

IDENTIFICATIONS OF LOCALLY ISOLATED REACTIVE BLACK 5
DECOLORIZING BACTERIA

LADEI YASEEN ABDULLAH

A thesis submitted in fulfillment of the
requirements for the award of the degree of
Master of Science (Biotechnology)

Faculty of Biosciences and Bioengineering
Universiti Teknologi Malaysia

JANUARY 2013

To my beloved mother, father and my dearest brother Dr. Hewa

ACKNOWLEDGEMENT

First of all alhamdulillah, all praise goes to Allah, who blessed and enabled me to achieve my research

I would like to express my sincere thanks and appreciation to my supervisor, Dr. Chong Chun Shiong, who continuously guided me throughout every step of my study and generously shared his time and knowledge with me.

Special thanks to my co-supervisor Associate Prof Dr. Zaharah Ibrahim for her remarkable assistance during conducting my experiments in the laboratory work and her guidance and encouragement during writing articles

My special thanks must be extended to all technical staff members at the Special Equipment Laboratory at UTM for their collaboration and assistance while carrying out my laboratory work. Millions words of thanks to fellow friends who showed their concern and support all the way. Their views and tips are useful indeed

Thanks are also extended to Dr. Hewa and Dr. Suhailah for their support and helpfulness otherwise it was difficult for me to complete this project.

Thanks again to all.....

ABSTRACT

Seven bacterial strains were successfully isolated from textile wastewater. They were screened for their ability to degrade azo dyes including Reactive Black 5 and Acid Orange 7. The screening showed that the decolourization of Reactive Black 5 was best performed under facultative anaerobic conditions with color removal ranging from 80% - 93% for the three bacterial strains (L1, L6 and L7) at 37°C after 96 hours incubation. However, no decolorization of Acid Orange 7 was observed in all of the isolated bacteria under facultative anaerobic condition. The L1, L6 and L7 were further tested for decolorization of Reactive Black 5 and Acid Orange 7 under aerobic conditions. No decolourization was observed in all sets of experiments. Inducibility studies showed Reactive Black 5 did not show an inducing effect on decolourization process. Two parameters (pH and temperature) were optimized for decolorization. The strains were incubated in medium with pH 4, 7 or 10 and it was found that the optimum pH for decolourization was 7 with 49%, 63% and 70% of decolourization by bacterial strains of L1, L6 and L7, respectively, after 48 hours of incubation. When the strains were incubated at 30°C, 37°C and 55°C, it was found that the optimum temperature was 37°C, which the highest decolourization percentage in the tested strains ranged from 53% - 71% after 48 hours of incubation. The 16S ribosomal ribonucleic acid (16S rRNA) sequences analysis revealed that strains L1 and L6 belonged to genus *Bacillus* with 99% identity to 16S rRNA gene of *Bacillus thuringiensis*. Strain L7 was found belong to genus *Escherichia* with 99% identity to 16S rRNA gene of *Escherichia coli* strain SCDC-1.

ABSTRAK

Tujuh strain bakteria telah berjaya dipencil daripada air sisa tekstil. Semua strain bakteria ini disaring untuk menguji keupayaan mereka untuk mengurai pewarna azo seperti Reactive Black 5 dan Asid Orange 7. Keputusan eksperimen menunjukkan pelunturan warna Reactive Black 5 adalah terbaik dalam keadaan fakultatif anaerobik, dengan penyingkiran warna antara 80% - 93% untuk tiga strain bakteria (L1, L6 dan L7), pada suhu 37 ° C selepas 96 jam inkubasi. Walau bagaimanapun, tiada sebarang penyahwarnaan Asid Orange 7 diperhatikan bagi semua strain bakteria dalam keadaan fakultatif anaerobik. Bakteria strain L1, L6 dan L7 turut diuji untuk penyahwarnaan Reactive Black 5 dan Asid Orange 7 dalam keadaan aerobik. Namun tiada sebarang penyahwarnaan direkodkan dalam semua set eksperimen. Kajian Indusibiliti menyatakan Reaktif Hitam 5 tidak memberi sebarang kesan yang mendorong kepada proses pelunturan warna. Dua parameter (pH dan suhu) telah dioptimumkan untuk memperoleh keputusan penyahwarnaan yang lebih baik. Ketiga-tiga bakteria strain dikulturkan dalam medium dengan pH 4, 7 dan 10. Selepas 48 jam inkubasi, keputusan eksperimen mempamerkan pH optimum untuk penyembunyian warna adalah 7, dengan peratus penyahwarnaan sebanyak 49%, 63% dan 70% oleh strain bakteria L1, L6 dan L7. Apabila ketiga-tiga bakteria strain dikulturkan pada suhu 30 ° C, 37 ° C dan 55 ° C, keputusan menunjukkan suhu optimum bagi L1, L6 dan L7 adalah 37 ° C, di mana kadar pelunturan warna tertinggi yang tercatat dari 53%- 71% selepas 48 jam inkubasi. Analisis 16S ribosomal asid ribonukleik (16S rRNA) menunjukkan strain L1 dan L6 adalah genus *Bacillus*, dengan 99% persamaan kepada gen 16S rRNA *Bacillus thuringiensis*. Manakala strain L7 adalah genus *Escherichia*, dengan persamaan 99% kepada gen 16S rRNA *Escherichia coli* strain SCDC-1.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	iii
	DEDICATION	iv
	ACKNOWLEDGEMENT	v
	ABSTRACT	vi
	ABSTRAK	vii
	TABLE OF CONTENT	viii
	LIST OF TABLES	xii
	LIST OF FIGURES	xiii
	LIST OF ABBREVIATIONS	xvi
	LIST OF APPENDICES	viii
1	INTRODUCTION	1
	1.1 Background of the Study	1
	1.2 Problem Statement	3
	1.3 Objectives of the Study	3
	1.4 Scope of the Study	3
2	LITERATURE REVIEW	4
	2.1 History of Dyes	4
	2.2 Classification of Dyes	5
	2.3 Azo Dyes	8
	2.3.1 Toxicity and Carcinogenicity of Azo Dyes	9
	2.4 Reactive Black	10
	2.5 Current Textile Waste Water Treatment System	10
	2.6 Chemical Methods	11
	2.6.1 Chemical Oxidation	12

