# DEGRADATION OF REMAZOL BLACK B BY *Bacillus cereus* STRAIN A FROM TEXTILE WASTEWATER

ZAINAB BINTI ARI

UNIVERSITI TEKNOLOGI MALAYSIA

# DEGRADATION OF REMAZOL BLACK B BY *Bacillus cereus* STRAIN A FROM TEXTILE WASTEWATER

ZAINAB ARI

A thesis submitted in fulfilment of the requirements for the award of the degree of Master of Science (Bioscience)

Faculty of Bioscience and Medical Engineering Universiti Teknologi Malaysia

MAY 2013

TO my beloveds family: Husband: Zulkefli Daud, Children: Nursyazwani, M Nazirul Amin and M Iliya Uzair

"YOU ARE MY INSPIRATION"

#### ACKNOWLEDGEMENT

#### In the Name of ALLAH the Almighty and the Merciful

First and foremost, I would like to extent my sincerest appreciation to my supervisor, Dr. Adibah Binti Yahya for her sincere guidance, help, inspiration, attention, encouragement and advices to accomplish this project. My extent appreciation goes to my co-supervisor, Assoc. Prof. Dr. Zaharah Binti Ibrahim for her invaluable assistance and continuous supports throughout this project. For both of them, their dedication had humbled me in many ways. Without their support and understanding, it is impossible for me to complete this project.

Additional thanks are due to all the academic staff of Biology Department, all laboratory staff (Pn Hjh Fatimah, Pn Radhiah, En Ruzaimi, En Hanif and EnYusman) and fellow friends especially MP2 members (Pn Khalida, Pn Norhayati, Azrimi, Nadirah, Mohd Saiful Zaini, Zaini, Nurul Izza, Norshalena, Salwa, Muhammad Faiz, Hafizan, Pamela, Satishkumar, Chung Siang Ling, Belinda, Gan Han Ming) who are always willing to lend a helping hand. I am indebted for the wonderful memories that we shared together throughout past years.

My gratitude is also extended to Kementerian Pelajaran Malaysia for giving the Master Scholarship Scheme and sponsoring the study. Special thanks also goes to Jabatan Perkhidmatan Awam for giving the study leave.

Furthermore, my utmost grateful goes to my beloveds family (Zulkefli Daud, Nursyazwani, M Nazirul Amin and M Iliya Uzair) and entire family for their support and understanding during these challenging years of study. Thank you very much.

#### ABSTRACT

Five bacterial strains designated as A, B1, B2, C and Y1 that were revived from the UTM glycerol stock were used for biodegradation study of azo dyes (SF BLACK EXA, SF RED 3BS, SFN BLUE 150 % and SF YELLOW EXF) at three different conditions; aerobic, facultative anaerobic and partial aerobic (without agitation). Screening test shows bacterium A, under facultative anaerobic condition demonstrated the highest decolourisation rate with removal rate  $(7.0\% h^{-1})$  of 0.1 gL<sup>-1</sup> SF BLACK EXA was observed. Relationship of growth and decolourization analysis shows decolourisation rate of the same azo dye by bacterium A in CDM medium under facultative anaerobic and aerobic conditions were 26.445% h<sup>-1</sup> and 24.56% h<sup>-1</sup>, respectively. The growth rate of bacterium A was found higher in aerobic culture (2.733 mgh<sup>-1</sup>) in comparison with that in facultative anaerobic culture (1.067 mgh<sup>-1</sup>). In other assessment, bacterium A was applied into optimized CDM medium (glucose (1 gL<sup>-1</sup>), NH<sub>4</sub>Cl (0.5 gL<sup>-1</sup>), K<sub>2</sub>HPO<sub>4</sub> (7 gL<sup>-1</sup>), KH<sub>2</sub>PO<sub>4</sub> (2 gL<sup>-1</sup>), MgSO<sub>4</sub>.7H<sub>2</sub>O (0.1 gL<sup>-1</sup>), CaCl<sub>2</sub> (0.02 gL<sup>-1</sup>), RBB (0.1 gL<sup>-1</sup>)), pH 6.5, incubated at 35 °C, with 10 % v/v inoculum under facultative anaerobic condition. Result showed that bacteria A successfully degraded more than 80 % RBB after 20 h incubation. Degradation of RBB molecules increased up to 90 % and 30 % COD removal before attaining stable condition within 164 h incubation. Azoreductase analysis revealed that enzyme was produced intracellularly (11.45 x 10<sup>-3</sup> Umin<sup>-1</sup>) and stabled at pH 6-8 and temperature ranged from 30 °C to 40 °C. HPLC analysis revealed that biodegradation of RBB under combined facultative anaerobic-aerobic condition produced sulfanilic acid ( $R_t = 3.988$  min) after 160 h incubation and two unknown metabolites ( $R_t = 6.071$  min and 7.480 min) after 164 h incubations. The partial 16S rRNA analysis showed that bacterium A closely related to Bacillus cereus (99 % sequence similarity). These findings suggest that this bacterium has the capability to assist in the degradation of wastewater containing azo dyes from industry of textilebased.

#### ABSTRAK

Lima strain bakteria bertanda A, B1, B2, C dan Y1 yang telah dipencilkan semula daripada stok kultur gliserol digunakan untuk mengkaji biodegradasi pewarna azo (SF BLACK EXA, SF RED 3BS, SFN BLUE 150 % dan SF YELLOW EXF) dalam tiga keadaan berbeza: aerobik, fakultatif anaerobik dan separa aerobik (tanpa goncangan). Kadar penyingkiran warna tertinggi daripada ujian penyaringan bakteria A di bawah keadaan fakultatif telah diperhatikan sebanyak 7.0% j<sup>-1</sup> dalam medium yang mengandungi 0.1 gL<sup>-1</sup> SF BLACK EXA. Analsis perkaitan antara pertumbuhan dan penyahwarnaan menunjukkan kadar penyahwarnaan pewarna azo tersebut oleh bakteria A dalam medium CDM di bawah keadaan fakultatif anaerobik dan aerobik adalah 26.445% j<sup>-1</sup> dan 24.56% j<sup>-1</sup>, masing-masing. Kadar pertumbuhan bakteria dalam keadaan aerobik (2.733 mgj<sup>-1</sup>) didapati lebih tinggi berbanding keadaan fakultatif anaerobik (1.067 mgj<sup>-1</sup>). Dalam kajian yang lain, bakteria A diaplikasikan dalam keadaan CDM optimum (glukosa (1 gL<sup>-1</sup>), NH<sub>4</sub>Cl (0.5 gL<sup>-1</sup>), K<sub>2</sub>HPO<sub>4</sub> (7 gL<sup>-1</sup>), KH<sub>2</sub>PO<sub>4</sub> (2 gL<sup>-1</sup>), MgSO<sub>4</sub>.7H<sub>2</sub>O (0.1 gL<sup>-1</sup>), CaCl<sub>2</sub> (0.02 gL<sup>-1</sup>), RBB (0.1 gL<sup>-1</sup>), pH 6.5, suhu eraman 35°C, 10% i/i inoculum) di bawah keadaan fakultatif anaerobik. Keputusan menunjukkan bakteria A berjaya menyingkirkan lebih daripada 80 % RBB dalam tempoh 20 jam pengeraman. Degradasi molekul RBB meningkat sehingga 90 % dan 30 % penyingkiran COD sebelum mencapai kestabilan dalam tempoh 164 jam pengeraman. Analisis azoreduktase mendapati enzim dihasilkan adalah intraselular (11.45 x 10<sup>-3</sup> Umin<sup>-1</sup>) dan stabil pada pH 6-8 serta dalam julat suhu dari 30 °C hingga 40 °C. Analisis HPLC menunjukkan biodegradasi pewarna azo RBB dalam fakultatif anaerobik-aerobik menghasilkan asid sulfanilik, ( $R_t$  = 3.988 min) setelah 160 jam pengeraman dan dua jenis metabolit tidak diketahui, (Rt = 6.071 min dan 7.480 min) setelah 164 jam pengeraman. Analisis separa jujukan 16S rRNA menunjukkan bakteria A berkait rapat dengan Bacillus cereus (99% kesamaan jujukan). Penemuan ini mencadangkan bahawa bakteria ini berkeupayaan membantu degradasi air sisa yang mengandungi pewarna azo dari industri tekstil.

