

PRODUCTION OF BIODIESEL FROM WASTE COOKING OIL BY
IMMOBILIZED LIPASE IN PVA-ALGINATE-SULFATE BEADS

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DEDICATION

Specially dedicated to my parents, Alhaji Ali Alhaji Deba and Hajiya Hafsat Mahdi, my relatives and all my well wishers whom aspired me to be a better person.

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Certainly all praise be to Allah, whom I praise and seek his forgiveness. I seek refuge with him from the evil of my soul and from the wickedness of my deeds. Surely, Whomever He has guided shall never go astray, and whomever He allows astray shall never be guided. I bear witness that there is no deity of being worshipped except Allah whom partnership shall never be associated with him. I bear witness Muhammad (S.A.W) is his Messenger and slave. May the peace and blessings of Allah be upon Him, his family and Companions and those that follow through his path till the last day.

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ABSTRACT

With the huge varieties of oil produced worldwide, management and disposal are the major challenges facing the production. This is due to contamination of water bodies and land resources as a result of incessant disposal. Thus embarking on waste oil utilization in biodiesel production by immobilized enzymatic transesterification is a triple step forward towards the waste management. Therefore in these studies, *Candida rugosa* (Type VII, 1176 units/mg) was immobilized in PVA-Alginate-Sulphate beads by entrapment and cross linking method. Transesterification process was conducted using the immobilized lipase. 32 treatment combinations were generated by the Design Expert software version 6.0.4 using 2 level fractional factorial designs. The treatment combination; 1:6 oil to methanol, 2.5 v/v, 60 U at 50 °C under 200 rpm was found to be the optimum condition that gave the highest production of 4.516g/L, 1.65 g/L and 1.6 g/L, corresponding to 45 %, 10.65 %, 10.6 %, oleic acid methyl ester, palmitic acid methyl ester and linoleic acid methyl ester respectively. In another vain the treatment combination used for the optimum production using immobilized lipase was applied to the free lipase resulting to 0.234 g/L, 0.130 g/L, 0.049 g/L, corresponding to 2.34%, 1.30%, 0.49 %, oleic acid methyl ester, palmitic acid methyl ester and linoleic acid methyl ester respectively. Approximately, the immobilized lipase performed 15 folds better than the free lipase. Enzyme assay was conducted and compared between free lipase and immobilized lipase over 3 hours of reaction using spectrophotometer, absorbance was read at 410nm. Immobilized lipase in general maintained their activity after an hour of the reaction. In other higher concentrations, both the free and the immobilized showed a steep fall with time. FESEM-EDX was also carried out to study the immobilization matrix morphology which revealed the native structure of the entrapped *Candida rugosa* lipase. Furthermore, reusability test of the immobilized lipase was conducted and found capable of maintaining a production of five consecutive cycles

ABSTRAK

Penghasilan pelbagai jenis minyak diseluruh dunia mendatangkan cabaran utama bagi mencari penyelesaian terbaik untuk menguruskan sisa sampingan hasil daripada industri tersebut. Ini adalah disebabkan oleh pencemaran air dan alam sekitar akibat pelupusan secara berterusan. Oleh itu, kaedah transesterifikasi dengan menggunakan enzim tersekat-gerak merupakan salah satu penyelesaian kepada rawatan sisa sampingan minyak. Dalam eksperimen ini, *Candida rugosa* (jenis VII, 1176 unit/mg) telah disekatgerak pada manik PVA-Alginat-Sulfat dengan menggunakan kaedah pengurangan dan hubung pangkah. Enzim lipase tersekatgerak telah digunakan dalam proses transesterifikasi. Sebanyak 32 kombinasi transesterifikasi telah dihasilkan dengan menggunakan reka bentuk factorial 2-level (2-level fractional factorial) oleh perisian *Design Expert* versi 6.0.4. Penghasilan biodiesel yang optima ialah 4.516g/L, 1.65 g/L dan 1.6 g/L merujuk kepada 45 % asid oleik metil ester, 10.65 % asid palmitik metil ester dan 10.6 % ester asid linoleik metil ester dengan keadaan nisbah minyak kepada metanol (1:6) , 2.5 v/v kandungan air, 60 U jumlah enzim, suhu 50 °C dan kelajuan 200 rpm. Manakala, penghasilan optima biodiesel bagi lipase bebas telah menghasilkan 0.234 g/L, 0.130 g/L dan 0.049 g/L, iaitu 2.34% asid oleik metil ester, 1.30% asid palmitik metil ester dan 0.49 % asid linoleik metil ester pada keadaan yang sama. Secara purata, lipase tersekatgerak menunjukkan prestasi penghasilan biodiesel 15 kali ganda lebih baik berbanding lipase bebas. Ujian enzim dilakukan bagi membandingkan aktiviti enzim antara lipase tersekatgerak dan lipase bebas dalam tindak balas selama tiga jam. Bacaan spektrofotometer ialah 410 nm. Aktiviti enzim lipase tersekatgerak pada 0.1-0.3 M menjadi stabil selepas sejam tindak balas berlaku. Dalam kepekatan yang lebih tinggi, aktiviti enzim bagi lipase bebas dan lipase tersekatgerak menunjukkan penurunan berkadar dengan masa. FESEM-EDX juga digunakan untuk mengkaji morfologi sekatgerak matriks dimana ia menonjolkan struktur natif lipase *Candida rugosa* yang terperangkap. Selain itu, lipase sekatgerak telah dibuktikan dalam eksperimen ini boleh digunakan bagi penghasilan biodiesel sebanyak lima kitaran secara berterusan.

