

**ISOLATION AND IDENTIFICATION OF A 3-CHLOROPROPIONIC ACID
(3CP) DEGRADING *ASPERGILLUS ACULEATUS* STRAIN MBS_179**

SALAR KADHUM ALI ZANGANA

**A dissertation submitted in partial fulfillment of the
requirements for the award of the degree of
Master of Science (Biotechnology)**

**Faculty of Biosciences and Bioengineering
Universiti Teknologi Malaysia**

JANUARY 2013

To my beloved father, mother, brothers, sisters, wife, and my daughters

ACKNOWLEDGEMENTS

First of all, I must be thankful to Allah for accomplishing the research and I would like to express my sincere thanks and appreciation to my supervisor Assoc. Prof. Dr. Fahrul Zaman Huyop, who continuously guided me throughout every step of my study and generously shared his time and knowledge with me. Million words of thanks to fellow friends who showed their collaboration, assistance, concern and support all the way while carrying out my laboratory work. Their views and tips are useful indeed.

ABSTRACT

The halogenated organic compounds which are used in agricultures and industries consider as xenobiotics. They cause the major group of environmental pollutions. They are potential toxicity and carcinogenicity effect and they pose health risks to humans and animals. In this study, the main objectives are to isolate and identify fungus from bread that has the ability to utilize 3-chloropropionic acid as a sole source of carbon and energy. The 3-chloropropionic acid (3-CP) is a synthetic halogenated compounds and classified as of β -chlorinated alcanoic acid (β -chloro substituted haloalkanoates). This compound is used as intermediates for the synthesis of pharmaceuticals, herbicides and pesticides. The MBS_179 fungus which was isolated has been identified by using conventional and molecular analysis techniques. The MBS_179 showed the ability to grow on minimal medium containing only 3-chloropropionic acid as a sole carbon source and energy. The growth was measured after inoculated into 20 mM 3-chloropropionate liquid minimal media (pH 7) and incubated at 30°C in a rotary incubator at 120 rpm over 20 days period. Sample was taken at 2 days intervals. The molecular analysis depended on the amplified 18S rRNA gene sequence of the MBS_179 and the phylogenetic relationship was constructed using MEGA4 software[®] in order to verify their evolutionary distance. From our current study, analysis of 18S rRNA gene and the evolutionary relationship described the subspecies and characteristic of the MBS_179 which had 100% identity to the *Aspergillus aculeatus*. This study demonstrates the ability of this MBS_179 to utilize 3-CP that indicates it may has a significant role to play in the removal of this environmental pollutant.

ABSTRAK

Sebatian organik berhalogen yang digunakan dalam sektor pertanian dan perindustrian adalah dianggap sebagai bahan xenobiotics. Bahan-bahan ini merupakan punca utama pencemaran alam sekitar. Ia berpotensi memberikan kesan kotoksidan dan kekarsinogenan yang memberi risiko kepada kesihatan manusia dan haiwan. Dalam kajian ini, objektif utama adalah untuk mengasingkan dan mengenalpasti kulat daripada roti yang mana mempunyai keupayaan dengan menggunakan 3-chloropropionik sebagai sumber utama karbon dan tenaga. Asid 3-chloropropionik (3CP) merupakan sebatian sintetik berhalogen dan diklasifikasikan sebagai asid β -chlorinated alkonik (β -chloro menggantikan haloalkanoates). Sebatian ini digunakan sebagai perantaraan untuk sintesis farmaseutikal, racun herba dan racun perosak. Kulat MBS_179 yang telah diasingkan telah dikenalpasti dengan menggunakan teknik konvensional dan molekul. MB_179 menunjukkan keupayaan untuk tumbuh dalam medium yang minimum yang hanya mengandungi asid 3-chloropropionik sebagai sumber tunggal kepada karbon dan tenaga. Pertumbuhan diukur berdasarkan penggunaan asid 3-chloropropionik dalam medium yang minimum. Analisis molekul bergantung kepada kekuatan urutan gen 18S rRNA daripada MBS_179 dan hubungan filogeni dibina menggunakan perisian MEGA4 untuk mengesahkan evolusi menunjukkan subspecies dan ciri-ciri MBS_179 yang mana 100% mempunyai identiti sebagai *Aspergillus aculeatus*. Kajian ini menunjukkan keupayaan MBS_179 menggunakan 3CP yang menunjukkan bahawa ia mungkin memainkan peranan penting untuk meningkatkan penyingkiran pencemaran pada alam sekitar.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	x
	LIST OF FIGURES	xi
	LIST OF APPENDICES	xiii
1	INTRODUCTION	1
	1.1 Overview	1
	1.2 Problem Statement	3
	1.3 Goal and Objectives	4
	1.4 Research Scope	5
	1.5 Research Significance	5
2	LITERATURE REVIEW	6
	1.1. Introduction	6
	1.2. What are xenobiotics	7
	1.3. The chemistry of halogenated compound	9
	1.4. Halogenated aliphatic compounds	9

