water effluents. *Journal of Hazardous Materials*. 149: 631-642.
52. Nakada, N., Shinohara, H., Murata, A., Kiri, K., Managaki, S., Sato, N. and Takada, H. (2007). Removal of selected pharmaceuticals and personal care products (ppcps) and endocrine-disrupting chemicals (edcs) during sand filtration and ozonation at a municipal sewage treatment plant. *Water Research*. 41: 4373-4382.
53. Carballa, M., Manterola, G., Larrea, L., Ternes, T., Omil, F. and Lema, J. M. (2007). Influence of ozone pre-treatment on sludge anaerobic digestion: Removal of pharmaceutical and personal care products. *Chemosphere*. 67: 1444-1452.
54. Jones, O. A., Lester, J. N. and Voulvoulis, N. (2005). Pharmaceuticals: A threat to drinking water? *Trends in Biotechnology*. 23: 163-167.
55. Castiglioni, S., Bagnati, R., Fanelli, R., Pomati, F., Calamari, D. and Zuccato, E. (2005). Removal of pharmaceuticals in sewage treatment plants in Italy. *Environ. Sci. Technol.* 40: 357-363.
56. Kim, S. D., Cho, J., Kim, I. S., Vanderford, B. J. and Snyder, S. A. (2007). Occurrence and removal of pharmaceuticals and endocrine disruptors in South Korean surface, drinking, and waste waters. *Water Res.* 41: 1013-1021.
57. Spongberg, A. L. and Witter, J. D. (2008). Pharmaceutical compounds in the wastewater process stream in Northwest Ohio. *Sci. Total Environ.* 397: 148-157.
58. Yoon, Y., Ryu, J., Oh, J., Choi, B.-G. and Snyder, S. A. (2010). Occurrence of endocrine disrupting compounds, pharmaceuticals, and personal care

- products in the han river (seoul, south korea). *Science Total Environ.* 408: 636-643.
59. Wang, C.-J., Li, Z., Jiang, W.-T., Jean, J.-S. and Liu, C.-C. (2010). Cation exchange interaction between antibiotic ciprofloxacin and montmorillonite. *J. Hazard. Mater.* 183: 309-314.
 60. Watkinson, A. J., Murby, E. J., Kolpin, D. W. and Costanzo, S. D. (2009). The occurrence of antibiotics in an urban watershed: From wastewater to drinking water. *Sci. Total Environ.* 407: 2711-2723.
 61. Lin, A. Y.-C., Yu, T.-H. and Lin, C.-F. (2008). Pharmaceutical contamination in residential, industrial, and agricultural waste streams: Risk to aqueous environments in taiwan. *Chemosphere.* 74: 131-141.
 62. Malaysian Standards. (2009). Environmental quality act 1974, environmental quality (industrial effluent regulations 2009).
 63. Gupta, V. K. and Suhas. (2009). Application of low-cost adsorbents for dye removal - a review. *Journal of Environmental Management.* 90: 2313-2342.
 64. Crini, G. (2006). Non-conventional low-cost adsorbents for dye removal: A review. *Bioresource Technology.* 97: 1061-1085.
 65. Pavoni, B., Drusian, D., Giacometti, A. and Zanette, M. (2006). Assessment of organic chlorinated compound removal from aqueous matrices by adsorption on activated carbon. *Water Research.* 40: 3571-3579.
 66. Gupta, V. K., Mittal, A., Kurup, L. and Mittal, J. (2006). Adsorption of a hazardous dye, erythrosine, over hen feathers. *Journal of Colloid and Interface Science.* 304: 52-57.
 67. Rafatullah, M., Sulaiman, O., Hashim, R. and Ahmad, A. (2010). Adsorption of methylene blue on low-cost adsorbents: A review. *Journal of Hazardous Materials.* 177: 70-80.
 68. Ramos, A. M., Otero, M. and Rodrigues, A. E. (2004). Recovery of vitamin b12 and cephalosporin-c from aqueous solutions by adsorption on non-ionic polymeric adsorbents. *Separation and Purification Technology.* 38: 85-98.
 69. Qi, S., Schideman, L., Mariñas, B. J., Snoeyink, V. L. and Campos, C. (2007). Simplification of the iast for activated carbon adsorption of trace organic compounds from natural water. *Water Research.* 41: 440-448.

