AZO DYES DECOLORIZATION BY BACTERIA ORIGINATED FROM TEXTILE WASTEWATER

BAKHTIYAR AHMED MOHAMMED

A dissertation submitted in partial fulfillment of the requirement for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Bioengineering Universiti Teknologi Malaysia

JANUARY 2013

To my beloved mother Tuba Husssain, my beloved wife Dlxwaz Mohammed and all my loved ones

ACKNOWLEGEMENT

First of all, I would like to thanks Allah, for giving me the strength, patience and health to go through all obstacles in order to complete this research.

With a deep sense of gratitude, I wish to express my sincere thanks to my supervisor, Dr. Chong Chun Shiong who has been helpful and offered invaluable assistance, advice, guidance and high level inspirations for me in completing this research. Special thanks to PhD students who always been there to help me and without their assistance this project would not have been successful.

My greatest appreciation goes to my beloved mother, Tuba Hussian Ali and my brother, Dara Ahmed, who have raised me with their greatest love to be the person I am today. You have been with me all the times including the difficult times. And also, I would like to thank my beloved wife, Dlxwaz for supporting me spiritually throughout my studies. My special appreciation extends to my beloved brothers and sisters. Thanks for everything.

Last but not the least, I wish to express my deepest appreciation and gratitude to all my friends, for their great contribution, responsiveness and valuable information.

ABSTRACT

This study investigated the ability of previously isolated bacteria Escherichia sp., Acinetobacter sp., Enterobacter sp. and Klebsiella sp. in decolorizing Acid Orange 7 and Reactive Black 5. Screening results indicated the decolorization of Acid Orange 7 under facultative anaerobic condition was faster in comparison to Reactive Black 5. No decolorization of Acid Orange 7 and Reactive Black 5 was observed under aerobic conditions in the tested strains. Due to the reason that two of the strains (*Klebsiella* sp. and *Escherichia* sp.) were more potent decolorizers and these strains were able to remove Acid Orange 7 faster than Reactive Black 5, they were selected for further characterization on decolorization of Acid Orange 7. The effects of temperature, pH and concentration of nitrogen (NH₄)₂SO₄ on Acid Orange 7 decolorization were studied. For both of the strains, the optimum pH, temperature and (NH₄)₂SO₄ concentration were found to be at pH7, 37°C and 0.5g/L respectively. At pH 7, 93% and 75% of Acid Orange 7 was decolorized by Klebsiella sp. and *Escherichia* sp., respectively. No decolorization was detected at pH 4 or 10. At optimum temperature (37°C), Klebsiella sp. and Escherichia sp. were found to be able to decolorize 84% and 68% of Acid Orange 7, respectively. The decolorization percentage was detected to be highest in both strains (Klebsiella sp., 83% and *Escherichia* sp., 68%) when the cells were incubated in medium contained 0.5g/l of (NH₄)₂SO₄. Inducibility studies indicated Acid Orange 7 did not show an inducing effect on decolorization in *Klebsiella* sp. However, Acid Orange 7 was found to have a positive inducing effect on decolourization in Esherichia sp. It was observed that the *Esherichia* sp., which was pre-exposed to Acid Orange 7 displayed a higher percentage of decolorization compared with non pre-exposed cells.

ABSTRAK

Kajian ini menyiasat kebolehan bakteria yang telah dipencilkan sebelum ini iaitu Escherichia sp., Acinetobacter sp., Enterobacter sp., dan Klebsiella sp., dalam penyahwarnaan 'Acid Orange 7' dan 'Reactive Black 5'. Keputusan penyaringan menunjukkan penyahwarnaan oleh 'Acid Orange 7 dengan kaedah anaerob fakultatif lebih cepat berbanding 'Reactive Black 5'. Tiada penyahwarnaan berlaku oleh stren tersebut dalam 'Acid Orange 7' dan 'Reactive Black 5' dengan keadaan aerobik. Berdasarkan keputusan di mana dua strain (Klebsiella sp. dan Escherichia sp.) penyahwarnaannya lebih kuat dan strain ini berupaya untuk menyahwarna 'Acid Orange 7'lebih cepat berbanding dengan 'Reactive Black 5', strain ini telah dipilih untuk pencirian seterusnya dalam penyahwarnaan 'Acid Orange 7'. Kesan suhu, pH dan kepekatan nitrogen $(NH_4)_2SO_4$ ke atas penyahwarnaan 'Acid Orange 7' juga telah dikaji. Untuk kedua-dua strain, masing-masing menunjukkan pH yang optimum, suhu and kepekatan nitrogen (NH₄)₂SO₄ menunjukkan ialah 7, 37^oC dan 0.5 g/l. Pada pH 7, sebanyak 93% dan 75% 'Acid Orange 7' telah dinyahwarna oleh *Klebsiella* sp. dan *Escherichia* sp. Tiada penyahwarnaan berlaku pada pH 4 atau pH 10. Pada suhu yang optimum (37°C), Klebsiella sp. dan Escherichia sp. masingmasing menunjukkan penyahwarnaan sebanyak 84% dan 68% oleh 'Acid Orange 7'. Peratusan penyahwarnaan paling tinggi telah dikesan dari (*Klebsiella* sp., 83% dan Escherichia sp., 68%) apabila sel tersebut diinkubasi didalam medium yang mengandungi 0.5 g/l (NH₄)₂SO₄. Kajian pendorongan menunjukkan 'Acid Orange 7' tidak bertindak balas atas kesan dorongan dalam penyahwarnaan oleh Klebsiella sp. Walau bagaimanapun, 'Acid Orange 7' telah menunjukkan kesan dorongan yang positif dalam penyahwarnaan yang dilakukan oleh Escherichia sp. Escherichia sp.yang terdedah kepada 'Acid Orange 7' menunjukkan peratusan yang lebih tinggi berbanding sel yang tidak terdedah.