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GLOSSARY OF TERMS

ANOVA	Analysis of variance
ASTM	American society of testing materials
COD	Chemical oxygen demand
EDX	Energy dispersive x-ray spectrometer
FAME	Fatty acid methyl ester
FE-SEM	Field emission scanning electron microscope
FFA	Free fatty acids
KOH	Potassium hydroxide
MGs	Monoglycerides
PBR	Packed bed reactor
p-NP	p-nitrophenol
p-NPP	p-nitrophenol palmitate
PUFAs	Poly unsaturated fatty acids
PVA	Poly vinyl alcohol
RSM	Response surface methodology
TGs	Triglycerides
WCO	Waste cooking oil

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CHAPTER 1

INTRODUCTION

1.0 Background of the study

Vegetable oils have become a focus of attention as a potential renewable source for the production of an alternative source of fuel for petroleum based diesel. This is owing to continuous world-wide consumption of petroleum reserves in addition to environmental consequences attached to the emission of exhaust gases from diesel engines (Noureddini *et al.*, 2001). Biodiesel is a renewable fuel that can be synthesized from edible, non-edible and waste oils. Vast varieties of starting materials come from edible oil, such as; soybean oil, rapeseed oil, cotton seed oil, sunflower oil, palm oil and restaurant kitchen wastes. In countries, such as India, such oils are not in abundance, choice of other starting materials like jatropha can be appropriate (Gayatri *et al.*, 2010).

The Methyl and ethyl esters of fatty acids, also well-known as biodiesel, are nontoxic, non-polluting and biodegradable fuel, and therefore considered as a good substitute for fossil petroleum diesel. Viscosity, cetane number, energy content, and other fuel characteristics of biodiesel are also similar to those of petroleum-based

diesel fuel (Mittelback and Tritthart, 1988). Furthermore, biodiesel is basically sulfur free and is considered ultra-sulfur fuel quality and the diesel engines fueled with the such fuel emit significantly fewer hydrocarbons, particulate, carbon monoxide and carbon dioxide than those operating on conventional fossil fuel diesel. NO_x emissions, however, are slightly higher than those of diesel engines operating on conventional diesel fuel (Schumacher *et al.*, 1996 and Ali *et al.*, 1995).

Transesterification of vegetable oils and animal fat can be catalyzed by chemical and/or biological catalysts, such as acid, base and enzymes such as lipase. However, the biodiesel produced by chemical catalyst has several drawbacks such as difficulty in recovery of glycerol, removal of acid or base catalysts from the product and the treatment of wastewater (Freeman *et al.*, 1984; Mittelbach, 1990; Basri *et al.*, 1997; Fukuda *et al.*, 2001). On the contrary, Enzymatic transesterification of triglycerides using immobilized lipase is a good substitute to chemical process owing to its; continuous operation, improved stability, reuse, chance of better control of reaction, high purity and product yields and hence more positive economic factors can be expected (Ruckenstein and Wang, 1993).

Therefore this study focuses on the production of biodiesel from waste cooking oil (WCO) by immobilized *Candida rugosa* lipase in PVA-alginate-sulfate beads. The immobilization would enhance the stability and re-usage of the enzyme while making the whole production process an eco friendly one.

1.3 Statement of the problem

Due to the fact that there are a lot of environmental problems associated with the use of fossil fuels, biodiesel can be expected as a replacement for conventional fuel. At present, biodiesel has been produced chemically using vegetable oil in USA

and Europe (Mamoru *et al.*, 2001). However, requirement of removal of catalyst and extreme energy requirement are the key drawbacks of such chemical process (Mamoru *et al.*, 2001).

Because the enzymatic method may overcome the problems for the reaction, numerous researches have been carried out using lipase (Nelson *et al.*, 1996; Selmi *et al.*, 1998; Shimada *et al.*, 1999; Charuchinda *et al.*, 2011). However the production of biodiesel fuel by enzymatic means has not yet been adopted industrially due to the high cost of enzymes (Mamoru *et al.*, 2001), hence the need for additional researches using immobilized enzymes to come up with improved production process that is cost reduced.

1.1 Objectives of the research

- To immobilize lipase from *Candida rugosa* on PVA-alginate-sulfate beads.
- To study the optimum conditions of transesterification process with the immobilized lipase by using 2-Level fractional factorial design.
- To study the performance of free and immobilized lipase in the enhancement of biodiesel production.

1.2 Scope of the research

In this study, the type of lipase chosen is *Candida rugosa* (Type VII, 1176 units/mg). The lipase was made more stable by entrapment and cross linking in

PVA-alginate sulfate beads. With this development biodiesel is produced by enzymatic transesterification using waste cooking oil as substrate. The optimum condition of this production is only considered based on the parameters selected for this study, namely, oil to methanol molar ratio, temperature, agitation, water content and enzyme concentration. The activity of the lipase would be determined by the degradation of p-NNP. Figure 1.1 illustrates the summary of the scope.

