# LIQUID AND SOLID PHASE MICROEXTRACTION METHODS FOR THE ANALYSIS OF ORGANIC ENVIRONMENTAL POLLUTANTS

LOH SAW HONG

A thesis submitted in fulfilment of the requirements for the award of the degree of Doctor of Philosophy (Chemistry)

> Faculty of Science Universiti Teknologi Malaysia

> > MAY 2013

#### ACKNOWLEDGEMENTS

First and foremost, I would like to give my sincere thankfulness to my main supervisor, Prof. Dr. Mohd Marsin Sanagi, for his guidance and priceless support, scientific excitement, inspiration and patient throughout this research. I would like to thank my co-supervisors, Prof. Dr. Wan Aini Wan Ibrahim and Prof. Dr. Mohamed Noor Hasan and visiting Prof. Dr. Hassan Y. Aboul Enein for giving me suggestions and critical comments during SepSTec group meeting as well as reviewing my draft thesis and manuscripts.

I would like to thank my family members for their moral support. Though far away from my family, but I was blessed to have their full support and spiritual strength to complete my research and thesis writing. I sincerely appreciate the companion and entertainment from my helpful and humorous lab-mates, especially when I was upset. Not forgetting the science officers and lab assistants in Department of Chemistry, Faculty of Science, UTM, I would like to give my complete gratitude towards them for their helps.

Special thanks to UMT and MOHE for the study leave and financial sponsorship given to me for three years. Finally yet importantly, I would like to express my gratitude to Assoc. Prof. Dr. Marinah Arrifin for her prompt advice and suggestion during my first semester study in UTM.

#### ABSTRACT

This work involves the investigation of new approaches and applications in miniaturized sample preparation techniques based on liquid phase and solid phase microextractions. A two-phase hollow fiber liquid phase microextraction (HF-LPME) method combined with gas chromatography-mass spectrometry was developed for the determination of selected polycyclic aromatic hydrocarbons (PAHs) in fresh milk. Under optimized conditions, low detection limits (LODs) were obtained ranging from 0.07-1.4  $\mu$ g L<sup>-1</sup> with relative recoveries of 85-110% which were higher than those obtained by conventional solvent extraction for the volatile PAHs. Agarose film liquid phase microextraction (AF-LPME) was developed for the extraction and preconcentration of PAHs in environmental water samples. Agarose, a green polymer, has been manipulated for different microextraction approaches. Agarose film was used as an interface between donor and acceptor phases which allowed for selective extraction of the analytes under optimum conditions. Under the optimum extraction conditions, the method showed good linearity in the range of 0.1–200  $\mu$ g L<sup>-1</sup>, low limits of detection (0.01-0.04  $\mu$ g L<sup>-1</sup>) and satisfactory relative recoveries (92.9-104.7%). AF-LPME device proved to be low-cost and thus reuse or recycle of the film was not required to eliminate the analytes carry-over between runs. A new microextraction technique termed agarose gel liquid phase microextraction (AG-LPME) was developed for the extraction of PAHs in water. Solvent-impregnated agarose gel disc used in AG-LPME was prepared by slicing gelled agarose and exchanging the solvent from water to ethanol and then to 1octanol that functioned as the extractant and impregnation solvent. The solvent impregnated AG-LPME was found to be comparable with HF-LPME in terms of extraction efficiencies without solvent dissolution problems observed. The method offered high enrichment factors in the range of 89-177 and trace level LODs in the range of 9-14 ng  $L^{-1}$ . This technique combines extraction and preconcentration approaches using an environmentally-compatible solvent holder that fulfils the green chemistry concept. Due to the hydrophilic property of agarose, the selectivity of AG-LPME was evaluated on hydrophilic triazine herbicides. The AG-LPME showed significantly higher extraction efficiencies as compared to HF-LPME. The method offered superior enrichment factors in the range of 115-300 and trace LODs in the range of 0.02-0.04  $\mu$ g L<sup>-1</sup>. Multi-walled carbon nanotube-impregnated agarose film microextraction (MWCNT-AFME) combined with micro high performance liquid chromatography-ultraviolet detection has also been developed. The method utilized MWCNTs immobilized in agarose film which served as the adsorbent holder. The technique achieved trace LODs in the range of 0.1-50 ng  $L^{-1}$  for selected PAHs. The new MWCNT-AFME method was successfully applied to the analysis of spiked green tea beverage samples with good relative recoveries. The results supported the feasibility of agarose to serve as adsorbent holder in solid phase microextraction, thus saving the cost of chemical and waste disposal.

## ABSTRAK

Kerja ini melibatkan kajian pendekatan dan aplikasi baru dalam teknik mini penyediaan sampel berdasarkan pengekstrakan mikro fasa cecair dan pepejal. Kaedah pengekstrakan mikro fasa cecair membran gentian berongga (HF-LPME) jenis dua fasa digabung dengan kromatografi gas-spektrometri jisim dibangunkan untuk menentukan hidrokarbon aromatik polisiklik (PAHs) dalam susu segar. Pada keadaan optimum, had pengesanan (LOD) rendah (0.07-1.4 µg L<sup>-1</sup>) diperoleh dengan perolehan balik relatif 85-110% yang lebih tinggi daripada pengekstrakan pelarut konvensional untuk PAHs meruap. Pengekstrakan mikro fasa cecair filem agarosa (AF-LPME) dibangunkan untuk mengekstrak dan pra-memekatkan PAHs dalam air persekitaran. Agarosa, sejenis polimer hijau, telah dimanipulasikan untuk pendekatan pengekstrakan mikro berlainan. Filem agarosa digunakan sebagai antaramuka di antara fasa penderma dan penerima untuk membolehkan pengekstrakan selektif analit pada keadaan optimum. Pada keadaan pengekstrakan optimum, kaedah ini menunjukkan kelinearan baik dalam julat 0.1-200  $\mu$ g L<sup>-1</sup>, LOD rendah (0.01-0.04  $\mu$ g L<sup>-1</sup>) dan perolehan balik relatif memuaskan (92.9-104.7%). Oleh kerana peralatan AF-LPME berkos rendah, penggunaan semula atau pengitaran semula filem agarosa tidak diperlukan bagi menghindari pencemaran analit antara larian. Satu teknik pengekstrakan mikro baru dinamakan pengekstrakan mikro fasa cecair gel agarosa (AG-LPME) dibangunkan bagi menentukan PAHs dalam air. Cakera gel agarosa terkandung pelarut yang diguna dalam AG-LPME disediakan dengan memotong agarosa yang telah membentuk gel dan menukar pelarut daripada air kepada 1-oktanol yang berfungsi sebagai pelarut pengekstrak dan impregnasi. Kaedah ini didapati setanding dengan HF-LPME dari segi keberkesanan pengekstrakan tanpa masalah kehilangan pelarut. Kaedah ini menawarkan faktor pengkayaan yang tinggi (89-177) dan LOD aras surih dalam julat 9-14 ng  $L^{-1}$ . Teknik ini menggabungkan pengekstrakan dan pra-pemekatan menggunakan pemegang pelarut yang serasi dengan persekitaran dan bersesuaian konsep kimia hijau. Disebabkan sifat agarosa yang hidrofilik, kepilihan AG-LPME seterusnya dinilai menggunakan racun rumpai triazina hidrofilik. AG-LPME menunjukkan keberkesanan pengekstrakan yang ketara lebih tinggi berbandingkan HF-LPME. Kaedah ini menawarkan faktor pengkayaan unggul (115-300) dan LOD aras surih dalam julat 0.02-0.04  $\mu$ g L<sup>-1</sup>. Pengekstrakan mikro karbon tiub-nano berbilang dinding yang terkandung dalam filem agarosa (MWCNT-AFME) digabung dengan kromatografi cecair prestasi tinggi mikro-pengesanan ultra lembayung telah dibangunkan. Kaedah ini menggunakan karbon tiub-nano berbilang dinding yang tidak bergerak dalam filem agarosa untuk berfungsi sebagai pemegang bahan penjerap. Teknik ini mencapai LOD aras surih dalam julat 0.1-50 ng L<sup>-1</sup> bagi PAHs terpilih. Kaedah MWCNT-AFME baru ini telah berjaya diaplikasi dalam analisis minuman teh hijau yang dipakukan dengan perolehan semula relatif vang baik. Keputusan kajian ini menyokong kebolehan agarosa untuk berfungsi sebagai pemegang bahan penjerap pengekstrakan mikro fasa pepejal dan menjimatkan kos bahan kimia dan pelupusan sisa.

# TABLE OF CONTENTS

CHAPTER	TITLE			PAGE	
	DECLARATION				
	ACI	KNOW	LEDGEMENTS	iii	
	ABS	STRAC'	Г	iv	
	ABS	STRAK		V	
	TAE	BLE OF	CONTENTS	vi	
	LIS	T OF T	ABLES	xii	
	LIS	T OF F	IGURES	xiv	
	LIS	ГOFА	BBREVIATIONS	xviii	
	LIS	T OF S	YMBOLS	xxi	
	LIS	ГOFА	PPENDICES	xxiii	
1			CTION	1	
	1.1	Backg	round of the Problem	1	
	1.2	Staten	nent of Problem	2	
	1.3	Resear	rch Objectives	3	
	1.4	Scope	of the Study	3	
	1.5	Signif	icance of Study	4	
2	LIT	FRATI	JRE REVIEW	5	
-	2.1		ccurrence and Physical Properties of	5	
	2.1		celic Aromatic Hydrocarbons	5	
	2.2		ity of PAHs	7	
		2.2.1	PAHs-Induced Carcinogenesis	, 7	
		2.2.1	Effects of PAHs on the Immune System	, 7	
	2.2		-	•	
	2.3	PAHS	Residues and Legislation	8	

2.	.4	Samp	le Preparation Techniques for PAHs Residues	9
		in Wa	ter and Food	
		2.4.1	Extraction and Clean-up of PAHs in Water	10
		2.4.2	Extraction and Clean-up of PAHs in Food	10
			2.4.2.1 Fatty Food	11
			2.4.2.2 Non Fatty Food	12
2.	.5	Altern	ative Green Microextraction Techniques	13
		2.5.1	Supercritical Fluid Extraction	14
		2.5.2	Subcritical or Superheated Water Extraction	14
		2.5.3	Solid Phase Microextraction	16
			2.5.3.1 Basic Principles of SPME	18
		2.5.4	Stir Bar Sorptive Extraction (SBSE)	18
		2.5.5	Microextraction in Packed Syringe (MEPS)	20
		2.5.6	Membrane Protected Micro Solid Phase	21
			Extraction	
		2.5.7	Liquid Phase Microextraction	22
			2.5.7.1 Basic Principles of LPME	25
		2.5.8	Dispersive Liquid-liquid Microextraction	27
2.	.6	Summ	nary of Past Reports on PAHs Analysis	28
2.	.7	Triazi	ne Herbicides	34
2.	.8	Physic	cal and Chemical Properties of Agarose	37
2.	.9	Appli	cation of Agarose	39
D	ЕТ	ERMI	NATION OF POLYCYCLIC AROMATIC	40
H	IYE	OROCA	ARBONS IN FRESH MILK BY HOLLOW	
F	IBI	ER LIQ	QUID PHASE MICROEXTRACTION-GAS	
С	HF	ROMA	TOGRAPHY-MASS SPECTROMETRY	
3.	.1	Introd	uction	40
3.	.2	Exper	imental	42
		3.2.1	Chemicals and Reagents	42
		3.2.2	Materials	42
		3.2.3	Chromatographic Conditions	43