2.6.2	Fenton's Reagent	11
2.6.3	Electrochemical Treatment	12
2.6.4	Ion Exchange	12
2.7	Physical Methods	13
2.7.1	Filtration	13
2.7.2	Adsorption Methods	13
2.8	Physical-Chemical Methods	15
2.9	Biological Techniques	15
2.9.1	Dyes Biodegradation by Fungi	15
2.9.2	Dyes Biodegradation by Algae	16
2.9.3	Dyes Biodegradation by Bacteria	17
2.10	Mechanisms of Microbial Color Removal under Anaerobic and Aerobic Conditions	21
2.11	Biodegradation by Pure Culture and Mixed Culture of Bacteria	22
3	MATERIALS AND METHODS	23
3.1	Textile Wastewater Sample Collection	23
3.2	Growth Medium Preparation	23
3.2.1	Nutrient Agar (NA)	23
3.2.2	Nutrient Broth (NB)	23
3.2.3	Synthetic Wastewater Medium: Chemically Defined Medium (CDM)	24
3.2.3.1	Preparation of Stock Solution for CDM media	24
3.3	Preparation of of Inoculum	25
3.3.1	Measurement of Cells	25
3.4	Isolation of Microorganisms	26
3.5	Preparation of Stock Culture	26
3.6	Screening of Bacteria for Color Removal	26
3.7	Inducibility Studies	27
3.8	Optimization of Azo Dye Decolourization	28
3.8.1	Effect of Temperature	28
3.8.2	Effect of pH	28

3.9	Gram Staining	28
3.10	16S rRNA Analysis	29
3.10.1	Genomic DNA Isolation	29
3.10.2	Agarose Gel Electrophoresis	30
3.10.3	Detection of genomic DNA and PCR Products	30
3.10.4	TAE Buffer	31
3.10.5	Polymerase Chain Reaction (PCR)	32
3.10.6	PCR Product Purification	34
3.11	Sequencing of the 16S rRNA Gene	35
3.12	Obtaining Full Sequence of Bacteria	35
3.13	Homology Search	35
3.14	Phylogenetic Analysis of 16S rDNA Gene	35
4	RESULTS AND DISCUSSION	36
4.1	Isolation of Color Removing Bacteria	36
4.2	Screening for Dye Decolorizing Bacteria	38
4.3	Decolorization of Reactive Black 5 (RB5) by strains L1, L6 and L7	43
4.4	Inducibilities Studies	44
4.5	Optimization	46
4.5.1	Effect of Temperature	46
4.5.2	Effect of pH	48
4.6	16S rRNA Analysis	51
4.6.1	Sequencing of the 16S rRNA Gene of L1 and L6	52
4.6.2	Sequence Analyses of Gene Encoding for the 16S rRNA from Bacterium (L1, L6)	53
4.6.3	Phylogenetic Tree Study of L1 and L6	54
4.6.4	Potential of Bacillus Thuringiensis Strains to Decolorize Azo Dyes	55
4.7	Identification of bacterium L7	56
4.7.1	Sequencing and Analysis of 16S rRNA Gene	56
4.7.2	Phylogenetic Tree Study of L7	57
4.7.3	Potential of Escherichia Coli Strain SCDC-16 for Azo Dye Decolorization	58

5	CONCLUSIONS AND SUGGESTIONS FOR FUTURE WORK	60
	5.1 Conclusion	60
	5.2 Suggestions for Future Work	61
	REFERENCE	62
	Appendices A-K	72-77

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Different class of dyes and their characteristics	7
2.2	Summary of physical and chemical methods	14
2.3	List of dye-degrading bacteria	18
3.1	Component of chemically defined medium (CDM)	24
3.2	50X TAE buffer compositions	31
3.3	Oligonucleotide primers 16S r RNA	32
3.4	Internal primers were designed by using the “oligoanalyser” tools from IDT web page from this link: https://eu.idtdna.com/scitools/scitools.aspx	32
3.5	PCR component reaction	33
3.6	Thermal profile of PCR reaction	33

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Indigo	4
2.2	Chromospheres, auxochrome and anthraquinone	6
2.3	Chemical structure of reactive black	10
2.4	Proposed mechanism for reduction of azo dyes by whole bacterial cells	21
3.1	Promega marker ladder	31
4.1	Sample of textile wastewater was incubated under facultative anaerobic condition at 37°C for 7 days	36
4.2	Serial dilution to obtain single colony	37
4.3	Bacterial colonies obtained from serial dilutions on agar plates	37
4.4	Seven pure isolates	38
4.5	Decolorization of Reactive Black 5 by 7 individual bacteria: All samples were incubated under facultative anaerobic conditions at 37°C and at pH 7 for 5 days	39
4.6	Decolorization of Reactive Black 5 by seven different strains of bacteria: All samples were incubated under aerobic conditions at 37°C, pH 7, for 5 days	40

4.7	Decolorization of Acid Orange 7 by 7 different bacteria strains: All samples were incubated under facultative anaerobic conditions at 37°C, pH 7 for 5 days	41
4.8	Specific decolorization acid orange 7 by 7 individual strains: All samples were incubated under aerobic condition, at 37°C, pH 7 for 5 days	41
4.9	Reactive Black 5 decolorization profile of strain L1, L6 and L7 at 37° C for 96 hour incubation	43
4.10	Decolorization of Reactive Black 5 using RB5 pre-exposed and non-pre-exposed inoculums of strain L1, L6 and L7 under partial anaerobic conditions at 37° C.	45
4.11	Effects of different temperatures (30°C, 37°C, and 55°C) on RB5 decolorization under facultative anaerobic condition for strain L1	46
4.12	Effects of different temperatures (30°C, 37°C, and 55°C) on RB5 decolorization under facultative anaerobic condition for strain L6	47
4.13	Effects of different temperatures (30°C, 37°C, and 55°C) on RB5 decolorization under facultative anaerobic conditions for strain L7	47
4.14	Effect of different pH levels on RB5 decolorization under facultative anaerobic conditions at 37°C for L1	49
4.15	Effect of different pH on RB5 decolorization under facultative anaerobic condition at 37°C by L6 strain	50
4.16	Effect of different pH on RB5 decolorization under facultative anaerobic conditions at 37°C for strain L7	50