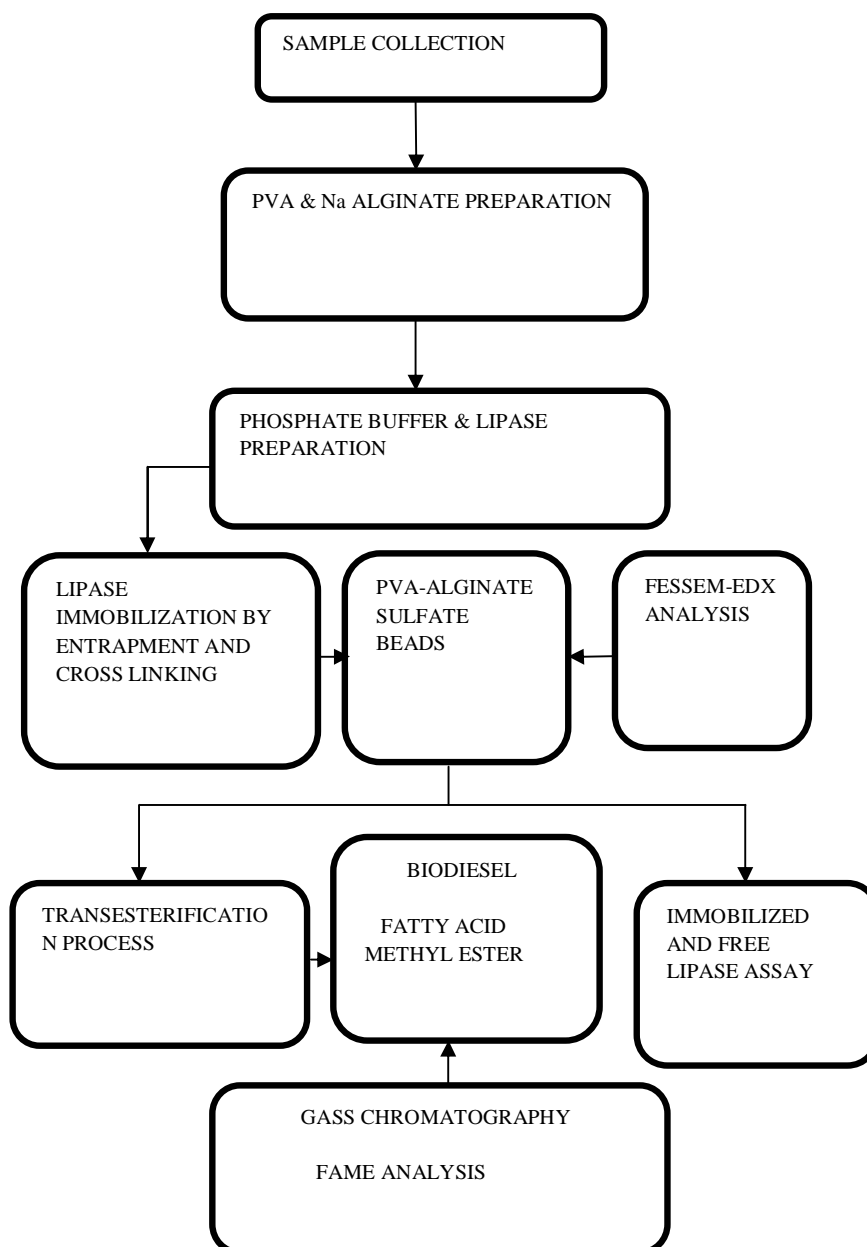


Figure 1.1 Scope of the research in summary

1.4 Research significance

From the invention of PVA-alginate-sulfate method to date no attempt had been made to transesterify waste cooking oil to produce biofuel. Therefore, this study emerges to establish the effectiveness of immobilized *Candida rugosa* lipase by this technique towards biodiesel production. Explored parameters from other researchers such as oil to methanol molar ratio, temperature, water content, agitation and enzyme concentration besides activity and reusability would also be studied.

The recycling process of waste cooking in order to produce biodiesel is of paramount importance considering the environmental concerned in the management of such oils as a result of pollution and contamination problems associated with its disposal (Arjun *et al.*, 2008), therefore embarking on the use of the waste cooking oil would serve as a good step towards management of such oil and at the same time attaining biodiesel production.

REFERENCES

- Abigor, R., Uadia, P., Foglia, T., Haas, M., Jones, K., Okpefa, E., Obibuzor, J. and Bafor, M., (2000). Lipase-catalysed production of biodiesel fuel from some Nigerian lauric oils. *Biochemical Society Transactions*. 28: 979–981.
- Akoh, C.C. and Min, D.B., .Microbial Lipases. and .Enzymatic Interesterification. *In Food Lipids- Chemistry, Nutrition and Biotechnology* (1998), Marcel Deccer, Inc, New York, p. 641-698.
- Akoh, C. C., Lee, G. C. and Shaw, J. F.(2004). Protein engineering and applications of *Candida rugosa* lipase isoforms. *Lipids*. 39: 513–526.
- Akoh, C.C., Chang, S.W., Lee, G.C. and Shaw, J.F. (2007). Enzymatic approach to biodiesel production. *Journal of Agricultural and Food Chemistry*. 55: 8995-9005.
- Alberghina, L., Schmid, R.D., Verger, R.(Eds.) (1991). Lipases: structure, mechanism and genetic engineering Weinheim: Wiley-VCH Verlag GmbH & Co. KGaA.
- Ali, Y., Hanna, M.A. and Leviticus, L.I. (1995). Emissions and power characteristics of diesel engines on methyl soyate and diesel fuel blends. *Technology*. 52(2): 185–195
- Antczak, M.S., Kubiak, A., Antczak T. and Bielecki, S. (2009). Enzymatic biodiesel synthesis-Key factors affecting efficiency of the process. *Renewable Energy*. 34:1185-1194.