70. Yu, Z., Peldszus, S. and Huck, P. M. (2008). Adsorption characteristics of selected pharmaceuticals and an endocrine disrupting compound--naproxen, carbamazepine and nonylphenol--on activated carbon. *Water Research*. 42: 2873-2882.
71. Robberson, K. A., Waghe, A. B., Sabatini, D. A. and Butler, E. C. (2006). Adsorption of the quinolone antibiotic nalidixic acid onto anion-exchange and neutral polymers. *Chemosphere*. 63: 934-941.
72. Bui, T. X. and Choi, H. (2009). Adsorptive removal of selected pharmaceuticals by mesoporous silica sba-15. *Journal of Hazardous Materials*. 168: 602-608.
73. Pocostales, J. P., Alvarez, P. M. and Beltrán, F. J. (2010). Kinetic modeling of powdered activated carbon ozonation of sulfamethoxazole in water. *Chemical Engineering Journal*. 164: 70-76.
74. Dantas, R. F., Contreras, S., Sans, C. and Esplugas, S. (2008). Sulfamethoxazole abatement by means of ozonation. *Journal of Hazardous Materials*. 150: 790-794.
75. Zhou, W. and Moore, D. E. (1997). Photosensitizing activity of the anti-bacterial drugs sulfamethoxazole and trimethoprim. *Journal of Photochemistry and Photobiology B: Biology*. 39: 63-72.
76. Cavallucci, S. (2007). What's topping the charts in prescription drugs this yea. *Pharmacypractice*.
77. Boreen, A. L., Arnold, W. A. and McNeill, K. (2004). Photochemical fate of sulfa drugs in the aquatic environment: Sulfa drugs containing five-membered heterocyclic groups. *Environmental Science & Technology*. 38: 3933-3940.
78. Watkinson, A. J., Murby, E. J., Kolpin, D. W. and Costanzo, S. D. (2009). The occurrence of antibiotics in an urban watershed: From wastewater to drinking water. *Science of The Total Environment*. 407: 2711-2723.
79. Dutta, M., Dutta, N. N. and Bhattacharya, K. G. (1999). Aqueous phase adsorption of certain beta-lactam antibiotics onto polymeric resins and activated carbon. *Separation and Purification Technology*. 16: 213-224.
80. Liu, H., Liu, W., Zhang, J., Zhang, C., Ren, L. and Li, Y. (2011). Removal of cephalexin from aqueous solutions by original and cu(ii)/fe(iii) impregnated

- activated carbons developed from lotus stalks kinetics and equilibrium studies. *Journal of Hazardous Materials*. 185: 1528-1535.
81. Kümmerer, K. (2003). Significance of antibiotics in the environment. *Journal of Antimicrobial Chemotherapy* 52:
 82. Al-Ahmad, A., Daschner, F. D. and Kümmerer, K. (1999). Biodegradability of cefotiam, ciprofloxacin, meropenem, penicillin g, and sulfamethoxazole and inhibition of waste water bacteria. *Archives of Environmental Contamination and Toxicology*. 37: 158-63.
 83. Hartig, C., Storm, T. and Jekel, M. (1999). Detection and identification of sulphonamide drugs in municipal waste water by liquid chromatography coupled with electrospray ionisation tandem mass spectrometry. *Journal of Chromatography A*. 854: 163-173.
 84. Dodd, M. C. and Huang, C.-H. (2004). Transformation of the antibacterial agent sulfamethoxazole in reactions with chlorine: Kinetics, mechanisms, and pathways. *Environmental Science & Technology*. 38: 5607-5615.
 85. Gagné, F., Blaise, C. and André, C. (2006). Occurrence of pharmaceutical products in a municipal effluent and toxicity to rainbow trout (*Oncorhynchus mykiss*) hepatocytes. *Ecotoxicology and Environmental Safety*. 64: 329-336.
 86. Kolpin, D. W., Furlong, E. T., Meyer, M. T., Thurman, E. M., Zaugg, S. D., Barber, L. B. and Buxton, H. T. (2002). Pharmaceuticals, hormones, and other organic wastewater contaminants in u.S. Streams , 1999-2000: A national reconnaissance. *Environmental Science Technology*. 36: 1202-11.