4.17	The 1.5kb of PCR product from the 16S rRNA fragment obtained using 27F and 1525 R primers via PCR amplification	51
4.18	Agar gel electrophoresis of purified PCR amplification product	52
4.19	BLASTn search results for strains L1 and L6	53
4.20	A phylogenetic tree showing the inter relationship between L1, L6 and the top 10 blast results from NCBI Bacteria 16S ribosomal RNA Sequences	55
4.21	Sequences producing significant alignments with L7 bacterium	57
4.22	The phylogenetic tree showed the interrelationship between L7 and top 12 blast results from NCBI bacteria 16S ribosomal RNA sequences	58

LIST OF ABBREVIATIONS

%	-	Percent
°C	-	Degree Centigrade Celsius
λ	-	Wavelength
A	-	Absorbance
AO7	-	Acid orange 7
ADMI	-	American Dye Manufacturers Institute
BLAST	-	Basic local alignment search tool
BOD	-	Biological oxygen demand
A595	-	Absorbance at 595nm
C.I	-	Colour Index
CaCl ₂	-	Calcium chloride
CDM	-	Chemically defined medium
COD	-	Chemical oxygen demand
DNA	-	Deoxyribonucleic acid
EDTA	-	Ethylene diamine tetra acetic acid
g	-	Gram
H	-	Hour
HCl	-	Hydrochloric acid
Kb	-	Kilo base
K ₂ HPO ₄	-	Potassium dichromate
KH ₂ PO ₄	-	Potassium dihydrogen phosphate
MgSO ₄ .7H ₂ O	-	Magnesium sulphate heptahydrate
MEGA4	-	Molecular evolutionary genetics analysis software version 4
mg	-	milligram
Nm	-	Nanometer
μ L	-	Microlitre
mg/ml	-	Miligram per milliliter
mg/L	-	Miligram per Liter
NCBI	-	National Center for Biotechnology Information
(NH ₄) ₂ SO ₄	-	Ammonium sulphate
NaCl	-	Sodium chloride
NAD	-	Nicotinamide adenine dinucleotide
NAD ⁺	-	Nicotinamide adenine dinucleotide(oxidized)
NADH	-	Nicotinamide adenine dinucleotide(reduced)
NADP	-	Nicotinamide adenine dinucleotide phosphate

NADPH	-	Nicotinamide adenine dinucleotide phosphate(reduced)
NaOH	-	Sodium Hydroxide
NA	-	Nutrient Agar
NB	-	Nutrient Broth
PCR	-	Polymerase chain reaction
L	-	Liter
rDNA	-	Ribosomal DNA
RNA	-	Ribonucleic acid
RB5	-	Reactive Black 5
RBB	-	Remazol Black B
rpm	-	Rotation per minute
rRNA	-	Ribosomal RNA
TAE	-	Tris-acetate buffer
Tris	-	2-hydroxymethyl-2-methyl-1,3-propanediol
v/v	-	Volume per volume
w/v	-	Weight per volume
UV	-	Ultraviolet

LIST OF APPENDICES

APPDENDIX	TITLE	PAGE
A	Bacillus thuringiensis (strain L6) gram -positive bacterium	74
B	Escherichia coli Strain SCDC-16 (strain L7) gram-negative bacterium	74
C	Bacillus thuringiensis (strain L1) gram -positive bacterium	75
D	The 16S rRNA full sequence of strain L1	75
E	Strain L1 with top ten blasts	76
F	The 16S rRNA full sequence of strain L6	76
G	Strain L6 with top ten blasts	76
H	The 16S rRNA full sequence of strain L7	77
I	Strain L7 with top ten blasts	77
J	Estimates of evolutionary divergence between sequences (L7)	78
K	Estimates of evolutionary divergence between sequences (L1and L6)	79

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Nowadays, one of the most critical global issues is environmental pollution. Due to the quick economic development, the developing countries such as Malaysia, China, India and some other countries are facing the same pollution problem. Environmental pollution, particularly in water pollution, has caused a number of negative impacts to the environment. These include harmful algal blooms, eutrophication and contamination of groundwater, which is the fresh water resources for drinking (Wu *et al.*, 2012).

Wastewater contains a variety of pollutants such as toxic organics, dissolved solids, heavy metals, and color. The presence of color in the waste water gives a basic indication of water being contaminated by dyes such as triarylmethane, anthraquinone and azo dyes. Dyes are chemical materials consist of chromophores, delocalized electron system with conjugated double bonds and auxochromes. The intensity of the dyes color can be altered by electron drawing or donating substances (Christie, 2001). It is estimated that almost 10^9 kg of dyes are produced annually in the world, of which azo dyes represent about 70% by weight (Zollinger, 1987). Textile-processing wastewaters, typically with dye content in the range $10\text{-}200\text{ mgL}^{-1}$ (O'Neill *et al.*, 1999), are usually highly coloured and when discharged in open waters presents an aesthetic problem. As dyes are designed to be chemically and photolytically stable, they are highly persistent in natural environments. The release of dyes may therefore present an ecotoxic hazard and introduces the potential danger

of bioaccumulation that may eventually affect man by transport through the food chain.

Amongst these dyes, azo dye is mostly used in the industries (Stolz 2001). In general, azo dyes are xenobiotic that are very rigid against bio-degradative processes (Stolz 2001). These compounds are characterized by aromatic rings linked to one or more azo groups. The metabolic pathway of azo dye may result in free aromatic amines productions, which have been shown to be carcinogenic and mutagenic to human (Banat *et al.*, 1996).

