

SUBCRITICAL WATER EXTRACTION OF 6-GINGEROL AND 6-SHOGAOL  
FROM *Zingiber Officinale*

MOHD SHARIZAN MD SARIP

UNIVERSITI TEKNOLOGI MALAYSIA

SUBCRITICAL WATER EXTRACTION OF 6-GINGEROL AND 6-SHOGAOL  
FROM *Zingiber Officinale*

MOHD SHARIZAN MD SARIP

A thesis submitted in fulfilment of the  
requirements for the award of the degree of  
Master of Engineering (Chemical)

Faculty of Chemical Engineering  
Universiti Teknologi Malaysia

DISEMBER 2012

To my mum, Ramlah Ibrahim, my dad Md Sarip Baba, my wife Nor Hasmi Abd  
Ghani and my family members.

With all love and gratitude

## ACKNOWLEDGEMENTS

‘In the name of Allah, the most gracious and the most merciful, selawat and salam to Prophet Muhammad s.a.w.’ I owe first and foremost my profound gratitude to almighty Allah s.w.t the source of all inspiration and help and without whose assistance; this study would not have come into existence.

Deep obligation and indebtedness and most sincere gratitude are offered to my supervisor Associate Professor Dr. Noor Azian Hj Morad and my co-supervisor Associate Professor Mustafa Kamal Abd Aziz for the continuous guidance during all stage of my research work and for this willingness to help. Without their continue support, interest, wisdom and ideas during our discussion, this thesis would not have been implemented and executed well.

I would like to acknowledge the Malaysian Ministry of Agriculture for the supporting and funding of this project on the ‘*Green Process for the Ginger Oil and Oleoresin Extraction*’ under eScience Fund and Malaysian Ministry of Higher Education for the financial support under My Brain 15.

I express my sincere thanks to Dr Norazah Mohamad Ali and Mr Radzi Ahmad from Natural Product Division, Forest Research Institute Malaysia (FRIM) for the use of analytical equipment and Prof Madya Dr Norashikin Saim for Universiti Teknologi Mara (UiTM) for the use of ASE 200.

I take pleasure in thanking my friends Mohd Shairazi, Huwaina, Syaripah Zaimah, Nor Baini Nabila, Nor Alia Anisa, Nik Fathurahman, Zuhaili, Nor Syuhana, Nurul Izzah, Manzuruddin, Syalwati, Tunku Hana, Bashirah and Maria deserve a special mention for being nice friends during all these years and for their support in completing my Master.

Finally, I wish to thank my mother, dad and other members of my family for their encouragement, support and love all through my ups and downs during my Master. This thesis would not have been possible without their support.

## ABSTRACT

Nowadays, natural products extract as a nutritional supplement becomes a part of healthy lifestyle. However, numerous scientific evidences suggested that the processing methods that mainly use organic solvents in extraction processes may result an undesired toxic residues in the product. Thus, the so called 'green' solvent that is water seems to be best alternative to substitute the organic solvent in the natural herb extraction process. In this study, the effect of subcritical water extraction (SWE) was employed for the extraction of bioactive compounds from *zingiber officinale* namely 6-gingerol, 6-shogaol and 10-gingerol. Two types of SWE equipment which are ASE 200 and CLEAR SWE prototype had been utilized. The ASE 200 with the capacity of 24 ml was used to evaluate the performance of the CLEAR SWE prototype with capacity of 1000 ml. Three parameters were manipulated in the SWE optimization process that are extraction temperature (100 to 200°C), static extraction time (10-60 minutes) and solvent to sample ratio (28/3-28/1 ml/g) at a fixed pressure of 3.5 MPa. The analysis was done using the High Performance Liquid Chromatography. Two main bioactive compounds namely 6-gingerol and 6-shogaol were extracted with the traces of 10-gingerol. The extraction and fractionation of 6-gingerol and 6-shogaol were obtained at the lower and higher temperature; respectively. The optimum conditions for the 6-gingerol was at the temperature of 130°C, in 30 minutes and solvent to sample ratio of 28/2 ml/g with the overall mass transfer coefficient of  $8.1179 \times 10^{-7}$  m/s. Meanwhile the optimum condition for the 6-shogaol was at the temperature of 170°C, in 20 minutes and solvent to ratio of 28/2 ml/g with the overall mass transfer coefficient of  $18.3764 \times 10^{-7}$  m/s. It is found that the ginger bioactive compounds will be started to degrade at a temperature above 180 °C.

