THE USE OF PROTEIN FROM PALM KERNEL MEAL IN DIETS OF SEABASS (*Lates calcarifer*)

MOHAMMED SUHAIMEE BIN ABD. MANAF

UNIVERSITI TEKNOLOGI MALAYSIA

THE USE OF PROTEIN FROM PALM KERNEL MEAL IN DIETS OF SEABASS (Lates calcarifer)

MOHAMMED SUHAIMEE BIN ABD. MANAF

A thesis submitted in fulfilment of the requirements for the award of the degree of Master of Science (Chemistry)

> Faculty of Science Universiti Teknologi Malaysia

> > SEPTEMBER 2005

ACKNOWLEDGEMENT

In preparing this thesis, I was in contact with many people, researchers, academicians, and practitioners. They have contributed towards my understanding and thoughts. In particular, I wish to express my sincere appreciation to my main thesis supervisor, Dr. Zainoha Zakaria, for encouragement, guidance, critics and friendship. I am also very thankful to my co-supervisor Associate Professor Dr. Ng Wing Keong (USM). Without their continued support and interest, this thesis would not have been the same as presented here.

I am also indebted to Department of Fisheries, Malaysia, for funding my M.Sc. study at Brackishwater Aquaculture Research Centre, Gelang Patah, Johor, and the Fisheries Research Institute of Penang also deserve special thanks for their assistance in supplying the relevant literature and providing the necessary equipment.

My sincere appreciation also extend to all my colleagues and others who have provided assistance at various occasions. Their views and tips are useful indeed. Unfortunately, it is not possible to list all of them in this limited space. I am also very grateful to all my family members.

ABSTRACT

Malaysia has an abundant supply of palm kernel meal (PKM) as a plant protein source as it contains up to 20 % crude protein. It has the potential to at least partly reduce the dependency on the soybean meal presently used in fish diets. However, the nutritive value of PKM limits its incorporation in fish diets due to the presence of high fibre content. A preliminary study on the use of raw PKM indicated that it could be included up to 15% inclusion level in the diets of juvenile seabass without affecting fish growth performance and feed efficiency. A series of problemsolving experiments were conducted to optimise the utilization of PKM as a feed ingredient in the diets of seabass (Lates calcarifer). Initially a chemical extraction of protein involving acid and bases was conducted. Less than 3% (w/w protein) soluble protein from PKM was obtained after 30 min in either acid (pH 2.7) or alkaline (pH 12.5) conditions. However, this amount increases to about 50% (w/w) when left for a longer period of time (3 h) at 28 °C and in alkaline condition (pH 10 and above). Increasing the extraction temperatures (60 °C, 80 °C, 105 °C or 121 °C) did not substantially improve the amount of extracted protein. The low recovery by the chemical extraction method could probably be due to the protein being hindered by layers of cell wall. Thus, pre-treatment on the raw PKM by physical elements such as pressure and heat using the fibre explosion process were attempted. However, it also did not substantially improve protein recovery compared to untreated raw PKM (P > 0.05). Hence, another strategy to expose the binding or entrapment of protein by polysaccharides was to treat PKM with urea at 80:750 g kg⁻¹ (urea:water). A series of feeding trial were then conducted to evaluate the effect of using raw PKM and chemically-treated PKM using urea (UTPKM) as feed ingredients in diets on growth performance of juvenile seabass L. calcarifer. Seabass juveniles fed 7.5% PKM and 15% PKM or 7.5% UTPKM, 15% UTPKM and 22.5% UTPKM diets showed similar growth performance (P > 0.05) with control diet. However, fish fed 22.5% PKM diet showed poor growth performance and feed efficiency (P < 0.05) than fish fed the control diet. On the other hand, there were significant differences (P < 0.05) in terms of protein efficiency ratio (PER) as fish fed the control diet giving the best PER compared to fish fed either 22.5% PKM, 15.0% UTPKM or 22.5% UTPKM. Net protein utilization (NPU) of fish fed 15.0% PKM and 22.5% PKM diets were substantially lower (P < 0.05) than fish fed the control diet. In general, fish growth performance and feed efficiency were shown to be better when fish fed diets with up to 22.5% inclusion of UTPKM than fish fed 22.5% PKM diets. It was concluded that raw PKM has the potential to partially replace soybean meal in the diets of juvenile seabass and that urea treatment on PKM may further improve the utilization of this by-product from the oil palm industry.

ABSTRAK

Malaysia mempunyai bekalan palm kernel meal (PKM) yang banyak sebagai sumber protein, yang mempunyai potensi untuk mengganti sekurang-kurangnya sebahagian tepung soya yang biasanya digunakan dalam makanan ikan. Walaupun begitu, kandungan serat kasar yang tinggi menghadkan penggunaannya dalam makanan ikan. Kajian awal dalam penggunaan PKM menunjukkan sehingga 15% tahap pencampuran dalam diet ikan siakap (Lates calcarifer) boleh dilakukan tanpa mengganggu kadar tumbesaran dan keberkesanan makanan ikan. Beberapa siri kajian telah dijalankan untuk mengoptimumkan penggunaan PKM sebagai ramuan dalam makanan ikan siakap. Kurang dari 3% (w/w protein) protein larut didapati daripada PKM setelah 30 min pencampuran samada dalam larutan berasid (pH 2.7) atau beralkali (pH 12.5). Walau bagaimanapun, jumlah protein larut didapati meningkat sekitar 50% (w/w) apabila PKM dicampurkan dalam larutan beralkali (pada pH 10 dan ke atas) dalam masa 180 min (3 jam) pada suhu bilik (28°C). Peningkatan suhu pengekstrakan kepada 60°C, 80°C, 105°C atau 121°C tidak dapat meningkatkan kandungan protein larut. Ini adalah kerana, kemungkinan protein PKM dihalangi oleh lapisan-lapisan dinding sel. Oleh itu, kaedah perawatan awal melalui elemen fizikal seperti tekanan tinggi dan pemanasan menggunakan proses letupan serat-amonia diuji. Hasilnya, penggunaan kaedah perawatan tersebut tidak dapat meningkatkan jumlah pengekstrakan protein berbanding dengan protein larut yang didapati sebelumnya (P>0.05). Maka, kaedah lain adalah untuk mendedahkan protein PKM melalui kaedah perawatan dengan urea pada kadar 80:750 g kg⁻¹ Satu siri percubaan makanan telah dijalankan untuk menilai kesan (urea:air). penggunaan PKM dan PKM selepas rawatan urea (UTPKM) sebagai ramuan terhadap pencapaian tumbesaran dan keberkesanan makanan ikan siakap. Ikan siakap yang diberi diet-diet 7.5% PKM dan 15% PKM atau diet-diet 7.5% UTPKM, 15% UTPKM dan 22.5% UTPKM menunjukkan pencapaian tumbesaran yang sama di antara satu dengan yang lain (P>0.05) berbanding diet kawalan tanpa PKM. Walaupun begitu, kadar tumbesaran dan keberkesanan makanan ikan yang diberi diet 22.5% PKM merosot (P<0.05) berbanding ikan yang diberi diet kawalan. Dalam lain hal, terdapat perbezaan ketara (P<0.05) dari segi nisbah keberkesanan protein (PER) di mana ikan yang diberi diet kawalan menunjukkan nilai PER yang terbaik (P<0.05) berbanding ikan yang diberi diet 22.5% PKM, 15.0% UTPKM atau 22.5% UTPKM. Penggunaan tetap protein (NPU) ikan yang diberi diet 15.0% PKM dan 22.5% PKM adalah rendah (P<0.05) berbanding ikan yang diberi diet kawalan. Pada amnya, pencapaian tumbesaran dan keberkesanan makanan siakap adalah lebih baik apabila diberi diet yang mengandungi sehingga 22.5% UTPKM berbanding diet yang mengandungi 22.5% PKM. Kesimpulannya, PKM berpotensi untuk menggantikan sebahagian dari tepung soya dalam diet ikan siakap dan perawatan dengan urea ke atas PKM berkemungkinan dapat meningkatkan lagi penggunaan bahan sampingan dari industri minyak sawit ini.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENT	iv
	ABSTRACT (ENGLISH)	V
	ABSTRAK (BAHASA)	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xi
	LIST OF FIGURES	xiii
	LIST OF SYMBOLS/ABBREVIATIONS	xvii
	LIST OF APPENDICES	xix