TABLE OF CONTENTS

CHAPTER	TITLE			
	DECLARATION	ii		
	DEDICATION	iii		
	ACKNOWLEDGEMENTS	iv		
	ABSTRACT	v		
	ABSTRAK	vi		
	TABLE OF CONTENTS	vii		
	LIST OF FIGURES			
	LIST OF TABLES			
	LIST OF SYMBOLS	XV		
	LIST OF ABBREVIATION	xvi		
1	INTRODUCTION	1		
	1.1 Introduction	1		
	1.2 Scopes of the study	2		
	1.3 Objective of the study	3		
2	LITERATURE REVIEW	4		

LITERATURE REVIEW		
2.1	History of Dyes	4
2.2	Natural and synthetic Dyes	5

2.3	Azo Dyes	5
2.4	Classification of Azo Dyes	6
	2.4.1 Disperse Azo Dyes	6
	2.4.2 Cationic Azo Dyes	7
	2.4.3 Metal-complex Dyes	7
	2.4.4 Direct Dyes	7
	2.4.5 Reactive Azo Dyes	7
	2.4.6 Anionic Azo Dyes	8
	2.4.7 Sulfur Dyes	9
	2.4.8 Solvent Dyes	9
	2.4.9 Mordant Dyes	10
	2.4.10 Vat Dyes	10
	2.4.11 Pigment Dyes	10
	2.4.12 Fluorescent brighteners	11
2.5	Contamination of azo dyes in industrial wastewater	11
2.6	Treatments of Azo Dyes	12
	2.6.1 Non biological treatments	13
	2.6.2 Biological treatments	14
	2.6.2.1 Algae	14
	2.6.2.2 Fungi	15
	2.6.2.3 Bacteria	16
2.7	Factors affecting bacterial azo dyes decolorization	19
	2.7.1 Oxygen and agitation	19
	2.7.2 Temperature	19
	2.7.3 pH	20
	2.7.4 Dye structure	21
	2.7.5 Carbon and Nitrogen source	21
	2.7.6 Redox mediator	22

2.8	Mechanism of Azo Dyes decolorization 23		
2.9	Azo Dyes decolorization by whole bacterial cell 2		
ME	THODOLOGY	25	
3.1	Experimental design	26	
3.2	Aseptic Technique	27	
3.3	Microorganisms	27	
3.4	Preparation of Medium	27	
	3.4.1 Nutrient broth (NB)	27	
	3.4.2 Nutrient agar (NA)	28	
	3.4.3 Chemically defined medium (CDM)	28	
3.5	Preparation of stock solution	28	
	3.5.1 Glycerol 1% (v/v)	29	
	3.5.2 Glucose 1% (w/v)	29	
	3.5.3 Acid Orange 7 1% (w/v)	29	
	3.5.4 Reactive Black 5 1% (w/v)	29	
3.6	Bacterial growth experiment on agar plates	30	
3.7	Screening for the ability of the selected bacteria on	30	
	decolorization of azo dyes		
	3.7.1 Preparation of bacteria inoculum	30	
	3.7.2 Measurements of optical density (OD)	31	
	3.7.3 Decolorization under facultative anaerobic	31	
	condition		
	3.7.4 Decolorization under aerobic condition	31	
3.8	Measurements of decolorization of azo dyes	32	
3.9	Optimization of Azo Dyes decolorization 32		
3.10	Inducibility study 33		

3

]	RESULI	FS AND D	ISCUSSION	35
4.1	Growt	h experime	ents	35
4.2	Decol	orization u	inder anaerobic condition	37
	4.2.1	Decoloriz	ation of Acid Orange 7 under	37
		facultativ	e anaerobic condition	
	4.2.2	Decolori	zation of Reactive Black 5 under	38
		facultativ	ve anaerobic condition	
4.3	Decol	orization u	inder aerobic condition	40
	4.3.1	Decolori	zation of Acid Orage 7	40
	4.3.2	Decolori	zation of Reactive Black 5	41
4.4	Acid (Orange 7 d	ecolorization pattern by	42
	Esche	<i>richia</i> sp. a	and <i>Klebsiella</i> sp.	
4.5	Optim	ization stu	ıdy	44
	4.5.1	Tempera	ture	44
		4.5.1.1	Effect of temperature on	44
			decolorization of Acid Orange 7	
			by <i>Escherichia</i> sp.	
		4.5.1.2	Effect of temperature on	45
			decolorization of Acid Orange 7	
			by <i>Klebsiella</i> sp.	
	4.5.2	Effect of	pН	46
		4.5.2.1	Effect of pH on decolorization of	47
			Acid Orange 7 by Esherichia sp.	
				48
		4.5.2.2	Effect of pH on decolorization of	
			Acid Orange 7 by Klebsiella sp.	
	4.5.3	Effect of	f nitrogen concentration	49
		4.5.3.1	Effect of nitrogen concentration	49
			on decolorization of Acid	
			Orange 7 by <i>Escherichia</i> sp.	