1.5.	Microbial degradation	11
1.6.	Involvement of dehalogenases in the microbial degradation of halogenated compounds	12
1.7.	Enzymatic dehalogenation of halogenated aliphatics	14
1.8.	Dehalogenating enzymes	16
1.9.	Dehalogenation mechanisms	17
1.10.	Properties of 3-chloropropionic acid	18
1.11.	Dehalogenation of β -halocarboxylic acid (3-chloropropionate)	19
1.12.	Fungi	22
1.13.	Identification of fungi	25
1.13.1	Morphological characterization	25
1.13.2	18S rRNA identification of fungi	25
1.14.	Phylogenetic study	29

3

	RESEARCH METHODOLOGY	31
3.1	Introduction	31
3.2	Source of Fungus	31
3.3	Source of Chemicals Substances	32
3.4	Preparation of Minimal Media	32
3.5	Sterilization Process	33
3.6	Measurement of the Fungal Growth	34
3.7	Halide Assay	34
3.8	Fungus Characterization	35
3.9	PDA and PDB Preparation	35
3.10	Staining for Fungus by Lactophenol Cotton Blue	35
3.11	Scanning Electron Microscope (SEM)	36
3.12	DNA Extraction	37
3.13	Polymerase Chain Reaction (PCR)	37
3.14	Agarose Gel Electrophoresis	39
3.15	PCR Purification	39
3.16	DNA Sequencing	40

3.17	Homology Search and Phylogenetic Tree Construction Using BLAST	40
3.18	DNA Sequences Analyzing by MEGA Software	40
4	RESULTS AND DISCUSSION	41
4.1	Isolation and Identification of the Fungi	41
4.2	Halide Ion Assay	46
4.3	Growth Curve Measurement of the MBS_179	47
4.4	Identification of the Fungus by 18S rRNA Analysis	48
4.5	DNA Sequencing	50
4.6	Phylogenetic Tree Construction	51
4.7	Properties of the Identified Fungus	52
4.8	Discussion	54
5	CONCLUSIONS	56
5.1	Conclusion Remarks	56
5.2	Future Studies	57
	REFERENCES	58
	Appendix A	65
	Appendix B	69
	Appendix C	70
	Appendix D	71
	Appendix E	72

LIST OF TABLES

TABLE No.	TITLE	PAGE
2.1	Dehalogenase and substrate of different microorganisms	15
3.1	Functions of the Lactophenol cotton blue components	36
3.2	Properties of the universal primers which were used	37
3.3	Components, which are used in PCR	38
3.4	Thermal cycle profile for PCR reaction	38
4.1	Summary of the genus	50
4.2	Scientific classification of <i>Aspergillus</i>	52