# **TABLE OF CONTENTS**

CHAPTER	TITLE			PAGE
	THESIS TITLE			i
	DEC	LARAT	TION	ii
	DED	ICATIO	DN	iii
	ACK	NOWL	EDGEMENTS	iv
	ABS	ГRACT		V
	ABS	ГRAK		vi
	TAB	LE OF	CONTENTS	vii
	LIST	OF TA	BLES	xii
	LIST	OF FI	GURES	xiii
	LIST	OF AB	BREVIATIONS	XV
	LIST	OF AP	PENDICES	xviii
1	INTF	RODUC	TION	1
	1.1	Backg	ground Study	1
	1.2	Scope	and Objectives of study	3
2	LITE	RATU	RE REVIEW	4
	2.1	Introd	uction	4
	2.2	Dyes	and Pigments	5
		2.2.1	History of Dyes	5
		2.2.2	The basic of Colour in Dyes	6
		2.2.3	Classification of Dyes	7
			2.2.3.1 Azo dyes	7
			2.2.3.2 Anthraquinone Dyes	9
	2.3 Colour Removal			10

	2.3.1	Measurement of Colour Removal	10
	2.3.2	Mechanisms of Colour Removal	10
	2.3.3	Factors Affecting Colour Removal	11
		2.3.3.1 Oxygen	12
		2.3.3.2 Temperature	12
		2.3.3.3 pH Value	13
		2.3.3.4 Dye concentration	13
		2.3.3.5 Dye structure	13
		2.3.3.6 Electron donor	14
		2.3.3.7 Redox potential	14
		2.3.3.8 Redox mediator	15
2.4	Waste	water Treatment Containing Dye	15
	2.4.1	Bacterial Decolourization and Biodegradation	16
		2.4.1.1 Anaerobic System	21
		2.4.1.2 Aerobic System	22
		2.4.1.3 Combined Anaerobic-Aerobic	
		Biodegradation	23
	2.4.2	Others microorganism in biodegradation	24
2.5	Azore	ductase as Dye degrading enzyme	25
2.6	Concl	uding Remarks	25
GEN	ERAL N	MATERIALS AND METHODS	27
3.1	Growt	th Medium Preparation	27
	3.1.1	Nutrient agar	27
	3.1.2	Nutrient Broth	27
3.2	Synthe	etic Wastewater Medium: Chemical Defined	
	Mediu	Im	28
3.3	Cultur	re Preparation	28
	3.3.1	Growing of Microorganisms	28
	3.3.2	Single Culture Preparation	29
	3.3.3	Preparation of Inoculum and Maintenance	
		of Bacteria	30
3.4	Growt	th Curve of Bacteria strain A	30
3.5	Decolourization of Azo Dyes		

3

3.6	Chem	icals and DNA Kits	31		
3.7	Gener	General Outline of Research Methodology			
SCR	EENIN	G AND CHARACTERIZATION OF DYE			
DEG	RADE	R BACTERIA FROM TEXTILE			
WAS	STEWA	TER	33		
4.1	Introd	luction	33		
4.2	Mater	ials and Methods	34		
	4.2.1	Microorganisms	34		
	4.2.2	Media and Reagents Preparation	34		
	4.2.3	Inoculum Preparation and Culture			
		Maintenance	35		
	4.2.4	Screening of Potential Azo Dye-Discoloring			
		Bacteria	35		
	4.2.5	Optimization of Azo Dye SF BLACK EXA			
		Decolourization	36		
	4.2.6	Relationship of Growth and Decolourization	37		
	4.2.7	Analytical Methods	37		
		4.2.7.1 Determination of Azo Dye			
		Decolourization	37		
		4.2.7.2 Determination of Bacterial Growth	38		
	4.2.8	Identification of Selected Bacteria using			
		16S rDNA	38		
		4.2.8.1 DNA Extraction	38		
		4.2.8.2 Agarosegal Electrophoresis	38		
		4.2.8.3 Polymerase Chain Reaction (PCR)			
		Procedure	39		
		4.2.8.4 Purification of PCR Product	41		
		4.2.8.5 Sequencing of the 16S rRNA Gene	41		
		4.2.8.6 Analysis of 16S rRNA Gene and			
		Phylogenetic Tree Construction	42		
4.3	Resul	t and Discussion	42		
	4.3.1	Screening of Bacteria for Azo Dye			
		Decolourization	42		

	4.3.2	Relationship of Growth and Decolourization	46
	4.3.3	Optimization of SF BLACK EXA	
		Decolourization	48
	4.3.4	Identification of Selected Strain of Bacterium A	55
		4.3.4.1Sequencing of the 16S rRNA Gene	55
		4.3.4.2 Sequencing Analysis of Gene	
		Encoding for the 16S rRNA	56
4.8	Concl	usion	59
LOC	ALIZA	TION OF AZOREDUCTASE AND	
DEG	RADA	TION ANALYSIS OF REMAZOL BLACK B	
(RBE	B) BY <i>B</i> (	acillus cereus STRAIN A	60
5.1	Introd	uction	60
5.2	Mater	ials and Methods	62
	5.2.1	Localization and Detection of Enzymatic	
		Activities	62
		5.2.1.1 Preparation of Bacterial Cell Fraction	62
		5.2.1.2 Azoreductase Assay	63
		5.2.1.3 Effects of pH value and Temperature	
		on theAzoreductase Activity and	

Х

63

64

64

65

65

65

66

66

5

5.3

5.2.1.2 Azoreductase Assay
5.2.1.3 Effects of pH value and Temperature on theAzoreductase Activity and Stability
5.2.2 Preparation of Inoculums and Biodegradation of Remazol Black B
5.2.2.1 Determination of CDW, ORP, pH, COD and Colour Removal
5.2.3 Product Detection and Determination using HPLC
5.2.3.1 Sample Preparation
5.2.3.2 Chromatographic Apparatus
Result and Discussion
5.3.1 Localization of Azoreductase Produced by Bacterium A
5.3.2 Optimization of Azoreductase Assay

5.3.2Optimization of Azoreductase Assay675.3.2.1 Effect of pH67

		5.3.2.2 Effect of Temperature	68
		5.3.2.3 Effect of Incubation Time	69
		5.3.2.4 Effect of NADH	70
		5.3.2.5 Effect of FAD	72
		5.3.2.6 Effect of Dye Concentration	73
	5.3.3	Decolourization and Degradation of	
		Remazol Black B by Bacillus cereus Strain A	72
		5.3.3.1 Time Course of RBB Degradation	
		in Batch Culture	74
		5.3.3.2 Kinetic Study on RBB Decolourisation	
		by Bacillus cereus Strain A	75
		5.3.3.3 Correlation between Specific	
		Decolourization Rate and Cell	
		Dry Weight	77
		5.3.3.4 Correlation between Colour Removal,	
		COD and pH value	78
	5.3.4	Product Detection and Determination using	
		HPLC under Facultative Anaerobic-Aerobic	
		Condition	79
		5.3.4.1 Product Degradation Detection Using	
		Reverse phase-HPLC	79
5.4	Concl	usion	83
CON	CLUSI	ON AND SUGGESTIONS	84
6.1	Concl	usion	84
6.2	Sugge	estion for Future Work	86
REF	ERENC	CES	87

APPENDICES 107

# LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Region of the electromagnetic spectrum	
	and relationship between wavelength and color	6
2.2	Biodegradation of dyes using single bacterial strain	17
3.1	Component of Chemically Defined Medium	28
4.1	Component mixes for electrophoresis	39
4.2	2X PCR Master Mix compositions	39
4.3	The universal primers used for the amplification of	
	16S rRNA	40
4.4	Components for PCR reaction	40
4.5	Thermal Profile for PCR Reaction	41
4.6	Decolourisation rate of azo dye under three different	
	conditions: aerobic with shaking, partial aerobic (without	
	shaking) and facultative anaerobic. The effect of azo dye	
	decolourisation by all bacteria was tested using four	
	different types of azo dyes: SF RED 3BS, SFN BLUE	
	150%, SF BLACK EXA and SF YELLOW EXF.	43
5.1	Azoreductase activity determined from different fractions	
	of cells assayed under anaerobic and aerobic condition	66

# LIST OF FIGURES

FIGURE NO	. TITLE	PAGE
2.1	Example of dye-auxochromes and dye-chromophores	
	of azo dyes	9
2.2	General overview of the fate of azo dyes and aromatic	
	amines during anaerobic-aerobic treatment	23
3.1	A general outline of research methodology	32
4.2	Time course of the bacterial growth and	
	decolourisation under (a) facultative anaerobic	
	and (b) aerobic conditions in the medium adjusted	
	to pH 7, incubated at 37 °C.	47
4.3	Decolourization of SF BLACK EXA by bacterium	
	A in CDM at 37 °C under facultative anaerobic	
	condition at different pH values.	49
4.4	Decolourization of SF BLACK EXA by bacterium	
	A in CDM at 37°C after 7 days incubation under	
	facultative anaerobic condition at different	
	SF BLACK EXA concentration.	51
4.5	Decolourization of SF BLACK EXA by bacterium	
	A in CDM at 37°C after 14 days incubation under	
	facultative anaerobic condition using different	
	nitrogen sources at varying concentrations.	52
4.6	Decolourization of SF BLACK EXA by bacterium	
	A in CDM at 37 °C after 7 days incubation under	
	facultative anaerobic condition using different	
	carbon sources at varying concentrations.	53