## ABSTRAK

Kini, pengambilan ekstrak produk herba sebagai makanan tambahan adalah kebiasaan dalam amalan gaya hidup sihat. Namun, banyak bukti saintifik menunjukkan bahawa kaedah pemprosesan yang kebanyakannya menggunakan pelarut organik dalam proses pengekstrakan boleh mengakibatkan sisa toksik yang tidak diinginkan dalam produk tersebut. Oleh itu, pelarut teknologi hijau, iaitu air menjadi alternatif terbaik untuk menggantikan pelarut organik dalam proses pengekstrakan herba semula jadi. Dalam kajian ini, pengekstrakan sub lampau genting (SWE) telah dikaji untuk mengekstrak sebatian bioaktif, iaitu 6-gingerol, 6-shogaol dan 10-gingerol daripada *zingiber officinale*. Dua jenis peralatan SWE, iaitu ASE 200 dan prototaip CLEAR SWE digunakan. ASE 200 yang berkapasiti 24 ml digunakan untuk menilai prestasi prototaip CLEAR SWE yang berkapasiti 1000 ml. Tiga parameter telah dimanipulasi dalam proses pengoptimuman SWE iaitu suhu (100 hingga 200°C), masa (10-60 minit) dan nisbah pelarut kepada sampel (28/3-28/1 ml / g) pada tekanan tetap iaitu 3.5 MPa. Hasil ekstrak dianalisa dengan menggunakan Kromatografi Cecair Prestasi Tinggi. Dua sebatian bioaktif utama iaitu 6-gingerol dan 6-shogaol dan hanya sedikit 10-gingerol dapat diekstrak. Dengan menggunakan proses SWE, pengekstrakan dan pemisahan 6-gingerol dan 6-shogaol dapat dicapai pada suhu yang berbeza. Keadaan optima untuk pengekstrakan 6-gingerol adalah pada suhu 130°C, dalam masa 30 minit dan nisbah pelarut kepada sampel adalah 28/2 ml/g dengan nilai pekali pemindahan jisim keseluruhan, iaitu  $8.1179 \times 10^{-7}$  m/s manakala keadaan optimum untuk 6-shogaol pula adalah pada suhu 170°C, dalam masa 20 minit dan nisbah pelarut kepada sampel adalah 28/2 ml/g dengan nilai pekali pemindahan jisim keseluruhan iaitu  $18.3764 \times 10^{-7}$  m/s. Sebatian bioaktif halia mula merosot pada suhu melebihi 180°C.

## TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	<b>DECLARATION</b>	ii
	<b>DEDICATION</b>	iii
	<b>ACKNOWLEDGEMENTS</b>	iv
	<b>ABSTRACT</b>	vi
	<b>ABSTRAK</b>	vii
	<b>TABLE OF CONTENTS</b>	viii
	<b>LIST OF TABLE</b>	xii
	<b>LIST OF FIGURES</b>	xiv
	<b>LIST OF ABBREVIATIONS</b>	xviii
	<b>LIST OF SYMBOLS</b>	xix
	<b>LIST OF APPENDICES</b>	xxi
<b>1</b>	<b>INTRODUCTION</b>	1
	1.1 Introduction	1
	1.2 Problem Statement	4
	1.3 Objective of the Study	6
	1.4 Scope of the Study	7
	1.5 Thesis Summary	8
	1.6 Significance Contribution	8
<b>2</b>	<b>LITERATURE REVIEW</b>	10
	2.1 Introduction	10
	2.2 Ginger	12



2.2.1	Ginger Constituents	
2.2.2	Ginger Application	14
2.3	Methods of Natural Product Extraction	16
2.3.1	Soxhlet Extraction	17
2.3.2	Hydro Distillation	18
2.3.3	Supercritical Fluid Extraction	19
2.3.4	Subcritical Water Extraction	22
2.3.4.1	Subcritical Water Extraction Application	25
2.3.4.2	Subcritical Water Extraction Instrumentation	28
2.3.5	Extraction Mechanism	31
2.4	Leveraging on Water Properties as a ‘Green’ Solvent	36
2.4.1	Molecular Structure of Water	37
2.4.2	Hydrogen Bonds	39
2.4.3	Physicochemical Properties of Water	40
2.4.3.1	Dielectric Constant of Water	41
2.4.3.2	Viscosity of Water	43
<b>3</b>	<b>METHODOLOGY</b>	46
3.1	Introduction	46
3.2	Material and Chemical.	46
3.3	Overall Methodology	47
3.4	Sample Preparation and Pre-treatment Process	50
3.4.1	Sample Size Optimization	51
3.4.2	Purification Process	52
3.4.3	Yield Calculation	54
3.5	Subcritical Water Extraction (SWE)	54
3.5.1	ASE 200	55
3.5.1.1	Equipment Description	55
3.5.1.2	Experimental Procedure	57
3.5.2	CLEAR SWE Prototype	58