Currently, there are different methods of textile wastewater treatment including physical and chemical process such as adsorption, membrane technologies, oxidation, and coagulation/flocculation. The advantages of these methods are that they are able to remove a wide range of dyes and rapid processes (Pearce *et al.*, 2003). On the other hands, these methods are costly and the accumulation of concentrated sludge may cause removal problems as well (Kamilaki, 2000; Pearce *et al.*, 2003). Biological treatment methods in general are more efficient and environmentally friendly (Pearce *et al.*, 2003). It has been reported that microorganisms are able to degrade azo dye efficiently under anoxic conditions and the intermediate products (amines) could be detoxified under aerobic environment (Grekova-Vasileva *et al.*, 2009). Under anaerobic condition bacteria secreted azoreductase enzyme, which could reductively cleavage the azo bond, yielding aromatic amines (Bibi *et al.*, 2012). The aromatic compounds were then degraded under aerobic condition (Stolz, 2001), to produced catechol compound and eventually to CO₂, water and ammonia (Van der Zee and Villaverde, 2005).

1.2 Problem Statement

The release of highly colored dye effluents has caused serious environmental damages. Therefore, color elimination in wastewater is one of the main issues in the textile industries. It has to be removed before discharging into receiving water body (Khadijah *et al.*, 2009). Azo dyes are considered as a toxic and undesirable substance in wastewater and their intermediate products are considered as a main cause of mutagenicity and carcinogenicity to living organisms. Hence, isolation and characterization of color removing bacteria is one of the important aspects as more competent isolated strains may enhance the efficiency of biodegradation of dyes.

1.3 Objectives of the Study

- 1- To isolate color removing bacteria from textile waste water
- 2- To optimize the decolorization process in a whole cell system
- 3- To identify the isolated bacteria using 16S rRNA analysis

1.4 Scope of the Study

The bacteria were first isolated from textile wastewater using serial dilution and spread plate method. They were screened for their ability to remove color in synthetic medium. The color removing bacteria were further characterized and the optimized pH and temperature were determined. In addition, the possible role of the azo dye as inducing agent for decolorization was also investigated. Finally, the color removing bacteria were identified using 16S rRNA analysis.

REFERENCES

- Aksu, Z. and Dönmez, G. (2003). A comparative study on the biosorption characteristics of some yeasts for Remazol Blue reactive dye. *Chemosphere*. 50 (8): 1075-1083
- Ali, H. (2010). Biodegradation of synthetic dyes—a review. *Water, Air, & Soil Pollution*. 213 (1): 251-273
- Allen, R.L.M. (1971). *Colour Chemistry*. Great Britain: Thomas Nelson Ltd.
- Banat, I. M., Nigam, P., Singh, D. and Marchant, R. (1996). Microbial decolorization of textile-dyecontaining effluents: a review. *Bioresource technology*. 58 (3): 217-227
- Barrett, Jc, Fry, B., Maller, J. and Daly, Mj (2005). Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics*. 21 (2): 263-265
- Bragger, JI, Lloyd, Aw, Soozandehfar, Sh, Bloomfield, Sf, Marriott, C. and Martin, Gp (1997). Investigations into the azo reducing activity of a common colonic microorganism. *International journal of pharmaceuticals*. 157 (1): 61-71
- Benson, D.A., Karsch-Mizrachi, I., Lipman, D.J., Ostell, J., Rapp, B.A. and Wheeler, D.L. (2002). *GenBank. Nucleic Acids Research*. 30 (1): 17-20
- Bes-Piá, A., Iborra-Clar, A., Mendoza-Roca, Ja, Iborra-Clar, Mi and Alcaina-Miranda, Mi (2004). Nanofiltration of biologically treated textile effluents using ozone as a pre-treatment. *Desalination*. 167: 387-392
- Beydili, Mi, Matthews, Rd and Pavlostathis, Sg (2001). Decolorization of a reactive copper-phthalocyanine dye under methanogenic conditions. *Water science and technology: a journal of the International Association on Water Pollution Research*. 43 (2): 333
- Bibi, R., Arshad, M. and Asghar, H.N. (2012). Optimization of factors for accelerated biodegradation of Reactive Black-5 azo dye. *International Journal of Agriculture and Biology*. 14 (3): 353-359
- Brown, D. and Laboureur, P. (1983). The degradation of dyestuffs: Part I Primary biodegradation under anaerobic conditions. *Chemosphere*. 12 (3): 397-404

- Brown, M.A. and De Vito, S.C. (1993). Predicting azo dye toxicity. *Critical Reviews in Environmental Science and Technology*. 23 (3): 249-324
- Carliell, C.M. (1993). Biological degradation of azo dyes in an anaerobic system. *Sc. Eng. Thesis. School of Chem. Eng., Univ. of Natal, Durban*
- Ciardelli, G. and Ranieri, N. (2001). The treatment and reuse of wastewater in the textile industry by means of ozonation and electroflocculation. *Water Research*. 35 (2): 567-572
- Chang, J.S., Chen, B.Y. and Lin, Y.S. (2004). Stimulation of bacterial decolorization of an azo dye by extracellular metabolites from *Escherichia coli* strain NO3. *Bioresource technology*. 91 (3): 243-248
- Chang, J.S., Kuo, T.S., Chao, Y.P., Ho, J.Y. and Lin, P.J. (2000). Azo dye decolorization with a mutant *Escherichia coli* strain. *Biotechnology letters*. 22 (9): 807-812
- Chang, J.S., Chou, C., Lin, Y.C., Lin, P.J., Ho, J.Y. and Lee Hu, T. (2001). Kinetic characteristics of bacterial azo-dye decolorization by *Pseudomonas luteola*. *Water Research*. 35 (12): 2841-2850
- Chang, J.S. and Kuo, T.S. (2000). Kinetics of bacterial decolorization of azo dye with *Escherichia coli*NO3. *Bioresource technology*. 75 (2): 107-111
- Chen, Brian E., Kondo, Masahiro, Garnier, Amélie, Watson, Fiona L., Püettmann-Holgado, Roland, Lamar, David R. and Schmucker, Dietmar (2006). The Molecular Diversity of Dscam Is Functionally Required for Neuronal Wiring Specificity in *Drosophila*. *Cell*. 125 (3): 607-620
- Chen, C., Xie, B., Ren, Y. and Wei, C. (2000). The mechanisms of affecting factors in treating wastewater by Fenton reagent. *Chinese Journal of Environmental Science*. 21 (3): 93-96
- Chen, B.Y., Zhang, M.M., Chang, C.T., Ding, Y., Chen, W.M. and Hsueh, C.C. (2011). Deciphering azo dye decolorization characteristics by indigenous *Proteus hauseri*: Chemical structure. *Journal of the Taiwan Institute of Chemical Engineers*. 42 (2): 327-333
- Chequer, F.M.D., Dorta, D.J. and De Oliveira, D.P. (2011). Azo Dyes and Their Metabolites: Does the Discharge of the Azo Dye into Water Bodies Represent Human and Ecological Risks? *Advances in treating textile effluent. In Tech, Croatia*: 28-48
- Claverie, J. M. and Notredame, C. (2003) *Bioinformatics for Dummies* ®. Wiley Publishing, Inc, 216, 382-405
- Cui, Daizong, Li, Guofang, Zhao, Dan, Gu, Xiaoxu, Wang, Chunlei and Zhao, Min (2012). Microbial community structures in mixed bacterial consortia for azo