 87. Goyne, K. W., Chorover, J., Kubicki, J. D., Zimmerman, A. R. and Brantley, S. L. (2005). Sorption of the antibiotic ofloxacin to mesoporous and nonporous alumina and silica. *Journal of Colloid and Interface Science*. 283: 160-170.
 88. Ji, L., Chen, W., Zheng, S., Xu, Z. and Zhu, D. (2009). Adsorption of sulfonamide antibiotics to multiwalled carbon nanotubes. *Langmuir*. 25: 11608-11613.
 89. Lee, J. W., Park, H. C. and Moon, H. (1997). Adsorption and desorption of cephalosporin c on nonionic polymeric sorbents. *Separation and Purification Technology*. 12: 1-11.

90. Shane A. Snyder, P. W., Yeomin Yoon, David L. Sedlak. (2003). Pharmaceuticals, personal care products, and endocrine disruptors in water: Implications for the water industry. *Environmental Engineering Science*. 20: 449-469.
91. Le-Minh, N., Khan, S. J., Drewes, J. E. and Stuetz, R. M. (2010). Fate of antibiotics during municipal water recycling treatment processes. *Water Research*. 44: 4295-4323.
92. Aksu, Z. and Tunç, Ö. (2005). Application of biosorption for penicillin G removal: Comparison with activated carbon. *Process Biochemistry*. 40: 831-847.
93. Kulshrestha, P., Giese, R. F. and Aga, D. S. (2004). Investigating the molecular interactions of oxytetracycline in clay and organic matter: Insights on factors affecting its mobility in soil. *Environmental Science & Technology*. 38: 4097-4105.
94. Zimnitsky, D. S., Yurkshtovich, T. L. and Bychkovsky, P. M. (2004). Adsorption of zwitterionic drugs on oxidized cellulose from aqueous and water/alcohol solutions. *The Journal of Physical Chemistry B*. 108: 17812-17817.
95. Claudius, J. S. and Neau, S. H. (1996). Kinetic and equilibrium characterization of interactions between glycopeptide antibiotics and sodium carboxymethyl starch. *International Journal of Pharmaceutics*. 144: 71-79.
96. Li, Z., Schulz, L., Ackley, C. and Fenske, N. (2010). Adsorption of tetracycline on kaolinite with pH-dependent surface charges. *Journal of Colloid and Interface Science*. 351: 254-260.
97. Wang, C.-J., Li, Z., Jiang, W.-T., Jean, J.-S. and Liu, C.-C. (2010). Cation exchange interaction between antibiotic ciprofloxacin and montmorillonite. *Journal of Hazardous Materials*. 183: 309-314.
98. Wu, Q., Li, Z., Hong, H., Yin, K. and Tie, L. (2010). Adsorption and intercalation of ciprofloxacin on montmorillonite. *Applied Clay Science*. 50: 204-211.
99. Putra, E. K., Pranowo, R., Sunarso, J., Indraswati, N. and Ismadji, S. (2009). Performance of activated carbon and bentonite for adsorption of amoxicillin

- from wastewater: Mechanisms, isotherms and kinetics. *Water Research*. 43: 2419-2430.
100. Khandal, R. K., Thoisy-Dur, J. C. and Terce, M. (1991). Adsorption characteristics of flumequine on kaolinitic clay. *Geoderma*. 50: 95-107.
 101. Chang, P.-H., Li, Z., Jiang, W.-T. and Jean, J.-S. (2009). Adsorption and intercalation of tetracycline by swelling clay minerals. *Applied Clay Science*. 46: 27-36.
 102. Akçay, G., Killınç, E. and Akçay, M. (2009). The equilibrium and kinetics studies of flurbiprofen adsorption onto tetrabutylammonium montmorillonite (tbam). *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 335: 189-193.
 103. IUPAC. (1997). Compendium of chemical terminology 2nd edition.
 104. Ania, C., Pelayo, J. and Bandosz, T. (2010). Reactive adsorption of penicillin on activated carbons. *Adsorption*. 1-9.
 105. Zhang, H. and Huang, C.-H. (2007). Adsorption and oxidation of fluoroquinolone antibacterial agents and structurally related amines with goethite. *Chemosphere*. 66: 1502-1512.
 106. SAKA, E. E. and GULER, C. (2006). The effects of electrolyte concentration, ion species and ph on the zeta potential and electrokinetic charge density of montmorillonite. *Clay Minerals*. 41: 853-861.