1	INTR	ODUCTION		
	1.1	The Importance of Aquaculture as an Alternative		
		Source of Protein for Human Consumption	1	
	1.2	The Need of Alternative Protein Source in Aquaculture		
		Feeds	2	
	1.3	Research Objectives	4	
2	LITE	RATURE REVIEW	6	
	2.1	Marine Fish Aquaculture in Malaysia	6	
	2.1.1	Cultured Species	10	
	2.2	Anatomy and Physiology of the Digestive System	11	
		2.2.1 Digestive System of Fish	11	

	2.2.	2 Digestion and Absorption	13
		2.2.2.1 Digestion and Absorption of Proteins	13
		2.2.2.2 Digestion and Absorption of Carbohydrates	14
		2.2.2.3 Digestion and Absorption of Lipids	15
2.3	Fee	ds in Aquaculture	15
	2.3.	1 Dietary Requirement of Fish	15
	2.3.	2 Feed Formulation	17
	2.3.	3 Feed Ingredients	17
2.4	Palr	n Kernel Meal	19
	2.4.	1 Nutrient Compositions of PKM	22
2.5	Prot	ein Extraction Process	24
2.6	Stra	tegies to Improve Nutritional Quality of Low Quality	
	Fora	ages	25
	2.6.	1 Acid Treatment	28
	2.6.	2 Alkaline Treatment	29
	2.6.	3 Biological Treatment	31
	2.6.	4 Physicochemical Treatment	32
MA	TERIA	LS AND METHODS	34
3.1	Mat	erials	34
3.2	Che	mical Analyses	34
	3.1.	1 Proximate Analysis	34
		3.2.1.1 Crude Protein	35
		3.2.1.2 Crude Fat	35
		3.2.1.3 Crude Fibre	36
		a. Acid Detergent Fibre	37
		b. Acid Detergent Lignin	38
		c. Neutral Detergent Fibre	38
		3.2.1.4 Ash / Organic Matter	39
		3.2.1.5 Nitrogen-Free Extract	40
		3.2.1.6 Dry Matter	40
	3.2.2	Amino Acids Analyses	41
	3.2.3	Gross Energy	41
	3.2.4	Non-Protein Nitrogen	41

3

3.3	Prelim	inary Study on the Potential of Raw PKM as a	
	Dietar	y Ingredient on the Growth Performance of Seabass	42
	3.3.1	Fish and Management	44
	3.3.2	Growth Performance and Feed Utilization	
		Efficiency	44
		3.3.2.1 Weight gain	44
		3.3.2.2 Specific Growth Rate	45
		3.3.2.3 Feed Conversion Ratio	45
		3.3.2.4 Protein Efficiency Ratio	45
		3.3.2.5 Net Protein Utilization	45
	3.3.3	Statistical Analysis	46
3.4	Prelim	inary Study on the Potential of Extracting Protein	
	from P	УКМ	46
	3.4.1	Protein Solubility of PKM	46
	3.4.2	Effect of Various pH Ranges on PKM Protein	
		Solubility	47
	3.4.3	Effect of Time and Temperature on the Rate of	
		Protein Extraction	48
3.5	Pre-Tr	reatment of PKM	48
	3.5.1	Ammonia Fibre Explosion (AFEX) Process	49
	3.5.2	Urea Treatment	50
3.6	Effect	Of Raw And Treated PKM As Feed Ingredient In	
	Seabas	ss Diet	50
	3.6.1	Feed Formulation	51
	3.6.2	Diet Preparation	52
	3.6.3	Feeding Trial on the Use of PKM and Treated	
		PKM as Feed Ingredients in Seabass Diets	53
RESU	ULTS A	ND DISCUSSION	54
4.1	Prelim	inary Study on the Potential of Raw PKM as a	
	Dietar	y Ingredient on the Growth Performance of Seabass	54
4.2	Prelim	inary Study on the Potential of Extracting Protein	
	from F	РКМ	59
	4.2.1	Protein Solubility of PKM	59
	 3.3 3.4 3.5 3.6 RESU 4.1 4.2 	 3.3 Prelim Dietary 3.3.1 3.3.1 3.3.2 3.4 3.5 3.4 3.4.1 3.4.2 3.4.3 3.5 3.5.1 3.5.1 3.5.1 3.5.2 3.6 Effect Seabas 3.6.1 3.6.2 3.6.3 RESULTS A 4.1 Prelim 5.6.2 3.6.3 RESULTS A 4.1 Prelim 10:etar 4.2 Prelim 10:etar 4.2 Prelim	 3.3 Preliminary Study on the Potential of Raw PKM as a Dietary Ingredient on the Growth Performance of Seabass 3.3.1 Fish and Management 3.3.2 Growth Performance and Feed Utilization Efficiency 3.3.2.1 Weight gain 3.3.2.2 Specific Growth Rate 3.3.2.3 Feed Conversion Ratio 3.3.2.4 Protein Efficiency Ratio 3.3.2.5 Net Protein Utilization 3.3.3 Statistical Analysis 3.4 Preliminary Study on the Potential of Extracting Protein from PKM 3.4.1 Protein Solubility of PKM 3.4.2 Effect of Various pH Ranges on PKM Protein Solubility 3.4.3 Effect of Time and Temperature on the Rate of Protein Extraction 3.5 Pre-Treatment of PKM 3.5.1 Ammonia Fibre Explosion (AFEX) Process 3.5.2 Urea Treatment 3.6 Effect OF Raw And Treated PKM As Feed Ingredient In Seabass Diet 3.6.1 Feed Formulation 3.6.3 Feeding Trial on the Use of PKM and Treated PKM as Feed Ingredients in Seabass Diets RESULTS AND DISCUSSION 4.1 Preliminary Study on the Potential of Raw PKM as a Dietary Ingredient on the Growth Performance of Seabass 4.2 Preliminary Study on the Potential of Extracting Protein from PKM 4.1 Preliminary Study on the Potential of Extracting Protein from PKM 4.2.1 Protein Solubility of PKM

		4.2.2	Effect of pH on PKM Protein Solubility	62
		4.2.3	Effect of Time on the Rate of Protein Extraction	65
		4.2.4	Effect of Temperature on the Rate of Protein	
			Extraction	68
	4.3	Ammo	onia Fibre Explosion (AFEX) Process	73
	4.4	Urea	Treatment of PKM	78
	4.5	The Ir	clusion of PKM and UTPKM as Feed	
		Ingred	lients in the Diet of Juvenile Seabass L. calcarifer.	83
5	CON	CLUSI	ONS AND SUGGESTIONS	95
	5.1	Concl	usions	95
	5.2	Sugge	stions for Further Work	97
REFERENC	CES		98 -	- 116
Appendices A – C		117 -	- 121	

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	General observations on feeding habits and relative	
	gut lengths (ratio of intestine to body length)	13
2.2	Summary on the performance of the Malaysian oil palm industry -2004	21
2.3	Chemical composition (g kg ⁻¹ DM) of palm kernel meal	
	according to some international databases	23
2.4	Yields and chemical composition of palm kernel meals	
	and the corresponding cell wall materials (CWM) (g kg ⁻¹)	24
2.5	Effect of pH, duration time (h) and temperatures (°C) on	
	extraction of various plant proteins as obtained by several	
	other researchers	26
3.1	Formulation (g kg ⁻¹ , dry) of the experimental diets	
	using local fish meal as the main protein source	43
3.2	Formulation (g kg ⁻¹ , dry) of the experimental diets	
	using Danish fish meal as the main protein source	52
4.1	Proximate composition (%, g dry weight) of various	
	protein and carbohydrate sources used in the	
	experimental diets	55