- Arjun, B., Chhetri, K. Watts, C. and Rafiqul Islam, M. (2008). Waste Cooking Oil as an Alternate Feedstock for Biodiesel Production. *Energies*. 1: 3-18.
- Balat, M. and Havva, B. (2010). Progress in biodiesel processing. *Applied Energy*. 6 (6):1815-1835.
- Ban, K., Kaieda, M., Matsumoto, T., Kondo, A. and Fukuda, H.(2001). Whole-cell biocatalyst for Biodiesel fuel production utilizing *Rhizopus oryzae* cells immobilized within biomass support particles. *Biochemical Engineering*. 8: 39–43.
- Basri, M., Heng, A.C., Razak, C.N.A., Wan Yunus, W. M. Z., Ahmad, M, Rahman, R.N.A., Ampon, K. and Salleh, A. B. (1997). Alcoholysis of palm oil midfraction by lipase from *Rhizopus rhizopodiformis*. *Journal of the American Oil Chemists' Society*. 74: 113-116.
- Biocyclopedia.com (2012). Section: General Biotechnology / Microbial Biotechnology(http://www.biocyclopedia.com/index/biotechnology/microbial_biotehnology/enzyme_technology/biotech_enzyme_immobilization.php#gg)
- Bornscheuer, U.T. (Ed.) (2000). Enzymes in lipid modification Weinheim: Wiley-VCH.
- Charuchinda, S., Suthianthong P. and Chulalaksananukul, W. (2011). Immobilization of lipase onto *Cyperus Papyrus L.* for biodiesel production by transesterification and hydrolysis esterification. *Journal of Metals, materials and minerals*. 22(1): 127-133.
- Chen, J.W., Wu, W.T., (2003). Regeneration of immobilized *Candida antarctica* lipase for transesterification. *Journal of Bioscience and Bioengineering*. 95: 466–469.

- Chiang C.J., Hsiau L.T. and Lee W.C. (2004) Immobilization of cell-associated enzymes by entrapment in polymethacrylamide beads. *Biotechnology Techniques*. 11(2): 121-125.
- Chen, Y., Bo, X., Jie, C., Yan, F., Pengmei, L., Xuwei, W., (2009). Synthesis of biodiesel from waste cooking oil using immobilized lipase in fixed bed reactor. *Energy Conversion and Management*. 50: 668–673
- Chiou, S. H. and Wu, W.T. (2004). Immobilization of *Candida rugosa* lipase on chitosan with activation of the hydroxyl groups. *Biomaterials*. 25(2): 197-204
- Chirvase A.A., Luminita, T., Nicoleta, R. and Irina L.(2007). Progress in Vegetable Oils Enzymatic Transesterification to Biodiesel - Case Study. Romanian National Agency for Research under the Second National Program, Project 61-032/2007.
- Coggon, R., Vasudevan, P.T. and Sanchez, F. (2007). Enzymatic transesterification of olive oil and its precursors. *Biocatalysis and Biotransformation*. 25: 135-143.
- Cygler M. and Schrag, J. D. (1999). Structure and conformational flexibility of *Candida rugosa* lipase. *Biochimica et Biophysica Acta* 1441: 205-214
- Database of Oil Yielding Plants. 2004. Retrieved from the World Wide Web July 15 2009. <http://www.mnre.gov.in/list/oil-plants.pdf>.
- Dean, A. and Voss, D. (1999). Design and Analysis of Experiment. Springer text in Statistics. New York: Springer -Verlag. New York Inc.

- Demirbas, A. (2005). Biodiesel production from vegetable oils via catalytic and non-catalytic supercritical methanol transesterification methods. *Progress in Energy and Combustion Science*. 31: 466–487.
- Demirbas, A. (2008). Comparison of transesterification methods for production of biodiesel from vegetable oils and fats. *Energy Conversion and Management*. 49: 125–130.
- Dizge, N., Aydiner, C., Imer, D.Y., Bayramoglu, M., Tanriseven, A. and Keskinler, B. (2009). Biodiesel production from sunflower, soybean, and waste cooking oils by transesterification using lipase immobilized onto a novel microporous polymer. *Bioresource Technology*. 100: 1983–91.
- Dossat, V., Combes, D. and Marty, A. (1999). Continuous enzymatic transesterification of high oleic sunflower oil in a packed bed reactor: influence of the glycerol production. *Enzyme and Microbial Technology*. 25: 194-200.
- Fakuda, H., Kondo, A. and Noda, H. (2001). Biodiesel fuel production by transesterification of oils. *Journal of Bioscience and Bioengineering*. 92: 405-416.
- Fakuda, H., Hama, S. and Tamalampudi, S. (2008). Whole-cell biocatalyst for biodiesel fuel production. *Trends in Biotechnology*. 26l: 668-673.
- Fakuda, H., Kondo, A. and Tamalampudi, S. (2009). Bioenergy: Sustainable fuels from biomass by yeast and fungal whole-cell biocatalyst *Biochemical Engineering Journal* 44: 2- 12.
- Farahat, S. M., Rabie, A. M. and Faras, A. A. (1990). Evaluation of the proteolytic and lipolytic activity of different *Penicillium roqueforti* strains. *Food Chemistry*. 36: 169–80.