	3.5.2.1 Equipment Description	58
	3.5.2.2 Experimental Procedure	60
	3.6 Analytical Procedure	62
	3.6.1 Standard Calibration Curve	
	Development	63
	3.6.1.1 Calibrated Solution Preparation	65
	3.6.2.2 Data Analysis and Linear	
	Regression Model for Standard	
	Calibration Curve	68
	3.7.2 SWE Analysis of SWE Extracts	70
	3.7 Mass Transfer Model	70
<b>4</b>	<b>RESULT AND DISCUSSION</b>	76
	4.1 Introduction	76
	4.2 Sample Size Optimization	77
	4.3 Quantitative Analytical Method	78
	4.3.1 Limit of Detection, LOD	82
	4.4 Ethanol Extraction of Ginger Oleoresin	83
	4.5 Subcritical Water Extraction	85
	4.5.1 Effect of Temperature to the Compounds	
	Concentration	85
	4.5.2 Effect of the Extraction Time on	
	Compound Concentration	92
	4.5.2.1 Effect of Extraction Time at a	
	Constants Temperature of 130°C	92
	4.5.2.2 Effect of Extraction Time at a	
	Constants Temperature of 170°C	94
	4.5.3 Effect of the Solvent to Sample Ratio	95
	4.5.4 Optimum Operating Condition for SWE	
	Process	96
	4.6 SWE Mechanism using Mass Transfer	
	Coefficient Model.	97
	4.6.1 Effect of Time to the Cumulative 6-	

	gingerol Concentration	99
	4.6.2 Effect of Time to the Cumulative 6- shagaol Concentration	100
	4.6.3 Mass Transfer Model	102
	4.7 The Comparison between SWE with Ethanol Extraction	105
	4.8 Ensuring Thermodynamics of CLEAR SWE Prototype	106
	4.9 Economical Study	109
<b>5</b>	<b>CONCLUSION AND RECOMMENDATIONS</b>	114
	5.1 Conclusion	114
	5.2 Recommendation	115
	5.3 List of Publications and Awards	116
	<b>REFERENCES</b>	118
	Appendices A- E	130-142

## LIST OF TABLES

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Structure and physical properties of the selected ginger constituent	14
2.2	The advantages and disadvantages of soxhlet extraction	18
2.3	Operating condition for SWE process for extraction of natural product	26
2.4	Physicochemical properties of water at 25°C and 1 Atm	40
3.1	Material and chemical used in the study	47
3.2	Miscellaneous items utilized in this study	47
3.3	Parameter for the SWE process	55
3.4	The valve types and it functions in CLEAR SWE prototype	59
3.5	The calibration points design for calibration curve development used for each ginger standard; 6-gingerol, 6-shogaol and 10-gingerol	66
3.6	Comparison between the diffusion model and other rate constants	72
4.1	The calculation of LOD for 6-gingerol, 6-shagoal and 10-gingerol	83
4.2	The concentration calculation for 6-gingerol, 6-shogaol and 10-gingerol	84
4.3	The calculated constants value involve in k value calculation. The calculation for the parameter are given in Appendix D	102

4.4	The calculated equilibrium temperature in cooling vessel	108
4.5	Calculated mass in reactor one and two	110
4.6	The batch production cost	112

## LIST OF FIGURES

<b>FIGURE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
1.1	Water phase diagram: Temperature versus pressure	2
2.1	a) Ginger Plant b) Ginger Rhizome	11
2.2	Core areas of Sabah Development Corridor. (Source: Institute for development Studies Sabah)	12
2.3	The percentage constituents of the fresh ginger rhizome	13
2.4	Pressure-temperature phase diagram of Carbon Dioxide	20
2.5	Generic SFE equipment	21
2.6	The 15 liter Supercritical CO <sub>2</sub> pilot plant extractor in Institute of Bioproduct, UTM	22
2.7	The accelerated solvent extraction ASE 200 ( Dionex, USA)	23
2.8	The dielectric constant, $\epsilon$ of sub critical water and organic solvent with function of temperature	24
2.9	The generic SWE equipment	29
2.10	The typical offline SWE schematic diagram.	29
2.11	The Online SWE coupled with the HPLC schematic diagram	30
2.12	Subcritical water extraction pilot plant schematic diagram	31
2.13	Phase diagram of water	36