4.2	Chemical composition of the experimental diets on dry	
	weight basis (g/100g diet).	56
4.3	Growth and feed utilization efficiency of seabass fed	
	palm kernel meal-based diets for 42 days.	57
4.4	Proximate compositions (%) of urea-treated palm kernel	
	meal at different urea concentration and moisture content	
	(g kg $^{-1}$ DM) after 7 days at 30 °C.	79
4.5	Proximate compositions (%) of urea-treated palm kernel	
	meal at different urea concentration and moisture content	
	(g kg $^{-1}$ DM) after 21 days at 30 $^{\circ}$ C.	80
4.6	Proximate composition (%, g dry weight) and amino acid	
	compositions of various protein and carbohydrate source	
	used in the experimental diets	84
4.7	Proximate composition (%, dry weight) and gross energy	
	level (kJg ⁻¹) of the experimental diets	85
4.8	Growth and feed utilization efficiency of seabass fed	
	different palm kernel meal-based diets	92
4.9	Whole-body composition (%, dry matter basis) of seabass	
	fed palm kernel meal-based diets for 56 days	94

LIST OF FIGURES

FIGURE NO	. TITLE	PAGE
1.1	World fish production and food use consumption 1976 – 2030 (FAO, 2002)	2
2.1	Trends of marine fish landings (million tonnes) in Malaysia from 1992 to 2003 (Annual Fisheries Statistics, 2004).	7
2.2	Estimated aquaculture production (thousand tonnes) and value (RM million) in Malaysia from 1992 to 2003 (Annual Fisheries Statistics, 2004).	8
2.3	Fisheries Districts with Marine Aquaculture System in Malaysia (Annual Fisheries Statistics 2004)	9
2.4	Seabass (Lates calcarifer)	10
2.5	Diagrammatic representation of the digestive system of four fishes arranged in order of increasing gut length. a. Carnivore (seabass); b. Omnivore, mainly eating animal food (catfish); c. Omnivore, mainly eating plant food (carp); d. Herbivore (milkfish) (De Silva and Anderson 1995)	12
2.6	Schematic illustration of palm kernel meal production	12
	from palm tree nut	20

2.7	Schematic diagram to show the influence of pH on	
	carboxyl and amino groups of protein	27
2.8	Schematic diagram to illustrate protein such as	
	β -lactoglobulin solubles at high and low pH and	
	aggregates at its isoelectric point (pI).	27
4.1	Soluble protein content (%) of palm kernel meal in 5	
	different solid to solvent ratios (w/v) after 30 minutes	
	of mixing in pH2.7 (■), 10.5 (■) and 12.5 (■)	
	respectively.	60
4.2	Effect of pH on soluble protein content (%) of PKM	
	after 30 minutes (\blacksquare) and after 3 hours (\blacklozenge) of protein	
	extraction at ambient temperature (28 °C)	63
4.3	Protein content (%, w/w dry basis) in palm kernel meal	
	protein extract (a) and its respective residue (b) after	
	various extraction time (min) in acid (pH 2.7) and alkali	
	treatment (pH 12.5) at 28 °C	66
4.4	Residues (g) of PKM protein extract (a) and PKM (b)	
	after various extraction time (min) in acid (pH 2.7) and	
	alkali treatment (pH 12.5) at 28 °C	67
4.5	Protein content (%, w/w dry basis) in palm kernel meal	
	protein extract at different temperatures (°C) after acid (a)	
	and alkali (b) treatment	69
4.6	Protein content (%, w/w dry basis) in palm kernel meal	
	residues at different temperatures (°C) after acid (a) and	
	alkali (b) treatment	70

4.7	Recovered amount (g) of PKMPE at different	
	temperatures (°C) after acid (a) and alkali (b) treatment	71
4.8	Recovered amount (g) of PKM residues at different	
	temperatures ($^{\circ}$ C) in acid (a) and alkali (b) treatment	72
4.9	Protein content (%, w/w dry basis) in ammonia-fibre-	
	explosion-treated palm kernel meal protein extract after	
	acid (a) and alkali (b) treatment at various temperatures	
	(°C).	74
4.10	Protein content (%, w/w dry basis) in AFEX-treated	
	PKM residues after acid (a) and alkali (b) treatment at	
	various temperatures (°C).	75
4.11	Recovered amount (g) of AFEXPKM protein extract	
	at different temperatures (°C) after acid (a) and alkali	
	(b) treatment	76
4.12	Recovered amount (g) of AFEX-treated PKM residues	
	at different temperatures (°C) after acid (a) and alkali	
	(b) treatment	77
4.13	Plot shows the mean weight (g) of fish fed PKM-based	
	diets at various inclusion levels for 56 days	87
4.14	Bar chart shows the mean final weight (g) of seabass	
	fed PKM-based diets at various inclusion levels of either	
	PKM or UTPKM	88
4.15	Bar chart shows the weight gain (%) of seabass fed	
	PKM-based diets at various inclusion levels of either	
	PKM or UTPKM	88

4.16	Bar chart shows the specific growth rates (% day ⁻¹) of			
	seabass fed PKM-based diets at various inclusion levels			
	of either PKM or UTPKM	89		
4.17	Bar chart shows the feed conversion ratios of seabass			
	fed PKM-based diets at various inclusion levels of			
	either PKM or UTPKM	89		
4.18	Bar chart shows protein efficiency ratios of seabass			
	fed PKM-based diets at various inclusion levels of			
	either PKM or UTPKM	90		
4.19	Bar chart shows net protein utilization of seabass			
	fed PKM-based diets at various inclusion levels of			
	either PKM or UTPKM	90		

LIST OF SYMBOLS / ABBREVIATIONS

ADF	-	Acid detergent fibre
ADL	-	Acid detergent lignin
AFEX	-	Ammonia fibre explosion
AFEXTPKM	-	Ammonia fibre explosion treated palm kernel meal
cm	-	centimetre
СР	-	Crude protein
DFM	-	Danish fish meal
DL	-	Dietary lipid
DM	-	Dry matter
DO	-	Dissolved oxygen
DP	-	Dietary protein
EAA	-	Essential amino acids
FCR	-	Feed conversion ratio
FO	-	Fish oil
h	-	hour
ht	-	height
pI	-	Isoelectric point
ME	-	Metabolizable energy
min	-	minute
um	-	micron
mm	-	millimetre
NDF	-	Neutral detergent fibre

NFE	-	Nitrogen-free extract
NPU	-	Net protein utilization
NPN	-	Non protein nitrogen
NSP	-	Non-starch polysaccharide
PER	-	Protein efficiency ratio
РКМ	-	Palm kernel meal
PKMPE		Palm kernel meal protein extract
ppm	-	Parts per million.
ppt	-	Parts per thousand.
SBM	-	Soybean meal
SGR	-	Specific growth rate
°C	-	Degrees Celsius
TCA	-	Trichloroacetic acid
UTPKM	-	Urea treated palm kernel meal

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	Steps in Diet Formulation.	119
В	Abstract of Paper Presented at the National Fisheries Symposium 2003.	121
С	Abstract of Paper Presented at the Asian Fisheries Forum 2004.	122

CHAPTER ONE INTRODUCTION

CHAPTER 1

INTRODUCTION

1.1 The Importance of Aquaculture as a Source of Protein for Human Consumption

Aquaculture is the fastest expanding food producing sector in the world. According to Food and Agriculture Organization of the United Nation, Rome (FAO) statistics, aquaculture's contribution to global supplies of fish, crustaceans and molluses continues to grow, increasing from 3.9 percent of total production by weight in 1970 to 27.3 percent in 2000. Worldwide, the sector has increased at an average compounded rate of 9.2 percent per year since 1970, compared with only 1.4 percent for capture fisheries and 2.8 percent for terrestrial farmed meat production systems [1]. In 2000, reported total aquaculture production (including aquatic plants) was 45.7 million tonnes by weight and USD56.5 billion by value. More than half of the total world aquaculture production in 2000 was finfish, and the growth of the major species groups continues to be rapid with no apparent slowdown in production to date. As production from wild fishery has been static at around 100 million tonnes for the last decade and unlikely to increase further, aquaculture production must at least double by the year 2030 if current per capita seafood consumption of between 19 and 21 kg is to be met [1 - 4]. Figure 1.1 shows the world fish production and food use consumption from 1976 and projected to 2030. Although much of this aquaculture expansion will have to come from inland water culture of herbivorous fish such as carp and tilapia, there will also be greater demand for high-value fish species that are grown on artificially-provided food. Global aquaculture feed requirements are estimated currently at between 3 and 4 million tonnes per annum and are predicted to double by the year 2010 with Asia alone expecting to consume at least 2.6 million tonnes [5, 6].