- Freeman, B., Pryde, E. H. and Mounts, T. L. (1984). Variables affecting the yields of fatty acid esters from transesterified vegetable oils. *Journal of the American Oil Chemists' Society*. 61: 1638-1643.
- Gayatri, Nahak., Chandra, K. M., Mohapatra, N. K. and Sahu, R. K. (2010). Enzymatic transesterification of lipases obtained from bacterial and plant sources. *Continental Journal of Biomedical Sciences*. 4: 21-36.
- Ghaly, A. E., Dave, D., Brooks, M.S. and Budge S. (2010). Production of biodiesel by Enzymatic Transesterification: Review. *American Journal of Biochemistry and Biotechnology*. 6(2): 54-76.
- Godfrey, T, Reichelt, J. (1983). *Industrial applications. In: Industrial enzymology applications of enzymes in industry*. London: The Nature Press London. pp 170-465.
- Gog, A., Marius, R., Monica, T., Csaba, P. and Florin, D. I. (2012). Biodiesel production using enzymatic transesterification - Current state and perspectives. *Renewable Energy*. 39: 10-16.
- Grant, W. D., Mwatha, W. E. and Jones, B. E. (1990). *FEMS Microbiology Letters*. 75: 255-269
- Gupta, R., Rathi, P. and Bradoo, S.(2003). Lipase mediated upgradation of dietary fats and oils. *Critical Reviews in Food Science and Nutrition*. 43: 635-644.
- Hakki Mevlut Ozcan & Ayten Sagiroglu (2009): Production of Ricinoleic Acid from Castor Oil by Immobilised Lipases, *Preparative Biochemistry and Biotechnology*. 39:2, 170-182.
- Hama, S., Yamaji, H., Kaieda, M., Oda, M., Kondo, A. and Fukuda, H. (2004). Effect of fatty acid membrane composition on whole cell biocatalysts for biodiesel-fuel production. *Biochemical Engineering Journal*. 21: 155-160.

- Hashimoto, S. and Furukawa, K. (1987). Immobilization of activated sludge by PVA-boric acid method. *Biotechnology and Bioengineering*. 30: 52-9.
- Hasan, F., Aamer, A. S., Abdul, H. (2006). Industrial applications of microbial lipases. *Enzyme and Microbial Technology*. 39: 235-251.
- Hsu, A. F., Jones, K., Foglia, T.A. and Marmer, W.N. (2002). Immobilized lipase-catalysed production of alkyl esters of restaurant grease as biodiesel. *Biotechnology and Applied Biochemistry*. 36(3): 181-186.
- Iso, M., Chen, B., Eguchi, M., Kudo, T. and Shrestha, S. (2001). Production of biodiesel fuel from triglycerides and alcohol using immobilized lipase. *Journal of Molecular Catalysis B: Enzymatic*. 16: 53-58.
- Jaeger, K.E. and Eggert, T. (2002). Lipases for biotechnology. *Current Opinion in Biotechnology*. 13: 390-397.
- Kaieda, M., Samukawa, T., Matsumoto, T, Ban, K., Kondo, A., Shimada, Y., Noda, H., Nomoto, F., Ohtsuka, K., Izumoto, E. and Fukuda, H. (1999). Biodiesel fuel production from plant oil catalysed by *Rhizopus oryzae* in a water-containing system without an organic solvent. *Journal of Bioscience and Bioengineering*. 88: 627- 631.
- Kaieda, M., Samukawa, T., Kondo, A. and Fukuda, H. (2001). Effect of methanol and water contents on production of biodiesel fuel from plant oil catalyzed by various lipases in a solvent-free system. *Journal of Bioscience and Bioengineering*. 91: 12-5.
- Katchalski-Katzir, E. and Kraemer, D.M. (2000). Eupergit C, a carrier for immobilization of enzymes of industrial potential., *Journal of molecular catalysis B: enzymatic*. 10 (1-3): 157-176.

- Kirk, O., Borchert, T. V. and Furlerng, C. C.(2002). Industrial enzyme applications. *Current Opinion in Biotechnology*. 13: 345-351.
- Klotzbach, T. L., Watt, M.M., Ansari, Y. and Minter, S. D. (2008). Improving the microenvironment for enzyme immobilization at electrodes by hydrophobically modifying chitosan and Nafion polymers. *Journal of Membrane Science*. 311: 81-88.
- Krakowiak, A., Trzcińska, M., Sieliwanowicz, B., Sawicka-Łukowska, R., Jedrychowska, B. and Ajzenberg, V. (2003). Properties of Immobilized and Free lipase from *Rhizopus cohnii*. *Polish Journal of Food and Nutrition Sciences*. 12/53, No 3, 39-44.
- Kuel, R.V., (2000). Design of experiment, statistical principles of research design and analysis; second edition, Duxbury press grove, CA, pp.2-255.
- Kynclova, E., Hartig, A. and Schalkhammer, T.(1995). Oligonucleotide labeled lipase as a new sensitive hybridization probe and its use in bioassays and biosensors. *Journal of Molecular Recognition*. 8: 139-145.
- Lam, M. K., Lee, K. T. and Mohamed, A. R. (2010). Homogeneous, Heterogeneous and Enzymatic Catalysis for Transesterification of High Free Fatty Acid Oil (waste cooking oil) to Biodiesel: A Review. *Biotechnology Advances*. 28: 500– 518.
- Lee, W. M., Kim, K. J., Kim, M. G. and Lee, S. B. (1995). Enzymatic resolution of racemic ibuprofen esters: Effect of organic cosolvents and temperature. *Journal of Fermentation and Bioengineering*. 6: 613-615.
- Li, L., Du, W., Liu, D., Wang, L. and Li, Z. (2006). Lipase-catalyzed transesterification of rapeseed oils for biodiesel production with novel organic solvent as the reaction medium. *Journal of Molecular Catalysis B: Enzymatic*. 43: 58–62.