4.7	Decolourization of SF BLACK EXA by bacterium	
	A in CDM at 37 °C after 7 days incubation under	
	facultative anaerobic condition using different	
	inoculum sizes.	54
4.8	The 1.5 kb of PCR product of 16S rRNA fragment	
	obtained via PCR amplification.	55
4.9	Phylogram show phylogenetic relationships of	
	bacterium A based on partial 16S rRNA sequences	
	retrieved from raw textile wastewater.	57
5.1	Effect of pH value on azoreductase activity.	68
5.2	Effect of temperature on azoreductase activity.	69
5.3	Effect of incubation time on azoreductase activity.	70
5.4	Effect of NADH concentration on azoreductase activity.	71
5.5	Effect of FAD concentration on azoreductase activity.	73
5.6	Effect of dye concentration on azoreductase activity.	74
5.7	Profile of decolourization rate, COD removal rate and	
	cell dry weight by Bacillus cereus Strain A against time.	75
5.8	Kinetic study on decolourisation of Remazol Black B	
	by Bacillus cereus Strain A in CDM at 35 °C	
	under facultative anaerobic condition.	76
5.9	Correlation between decolourization rate and cell dry	
	weight.	77
5.10	Time course of RBB biodegradation by Bacillus cereus	
	Strain A in CDM under facultative anaerobic-aerobic	
	condition at 35 °C.	79
5.11	The HPLC chromatogram of combined facultative	
	anaerobic-aerobic treatment.	81
5.12	Chromatogram of sulfanilic acid standard in HPLC	
	analysis.	82

# LIST OF ABBREVIATIONS

°C	-	Degree Celsius
%	-	Percentage
$\% h^{-1}$	-	percentage per hour
λ	-	Wavelength
ABS	-	Aminobenzene sulphate
3-ABS	-	3-Aminobenzene sulphate
4-ABS	-	4-aminobenzene sulphate
5-ABS	-	5-aminobenzene sulphate
ADMI	-	American Dye Manufacturer Institute
АРНА	-	American Public Health Association
4-AP	-	4-Aminophenol
ANS	-	Aminonaphythyl sulphate
BC	-	Before Century
BOD	-	Biological Oxygen Demand
C.I	-	Colour Index
CaCl <sub>2</sub>	-	calcium chloride
CDM	-	Chemically Defined Medium
CDW	-	cell dry weight
Cl	-	chloride
Cl <sup>-1</sup>	-	ion chloride
$CO_2$	-	Carbon dioxide
COD	-	chemical oxygen demand
$\mathbf{d}^{\cdot_1}$	-	per day
dATP	-	deoxyadenosine 5'-triphosphate
dCTP	-	deoxycytosine 5' triphosphate
dGTP	-	deoxyguanosine 5' triphosphate
DNA	-	deoxyribonucleic acid

dNTP	-	deoxynucleotide triphosphate
dTTP	-	deoxythymidine 5'-triphosphate
EDTA	-	ethylene diamine tetra acetic acid
Et Br	-	Ethidium bromide
FAD	-	flavin adenine dinucleotide(oxidized)
$\operatorname{FAD}^+$	-	ion flavin adenine dinucleotide
FADH <sub>2</sub>	-	flavin adenine dinucleotide(reduced)
Fe <sup>2+</sup>	-	ion ferum
FMNH <sub>2</sub>	-	flavin adenine mononucleotide (reduced)
g	-	gram
gL-1	-	gram per litre
h	-	hour
$\mathbf{h}^{-1}$	-	per hour
H <sub>2</sub> O	-	hydrogen dioxide
HPLC	-	high performance liquid chromatography
K2HPO4	-	dipotassium hydrogen phosphate
kb	-	kilobase
KH2PO4	-	potassium dihydrogen phosphate
kg	-	kilogram
mg	-	milligram
${\sf mgh}^{-1}$	-	milligram per hour
min	-	minute
mol	-	mole
mgL-1	-	milligram per litre
MgCl <sub>2</sub>	-	magnesium chloride
MgSO4.7H2O	-	magnesium sulphate heptahydrate
mL	-	milliliter
mm	-	millimeter
mM	-	milimol
mV	-	miliVolt
NA	-	nutrient agar
$\mathbf{NAD}^{\scriptscriptstyle +}$	-	nicotinamide adenine dinucleotide(oxidized)
NADH	-	nicotinamide adenine dinucleotide(reduced)
NADP	-	nicotinamide adenine dinucleotide phosphate

NADPH	-	nicotinamide adenine dinucleotide phosphate(reduced)
NB	-	nutrient broth
NH4Cl	-	ammonium chloride
(NH4) <sub>2</sub> SO4	-	ammonium sulphate
ngmL <sup>-1</sup>	-	nanogram per liter
nm	-	nanometer
NO <sub>3-</sub>	-	nitrate
OD600nm	-	optical density at 600nm
PAAB	-	p-Aminoazobenzene
PCR	-	polymerase chain reaction
pН	-	potential ion hydrogen
ppm	-	part per million
Pt-Co	-	platinum cobalt
RBB	-	Remazol Black B
RM	-	redox mediator
rpm	-	rotation per minute
rRNA	-	ribosomal RNA
<b>SO</b> 4 <sup>2-</sup>	-	sulphate
TAE	-	tris-acetate buffer
Tris	-	2-hydroxymethyl-2-methyl-1,3-propanediol
TSS	-	total suspended solid
U	-	enzyme unit
UV	-	ultraviolet
UV-vis	-	ultraviolet-visible
<i>v/v</i>	-	volume per volume
w/v	-	weight per volume
µgmL⁻¹	-	microgram per milliliter
μL	-	microliter
рМ	-	picomolar

## xviii

# LIST OF APPENDICES

# APPENDIX

## TITLE

# PAGE

А	Full sequences of 16S rDNA for Bacillus cereus	108
В	Alignment score of full sequence of bacterium A	
	obtained using BLASTn	109
С	Standard Curve of Remazol Black B concentration	
	at $\lambda_{\text{max}} = 597$ nm	113
D	Correlation between OD 600nm and cell dry	
	weight of Bacillus sp strain A	114
E	Standard Methods for the Examination of Water	
	and Wastewater	115
F	Reversed phase-HPLC analytical parameters	118
G	Ln Cell Dry Weight $\mu 1$ and $\mu 2$ in Facultative	
	Anaerobic and Aerobic Conditions	119

#### **CHAPTER 1**

#### **INTRODUCTION**

#### **1.1 Background Study**

Synthetic dyes are one of the main wastewater pollutants. It is estimated about  $10^9$  kg of dyes are produced annually in the world, which azo dyes represent about 70% by weight (Zollinger, 1987). Azo dyes characterized by one or more azo groups (-N=N-) (Selvam *et al.*, 2003) are considered as the most important group of synthetic colorants used in textile, printing, pharmaceutical, cosmetic and food industries (Beydilli and Pavlostathis, 2005). It's generally xenobiotic compounds that are very recalcitrant against biodegradation processes (Van der Zee, 2002). Azo dyes are designed to resist chemical and microbial attacks (Ramalho *et al.*, 2002) and to be stable in light during material washing (Rajaguru *et al.*, 2000).

As azo dyes are widely used in textile industry, its wastewater are characterised by their highly visible colour with range of 10-200 mgL<sup>1</sup> (O'Neill *et al.*, 1999), high chemical oxygen demand (COD), suspended solids and alkaline pH (9-11) (Manu and Chaudhari, 2002). According to O'Neil (1999), textile processing wastewater typically containing dye in a range of 10-200 mgL<sup>-1</sup> due to its disability to bind to cloth during the dyeing process. Reish (1996) also reported that over 10% of the dye used in textile processing does not bind to the fibres and is therefore released to environment. Fixation efficiency for basic dyes and reactive dyes was reported to vary around 98% and 50%, respectively (O'Neil, 1999). Thus, coloured effluent that was released into the environment is undesirable, not only because of its

colour, but also many dyes and their breakdown products are toxic and/or mutagenic to living things (Chung and Cerniglia, 1992). Without adequate treatment, these dyes are stable and can remain in the environment for an extended period of time. For example the half-life of hydrolysed Reactive Blue 19 is about 46 years at pH 7 at 25°C (Hao *et al.*, 2000).