 107. Bekçi, Z., Seki, Y. and Yurdakoç, M. K. (2006). Equilibrium studies for trimethoprim adsorption on montmorillonite ksf. *Journal of Hazardous Materials*. 133: 233-242.
 108. Ötker, H. M. and Akmehmet-Balcıoğlu, I. (2005). Adsorption and degradation of enrofloxacin, a veterinary antibiotic on natural zeolite. *J. Hazard. Mater.* 122: 251-258.
 109. Vergili, I. and Barlas, H. (2009). Removal of selected pharmaceutical compounds from water by an organic polymer resin. *Journal Scientific Industrial Research*. 68: 417-425.
 110. Rossner, A., Snyder, S. A. and Knappe, D. R. U. (2009). Removal of emerging contaminants of concern by alternative adsorbents. *Water Research*. 43: 3787-3796.

111. Ribeiro, M. L. and Ribeiro, I. C. (2003). Modelling the adsorption kinetics of erythromycin onto neutral and anionic resins. *Bioprocess and Biosystems Engineering*. 26: 49-55.
112. Li, G., Li, H., Li, Y., Chen, J., Zhu, M. and Zhang, X. (2010). Adsorption of tetracycline by activated carbon fiber. *Bioinformatics and Biomedical Engineering (iCBBE), 4th International Conference*. Chengdu, China.
113. Xu, Z., Kuang, D., Liu, L. and Deng, Q. (2007). Selective adsorption of norfloxacin in aqueous media by an imprinted polymer based on hydrophobic and electrostatic interactions. *Journal of Pharmaceutical and Biomedical Analysis*. 45: 54-61.
114. Choi, K.-J., Son, H.-J. and Kim, S.-H. (2007). Ionic treatment for removal of sulfonamide and tetracycline classes of antibiotic. *Science of The Total Environment*. 387: 247-256.
115. El-Shaboury, S. R., Saleh, G. A., Mohamed, F. A. and Rageh, A. H. (2007). Analysis of cephalosporin antibiotics. *Journal of Pharmaceutical and Biomedical Analysis*. 45: 1-19.
116. Qtaitat, M. A. (2004). Study of the interaction of trimethoprim-montmorillonite by infrared spectroscopy. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 60: 673-678.
117. Parolo, M. E., Savini, M. C., Vallés, J. M., Baschini, M. T. and Avena, M. J. (2008). Tetracycline adsorption on montmorillonite: Ph and ionic strength effects. *Applied Clay Science*. 40: 179-186.
118. Turku, I., Sainio, T. and Paatero, E. (2007). Thermodynamics of tetracycline adsorption on silica. *Environmental Chemistry Letters*. 5: 225-228.
119. Lin, S. H. and Lai, C. L. (2000). Kinetic characteristics of textile wastewater ozonation in fluidized and fixed activated carbon beds. *Water Research*. 34: 763-772.
120. Nawrocki, J. and Kasprzyk-Hordern, B. (2010). The efficiency and mechanisms of catalytic ozonation. *Applied Catalysis B: Environmental*. 99: 27-42.
121. Jans, U. and Hoigné, J. (1998). Activated carbon and carbon black catalyzed transformation of aqueous ozone into oh-radicals. *Ozone Science and Engineering*. 20 67-90.

122. Beltrán, F. J., Rivas, J., Álvarez, P. and Montero-de-Espinosa, R. (2002). Kinetics of heterogeneous catalytic ozone decomposition in water on an activated carbon. *Ozone: Science & Engineering: The Journal of the International Ozone Association*. 24: 227 - 237.
123. Beltrán, F. J., Pocostales, P., Alvarez, P. and Oropesa, A. (2009). Diclofenac removal from water with ozone and activated carbon. *Journal of Hazardous Materials*. 163: 768-776.
124. Beltrán, F. J., Pocostales, P., Álvarez, P. M. and López-Piñeiro, F. (2009). Catalysts to improve the abatement of sulfamethoxazole and the resulting organic carbon in water during ozonation. *Applied Catalysis B: Environmental*. 92: 262-270.
125. Andreozzi, R., Caprio, V., Ciniglia, C., de Champdoré, M., Lo Giudice, R., Marotta, R. and Zuccato, E. (2004). Antibiotics in the environment: Occurrence in Italian streams, fate, and preliminary assessment on algal toxicity of amoxicillin. *Environmental Science & Technology*. 38: 6832-6838.