4

			4.5.3.2	Effect of nitrogen source	50
				concentration on decolorization	
				of Acid Orange 7 by Klebsiella	
				sp.	
	4.6	Inducib	ility stud	ies	51
		4.6.1	Inducibil	ity study in <i>Esherichia</i> sp.	52
		4.6.2	Inducibi	lity study in <i>Klebsiella</i> sp.	53
5	CON	ICLUSIC	ONS ANI	D FUTURE WORK	54
	5.1	Conclusio	ons		54
	5.2	Future we	ork		55
	REF	ERENCI	ES		56-66

	56-66

LIST OF FIGURE

FIGURE NO	TITLE	PAGE
2.1	An example of reactive dye (Reactive Black 5)	8
2.2	chemical structure of acid orange 7	9
2.3	Treatment of textile wastewater by different methods	13
2.4	Decolorization and complete mineralization of azo dyes during anaerobic-aerobic treatment	24
3.1	An overall flow of experiments	26
4.1	Single culture of bacteria (a) <i>Enterobacter</i> sp. (b) <i>Acinetobacter</i> sp. (c) <i>Escherichia</i> sp. (d) <i>Klebsiella</i> sp.	36
4.2	Decolorization of Acid Orange 7 by bacteria (a) <i>Enterobacter</i> sp., (b) <i>Acinetobacter</i> sp., (c) <i>Escherichia</i> sp. and (d) <i>Klebsiella</i> sp. after 60 hours	37
4.3	Decolorization of Reactive Black 5 by (a) <i>Enterobacter</i> sp. (b) <i>Acinetobacter</i> sp. (c) <i>Escherichia</i> sp. and (d) <i>Klebsiella</i> sp. after 6 days.	39
4.4	Decolorization of acid Orange 7 under aerobic condition by (a) <i>Enterobacter</i> sp. (b) <i>Acinetobacter</i> sp. (c) <i>Escherichia</i> sp. (d) <i>Klebsiella</i> sp.	40
4.5	Decolorization of Reactive Black 5 under aerobic condition by (a) <i>Enterobacter</i> sp. (b) <i>Acinetobacter</i> sp. (c) <i>Escherichia</i> sp. (d) <i>Klebsiella</i> sp.	41

- 4.6 Percentage of decolorization of Acid Orange 7 using 43 single culture of bacteria
- 4.7 Percentage of Acid Orange 7 decolorization (%) by 45 *Escherichia* sp. at different temperature.
- 4.8 Percentage of Acid Orange 7 decolorization (%) by 46 *Klebsiella* sp. under different temperature of incubation.
- 4.9 Percentage of Acid Orange 7 decolorization (%) by 47 *Escherichia* sp. at different pH.
- 4.10 Percentage of Acid Orange 7 decolorization (%) by 48 *Klebsiella* sp. with different pH of medium.
- 4.11 Percentage of Acid Orange 7 decolorization (%) by 50 *Escherichia* sp. with different concentration of nitrogen source (NH₄)₂ SO₄.
- 4.12 Percentage of Acid Orange 7 decolorization (%) by 51 *Klebsiella* sp. with different concentration of nitrogen source (NH₄)₂ SO₄.
- 4.13 Time course of the decolorization of Acid Orange 7 by 52 pre-exposed and non-exposed cells of *Escherichia* sp.
- 4.14 Time course of the decolorization of Acid Orange 7 by 53 preadapted and nonadapted cells *Klebsiella* sp.

LIST OF TABLE

TABLE NO	TITLE	PAGE
2.1	List of different species of azo dye decolorizing bacteria	18

LIST OF SYMBOLS

μm Micro milliliter -°C Celcius _ G Gram -L Liter -Milliliter mL g/L Gram per liter -Mg/L Milligram per liter -Nanometer nm -Rotation per minute rpm -Volume per volume v/v -Weight per volume W/Vh Hour -

LIST OF ABBREVIATIONS

AO 7	-	Acid Orange 7
RB5	-	Reactive Black 5
NaOH	-	Sodium Chloride
HCl	-	Hydrochloric acid
K ₂ HPO ₄	-	Dipotassium phosphate
KH ₂ PO ₄	-	Mono potassium phosphate
MgSO ₄ 7H ₂ O	-	Magnesium Sulphate Heptahydrate
CaCl ₂	-	calcium chloride
(NH ₄) ₂ SO ₄	-	Ammonium sulphate
OD	-	Optical density
UV	-	Ultraviolet