		3.2.4	Hollow Fiber Liquid Phase Microextraction	43
			(HF-LPME)	
		3.2.5	Saponification prior to Solvent Extraction	44
			Validation of Analytical Method	45
	3.3		ts and Discussion	45
	3.4	Concl	usions	51
4	AGA	AROSE	E FILM LIQUID PHASE	52
	MIC	CROEX	<b>TRACTION OF POLYCYCLIC</b>	
	ARC	OMAT	IC HYDROCARBONS IN WATER	
	4.1	Introd	uction	52
	4.2	Exper	imental	54
		4.2.1	Chemicals and Reagents	54
		4.2.2	Materials	54
		4.2.3	Chromatographic Conditions	55
		4.2.4	Preparation of Agarose Film	55
		4.2.5	Agarose Film Liquid Phase Microextraction	56
			(AF-LPME)	
		4.2.6	Validation of Analytical Method	57
		4.2.7	Sample Analysis	57
	4.3	Resul	ts and Discussion	58
		4.3.1	Optimization of AF-LPME	58
			4.3.1.1 Stirring Speed	58
			4.3.1.2 Extraction Time	59
			4.3.1.3 Salting Out Effect	60
			4.3.1.4 Agarose Concentration	61
			4.3.1.5 Acceptor Phase Volume	63
		4.3.2	Validation of AF-LPME	64
		4.3.3	Application of AF-LPME on Environmental	66
			Water Samples	
		4.3.4	Comparison with Other Reported Methods	67
	4.4	Concl	usions	70

SOI	LVENT	-IMPREGNATED AGAROSE GEL	71
LIQ	UID Pl	HASE MICROEXTRACTION FOR THE	
ANA	ALSIS	OF POLYCYCLIC AROMATIC	
HYI	DROCA	ARBONS AND TRIAZINE HERBICIDES	
5.1	Introd	luction	71
5.2	Exper	imental	73
	5.2.1	Chemicals and Reagents	73
	5.2.2	Materials	74
	5.2.3	Chromatographic Conditions	74
	5.2.4	Preparation of Solvent Impregnated Agarose	75
		Gel Disc	
	5.2.5	Solvent-Impregnated Agarose Gel Liquid	76
		Phase Microextraction (AG-LPME)	
	5.2.6	Polypropylene Hollow Fiber Liquid Phase	77
		Microextraction (HF-LPME)	
	5.2.7	Agarose Film Liquid Phase Microextraction	78
		(AF-LPME)	
	5.2.8	Sample Analysis	78
5.3	Result	ts and Discussion	78
	5.3.1	Optimization of Solvent-Impregnated AG-	78
		LPME for the Analysis of PAHs	
		5.3.1.1 Stirring Speed	79
		5.3.1.2 Extraction Time	80
		5.3.1.3 Agarose Concentration	81
		5.3.1.4 Length of Agarose Gel Disc	83
	5.3.2	Optimization of HF-LPME for the Analysis	84
		of PAHs	
	5.3.3	Comparison of Extraction Efficiencies of	86
		PAHs Among AG-LPME, AF-LPME and	
		HF-LPME.	
	5.3.4	Validation of AG-LPME for the Analysis of	88
		PAHs	

5

ix

	5.3.5	Application of AG-LPME on Drinking	89
		Water Samples for the Analysis of PAHs	
	5.3.6	Optimization of Solvent-Impregnated AG-	90
		LPME for the Analysis of Triazine	
		Herbicides	
		5.3.6.1 Stirring Speed	91
		5.3.6.2 Extraction Time	92
		5.3.6.3 Sample pH	93
		5.3.6.4 Salting Out Effect	94
	5.3.7	Optimization of HF-LPME for the Analysis	95
		of Triazine Herbicides	
	5.3.8	Comparison of Extraction Efficiencies of	97
		Triazine Herbicides between AG-LPME and	
		HF-LPME	
	5.3.9	Validation of AG-LPME for the Analysis of	99
		Triazine Herbicides	
5.4	Concl		101
MUI	LTI-W	usions	
MUI IMP	LTI-W. PREGN	usions ALLED CARBON NANOTUBE-	
MUI IMP MIC	LTI-W PREGN CROEX	usions ALLED CARBON NANOTUBE- ATED AGAROSE FILM	
MUI IMP MIC ARC	LTI-W PREGN CROEX	usions ALLED CARBON NANOTUBE- ATED AGAROSE FILM CTRACTION OF POLYCYCLIC IC HYDROCARBONS IN GREEN TEA	
MUI IMP MIC ARC	LTI-W PREGN CROEX DMATI VERAG	usions ALLED CARBON NANOTUBE- ATED AGAROSE FILM CTRACTION OF POLYCYCLIC IC HYDROCARBONS IN GREEN TEA	102
MUI IMP MIC ARC BEV	LTI-W PREGN CROEX DMATI VERAG Introd	usions ALLED CARBON NANOTUBE- ATED AGAROSE FILM TRACTION OF POLYCYCLIC IC HYDROCARBONS IN GREEN TEA	102 102
MUI IMP MIC ARC BEV 6.1	LTI-W PREGN CROEX DMATI VERAG Introd	usions ALLED CARBON NANOTUBE- ATED AGAROSE FILM CTRACTION OF POLYCYCLIC IC HYDROCARBONS IN GREEN TEA E Uction	102 102 103
MUI IMP MIC ARC BEV 6.1	LTI-W PREGN CROEX DMATI VERAG Introd Exper	ALLED CARBON NANOTUBE- ATED AGAROSE FILM CTRACTION OF POLYCYCLIC IC HYDROCARBONS IN GREEN TEA E uction imental Chemicals and Materials	102 102 103 103
MUI IMP MIC ARC BEV 6.1	LTI-W PREGN CROEX DMATI VERAG Introd Exper 6.2.1	ALLED CARBON NANOTUBE- ATED AGAROSE FILM TRACTION OF POLYCYCLIC IC HYDROCARBONS IN GREEN TEA E uction imental Chemicals and Materials	102 102 103 103
MUI IMP MIC ARC BEV 6.1	LTI-W. PREGN CROEX DMATI VERAG Introd Exper 6.2.1 6.2.2	ALLED CARBON NANOTUBE- ATED AGAROSE FILM TRACTION OF POLYCYCLIC IC HYDROCARBONS IN GREEN TEA E uction imental Chemicals and Materials Chromatographic Conditions	102 102 103 104
MUI IMP MIC ARC BEV 6.1	LTI-W. PREGN CROEX DMATI VERAG Introd Exper 6.2.1 6.2.2	ALLED CARBON NANOTUBE- ATED AGAROSE FILM TRACTION OF POLYCYCLIC C HYDROCARBONS IN GREEN TEA E uction imental Chemicals and Materials Chromatographic Conditions Preparation of Multi-Walled Carbon	102 102 103 103
MUI IMP MIC ARC BEV 6.1	LTI-W. PREGN CROEX DMATI VERAG Introd Exper 6.2.1 6.2.2	ALLED CARBON NANOTUBE- ATED AGAROSE FILM TRACTION OF POLYCYCLIC C HYDROCARBONS IN GREEN TEA C HYDROCARBONS IN GREEN TEA E uction imental Chemicals and Materials Chromatographic Conditions Preparation of Multi-Walled Carbon Nanotube-Impregnated Agarose Film	101 102 103 103 104 104
MUI IMP MIC ARC BEV 6.1	LTI-W. PREGN CROEX DMATI VERAG Introd Exper 6.2.1 6.2.2 6.2.3	ALLED CARBON NANOTUBE- ATED AGAROSE FILM TRACTION OF POLYCYCLIC C HYDROCARBONS IN GREEN TEA C HYDROCARBONS IN GREEN TEA E uction imental Chemicals and Materials Chromatographic Conditions Preparation of Multi-Walled Carbon Nanotube-Impregnated Agarose Film (MWCNT-AF)	102 102 103 104 104

6

		6.2.5	Validati	on of Analytical Method	106
		6.2.6	Sample	Analysis	106
	6.3	Result	ts and Dis	cussion	107
		6.3.1	Optimiz	ation of MWCNT-AFME	107
			6.3.1.1	Conditioning Solvent	107
			6.3.1.2	Sample Volume, Concentration of	107
				MWCNTs and Number of Films	
			6.3.1.3	Desorption Time and Desorption	111
				Solvent	
			6.3.1.4	Stirring Speed and Extraction Time	113
		6.3.2	Validati	on of MWCNT-AFME	115
		6.3.3	Applicat	tion of MWCNT-AFME on Green	117
			Tea Bev	verage Samples	
		6.3.4	Compar	ison with Other Reported Methods	117
	6.4	Concl	usions		119
7	CO	NCLUS	SIONS AI	ND FUTURE DIRECTIONS	120
	7.1	Concl	usions		120
	7.2	Future	e Directio	ns	122
REFERENC	CES				124

Appendices A - D

xi

143-146

## LIST OF TABLES

<b>TABLE</b>	NO.
--------------	-----

# TITLE

## PAGE

2.1	Physical properties of several PAHs.	6
2.2	Published methods for the extraction and determination of	29
	PAHs from milk samples.	
2.3	Published methods for the extraction and determination of	31
	PAHs from aqueous samples.	
2.4	Published methods for the extraction and determination of	34
	PAHs from tea samples.	
2.5	Published methods for the extraction and determination of	36
	triazine herbicides from aqueous samples.	
3.1	Validation data of HF-LPME of PAHs from milk.	49
3.2	Fat contents and slopes of calibration plots of different	49
	milk samples.	
3.3	PAHs residues in commercial fresh milk products (n=3).	50
3.4	Relative recovery study.	51
4.1	Characterization of agarose film.	63
4.2	Validation data of AF-LPME of PAHs from spiked river	65
	water sample, n=3	
4.3	Relative recovery study of AF-LPME on river water	66
4.4	Application of AF-LPME on environmental water	67
	samples.	
4.5	Comparison of the AF-LPME with other published	69
	methods for the extraction and determination of PAHs	
	from water samples.	
5.1	Validation data of AG-LPME of PAHs from spiked	89
	drinking water samples (n=3).	

5.2	Relative recovery studies of AG-LPME using PAHs	89
	spiked drinking water.	
5.3	Application of AG-LPME on drinking water samples for	90
	the analysis of PAHs (n=3).	
5.4	Validation data of AG-LPME of triazine herbicides using	100
	spiked drinking water samples (n=3).	
5.5	Relative recovery studies of AG-LPME using triazine	101
	herbicides spiked drinking and river water samples (n=3).	
6.1	Validation data of MWCNT-AFME of PAHs from green	115
	tea beverage samples (n=3).	
6.2	Relative recovery studies of MWCNT-AFME using spiked	116
	green tea beverage samples (n=3).	
6.3	Application of MWCNT-AFME on green tea beverage	117
	samples (n=3).	
6.4	Comparison of the MWCNT-AFME with other published	118
	methods for the extraction and determination of PAHs	
	from tea samples.	