dye treatment under aerobic and anaerobic conditions. *Journal of hazardous materials*. 221–222 (0): 185-192

- Dafale, N., Rao, N.N., Meshram, S.U. and Wate, S.R. (2008). Decolorization of azo dyes and simulated dye bath wastewater using acclimatized microbial consortium–biostimulation and halo tolerance. *Bioresource technology*. 99 (7): 2552-2558
- Deng, D., Guo, J., Zeng, G. and Sun, G. (2008). Decolorization of anthraquinone, triphenylmethane and azo dyes by a new isolated *Bacillus cereus* strain DC11. *International Biodeterioration & Biodegradation*. 62 (3): 263-269
- Dos Santos, A.B., Cervantes, F.J. and Van Lier, J.B. (2007). Review paper on current technologies for decolourisation of textile wastewaters: perspectives for anaerobic biotechnology. *Bioresource technology*. 98 (12): 2369-2385
- Fewson, Charles A. (1988). Biodegradation of xenobiotic and other persistent compounds: the causes of recalcitrance. *Trends in biotechnology*. 6 (7): 148-153
- Fitch, W. M. (2000). "Homology: a personal view on some of the problems." *Trends in genetics*. 16(5): 227-231 Elsevier Science Ltd
- Forgacs, E., Cserhati, T. and Oros, G. (2004). Removal of synthetic dyes from wastewaters: a review. *Environment International*. 30 (7): 953-971
- Franciscon, E., Grossman, M.J., Paschoal, J.A., Reyes, F.G. and Durrant, L.R. (2012). Decolorization and biodegradation of reactive sulfonated azo dyes by a newly isolated *Brevibacterium* sp. strain VN-15. *SpringerPlus*. 1 (1): 37
- Fu, Y. and Viraraghavan, T. (2001). Fungal decolorization of dye wastewaters: a review. *Bioresource technology*. 79 (3): 251-262
- Ganesh, R., Boardman, G.D. and Michelsen, D. (1994). Fate of azo dyes in sludges. *Water Research*. 28 (6): 1367-1376
- Glenn, J.K. and Gold, M.H. (1983). Decolorization of several polymeric dyes by the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology*. 45 (6): 1741-1747
- Gomes, Arlindo C., Fernandes, Luis R. and Simões, Rogério M. S. (2012). Oxidation rates of two textile dyes by ozone: Effect of pH and competitive kinetics. *Chemical Engineering Journal*. 189–190 (0): 175-181
- Grekova-Vasileva, M., Popov, I., Vassilev, D. and Topalova, Y. (2009). Isolation and Characterisation of microbial strain azo 29 capable of azo dye decolourization. *Biotechnology and Biotechnological Equipments*. 23: 318-322

- Ghoreishi, Sm and Haghghi, R. (2003). Chemical catalytic reaction and biological oxidation for treatment of non-biodegradable textile effluent. *Chemical Engineering Journal*. 95 (1): 163-169
- Ibrahim, Z. (2011). Decolourisation of Reactive Black 5 Using paenibacillus sp. Immobilised Onto Macrocomposite. *Journal of Bioremediation & Biodegradation*. S1:004. doi:10.4172/2155-6199.S1-004
- Ibrahim, Z., Md Salleh, M., Abdulrashid, N.A., Yahya, A. and Chong, C.S. (2006). Decolourization of Azo Dye Direct Blue 15 Using Batch Culture of *Klebsiella* sp, School of Postgraduate Studies, UTM.
- Işik, M. and Sponza, D.T. (2003). Aromatic amine degradation in a UASB/CSTR sequential system treating Congo Red dye. *Journal of Environmental Science and Health, Part A*. 38 (10): 2301-2315
- Itoh, K., Kitade, Y., Nakanishi, M. and Yatome, C. (2002). Decolorization of methyl red by a mixed culture of *Bacillus* sp. and *Pseudomonas stutzeri*. *Journal of Environmental Science and Health, Part A*. 37 (3): 415-421
- Inamuddin, I. and Luqman, M. (2012). Ion-Exchange Technology I: Theory and Materials. Springer.DOI :10.1007/978-94-007-1700-8_1
- Igunnu, E.T. and Chen, G.Z. (2012). Produced water treatment technologies. *International Journal of Low-Carbon Technologies* doi: 10.1093/ijlct/cts049
- Jadhav, U.U., Dawkar, V.V., Ghodake, G.S. and Govindwar, S.P. (2008). Biodegradation of Direct Red 5B, a textile dye by newly isolated *Comamonas* sp. UVS. *Journal of hazardous materials* 158 (2): 507-516
- Jain, Kunal, Shah, Varun, Chapla, Digantkumar and Madamwar, Datta (2012). Decolorization and degradation of azo dye – Reactive Violet 5R by an acclimatized indigenous bacterial mixed cultures-SB4 isolated from anthropogenic dye contaminated soil. *Journal of hazardous materials* 213–214 (0): 378-386
- Jiménez, B., Noyola, A. and Capdeville, B. (1988). Selected dyes for residence time distribution evaluation in bioreactors. *Biotechnology techniques*. 2 (2): 77-82
- Jinqi, L. and Houtian, L. (1992). Degradation of azo dyes by algae. *Environmental Pollution* 75 (3): 273-278
- Kamilaki, A. (2000). The removal of reactive dyes from textile effluents-a bioreactor approach employing whole bacterial cells, PhD thesis, UK: University of Leeds.
- Keck, A., Klein, J., Kudlich, M., Stolz, A., Knackmuss, H.J. and Mattes, R. (1997). Reduction of azo dyes by redox mediators originating in the naphthalenesulfonic acid degradation pathway of *Sphingomonas* sp. strain BN6. *Applied and Environmental Microbiology*. 63 (9): 3684-3690