Several physicochemical techniques, such as carbon absorption, ozonation, coagulation/adsorption that have been used for treating wastewater-containing dyes (Somasiri *et al.*, 2006), are in general inefficient, costly and limited applicability, while sometimes producing large amount of difficult to disposed toxic waste (Sanayei *et al.*, 2010). Recent researchers focus on bacteria as produce nontoxic waste (Somasiri *et al.*, 2006), cheep and effective dyes degrader using anaerobic-aerobic treatment system (Dos Santos *et al.*, 2005). Azo dyes are reduced by the cleavage of the azo bond in anaerobic system to formed carcinogenic aromatic amines which need to be further degraded (Haugh *et al.*, 1991; Coughlin *et al.*, 1997). Most azo dyes are reduced anaerobically to the corresponding amines but they are difficult to degrade aerobically (Zimmerman *et al.*, 1982; Banat *et al.*, 1996). Under aerobic conditions, bacteria are able to mineralise some aromatic amines (Stolz, 2001; Pearce *et al.*, 2003). Rajaguru *et al.*, (2000) revealed that the performance of sequential anaerobic aerobic bacterial degradation system has been shown to be efficient in the degradation of azo dyes.

The effectiveness of microbial decolourisation depends on the adaptability and the activity of selected microorganisms (Chen *et al.*, 2003). Kodam *et al.*, (2005) found that KMK 48 bacterium able to decolourize reactive azo dyes under aerobic conditions with high effectiveness. To date, the ability of microorganisms to carry out dye decolourisation has received much attention. Microbial decolourisation and degradation of dyes is seen as a cost effective method for removing these pollutants from the environment. Therefore in this study, specialised strains of anaerobic aerobic bacteria would be acclimatized its ability to reduce the azo groups of azo dyes to non-colored intermediates and/or even to partially mineralize them, which are safe and less toxic to the environment.

### 1.2 Scope and Objectives of Study

This study was focused on the optimization of azo dye degradation by selected bacteria locally isolated from acclimatized bacterial consortium used in textile wastewater treatment. Five bacterial strains coded A, B1, B2, C and Y1 were initially acclimatized in chemically defined medium containing pure azo dye. This is to ensure that the bacteria have consistent growth and activity on the dye used. The bacteria were then screened based on their ability to decolorize selected type of pure dye provided by the textile industry. Optimization is carried out under facultative anaerobic condition as the factors of temperature, types and concentrations of carbon and nitrogen sources, inoculums size, rate of agitation and dye concentration. The intermediates of azo dye biodegradation will be determined by carrying out the experiment under sequential anaerobic-aerobic condition.

This study is carried out with the specific objectives:

- i. To screen and characterize potential dye degrading bacteria using four different azo dyes in chemical defined medium (CDM).
- ii. To optimize physical and chemical condition for azo dye decolonization by selected bacteria in chemical defined medium.
- iii. To determine enzyme localization and azoreductase activity for RemazolBlack B color removal using selected bacteria.
- iv. To analyze biodegradation products of Remazol Black B and its intermediate in synthetic medium using selected bacteria.

#### REFERENCES

- Adedayo, O., Javadpour, S., Taylor, C., Anderson, W.A. and Moo-Young, M.
  (2004). Decolourization and Detoxification of Methyl Red by Aerobic Bacteria from a Wastewater Treatment Plant. *World Journal of Microbiology and Biotechnology*. 20, 545-550.
- Albuquerque, M. G. E., Lopes, A. T., Serralheiro, M. L., Novais, J. M. and Pinheiro,
  H. M. (2005). Biological Sulphate Reduction and Redox Mediator Effects on
  Azo Dye Decolorization in Anaerobic-Aerobic Sequencing Batch Reactors. *Enzyme and Microbiol Technology*. 36, 790-799.
- American Public Health Association (2005). Standard Methods for the Examination of Water and Wastewater. (21<sup>th</sup> ed). Washington, DC, USA American Public Health Association.
- An, S. Y., Min, S. K., Cha, I. H., Choi, Y. L., Cho, Y. S., Kim, C. H. and Lee, Y. C. (2002). Decolorization of Triphenylmethane and Azo Dyes by *Citrobacter* sp. *Biotechnology Letters*. 24, 1037-1040.
- Asad, S., Amoozegar, M. A., Pourbabaee, A. A., Sarbolouki, M. N. and Dastgheib,
  S. M. M. (2006). Decolorization of Textile Azo Dyes by Newly Isolated Halophilic and Halotolerant Bacteria. *Bioresource Technology*. 15, 21-34.
- Bafana, A., Chakrabarti, T. and Devi, S. S. (2008). Azoreductase and Dye Detoxification Activities of *Bacillus Velezensis* Strain AB. *Applied Microbiology and Biotechnology*. 77, 1139-1144.

- Banat, I. M., Nigam, P., Singh, H. D. and Marchant, R. (1996). Microbial Decolorization of Textile-Dyecontaining Effluents: A Review. *Bioresource Technology*. 58, 217-227.
- Baughman, G. L. and Weber, E. J. (1994). Transformation of dyes and related compounds in anoxic sediment: Kinetics and products. *Environmental Science and Technology*. 28, 267-276.
- Beydilli, M. I. and Pavlostathis, S. G. (2005). Decolorization Kinetics of the Azo Dye Reactive Red 2 under Methanogenic Conditions: Effect of Long-Term Culture Acclimation. *Biodegradation*. 16, 135-146.
- Blumel, S., Contzen, M., Lutz, M., Stolz, A. and Knackmuss, H. J. (1998). Isolation of a Bacterial Strain with the Ability to Utilize the Sulfonated Azo Compound 4-Carboxy-4'-Sulfoazobenzene as the Sole Source of Carbon and Energy. *Applied and Environmental Microbiology*. 64(6), 2315-2317.
- Bragger, J. L., Lloyd, A. W., Soozandehfar, S. H., Bloomfield, S. F., Marriott, C. and Martin, G. P. (1997). Investigations into the Azo Reducing Activity of a Common Colonic Microorganism. *International Journal of Pharmaceutics*. 157, 61-71.
- Brar, S. K., Verma, M., Tyagi, R. D. and Valero, J. R. (2006). Recent advances in downstream processing and formulations of *Bacillus thuringiensis* based biopesticides. *Process Biochemistry*. 41(2), 323-342.
- Bras, R., Ferra, M. I. A., Pinheiro, H. M. and Goncalves, I. C. (2001). Batch Tests For Assessing Decolourisation Of Azo Dyes By Methanogenic And Mixed Cultures. *Journal of Biotechnology*. 89, 155-162.
- Bromley-Challenor, K. C. A., Knappi, J. S., Zang, Z., Gray, N. C. C., Hetheridge, M. J., and Evans, M. R. (2000). Decolorization of an Azo Dye by Unacclimated Activated Sludge under Anaerobic Conditions. *Water Research*. 34(18), 4410-4418.

- Brown, D. and Laboureur, P. (1983). The aerobic biodegradability of primary aromatic amines. *Chemosphere*. 12: 405-414.
- Bumpus, J. A. (1995). Microbial degradation of azo dyes. Progress in Industrial Microbiology. 32, 157–176.
- Byrns, G. (2001). The Fate of Xenobiotic Organic Compounds in Wastewater Treatment Plants. *Water Research*. 35(10), 2523-2533.
- Carliell, C. M., Barclay, S. J., Naidoo, N., Buckley, C. A., Mulholland, D. A. and Senior, E. (1994). Anaerobic decolorisation of reactive dyes in conventional sewage treatment processes. *Water SA*. 20:341-344
- Chagas, E. P. and Durrant, L. R. (2001). Decolourization of azo dyes by Phanerochaete chrysosporium and Pleurotus sajorcaju. Enzyme and Microbial Technology. 29, 473-477.
- Chan, G. F. (2004). Studies on the decolourization mechanism of azo dye by Citrobacter freundii A1 from the molecular and enzymatic aspects, PhD Doctoral Thesis, Universiti Teknologi Malaysia, Skudai, Johor.
- Chang, J. S. and Lin, Y. C. (2000). Fed-batch Bioreactor Strategies for Microbial Decolorization of Azo dye using a *Pseudomonas luteola* Strain. *Biotechnology Progress*. 16, 979-985.
- Chang, J. S., Chou, C., and Chen, S. Y. (2001). Decolorization of Azo Dyes with Immobilized *Pseudomonas luteola*. *Process Biochemistry*. 36, 757-763.
- Chatterjee, S., Chatterjee, S., Chatterjee, B. P. and Guha, A. K. (2006). Adsorptive
  Removal of Congo Red, A Carcinogenic Textile Dye by Chitosan
  Hydrobeads: Binding Mechanism, Equilibrium and Kinetics, Colloids and
  Surfaces. A Physicochemical and Engineering Aspects. 6, 33-45.