## LIST OF FIGURES

## FIGURE NO.

## TITLE

## PAGE

2.1	Schematic of SPME procedures.	16
2.2	Stir bar coated with PDMS.	19
2.3	Schematic of SPMTE (See et al., 2010).	20
2.4	Design of membrane protected micro solid phase	21
	extraction.	
2.5	Configuration A and B in LPME (Psillakis and	23
	Kalogerakis, 2003).	
2.6	Two-phase and three-phase sampling modes LPME	24
	(Psillakis and Kalogerakis, 2003).	
2.7	Schematic of cone-shaped LPME (Sanagi et al., 2007).	25
2.8	Schematic of DLLME procedures.	27
2.9	Chemical structure of agarose.	37
2.10	(a) Basic repeat units of agarose; (b) Schematic of gelling	39
	process of agarose (Zhou et al., 2006a).	
3.1	Schematic of HF-LPME.	44
3.2	Effect of stirring speed on HF-LPME of PAHs in milk.	46
3.3	Effect of salting out on HF-LPME of PAHs in milk.	47
3.4	Effect of extraction time on HF-LPME of PAHs in milk.	48
4.1	(a) Agarose film and (b) glass tube that is used in agarose	56
	film liquid phase microextraction.	
4.2	Schematic of AF-LPME system.	57
4.3	Effect of stirring speed on AF-LPME of PAHs from	59
	water sample.	
4.4	Effect of extraction time on AF-LPME of PAHs from	60
	water sample.	

4.5	Effect of salting out on AF-LPME of PAHs from water sample.	61
4.6	Effect of agarose concentration on AF-LPME of PAHs from water samples.	62
4.7	FESEM image of the 0.8% agarose film.	63
4.8	Effect of acceptor phase volume on AF-LPME of PAHs	64
	from water sample.	
4.9	GC-MS analysis of four polycyclic aromatic	66
	hydrocarbons spiked at 100 $\mu$ g L <sup>-1</sup> of river water on	
	Agilent HP5 MS column (30 m $\times$ 0.25 mm i.d., 0.25 $\mu$ m	
	film thickness).	
5.1	(a) Agarose gel disc; (b) Agarose gel discs dipped in 70%	76
	ethanol solution.	
5.2	Schematic of AG-LPME system.	77
5.3	Effect of stirring speed on AG-LPME of PAHs from	80
	water sample.	
5.4	Effect of extraction time on AG-LPME of PAHs from	81
	water sample.	
5.5	Effect of agarose concentration on AG-LPME of PAHs	82
	from water sample.	
5.6	Effect of agarose disc length on AG-LPME of PAHs	83
	from water sample.	
5.7	Effect of stirring speed on HF-LPME of PAHs from	84
	water sample.	
5.8	Effect of extraction time on HF-LPME of PAHs from	85
	water sample.	
5.9	Comparison of extraction efficiencies among AG-LPME,	87
	AF-LPME and HF-LPME. Error bars followed by same	
	letter without string showed no significant difference	
	according to Anova, Tukey test p>0.05.	

5.10	GC-MS total ion chromatogram (TIC) of four PAHs	88
	obtained after (a) AG-LPME and (b) HF-LPME spiked at	
	40 $\mu$ g L <sup>-1</sup> of deionized water. Conditions: helium	
	constant flowrate of 1 mL min <sup>-1</sup> , oven temperature profile	
	was programmed at 150°C for 3 min, and then increased	
	to 250°C at 10°C min <sup>-1</sup> using Agilent HP5 MS column	
	(30 m $\times$ 0.25 mm i.d., 0.25 $\mu$ m film thickness).	
5.11	Effect of stirring speed on AG-LPME of triazines from	91
	water sample.	
5.12	Effect of extraction time on AG-LPME of triazines from	92
	water sample.	
5.13	Effect of sample pH on AG-LPME of triazines from	93
	water sample.	
5.14	Effect of salting out on AG-LPME of triazines from	94
	water sample.	
5.15	Effect of stirring speed on HF-LPME of triazines from	95
	water sample.	
5.16	Effect of extraction time on HF-LPME of triazines from	96
	water sample.	
5.17	Comparison of extraction efficiencies between AG-	98
	LPME and HF-LPME. Error bars followed by same letter	
	without string showed no significant difference according	
	to t-test, p>0.05.	
5.18	GC-MS total ion chromatogram (TIC) of three triazine	99
	herbicides obtained after (a) AG-LPME and (b) HF-	
	LPME spiked at 4 $\mu$ g L <sup>-1</sup> of deionized water. Conditions:	
	helium constant flowrate of 1 mL min <sup>-1</sup> , oven	
	temperature profile was programmed at 170°C, and then	
	increased to 206°C at 3°C min <sup>-1</sup> using Agilent HP5 MS	
	column (30 m $\times$ 0.25 mm i.d., 0.25 µm film thickness).	
5.19	GC-MS TIC of (a) drinking and (b) river water samples	100
	that were free from triazines and used for relative	
	recovery studies.	

6.1	Schematic drawing of MWCNT-AFME system.	106
6.2	Effect of sample volume on MWCNT-AFME of PAHs	108
	from water sample.	
6.3	Effect of concentration of MWCNTs on MWCNT-	109
	AFME of PAHs from water sample. The surface area of	
	the 0.1, 0.3 and 0.6% of MWCNTs impregnated within	
	the agarose film were 21.4, 37.6 and 52.0 m <sup>2</sup> g <sup>-1</sup> ,	
	respectively.	
6.4	FESEM image of the (a) 0.3% MWCNT-AF and (b) AF;	110
	the pictures on the right top corner of each image were	
	the actual image of the circular films.	
6.5	Effect of pieces of MWCNTs-AF on MWCNT-AFME of	111
	PAHs from water sample.	
6.6	Effect of desorption solvent on MWCNT-AFME of	112
	PAHs from water sample.	
6.7	Effect of stirring speed on MWCNT-AFME of PAHs	113
	from water sample.	
6.8	Effect of extraction time on MWCNT-AFME of PAHs	114
	from water sample.	
6.9	$\mu$ -HPLC-UV analysis of three PAHs spiked at 0.008 $\mu$ g	116
	$L^{-1}$ for both PHE and FLA and 5 µg $L^{-1}$ for BaP of green	
	tea beverage on Agilent ZORBAX Eclipse Plus $C_{18}$	
	column (2.1 $\times$ 100 mm, 3.5 $\mu m$ ). $\mu \text{-HPLC}$ conditions:	
	isocratic mobile phase ACN-water (80:20) (v/v), column	
	temperature at 25°C, flowrate at 0.2 mL min <sup>-1</sup> , injection	
	volume of 2 $\mu$ L and detection wavelength at 254 nm.	

# LIST OF ABBREVIATIONS

ACE	-	Acetone
ACT	-	Acetonitrile
AF	-	Agarose film
AF-LPME	-	Agarose film liquid phase microextraction
AG	-	Agarose gel
AG-LPME	-	Agarose gel liquid phase microextraction
AOAC	-	Association of Analytical Communities
ATR	-	Atrazine
BaP	-	Benzo[a]pyrene
BET	-	Brunauer Emmett Teller
CYN	-	Cyanazine
DAD	-	Diode array detector
DCM	-	Dichloromethane
DLLME	-	Dispersive liquid-liquid microextraction
DNA	-	Deoxyribonucleic acid
EF	-	Enrichment factor
EME	-	Electromembrane extraction
EtOH	-	Ethanol
EU	-	European Union
FD	-	Fluorescence detector
FESEM	-	Field emission scanning electron microscope
FLA	-	Fluoranthene
FLU	-	Fluorene
GC	-	Gas chromatography
HF	-	Hollow fiber
HF-LPME	-	Hollow fiber liquid phase microextraction
HPLC	-	High performance liquid chromatography

I.DInternal diameterIL-Ionic liquidIPA-Isopropyl alcoholLLE-Liquid-liquid extraction	
IPA - Isopropyl alcohol	
I IJ	
LLE - Liquid-liquid extraction	
LOD - Limit of detection	
LOQ - Limit of quantification	
LPME - Liquid phase microextraction	
LVI - Large volume injection	
MASE - Membrane assisted solvent extraction	
ME - Microextraction	
MEPS - Microextraction in packed syringe	
MRL - Maximum residue level	
MS - Mass spectrometry	
MWCNT-AF Multi-walled carbon nanotube-impregnated agarose	film
MWCNT-AFME - Multi-walled carbon nanotube-impregnated agarose	film
microextraction	
MWCNTs - Multi-walled carbon nanotubes	
NaCl - Sodium chloride	
PAHs - Polycyclic aromatic hydrocarbons	
PCBs - Polychlorinated biphenyls	
PDA - Photodiode array	
PDMS - Polydimethylsiloxane	
PE - Polyethylene	
PHE - Phenanthrene	
PP - Polypropylene	
PTV - Programmed temperature vaporization	
PYR - Pyrene	
RSD - Relative standard deviation	
RTP - Room temperature phosphorimetry	
SBSE - Stir bar sorptive extraction	
SDME - Single drop microextraction	
SE - Solvent extraction	

SEC	-	Secbumeton
SFE	-	Subcritical fluid extraction
SFO	-	solidification of floating organic drop
SIM	-	Selected ion monitoring
SIR	-	Solvent impregnated resin
SPE	-	Solid phase extraction
SPME	-	Solid phase microextraction
SPMTE	-	Solid phase membrane tip extraction
SWE	-	Subcritical or superheated water extraction
THF	-	Tetrahydrofuran
USEPA	-	United States Environmental Protection Agency
UV	-	Ultraviolet
WHO	-	World Health Organization
μHPLC	-	Micro high performance liquid chromatography
µ-SPE	-	Micro solid phase extraction

# LIST OF SYMBOLS

pK <sub>a</sub>	-	Acid dissociation constant
n	-	Amount of analyte extracted by the coating
$CO_2$	-	Carbon dioxide
$\mathbf{R}^2$	-	Coefficient of determination
$C_{W}$	-	Concentration of analyte in the aqueous sample solution
Co	-	Concentration of analyte in the organic extraction solvent
r	-	Correlation coefficient
°C	-	Degree Celsius
eV	-	Electronvolt
Κ	-	Equilibrium distribution coefficient
K <sub>fs</sub>	-	Fiber coating/sample matrix distribution constant
$V_{\mathrm{f}}$	-	Fiber coating volume
g	-	Gram
g mol <sup>-1</sup>	-	Gram per mole
h	-	Hour
$C^0_{w}$	-	Initial amount of the analyte
$C_0$	-	Initial concentration of the analyte in the sample
kV	-	Kilovolts
L	-	Liter
m	-	Meter
μ	-	Micro
μg	-	Microgram
µg g <sup>-1</sup>	-	Microgram per gram
µg kg <sup>-1</sup>	-	Microgram per kilogram
μg L <sup>-1</sup>	-	Microgram per liter
μm	-	Micrometer
mg	-	Milligram

$mg L^{-1}$	-	Milligram per liter
mL	-	Milliliter
mm	-	Millimeter
min	-	Minutes
ng	-	Nanogram
ng L <sup>-1</sup>	-	Nanogram per liter
K <sub>ow</sub>	-	1-octanol/water partitioning coefficients
ppb	-	Part per billion
%	-	Percent
р	-	Probability
rpm	-	Revolutions per minute
Vs	-	Sample volume
S	-	Seconds
$V_{w}$	-	Volume of aqueous sample solution
Vo	-	Volume of organic extraction solvent
w/v	-	Weight per volume

# LIST OF APPENDICES

## APPENDIX

## TITLE

### PAGE

А	List of Publications from this Study	143
В	List of Awards from this Study	144
С	List of Presentations Related to this Study	145
D	List of Patents Related to this Study	146

#### **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of the Problem**

Green Chemistry, a phrase first coined by the United States Environmental Protection Agency (USEPA) in the early 1990s as 'to promote innovative chemical technologies that reduce or eliminate the use or generation of hazardous substances in the design, manufacture and use of chemical products' (Environmental Protection Agency, 2008). Chemists are concerned about the future of the environment and are worried about what will happen if we continue to suffocate the earth with all the hazardous substance in chemical process. Therefore, it is important to design or develop greener chemical processes and to utilize sustainable development practices in order to eliminate environmental degradation and pollution.