LIST OF FIGURES

FIGURES NO	TITLE	PAGE
2.1	Possible environmental fate of a xenobiotic compound	8
2.2	Types of dehalogenases	13
2.3	Chemical structure of 3-chloropropionic acid	18
2.4	The formation of acrylic acid from 3-chloropropionic acid	21
2.5	Adapted from Alexopoulos et al. (1996)	22
2.6	The evolution of the fungi and allied fungi-like microorganisms based on a concatenated neighbor-joining analysis	23
2.7	The component of ribosome in eukaryotic cell	26
2.8	Ribosomal DNA in fungal identification	28
2.9	Universal phylogenetic tree shows the three major domains	29
2.10	Detailed tree for eukaryotic shows the trunk branching to kingdoms	30
4.1	Result of spread plate on minimal medium (20 mM 3-chloropropionic acid)	42
4.2	Photographs of the isolated MBS_179 on PDA	43
4.3	Photographs shown the morphology of the isolated MBS_179 by optical microscope	44
4.4	Photographs shown the morphology of the isolated MBS_179	45
4.5	Growth curves of the fungal culture (MBS_179) measured using chloride ion released in the growth medium containing 20 mM 3CP.	47

4.6	The growth curve of the MBS_179 on a 3-chloropropionic acid liquid media	48
4.7	Agarose gel analysis of PCR amplified band from fungus MBS_179 in 1% Agarose	49
4.8	A sequence of strain MBS_179	50
4.9	Neighbour- joining distance ree of the <i>Aspergillus aculeatus</i> MBS_179	51

LIST OF APPENDICES

APPENDIX NO.	TITLE	PAGE
A	External view of TM3000 Tabletop Microscope	65
B	Properties of the ITS primers	69
C	1KB plus DNA ladder	70
D	Graphic Summay and Description of the Sequences producing significant alignments (By BLAST)	71
E	Estimates of Evolutionary Divergence between Sequences	72

CHAPTER 1

INTRODUCTION

1.1 Overview

The dawn of the 20th century, marked the beginning of the use of haloaliphatic compounds in industries and agriculture. After some decades of their discovery, the hazard associated with the use of these compounds started to manifest in the environment and in living organisms. The quest by people for a cleaner lifestyle lead to the use of man-made chemical such as haloaliphatic hydrocarbon compounds as herbicide, pesticide, fungicides, paint, solvents and industrial chemicals which cause harmful damage to the environment via a pollution (Rasul and Chapalamaduqu, 1991).

Halogenated compounds are very essential group of xenobiotics. 3-chloropropionic acid (3CP) that belongs to the class of chlorinated mono carboxylic acid or β -chloro substituted haloalkanotes. They are considered as a possible chemical inclusion in pesticides. This compound has a carcinogenic and genotoxic effect on animals and humans (Jing and Huyop, 2007).

A major process through which xenobiotic chemicals such as chloroaliphatic compounds is removed from the environment by microorganisms is called biodegradation (Sinha *et al.*, 2009). The microbial degradation of a contaminant usually occurs because microorganisms derive support for growth through the use of the contaminant as an electron donor and carbon source. Hydrolytic dehalogenases typifies the major step in the degradation of haloaliphatic compounds. These enzymes propel the cleavage of halogen-carbon bonds by nucleophilic substitution, and change the halogen ion by a hydroxyl group obtained from water (Schwarze *et al.*, 1996). Field and Alvarez (2004) noted that by using the dehalogenase enzyme, compounds containing chlorine are being metabolized and chlorine substituent are enzymatically removed to form non-halogenated compounds.

The decontamination of these substances can be done by using biological or non-biological degradation process. Since the biological methods are reasonable, harmless, and environmentally friendly, they are favored (Janssen *et al.*, 2005). Bioremediation is a process that involves the use of biological methods like microbial or enzymatic biocatalysts (Janssen *et al.*, 2001). Many years ago, studies on microbial degradation of harmful waste started, due to growing of xenobiotic compounds in industrial and agricultural uses. Microbial degradation of xenobiotic chemicals such as chloroaliphatic compounds involves the ways and processes of biodegradation that assist in removing these compounds from the environment. There are so many studies had been done on the microbial catabolism of dehalogenase (Olaniran *et al.*, 2001, 2004; Jing and Huyop, 2007; Jing *et al.*, 2008). Most of the degradation pathway of halogenated compounds includes dehalogenation, the cleavage of the carbon-halogen bond to release halogen, catalyzed by 'dehalogenase' (Fetzner and Lingens, 1994; Janssen *et al.*, 2005).