- Khadijah, O., Lee, K. K. and Abdullah, M. F. F. (2009). Isolation, screening and development of local bacterial consortia with azo dyes decolourising capability. *Malaysian Journal of Microbiology*. 5: 25-32
- Khalid, A., Arshad, M. and Crowley, D.E. (2008). Decolorization of azo dyes by *Shewanella* sp. under saline conditions. *Applied Microbiology and Biotechnology*. 79 (6): 1053-1059
- Khalid, A., Arshad, M. and Crowley, D.E. (2008). Accelerated decolorization of structurally different azo dyes by newly isolated bacterial strains. *Applied Microbiology and Biotechnology*. 78 (2): 361-369
- Khalid, A., M. Arshad, and Crowley, D. (2010). Bioaugmentation of Azo Dyes. In: H. Atacag Erkurt (ed.). Biodegradation of Azo Dyes. Pakistan. *Springer-Verlag*. 9: (0:) 1-37
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S. and Chimni, S.S. (2005a). Comparative studies on potential of consortium and constituent pure bacterial isolates to decolorize azo dyes. *Water Research*. 39 (20): 5135-5141
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S. and Chimni, S.S. (2005b). Decolorization of various azo dyes by bacterial consortium. *Dyes and pigments*. 67 (1): 55-61
- Kirk, T.K. and Farrell, R.L. (1987). Enzymatic "combustion": the microbial degradation of lignin. *Annual Reviews in Microbiology*. 41 (1): 465-501
- Kuhad, R.C., Kapoor, M. and Rustagi, R. (2004). Enhanced production of an alkaline pectinase from *Streptomyces* sp. RCK-SC by whole-cell immobilization and solid-state cultivation. *World Journal of Microbiology and Biotechnology*. 20 (3): 257-263
- Kodam, Km, Soojhawon, I, Lokhande, Pd and Gawai, Kr (2005). Microbial decolorization of reactive azo dyes under aerobic conditions. *World Journal of Microbiology and Biotechnology*. 21 (3): 367-370
- Lane, D.J. 1991. 16S/23S rRNA sequencing. In: Nucleic acid techniques in bacterial systematics. Stackebrandt, E., and Goodfellow, M., eds., *John Wiley and Sons, New York, NY*, pp. 115-175
- Ledakowicz, S., Bilińska, L. and Żyłła, R. (2012). Application of Fenton's Reagent in the Textile Wastewater Treatment Under Industrial Conditions. 19(2), pp. 133-279. Retrieved 22 Nov. 2012, from doi: 10.2478/v10216-011-0013-z
- Lin, Jun, Zhang, Xingwang, Li, Zhongjian and Lei, Lecheng (2010). Biodegradation of Reactive blue 13 in a two-stage anaerobic/aerobic fluidized beds system with a *Pseudomonas* sp. isolate. *Bioresource technology*. 101 (1): 34-40

- Li, T. and Guthrie, J.T. (2010). Colour removal from aqueous solutions of metal-complex azo dyes using bacterial cells of *Shewanella* strain J18 143. *Bioresource technology*. 101 (12): 4291-4295
- Marchesi, J.R., Sato, T., Weightman, A.J., Martin, T.A., Fry, J.C., Hiom, S.J. and Wade, W.G. (1998). Design and evaluation of useful bacterium-specific PCR primers that amplify genes coding for bacterial 16S rRNA. *Applied and Environmental Microbiology*. 64 (2): 795-799
- Mahmood, S., Arshad, M., Khalid, A., Nazli, Z.H. and Mahmood, T. (2011). Isolation and screening of azo dye decolorizing bacterial isolates from dye-contaminated textile wastewater. *Soil and Environment*. 30 (1): 7-12
- Maier, J., Kandelbauer, A., Erlacher, A., Cavaco-Paulo, A. and Gübitz, G.M. (2004). A new alkali-thermostable azoreductase from *Bacillus* sp. strain SF. *Applied and Environmental Microbiology*. 70 (2): 837-844
- Meehan, C., Bjourson, A.J. and McMullan, G. (2001). *Paenibacillus azoreducens* sp. nov., a synthetic azo dye decolorizing bacterium from industrial wastewater. *International journal of systematic and evolutionary microbiology* 51 (5): 1681-1685
- Masyithah Aalia. (2012). Characterisation of Azoreductase Produced by *Brevibacillus panacihumi* during the decolourization of Reactive Black 5. *Applied Microbiology and Biotechnology* 56 (1): 81-87
- McMullan, G., Meehan, C., Conneely, A., Kirby, N., Robinson, T., Nigam, P., Banat, Im, Marchant, R. and Smyth, Wf (2001). Microbial decolourisation and degradation of textile dyes. *Applied Microbiology and Biotechnology*. 56 (1): 81-87
- Mergaert, J., Lednická, D., Goris, J., Cnockaert, M.C., De Vos, P. and Swings, J. (2003). Taxonomic study of *Cellvibrio* strains and description of *Cellvibrio ostraviensis* sp. nov., *Cellvibrio fibrivorans* sp. nov. and *Cellvibrio gandavensis* sp. nov. *International journal of systematic and evolutionary microbiology*. 53 (2): 465-471
- Mishra, G. and Tripathy, M. (1993). A critical review of the treatments for decolourization of textile effluent. *Colourage*. 40: 35-35
- Moosvi, S., Keharia, H. and Madamwar, D. (2005). Decolourization of textile dye Reactive Violet 5 by a newly isolated bacterial consortium RVM 11.1. *World Journal of Microbiology and Biotechnology*. 21 (5): 667-672
- Nachiyar, C.V. and Rajakumar, G.S. (2005). Purification and characterization of an oxygen insensitive azoreductase from *Pseudomonas aeruginosa*. *Enzyme and Microbial Technology*. 36 (4): 503-509
- Nam, S. (1999). Azo dye transformation by enzymatic and chemical systems. Dissertation. Faculty of the Oregon Graduate Institute of Science and Technology. 1-145