- Li, N., Zong, M. and Wu, H. (2009) Highly efficient transformation of waste oil to biodiesel by immobilized lipase from *Penicillium expansum*. *Process Biochemistry*. 44(6): 685-688.
- Li, Y., Gao, F., Wei, W., Qua, J-B., Maa, G-H., Zhou, W-Q. (2010). Pore size of macroporous polystyrene microspheres affects lipase immobilization. *Journal of Molecular Catalysis B: Enzymatic*. 66. 182–189.
- Linko, Y.Y., Liimsii, M., Wu, X., Uosukainen, W., Sappilii, J. and Linko, P.(1998). Biodegradable products by lipase biocatalysis. *Journal of Biotechnology*. 66: 41–50.
- Linko, Y. Y. and Wu, X. Y.(1996). Biocatalytic production of useful esters by two forms of lipase from *Candida rugosa*. *Journal of Chemical Technology and Biotechnology*. 65: 163- 170.
- Loboret, F. and Perraud, R. (1999). Lipase-catalyzed production of shortchain acids terpenyl esters of interest to the food industry. *Applied Biochemistry and Biotechnology*. 82: 185-198.
- Lu, J., Chen, Y., Wang, F. and Tan, T. (2009). Effect of water on methanolysis of glycerol trioleate catalyzed by immobilized lipase *Candida* sp. 99-125 in organic solvent system. *Journal of Molecular Catalysis B: Enzymatic*. 56: 122-125.
- Ma, F. and Hanna, M.A. (1999). Biodiesel production: a review. *Bioresource Technology*. 70: 1–15.
- Mamoru, I., Baoxue, C. Masashi, E. Takashi, K. and Surekina, S. (2001). Production of biodiesel from triglycerides and alcohol using immobilized lipase. *Journal of molecular catalysis B: Enzymatic*. 16: 53-58.

- Matassoli, A.L.F., Correa, I.N.S, Ortilho, M.F., Veloso, C.O. and Langone, M.A.P. (2009). Enzymatic synthesis of biodiesel via alcoholysis of palm oil. *Applied Biochemistry and Biotechnology* 155: 347-355.
- Matsumoto, T., Takahashi, S., Kaieda, M., Ueda, M., Tanaka, A., Fukuda, H., Kondo, A. (2001). Yeast whole-cell biocatalyst constructed by intracellular over production of *Rhizopus oryzae* lipase is applicable to biodiesel fuel production. *Applied Microbiology and Biotechnology*. 57 (4): 515-20.
- Mauvernay, R.Y., Labreur, P. and Labrousse, M. (1970). United States Patent 3,513,073. Novel Lipase Composition and Method for Producing Same. United States Patent.
- Mendes, A., Giordano, R., Giordano, R. and de Castro, H. (2011). Immobilization and stabilization of microbial lipases by multipoint covalent attachment on aldehydesin affinity: Application of the biocatalysts in biodiesel synthesis. *Journal of Molecular Catalysis B: Enzymatic*. 68(1): 109-115.
- Meunier, S. and Legge, R. (2010) Evaluation of diatomaceous earth as a support for sol-gel immobilized lipase for transesterification. *Journal of Molecular Catalysis B: Enzymatic*. 62 (1): 54-58.
- Mittelbach, M. (1990). Lipase catalyzed alcoholysis of sunflower oil. *Journal of the American Oil Chemists' Society*. 67: 168-170.
- Mittelback, M. and Tritthart, P. (1988). Diesel fuel derived from vegetable oils. III. Emission tests using methyl esters of used frying oil. *Journal of American Oil Chemists Society*. 65 (7): 1185–1187.
- Mrin, M., Pedregosa, A., Rios, S., Ortiz, M. L. and Laborda, F. (1995). *International Biodeterioration and Biodegradation* 35: 269–285.

- Murty, V.R., Jayadev, B. and Muniswaran, P.K.A. (202). Hydrolysis of Oils by Using Immobilized Lipase Enzyme: A Review. *Biotechnology and Bioprocess Engineering* 7: 57-66.
- Nakajima, M., Snape, J., and Khare, S.K. In: Gupta MN, editor. Method in non-aqueous enzymology. Basel: Birkhauser Verlag; 2000. p. 52– 69.
- Nelson, L.A., Foglia, T.A., Marmer, W.N., (1996). Lipase-catalyzed production of biodiesel. *Journal of the American Oil Chemists' Society*. 73(8): 1191–1195.
- Noureddini, H., Gao, X. and Phikana, R.S. (2001). Immobilized *Pseudomonas cepacia* lipase for biodiesel fuel production from Soyabean oil. *Bioresource Technology*. 96: 767–777.
- Ozturk, B. (2001): *Immobilization of lipase from Candida rugosa on Hydrophobic and Hydrophilic Supports*. Master of Science Dissertation, Izmir Institute of Technolog, Turkey.
- Pahoja, M.V. and mumtaz A. S. (2002). A Review of enzymatic propereties of lipases in plants, animals and microorganisms. *Pakistan journal of applied sciences*. 2(4): 474-484.
- Pandey, A., Sailas, B., Carlos, R. S., Poonam, N., Nadia, K. and Vanete, T. S. (1999). The realm of microbial lipases in biotechnology. *Applied Biochemistry and Biotechnology*. 29: 119–131.
- Piraj`n, M. JC. and Liliana, G. (2001). Study of immobilized *candida rugosa* lipase for biodiesel fuel production from palm oil by flow microcalorimetry. *Arabian Journal of Chemistry*. 4: 55–62.
- Ray, A. (2012). Application of Lipase in Industry. *Asian Journal of Pharmaceutical*. 2(2): 33-37.