Conventional extraction methods such as liquid-liquid extraction or solvent extraction based on organic solvents are generating hazardous wastes to the environment, and this has contradicted the concept of green chemistry which emphasises on chemical process or technology that improves the environment and quality of life (Sanghi and Srivastava, 2003).

In order to meet green chemistry requirements, eco-friendly extraction and innovations for a cleaner analysis is urged. Of this, solid phase microextraction (SPME) and liquid phase microextraction (LPME) have emerged as the latest green analysis to replace solid phase extraction (SPE) and liquid-liquid extraction (LLE).

The techniques are categorized as 'prevention' under the twelve principles of green chemistry proposed by Anastas and Warner in 1991 (Anastas and Warner, 2000).

#### **1.2** Statement of Problem

Although the green techniques such as SPME and LPME have been increasingly developed and modified, the application of the techniques are still confined to the research institutes and universities. This is mainly because most of the green techniques were in-house validated. The food manufacturers and private industries' chemists are still employing the conventional technique such as LLE that was declared as standard reference method by some of the associations who established the maximum residue levels of food and environmental pollutants. Therefore, the publication of the green techniques should be made available to those associations and the public.

As fresh milk is used as one of the substitutes for breast milk during infancy, a rapid, selective, sensitive method is required to ensure the fresh milk is safe to be consumed. The monitoring of environmental pollutants residue levels is important to establish new maximum permitted levels. Existing method that uses conventional solvent extraction and fat saponification to extract environmental pollutants from milk consumes large amount of organic solvent and the procedures are tedious. Therefore, an environmentally green extraction method shall be developed to replace the existing solvent extraction method in order to reduce organic solvent usage and disposal costs. The liquid phase microextraction (LPME) promotes the reduction of hazardous substances in chemical processes for sample extraction and analysis.

In order to resolve the solvent dissolution problem that occurs during LPME, hollow fiber liquid phase microextraction (HF-LPME) has been developed to replace the core single drop microextraction (SDME). The extraction solvent or acceptor phase is well protected under impregnated hollow fiber made of polypropylene. However, solvent depletion problems are still frequently reported even though hollow fiber is used. In addition, although polypropylene is a recycled thermoplastic, the economic and environmental benefit of recycling is always an argument issue of the environmentalists. Therefore, an alternative material to protect acceptor phase in LPME is an urge to solve those problems.

#### **1.3** Research Objectives

The objectives of this research are:

- a. To develop and apply hollow fiber liquid microextraction (HF-LPME) method combined with gas chromatography–mass spectrometry (GC-MS) for the analysis of polycyclic aromatic hydrocarbons (PAHs) in fresh milk;
- b. To develop an innovative LPME system using biodegradable material termed agarose film (AF) and solvent impregnated agarose gel (AG) LPME coupled with GC-MS for the analysis of PAHs in water samples;
- To evaluate the selectivity of hydrophilic agarose towards hydrophilic triazine herbicides using solvent impregnated AG-LPME;
- d. To develop an innovative multi-walled carbon nanotube-impregnated agarose film microextraction (MWCNT-AFME) combined with micro high performance liquid chromatography (µHPLC)-ultraviolet (UV) detection for the analysis of PAHs in green tea beverage samples.

### **1.4** Scope of the Study

Liquid phase microextraction techniques were thoroughly studied where the extraction performances of commercially available hollow fiber and innovations on biodegradable agarose were evaluated. Hollow fiber liquid phase microextraction (HF-LPME) technique was studied for the first time to extract lipophilic PAHs from fresh milk samples. Several important extraction parameters were optimized and sample pretreatment steps such as pH adjustment and ultrasonification of sample solutions have been omitted. Agarose which is generally used in biotechnology

analysis as a medium of deoxyribonucleic acid (DNA) electrophoresis separation were prepared into film and solvent impregnated gel forms to serve as barriers or membranes of LPME to replace the hollow fiber. Several extraction parameters were explored and the newly developed technique was compared with LPME using commercially available HF for PAHs analysis as the reference method. The newly developed solvent-impregnated agarose gel LPME was thoroughly studied on its selectivity towards analytes which are more hydrophilic. Triazine herbicides were selected as model compounds. Several extraction parameters were comprehensively optimized and the optimum conditions were applied for the analysis of environmental and drinking water samples. The results were compared with those obtained by HF-LPME. An innovative agarose-immobilized sorbent film microextraction was studied, where multi-walled carbon nanotube (MWCNT)impregnated agarose film was synthesized and applied as adsorbent film in solid phase microextraction (SPME). Several extraction parameters were thoroughly investigated and optimized for the analysis PAHs in green tea beverage samples.

#### 1.5 Significance of Study

Modern life depends on the petrochemical and chemical related industries most drugs, food, transportation, office equipments, paints, analysis and plastics are derived from oil and chemicals. It is difficult to imagine what will happen when petroleum-based starting materials become more expensive when the population growth is accelerated against world oil production. At the commencement of the new century, a shift in emphasis in green chemistry is apparent with the desire to develop more environmentally friendly routes. From a green chemistry viewpoint, the uses of green solvent and material have many advantages. The idea of "green" solvents or materials expresses the goal to minimize the environmental impact and reduce the waste produced and chemical related impact on human health resulting from the use of solvents in chemical process and analysis. For this reason, finding environmentally green-solvent and material have become a top priority of the chemists. Therefore, the use of less organic solvent and biodegradable material as extraction tools have formed the main thrust of a movement.

#### REFERENCES

- Abdel-Rehim, M. (2004). New Trend in Sample Preparation : On-Line Microextraction in Packed Syringe for Liquid and Gas Chromatography Applications I. Determination of Local Anaesthetics in Human Plasma Samples using Gas Chromatography-Mass Spectrometry. J. Chromatogr. B. 801, 317-321.
- Aguinaga, N., Campillo, N., Vinas, P., and Hernandez-Cordoba, M. (2007). Determination of 16 Polycyclic Aromatic Hydrocarbons in Milk and Related Products using Solid Phase Microextraction Coupled to Gas Chromatography-Mass Spectrometry. *Anal. Chim. Acta.* 596, 285-290.
- Aguinaga, N., Campillo, N., Vinas, P., and Hernandez-Cordoba, M. (2008). A Headspace Solid-Phase Microextraction Procedure Coupled with Gas Chromatography-Mass Spectrometry for the Analysis of Volatile Polycyclic Aromatic Hydrocarbons in Milk Samples. *Anal. Bioanal. Chem.* 391, 753-758.
- Alcudia-Leon, M.C., Lucena, R., Cardenas, S., and Valcarcel, M. (2011a). Stir Membrane Liquid-liquid Microextraction. J. Chromatogr. A. 1218, 869-874.
- Alcudia-Leon, M.C., Lucena, R., Cardenas, S., and Valcarcel, M. (2011b). Determination of Phenols in Water by Stir Membrane Liquid-Liquid-Liquid Microextraction Coupled to Liquid Chromatography with Ultraviolet Detection. J. Chromatogr. A. 1218, 2176-2181.
- Almeida, C., and Nogueira, J.M.F. (2012). Comparison of the Selectivity of Different Sorbent Phases for Bar Adsorptive Microextraction – Application to Trace Level Analysis of Fungicides in Real Matrices. J. Chromatogr. A. 1265, 7-16.
- Anastas, P.T., and Warner, J.C. (2000). *Green Chemistry: Theory and Practice*. (1<sup>st</sup> ed.) United States: Oxford University Press.
- Angels Olivella, M. (2006). Isolation and Analysis of Polycyclic Aromatic Hydrocarbons from Natural Water using Accelerated Solvent Extraction followed by Gas Chromatography-Mass Spectrometry. *Talanta*. 69, 267-275.

- Araki, R.Y., Dodo, G.H., Reimer, S.H., and Knight, M.M. (2001). Protocol for the Determination of Selected Neutral and Acidic Semi-Volatile Organic Contaminants in Fish Tissue. J. Chromatogr. A. 923(1-2), 177-185.
- Arthur, C.L., and Pawliszyn, J. (1990). Solid Phase Microextraction with Thermal Desorption using Fused Silica Optical Fibers. *Anal. Chem.* 62, 2145-2148.
- Asai, S., Watanabe, K., Sugo, T., and Saito, K. (2005). Preparation of an Extractant-Impregnated Porous Membrane for the High-Speed Separation of a Metal Ion. J. *Chromatogr. A.* 1094, 158-164.
- Aymard, P., Martin, D.R., Plucknett, K., Foster, T.J., Clark, A.H., and Nortn, I.T. (2001). Influence of Thermal History on the Structural and Mechanical Properties of Agarose Gels. *Biopolymers*. 59, 131-144.
- Babic, K., Driessen, G.H.M., van der Ham, A.G.J., and de Haan, A.B. (2007). Chiral Separation of Amino-Alcohols using Extractant Impregnated Resins. J. Chromatogr. A. 1142, 84-92.
- Babic, K., van der Ham, L., and de Haan, A. (2006). Recovery of Benzaldehyde from Aqueous Streams using Extractant Impregnated Resins. *React. Funct. Polym.* 66, 1494-1505.
- Balchen, M., Reubsaet, L., and Pedersen-Bjergaard, S. (2008). Electromembrane Extraction of Peptides. J. Chromatogr. A. 1194, 143-149.
- Baltussen, E., Sandra, P., David, F., and Cramers, C. (1999). Stir Bar Sorptive Extraction (SBSE), a Novel Extraction Technique for Aqueous Samples: Theory and Principles. J. Microcol. Sep. 11(10), 737-747.
- Barri, T., and Jonsson, J.A. (2008). Advances and Developments in Membrane Extraction for Gas Chromatography – Techniques and Applications. J. Chromatogr. A. 1186, 16-38.
- Basheer, C., Alnedhary, A.A., Madhava Rao, B.S., Valliyaveettil, S., and Lee, H.K. (2006). Development and Application of Porous Membrane-Protected Carbon Nanotube Micro-Solid-Phase Extraction Combined with Gas Chromatography/Mass Spectrometry. *Anal. Chem.* 78(8), 2853-2858.
- Basile, A., Jimenez-Carmona, M.M., and Clifford, A.A. (1998). Extraction of Rosemary by Superheated Water. J. Agric. Food Chem. 46, 5205-5209.