126. Ternes, T. A., Stüber, J., Herrmann, N., McDowell, D., Ried, A., Kampmann, M. and Teiser, B. (2003). Ozonation: A tool for removal of pharmaceuticals, contrast media and musk fragrances from wastewater? *Water Research*. 37: 1976-1982.
127. Garoma, T., Umamaheshwar, S. K. and Mumper, A. (2010). Removal of sulfadiazine, sulfamethizole, sulfamethoxazole, and sulfathiazole from aqueous solution by ozonation. *Chemosphere*. 79: 814-820.
128. Rodayan, A., Roy, R. and Yargeau, V. (2010). Oxidation products of sulfamethoxazole in ozonated secondary effluent. *Journal of Hazardous Materials*. 177: 237-243.
129. Beltrán, F. J., Aguinaco, A. and García-Araya, J. F. (2009). Mechanism and kinetics of sulfamethoxazole photocatalytic ozonation in water. *Water Research*. 43: 1359-1369.
130. Abellán, M. N., Bayarri, B., Giménez, J. and Costa, J. (2007). Photocatalytic degradation of sulfamethoxazole in aqueous suspension of TiO_2 . *Applied Catalysis B: Environmental*. 74: 233-241.
131. Bahdod, A., El Asri, S., Saoiabi, A., Coradin, T. and Laghizil, A. (2009). Adsorption of phenol from an aqueous solution by selected apatite

- adsorbents: Kinetic process and impact of the surface properties. *Water Research*. 43: 313-318.
132. Faria, P. C. C., Órfão, J. J. M. and Pereira, M. F. R. (2008). A novel ceria-activated carbon composite for the catalytic ozonation of carboxylic acids. *Catalysis Communications*. 9: 2121-2126.
 133. Santhy, K. and Selvapathy, P. (2006). Removal of reactive dyes from wastewater by adsorption on coir pith activated carbon. *Bioresource Technology*. 97: 1329-1336.
 134. Montgomery, D. C. (2001). Design and analysis of experiments. *New York: John Wiley & Sons*.
 135. Clarke, G. M. and Kempson, R. E. (1997). Introduction to the design and analysis of experiments. *London: Arnold*.
 136. Wu, D., Li, Y., Shi, Y., Fang, Z., Wu, D. and Chang, L. (2002). Effects of the calcination conditions on the mechanical properties of a pcom/al₂o₃ hydrotreating catalyst. *Chemical Engineering Science*. 57: 3495-3504.
 137. Cornell, J. A. (1990). How to apply response surface methodology. *Wisconsin: American Society for Quality Control*.
 138. Box, G. E. P., Hunter, W. G. and Hunter, J. S. (1978). Statistics for experimenters: An introduction to design, data analysis, and model building. *New York: John Wiley & Sons*.
 139. Al-Ghouti, M. A., Yousef, I., Ahmad, R., Ghrair, A. M. and Al-Maaitah, A. A. (2010). Characterization of diethyl ether adsorption on activated carbon using a novel adsorption refrigerator. *Chemical Engineering Journal*. 162: 234-241.
 140. Amin, N. A. S., Akhtar, J. and Rai, H. K. (2010). Screening of combined zeolite-ozone system for phenol and cod removal. *Chemical Engineering Journal*. 158: 520-527.
 141. Istadi and Amin, N. A. S. (2004). Screening of mgo- and ceo₂-based catalysts for carbon dioxide oxidative coupling of methane to c₂⁺ hydrocarbon. *Journal of Natural Gas Chemistry* 13: 23-35.
 142. Leofanti, G., Tozzola, G., Padovan, M., Petrini, G., Bordiga, S. and Zecchina, A. (1997). Catalyst characterization: Characterization techniques. *Catalysis Today*. 34: 307-327.

143. Rakness, K. L., Wert, E. C., Elovitz, M. and Mahoney, S. (2010). Operator-friendly technique and quality control considerations for indigo colorimetric measurement of ozone residual. *Ozone: Science & Engineering: The Journal of the International Ozone Association*. 32: 33 - 42.
144. Li, W., Gibbs, G. V. and Oyama, S. T. (1998). Mechanism of ozone decomposition on a manganese oxide catalyst. 1. In situ raman spectroscopy and ab initio molecular orbital calculations. *Journal of the American Chemical Society*. 120: 9041-9046.