- Ranganathan, S.V., Narasimhan, S.L. and Muthukumar, K. (2008). An overview of enzymatic production of biodiesel. *Bioresource Technology*. 99: 3975-3981.
- Roberto, D.V., Damianni, S., Graca, M. N. and Valdir, S. (2005). Carboxymethylcellulose and poly (vinyl alcohol) used as a film support for lipase immobilization. *Process Biochemistry*. 40(8): 2677-2682.
- Robles-Medina, A., Gonzalez-Moreno, P.A., Esteban-Cerdana, L. and Molina-Grima, E. (2009). Biocatalysis: Towards ever greener biodiesel production. *Biotechnology Advances*. 27: 398-408.
- Royon, D., Daz, M., Ellenrieder, G. and Locatelli, S. (2007) Enzymatic of biodiesel from cottonseed using t-butanol as a solvent. *Bioresource Technology*. 98: 648–53.
- Ruckenstein, E. and Wang, X. (1993). Lipase immobilized on hydrophobic porous polymer supports prepared by concentrated emulsion polymerization and their activity in the hydrolysis of triacylglycerides. *Biotechnology and Bioengineering*. 42: 821-828.
- Saifuddin N. and Razia A. Z., (2008). Enhancement of Lipase Enzyme Activity in Non-Aqueous Media through a Rapid Three Phase Partitioning and Microwave Irradiation. *E-Journal of Chemistry*. 5(4): 864-871
- Samukawa, T., Kaieda, M., Matsumoto, T., Ban, K., Kondo, A., Shimada, Y., Noda, H., Fukuda, H., (2000). Pretreatment of Immobilized *Candida antarctica* lipase for Biodiesel fuel production from plant oil. *Journal of Bioscience and Bioengineering*. 90: 180–183.
- Sanchez, M., Prim, N., Rendez-Gil, F., Pastor, F I J. and Diaz, P.(2002). Engineering of baker's yeasts, *Escherichia coli* and *Bacillus* hosts for the production of *B. subtilis* lipase A. *Biotechnology and Bioengineering* 78: 339-345.

- Saraiva Silva, G., Fernandez, L.R.V., Higa, O.Z., Vítolo, M., De Queiroz, M. A.A.A (2004). Alginate-Poly(Vinyl Alcohol) Core-Shell Microspheres For Lipase Immobilization. XVI Congressor Brasileiro de Engenharia e Ciencia dos Materiais. *Porto Alegre,-RS* de 28 de novembro a 02 de dezembro de 2004.
- Saxena S.K. (2004). Polyvinyl Alcohol (PVA). *Chemical and Technical Assessment (CTA), 61st JECFA*. FAO
- Schumacher, L. G., Borget, S. C., Fosseen, D., Goetz, W. and Hires, W.G. (1996). Heavy-duty engine exhaust emission test using methyl ester soybean oil/diesel fuel blends. *Bioresourse Technology*. 7(1): 31-36.
- Schumacher, L.G., Gerpen, J.V. and Adams, B. (2004). *Biofuels. Encyclopedia of Energy*, Volume 1. r 2004 Elsevier Inc.
- Sellappan, S. and Akoh, C. C. Applications of lipases in modification of food lipids. In *Handbook of Industrial Catalysis*; Hou, C. T., Ed.; Taylor and Francis: Boca Raton, FL, 2005; pp 9-1–9-39.
- Selmi, B. and Thomas, D. (1998). Immobilized-lipase-catalyzed ethanolysis of sunflower oil in solvent-free medium. *Journal of the American Oil Chemists' Society*. 75: 691–5.
- Sharma, R., Chisti, Y. and Banerjee, U. C.(2001). Production, purification, characterization and applications of lipases. *Biotechnology Advances*. 19: 627-662.
- Shay, L.K., Fisher, T.J., Banasiak, D.S. and Wegner, E.H. (1990). *European Patent 357,812* Enhancing the flavour of proteinaceous products derived from microorganisms, European Patent.
- Shah, S., Sharma, S. and Gupta, M.N.(2004). Biodiesel preparation by lipase-catalyzed transesterification of jatropha oil. *Energy Fuel*. 18: 154–9.