- Bianchi, F., Careri, M., Mangia, A., Mattarozzi M., and Musci, M. (2008). Experimental Design for the Optimization of the Extraction Conditions of Polycyclic Aromatic Hydrocarbons in Milk with a Novel Diethoxydiphenylsilane Solid Phase Microextraction Fiber. J. Chromatogr. A. 1196-1197, 41-45.
- Bishnoi, N.R., Mehta, U., Sain, U., and Pandit, G.G. (2005). Quantification of Polycyclic Aromatic Hydrocarbons in Tea and Coffee Samples of Mumbai City (India) by High Performance Liquid Chromatography. *Environ. Monit. Assess.* 107, 399-406.
- Bjorhovde, A., Halvorsen, T.G., Rasmussen, K.N., and Pedersen-Bjergaard, S. (2003). Liquid-Phase Micrextraction of Drugs from Human Breast Milk. *Anal. Chim. Acta.* 491, 155-161.
- Bjorseth, A., and Ramdahl, T. (1985). Handbook of Polycyclic Aromatic Hydrocarbon Volume 2 : Sources and Emission of PAH. (1<sup>st</sup> ed.) New York: Marcel Dekker.
- Blahusiak, M., Schlosser, S., and Martak, J. (2011). Extraction of Butyric Acid by a Solvent Impregnated Resin Containing Ionic Liquid. *React. Funct. Polym.* 71, 736-744.
- Bourdat-Deschamps, M., Davdin, J.J., and Barriuso, E. (2007). An Experimental Design Approach to Optimise the Determination of Polycyclic Aromatic Hydrocarbons from Rainfall Water using Stir Bar Sorptive Extraction and High Performance Liquid Chromatography-Fluorescence Detection. *J. Chromatogr. A.* 1167(2), 143-153.
- Burghoff, B., Cuypers, R., van Ettinger, M., Sudholter, E.J.R., Zuilhof, H., and de Haan, A.B. (2009). Evaluation of Tri-n-Octylamine Oxide as Phenol Extractant in a Solvent Impregnated Resin. *Sep. Purif. Technol.* 67, 117-120.
- Burghoff, B., Goetheer, E.L.V., and de Haan, A.B. (2008). Solvent Impregnated Resin for the Removal of Low Concentration Phenol from Water. *React. Funct. Polym.* 68, 1314-1324.
- Burghoff, B., Marques, J.S., van Lankvelt, B.M., and de Haan, A.B. (2010). Solvent Impregnated Resins for MTBE Removal from Aqueous Environments. *React. Funct. Polym.* 70, 41-47.
- Cejpek, K., Hajsova, J., Jehllckova, Z., and Merhaut, J. (1995). Simplified Extraction and Cleanup Procedure for the Determination of PAHs in Fatty and Protein-Rich Matrices. *Int. J. Environ. Anal. Chem.* 61(1), 65-80.

- Celik, Z.C., Can, B.Z., and Kocakerim, M.M. (2008). Boron Removal from Aqueous Solutions by Activated Carbon Impregnated with Salicylic Acid. J. *Hazard. Mater.* 152, 415-422.
- Charalabaki, M., Psillakis, E., Mantzavinos, D., and Kalogerakis, N. (2005). Analysis of Polycyclic Aromatic Hydrocarbons in Wastewater Treatment Plant Effluents using Hollow Fibre Liquid-Phase Microextraction. *Chemosphere*. 60, 690-698.
- Chimuka, L., Cukrowska, E., and Jonsson, J.K. (2004). Why Liquid Membrane Extraction is an Attractive Alternative in Sample Preparation. *Pure Appl. Chem.* 76, 707-722.
- Chung, T.L., Liao, C.J., and Chen, M.F. (2010). Comparison of Liquid-Liquid Extraction and Solid Phase Extraction for the Determination of Polycyclic Aromatic Hydrocarbons in the Milk of Taiwan. J. Taiwan Inst. Chem. Engrs. 41, 178-183.
- Cortina, J.L., Miralles, N., Sastre, A.M., Aguilar, M., Profumo, A., and Pesavento, M. (1993). Solvent-Impregnated Resins Containing Di-(2,4,4-Trimethylpentyl) Phosphinic Acid- II. Study of the Distribution Equilibria of Zn(II), Cu(II) and Cd(II). *React. Polym.* 21, 103-116.
- Cox, M., Rus-Romero, J.R., and Sheriff, T.S. (2004). The Application of Monmorillonite Clays Impregnated with Organic Extractants for the Removal of Metals from Aqueous Solution. Part II. The Preparation of Clays Impregnated with Commercial Solvent Extraction Reagents and Their Use for the Removal of Copper(II). *React. Funct. Polym.* 60, 215-222.
- Davila, D.R., Romero, D.L., and Burchiel, S.W. (1996). Human T Cells are Highly Sensitive to Suppression of Mitogenesis by Polycyclic Aromatic Hydrocarbons and This Effect is Differentially Reversed by α-Naphthoflavone. *Toxicol. Appl. Pharm.* 139(2), 333-341.
- Doong, R.A., Chang, S.M., and Sun, Y.C. (2000). Solid-Phase Microextraction for Determining the Distribution of Sixteen US Environmental Protection Agency Polycyclic Aromatic Hydrocarbons in Water Samples. J. Chromatogr. A. 879, 177-188.
- Ebrahimpour, B., Yamini, Y., and Esrafili, A. (2011). Extraction of Azole Antifungal Drugs from Milk and Biological Fluids using a New Hollow Fiber Liquid-Phase Microextraction and Analysis by GC-FID. *Chromatographia*. 74, 281-289.

- Eibak, L.E.E., Gjelstad, A., Rasmussen, K.E., and Pedersen-Bjergaard, S. (2012). Exhaustive Electromembrane Extraction of Some Basic Drugs from Human Plasma Followed by Liquid Chromatography-Mass Spectrometry. *J. Pharmaceut. Biomed. Anal.* 57, 33-38.
- Einsle, T., Paschke, H., Bruns, K., Schrader, S., Popp, P., and Moeder, M. (2006).
  Membrane-Assisted Liquid-Liquid Extraction Coupled with Gas Chromatography-Mass Spectrometry for Determination of Selected Polycyclic Musk Compounds and Drugs in Water Samples. J. Chromatogr. A. 1124, 196-204.
- Environmental Protection Agency. (2008). *Green Chemistry*. Retrieved December 28, 2011, from http://www.epa.gov/gcc/
- European Commission. (2002). Directorate C- Scientific Opinions SCF/CS/CNTM/PAH/29 ADD1 Final: Opinion of the Scientific Committee on Food on the Risks to Human Health of Polycyclic Aromatic Hydrocarbons in Food. Retrieved November 22, 2011, from http://ec.europa.eu/food/fs/sc/scf/out 153\_en.pdf
- European Union. (2006). *Commission Regulation (CE) No 208/2005*. Retrieved July 14, 2011, from http://www.ihta.org/word-documents/EU\_sets\_maximum\_levels\_ of\_PAHs\_in food.doc
- Farre, M., Perez, S., Goncalves, C., Alpendurada, M.F., and Barcelo, D. (2010). Green Analytical Chemistry in the Determination of Organic Pollutants in the Aquatic Environment. *Trends Anal. Chem.* 29(11), 1347-1362.
- Fetzer, J.C. (2000). Large (C≥24) Polycyclic Aromatic Hydrocarbons : Chemistry and Analysis. (1<sup>st</sup> ed.) New York: Wiley Interscience.
- Fledler, H., Cheung, C.K., and Wong, M.H. (2002). PCDD/PCDF, Chlorinated Pesticides and PAH in Chinese Teas. *Chemosphere*. 46(9), 1429-1433.
- Fotouhi, L., Yamini, Y., Molaei, S., and Seidi, S. (2011). Comparison of Conventional Hollow Fiber Based Liquid Phase Microextraction and Electromembrane Extraction Efficiencies for the Extraction of Ephedrine from Biological Fluids. J. Chromatogr. A. 1218, 8581-8586.
- Gao, Y., Zeng, Y., Zheng, L., and Li, L. (2009). Determination of Triazine Herbicides in Aqueous Samples using Solidification of a Floating Drop for Liquid-Phase Microextraction with Liquid Chromatography. *Anal. Lett.* 42, 1620-1631.

- Garcia-Falcon, M.S., Cancho-Grande, B., and Simal-Gandara, J. (2004a). Stirring Bar Sorptive Extraction in the Determination of PAHs in Drinking Waters. *Water Res.* 38, 1679-1684.
- Garcia-Falcon, M.S., Perez-Lamela, M., and Simal-Gandara, J. (2004b). Comparison of Strategies for Extraction of High Molecular Weight Polycyclic Aromatic Hydrocarbons from Drinking Waters. J. Agric. Food Chem. 52, 6897-6903.
- Ge, D., and Lee, H.K. (2011). Water Stability of Zeolite Imidazolate Framework 8 and Application to Porous Membrane-Protected Micro-Solid-Phase Extraction of Polycyclic Aromatic Hydrocarbons from Environmental Water Samples. J. Chromatogr. A. 1218, 8490-8495.
- Grova, N., Feidt, C., Crepineau, C., Laurent, C., Lafargue, P.E., Hachimi, A., and Rychen, G. (2002). Detection of Polycyclic Aromatic Hydrocarbon Levels in Milk Collected Near Potential Contamination Sources. J. Agric. Food Chem. 50, 4640-4642.
- Gupta, S., Saha, B., and Giri, A.K. (2002). Comparative Antimutagenic and Anticlastogenic Effects of Green Tea and Black Tea: A Review. *Mutat. Res.* 512, 37-65.
- Hagestuen, E.D., Arruda, A.F., and Campiglia, A.D. (2000). On the Improvement of Solid-Phase Extraction Room-Temperature Phosphorimetry for the Analysis of Polycyclic Aromatic Hydrocarbons in Water Samples. *Talanta*. 52, 727-737.
- Hall, B. (2008). Single Laboratory Validation : A Key Step along the Path to Official Methods of Analysis. Retrieved May 18, 2011, from http://www.aoac.org/ OMB/slv.pdf
- Harrak, R.E., Calull, M., Marce, R.M., and Borrull, F. (1996). Determination of Polycyclic Aromatic Hydrocarbon in Water by Solid-Phase Extraction Membranes. *Inter. J. Environ. Anal. Chem.* 64, 47-57.
- Hartonen, K., Inkala, K., Kangas, M., and Riekkola, M.L. (1997). Extraction of Polychlorinated Biphenyls with Water under Subcritical Conditions. J. Chromatogr. A. 785, 219-226.
- Hashemi, P., and Abolghasemi, M.M. (2006). Preparation of a Novel Optical Sensor for Low pH Values using Agarose Membranes as Support. *Sensor. Actuat. B-Chem.* 115, 49-53.