145. Sing, K. S. W., Everett, D. H., Haul, R. A. W., Moscou, L., Pierotti, R. A., Rouquerol, J. and Siemieniewska, T. (1984). Reporting physisorption data for gas/solid systems with special reference to the determination of surface area and porosity. *Pure and Applied Chemistry*. 57: 603–619.
146. Xue, K., Chen, D. and Jiao, X. (2009). Fabrication of crystalline mesoporous metal oxides and sulfides. *Inorganic Chemistry*. 49: 1191-1197.
147. Sing, K. S. W., Everett, D. H., Haul, R. A. W., Moscou, L., Pierotti, R. A., Rouquerol, J. and Siemieniewska, T. (1985). Reporting physisorption data for gas/solid systems with special reference to the determination of surface area and porosity. *Pure Appl. Chem.* . 57: 603-619.
148. Hameed, B. H., Chin, L. H. and Rengaraj, S. (2008). Adsorption of 4-chlorophenol onto activated carbon prepared from rattan sawdust. *Desalination*. 225: 185-198.
149. Bautista-Toledo, M. I., Méndez-Díaz, J. D., Sánchez-Polo, M., Rivera-Utrilla, J. and Ferro-García, M. A. (2008). Adsorption of sodium dodecylbenzenesulfonate on activated carbons: Effects of solution chemistry and presence of bacteria. *Journal of Colloid and Interface Science*. 317: 11-17.
150. Ania, C. O. and Bandosz, T. J. (2006). Metal-loaded polystyrene-based activated carbons as dibenzothiophene removal media via reactive adsorption. *Carbon*. 44: 2404-2412.
151. Lillo-Ródenas, M. A., Cazorla-Amorós, D. and Linares-Solano, A. (2005). Behaviour of activated carbons with different pore size distributions and surface oxygen groups for benzene and toluene adsorption at low concentrations. *Carbon*. 43: 1758-1767.

152. Ngah, W. S. W. and Fatinathan, S. Adsorption characterization of pb(ii) and cu(ii) ions onto chitosan-tripolyphosphate beads: Kinetic, equilibrium and thermodynamic studies. *Journal of Environmental Management*. 91: 958-969.
153. Çalışkan, E. and Göktürk, S. (2010). Adsorption characteristics of sulfamethoxazole and metronidazole on activated carbon. *Separation Science and Technology*. 45: 244 - 255.
154. Senthilkumaar, S., Kalaamani, P., Porkodi, K., Varadarajan, P. R. and Subburaam, C. V. (2006). Adsorption of dissolved reactive red dye from aqueous phase onto activated carbon prepared from agricultural waste. *Bioresource Technology*. 97: 1618-1625.
155. Kushwaha, J. P., Srivastava, V. C. and Mall, I. D. (2010). Treatment of dairy wastewater by commercial activated carbon and bagasse fly ash: Parametric, kinetic and equilibrium modelling, disposal studies. *Bioresource Technology*. 101: 3474-3483.
156. Li, X., Hai, F. I. and Nghiem, L. D. (2011). Simultaneous activated carbon adsorption within a membrane bioreactor for an enhanced micropollutant removal. *Bioresource Technology*. 102: 5319-5324.
157. Özcan, A., Özcan, A. S., Tunali, S., Akar, T. and Kiran, I. (2005). Determination of the equilibrium, kinetic and thermodynamic parameters of adsorption of copper(ii) ions onto seeds of capsicum annum. *Journal of Hazardous Materials*. 124: 200-208.
158. Çalışkan, E. and Göktürk, S. (2010). Adsorption characteristics of sulfamethoxazole and metronidazole on activated carbon. *Separation Science and Technology*. 45: 244-255.
159. Li, L., Ye, W., Zhang, Q., Sun, F., Lu, P. and Li, X. (2009). Catalytic ozonation of dimethyl phthalate over cerium supported on activated carbon. *Journal of Hazardous Materials*. 170: 411-416.
160. González, O., Sans, C. and Esplugas, S. (2007). Sulfamethoxazole abatement by photo-fenton: Toxicity, inhibition and biodegradability assessment of intermediates. *Journal of Hazardous Materials*. 146: 459-464.