Figure 1.1: World fish production and food use consumption 1976 – 2030 (FAO, 2002)

1.2 The Need for Alternative Protein Sources in Aquaculture Feeds

One of the major factors limiting the expansion of aquaculture is the development of nutritionally adequate, cost-effective diets. Feeds and feeding can contribute up to 70% of the total operating costs for fish and shrimp farms [7]. The most expensive component of pelleted feed is protein, of which 25-55% is required, depending upon whether the species is herbivorous, omnivorous or carnivorous [8, 9]. The major protein source for most aquaculture diets is fishmeal [9, 10] and formulated diets can contain up to 60% fishmeal [7, 11]. Almost 30% of fishmeal available globally for export is being consumed by the aquaculture sector [5, 12, 13]. However, there are some major problems with fishmeal. Fishmeal and fish oil production is declining [14] and the aquaculture feed industry currently uses more than 3 million tonnes of the global fisheries catch [15] excluding 'trash fish' which is fed directly to aquaculture species. As aquaculture production increases, demand for fishmeal will also increase, inevitably forcing prices to rise. As higher quality

fishmeal is generally required for aquaculture feeds, species of fish currently used for human consumption will increasingly be targeted by manufacturers for fishmeal production. In Malaysia, much of the cheap fish used to produce salted fish for human consumption is now instead used for aquaculture [12]. It is evident from the statistics that continued expansion of aquaculture would be curtailed unless suitable alternatives to fishmeal are found.

The development of cost-effective diets, with reduced contents of fish- and other aquatic- meals is an urgent priority for most fish and crustacean aquaculture industries. The problem is particularly important in Malaysia, which has very poor supplies of aquatic meals. Malaysia is particularly vulnerable to any world shortage of fishmeal because of our reliance on imported fishmeal and other non-edible marine product. In Malaysia, under the Third National Agriculture Plan, aquaculture industry is going to be a major contributor of fish and shrimp to fulfil the protein demand of the nation. Aquaculture production of brackishwater fish species in year 2010 has been targeted at 110,000 metric tonnes from only around 8,000 metric tonnes in year 2001 [16]. Among the high-valued marine species are Asian seabass (*Lates calcarifer*), mangrove snapper (*Lutjanus argentimaculatus*) and grouper (*Epinephelus sp.*). At present, the aquaculture industry in Malaysia is still dependent on either trash fish or costly imported diets. The use of cost-effective aquaculture diets with reduced fishmeal usage will significantly enhance the aquaculture industry in Malaysia.

Future aquaculture expansion will be primarily dependent on intensification and feeding with nutritionally complete diets. With increasing costs and the uncertainty in the future supply of fishmeal, it is thus desirable for nutritionists and feed manufacturers to opt for less expensive, readily available plant protein as a substitute for fishmeal. Oilseed proteins such as soybean meal has been used with varying degrees of success as a replacement for fishmeal in the diets for many cultured fish species. Freshwater fish such as channel catfish (*Ictalurus punctatus*) and blue catfish (*I. furcatus*) [17, 18] as well as marine fish such as rainbow trout (*Oncorhynchus mykiss*), Atlantic salmon (*Salmo salar*) [19], Australian snapper (*Pagrus auratus*) [20], cobia (*Rachycentron canadum*) [21] and Asian seabass (*L. calcarifer*) [22 - 24] were reported to be able to accept certain levels of soybean meal in their diets without affecting growth performance. However, the rising costs of conventional feed ingredients such as fishmeal and soybean meal, which are usually imported into tropical countries have stimulated much research into the use of alternative, locally available, plant protein sources [25].

Malaysia has an abundant supply of plant protein feed, which has the potential to at least partially reduce soybean meal presently used in compounded aquaculture diets. Palm kernel meal (PKM) is an oilseed protein source available in large quantities. The global production of PKM is increasing due to the remarkable growth of the oil palm industry in many parts of Asia and Africa. About 2 million tonnes of PKM are produced in Malaysia annually, the current global leader in the oil palm industry [26].

PKM has been successfully used as a diet ingredient for ruminants [27, 28] and can be incorporated at low levels (10-30%) in the diets of non-ruminant livestock such as poultry [29,30] and swine [31 - 34]. Generally, studies on the use of PKM by fish are very few and PKM is therefore seldom used in fish diets. In addition, the nutritive value of PKM limits its incorporation in fish diets due to the presence of high fibre content. It is worth to note that since PKM is very cheap, its incorporation in fish diets could considerably reduce the cost of feed. Thus, series of problem-solving experiments were conducted to determine the potential and improve the nutritional quality of PKM so that its use as a feed ingredient in the diets of seabass (*L. calcarifer*) could be optimised. This project provides the first attempt on isolating protein from PKM as the use of chemicals in protein isolation process from oilseeds such as soybean, cereal such as wheat and leaves such as alfalfa have successfully been practiced for decades. At present, the use of PKM in the diet of carnivorous marine fish species has not been reported.

1.3 Research Objectives

The main objective of this research is to evaluate the potential of PKM as a protein source in practical diets of seabass *Lates calcarifer* (Siakap) by enhancing its nutritional quality. This includes a series of experiments to determine:

1.3.1 The possibility of extracting protein from raw PKM in either acidic or alkaline condition,

- 1.3.2 The effect of pre-treating the cell wall of PKM using ammonia-fibre explosion method prior to protein extraction,
- 1.3.3 The effect of urea treatment on PKM.
- 1.3.4 The effect of using raw and treated PKM as feed ingredient in diets to partially replace soybean meal on growth performance and feed efficiency of seabass.

REFERENCES

- 1. FAO. *The State of World Fisheries and Aquaculture 2002*. Rome: Food and Agriculture Organization of the United Nation. 2002.
- 2. Chamberlain, G.W. Aquaculture trends. *World Aquaculture*. 1993. 24: 19-29.
- 3. Csavas, I. World Aquaculture status and outlook. *Infofish International*. 1994. 5/94: 47-54.
- 4. Gjedrem, T. Selective breeding to improve aquaculture production. *World Aquaculture*. 1997. Mar: 33-45.
- 5. New, M.B. and Csavas, I. Will there be enough fish meal for fish feeds? *Aquaculture Europe*. 1995. 19: 6-13.
- 6. Tacon, A.G.J. Global trends in aquaculture and aquafeed production. FAO/Globefish Research Program Report, FAO, Rome. 1996.
- Tacon, A.G.J. Feed ingredients for carnivorous fish species Alternatives to fishmeal and other fishery resources. In: FAO Fisheries Circ. FIRI/C881.
 Food and Agriculture Organisation of the United Nations, Rome. 35. 1994.
- Wee, K.L. An overview of fish digestive physiology and the relevance to the formulation of artificial fish feeds. In: G.L. Allan and W. Dall (Eds), Proceedings of the Aquaculture Nutrition Workshop. NSW Fisheries, Salamandar Bay, NSW, Australia. 1992.
- NRC. In: Nutrient Requirement of Fish. National Research Council, Washington: National Academy Press. 1993.
- Lovell, T. Canola meal in catfish feeds. Aquaculture Magazine, 1989. Sep/Oct: 68-70.