The ability of the enzyme to catalyze the dehalogenation of various halogen-substituted organic acids was investigated and the highest activity was found with 3-chloropropionic acid as a sole carbon source in the growth medium

(Jing *et al.*, 2008). There are many different microorganisms (like bacteria and fungi) which are capable of utilizing halogenated compound as the only carbon source and consequently, are supposed to have a significant role in their natural detoxification (Janssen *et al.*, 2001; Song *et al.*, 2003; Park *et al.*, 2003), and is a major step in aerobic mineralization pathways of many halogenated compounds that occur as environmental pollutants (Janssen *et al.*, 2005). A number of these microorganisms were originally isolated from polluted soil and sewage oxidation ponds (Olaniran *et al.*, 2001, 2004). So far, fungi had been found to be isolated to degrade halogenated compound particle (α -haloalkanoic acid).

The current study focuses on the isolation and identification of other environmental microorganisms to dissociate the presence of 3-chloropropionic acid. In detail, the aim of this study is identification of the fungus isolated from the bread that can degrade 3-chloropropionic acid based on the 18S rRNA molecular approach. This will be a genotypic dependable approach to characterize the isolated fungi, compared to conventional methods like morphological procedures.

1.2 Problem statement

The widespread usage of chloroaliphatic hydrocarbon compounds in industries has resulted in extensive pollution of environment and the ground water (Fliermans, 1989). Chlorinated aliphatic compounds form one of the most vital groups of industrially produced chemicals. Several of these compounds are weakly degraded in the environment (Reineke, 2001). The 3-chloropropionic acid which is one of these compounds, considered as a possible chemical inclusion in certain herbicide and carcinogenic and genotoxic to the animal and human. The uses of herbicides in general (2,2DCP and 3CP) are the main contributor of the environmental pollution (Jing *et al.*, 2005). Aerobic biodegradation by using microorganisms such as fungi, to

degrade the chloroaliphatic hydrocarbon compounds to carbon dioxide, water, and hydrogen chloride is more efficient than anaerobic or chemical processes. Therefore, the aerobic biodegradation by using microorganism such as fungi is preferable as it is costly and biologically effective.

1.3 Goals and Objectives

The main goals of the current research were to isolate and identify 3-chloropropionic acid degrading fungi. Therefore, to achieve these goals, the following objectives were considered:

- (i) To isolate of a single fungus from the Bread that can degrade 3CP.
- (ii) To identify and characterize the isolated fungus by determining the morphological appearances and genetic sequence analysis.
- (iii) To study the growth capability of isolated fungus on 3CP.

1.4 Research Scope

The scope of this research includes the molecular identification of the fungus that isolated from bread, which has ability to degrade 3-chloropropionic acid by amplifying the ITS region, whereas the universal ITS primers (ITS1 and ITS4) used to run the PCR (White *et al.*, 1990). In addition, the conventional methods that used for further identification and characterization.

1.5 Research Significance

The outcome of this study tends to identify fungal species from bread capable of degrading 3CP compounds. Novel fungal species was isolated, which has the ability of degrading the chloroaliphatic compounds and this new strain of fungus, which is previously unknown for its reaction of chloroaliphatic compounds.

REFERENCES

- Alexopoulos, C. J., Mims, C. W., and Blackwell, M. (1996). *Introductory Mycology*. (4th ed.). USA: John Wiley and Sons Inc.
- Allison N., Skinnerb, A. J., Cooper, R. A. (1983). The dehalogenases of a 2,2dichloropropionate degrading bacterium. *Journal of. gen. microbial.* 192:1283-1293.
- Asmara, W, Murdiyatmo U, Baines AJ, Bull AT, Hardman DJ (1993). Protein engineering of the 2-haloacid halidohydrolase Iva from *Pseudomonas cepacia* MBA4. *Biochem. Journal.* 292: 69-74.
- Bachas-Daunert PG, Sellers ZP, Wei Y (2009). Detection of halogenated organic comopounds using immobilized thermophilic dehalogenase. *Anal. Bioanal. Chem.*, 395(4): 1173-1178.
- Bergmann, J.G. and Sanik, J. (1957). Determination of trace amounts of chloride in naphtha. *Anal. Chem.* (29): 241-243.
- Berry E. K. M., Allison N, Skinner AJ, Cooper RA (1979). Degradation of the selective herbicide 2,2-dichloropropionate (Dalapon) by a soil bacterium. *Journal of. Gen. Microbiol.*, 110: 39-45.
- Bollag JM, Alexander M (1971) Bacterial dehalogenation of chlorinated aliphatic acids. *Soil Biol Biochem* 3:91–96.
- Bridge, P. D., Hawksworth, D. L., Kozakiewicz, Z., Onions, A. H. S., Paterson, R. R. M., and Sackin, M. J. (1985). In Samson, R. A., and Pitt , J. I. (Editors), *Advances in Penicillium and Aspergillus Systematics*. (pp.281). New York, London: Plenum Press.