- Chen, K. C., Wua, J. Y., Liou, D. J., Hwang, S. C. J. (2003) Decolorization of the Textile Dyes by Newly Isolated Bacterial Strains. *Journal of Biotechnology*. 101, 57-68.
- Chinwetkitvanich, S., Tuntoolvest, M. and Panswad, T. (2000). Anaerobic Decolorization of Reactive Dyebath Effluents by a Two-Stage with Tapioca as a Co-Substrate. *Water Research*. 34, 2223-2232.
- Christie, R. (2001). Colour Chemistry. *The Royal Society of Chemistry*, Cambridge, United Kingdom.
- Clarke, E. A. and Anliker, R. (1980). In Zollinger, H. (ed): Color Chemistry: Syntheses, properties and application of organic dyes and pigments. 2<sup>nd</sup> Rev. Edition.Weinheim; New York; Basel; Chambridge. 496pp.
- Coughlin, M. F., Kinkle, B. K., Tepper, A., Bishop, P. L. (1997). Characterization of aerobic azo dye-degrading bacteria and their activity in biofilms. *Water Science and Technology*. 36, 215-220.
- Cripps, C., Bumpus, J. A. and Aust, S. D. (1990). Biodegradation of Azo and Heterocyclic Dyes by *Phanerochaete chrysosporium*. Applied and Environmental Microbiology. 56(4), 1114-1118.
- Cruz, A. and Buitron, G. (2001). Biodegration of Disperse Blue 79 Using Sequenced Anaerobic/Aerobic Biofilters. *Water Science and Technology*. 44, 159-166.
- Dafale, N., Wate, S., Meshram, S. and Nandy, T. (2008). Kinetic Study Approach of Remazol Black-B Use for the Development of Two-Stage Anoxic-Oxic Reactor for Decolorization/Biodegradation of Azo Dyes by Activated Bacterial Consortium. *Journal of Hazardous Materials*. 159, 319-328.
- De Souza, S. M. A. G. U., Forgiarini, E. and de Souza, A. A. U. (2007). Toxicity of Textile Dyes and their Degradation by the Enzyme Horseradish Peroxidase (HRP) *Journal of Hazardous Materials*. 147, 1073-1078.

- Delee, W., O'Neill, C., Hawkes, F. R. and Pinheiro, H. M. (1998). Anaerobic Treatment of Textile Effluents: A Review. *Journal of Chemical Technology* and Biotechnology. 73, 323-335.
- Di´az, E., Ferra´ndez, A., Prieto, M. A. and Garci´a, J. L. (2001). Biodegradation of Aromatic Compounds by *Escherichia coli*. *Microbiology and Molecular Biology Reviews*. 65(4), 523-569.
- Dos Santos, A.B., Cervantes, F.J. and van Lier, J.B. (2004). Azo Dye Reduction by Thermophilic Anaerobic Granular Sludge and the Impact of the Redox Mediator AQDS on the Reductive Biochemical Transformation. *Applied Microbiology and Biotechnology*. 64, 62-69.
- Dos Santos, A. B., Bisschops, I. A. E., Cervantes, F. J., Van Lier, J. B. (2005). The Transformation and Toxicity of Anthraquinone Dyes during Thermophilic (55°C) and Mesophilic (30°C) Anaerobic Treatments. *Journal of Biotechnology*. 15, 345-353.
- Dos Santos, A. B., Cervantes, F. J. and van Lier, J. B. (2007). Review Paper on Current Technologies for Decolourisation of Textile Wastewaters: Perspectives for Anaerobic Biotechnology. *Bioresource Technology*. 98, 2369-2385.
- Ekici, P., Leupold, G. and Parlar, H. (2001). Degradability of selected azo dye metabolits in activated sludge systems. *Chemosphere*. 44: 721-728.
- Enayatzamir, K., Alikhani, H. A. and Couto, S. R. (2009). Simultaneous Production of Laccase and Decolouration of the Diazo Dye Reactive Black 5 in a Fixed-Bed Bioreactor. *Journal of Hazardous Materials*. 164, 296-300.
- Field, J. A., Stams, A. J. M, Kato, M. and Schraa, G. (1995). Enhanced biodegradation of aromatic pollutant in co-culture of anaerobic and aerobic bacterial consortia. *Antonie van Leeuwenhoek*. 67, 47-77.

- Fontenot, E. J., Lee, Y. H., Matthews, R. D., Zhu, G. and Pavlostathis, S. G. (2003). Reductive decolorization of a textile reactive dyebath under methanogenic conditions. *Applied Biochemistry and Biotechnology*. 109, 207-225.
- Forgacs, E., Cserha´ti, T. and Oros, G. (2004). Removal of Synthetic Dyes from Wastewaters: A Review. *Environment International*. 30, 953-971.
- Fu, Y. and Viraraghavan, T. (2001). Fungal Decolorization of Dye Wastewater: A Review. *Bioresource Technology*. 79, 251-262.
- Gheewala, S. H. and Annachhatre, A. P. (1997). Biodegradation of aniline. *Water Science and Technology*. 36(10), 53-63.
- Gingell, R. and Walker, R. (1971). Mechanisms of Azo Reduction by Streptococcus faecalis. II. The Role of Soluble Flavins. Xenobiotica. 1(3), 231-239.
- Goncalves, I., Gomes, A., Brás, R., Ferra, M. I. A., Amorim, M. T. P. and Porter, R.
  S. (2000). Biological treatment of effluent containing textile dyes. *Journal of The Society of Dyers and Colourists*. 6, 393-397.
- Gordon, R. E., Haynes, W. C. and Pang, C. H. (1973). The Genus Bacillus. Agriculture Handbook no 427 Washington, DC: US Department of Agriculture.
- Gupta, V. K. and Suhas. (2009). Application of Low-cost Adsorbents for Dye Removal: A Review. *Journal of Environmental Management*. 12, 1-30.
- Hao, O. J. and Chang, P. C. (2000). Decolorization of Wastewater. *Critical Reviews* in Environmental Science and Technology. 30, 449-505.
- Haug, W., Schmidt, A., Nortemann, B., Hempel, D. C., Stolz, A. and Knackmuss H.
  J. (1991). Mineralization of the Sulfonated Azo Dye Mordant Yellow 3 by a 6-Aminonaphthalene-2-Sulfonate-Degrading Bacterial Consortium. *Applied Environmental Microbiology*. 57(11), 3144-3149.

- Heyndrickx, M., Vandemeulebroecke, K., Scheldeman, P., Kersters, K., De Vos, P., Logan, N.A., Aziz, A.M., Ali, N. and Berkeley, R.C.W. (1996). A polyphasic reassessment of the genus *Paenibacillus*, reclassification of *Bacillus lautus* (Nakamura, 1984) as *Paenibacillus lautus* comb. nov. and of *Bacillus peoriae* (Montefusco *et al.*, 1993) as *Paenibacillus peoriae comb. nov.*, and emended descriptions of *P. lautus* and of *P. peoriae*, *International Journal of Systematic Bacteriology*. 46 (4), 988-1003.
- Holme, I. (1984). In Zollinger, H. (ed): Color Chemistry: Syntheses, properties and application of organic dyes and pigments. 2<sup>nd</sup> Rev. Edition.Weinheim; New York; Basel; Chambridge. 496pp.
- Horitsu, H., Takada, M., Idaka, E., Tomoyeda, M. and Ogawa, T. (1977).
  Degradation of p-Aminoazobenzene by *Bacillus subtilis*. *European Journal* of Applied Microbiology. 4, 217-224.
- Hsueh, C.C. and Chen, B.Y. (2007). Comparative Study on Reaction Selectivity of Azo Dye Decolorization by *Pseudomonas luteola*. *Journal of Hazardous Materials*. 141, 842-849.

http://www.straw.com/sig/dyehist.html (18.2.2010 11.04am)

- Hu, T.L. (1998). Degradation of azo dye RP2B by *Pseudomonas luteola*. Water Science and Technology. 38, 299-306.
- Hu, T. L. (2001). Kinetics of Azoreductase and Assessment of Toxicity of Metabolic Products from Azo dyes by *Pseudomonas luteola*. *Water Science and Technology*. 43(2), 261-9.
- Idaka, E., Ogawa, T., Horitsu, H. and Tomoyeda, M. (1978). Degradation of Azo Compounds by Aeromonas hydrophilia var. 24B. Journal of The Society of Dyers and Colourists. 9, 1-4.