- Shah, S. & Gupta, M.N., (2007) Lipase catalyzed preparation of biodiesel from *Jatropha* oil in asolvent free system. *Process Biochemistry*. 42(2): 409-414.
- Shimada, Y., Watanabe, Y., Samukawa, T., Sugihara, A., Noda, H., Fakuda, H. and Tominaga, Y. (1999). Conversion of vegetable oil to biodiesel using immobilized *Candida antarctica* lipase . *Journal of American oil chemists society*. 76(7): 789-793.
- Shimada Y., Watanabe Y., Sugihara A., and Tominaga Y., (2002). Enzymatic alcoholysis for Biodiesel fuel production and application of the reaction to oil processing. *Journal of Molecular Catalysis B: Enzymatic*. 17(3-5): 133-142.
- Silva, W.O.B., Mitidieri, S., Schrank, A. and Vainstein, M.H. (2005). Production and extraction of an extracellular lipase from the entomopathogenic fungus *Metarhiziumanisopliae*. *Process Biochemistry*. 40: 321-326.
- Soumanou, M. M. and Bornscheuer, U. T. (2003). Improvement in lipase-catalyzed synthesis of fatty acid methyl esters from sunflower oil. *Enzyme and Microbial Technology*. 33: 97-103.
- Spahn C. and Shelley D. M.(2008). Enzyme Immobilization in Biotechnology. *Recent Patents on Engineering*. 2: 195-200.
- Sumner, C., Krause, S., Sabot, A., Turner, K. and McNeil C.J. (2001). Biosensor based on enzyme-catalysed degradation of thin polymer films. *Biosensors and Bioelectronics*. 16(9-12): 709-14.
- Tamalampudi, S., Talukder, M. R., Hamad, S., Numata, T., Kondo, A., and Fukuda, H. (2008). Enzymatic production of biodiesel from *Jatropha* oil: A comparative study of immobilized-whole cell and commercial lipases as a biocatalyst. *Biochemical Engineering Journal* 39, 185-189.

- Tcacenco, L., Ana, A. C., Camelia, U. and Elena, B. (2010). The preparation and immobilization of some yeast lipases for rapeseed oil transesterification to biodiesel. *Romanian Biotechnological Letter*. 15: 5.
- Uhlich, T., Ulbricht, M. and Tomaschewski G.(1999). Immobilization of enzymes in photochemically cross-linked poly(vinyl alcohol). *Enzyme and Microbial Technology*. 19: 124-31.
- Ushio, K., Hirata, T., Yoshida, H., Sakaue, M., Hirose, C., Suzuki, T. and Ishizuka, M. (1996). *Biotechnology Techniques*. 10: 267–272.
- Vasileva-Tonkova, E. and Galabova, D.(2003). Hydrolytic enzymes and surfactants of bacterial isolates from lubricant-contaminated wastewater. *Zeitschrift fur Naturforschung*. 58(1–2): 87–92.
- Vasudevan, P.T. and Briggs, M. (2008). Biodiesel production-current state of the art and challenges. *Journal of Industrial Microbiology and Biotechnology*. 35: 421-430.
- Verger, R. (1997). Interfacial activation of lipases: facts and artifacts., *TIBTECH reviews*. 15: 32-38.
- Vitolo, S., Petarca, L. and Bresci, B. (1998). *Bioresource Technology*. 67: 129–137.
- Vulfson, EN.(1994). Industrial applications of lipases. In: Wooley P, Petersen SB, editors. *Lipases*. Cambridge, Great Britain: Cambridge University Press, 271.
- Wan Omar, W. N. N., Nordin, N., Mohamed, M. and Amin, N. A. S. (2009). A TwoStep Biodiesel Production from Waste Cooking Oil: Optimization of Pre Treatment Step. *Journal of Applied Science*. 9: 3098-3103.
- Wang, P. G., Nahalka, J., Ziye, L. and Xi, C. (2003). Superbeads: Immobilization in “Sweet” Chemistry. *Chemistry - A European Journal*. 9 : 2

- Watanabe, Y., Shimada, Y., Sugihara, A. and Tominaga Y. (2001). Enzymatic conversion of waste edible oil to biodiesel fuel in a fixed-bed bioreactor. *Journal of the American Oil Chemists' Society*. 78: 703-7.
- Xie, Y. C., Liu, H. Z. and Chen, J. Y.(1998) *Candida rugosa* lipase catalyzed esterification of racemic ibuprofen and chemical hydrolysis of S-ester formed, *Biotechnology Letters*. 20: 455-458.
- Xu, G. and Wu, G. Y. (2003). The investigation of blending properties of biodiesel and no. 0 fuel. L. *Jiangsu Polytechnic University*. 15:16-18
- Xu, Y. Y., Du, W., Zeng, J. and Liu, D.H. (2004). Conversion of soybean oil to biodiesel fuel using lipozymeTL IM in a solvent-free medium. *Biocatalysis and Biotransformation*. 22: 45-8.
- Yan, J., Yunjun, Y., Sanxiong, L., Jiang, H. and Guilong, W. (2011). Preparation of cross-linked lipase-coated micro-crystals for biodiesel production from waste cooking oil. *Bioresource Technology*. 102: 4755-4758.
- Ye, P., Jiang, J. and Xu, Z. (2007) Adsorption and activity of lipase from *Candida rugosa* on the chitosan-modified poly(acrylonitrile-co-maleic acid) membrane surface. *Colloids and Surfaces B: Biointerfaces*. 60: 62-67.
- Zain, M. A., Atika, H.A.N. and Noorfadilatulshilla, M.A. (2011). Preliminary Study on Synthetic Textiles Effluents Discoloration by Immobilized *Phanerochaete chrysosporium* in PVA-Alginate beads (PI 20081825). UMTAS.
- Zain, A.M., Suhaimi, M. A., and Ani, I. (2010). Hydrolysis of liquid pineapple waste by invertase immobilized in PVA-alginate matrix. *Biochemical Engineering Journal*. 50: 83-89.
- Zalacain, I., Zapelena, M. J., Astiasaran, I. and Bello, J. (1995). *Meat Science*. 40: 55-61.