- Busto M.D., Smith P.P., Mateos-Perez M., Burns R.G. (1992). Degradation of aliphatic halogen-substituted pesticides by dehalogenase isolated from *Pseudomonas alcaligenes*. Identification and properties of the enzyme. *Sci. Total Environ.*, 123/124, 267-277.
- Cairns SS, Cornish A, Cooper RA (1996). Cloning, sequencing and expression in *Escherichia coli* of two *Rhizobium sp.* genes encoding haloalkanoate dehalogenases of opposite stereospecificity. *Eur. J. Biochem.*, 253: 744-749.
- Carlile, M. J., Watkinson, S. C. and Graham. (2001). *The Fungi*. London UK : Academic Press a Harcourt Science and Technology Company.
- Commandeur, L.C. and Parsons, J.R.(1990) Degradation of Halogenated Aromatic Compounds. *Biodegradation*.1(2-3):207-20.
- Davies, J. L., and Evans, W. C. (1962). The elimination of halide ions from aliphatic halogen-substituted organic acids by an enzyme preparation from *Pseudomonas dehalogens*. *Biochemical Journal*. 82, 50.
- Colquhoun, J.A., Heald, S.C., Li, L., Tamaoka, J., Kato, C., Horikoshi, K., and Bull, A.T. (1998). Taxonomy and biotransformation activities for some deep-sea actinomycetes. *Extremophiles*, (2): 269-277.
- El Fantroussi, S., Naveau, H., and Agathos, S. N. (1998). Anaerobic dechlorinating bacteria. *Biotechnol Prog*. 14: 167-188.
- Fetzner, S. (1998). Bacterial dehalogenation. *Appl. Microbiol. Biotechnol.* 50, 633–657.
- Fetzner, S. R., Lingens F. (1994). Bacterial Dehalogenases. *Microbiol. Rev.* 58(4):641-685.
- Field, J. A., Alvarez, S. R. (2004). Biodegradability of chlorinated solvents and related chlorinated aliphatic compounds. *Reviews in Environmental Science & Bio/Technology*. (3): 185–254.
- Fowden, L. (1968). The occurrence and metabolism of carbon halogen compound. *Proc. Royal Soc.*, 171: 5-18.
- Frisvad, J. C. (1989b) The use of high-performance liquid chromatography and diode-array detection in fungal chemotaxonomy based on profiles of secondary metabolites. *Botanical Journal of the Linnean Society* 99: 81-95.
- Goldman, P., Milne, G. W. A. & Keister, B. (1968). Carbon-halogen bond cleavage. 111. Studies on bacterial halidohydrolases. *Journal of Biological Chemistry* 243,428-434.