CHAPTER 1

INTRODUCTION

1.1 Introduction

Water pollution is a global environmental problem. It occurs when harmful compounds, which include organic substances and heavy metals, are discharged from the industries into water bodies without proper treatments (Joshni *et al.*, 2011). Among the pollutants, artificial dyes are one of them. The uses of artificial dyes in the textile industries have increased due to their ease and it is cheap to synthesis, high stability to temperature and has more variety in color compared with natural dyes (Ganem *et al.*, 2011)

Azo dyes constitute the largest class of synthetic dyes, which are aromatic compounds that contain one or more -N=N- groups (Pandy *et al.*, 2007). In dyes industries, it is recorded that 80% of total amount of dyes, which are produced annually, are composed of azo dye (Tripathi *et al.*, 2011). It is found that azo dyes are toxic, highly colored and can cause water contamination (Zolliger, 1991; Ganghua *et al.*, 2011).

2

Azo dyes are considered to be organic pollutant (Jonstrup *et al.*, 2011). During the dyeing process, a significant amount of azo dyes cannot bind with the fiber (Forgacs *et al.*, 2004). Subsequently they affect the aquatic life. In addition, these dyes were found to have mutagenic and carcinogenic effect on human body and other organisms (Robinson *et al.*, 2001). Therefore, the suitable treatment to remove this dye is very important.

Several physical-chemical methods such as photo catalytic degradation, filtration, and activated carbon could be used to eliminate the color from the waste water. The problems with these methods are that these methods caused the formation of some harmful side products, producing a lot of sludge and generally expensive (Tripathi *et al.*, 2011). The use of microbes to biodegrade azo dye is a better alternative because this method is cheaper and less accumulation of sludge. Many of microorganisms have been found to have the ability to remove color. These included bacteria, fungi and yeasts (Robinson *et al.*, 2001).

1.3 Objectives of the study

Three objectives of this study were:

- 1. To screen the ability of the selected bacteria on decolorization of azo dyes.
- 2. To optimize the decolorization process in the selected bacteria's whole cells, specifically in pH, temperature and concentration of nitrogen source.
- 3. To determine whether azo dye (substrate) can induce the decolorization process in the tested bacteria.

1.2 Scope of the study

This research focused on biological decolorization of azo dyes Acid Orange 7 and Reactive Black 5 using individual culture of bacteria (*Escherichia* sp., *Acinetobacter* sp. *Enterobacter* sp. and *Klebsiella* sp.), in order to obtain the best candidate that was able to remove color. The effect of three parameters (temperature, pH and nitrogen source $(NH_4)_2$ SO₄ concentration) on decolourization were investigated. Inducibility studies were carried out to determine if the azo dye (substrate) is the inducing agent for decolorization process in the tested bacteria.

References

- Acuner, E., and Dilek, F. B. (2004). Treatment of tectilon yellow 2G by Chlorella vulgaris. *Process Biochemistry*, *39*(5), 623-631.
- Al-Kdasi, A., Idris, A., Saed, K., and Guan, C. T. (2004). Treatment of textile wastewater by advanced oxidation processes-a review. *Global nest: the Int. J*, 6(3), 222-230.
- Allègre, C., Moulin, P., Maisseu, M., and Charbit, F. (2006). Treatment and reuse of reactive dyeing effluents. *Journal of Membrane Science*, 269(1–2), 15-34.
- Asad, S., Amoozegar, M., Pourbabaee, A., Sarbolouki, M., and Dastgheib, S. (2007). Decolorization of textile azo dyes by newly isolated halophilic and halotolerant bacteria. *Bioresource Technology*, 98(11), 2082-2088.
- Barragán, B. E., Costa, C., and Carmen Márquez, M. (2007). Biodegradation of azo dyes by bacteria inoculated on solid media. *Dyes and Pigments*, 75(1), 73-81.
- Bayoumi, R.A1. Musa, S.M.2, Bahobil, A.S.3, Louboudy, S.S.4 and El-Sakawey,
 T.A. (2010). Biodecolorization and Biodegradation of Azo dyes by Some
 Bacterial isolates. J. Appl. Environ. Biol. Sci., 1(1), 1-25.
- Brás, R., Isabel A. Ferra, M., Pinheiro, H. M., and Gonçalves, I. C. (2001). Batch tests for assessing decolourisation of azo dyes by methanogenic and mixed cultures. *Journal of Biotechnology*, 89(2–3), 155-162.
- Burkinshaw, S. M., and Lagonika, K. (2006). Sulphur dyes on nylon 6,6. Part 3.
 Preliminary studies of the nature of dye–fibre interaction. *Dyes and Pigments*, 69(3), 185-191.