- Hashemi, P., Zarjani, R.A., Abolghasemi, M.M., and Olin, A. (2007). Agarose Film Coated Glass Slides for Preparation of pH Optical Sensors. *Sensor. Actuat. B-Chem.* 121, 396-400.
- Hauser, B., Popp, P., and Kleine-Benne, E. J. (2002). Membrane-Assisted Solvent Extraction of Triazines and other Semi Volatile Contaminants Directly Coupled to Large-Volume-Injection-Gas Chromatography-Mass Spectrometric Detection. J. Chromatogr. A. 963, 27-36.
- Hawthorne, S.B., Grabanski, C.B., Martin, E., and Miller, D.J. (2000). Comparisons of Soxhlet Extraction, Pressurized Liquid Extraction, Supercritical Fluid Extraction and Subcritical Water Extraction for Environmental Solids: Recovery, Selectivity and Effects on Sample Matrix. J. Chromatogr. A. 892, 421-433.
- Hawthorne, S.B., Yang, Y., and Miller, D.J. (1994). Extraction of Organic Pollutants from Environmental Solids with Sub- and Supercritical Water. *Anal. Chem.* 66(18), 2912-2920.
- Heemken, O.P., Theobald, N., and Wenclawiak, B.W. (1997). Comparison of ASE and SFE with Soxhlet, Sonication and Methanolic Saponification Extractions for the Determination of Organic Micropollutants in Marine Particulate Matter. *Anal. Chem.* 69(11), 2171-2180.
- Hou, L., and Lee, H.K. (2002). Application of Static and Dynamic Liquid Phase Microextraction in the Determination of Polycyclic Aromatic Hydrocarbons. J. Chromatogr. A. 976, 377-385.
- Hylton, K., and Mitra, S. (2007). Barrier Film Protected, and Mixed Solvent Optimized Micro-Scale Membrane Extraction of Methyl Carbamate Pesticides. J. Chromatgr. A. 1154, 60-65.
- Jarvenpaa, E., Huopalahti, R., and Tapanainen, P. (1996). Use of Supercritical Fluid Extraction-High Performance Liquid Chromatography in the Determination of Polynuclear Aromatic Hydrocarbons from Smoked and Broiled Fish. J. Liq. Chromatogr. Related Technol. 19(9), 1473-1482.
- Jeannot, M. A., and Cantwell, F. F. (1996). Solvent Microextraction into a Single Drop. *Anal. Chem.* 68(13), 2236-2240.
- Jiang, X., Basheer, C., Zhang, J., and Lee, H.K. (2005). Dynamic Hollow Fiber Supported Headspace Liquid Phase Microextraction. J. Chromatogr. A. 1087, 289-294.

- Kah, M., Zhang, X., Jonker, M.T.O., and Hofmann, T. (2011). Measuring and Modelling Adsorption of PAHs to Carbon Nanotubes Over a Six Order of Magnitude Wide Concentration Range. *Environ. Sci. Technol.* 45, 6011-6017.
- Kanimozhi, S., Basheer, C., Narasimhan, K., Liu, L., Koh, S., Xue, F., Choolani, M., and Lee, H.K. (2011). Application of Porous Membrane Protected Micro-Solid-Phase-Extraction Combined with Gas Chromatography-Mass Spectrometry for the Determination of Estrogens in Ovarian Cyst Fluid Samples. *Anal. Chim. Acta.* 687, 56-60.
- Kayali-Sayadi, M.N., Rubio-Barroso, S., Cuesta-Jimenez, M.P., and Polo-Diez, L.M. (1998). Rapid Determination of Polycyclic Aromatic Hydrocarbons in Tea Infusion Samples by High-Performance Liquid Chromatography and Fluorimetric Detection based on Solid-Phase Extraction. *Analyst.* 123, 2145-2148.
- Khalili Zanjani, M.R., Yamini, Y., Shariati, S., and Jonsson, J.A. (2007). A New Liquid-Phase Microextraction Method based on Solidification of Floating Organic Drop. Anal. Chim. Acta. 585, 286-293.
- King, A.J., Readman, J.W., and Zhou, J.L. (2004). Determination of Polycyclic Aromatic Hydrocarbons in Water by Solid-Phase Microextraction-Gas Chromatography-Mass Spectrometry. *Anal. Chim. Acta*. 523, 259-267.
- King, S., Meyer, J.S., and Andrews, A.R.J. (2002). Screening Method for Polycyclic Aromatic Hydrocarbon in Soil using Hollow Fiber Membrane Solvent Microextraction. J. Chromatogr. A. 982, 201-208.
- Kishikawa, N., Wada, M., Kuroda, N., Akiyama, S., and Nakashima, K. (2003).
  Determination of Polycyclic Aromatic Hydrocarbons in Milk Samples by High Performance Liquid Chromatography with Fluorescence Detection. J. Chromatogr. B. 789, 257-264.
- Kjelsen, I.J.O., Gjelstad, A., Rasmussen, K.E., and Pedersen-Bjergaard, S. (2008). Low-Voltage Electromembrane Extraction of Basic Drugs from Biological Samples. J. Chromatogr. A. 1180, 1-9.
- Kokosa, J.M., Przyjazny, A., and Jeannot, M.A. (2009). *Solvent Microextraction: Theory and Practice*. (1<sup>st</sup> ed.) New Jersey: John Wiley & Sons.
- Kolarovic, L., and Traitler, H. (1982). Determination of Polycyclic Aromatic Hydrocarbon in Vegetables Oils by Caffeine Complexation and Glass Capillary Gas Chromatography. J. Chromatogr. A. 237(2), 263-272.

- Lancas, F.M., Queiroz, M.E.C., Grossi, P., and Olivares, I.R.B. (2009). Recent Developments and Applications of Stir Bar Sorptive Extraction. J. Sep. Sci. 32, 813-824.
- Larsson, B.K., Eriksson, A.T., and Cervenka, M. (1987). Polycyclic Aromatic Hydrocarbon in Crude and Deodorized Vegetable Oils. *JAOCS*. 64(3), 365-370.
- Lee, T.P., Saad, B., Khayoon, W.S., and Salleh, B. (2012). Molecular Imprinted Polymer as Sorbent in Micro-Solid Phase Extraction of Ochratoxin A in Coffee, Grape Juice and Urine. *Talanta*. 88, 129-135.
- Lin, D., Tu, Y., and Zhu, L. (2005). Concentrations and Health Risk of Polycyclic Aromatic Hydrocarbons in Tea. *Food Chem. Toxicol.* 43, 41-48.
- Lin, D., Zhu, L., He, W., and Tu, Y. (2006). Tea Plant Uptake and Translocation of PAHs from Water and Around Air. *J. Agric. Food Chem.* 54, 3658–3662.
- Liu, H.H., and Dasgupta, P.K. (1996). Analytical Chemistry in a Drop. Solvent Extraction in a Microdrop. *Anal. Chem.* 68(11), 1817-1821.
- Liu, J.F., Jiang, G.B., Chi, Y.G., Cai, Y.Q., Zhou, Q.X., and Hu, J.T. (2003). Use of Ionic Liquids for Liquid-Phase Microextraction of Polycyclic Aromatic Hydrocarbon. *Anal. Chem.* 75, 5870-5876.
- Liu, Y., Zhao, E., Zhu, W., Gao, H., and Zhou, Z. (2009). Determination of Four Heterocyclic Insecticides by Ionic Liquid Dispersive Liquid–Liquid Microextraction in Water Samples. J. Chromatogr. A. 1216(6), 885-891.
- Lund, M., Duedahl-Olesen, L., and Christensen, J.H. (2009). Extraction of Polycyclic Aromatic Hydrocarbons from Smoked Fish using Pressurized Liquid Extraction with Integrated Fat Removal. *Talanta*. 79, 10-15.
- Lutz, S., Feidt, C., Monteau, F., Rychen, G., Bizec, B.L., and Jurjanz, S.J. (2006). Effect of Exposure to Soil-Bound Polycyclic Aromatic Hydrocarbons on Milk Contaminations of Parent Compounds and Their Monohydroxylated Metabolites. *J. Agric. Food Chem.* 54, 263-268.
- MacLennan, P.A., Delzell, E., Sathiakumar, N., Myers, S.L., Cheng, H., Grizzle, W., Chen, V.W., and Wu, X.C. (2002). Cancer Incidence Among Triazine Herbicide Manufacturing Workers. J. Occup. Environ. Med. 44(11), 1048-1058.
- Madhavan, N.D., and Naidu, K.A. (1995). Polycyclic Aromatic Hydrocarbons in Placenta, Maternal blood, Umbilical Cord Blood and Milk Indian Woman. *Hum. Exposure Toxicol.* 14, 503-506.

- Majors, R.E. (2003). Trends in Sample Preparation. LC GC Europe. Retrieved December 2, 2012, from http://www.chromatographyonline.com/lcgc/data/ articlestandard/lcgceurope/052003/45011/article.pdf
- Meharg, A.A., Dyke, W.H., and Osborn, D. (1998). Polycyclic Aromatic Hydrocarbon (PAH) Dispersion and Deposition to Vegetation and Soil Following a Large Scale Chemical Fire. *Environ. Pollut.* 99, 29-36.
- Miller, D.J., and Hawthorne, S.B. (1998). Solubility of Polycyclic Aromatic Hydrocarbon in Subcritical Water from 298K to 498K. *J. Chem. Eng. Data.* 43, 1043-1047.
- Miloudi, H., Boos, A., Bouazza, D., Ali-Dahmane, T., Tayeb, A., Goetz-Grandmont, G., and Bengueddach, A. (2007). Acylisoxazolone-Impregnated Si-MCM-41
  Mesoporous Materials as Promising Liquid–Solid Extractants of Metals. *Mater. Res. Bull.* 42, 769-775.
- Moffat, C.F., and Whittle, K.J. (1999). Environmental Contaminants in Food : Polycyclic Aromatic Hydrocarbons, Petroleum and Other Hydrocarbon Contaminants. (1<sup>st</sup> ed.) England: CRC Press.
- Moradi, M., Yamini, Y., Vatanara, A., Saleh, A., Hojati, M., and Seidi, S. (2010). Monitoring of Trace Amounts of Some Anti-Fungal Drugs in Biological Fluids by Hollow Fiber Based Liquid Phase Microextraction followed by High Performance Liquid Chromatography. *Anal. Methods.* 2, 387-392.
- Moret, S., Grob, K., and Conte, L.S. (1996). On-Line High-Performance Liquid Chromatography-Solvent Evaporation-High-Performance Liquid Chromatography -Capillary Gas Chromatography-Flame Ionisation Detection for the Analysis of Mineral Oil Polyaromatic Hydrocarbons in Fatty Foods. J. Chromatogr. A. 750(1-2), 361-368.
- Nagaraju, D., and Huang, S.D. (2007). Determination of Triazine Herbicides in Aqueous Samples by Dispersive Liquid-Liquid Microextraction with Gas Chromatography-Ion trap Mass Spectrometry. J. Chromatogr. A. 1161, 89-97.
- Narayanan, J., Xiong, J.Y., and Liu, X.Y. (2006). Determination of Agarose Gel Pore Size: Absorbance Measurement Vis a Vis Other Techniques. J. Phys. : Conf. Ser. 28, 83-86.