- Guarro, J., Gene, J., and Stchigel, M. A. (1999). Development In Fungal Taxonomy. *Clinical Microbiology*, 11, 1-40.
- Hardman, D. J. (1991). Biotransformation of halogenated compounds. *Crit. Rev. Biotechnol.*, 11: 1-40.
- Hardman, D. J, Slater, J. H. (1981). Dehalogenases in soil bacteria. *J. Gen. Microbiol.*, 123: 117-128.
- Hareland, W. A., Crawford, R. L., Chapman, P. J., and Dagley, S. (1975). Metabolic function and properties of a 4-hydroxyphenyl-acetic acid 1-hydroxylase from *Pseudomonas acidovorans*. *J. Bacteriology*. 121: 272-285.
- Hill, K. E., Marchesi, J. R., and Weightman A. J. (1999). Investigation of two evolutionary unrelated halocarboxylic acid dehalogenase gene families. *Journal of . Bacteriol.* 181: 2535-2547.
- Hirsch, P., and Alexander, M. (1960). Microbial decomposition of halogenated propionic and acetic acids. *Can. J. Microbiol.*, 6 (3):241-249.
- Huyop, F., and Nemati, M. (2010). Properties of dehalogenase from *Rhizobium* sp. RC1. *African Journal of Microbiology Research* Vol. 4(25), pp. 2836-2847.
- Janssen, D. B., Dinkla, I. J. T., Poelarends, G. J. and Terpstra, P. (2005). Bacterial degradation of xenobiotic compounds: evolution and distribution of novel enzyme activities. *Environmental Microbiology* 7(12), 1868–1882.
- Janssen, D. B., Oppentocht, J. E., and Poelarends, G. J. (2001). Microbial dehalogenation. *Curr. Opin. Biotechnol.*, 12: 254-258.
- Jensen, H. L. (1960) Decomposition of chloroacetates and chloropropionates. by bacteria. *Acta Agriculturae Scandinavica* 10:83-103.
- Jensen, H. L. (1959) Decomposition of chlorine-substituted organic acids by fungi *Acta. Agriculturae Scandinavica* 9:421-434.
- Jensen, H. L. (1957). Decomposition of chloro-organic acids by fungi. *Nature*, 180: 1416.
- Jensen, H. L. (1951) Decomposition of chlorosubstituted aliphatic acids by soil bacteria. *Can. J. Microbiol.* 3:151-164.
- Jing, N. H., Wahab, R. A. b., Hamdan, S., Huyop, F. (2010). Cloning and DNA sequence analysis of the haloalkanoic permease uptake gene from *Rhizobium* sp. RC1. *Biotechnol.*, 9(3): 319-325.

- Jing, N. H., Wahab R. A., Taha A. M., Rashid N. A. A., Huyop AM (2008) A further characterization of 3-chloropropionic acid dehalogenase from *Rhodococcus sp.* HJ1. Res J Microbiol 3:482–488.
- Jing, N. H. and Huyop, F.(2007). Dehalogenation of Chlorinated Aliphatic Acid By *Rhodococcus Sp.* Asia Pacific Journal of Molecular Biology and Biotechnology. 15, 147-151.
- Jing, N. H., Wahab, R., Cooper R. A., and Huyop, F. (2005). Degradation of herbicide (3-CP) by bacterial dehalogenases. Proc. KUSTEM 4th Annual seminar. 2-5 May. Primula Beach Resort, Kuala Terengganu.
- Kawasaki, H., Tone, H., and Tonomura, K. (1981). Purification and properties of haloacetate halidohydrolase specified by plasmid from *Moraxella sp.* strain. B. Agric. Biol. Chem., 45: 35-42.
- Kwon, O. Y., Ogino, K., and Ishikawa, H. (1991). The Longest 18s Ribosomal RNA Ever Known .Eur J. Biochem. 202,827-833.
- Lee, R. E. (2008). Phycology (4th Edition).(pp 22-25). USA: Cambridge University Press .
- Leisinger, T. (1996). Biodegradation of chlorinated aliphatic compounds. Curr. Opin. Biotechnol. 7, 295-300.
- Little, M., and Williams, P. A. (1971). A bacterial halidohydrolase. Its purification, some properties and its modification by specific amino acid reagents. Eur. J. Biochem. 21:99-109.
- Marais, J. S. C. (1944). Monofluoroacetic acid, the toxic principle of "Gifblaar" *Dichapetalum cymosum* (Hook) Engl. Onderstepoort J. Vet. Sci. Anim. Ind., 20: 67-73.
- Marchesi, J. R., Weightman, A. J. (2003). Comparing the dehalogenase gene pool in cultivated alpha-halocarboxylic aciddegrading bacteria with the environmental metagene pool. Appl. Env. Microbiol., 69: 4357-4382.
- McConell, G., Ferguson, D. M., and Pearson, C. R. (1975). Chlorinated hydrocarbons and the environment. Endeavour 34, 34-18.
- McGrath, J. E., Harfoot, C. G. (1997). Reductive dehalogenation of halocarboxylic acids by the phototrophic genera *Rhodospirillum* and *Rhodopseudomonas*. Appl. Env. Microbiol. 63 (8): 3333-3335.