- Isik, M. and Sponza, D. T. (2003). Effect of Oxygen on Decolorization of Azo Dyes by *Escherichia coli* and *Pseudomonas* sp. and Fate of Aromatic Amines. *Process Biochemistry*. 38, 1183-1192.
- Isik, M. and Sponza, D. T. (2004). Monitoring of Toxicity and Intermediates of C.I. Direct Black 38 Azo Dye through Decolorization in an Anaerobic/Aerobic Sequential Reactor System. *Journal of Hazardous Materials*. 114, 29-39.
- Kalme, S. D., Parshetti, G. K., Jadhav, S. U. and Govindwar, S. P. (2007).
   Biodegradation of Benzidine Based Dye Direct Blue-6 by *Pseudomonas desmolyticum* NCIM 2112. *Bioresource Technology*. 98, 1405-1410.
- Kamilaki, A. (2000). The Removal of Reactive Dyes from Textile Effluents-ABioreactor Approach Employing Whole Bacterial Cells. PhD thesis, UK:University of Leeds.
- Kapdan, I. K., Kargi, F., McMullan, G. and Marchant, R. (2000). Effect of environmental conditions on biological decolorization of textile dyestuff by *C. versicolor . Enzyme and Microbial Technology*. 26, 381-387.
- Kapdan I. K. and Oztekin, R. (2003). Decolorization of Textile Dyestuff Reactive Orange 16 in Fed-Batch Reactor under Anaerobic Condition. *Enzyme and Microbial Technology*. 33, 231-235.
- Kapdan, I. K. and Alparslan, S. (2005). Application of Anaerobic-Aerobic Sequential Treatment System to Real Textile Wastewater for Color and COD Removal. *Enzyme and Microbial Technology*. 36, 273-279.
- Kaushik, P. and Malik, A. (2009). Review Article: Fungal Dye Decolourization: Recent and Advances Future Potential. *Environment International*. 35, 127-141.

- Keharia, H., Patel, H. and Madamwar, D. (2004). Decolorization Screening of
  Synthetic Dyes by Anaerobic Methanogenic Sludge using a Batch
  Decolorization Assay. World Journal of Microbiology and Biotechnology. 20, 365-370
- Khalid, A., Arshad, M. and Crowley, D. E. (2010). Bioaugmentation of Azo Dyes.H. Atacag Erkurt (ed.), Biodegradation of Azo Dyes. *The Handbook of Environmental Chemistry*. 9, 1-37.
- Khehra, M. S., Saini, H. S., Sharma, D. K., Chadha, B. S. and Chimni, S. S. (2006).Biodegradation of Azo Dye C.I. Acid Red 88 by an Anoxic-Aerobic Sequential Bioreactor. *Dyes and Pigments*. 70, 1-7.
- Kirby, N., Marchant, R. and McMullan, G. (2000). Decolourisation Of Synthetic Textile Dyes by *Phlebia tremellosa*. *FEMS Microbiology Letters*. 188, 93-96.
- Knackmuss, H. J. (1996). Basic Knowledge and Perspectives of Bioelimination of Xenobiotic Compounds. *Journal of Biotechnology*. 51, 287-295.
- Knapp, J. S. and Newby, P. S. (1995). The Microbiological Decolorization of an Industrial Effluent Containing a Diazo-Linked Chromophore. Water Research. 29(7), 1807-1809.
- Kodam, K. M., Soojhawon, I., Lokhande, P. D. and Gawai, K. R. (2005). Microbial Decolorization of Reactive Azo Dyes under Aerobic Conditions. World Journal of Microbiology and Biotechnology. 21, 367-370.
- Kudlich, M., Keck, A., Klein, J. and Stolz, A. (1997). Localization of The Enzyme System Involved in Anaerobic Reduction of Azo Dyes by *Sphingomonas* sp. Strain BN6 and Effect of Artificial Redox Mediators on the Rate of Azo Dye Reduction. *Applied and Environmental Microbiology*. 63(9), 3691-3694.

- Kulkarni, M. and Chaudhari, A. (2007). Microbial Remediation of Nitro-Aromatic Compounds: An Overview. *Journal of Environmental Management*. 85, 496-512.
- Kulla, H.G., Klausener, F., Meyer, U., Lfideke, B. and Leisinger, T. (1983).
   Interference of Aromatic Sulfo Groups in the Microbial Degradation of the Azo Dyes Orange I and Orange II. *Archives of Microbiology*. 135, 1-7.
- Lee, Y. H., Matthews, R. D. and Pavlostathis, S. G. (2006). Biological decolorization of reactice anthraquinone and phthalocyanine dyes under various oxidationreduction conditions. *Water Environment Research*. 78, 156-169.
- Lorenco, N. D., Novais, J. M., Pinheiro, H. M. (2001). Effect of some operational parameters on textile dye biodegradation in a sequential batch reactor. *Journal of Biotechnology*. 89,163-174.
- Lourenco, N. D., Novais, J. M. and Pinheiro, H.M. (2006). Kinetic Studies of Reactive Azo Dye Decolorization in Anaerobic/Aerobic Sequencing Batch Reactors. *Biotechnology Letters*. 28, 733-739.
- Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. (1951). Protein Measurement with the Folin Phenol Reagent. *Journal of Biological Chemistry*.12, 21-43.
- Maas, R. and Chaudhari, S. (2005). Adsorption and Biological Decolourization of Azo Dye Reactive Red 2 in Semi Continuous Anaerobic Reactors. *Process Biochemistry* 40, 699-705.
- Madigan, J. E., Pusterla, N., Johnson, E., Chae, J. S., Berger Pusterla, J., DeRock, E. and Lawler, S.P. (2000). Transmission of *Ehrlichia risticii*, the Agent of Potomac Horse Fever, using Naturally Infected Aquatic Insects and Helminth Vectors: Preliminary Report. *Equine Veterinary Journal*. 32, 275-279.

- Maier, J., Kandelbauer, A., Erlancher, A., Cavaco-Paulo, A. and Gubits, G. M. (2004). A new alkali thermostable azoreductase from *bacillus sp.* Strain SF. *Applied and Environmental Microbiology*. 70, 837-844.
- Manu, B. and Chaudhari, S. (2002). Anaerobic Decolorization of Simulated Textile Wastewater Containing Azo Dyes. *Bioresource Technology*. 82, 225-231.
- Manu, B. and Chaudhari, S. (2003). Decolorization of indigo and azo dyes in semicontinuous reactors with long hydraulic retention time. *Process Biochemistry*. 38,1213-1221.
- Mc Mullan, G., Meehan, C., Conneely, A., Kirby, N., Robinson, T., Nigam, P.,
  Banat, I. M., Marchant, R. and Smyth, W. F. (2001). Microbial Decolourisation And Degradation Of Textile Dyes. *Applied and Environmental Microbiology*. 56, 81-87.
- Meehan, C., Bjourson, A. J. and McMullan, G. (2001). Paenibacillus azoreducens sp. nov., A Synthetic Azo Dye Decolorizing Bacterium from Industrial Wastewater. International Journal of Systematic and Evolutionary Microbiology. 51, 1681-1685.
- Mendez-Paz, D., Omil, F. and Lema, J. M. (2005). Anaerobic Treatment of Azo Dye Acid Orange 7 Under Batch Conditions. Enzyme and Microbial Technology. 36, 264-272.
- Mohan, S. V., Vijaya Bhaskar, Y. and Karthikeyan, J. (2004): Biological decolorization of azo dyes by algal spirogyra species. *International Journal* of Environment and Pollution. 21 (3), 210-222.
- Mohana, S., Shrivastava,S., Divecha, J. and Madamwar, D. (2007). Response Surface Methodology for Optimization of Medium for Decolorization of Textile Dye Direct Black 22 by a Novel Bacterial Consortium. *Bioresource Technology*. 23, 55-66.