- National Associations of Testing Authorities (NATA). (2012). *Technical Note 17 : Guidelines for the validation and verification of quantitative and qualitative test methods*. Australia : National Associations of Testing Authorities. Retrieved April 9, 2012, from http://www.nata.asn.au/phocadownload/publications/Technical\_publications/Technical\_note\_17.pdf
- Negrao, M.R., and Alpendurada, M.F. (1998). Solvent-Free Method for the Determination of Polynuclear Aromatic Hydrocarbons in Waste Water by Solid-Phase Microextraction-High-Performance Liquid Chromatography with Photodiode-Array Detection. J. Chromatogr. A. 823, 211-218.
- Neng, N.R., Mestre, A.S., Carvalho, A.P., and Nogueira, J.M.F. (2011a). Powdered Activated Carbons as Effective Phases for Bar Adsorptive Micro-Extraction (BaµE) to Monitor Levels of Triazine Herbicides in Environmental Water Matrices. *Talanta*. 83, 1643-1649.
- Neng, N.R., Mestre, A.S., Carvalho, A.P., and Nogueira, J.M.F. (2011b). Cork-Based Activated Carbons as Supported Adsorbent Materials for Trace Level Analysis of Ibuprofen and Clofibric Acid in Environmental and Biological Matrices. J. Chromatogr. A. 1218, 6263-6270.
- Nerin, C., Salafranca, J., Aznar, M., and Batlle, R. (2009). Critical Review on Recent Development in Solventless Techniques for Extraction of Analytes. *Anal. Bioanal. Chem.* 393, 809-833.
- Newman, A. (1995). More Bad News for Triazine Herbicides. *Environ. Sci. Technol.* 29(10), 450A-450A.
- Nii, S., Okumura, S., Kinoshita, T., Ishigaki, Y., Nakano, K., Yamaguchi, K., and Akita, S. (2010). Extractant-Impregnated Organogel for Capturing Heavy Metals from Aqueous Solutions. *Sep. Purif. Technol.* 73, 250-255.
- Ozcan, S., Tor, A., and Aydin, M.E. (2010). Determination of Polycyclic Aromatic Hydrocarbons in Waters by Ultrasound-Assisted Emulsification-Microextraction and Gas Chromatography-Mass Spectrometry. *Anal. Chim. Acta*. 665, 193-199.
- Pawliszyn, J. (1997). Solid Phase Microextraction : Theory and Practise. (1<sup>st</sup> ed.) New York : Wiley-VCH.
- Pedersen-Bjergaard, S., and Rasmussen, K.E. (1999). Liquid-Liquid-Liquid Microextraction for Sample Preparation of Biological Fluids Prior to Capillary Electrophoresis. *Anal. Chem.* 71, 2650-2656.

- Pedersen-Bjergaard, S., and Rasmussen, K.E. (2006). Electrokinetic Migration Across Artificial Liquid Membranes: New Concept for Rapid Sample Preparation of Biological Fluids. J. Chromatogr. A. 1109(2), 183-190.
- Pena-Pereira, F., Costas-Mora, I., Lavilla, I., and Bendicho, C. (2012). Rapid Screening of Polycyclic Aromatic Hydrocarbons (PAHs) in Waters by Directly Suspended Droplet Microextraction-Microvolume Fluorospectrometry. *Talanta*. 89, 217-222.
- Popp, P., Bauer, C., Hauser, B., Keil, P., and Wennrich, L. (2003). Extraction of Polycyclic Aromatic Hydrocarbons and Organochlorine Compounds from Water: A Comparison Between Solid-Phase Microextraction and Stir Bar Sorptive Extraction. J. Sep. Sci. 26, 961-967.
- Popp, P., Bauer, C., Moder, M., and Paschke, A. (2000). Determination of Polycyclic Aromatic Hydrocarbons in Waste Water by Off-Line Coupling of Solid-Phase Microextraction with Column Liquid Chromatography. J. Chromatogr. A. 897, 153-159.
- Popp, P., Bauer, C., and Wennrich, L. (2001). Application of Stir Bar Sorptive Extraction in Combination with Column Liquid Chromatography for the Determination of Polycyclic Aromatic Hydrocarbons in Water Samples. *Anal. Chim. Acta.* 436, 1-9.
- Porto, S. (2003). *Agargel*. Retrieved December 5, 2011, from http://www.agargel.com.br/agar-tec-en.html
- Pourreza, N., and. Ghanemi, K, J. (2009). Determination of Mercury in Water and Fish Samples by Cold Vapour Aomic Absorption Spectrometry after Solid Phase Extraction on Agar Modified with 2-Mercaptobenzimidazole. *Hazard. Mater.* 161, 982-987.
- Prieto, A., Telleria, O., Etxebarria, N., Fernandez, L.A., Usobiaga, A., and Zuloaga,
  O. (2008). Simultaneous Preconcentration of a Wide Variety of Organic
  Pollutants in Water Samples : Comparison of Stir Bar Sorptive Extraction and
  Membrane Assisted Solvent Extraction. J. Chromatogr. A. 1214, 1-10.
- Psillakis, E., and Kalogerakis, N. (2003). Developments in Liquid-Phase Microextraction. *Trends in Anal. Chem.* 22(9), 565-574.

- Pun ń Crespo, M.O., Cam, D., Gagni, S., Lombardi, N., and Lage Yusty, M.A. (2006). Extraction of Hydrocarbons from Seaweed Samples using Sonication and Microwave-Assisted Extraction: A Comparative Study. J. Chromatogr. Sci. 44(10), 615-618.
- Purcaro, G., Moret, S., and Conte, L.S. (2007). Rapid Validated Method for the Analysis of Benzo[a]pyrene in Vegetable Oils by Using Solid Phase Microextraction–Gas Chromatography-Mass Spectrometry. J. Chromatogr. A. 1176, 231-235.
- Qin, Z., Bragg, L., Ouyang, G., and Pawliszyn, J. (2008). Comparison of Thin-Film Microextraction and Stir Bar Sorptive Extraction for the Analysis of Polycyclic Aromatic Hydrocarbons in Aqueous Samples with Controlled Agitation Conditions. J. Chromatogr. A. 1196-1197, 89-95.
- Ratola, N., Alves, A., Kalogerakis, N., and Psillakis, E. (2008). Hollow-Fibre Liquid-Phase Microextraction: A Simple and Fast Cleanup Step used for PAHs Determination in Pine Needles. *Anal. Chim. Acta.* 618, 70-78.
- Ren, R., Wang, Y., Zhang, R., Gao, S., Zhang, H., and Yu, A. (2011). Solvent (Ionic Liquid) Impregnated Resin-Based Extraction Coupled with Dynamic Ultrasonic Desorption for Separation and Concentration of Four Herbicides in Environmental Water. *Talanta*. 83, 1392-1400.
- Rey-Salgueiro, L., Martinez-Carballo, E., Garcia-Falcon, M.S., Gonzalez-Barreiro, C., and Simal-Gandara, J. (2009). Occurrence of Polycyclic Aromatic Hydrocarbons and Their Hydroxlated Metabolites in Infant Foods. *Food Chem.* 115, 814-819.
- Rezaee, M., Assadi, Y., Milani Hosseini, M.R., Aghaee, E., Ahmadi, F., and Berijani,
  S. (2006). Determination of Organic Compounds in Water using Dispersive
  Liquid–Liquid Microextraction. J. Chromatogr. A. 1116, 1-9.
- Rianawati, E., and Balasubramanian, R. (2009). Optimization and Validation of Solid Phase Micro-Extraction (SPME) Method for Analysis of Polycyclic Aromatic Hydrocarbons in Rainwater and Stormwater. *Phys. Chem. Earth.* 34, 857-865.
- Risticevic, S., Niri, V.H., Vuckovic, D., and Pawliszyn, J. (2009). Recent Developments in Solid-Phase Microextraction. *Anal. Bioanal. Chem.* 393, 781-795.

- Rodrigues Chaves, A., Chiericato Jr., G., and Costa Queiroz, M.E. (2009). Solid-Phase Microextraction using Poly(pyrrole) Film and Liquid Chromatography with UV Detection for Analysis of Antidepressants in Plasma Samples. *J. Chromatogr. B.* 877, 587-593.
- Ruiz, M.O., Cabezas, J.L., Escudero, I., Alvarez, J.R., and Coca, J. (2002). α Phenylglycine Extraction with Trialkylmethylammonium Chloride Free and
   Immobilized in a Macroporous Resin: 1. Equilibria. *Trans IChemE*. 80, 529-536.
- Saaid, M., Saad, B., Mohamed Ali, A.S., Saleh, M.I., Basheer, C., and Lee, H.K. (2009). In Situ Derivatization Hollow Fibre Liquid-Phase Microextraction for the Determination of Biogenic Amines in Food Samples. J. Chromatogr. A. 1216, 5165-5170.
- Sambrook, J., and Russell, D.W. (2001). *Molecular Cloning*. (3<sup>rd</sup> ed.) New York: Cold Spring Harbor.
- Sanagi, M.M., Hassan Abbas, H., Wan Ibrahim, W.A., and Aboul-Enien, H.Y. (2012a). Dispersive Liquid-Liquid Microextraction Method Based on Solidification of Floating Organic Droplet for the Determination of Triazine Herbicides in Water and Sugarcane Samples. *Food Chem.* 133, 557-562.
- Sanagi, M.M., Loh, S.H., Wan Ibrahim, W.A., and Hasan, M.N. (2012b). Agarose Film Liquid Phase Microextraction Combined with Gas Chromatography–Mass Spectrometry for the Determination of Polycyclic Aromatic Hydrocarbons in Water. J. Chromatogr. A. 1262, 43-48.
- Sanagi, M.M., See, H.H., Wan Ibrahim, W.A., and Abu Naim, A. (2007). Determination of Pesticides in Water by Cone-Shaped Membrane Protected Liquid Phase Microextraction prior to Micro-Liquid Chromatography. J. Chromatogr. A. 1152, 215-219.
- Sanghi, R., and Srivastava, M.M. (2003). *Green Chemistry: Environment Friendly Alternatives*. (1<sup>st</sup> ed.) Pangbourne England: Alpha Science International Ltd.
- See, H.H., and Hauser, P.C. (2011). Electric Field-Driven Extraction of Lipophilic Anions Across a Carrier-Mediated Polymer Inclusion Membrane. *Anal. Chem.* 83, 7507-7513.
- See, H.H., Sanagi, M.M., Wan Ibrahim, W.A., and Naim, A.A. (2010). Determination of Triazine Herbicides using Membrane Protected Carbon Nanotubes Solid Phase Membrane Tip Extraction prior to Micro-Liquid Chromatography. J. Chromatogr. A. 1217, 1767-1772.