- Chacko, J. T., and Subramaniam, K. (2010). Enzymatic Degradation of Azo Dyes–A Review.
- Chang, J. S., Chen, B.Y., and Lin, Y. S. (2004). Stimulation of bacterial decolorization of an azo dye by extracellular metabolites from Escherichia coli strain NO₃. *Bioresource Technology*, 91(3), 243-248.
- Chang, J. S., Chou, C., Lin, Y. C., Lin, P. J., Ho, J. Y., and Lee Hu, T. (2001). Kineti characteristics of bacterial azo-dye decolorization by Pseudomonas luteola. *Water Research*, 35(12), 2841-2850.
- Chang, J. S., and Lin, Y. C. (2008). Fed-Batch Bioreactor Strategies for Microbial Decolorization of Azo Dye Using a Pseudomonasluteola Strain. *Biotechnology* progress, 16(6), 979-985.
- Chang, J.S., and Kuo, T.S. (2000). Kinetics of bacterial decolorization of azo dye with Escherichia coli NO3. *Bioresource Technology*, 75(2), 107-111.
- Chen, B.Y., and Chang, J.S. (2007). Assessment upon species evolution of mixed Consortia for azo dye decolorization. *Journal of the Chinese Institute of Chemical Engineers*, 38(3–4), 259-266.
- Chen, C.Y., Cheng, M. C., and Chen, A.H. (2012). Photocatalytic decolorization of Remazol Black 5 and Remazol Brilliant Orange 3R by mesoporous TiO2. *Journal of Environmental Management*, 102(0), 125-133.
- Chen, K.C., Wu, J.Y., Liou, D.J., and Hwang, S.C. J. (2003). Decolorization of the textile dyes by newly isolated bacterial strains. *Journal of Biotechnology*, *101*(1), 57-68.
- Chen, K.M., Lin, L.H., Wang, C. F., and Hwang, M.C. (2010). Interactions between new multi-anionic surfactants and direct dyes and their effects on the dyeing of cotton fabrics. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 356(1–3), 46-50.
- Chompuchan, C., Satapanajaru, T., Suntornchot, P., and Pengthamkeerati, P. (2009).
 Decolorization of Reactive Black 5 and Reactive Red 198 using nanoscale
 Zero valent iron. *Proceedings of the World Academy Science, Engineering and Technology*, 37, 130-134.

- Dafale, N., Wate, S., Meshram, S., and Nandy, T. (2008). Kinetic study approach of remazol black-B use for the development of two-stage anoxic–oxic reactor for decolorization/biodegradation of azo dyes by activated bacterial consortium. *Journal of Hazardous Materials*, 159(2–3), 319-328.
- Daneshvar, N., Aber, S., and Hosseinzadeh, F. (2008). Study Of CI acid orange 7 removal In contaminated water by photo oxidation processes. *global nest journal*, *16*, 23-10.
- Daneshvar, N., Ayazloo, M., Khataee, A. R., and Pourhassan, M. (2007). Biological decolorization of dye solution containing Malachite Green by microalgae Cosmarium sp. *Bioresource Technology*, 98(6), 1176-1182.
- Daneshvar, N., Salari, D., and Khataee, A. R. (2003). Photocatalytic degradation of azo dye acid red 14 in water: investigation of the effect of operational parameters. *Journal of Photochemistry and Photobiology A: Chemistry*, 157(1), 111-116.
- Dong. X., Zhou, J., Liu, Y. (2003). Peptone-induced biodecolorization of reactive Brilliant blue (KN-R) by Rhodocycus gelatinosus XL-1. Process Biochem., 39: 89–94.
- Dos Santos, A. B., Cervantes, F. J., and van Lier, J. B. (2007). Review paper on current technologies for decolourisation of textile wastewaters: perspectives for anaerobic biotechnology. *Bioresource Technology*, 98(12), 2369-2385.
- Elmorsi, T. M., Riyad, Y. M., Mohamed, Z. H., and Abd El Bary, H. M. H. (2010). Decolorization of Mordant red 73 azo dye in water using H2O2/UV and photo-Fenton treatment. *Journal of Hazardous Materials*, *174*(1–3), 352-358.
- Forgacs, E., Cserháti, T., and Oros, G. (2004). Removal of synthetic dyes from wastewaters: a review. *Environment International*, *30*(7), 953-971.
- Franciscon, E., Zille, A., Fantinatti-Garboggini, F., Silva, I. S., Cavaco-Paulo, A., and Durrant, L. R. (2009). Microaerophilic–aerobic sequential decolourization/biodegradation of textile azo dyes by a facultative Klebsiella sp. strain VN-31. *Process Biochemistry*, 44(4), 446-452.