- Mohaney, S., Dafale, N. and Rao, N. N. (2006). Microbial Decolorization of Reactive Black-5 in a Two-Stage Anaerobic–Aerobic Reactor using Acclimatized Activated Textile Sludge. *Biodegradation*. 17, 403-413.
- Moosvi, S. and Madamwar, D. (2007). An Integrated Process for the Treatment of CETP Wastewater using Coagulation, Anaerobic and Aerobic Process. *Bioresource Technology*. 98, 3384-3392.
- Moosvi, S., Keharia, H. and Madamwar, D. (2005). Decolourization of Textile Dye Reactive Violet 5 by a Newly Isolated Bacterial Consortium RVM 11.1. *World Journal of Microbiology and Biotechnology*. 21, 667-672.
- Moutaouakkil, A., Zeroual, Y., Dzayri, F. Z., Talbi, M., Lee, K. and Blaghen, M.
  (2004). Decolorization of Azo Dyes with *Enterobacter agglomerans* Immobilized in Different Supports by Using Fluidized Bed Bioreactor. *Current Microbiology*. 48, 124-129.
- Nachiyar, C. V. and Rajakumar, G. S. (2005). Purification And Characterization of an Oxygen Insensitive Azoreductase from *Pseudomonas aeruginosa*. *Enzyme* and Microbial Technology. 36, 503-509.
- Nachiyar, C. V. and Rajkumar, G. S. (2003). Degradation of a Tannery and Textile Dye, Navitan Fast Blue S5R by *Pseudomonas Aeruginosa*. World Journal of Microbiology and Biotechnology. 19, 609-614.
- Najafi, H. and Movahed, H. R. (2009). Improvement of COD and TOC Reactive Dyes in Textile Wastewater by Coagulation Chemical Material. *African Journal of Biotechnology*. Vol. 8 (13), 3053-3059.
- Nigam, P., Banat, I. M., Singh, D. and Marchant, R. (1996a). Microbial Process for the Decolorization of Textile Effluent Containing Azo, Diazo and Reactive Dyes. *Process Biochemistry*. 31(5), 435-442.

- Nigam, P., Mullan, M.C., Banat, I.M. and Marchant, R. (1996). Decolourisation of Effluent from the Textile Industry by a Microbial Consortium. *Biotechnology Letters*. 18(1), 117-120.
- Nortemann, B., Baumgarten, J., Rast, H. G. and Knackmuss, H. J. (1986). Bacterial communities degrading amino- and hydroxynaphthalenesulfonates. *Applied and Environmental Microbiology*. 52, 1195–1202.
- Novotny, C., Svobodova, K., Kasinath, A. and Erbanova, P. (2004). Biodegradation of Synthetic Dyes by *Irpex Lacteus* under Various Growth Conditions. *International Biodeterioration and Biodegradation*. 54, 215- 223.
- Ogawa, T., Yatome, C. and Idaka, E. (1981). Biodegradation of p-Aminoazobenzene by Continuous Cultivation of *Pseudomonas pseudomallei* 13NA. *Journal of the Society of Dyers and Colourists*. 97, 435-437.
- O'Neill, C., Freda, R. H., Dennis, L. H., Nidia, D. L., Helena, M. P. and Wouter, D. (1999). Colour in Textile Effluents-Sources, Measurements, Discharge Contents and Simulation: A Review. *Journal of Chemical Technology and Biotechnology*. 74, 1009-1018.
- O'Neill, C., Lopez, A., Esteves, S., Hawkes, F. R., Hawkes, D. L. and Wilcox, S.
  (2000). Azo-dye Degradation in an Anaerobic-Aerobic Treatment System Operating on Simulated Textile Efluent. *Applied Microbiology and Biotechnology*. 53, 249-254.
- Ooi, T., Shibata, T., Sato, R., Ohno, H., Kinoshita, S., Thuoc, T. L. and Taguchi, S. (2007). An Azoreductase, Aerobic NADH-Dependent Flavoprotein Discovered from *Bacillus* sp.: Functional Expression and Enzymatic Characterization. *Applied Microbiology and Biotechnology*. 75, 377-386.

- Oxspring, D. A., McMullan', G., Smyth, W. F. and Marchant, R. (1996).
  Decolourisation And Metabolism Of The Reactive Textile Dye, Remazol Black B, by an Immobilized Microbial Consortium. *Biotechnology Letters*. 18(5), 527-530.
- Pala, A. and Tokat, E. (2002). Technical Note: Color Removal from Cotton Textile Industry Wastewater in an Activated Sludge System with Various Additives. *Water Research.* 36, 2920-2925.
- Pandey, A., Singh, P. and Iyengar, L. (2007). Review: Bacterial Decolorization and Degradation of Azo Dyes. *International Biodeterioration and Biodegradation*. 59, 73-84.
- Panswad, T. and Luangdilok, W. (2000). Decolorization of Reactive Dyes with Different Molecular Structures under Different Environmental Conditions. *Water Research.* 34(17), 4177- 4180.
- Park, C., Lee, M., Lee, B., Kim, S-W., Chase, H. A., Lee, J. and Kim, S. (2006).
  Biodegradation and biosorption for decolorization of synthetic dyes by *Funalia trogii. Biochemical Engineering Journal.* 18, 21-35.
- Pearce, C.I., Lloyd, J.R. and Guthrie, J.T. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments* 58, 179-196.
- Pereira, L., Viegas, C., Coelho, A., Robalo, M.P. and Martins, L. (2007). Mechanisms of enzymatic degradation of azo and anthraquinone dyes by bacterial CotA-laccase. *Journal of Biotechnology*. 131, 98-121.
- Pielesz, A., Baranowska, I., Rybak, A. and Wochowicz, A. (2002). Detection and Determination of Aromatic Amines as Products of Reductive Splitting from Selected Azo Dyes. *Ecotoxicology and Environmental Safety*. 53, 42-47.

- Pinheiro, H.M., Touraud, E. and Thomas, O. (2004). Aromatic amines from azo dye reduction: status review with emphasis on direct UV spectrophotometric detection in textile industry wastewaters. *Dyes and Pigments*. 61, 121-139.
- Pricelius, S., Held, C., Murkovic, M., Bozic, M., Kokol, V., Cavaco-Paulo, A. and Guebitz, G.M. (2007a). Enzymatic reduction of azo and indigoid compounds. *Applied Microbiology and Biotechnology*. 77, 321-327.
- Rafii, F., Embden, J. G. H. R. V. and Asad, Y.F. (1997). Azoreductase and Nitroreductase Activity of Bacteria in Feces from Patients with an Ileal Reservoir. *Digestive Diseases and Sciences*. 42(1), 133-136.
- Rajaguru, P., Kalaiselvi, K., Palanevil, M. and Subburam, V. (2000). Biodegradation of azo dyes in a sequential anaerobic-aerobic system. *Applied Microbiology* and Biotechnology. 54, 268-273.
- Robinson, T., Chandran, B. and Nigam, P. (2002). Removal of dyes from an artificial textile dye effluent by two agricultural waste residues, corncob and barley husk. *Environment International*. 28, 29-33.
- Robinson, T., McMullan, G., Marchant, R. and Nigam, P. (2001). Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. *Bioresource Technology*. 77, 247-255.
- Russ, R., Rau, J. and Stolz, A. (2000). The Function of Cytoplasmic Flavin Reductases in The Reduction of Azo Dyes by Bacteria. *Applied and Environmental Microbiology*. 66(4), 1429-1434.
- Sahel, K., Perol, N., Chermette, H., Bordes, C., Derriche, Z. and Guillard, C. (2007).
  Photocatalytic decolorization of Remazol Black 5 (RB5) and Procion Red MX-5B -Isotherm of adsorption, kinetic of decolorization and mineralization. *Applied Catalysis B: Environmental.* 77, 100-109.

- Sanayei, Y., Ismail, N., Teng, T. T., and Morad, M. (2010). Studies on Flocculating Activity of Bioflocculant from Closed Drainage System (CDS) and Its Application in Reactive Dye Removal. *International Journal of Chemistry*. Vol. 2, No. 1, 168-173.
- Sani, R. K. and Banerjee, U. C.(1999). Decolorization of triphenylmethane dyes and textile and dye-stuff effluent by *Kurthia* sp.. *Enzyme and Microbial Technology*. 24, 433-437.
- Semde, R., Pierre, D., Geuskens, G., Devleeschouwer, M. and Moe, A. J. (1998). Study of Some Important Factors Involved in Azo Derivative Reduction by *Clostridium perfringens. International Journal of Pharmaceutics.* 161, 45-54.
- Shabir, G. A. (2003). Validation of HPLC Chromatography Methods for Pharmaceutical Analysis. Understanding the Differences and Similarities between Validation Requirements of FDA, the US Pharmacopeia and the ICH. Journal of Chromatography A. 987(1-2), 57-66.
- Singh, P., Mishra, L. C., Pandey, A. and Iyengar, L. (2006). Degradation of 4-Aminobenzenesulfonate by Alginate Encapsulated Cells of Agrobacterium sp. PNS-1. *Bioresource Technology*. 97, 1655-1659.
- Slepecky, R. A., Hemphill, H. E. (1992). The genus *Bacillus* Nonmedical the Prokaryotes (2<sup>nd</sup> ed). Edited by Balows, A., Trüper, H. G., Dworkin, M., Harder, W. and Schleifer, K. H. 1663-1696. New York: Springer.
- Slokar, Y. M. and Marechal, A. M. L. (1998). Methods of Decoloration of Textile Wastewaters. *Dyes and Pigments*. 37(4), 335-356.
- Somasiri, W., Ruan, W., Li Xiufen, and Chen Jian. (2006). Decolourization of Textile Wastewater Containing Acid Dyes in UASB Reactor System under Mixed Anaerobic Granular Sludge. *Electronic Journal of Environmental*, *Agricultural and Food Chemistry*. 5 (1), 1224-1234.