- Mesri, S., Wahab, R. A., Huyop, F. (2009). Degradation of 3-chloropropionic acid (3CP) by *Pseudomonas* sp. B6P isolated from a rice paddy field. *Ann Microbiol* 59:447–451.
- Mohn, W. W., and Tiedje, J. M. (1992). Anaerobic biodegradation of chlorinated benzoates. *Microbiol. Rev.* 56, 482-501.
- Morgan, P. and Watkinson, R. J. (1989). Microbiological methods for the cleanup of soil and ground water contaminated with halogenated organic compounds. *FEMS Microbiol. Lett.* 63, 277-300.
- Motosugi, K., Esaki, N., and Soda, K. (1982). Purification and properties of 2-halo acid dehalogenase from *Pseudomonas putida*. *Agric. Biol.Chem.* 46: 837-838.
- Nardi-Dei, V., Kurihara, T., Chung, P., Masaru, M., Susumu, T., Soda, K., and Nobuyoshi, E. (1999). DL-2Haloacid Dehalogenase from *Pseudomonas* sp. 113 is a new class of dehalogenase catalyzing hydrolytic dehalogenation not involving enzyme–substrate ester intermediate. *J. Biol. Chem.*, 30: 20977-20981.
- Ng, H. J., Roswanira, A., Ronald, A. C., and Fahrul, H. (2005). Degradation of Herbicide (3-Chloropropionic Acid) By Bacterial Dehalogenases.
- Olaniran, A. O., Babalola, G. O., Okoh, A. I. (2001). Aerobic dehalogenation potentials of four bacterial species isolated from soil and sewage sludge. *Chemosphere*, 45(1): 45-50.
- Olaniran, A. O., Pillay, D., Pillay, B. (2004). Haloalkane and haloacid dehalogenases from aerobic bacterial isolates indigenous to contaminated sites in Africa demonstrate diverse substrate specificities. *Chemosphere* 55(1):2733.
- Park, C., Kurihara, T., Yoshimura, T., Soda, K., and Esaki, N. (2003). A new D,L-2-haloacid dehalogenase acting on 2-haloacid amides: purification, characterization, and mechanism. *J. Mol. Catal. B.* 23(1):329-336.
- Paterson, R. R. M. (1986). Standardized One- and Two-Dimensional Thin-Layer Chromatographic Methods for the Identification of Secondary Metabolites in *Penicillium* and Other Fungi. *Journal of Chromatography.* 368, 249-264.
- Rasul, G. C., and Chapalamadugu, S. (1991). Biodegradation of Halogenated Organic Compounds. *American Society for Microbiology* 55(1): 59-79.
- Rittmann, B. and McCarthy, P. (2001). *Environmental Biotechnology: Principles and Applications*, McGraw-Hill.