2.14	(a) Molecular structure of water. (b) Model of water molecule highlighting the bond angle and polar charges (c) Four point charge model of water	38
2.15	Hydrogen bonds among water molecules	39
2.16	The dielectric constants, $\epsilon$ of water as the function of pressure, P water at different temperature for water	42
2.17	The effect of temperature on water viscosity at different pressures for water. Data taken from IAPWS	43
3.1	Overall methodology	49
3.2	Flowchart for the sample preparation.	50
3.3	The experimental procedure for ethanol extraction of ginger	52
3.4	The rotary evaporator at the Centre of Lipid Engineering Applied Research (CLEAR), UTM	53
3.5	The purification process of ethanol and water based extracts using rotary evaporator.	53
3.6	a) The accelerated solvent extraction, ASE 200 in Universiti Teknologi Mara, UiTM b.) Extraction cell of ASE 200 (22 ml)	54
3.7	The schematic diagram of ASE 200	54
3.8	Experimental procedure for ASE 200	57
3.9	CLEAR SWE prototype in Centre of Lipid Engineering Applied Research (CLEAR), UTM	58
3.10	The schematic diagram of CLEAR SWE prototype	60
3.11	a) The dimension of the stainless steel mesh geometry b) The orientation of the stainless steel mesh in the extraction cell	61
3.12	Experimental Procedure for CLEAR SWE Prototype	62
3.13	The calibration curve development process	64
3.14	Dilution of ginger standards (6-gingerol, 6-shogaol and 10-gingerol) and storage procedure	65
3.15	Procedure for preparation and injection of the	

	establishment calibration curve for ginger standards; 6-gingerol, 6-shogaol and 10-gingerol	67
3.16	The experimental procedure for mass transfer coefficient determination	73
3.17	The experimental procedure for true density measurement.	74
4.1	Effect of mean particles size, MPS to the yield percentage, % (w/w)	77
4.2	The ginger bioactive standard chromatogram at different concentration	79
4.3	The standard calibration curve for ginger bioactive compound	80
4.4	HPLC profile for ethanol extraction	84
4.5	The effect of SWE temperature, °C to the ginger bioactive compounds concentration, µg/g using CLEAR SWE prototype at constants pressure (3.5MPa), time (30 minutes) and solvent to ratio (28/3 ml/g)	86
4.6	HPLC profile for SWE extract at temperature 130 oC and constants pressure (3.5 MPa), constants extraction time (30 minute) and constant solvent to sample ratio (28/3 ml/g)	87
4.7	HPLC profile for SWE extract at temperature 170 oC and constants pressure (3.5 MPa), constants extraction time (30 minute) and constant solvent to sample ratio (28/3 ml/g)	88
4.8	The dehydration of 6-gingerol to 6-shogaol stoichiometry	88
4.9	The carbon formation in SWE extract at 200°C, 3.5 MPa and 30 minute extraction time	89
4.10	The effect temperature on the overall yield percentage of ginger	90
4.11	The effect of SWE temperature, °C to the ginger bioactive compounds concentration, µg/g using ASE	



	200 at a constants pressure (3.5MPa), constant time (30) minutes and constant solvent to sample ratio (28/3)	91
4.12	The effect of the extraction time to the bioactive compound at temperature of 130°C, constant pressure of 3.5MPa and constant solvent to sample ratio of 28/3 ml/g	93
4.13	The effect of the extraction time to the bioactive compound at temperature of 170oC (3.5 MPa , 28/3 ml/g)	94
4.14	The effect of the solvent to sample ratio to the overall yield percentage and compound concentration at constants temperature of 130°C, constants pressure of 3.5 MPa and constants extraction of 30 minutes	95
4.15	The mechanism of ginger extraction process	98
4.16	The concentration of the 6-gingerol extracted with time at different temperature	100
4.17	The concentration of the 6-shogaol extracted per unit time at different temperatures	101
4.18	The relationship between $-\ln(c_1(\text{sat})-c_1)$ and extraction time, t for 6-gingerol extraction at different temperatures	103
4.19	The relationship between $-\ln(c_1(\text{sat})-c_1)$ and extraction time, t for 6-shogaol extraction at different temperature	103
4.20	The effect of the temperature to the overall mass transfer coefficient, k	104
4.21	The comparison between ethanol and SWE extraction for the 6-gingerol and 6-shogaol concentration	106
4.22	Stage 1: The extraction process Stage 2: Cooling process	107
4.23	The mass distribution in the process	109