- Ganghua, Y., Yucai, H., Zhiqiang, C., Xiyue, Z., Liqun, W. and Li, W. (2011). Isolation and characterization of *Pseudomonas putida* WLY for reactive brilliant red x-3bdecolorization. African Journal of Biotechnology Vol. 10(51), pp. 10456-10464.
- Gessesse, B. (2009). Dye Biodegradation Using Alkalophillic Consortia in Anaerobic-Aerobic Bioprocess. Master Thesis. *School of Graduate Studies Faculty of Science Biotechnology Program.* Addis Ababa University.
- Ghanem, K. M., Al-Garni, S. M., and Biag, A. K. (2011). Statistical optimization of cultural conditions for decolorization of methylene blue by mono and mixed bacterial culture techniques. *Afr. J. Microbiol. Res*, 5(15), 2187-2197.
- Ghanem, K. M., Al-Fassi, F. A., and Biag, A. K. (2012). Optimization of methyl orange decolorization by mono and mixed bacterial culture techniques using statistical designs. *African Journal of Microbiology Research*, 6(12), 2918-2928.
- Gou, M., Qu, Y., Zhou, J., Ma, F., and Tan, L. (2009). Azo dye decolorization by a new fungal isolate, Penicillium sp. QQ and fungal-bacterial cocultures. *Journal* of Hazardous Materials, 170(1), 314-319.
- Guivarch, E., Trevin, S., Lahitte, C., and Oturan, M. A. (2003). Degradation of azo dyes in water by electro-Fenton process. *Environmental Chemistry Letters*, 1(1), 38-44.
- Gurulakshmi. M, Sudarmani. D.N.P and Venba. R. (2008). Biodegradation of Leather Acid dye by *Bacillus subtilis*. Advanced Biotech.
- Hosseini Koupaie, E., Alavi Moghaddam, M. R., and Hashemi, S. H. (2011). Posttreatmentof anaerobically degraded azo dye Acid Red 18 using aerobic moving bed biofilm process: Enhanced removal of aromatic amines. *Journal of Hazardous Materials*, 195(0), 147-154.
- Ibrahim, Z. (2011). Decolourisation of Reactive Black 5 Using paenibacillus sp. Immobilised Onto Macrocomposite. *Journal of Bioremediation & Biodegradation*.

- Işik, M., and Sponza, D. T. (2003). Effect of oxygen on decolorization of azo dyes by Escherichia coli and Pseudomonas sp. and fate of aromatic amines. *Process Biochemistry*, 38(8), 1183-1192.
- Işik, M., and Sponza, D. T. (2007). Fate and toxicity of azo dye metabolites under batch long-term anaerobic incubations. *Enzyme and Microbial Technology*, 40(4), 934-939.
- Jadhav, S. U., Jadhav, M. U., Kagalkar, A. N., and Govindwar, S. P. (2008).
 Decolorization of Brilliant Blue G dye mediated by degradation of the microbial consortium of Galactomyces geotrichum and Bacillus sp. *Journal of the Chinese Institute of Chemical Engineers*, 39(6), 563-570.
- Jin, X. C., Liu, G. Q., Xu, Z. H., and Tao, W. Y. (2007). Decolorization of a dye industry effluent by Aspergillus fumigatus XC6. *Applied Microbiology and Biotechnology*, 74(1), 239-243.
- Jonstrup, M., Kumar, N., Murto, M., and Mattiasson, B. (2011). Sequential anaerobic–aerobic treatment of azo dyes: Decolourisation and amine degradability. *Desalination*, 280(1–3), 339-346.
- Joshni, T. C., Kalidass, S. (2011). Enzymatic degradation of azo dyes. Enternational J. of environmental sciences Volume 1, No 6, 2011.
- Kalyani, D. C., Telke, A. A., Dhanve, R. S., and Jadhav, J. P. (2009). Ecofriendly biodegradation and detoxification of Reactive Red 2 textile dye by newly isolated Pseudomonas sp. SUK1. *Journal of Hazardous Materials*, 163(2–3), 735-742.
- Khalid, A., Arshad, M., and Crowley, D. E. (2008). Accelerated decolorization of structurally different azo dyes by newly isolated bacterial strains. *Applied Microbiology and Biotechnology*, 78(2), 361-369.
- Khalid, A., Arshad, M., and Crowley, D. (2010). Bioaugmentation of azo dyes. *Biodegradation of Azo Dyes*, 1-37.
- Khehra, M. S., Saini, H. S., Sharma, D. K., Chadha, B. S., and Chimni, S. S. (2005). Comparative studies on potential of consortium and constituent pure bacterial isolates to decolorize azo dyes. *Water Research*, 39(20), 5135-5141.

- Khehra, M. S., Saini, H. S., Sharma, D. K., Chadha, B. S., and Chimni, S. S. (2006).
 Biodegradation of azo dye C.I. Acid Red 88 by an anoxic–aerobic sequential bioreactor. *Dyes and Pigments*, 70(1), 1-7.
- Kumar Garg, S., Tripathi, M., Singh, S. K., and Tiwari, J. K. (2012).
 Biodecolorization of textile dye effluent by Pseudomonas putida SKG-1 (MTCC 10510) under the conditions optimized for monoazo dye orange II color removal in simulated minimal salt medium. *International Biodeterioration & amp; Biodegradation*, 74(0), 24-35.
- Liu, G., Zhou, J., Wang, J., Zhou, M., Lu, H., and Jin, R. (2009). Acceleration of azo dye decolorization by using quinone reductase activity of azoreductase and quinone redox mediator. *Bioresource Technology*, 100(11), 2791-2795.
- Lucas, M., Mertens, V., Corbisier, A. M., and Vanhulle, S. (2008). Synthetic dyes decolourisation by white-rot fungi: Development of original microtitre plate method and screening. *Enzyme and Microbial Technology*, 42(2), 97-106.
- Mahmood, S., Arshad, M., Khalid, A., Nazli, Z. H., and Mahmood, T. (2011). Isolation and screening of azo dye decolorizing bacterial isolates from dyecontaminated textile wastewater. *Soil and Environment*, 30(1), 7-12.
- Mane, U., Gurav, P., Deshmukh, A., and Govindwar, S. (2008). Degradation of textile dye reactive navy–blue Rx (Reactive blue–59) by an isolated Actinomycete Streptomyces krainskii SUK–5. *Malaysian Journal of Microbiology*, 4(2), 1-5.
- Martins, M. A. M., Ferreira, I. C., Santos, I. M., Queiroz, M. J., and Lima, N. (2001). Biodegradation of bioaccessible textile azo dyes by Phanerochaete chrysosporium. *Journal of Biotechnology*, 89(2–3), 91-98.
- Méndez-Paz, D., Omil, F., and Lema, J. M. (2005). Anaerobic treatment of azo dye Acid Orange 7 under batch conditions. *Enzyme and Microbial Technology*, 36(2–3), 264-272.
- Mielgo, I., Moreira, M. T., Feijoo, G., and Lema, J. M. (2001). A packed-bed fungal bioreactor for the continuous decolourisation of azo-dyes (Orange II). *Journal* of Biotechnology, 89(2–3), 99-106.