- Lovell, R.T. Nutrition and feeding of channel catfish. In: G.L. Allan and W. Dall (Eds), Proceedings of the Aquaculture Nutrition Workshop. NSW Fisheries, Salamandar Bay, NSW, Australia. 1992.
- New, M.B. Where will feeds be in year 2000? Fish Farmer. 1991. May/June: 38-41.
- Starkey, T.J. Status of fish meal supplies and market demand. Miscellaneous report. Stamford, CT, USA: H.J. Baker and Bro. Inc. 1994.
- Barlow, S. Fishmeal world outlook to the year 2000. Fish Farmer. 1989. Sep/Oct: 40-43.
- 15. New, M.B. and Wijkstrom, U. Feed for thought Some observations on aquaculture feed production in Asia. *World Aquaculture*. 1990. 21: 17-23.
- Annual Fisheries Statistics 2003, Vol. I. Department of Fisheries, Malaysia. Perniagaan Normahs. 2004.
- Webster, C.D., J.H. Tidwell, L.S. Goodgame, D.H. Yancey and L. Mackey. Use of soybean meal and distillers grains with solubles as partial or total replacement of fish meal in diets for channel catfish, *Ictalurus punctatus*. *Aquaculture*. 1992. 106: 301-309.
- Webster, C. D., Goodgame-Tiu, L. S. and Tidwell, J. H. Total replacement of fish meal by soybean meal, with various percentages of supplemental Lmethionine, in diets for blue catfish, *Ictalurus furcatus* (Lesueur). *Aquaculture Research*. 1995. 26: 299-306.
- Refstie, S., Korsøen, O.J., Storebakken, T., Baeverfjord, G., Lein, I. and Roem, A.J. Differing nutritional responses to dietary soybean meal in rainbow trout (*Oncorhynchus mykiss*) and Atlantic salmon (*Salmo salar*). *Aquaculture*. 2000. 190: 49–63.

- 20. Quartararo, N., Allan, G.L. and Bell, J.D. Replacement of fish meal in diets for Australian snapper, *Pagrus auratus*. *Aquaculture*. 1998. 166: 279–295.
- 21. Chou, R.L, Her, B.Y., Su, M.S., Hwang, G Wu, Y.H. Chen, H.Y. Substituting fish meal with soybean meal in diets of juvenile cobia *Rachycentron canadum*. *Aquaculture*. 2004. 229: 325–333.
- Boonyaratpalin, M., Suraneiranat, P. and Tunpibal, T. Replacement of fish meal with various types of soybean products in diets for the Asian seabass, *Lates calcarifer. Aquaculture.* 1998. 161: 67–78.
- Suhaimee, A.M., Che Utama, C.M., Mohd. Khan, B., and Ahmad Daud, O. Substitution of Fishmeal in Seabass (*Lates calcarifer*) diet with Soybean Meal. Fisheries Research Institute, Penang, Malaysia. *FRI Newsletter*. 1999. 4(1): 16-17.
- Suhaimee, A.M., Che Utama, C.M. and Mohd. Khan, B. The Effect of Heattreated Soybean on the Growth Performance of Seabass (*Lates calcarifer*), Fed Soybean Meal-based Diets. *Malaysian Fisheries Journal*. 2000. 1(2): 150-156.
- 25. El-Sayed, A.F.M. Alternative dietary protein sources for farmed tilapia, *Oreochromis* spp. *Aquaculture*. 1999. 179: 149-168.
- PORLA (Palm Oil Registration and Licensing Authority). Palm Oil Statistics. Ministry of Primary Industries, Kuala Lumpur, Malaysia. 2000.
- Siew, W.L. Characteristics and Uses of Malaysian Palm Kernel Cake.
 PORIM Technology Series, No. 14, Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia. 1989.
- 28. Hassan, O.A, and Yeong, S.W. By-products as animal feedstuffs. In: Oil Palm and The Environment: A Malaysian Perspective. Eds. Gurmit, S., Lim,

K.H., Teo, L. and Lee, D.K.), Malaysian Oil Palm Growers' Council, Malaysia. 225-239. 1999.

- Yeong, S.W. Palm oil by-products as feeds for poultry. Proceedings of the National Symposium on Oil Palm By-products for Agro-based Industries, Palm Oil Research Institute of Malaysia, Malaysia. 175-186. 1985.
- Yeong, S.W., Mukerjee, T.K. and Hutagalung, R.I., 1981. The nutritive value of palm kernel cake as feedstuffs for poultry. Proc. National Workshop of oil Palm By-product Utilization. Palm oil Res. of Malaysia (PORIM), Kuala Lumpur, pp. 100-107.
- Babatunde, G.M., Fetuga, B.L., Odumosu, O. and Oyenuga, V.A. Palm kernel meal as a major protein concentrate in the diets of pigs in the tropics. *Journal of the Science of Food and Agriculture*. 1975. 26: 1279-1291.
- Rhule, S.W.A. Growth rate and carcass characteristics of pigs fed on diets containing palm kernel cake. *Animal Feed Science and Technology*. 1996. 61: 167-172.
- 33. Agunbiade, J.A., Wiseman, J. and Cole, D.J.A. Energy and nutrient use of palm kernels, palm kernel meals and palm kernel oil in diets for growing pigs. *Animal Feed Science and Technology*. 1999. 80: 165-181.
- 34. Fetuga, B.L., Babatunde, G.M. and Oyenuga, V.A. The value of palm kernel meal in finishing diets for pigs. The effect of varying the proportion of protein contribution from blood meal and palm kernel meal on performance and carcass quality of finishing pigs. *Journal of Agricultural Science*. 1977. 88: 655-661.
- Yamamoto, T. Aquaculture in Asia: The 1989 APO Aquaculture Survey. In Liao et al. (eds.) Aquaculture in Asia: Proceedings of the 1990 APO Symposium on Aquaculture TFRI Conference. 1992.

- 36. Ong, K. S. Aquaculture development in Malaysia in the 80's. Risalah Perikanan Bil. 18. Jabatan Perikanan, Kementerian Pertanian Malaysia. 1988.
- Chua, T. E. and Tong, S. E. Floating fishpens for rearing fishes in coastal waters, reservoir and mining pools in Malaysia. Fisheries Bull. no. 20, Malaysia, Ministry of Agriculture. 1977.
- Sukarno, W., Raja Muhamad Nordin, R. O., Che Omar, M. H., and Rosdi, M. N. Tukun tiruan Malaysia. Jabatan Perikanan Malaysia. Kuala Terengganu: Percetakan Nasional Malaysia Berhad. 1994.
- Anonymous. Asian markets for reef fish. Aqua Farm News. 1992. X(3): 15-16.
- 40. UNDP/FAO. *Fish Feed Technology*. Rome: United Nations Development Programme, Food and Agriculture Organization of the United Nations. 1980.
- 41. De Silva, S.S. and Anderson, T.A. Fish nutrition in aquaculture, London: Chapman & Hall, 319 pp. 1995.
- 42. Voet, D. and Voet, J.G. Biochemistry. London: John Wiley & Sons, Inc. 1177pp. 1990.
- 43. Morrison, R.T. and Boyd, R.N. Organic chemistry. New York: Allyn and Bacon, Inc. 1320pp. 1983.
- 44. Mertz, E.T. Fish Nutrition. In, Halver, J.E (Ed.), New York: Academic Press, Inc. 1972.
- Cowey, C.B. and Luquet, P. Physiological basis of protein requirements of fishes. Critical analysis of allowances. In: IVth International Symposium of Protein Metabolism and Nutrition, France: Clermont-Ferrand. 365-384. 1983.

- Cuzon, G. and Fuchs, J. Preliminary nutritional studies of seabass *Lates calcarifer* (Bloch): protein and lipid requirements. In: Program and Abstracts, 19th Annual Conf. & Expo, Hawaii: World Aquaculture Society. 15-16. 1988.
- Tucker, J., MacKinnon, M., Russel, D., O'Brien, J and Cazzola, E. Growth of juvenile barramundi (*Lates calcarifer*) on dry feeds. *Progressive Fish-Culturist*. 1988. 50: 81-85.
- 48. Sakaras, W.M., Boonyaratpalin, M., Unprasert, N. and Kumpang, P. Optimum dietary protein/energy ratio in seabass feed. I. Technical Paper No. 7. Rayong Brackishwater Fisheries Station, Thailand. 1988.
- Sakaras, W.M., Boonyaratpalin, M. and Unprasert, N. Optimum dietary protein/energy ratio in seabass feed. II. Technical Paper No. 8. Rayong Brackishwater Fisheries Station, Thailand. 1989.
- Wong, F.J. and Chou, R. Dietary protein requirement of early grow-out seabass (*Lates calcarifer*), Bloch) and some observations on the performance of two practical formulated feeds. *Journal of Primary Industry*. 1989. 17: 98-111.
- Boonyaratpalin, M. Nutritional studies on seabass (*Lates calcarifer*). In: (S.S. De Silva, Editor), Fish Nutrition in Asia, Manila, Philippines: Asian Fisheries Society. 33-41. 1991.
- SEAFDEC. In: (M.N., Bautista, Editor), Feeds and Feeding of Milkfish, Nile Tilapia, Asian Sea Bass and Tiger Shrimp. SEAFDEC Aquaculture Department, Iloilo, Philippines. 1994.
- 53. Boonyaratpalin, M. Nutrient requirements of marine food fish cultured in Southeast Asia. *Aquaculture*. 1997. 151: 283-313.