161. Legube, B. and Karpel Vel Leitner, N. (1999). Catalytic ozonation: A promising advanced oxidation technology for water treatment. *Catalysis Today*. 53: 61-72.

162. Kastner, J. R., Ganagavaram, R., Kolar, P., Teja, A. and Xu, C. (2007). Catalytic ozonation of propanal using wood fly ash and metal oxide nanoparticle impregnated carbon. *Environmental Science & Technology*. 42: 556-562.
163. Faria, P. C. C., Monteiro, D. C. M., Órfão, J. J. M. and Pereira, M. F. R. (2009). Cerium, manganese and cobalt oxides as catalysts for the ozonation of selected organic compounds. *Chemosphere*. 74: 818-824.
164. Muruganandham, M., Chen, S. H. and Wu, J. J. (2007). Evaluation of water treatment sludge as a catalyst for aqueous ozone decomposition. *Catalysis Communications*. 8: 1609-1614.
165. Villaseñor, J., Reyes, P. and Pecchi, G. (2002). Catalytic and photocatalytic ozonation of phenol on mno₂ supported catalysts. *Catalysis Today*. 76: 121-131.
166. Dong, Y., Yang, H., He, K., Song, S. and Zhang, A. (2009). [beta]-mno₂ nanowires: A novel ozonation catalyst for water treatment. *Applied Catalysis B: Environmental*. 85: 155-161.
167. Nghiem, L. D., Schäfer, A. I. and Elimelech, M. (2005). Pharmaceutical retention mechanisms by nanofiltration membranes. *Environmental Science & Technology*. 39: 7698-7705.
168. Ji, L., Shao, Y., Xu, Z., Zheng, S. and Zhu, D. (2010). Adsorption of monoaromatic compounds and pharmaceutical antibiotics on carbon nanotubes activated by koh etching. *Environmental Science & Technology*. 44: 6429-6436.
169. Ismadji, S. and Bhatia, S. K. (2001). A modified pore-filling isotherm for liquid-phase adsorption in activated carbon. *Langmuir*. 17: 1488-1498.
170. Zhu, D. and Pignatello, J. J. (2005). Characterization of aromatic compound sorptive interactions with black carbon (charcoal) assisted by graphite as a model. *Environmental Science & Technology*. 39: 2033-2041.
171. Nasuhoglu, D., Yargeau, V. and Berk, D. (2011). Photo-removal of sulfamethoxazole (smx) by photolytic and photocatalytic processes in a batch reactor under uv-c radiation ($[\lambda]_{\max} = 254 \text{ nm}$). *Journal of Hazardous Materials*. 186: 67-75.

172. Hoigné, J. and Bader, H. (1983). Rate constants of reactions of ozone with organic and inorganic compounds in water--i: Non-dissociating organic compounds. *Water Research*. 17: 173-183.
173. Álvarez, P. M., Pocostales, J. P. and Beltrán, F. J. (2011). Granular activated carbon promoted ozonation of a food-processing secondary effluent. *Journal of Hazardous Materials*. 185: 776-783.
174. Gómez-Ramos, M. d. M., Mezcuca, M., Agüera, A., Fernández-Alba, A. R., Gonzalo, S., Rodríguez, A. and Rosal, R. Chemical and toxicological evolution of the antibiotic sulfamethoxazole under ozone treatment in water solution. *Journal of Hazardous Materials*. In Press, Accepted Manuscript:
175. Ledakowicz, S. and Gonera, M. (1999). Optimisation of oxidants dose for combined chemical and biological treatment of textile wastewater. *Water Research*. 33: 2511-2516.
176. Li, K., Yediler, A., Yang, M., Schulte-Hostede, S. and Wong, M. H. (2008). Ozonation of oxytetracycline and toxicological assessment of its oxidation by-products. *Chemosphere*. 72: 473-478.
177. Istadi and Amin, N. A. S. Co-generation of synthesis gas and c2+ hydrocarbons from methane and carbon dioxide in a hybrid catalytic-plasma reactor: A review. *Fuel*. 85: 577-592.
178. Farré, M. J., Franch, M. I., Ayllón, J. A., Peral, J. and Domènech, X. (2007). Biodegradability of treated aqueous solutions of biorecalcitrant pesticides by means of photocatalytic ozonation. *Desalination*. 211: 22-33.