- Mohan, S. V., and Sarma, N. C. (2009). Simulated acid azo dye wastewater Treatment using suspended growth configured sequencing batch reactor (SBR) under anoxic-aerobic-anoxic microenviroment. *Applied Ecology* and Environmental Research, 7(1), 25-34.
- Mohorčič, M., Teodorovič, S., Golob, V., and Friedrich, J. (2006). Fungal and enzymaticdecolourisation of artificial textile dye baths. *Chemosphere*, *63*(10), 1709-1717.
- Öztürk, A., and Abdullah, M. I. (2006). Toxicological effect of indole and its azo dye derivatives on some microorganisms under aerobic conditions. *Science of The Total Environment*, 358(1–3), 137-142.
- Pandey, A., Singh, P., and Iyengar, L. (2007). Bacterial decolorization and degradation of azo dyes. *International Biodeterioration & amp; Biodegradation*, 59(2), 73-84.
- Pearce, C. I., Lloyd, J. R., and Guthrie, J. T. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments*, 58(3), 179-196.
- Prasad, A. S. A., and KVB, R. (2010). Physico chemical characterization of textile effluent and screening for dye decolorizing bacteria. *Global Journal Of Biotechnology &Biochemistry*, 5(2), 80-86.
- Rai, H. S., Bhattacharyya, M. S., Singh, J., Bansal, T., Vats, P., and Banerjee, U. (2005). Removal of dyes from the effluent of textile and dyestuff manufacturing industry: a review of emerging techniques with reference to biological treatment. *Critical reviews in environmental science and technology*, 35(3), 219-238.
- Ramalho, P. A., Scholze, H., Cardoso, M. H., Ramalho, M. T., and Oliveira-Campos, A. (2002). Improved conditions for the aerobic reductive decolourisation of azo dyes by Candida zeylanoides *Enzyme and Microbial Technology*, *31*(6), 848-854.
- Robinson, T., McMullan, G., Marchant, R., and Nigam, P. (2001). Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. *Bioresource Technology*, 77(3), 247-255.

- Rungruangkitkrai, N., and Mongkholrattanasit, R. Eco-Friendly of Textiles Dyeing and Printing with Natural Dyes.
- Samantaa, A. K., and Agarwal, P. (2009). Application of natural dyes on textiles. *Indian Journal of Fibre & Textile Research*, *34*, 384-399.
- Sani, R. K., and Banerjee, U. C. (1999). Decolorization of triphenylmethane dyes and textile and dye-stuff effluent by Kurthia sp. *Enzyme and Microbial Technology*, 24(7), 433-437.
- Saratale, R. G., Saratale, G. D., Chang, J. S., and Govindwar, S. P. (2009).
 Decolorization and biodegradation of textile dye Navy blue HER by
 Trichosporon beigelii NCIM- 3326.. *Journal of Hazardous Materials*, 166(2–3), 1421-1428.
- Saratale, R. G., Saratale, G. D., Chang, J. S., and Govindwar, S. P. (2011). Bacterial decolorization and degradation of azo dyes: A review. *Journal of the Taiwan Institute of Chemical Engineers*, 42(1), 138-157.
- Seesuriyachan, P., Takenaka, S., Kuntiya, A., Klayraung, S., Murakami, S., and Aoki, K. (2007). Metabolism of azo dyes by Lactobacillus casei TISTR 1500 and effects of various factors on decolorization. *Water Research*, 41(5), 985-992.
- Singh, S.V. and Purohit, M.C. (2012). Application of eco-friendly naturaldye on wool fibers using combination of natural and chemical mordants. Journal of environmental research and technology, 2(2), 48-55.
- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of azo dyes. *Applied Microbiology and Biotechnology*, *56*(1), 69-80.
- Sugiura, W., Miyashita, T., Yokoyama, T., and Arai, M. (1999). Isolation of azodye-degrading microorganisms and their application to white discharge printing of fabric. *Journal of bioscience and bioengineering*, 88(5), 577-581.
- Syed, M., Sim, H., Khalid, A., and Shukor, M. (2009). A simple method to screen for azo-dye-degrading bacteria. *Journal of Environmental Biology*, 30(1), 89-92.