- Schwarze, R., Brokamp, A., and Schmidt, F. R. J. (1997). Isolation and Characterization of Dehalogenases from 2,2-Dichloropropionate-Degrading Soil Bacteria. (34): 103–109.
- Sinha, S., Chattopadhyay, P., Pan, L., Chatterjee, S., Chanda, P., Bandyopadhyay, D., Das, K. and Sen, S. (2009). Microbial transformation of xenobiotics for environmental bioremediation. *African Journal of Biotechnology*. 8 (22):6016-6027.
- Slater, J. H., Bull, A. T., Hardman, D. J. (1997). Microbial dehalogenation of halogenated alkanic acids. *Adv. Microb. Physiol.*, 38:133-176.
- Slater, J. H., Bull, A. T., and Hardman, D. J. (1995). Microbial Dehalogenation. *Biodegradation*. 6(3):181-189.
- Slater, J. H. (1994) Microbial dehalogenation of haloaliphatic compounds. In: Ratledge, C. (Ed.) *Biochemistry of Microbial Degradation*. (pp 379--421). Dordrecht, Netherlands: Kluwer Academic Publ.
- Slater, J. H., Weightman, A. J., and Hall, B. G. (1985). Dehalogenase genes of *Pseudomonas putida* PP3 on chromosomally located transposable elements. *Mol. Biol. Evol.*, 2: 557-567.
- Slater, J. H., Lovatt, D., Weightman, A. J., Senior, E. & Bull, A. T. (1979). The growth of *Pseudomonas putida* on chlorinated aliphatic acids and its dehalogenase activity. *Journal of General Microbiology* 114, 125-136.
- Smit, E., Leeflang, P., Glandorf, B., Elsas, J. D., and Wernars, K. (1999). Analysis of Fungal Diversity in the Wheat Rhizosphere by Sequencing of Cloned PCR-Amplified Genes Encoding 18S rRNA and Temperature Gradient Gel Electrophoresis. *Applied and Environmental Microbiology*. 65 (6): 2614-2621.
- Song, J. S., Lee, D. H., Lee, K., and Kim, C.K. (2003). Characteristics of several bacterial isolates capable of degrading chloroaliphatic compounds via hydrolytic dechlorination. *J. Microbiol.* 41(4):277-283.
- Staub, D. K. and Kohler, H. P. E. (1989). Microbial Degradation of β -Chlorinated Four-Carbon Aliphatic Acids. *Journal of Bacteriology*. 1428-1434.
- Stringfellow, J. M., Cairns, S. S., Cornish, A., and Cooper, R. A. (1997). Haloalkanoate dehalogenase II (DehE) of a *Rhizobium* sp. Molecular analysis of the gene and formation of carbon monoxide from trihaloacetate by the enzyme. *Eur. J. Biochem.*, 250: 789-793.

- Suida, J. F., and De Bernadis, J. F. (1973). Naturally occurring halogenated organic compounds. *Lloydia*, 36(2): 107-143.
- Thasif, S., Hamdan, S., and Huyop, F. (2009). Degradation of DL-2chloropropionic acid by bacterial dehalogenases that shows stereospecificity and its partial enzymatic characteristics. *Biotechnology*, 8(2): 264-269.
- Towner, K. J. and Cockayayne, A. (1993). *Molecular Methods for microbial Identification and Typing*. Chapman & Hall.
- Van Pée, K. H., and Unversucht, S. (2003). "Biological Dehalogenation and Halogenation Reactions." *Chemosphere*. 52:299–312.
- Weightman, A. J., Weightman, A. L., and Slater, J. H. (1982). Stereospecificity of 2-monochloropropionate dehalogenation by the two dehalogenases of *Pseudomonas putida* PP3: evidence of two different dehalogenation mechanisms. *J. Gen. Microbiol.*, 131: 1755-1762.
- White, T. J., Bruns, T., Lee, S., and Taylor, J. (1990). Amplification and direct sequencing of fungal ribosome RNA genes for phylogenetics. In Innis M. A., Gelfand D. H., Sninsky, J. J., and White T. J. (ed.). *PCR Protocols: a guide to methods and applications*. 315-322. New York, USA: Academic Press.
- Yan, S., Subramaniam, S. B., Tyagi, R. D., Surampalli, R. Y., and Zhang, T. C. (2010). Emerging contaminants of environmental concern: Source, transport, fate, and treatment. *Pract. Periodical Haz. Toxic Radioactive Waste Manage.*, 14(1): 2-20.
- Yokota, T., Fuse, H., Omori, T., and Minoda, Y. (1986). "Microbial Dehalogenation of Haloalkanes Mediated by Oxygenases or Halohydrolyase." *Agricul. And Biol. Chem.* 50:453 – 460.
- Yong W. W. (2010). Isolation, Characterization and Identification of 2,2-dichloropropionate degrading soil microorganism from Melaka rubber estate. Bachelor. Universiti Teknologi Malaysia, Skudai.
- Yu, M., Faan, Y. W., Chung, W. Y. K., and Tsang, J. H. S. (2007). Isolation and characterization of a novel haloacid permease from *Burkholderia cepacia* MBA4. *Appl. Environ. Microbiol.*, 73(15): 4874-4880.