- 54. Catacutan, M.R. and Coloso, R.M. Effect of dietary protein to energy ratios on growth, survival and body composition of juvenile Asian sebass, *Lates calcarifer*. *Aquaculture*. 1994. 131: 125-133.
- 55. Ng, W-K. and Chen, M-L. Replacement of Soybean Meal with Palm Kernel Meal in Practical Diets for Hybrid Asian-African Catfish, *Clarias macrocephalus* X *C. gariepinus. Journal of Applied Aquaculture.* 2002. 12(4): 67-76.
- Minson, D.J. Forage in Ruminant Nutrition. Academic Press: San Diego, 483 pp. 1990.
- Thomson, N.S. In: Wood and Agricultural Residues. Soltes, E.J. Ed., Academic Press: New York. 1983.
- 58. Barros, M.M., Lim C. and Klesius, P.H. Effect of soybean meal replacement by cottonseed meal and iron supplementation on growth, immune response and resistance of Channel Catfish (*Ictalurus punctatus*) to *Edwardsiella ictaluri* challenge. *Aquaculture*. 2002. 207: 263–279.
- De la Higuera, M., Garcia-Gallego, M., Sanz, A., Cardenete, G., Suarez, M.D., Moyano, F.J. Evaluation of lupin seed meal as an alternative protein source in feeding of rainbow trout (*Salmo gairdneri*). *Aquaculture*. 1988. 71: 37–50.
- 60. Borlongan, I.G., Eusebio, P.S. and Welsh, T. Potential of feed pea (*Pisum sativum*) meal as a protein source in practical diets for milkfish (*Chanos chanos* Forsskal). *Aquaculture*. 2003. 225: 89–98.
- 61. Food and Agricultural Organisation, 1998. Quarterly bulletin of statistics, vol. 11, no. 1/2.
- 62. Food and Agricultural Organization, 1996. Trade Yearbook, vol. 50.

- Ministry of Agriculture, Fisheries and Food. In: Feed Composition. UK Tables of Feed Composition and Nutritive Value for Ruminants, 2nd ed, Chalcombe Publications, Nr. Canterbury, UK. 1992.
- 64. Panigrahi, S. and Powell, C.J. Effect of substituting fish meal with palm kernel meal on growth and food utilization of the Nile tilapia, *Oreochromis niloticus*. *The Israeli Journal of Aquaculture-Bamidegh*. 1991. 45: 113-119.
- 65. Saad, C.R., Cheah, S.H. and Kamarudin, M.S. The use of Palm Kernel Cake (PKC) in diets of Red Tilapia (*Oreochromis niloticus*). Eds. Japar Sidik, F.M., Yussoff, M.S., Mohd Zaki and T. Petr. In: Fisheries and the Environment: Beyond 2000. Universiti Putra Malaysia, Serdang, Malaysia. 269-274. 1997.
- Lim, H-A., Ng, W-K., Lim, S-L. and Ibrahim, C.O. Contamination of palm kernel meal with *Aspergillus flavus* affects its nutritive value in pelleted feed for tilapia, *Oreochromis mossambicus*. *Aquaculture Research*. 2001. 32(11): 895-905.
- 67. Omoregie, E. and Ogbemudia, F.I. Effect of substituting fish meal with palm kernel meal on growth and food utilization of the Nile tilapia, *Oreochromis niloticus*. *The Israeli Journal of Aquaculture-Bamidegh*. 1993. 45: 113-119.
- Saad, C.R., Cheah S.H. and Hashimi, M.A.A. The Use of Palm Kernel Cake (PKC) in Catfish Practical Diets. Eds. L.M. Chou, A.D. Munro, T.J. Lam, T.W. Chen, L.K.K. Cheong, J.K. ding, K.K. Hooi, H.W. Khoo, V.P.E. Phang, K.F. Shim and C.H. Tan. In Proceedings of the Third Asian Fisheries Forum, Asian Fisheries Society, Manila, Philippines. 653-655. 1994.
- Omoregie, E. Utilization and nutrient digestibility of mango seeds and palm kernel meal by juvenile *Labeo senegalensis* (Antheriniformes: Cyprinidae). *Aquaculture Research.* 2001. 32: 681-687.

- 70. O'Mara, F.P., Mulligan, F.J., Cronin, E.J., Rath, M. and Caffrey, P.J. The nutritive value of palm kernel meal measured in vivo and using rumen fluid and enzymatic techniques. *Livestock Production Science*. 1999. 60: 305–316.
- 71. Widart, W. and Gascon, J.P. Study of the composition of *Elaeis guineensis* Jacq.oil. Possibilities for improvement. *Oleagineux*. 1975. 30(10): 401-408.
- 72. George, S. and Arumughan, C. Distribution of lipids in exocarp and mesocarp of three varieties of oil palm fruit (*Elaeis guineensis*). *Journal of the Science of Food and Agriculture*. 1991. 56(2): 210-222.
- George, S. and Arumughan, C. Positional distribution of fatty acids in the triacylglycerols of the developing palm fruit. *Journal of American Oil Chemistry Society*. 1993. 70(12): 1255-1258.
- Boraa, P.S, Rochaa, R.V.M., Narain, N, Moreira-Monteirob, A.C. and Moreira, R.A. Characterization of principal nutritional components of Brazilian oil palm (*Elaeis guineensis*) fruits. *Bioresource Technology*. 2003. 87:1-5.
- 75. Dusterhoft, E.M., Voragen, A.G.J. and Engels, F.M. Non-starch polysaccharides from sun-flower (*Helianthus annuus*) and palm kernel (*Elaies guineensis*) meal-preparation of cell wall material and extraction of polysaccharide fractions. *Journal of the Science of Food and Agriculture*. 1991. 55: 411-422.
- 76. Hutagalung R.I., Mohd-Mahyudin and Syed-Jalaludin. Feeds for farm animals from oil palm. Proceedings of the International Conference Oil Palm in Agriculture in Eighties. Kuala Lumpur, Malaysia. Vol. 2: 609-622. 1982.
- Mc Donald, P., Edwards, R.A. and Greenhalgh, J.F.G. Palm Kernel Meal. In Animal Nutrition. 4th ed., Harlow: Longman. 462-463. 1988.

- 78. Duran, A.O., Lozano, E. and Reyes, E. Use of African palm residue as an energy source in starter, grower and finishing phases of pigs. *Livestock Research and Rural Development*. 1990. 2: 43-50.
- Maltz, M.A. Protein Food Supplements Recent Advances Food Technology Review No.54. Park Ridge, New Jersey: Noyes Data Corporation. 82 – 250. 1981.
- Hofland, G.W., de Rijke, A., Thiering, R., van der Wielen, L.A.M and Witkamp, G.-J. Isoelectric precipitation of soybean protein using carbon dioxide as a volatile acid. *Journal of Chromatography B*. 2000. 743: 357– 368.
- Buentello, J. A., Gatlin III, D. M. and Dale, B. E. Evaluation of Coastal Bermuda Grass Protein Isolate as a Substitute for Fishmeal in Practical Diets for Channel catfish *Ictalurus punctatus*. *Journal of the World Aquaculture Society*. 1997. 28: 52-61.
- Little, E.C.S. Handbook of utilization of aquatic plants. FAO Fisheries Technical Paper. Food and Agriculture Organization of the United Nation, Rome. (187). 176. 1979.
- Ma, C.-Y., Liu, W.-S., Kwok, K.C. and Kwok, F. Isolation and characterization of proteins from soymilk residue (okara). *Food Research International*. 1997. 29(8): 799-805.
- Molina Ortiz, S.E. and Wagner, J.R. Hydrolysates of native and modified soy protein isolates: structural characteristics, solubility and foaming properties. *Food Research International*. 2002. 35: 511-518.
- Sorgentini, D.A. and Wagner, J.R. Comparative study of foaming properties of whey and isolate soybean proteins. *Food Research International*. 2002. 35: 721-729.