- Nigam, P., Armour, G., Banat, Im, Singh, D. and Marchant, R. (2000). Physical removal of textile dyes from effluents and solid-state fermentation of dye-adsorbed agricultural residues. *Bioresource technology*. 72 (3): 219-226
- Nigam, P., Banat, I.M., Singh, D. and Marchant, R. (1996). Microbial process for the decolorization of textile effluent containing azo, diazo and reactive dyes. *Process Biochemistry*. 31 (5): 435-442
- Ogawa, T., Yatome, C., Idaka, E. and Kamiya, H. (1986). Biodegradation of azo acid dyes by continuous cultivation of *Pseudomonas cepacia* 13NA. *Journal of the Society of Dyers and Colourists*. 102 (1): 12-14
- Ogugbue, Chimeziejason, Morad, Norhashimah, Sawidis, Thomas and Oranusi, Nathaniela (2012). Decolorization and partial mineralization of a polyazo dye by *Bacillus firmus* immobilized within tubular polymeric gel. *3 Biotech*. 2 (1): 67-78
- Ong, Siew-Teng, Keng, Pei-Sin, Lee, Weng-Nam, Ha, Sie-Tiong and Hung, Yung-Tse (2011). Dye Waste Treatment. *Water*. 3 (1): 157-176
- Padamavathy, S. (2003). Aerobic decolorization of reactive azo dyes in presence of various cosubstrates. *Chemical and biochemical engineering quarterly*. 17 (2): 147-152
- Pagga, U. and Taeger, K. (1994). Development of a method for adsorption of dyestuffs on activated sludge. *Water Research*. 28 (5): 1051-1057
- Pan, Hongmiao, Feng, Jinhui, He, Gui-Xin, Cerniglia, Carl E. and Chen, Huizhong (2012). Evaluation of impact of exposure of Sudan azo dyes and their metabolites on human intestinal bacteria. *Anaerobe*. 18 (4): 445-453
- Pandey, A., Singh, P. and Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *International Biodeterioration & Biodegradation*. 59 (2): 73-84
- Pearce, Ci, Lloyd, Jr and Guthrie, Jt (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and pigments*. 58 (3): 179-196
- Prabakaran, V., Arthysurendren, S. and Vigneewaran, M. (2012). Biodegradation & Bioremediation of Azo Dye entrenched soil by *Pseudomonas* sp. *International Journal of Chemical and Analytical Science*. 3 (5): 1381-1384
- Ponraj, M., Gokila, K. and Zambare, V. (2011). Bacterial decolorization of textile dye-Orange 3R. *International Journal of Advanced Biotechnology and Research* 2: 168-177

- Qu, M.H., Wang, Y.Z., Wang, C., Ge, X.G., Wang, D.Y. and Zhou, Q. (2005). A novel method for preparing poly (ethylene terephthalate)/BaSO₄ nanocomposites. *European polymer journal*. 41 (11): 2569-2574
- Rai, H.S., Bhattacharyya, M.S., Singh, J., Bansal, Tk, Vats, P. and Banerjee, Uc (2005). Removal of dyes from the effluent of textile and dyestuff manufacturing industry: a review of emerging techniques with reference to biological treatment. *Critical Reviews in Environmental Science and Technology*. 35 (3): 219-238
- Robinson, T., McMullan, G., Marchant, R. and Nigam, P. (2001). Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. *Bioresource technology*. 77 (3): 247-255
- Sala Gastón, M. and Gutiérrez Bouzán, M.C. (2012). Electrochemical techniques in textile processes and wastewater treatment. , vol. 2012, Article ID 629103, 12 pages, 2012. doi:10.1155/2012/629103
- Sani, R.K. and Banerjee, U.C. (1999). Decolorization of triphenylmethane dyes and textile and dye-stuff effluent by *Kurthia* sp. *Enzyme and Microbial Technology*. 24 (7): 433-437
- Saratale, Rg, Saratale, Gd, Chang, Js and Govindwar, Sp (2010). Decolorization and biodegradation of reactive dyes and dye wastewater by a developed bacterial consortium. *Biodegradation*. 21 (6): 999-1015
- Saratale, Rg, Saratale, Gd, Chang, Js and Govindwar, Sp (2011). Bacterial decolorization and degradation of azo dyes: A review. *Journal of the Taiwan Institute of Chemical Engineers*. 42 (1): 138-157
- Saratale, Rg, Saratale, Gd, Kalyani, Dc, Chang, J.S. and Govindwar, Sp (2009). Enhanced decolorization and biodegradation of textile azo dye Scarlet R by using developed microbial consortium-GR. *Bioresource technology*. 100 (9): 2493-2500
- Sarayu, K. and Sandhya, S. (2010). Aerobic biodegradation pathway for Remazol Orange by *Pseudomonas aeruginosa*. *Applied biochemistry and biotechnology*. 160 (4): 1241-1253
- Sarayu, K. and Sandhya, S. (2012). Current Technologies for Biological Treatment of Textile Wastewater—A Review. *Applied biochemistry and biotechnology*. 167 (3): 645-661
- Sandhya, S. (2010). Biodegradation of Azo Dyes Under Anaerobic Condition: Role of Azoreductase. *Biodegradation of Azo Dyes*. Atacag Erkurt, Springer Berlin Heidelberg. 9: 39-57
- Semple, K.T., Cain, R.B. and Schmidt, S. (2006). Biodegradation of aromatic compounds by microalgae. *FEMS Microbiology letters*. 170 (2): 291-300