- Shariati-Feizabadi, S., Yamini, Y., and Bahramifar, N. (2003). Headspace Solvent Microextraction and Gas Chromatographic Determination of Some Polycyclic Aromatic Hydrocarbons in Water Samples. *Anal. Chim. Acta*. 489, 21-31.
- Shen, G., and Lee, H.K. (2002). Hollow Fiber-Protected Liquid-Phase Microextraction of Triazine Herbicides. *Anal. Chem.* 74, 648-654.
- Shen, G., and Lee, H.K. (2003). Headspace Liquid-Phase Microextraction of Chlorobenzenes in Soil with Gas Chromatography-Electron Capture Detection. *Anal. Chem.* 75, 98-103.
- Silva, A.R.M., Portugal, F.C.M., and Nogueira, J.M.F. (2008). Advances in Stir Bar Sorptive Extraction for the Determination of Acidic Pharmaceuticals in Environmental Water Matrices – Comparison between Polyurethane and Polydimethylsiloxane Polymeric Phases. J. Chromatogr. A. 1209, 10-16.
- Simonich, S.T., and Hites, R.A. (1995). Organic Pollutant Accumulation in Vegetation. *Environ. Sci. Technol.* 29, 2905-2913.
- Smith, R.M. (2008). Superheated Water Chromatography A Green Technology for the Future. J. Chromatogr. A. 1184 (1-2), 441-455.
- UK Legislation. (1989). The Water Supply (Water Quality) Regulations 1989. UK Statutory Instrument No. 1147. Retrieved December 20, 2012, from http://www.legislation.gov.uk/1989/1147
- Strikovsky, A.G., Jerabek K., Cortina, J.L., Sastre, A.M., and Warshawsky, A. (1996). Solvent Impregnated Resin (SIR) Containing Dialkyl Dithiophosphoric Acid on Amberlite XAD-2: Extraction of Copper and Comparison to the Liquid-Liquid Extraction. *React. Funct. Polym.* 28, 149-158.
- Tan, F., Zhao, H., Li, X., Quan, X., Chen, J., Xiang, X., and Zhang, X. (2009). Preparation and Evaluation of Molecularly Imprinted Solid-Phase Microextraction Fibers for Selective Extraction of Bisphenol A in Complex Samples. J. Chromatogr. A. 1216, 5647-5654.
- Teo, C.C., Tan, S.N., Yong, J.W.H., Hew, C.S., and Ong, E.S. (2008). Evaluation of the Extraction Efficiency of Thermally Labile Bioactive Compounds in Gastrodia Elata Blume by Pressurized Hot Water Extraction and Microwave Assisted Extraction. J. Chromatogr. A. 1182(1), 34-40.

- Teresa Pena, M., Carmen Casais, M., Carmen Mejuto, M., and Cela, R. (2009). Development of an Ionic Liquid Based Dispersive Liquid–Liquid Microextraction Method for the Analysis of Polycyclic Aromatic Hydrocarbons in Water Sample. J. Chromatogr. A. 1216, 6356-6364.
- Titato, G.M., and Lancas, F.M. (2005). Comparison between Different Extraction (LLE & SPE) and Determination (HPLC & Capillary-LC) Techniques in the Analysis of Selected PAHs in Water Samples. J. Liq. Chromatogr. Rel. Technol. 28, 3045-3056.
- Titato, G.M., and Lancas, F.M. (2006). Optimization and Validation of HPLC-UV-DAD and HPLC-APCI-MS Methodologies for the Determination of Selected PAHs in Water Samples. *J. Chromatogr. Sci.* 44, 35-40.
- Traving, M., and Bart, H.J. (2002). Recovery of Organic Acids using Ion-Exchanger-Impregnated Resins. *Chem. Eng. Technol.* 25, 997-1003.
- Tuteja, G., Rout, C., and Bishnoi, N.R. (2011). Quantification of Polycyclic Aromatic Hydrocarbons in Leafy and Underground Vegetables: A Case Study Around Panipat City, Haryana, India. J. Environ. Sci. Technol. 4(6), 611-620.
- Villaescusa, I., Salvado, V., and . de Pablo, J. (1996). Liquid-Liquid and Solid-Liquid Extraction of Gold by Triotylmethylammonium Chloride (TOMACI) Dissolved in Toluene and Impregnated on Amberlite XAD-2 Resin. *Hydrometallurgy*. 41, 303-311.
- Vinas, P., Campillo, N., Aguinaga, N., Perez-Canovas, E., and Hernandez-Cordoba, M. (2007). Use of Headspace Solid-phase Microextraction Coupled to Liquid Chromatography for the Analysis of Polycyclc Aromatic Hydrocarbons in Tea Infusions. J. Chromatogr. A. 1164, 10-17.
- Vreuls, J.J., Jong, G.J.D., and Brinkman, U.A.T. (1991). On-Line Coupling of Liquid Chromatography, Capillary Gas Chromatography and Mass Spectrometry for the Determination and Identification of Polycyclic Aromatic Hydrocarbons in Vegetables Oils. *Chromatographia*. 31(3-4), 113-118.
- Wang, G.Y., Chen, J., and Shi, Y.P. (2010). Hollow Fiber Liquid Phase Microextraction Combined with High Performance Liquid Chromatography for Analysis of Melamine in Milk Products. *Acta Chromatographica*. 22(2), 307-321.
- Wang, X., and Mitra, S. (2006). Enhancing Micro-Scale Membrane Extraction by Implementing a Barrier Film. J. Chromatgr. A. 1122, 1-6.

- Wei, M.C., and Jen, J.F. (2007). Determination of Polycyclic Aromatic Hydrocarbons in Aqueous Samples by Microwave Assisted Headspace Solid-Phase Microextraction and Gas Chromatography/Flame Ionization Detection. *Talanta*. 72, 1269-1274.
- Wei, F., Zhang, F.F., Liao, H., Dong, X.Y., Li, Y.H., and Chen, H. (2011). Preparation of Novel Polydimethylsiloxane Solid-Phase Microextraction Film and Its Application in Liquid Sample Pretreatment. J. Sep. Sci. 34, 331-339.
- World Health Organization. (2003a). Atrazine in Drinking-Water. Background Document for Preparation of WHO Guidelines for Drinking-Water Quality (WHO/SDE/WSH/03.04/32). Geneva: World Health Organization.
- World Health Organization. (2003b). Polynuclear Aromatic Hydrocarbon in Drinking-Water. Background Document for Preparation of WHO Guidelines for Drinking-Water Quality (WHO/SDE/WSH/03.04/59). Geneva: World Health Organization.
- Wu, C., Liu, Y., Wu, Q., Wang, C., and Wang, Z. (2011). Combined Use of Liquid-Liquid Microextraction and Carbon Nanotube Reinforced Hollow Fiber Microporous Membrane Solid-Phase Microextraction for the Determination of Triazine Herbicides in Water and Milk Samples by High-Performance Liquid Chromatography. *Food Anal. Methods.* 5(3), 540-550.
- Wu, Y., Xia, L., Chen, R., and Hu, B. (2008). Headspace Single Drop Microextraction Combined with HPLC for the Determination of Trace Polycyclic Aromatic Hydrocarbons in Environmental Samples. *Talanta*. 74, 470-477.
- Xu, H., Ding, Z., Lv, L., Song, D., and Feng, Y.Q. (2009). A novel Dispersive Liquid-Liquid Microextraction based on Solidification of Organic Droplet Method for Determination of Polycyclic Aromatic Hydrocarbons in Aqueous Samples. *Anal. Chim. Acta.* 636, 28-33.
- Xu, L., and Lee, H.K. (2009). Solvent-Bar Microextraction Using a Silica Monolith as the Extractant Phase Holder. *J. Chromatogr. A.* 1216, 5483-5488.
- Yang, K., and Xing, B. (2007). Desorption of Polycyclic Aromatic Hydrocarbons from Carbon Nanomaterials in Water. *Environ. Pollut.* 145, 529-537.
- Yang, K., and Xing, B. (2010). Adsorption of Organic Compounds by Carbon Nanomaterials in Aqueous Phase: Polanyi Theory and Its Application. *Chem. Rev.* 110, 5989-6008.

- Yang, Y., Belgazi, M., Lagadec, A., Miller, D.J., and Hawthorne, S.B. (1998). Elution of Organic Solutes from Different Polarity Sorbents using Subcritical Water. J.Chromatogr. A. 810, 149-159.
- Yao, C., Twu, P., and Anderson, J.L. (2010). Headspace Single Drop Microextraction using Micellar Ionic Liquid Extraction Solvents. *Chromatographia*. 72, 393-402.
- Yoshikawa, M., Masaki, K., and Ishikawa, M. (2002). Pervaporation Separation of Aqueous Organic Mixtures through Agarose Membranes. J. Membrane Sci. 205, 293-300.
- Yoshikawa, M., Yoshioka, T., Fujime, J., and Murakami, A. (2000). Pervaporation Separation of MeOH/MTBE through Agarose Membranes. J. Membrane Sci. 178, 75-78.
- Yudthavorasit, S., Chiaochan, C., and Leepipatpiboon, N. (2011). Simultaneous Determination of Multi-Class Antibiotic Residues in Water using Carrier-Mediated Hollow-Fiber Liquid-Phase Microextraction Coupled with Ultra-High Performance Liquid Chromatography Tandem Mass Spectrometry. *Microchim. Acta.* 172, 39-49.
- Zang, X.H., Wu, O.H., Zhang, M.Y., Xi, G.H., and Wang Z. (2009). Developments of Dispersive Liquid-Liquid Microextraction Technique. *Chinese J. Anal. Chem.* 37(2), 161-168.
- Zhang, H., and Andrews, A.R.J. (2000). Preliminary Studies of a Fast Screening Method for Polycyclic Aromatic Hydrocarbons in Soil by using Solvent Microextraction-Gas Chromatography. J. Environ. Monit. 2, 656-661.
- Zhao, J.J., Buldum, A., Han, J., and Lu, J.P. (2002). Gas Molecule Adsorption in Carbon Nanotubes and Nanotube Bundles. *Nanotechnology*. 13, 195-200.
- Zhao, L., and Lee, H.K. (2002). Liquid Phase Microextraction Combined with Hollow Fiber as a Sample Preparation Technique prior to Gas Chromatography/Mass Spectrometry. *Anal. Chem.* 74, 2486-2492.
- Zhou, J., Zhou, M., and Caruso, R.A. (2006a). Agarose Template for the Fabrication of Macroporous Metal Oxide Structures. *Langmuir*. 22, 3332-3336.

- Zhou, Q., Xiao, J., Wang, W., Liu, G., Shi, Q., and Wang, J. (2006b). Determination of Atrazine and Simazine in Environmental Water Samples using Multiwalled Carbon Nanotubes as the Adsorbents for Preconcentration prior to High Performance Liquid Chromatography with Diode Array Detector. *Talanta*. 68, 1309-1315.
- Zhu, L., Ee, K.H., Zhao, L., and Lee, H.K. (2002). Analysis of Phenoxy Herbicides in Bovine Milk by Means of Liquid-Liquid-Liquid Microextraction with a Hollow-Fiber Membrane. J. Chromatogr. A. 963, 335-343.
- Zuin, V.G., Montero, L., Bauer, C., and Popp, P. (2005). Stir Bar Sorptive Extraction and High-Performance Liquid Chromatohraphy-Fluorescence Detection for the Determination of Polycyclic Aromatic Hydrocarbon in Mate Teas. *J. Chromatogr. A.* 1091, 2-10.