- Chove, B.E., Grandison, A.S. and Lewis, M.J. Emulsifying properties of soy protein isolate fractions obtained by isoelectric precipitation. *Journal of the Science of Food and Agriculture*. 2001. 81: 759-763.
- Bejosano, F.P and Corke, H. Properties of protein concentrates and hydrolysates from *Amaranthus* and Buckwheat. *Industrial Crops and Products*. 1999. 10: 175–183.
- Lqari, H. Vioque, J. Pedroche, J. and Millan, F. *Lupinus angustifolius* protein isolates:chemical composition, functional properties and protein characterization. *Food Chemistry*. 2002. 76: 349-356.
- El-Adawy, T.A., Rahma, E.H., El-Bedawey, A.A. and Gafar, A.F. Nutritional potential and functional properties of sweet and bitter lupin seed protein isolates. *Food Chemistry*. 2001. 74: 455-462.
- Moure, A., Sineiro, J. and Dominguez, H. Extraction and functionality of membrane-concentrated protein from defatted *Rosa rubiginosa* seeds. *Food Chemistry*. 2001. 74: 327-339.
- Ng, W-K, Lim, H-A, Lim, S-L and Ibrahim, C-O. Nutritive value of palm kernel meal pretreated with enzyme or fermented with *Trichoderma koningii* (Oudemans) as a dietary ingredient for red hybrid tilapia (*Oreochromis* sp.). *Aquaculture Research*. 2002. 33: 1199-1207.
- 91. Ng, W-K and Chong, K-K. The nutritive value of palm kernel meal and the effect of enzyme supplementation in practical diets for red hybrid tilapia (*Oreochromis* sp.). *Asian Fisheries Science*. 2002. 15: 167-176.
- 93. Schell, D.J., Torget, R., Power, A., Walter, P.J., Grohmann, K. and Hinman, N.D. A technical and economic analysis of acid-catalyzed steam explosion and dilute sulfuric acid pretreatments using wheat straw or aspen wood chips. *Applied Biochemistry and Biotechnology*. 1991. 28/29: 87-97.

- 94. Torget, R., Werdene, P., Himmel, M. and Grohmann, K. Dilute acid pretreatment of short rotation woody and herbaceous crops. *Applied Biochemistry and Biotechnology*. 1990. 24/25: 115-126.
- 95. Torget, R., Himmel, M. and Grohmann, K. Dilute sulfuric acid pretreatment of hardwood bark. *Bioresource Technology*. 1991. 35: 239-246.
- 96. Torget, R., Walter, P., Himmel, M. and Grohmann, K. Dilute-acid pretreatment of corn residues and short rotation woody crops. *Applied Biochemistry and Biotechnology*. 1991. 28/29: 75-86.
- Torget, R., Himmel, M. and Grohmann, K. Dilute-acid pretreatment of two short-rotation herbaceous crops. *Applied Biochemistry and Biotechnology*. 1992. 34/35: 115-123.
- 98. Sreedhara, N and Kurup, P.A. Effect of Hydrochloric Acid Treatment of Palm Kernel for Dehulling on the Nutritional Quality of Kernel protein. *Journal of the Science of Food and Agriculture*. 1998. 77: 435-440.
- 99. Sunstøl, F. Straw and other fibrous by-products. *Livestock Production and Science*. 1988. 19: 137-158.
- 100. Waiss, AC., Guggolz, J., Kohler, G.O., Walker Jr., H.G. and Garrett, W.N. Improving digestibility of straws for ruminant feed by aqueous ammonia. *Journal of Animal Science*. 1972. 35: 109-112.
- 101. Mason, V.C., Dhanoa, M.S., Hartley, R.D. and Keene, A.S. Relationships between chemical composition, digestibility in vitro and cell-wall degradability of wheat straw treated with different amount of ammonia and water at elevated temperature. *Animal Feed Science and Technology*. 1990. 27: 293-306.
- 102. Zaman, M.S., Owen, E. and Pike, D.J. The calculations method used for optimising conditions of treatment of barley straw with calcium hydroxide;

effects of level of calcium hydroxide and urea, moisture, treatment time and temperature on in vitro digestibility. *Animal Feed Science and Technology*. 1993. 45: 271-282.

- 103. Williams, P.E.V., Innes, G.M. and Brewer, A. Ammonia treatment of straw via hydrolysis of urea. I. Effect of dry matter and urea concentration on the rate of hydrolysis of urea. *Animal Feed Science and Technology*. 1984a. 11: 103–113.
- 104. Williams, P.E.V., Innes, G.M. and Brewer, A. Ammonia treatment of straw via the hydrolysis of urea. II- Additions of soya bean (urease), sodium hydroxide and molasses. Effects on the digestibility of urea-treated straw. *Animal Feed Science and Technology*. 1984b. 11: 115–124.
- Sahnoune, S., Besle, J.M. and Jouany, J.P. Treatment of wheat straw by hydrolysis of urea at low level of water addition. *Journal of Animal Science*. 1989. 2: 533–534.
- 106. Sahnoune, S., Besle, J.M., Chenost, M., Jouany, J.P. and Combes D. Treatment of straw with urea. I-Ureolysis in low water medium. *Animal Feed Science and Technology*. 1991. 34: 75–93.
- 107. Jackson, M.G. Review article: the alkali treatment of straws. *Animal Feed Science and Technology*. 1977. 2: 105-130.
- 108. Oji, U.I. and Mowat, D.N. Nutritive value of thermoammoniated and stemtreated maize stover. 1. Intake, digestibility and nitrogen retention. *Animal Feed Science and Technology*. 1979. 4: 177-186.
- Choct, M and Annison, G. Anti-nutritive effect of wheat pentosans in broiler chickens: roles of viscosity and gut microflora. *British Poultry Science*. 1992. 33: 821-834.

- Sundstol, F., Coxworth, E. and Mowat, D.N. Improving the nutritive value of straw and other low quality forages by treatment with ammonia. *World Animal Review*. 1978. 26: 13-21.
- 111. Park, D.L., Lee, L.S. and Price, R.L. Review of the decontamination of aflatoxins by ammoniation: current status and regulation. *Journal of Association of Official Analytical Chemists*. 1988. 71: 685-703.
- 112. Wylie, A.R.G. and Steen, R.W.J. Effect of anhydrous ammonia treatment on the chemical composition and nutritive value of grass hay and on intake and performance in beef steers. *Grass Forage Science*. 1988. 43: 79-86.
- Zorrila-Rios, J., Horn, G.W. and McNew, R.W. Nutritive value of ammoniated wheat straw fed to cattle. *Journal of Animal Science*. 1991. 69: 283-294.
- Dale, B. E., Henk, L.L. and Shiang, M. Fermentation of lignocellulosic materials treated by ammonia freeze-explosion. *Development of Industrial Microbiology*. 1985. 26: 223-233.
- 115. Holtzapple, M.T., Jun, J.-H., Ashok, G., Patibandla, S.L. and Dale, B.E. The ammonia freeze explosion process: a practical lignocellulose pretreatment. *Applied Biochemistry and Biotechnology*. 1991. 28/29: 59-74.
- Holtzapple, M.T. Lundeen, J.E., Sturgis, R., Lewis, J.E. and Dale, B.E. Pretreatment of lignocellulosic municipal solid waste by ammonia fiber explosion (AFEX). *Applied Biochemistry and Biotechnology*. 1992. 34/35: 5-21.
- 117. Dale , B.E. and Moreira, M.J. A freeze-explosion technique for increasing cellulose hydrolysis. *Biotechnology and Bioengineering Symposium*. 1982. 12: 31-34.