- Sponza, D. T. and Isik, M. (2002). Decolorization and Azo Dye Degradation by Anaerobic/Aerobic Sequential Process. *Enzyme and Microbial Technology*. 31, 102-110.
- Stackebrandt, E., Liesack, W. (1993). Nucleic acids and classification. *Handbook of New Bacterial Systematics*: 152-189.
- Steffan, S., Bardi, L. and Marzona, M. (2005). Azo Dye Biodegradation by Microbial Cultures Immobilized in Alginate Beads. *Environment International*. 31, 201-205.
- Stolz, A., Nörtemann, B. and Knackmuss, H. J. (1992). Bacterial metabolism of 5aminosalicylic acid. Initial ring cleavage. *Biochemical Journal*. 282, 675-680.
- Stolz, A. (2001). Basic and Applied Aspects in The Microbial Degradation of Azo Dyes. Applied Microbiology and Biotechnology. 56, 69-80.
- Sugiura, W., Miyashita, T., Yokoyama, T., Arai, M. (1999). Isolation of azo-dye degrading microorganisms and their application to white discharge printing of fabric. *Journal of Bioscience and Bioengineering*. 88: 577-581.
- Supaka, N., Juntongjin, K., Damronglerd, S., Delia, M. L. and Strehaiano, P. (2004). Microbial Decolorization of Reactive Azo Dyes in a Sequential Anaerobic-Aerobic System. *Chemical Engineering Journal*. 99, 169-176.
- Tan, N.C.G. and Field, J.A. (2000). Biodegradation of Sulfonated Aromatic Compounds. In: Environmental Technologies to Treat Sulfur Pollution. *Principles and Engineering*. IWA Publishing, London.
- Tan, N.C.G., Prenafeta-Boldu, F. X., Opsteeg, J. L., Lettinga, G. and Field, J. A.
  (1999). Biodegradation of Azo Dyes in Cocultures of Anaerobic Granular Sludge with Aerobic Aromatic Amine Degrading Enrichment Cultures. *Applied Microbiology and Biotechnology*. 51, 865-871.

- Ting, T. M. and Jamaludin, N. (2008). Decolorization and Decomposition of Organic Pollutants for Reactive and Disperse Dyes using Electron Beam Technology: Effect of the Concentrations of Pollutants and Irradiation Dose. *Chemosphere*. 73, 76-80.
- Todd, M. J. and Gomez, J. (2001). Enzyme Kinetics Determined using Calorimetry: A General Assay for Enzyme Activity? *Analytical Biochemistry*. 296, 179-187.
- Vajnhandl, S. and Marechal, A. M. L. (2005). Ultrasound in Textile Dyeing and the Decolouration/Mineralization of Textile Dyes. *Dyes and Pigments*. 65, 89-101.
- Van der Zee, F.P., Lettinga, G. and Field, J.A. (2001). Azo Dye Decolourisation by Anaerobic Granular Sludge. *Chemosphere*. 44(5), 1169-1176.
- Van der Zee, F. P. and Villaverde, S. (2005). Review: Combined Anaerobic-Aerobic Treatment of Azo Dyes - A Short Review of Bioreactor Studies. Water Research. 39, 1425-1440.
- Van der Zee, F. P. (2002). Anaerobic Azo Dye Reduction. Doctoral Thesis, Wageningen University. Wageningen, The Netherlands, 142 pp.
- Vijaya, P. P. and Sandhya, S. (2003). Decolorization and Complete Degradation of Methyl Red by a Mixed Culture. *The Environmentalist*. 23, 145-149.
- Wang, J., Lu, H., Jin, R., Zhou, J., Liu, G. and Xing, L. (2009). Decolorization of 1amino-4-bromoanthraquinone-2-sulfonic Acid in Bioaugmented Membrane Bioreactor. *Process Biochemistry*. 10, 1016-1025.
- Wang, X., Yin, C. and Wang, L. (2002). Structure-Activity Relationships and Response-Surface Analysis of Nitroaromatics Toxicity to the Yeast (Saccharomyces cerevisiae). Chemosphere. 46, 1045-1051.

- Welham, R. D. (1963). In Zollinger, H. (ed): Color Chemistry: Syntheses, properties and application of organic dyes and pigments. 2<sup>nd</sup> Rev. Edition.Weinheim; New York; Basel; Chambridge. 496pp.
- Whiteley, C. G. and Lee, D. J. (2006). Review Enzyme Technology and Biological Remediation. *Enzyme and Microbial Technology*. 38, 291-316.
- Willmott, N. J. (1997). The use of Bacteria-polymer Composites for The Removal of Colour from Reactive Dye Effluents. PhD thesis, UK: University of Leeds.
- Won, P. K. and Yuen, P. Y. (1996). Decolorization and degradation of Methyl Red by *Klebsiella pneumonia* R5-13. *Water Resources*. 30(7), 1736-1744.
- Wuhrmann, K., Mechsner, K. and Kappeler, T. (1980). Investigation on Rate Determining Factors in the Microbial Reduction of Azo Dyes. *European Journal of Applied Microbiology and Biotechnology*. 9, 325-338.
- Xu, M., Guo, J., Zeng, G., Zhong, X. and Sun, G. (2006). Decolorization of Anthraquinone Dye by Shewanella decolorationis S12. Applied Microbiology and Biotechnology. 71, 246-251.
- Yang, Q., Yediler, A., Yang, M. and Kettrup, A. (2005). Decolorization of an Azo dye, *Reactive Black 5* and MnP Production by Yeast Isolate: *Debaryomyces* polymorphus. Biochemical Engineering Journal. 24, 249-253.
- Yatome, C., Ogawa, T., Koga, D. and Idaka, E. (1981). Biodegradability of Azo and Triphenylmethane Dyes by *Pseudomonas pseudomallei* 13NA. *Journal of the Society of Dyers and Colourists*. 97, 166-9.
- Yatome, C., Matsufuru, H., Taguchi, T., Ogawa, T. (1993). Degradation of 4'dimethylaminoazobenzene-2-carboxylic acid by *Pseudomonas stutzeri*. *Applied Microbiology and Biotechnology*. 39, 778-781.

- Yoo, E.S. (2002). Chemical Decolorization of the Azo Dye CI Reactive Orange 96 by Various Organic/Inorganic Compounds. *Journal of Chemical Technology* and Biotechnology. 77, 481-485.
- Yoo, E. S., Libra, J. and Adrian, L. (2001). Mechanism of Decolorization of Azo Dyes in Anaerobic Mixed Culture. *Journal of Environmental Engineering*. 127(9), 844-849.
- Yu, Z. and Wen, X. (2005). Screening and Identification of Yeasts for Decolorizing Synthetic Dyes in Industrial Wastewater. *International Biodeterioration and Biodegradation*. 56, 109-114
- Zhao, X. and Hardin, I.R. (2007). HPLC and Spectrophotometric Analysis of Biodegradation of Azo dyes by *Pleurotus ostreatus*. *Dyes and Pigments*. 73, 322-325.
- Zheng, Z., Levin, R. E., Pinkham, J. L. and Shetty, K. (1999). Decolorization of Polymeric Dyes by A Novel *Penicillium* Isolate. *Process Biochemistry*. 34, 31-37.
- Zimmermann, T., Kulla, H. G. and Leisinger, T. (1982). Properties of Purified Orange I1 Azoreductase, the Enzyme Initiating Azo Dye Degradation by *Pseudumunas* KF46. *European Journal of Biochemistry*. 129, 197-203.
- Zimmerman, T, Gasser, F., Kulla, H. G. and Leisinger, T. (1984). Comparison of two bacterial azoreductases acquired during adaptation to growth on azo dyes. *Archives of Microbiology*. 138, 37-43.
- Zollinger, H. (1991). Color Chemistry: Syntheses, Properties and Application of Organic Dyes and Pigments. 2<sup>nd</sup> Rev. Edition.Weinheim; New York; Basel; Chambridge. 496pp.