- Shaul, G.M., Holdsworth, T.J., Dempsey, C.R. and Dostal, K.A. (1991). Fate of water soluble azo dyes in the activated sludge process. *Chemosphere*. 22 (1): 107-119
- Singh, P., Iyengar, L. and Pandey, A. (2012). Bacterial Decolorization and Degradation of Azo Dyes. *Microbial Degradation of Xenobiotics*: 101-133 doi10.1007/978-3-642-23789-8_4
- Sirianuntapiboon, S. and Srisornsak, P. (2007). Removal of disperse dyes from textile wastewater using bio-sludge. *Bioresource technology* 98 (5): 1057-1066
- Shuler, MI and Kargi, F. (2002). *Bioprocess Engineering—Basic Principles*, Prentice Hall PTR, USA.
- Solís, M., Solís, A., Pérez, H.I., Manjarrez, N. and Flores, M. (2012). Microbial Decolouration of Azo Dyes: A Review. *Process Biochemistry* ,<http://dx.doi.org/10.1016/j.procbio.2012>.
- Spadaro, J.T., Isabelle, L. and Renganathan, V. (1994). Hydroxyl radical mediated degradation of azo dyes: evidence for benzene generation. *Environmental science & technology*. 28 (7): 1389-1393
- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of azo dyes. *Applied Microbiology and Biotechnology*. 56 (1): 69-80
- Swamy, J. and Ramsay, Ja (1999). The evaluation of white rot fungi in the decoloration of textile dyes. *Enzyme and Microbial Technology*. 24 (3): 130-137
- Tan, N. (2001). *Integrated and Sequential Anaerobic/aerobic Biodegradation of Azo Dyes*, Ph.D. Thesis, Agrotechnology and Food Sciences, Sub-department of *Environmental Technology*, Wageningen University, The Netherlands.
- Tamura, K., Dudley, J., Nei, M. and Kumar, S. (2007). MEGA4: molecular evolutionary genetics analysis (MEGA) software version 4.0. *Molecular biology and evolution*. 24 (8): 1596-1599
- Tony, B.D., Goyal, D. and Khanna, S. (2009a). Decolorization of Direct Red 28 by mixed bacterial culture in an up-flow immobilized bioreactor. *Journal of industrial microbiology & biotechnology*. 36 (7): 955-960
- Tony, B.D., Goyal, D. and Khanna, S. (2009b). Decolorization of textile azo dyes by aerobic bacterial consortium. *International Biodeterioration & Biodegradation*. 63 (4): 462-469
- Van Der Zee, F.P. and Villaverde, S. (2005). Combined anaerobic–aerobic treatment of azo dyes—a short review of bioreactor studies. *Water Research*. 39 (8): 1425-1440
- Walker, Gm and Weatherley, Lr (2001). COD removal from textile industry effluent: pilot plant studies. *Chemical Engineering Journal*. 84 (2): 125-131

- Wallace, T.H. (2001). Biological treatment of a synthetic dye water and an industrial textile wastewater containing azo dye compounds, Virginia Polytechnic Institute and State University.
- Wang, H., Zheng, X.W., Su, J.Q., Tian, Y., Xiong, X.J. and Zheng, T.L. (2009). Biological decolorization of the reactive dyes Reactive Black 5 by a novel isolated bacterial strain *Enterobacter* sp. EC3. *Journal of hazardous materials*. 171 (1): 654-659
- Wu, Yonghong, Li, Tianling and Yang, Linzhang (2012). Mechanisms of removing pollutants from aqueous solutions by microorganisms and their aggregates: A review. *Bioresource technology*. 107 (0): 10-18
- Willmott, N.J. (1997). The use of bacteria-polymer composites for the removal of colour from reactive dye effluents, University of Leeds.
- Wuhrmann, K., Mechsner, Kl and Kappeler, Th (1980). Investigation on rate—Determining factors in the microbial reduction of azo dyes. *Applied Microbiology and Biotechnology* 9 (4): 325-338
- Zabłocka-Godlewska, Ewa, Przystaś, Wioletta and Grabińska-Sota, Elżbieta (2012). Decolourization of Diazo Evans Blue by Two Strains of *Pseudomonas fluorescens* Isolated from Different Wastewater Treatment Plants. *Water, Air, & Soil Pollution*. 223 (8): 5259-5266
- Zee, F.P. (2002). Anaerobic azo dye reduction, Ph. D. thesis. Wageningen University, Wageningen-Netherlands
- Zhang, F. and Yu, J. (2000). Decolourisation of Acid Violet 7 with complex pellets of white rot fungus and activated carbon. *Bioprocess and Biosystems Engineering*. 23 (3): 295-301
- Zhang, Z., Schwartz, S., Wagner, L. and Miller, W. (2000). A greedy algorithm for aligning DNA sequences. *Journal of Computational Biology*. 7 (1-2): 203-214
- Zimmermann, T., Gasser, F., Kulla, H.G. and Leisinger, T. (1984). Comparison of two bacterial azoreductases acquired during adaptation to growth on azo dyes. *Archives of Microbiology*. 138 (1): 37-43
- Zimmermann, T., H. Kulla, and Leisinger, T. (1982). ‘‘Properties of Purified Orange II Azoreductase, the Enzyme Initiating Azo Dye Degradation by *Pseudomonas* KF46,’’ *Eur.J. Biochem.* 129: 197–203
- Zissi, U. and Lyberatos, G. (1996). Azo-dye biodegradation under anoxic conditions. *Water science and technology*. 34 (5): 495-500
- Zollinger H. (1991) Color Chemistry: Syntheses, Properties and Applications of Organic Dyes and Pigments. VCH Publishers, 2nd edn., New York, U.S.A