- Telke, A. A., Kalyani, D. C., Dawkar, V. V., and Govindwar, S. P. (2009). Influence of organic and inorganic compounds on oxidoreductive decolorization of *172*(1), 298-309.
- Telke, A., Kalyani, D., Jadhav, J., and Govindwar, S. (2008). Kinetics and mechanism of Reactive Red 141 degradation by a bacterial isolate Rhizobium radiobacter MTCC 8161. Acta Chimica Slovenica, 55(2), 320.
- Toh, Y. C., Yen, J. J. L., Obbard, J. P., and Ting, Y. P. (2003). Decolourisation of azo dyes by white-rot fungi (WRF) isolated in Singapore. *Enzyme and Microbial Technology*, 33(5), 569-575.
- Tony, B. D., Goyal, D., and Khanna, S. (2009). Decolorization of textile azo dyes by aerobic bacterial consortium. *International Biodeterioration & amp; Biodegradation*, 63(4), 462-469.
- Tripathi, A., and Srivastava, S. (2011). Ecofriendly Treatment of Azo Dyes:
 Biodecolorization using Bacterial Strains. *International Journal of Bioscience*, *Biochemistry and Bioinformatics*, 1, 37-40.
- Turhan, K., Durukan, I., Ozturkcan, S. A., and Turgut, Z. (2012). Decolorization of textile basic dye in aqueous solution by ozone. *Dyes and Pigments*, 92(3), 897-901.
- Van der Zee, F. P., and Villaverde, S. (2005). Combined anaerobic–aerobic treatment of azo dyes—A short review of bioreactor studies. *Water Research*, 39(8), 1425-1440.
- Venkata Mohan, S., Chandrasekhar Rao, N., Krishna Prasad, K., and Karthikeyan, J. (2002).Treatment of simulated Reactive Yellow 22 (Azo) dye effluents using Spirogyra species. *Waste Management*, 22(6), 575-582
- Vijayaraghavan, K., and Yun, Y. S. (2007). Utilization of fermentation waste (Corynebacterium glutamicum) for biosorption of Reactive Black 5 from aqueous solution. *Journal of Hazardous Materials*, 141(1), 45-52.
- Wang, C., Yediler, A., Lienert, D., Wang, Z., and Kettrup, A. (2003). Ozonation of an azo dye C.I. Remazol Black 5 and toxicological assessment of its oxidation products. *Chemosphere*, 52(7), 1225-1232.

- Wang, H., Zheng, X. W., Su, J. Q., Tian, Y., Xiong, X. J., and Zheng, T. L. (2009). Biological decolorization of the reactive dyes Reactive Black 5 by a novel isolated bacterial strain Enterobacter sp. EC3. *Journal of Hazardous Materials*, 171(1–3), 654- 659.
- Wang, H., Su, J. Q., Zheng, X. W., Tian, Y., Xiong, X. J., and Zheng, T. L. (2009).
 Bacterial decolorization and degradation of the reactive dye Reactive Red 180 by Citrobacter sp.CK3. *International Biodeterioration & amp; Biodegradation*, 63(4), 395-399.
- Wang, X., Cheng, X., Sun, D., and Qi, H. (2008). Biodecolorization and partial mineralization of Reactive Black 5 by a strain of Rhodopseudomonas palustris. *Journal of Environmental Sciences*, 20(10), 1218-1225.
- Xu, M., Guo, J., Zeng, G., Zhong, X., and Sun, G. (2006). Decolorization of anthraquinone dye by Shewanella decolorationis S12. *Applied Microbiology* and Biotechnology, 71(2), 246-251.
- Yan, H., and Pan, G. (2004). Increase in biodegradation of dimethyl phthalate by Closterium lunula using inorganic carbon. *Chemosphere*, *55*(9), 1281-1285.
- Yang, G., He, Y., Cai, Z., Zhao, X., and Wang, L. (2011). Isolation and characterization of Pseudomonas putida WLY for reactive brilliant red x-3b decolorization. *Afr J Biotechnol*, 10(51), 10456-10464.
- Yoo, E., Libra, J., and Adrian, L. (2001). Mechanism of decolorization of azo dyes in anaerobic mixed culture. *Journal of environmental engineering*, 127(9), 844-849.
- You, S. J., and Teng, J. Y. (2009). Anaerobic decolorization bacteria for the treatment of azo dye in a sequential anaerobic and aerobic membrane bioreactor. *Journal of the Taiwan Institute of Chemical Engineers*, 40(5), 500-504.
- Yu, J., Wang, X., and Yue, P. L. (2001). Optimal Decolorization and Kinetic Modeling of Synthetic Dyes by Pseudomonas Strains. *Water Research*, 35(15), 3579-3586.
- Zee, F. P. (2002). *Anaerobic azo dye reduction*. Ph. D. thesis. Wageningen University, Wageningen-Netherlands.

- Zollinger, H. (1987). Color chemistry: syntheses, properties, and applications of organic dyes and pigments: Wiley-VCH.
- Zollinger, H. (1991). Color chemistry: syntheses, properties, and applications of organic dyes and pigments: Wiley-VCH.