- Ashok, G. Ammonia fiber explosion (AFEX) treatment of grass. M.S. Thesis. Texas A&M University, College Station, TX. 1991.
- 119. Turner, N.D., Schelling, G.T., Greene, L.W. and Byers, F.M. Modification of in-vitro and in-situ dry matter digestibility of forages with chemical treatment. *Journal on Product of Agriculture*. 1990. 3: 83-87.
- Hagevoort, G. R., Byers, F.M., Holtzapple, M.T., Jun, J.-H., Greene, L.W. and Carstens, G.E. Enhancing the nutritive value of forages with an ammonia fiber explosion (AFEX) technique. *Journal of Animal Science*. 1990. 68(1): 584.
- 121. De la Rosa, L.B., Reshamwala, S., Latimer, V. Shawky, B.T., Dale, B.E. and Stuart, E.D. Integrated production of ethanol fuel and protein from coastal Bermuda grass. *Applied Biochemistry and Biotechnology*. 1994. 45/46: 483-497.
- Dale, B.E., Henk, L.L. and Shiang, M. Fermentations of lignocellulosic materials treated by ammonoa-freeze explosion. *Developments in Industrial Microbiology*. 1985. 26: 233.
- AOAC (Association of Official Analytical Chemists) Official Methods of Analysis (17th Ed.) W. Hormitz (Editor), Washington, DC: Association of Analytical Chemists, Inc. 2002.
- 124. Allred, M.C. and MacDonald, J.L. Vitamins and other nutrients Determination of sulphur amino acids and tryptophan in foods and food and feed ingredients: Collaborative study. *Journal of Association of Official Analytical Chemists.* 1988. 71: 603-606.
- 125. Greenberg, N.A. and Shipe, W.P. Comparison of the abilities of trichloroacetic, picric, sulfosalicyclic and tungstic acids to precipitate protein hydrolysates and proteins. *Journal of Food Science*. 1979. 44: 735-737.

- 126. Williams, K. and Barlow, C. Dietary requirement and optimal feeding practices for barramundi (*Lates calcarifer*). Final Report to Fisheries Research and Development Corporation, Australia. Project 92/63. 91. 1999a.
- 127. Strickland, J.D.H. and Parsons, T.R. A practical handbook of seawater analysis. Bulletin 167 (2nd Ed). Department of Fisheries and the Environment, Fisheries Research Board of Canada, Ottawa. 1972.
- 128. Araba, M. and Dale, N. Evaluation of protein solubility as an indicator of over-processing soybean meal. *Poultry Science*. 1990. 69: 76-83.
- Garling, D.L., Jr and Wilson, R.P. Optimum dietary protein to energy ratio for channel catfish fingerlings, *Ictalurus punctatus. Journal of Nutrition*. 1976. 106: 1368-1375.
- Buhler, D.R. and Halver, J.E. Carbohydrate requirements of Chinook salmon. Journal of Nutrition. 1961. 74: 307-317.
- Leary, D.F. and Lovell, R.T. Value of fibre in production-type diet for channel catfish. *Transactions of the American Fisheries Society*. 1975. 104: 328-332.
- 132. Hilton, J.W., Atkinson, J.L. and Slinger, S.J. Effect of increased dietary fibre on the growth of rainbow trout (*Salmo gairdneri*). *Canadian Journal of Fish and Aquatic Science*. 1983. 40: 81-85.
- 133. Williams, K. and Barlow, C. Fishmeal replacement in aquaculture feeds for barramundi: (i) Nutritive value of crystalline amino acids (ii) potential of meat meal to replace fishmeal. Final Report to Fisheries Research and Development Corporation, Australia. Project 95/069. 20-51. 1999b.
- Shiau, S.Y., J.L. Chuang and C.L. Sun. Inclusion of soybean meal in tilapia (Oreochromis niloticus X O. aureaus) diets at two protein levels. Aquaculture. 1987. 65: 251-261.

- 131. Andrews, J.W. and Page, J.W. Growth factors in the fish meal component of catfish diets. *Journal of Nutrition*. 1974. 104:1091-1096.
- 132. Dabrowski, K., and B. Kozak. The use of fish meal and soybean meal as a protein source in the diet of grass carp fry. *Aquaculture*. 1979. 18:107-114.
- 134. Shamshuddin, J., Muhrizal, S., Fauziah, I. and Husni, M. H. A. Effects of adding organic materials to an acid sulfate soil on the growth of cocoa (*Theobroma cacao* L.) seedlings. *Science of The Total Environment*. 2004. 323(1-3): 33-45.
- 135. Onuora, J.O and King, R.D. Enzymic solubilization of nitrogenous constituents of palm kernel cake. *Food Chemistry*. 1985. 17: 297-302.
- Wolf, W.J. and Cowan, J.C. Soybeans as a food source. Cleveland, Ohio: CRC Press. 1975.
- Pearson, A.M. Soy proteins. In, B.J.F. Hudson (Ed.), Developments in food proteins – 2. Essex: Applied Science Publishers. 67-108. 1983.
- Aghazu, B. U. C., Meissner, H. P., Ngoui, C. I. and Tannenbaum, S. R. Protein-rich flours from palm kernels. *Journal of Food Technology*. 1979. 14: 1-8.
- 139. Cornelius, J.A. Some technical aspects influencing the quality of palm kernels. *Journal of the Science of Food and Agriculture*. 1966. 17: 57-61.
- Cornelius, J.A. Processing of Oilpalm Fruit and its Products. Tropical Products Institute, Grays Inn Road, London, UK, pp 22-34. 1983.
- 141. Tang, T.S. and Teoh, P.K. Palm kernel oil extraction: The Malaysian experience. *Journal of American Oil Chemistry Society*. 1985. 62: 254-258.

- Barry, T.N. and Duncan, S.J. The role of condensed tannins in the nutritional value of *Lotus pedunclatus* for sheep. *British Journal of Nutrition*. 1984. 51: 485-491.
- 143. Barry, T.N. Condensed tannins: their role in ruminant protein and carbohydrate digestion and possible effects upon the rumen ecosystem. In: The Roles of Protozoa and Fungi in Ruminant Digestion. Eds. Nolan, J.V., Leng, R.A. and Demeyer, D.I. Penambul Books, Armidale, NSW, Australia, pp 153-169. 1989.
- 144. Yu, F., Barry, T.N., McNabb, W.C., Moughan, P.J. and Wilson, G.F. Effect of bound condensed tannin from cottonseed upon *in situ* protein solubility and dry matter digestion in the rumen. *Journal of the Science of Food and Agriculture*. 1995. 69: 311-320.
- 145. Pierpoint, W.S. *o*-Quinones formed in plant extracts. Their reactions with amino acid and peptides. *Biochemistry Journal*. 1969. 112:609-618.
- Sosulski, F.W. Organoleptic and nutritional effects of phenolic compounds on oilseed protein products: A review. *Journal of American Oil Chemistry Society.* 1979. 56: 711-715.
- Siebert, K.J., Troukhanova, N.V. and Lynn, P.Y. Nature of polyphenolprotein interactions. *Journal of Agriculture and Food Chemistry*. 1996. 44: 80-85.
- 148. Dabrowski, K., Hassard, S., Quinn, J., Pitcher, T.J. and Flinn, A.M. Effect of *Geotrichum candidum* protein substitution in pelleted fish feed on the growth of rainbow trout (*Salmo gairdneri* Rich.) and on utilization of the diet. *Aquaculture*. 1980. 21: 213-232.
- 149. Fickler, J. Fish meal; High protein does not stand for high quality! *Feed International*, 2002. 23(7): 13-16.

150. Boonyaratpalin, M., Promkunthong, W. and Hunter, B. Effects of enzyme pre-treatment on in vitro glucose solubility of Asian plant by-products and growth and digestibility of oil palm expeller meal by *Oreochromis niloticus* (Nile tilapia). In: W.V. Hartingsveldt (Ed), Proceedings of the Third European Symposium on Feed Enzymes, TNO Voeding, The Netherlands. 86-92. 2000.