**LIST OF ABBREVIATIONS**

ASE	-	Accelerated solvent extraction
DNA	-	Deoxyribonucleic acid
EPSM	-	Environmental protection society of malaysia
FOA	-	Food and agriculture organization of the united nations
HPLC	-	High performance liquid chromatography
H	-	Enthalpy
H- bonds	-	Hydrogen bond
IAPWS.	-	The international association for the properties of water and steam
MPS	-	Mean particles size
MOA	-	Malaysia ministry of agriculture
P-T diagram	-	Pressure-temperature diagram
P-v diagram	-	Pressure-volume diagram
P		Pressure
RT	-	Retention time
SWE	-	Subcritical water extraction
SFE	-	Supercritical fluid extraction
T-v diagram	-	Temperature-volume diagram
T <sub>cr</sub>		Critical temperature
T		Temperature
UTM		Universiti Teknologi Malaysia
UiTM		Universiti Teknologi MARA
U	-	Internal energy

**LIST OF SYMBOLS**

$A_T$	- Particle surface area
$c$	- Coefficient representing the intercept-y.
$C_1$	- Compound concentration in liquid
$C_s$	- Solid concentration.
$C_{1(sat)}$	- Initial concentration in the solid
$C_{1(sat)6\text{-gingerol}}$	- 6-gingerol initial concentration
$C_{1(sat)6\text{-shagaol}}$	- 6-shagaol initial concentration
$CO_2$	- Carbon dioxide
$C_p$	- Heat capacity
$D_p$	- Particle diameter
$D$	- Diameter of bed
$H_2O$	- Water
$h$	- Bed height
$k$	- Overall mass transfer coefficient
$k_i$	- Intra-particle diffusion coefficient
$k_e$	- External mass transfer coefficient
$m_s$	- Total mass of solid
$m$	- Coefficient representing the slope
$O$	- Oxygen
$\rho$	- Particle density
$\rho_e$	- Density of ethanol
RSD %	- Relative standard deviation
$r^2$	- Coefficient of determination
$r$	- Correlation coefficient

$S_{\text{upper}}$	- Upper sieve size
$S_{\text{lower}}$	- Lower sieve size
$S_{\text{bl}}$	- The standard deviation of the blank measurement
$\mu$	- Mean
$V_L$	- Volume of the solution or bed
$W$	- Weight of mixture of ethanol and ginger particles
$W_f$	- Weight of the round bottom flask after purification process, g.
$W_b$	- Weight of the round bottom flask before purification process, g.
$W_s$	- Weight of the sample matrix, g.
$W_i$	- Initial weight of volumetric flask
$x_1$	- The lowest measurement
$x_{\text{bl}}$	- Mean of the blank measurement
$Y_{\%}$	- Overall yield percentage, % (w/w)
$\epsilon$	- Dielectric constant
$\Phi_s$	- Particles sphericity
$\sigma$	- Standards deviation
$\sigma$	- Surface tension

## LIST OF APPENDICES

<b>APPENDIX</b>	<b>TITLE</b>	<b>PAGE</b>
A	Calculation for the concentration of SWE ginger extract	129
B	Calculation of LOD	132
C	Calculation for constants in overall mass transfer coefficient measurement	133
D	Significance test using standard normal distribution	134
E-1	Journal Publication- Md Sarip, M.S and Morad, N.A.2012.Determination of Overall Mass Transfer Coefficient for 6-Gingerol and 6-Shagoal in Subcritical Water Extraction. Advanced Materials Research.550-553:1900-1903	136
E-2	Oral presentation - Mohd Sharizan Md Sarip, Noor Azian Morad, Nor Azah Mohd Ali, Mohd Radzi Ahmad and Mohd Shairazi Romainor. Subcritical Water Extraction of the Zingiber officinale: The Quantitative Analysis Method using High Performance Liquid Chromatography. Presented at 24th Regional Symposium of Malaysia Analytical Sciences (SKAM 24), 21st – 23rd November 2011, One Hotel Helang, Langkawi, Kedah	141

- E-3                      Poster Presentation - Md Sarip, M.S, Morad, N.A,  
Abd Aziz, M.K, Mohd Ali,N.A, Mohd Azizi,  
C.Y,Dhilon, A. and Romainor,M.S. Ginger  
oleoresin extraction: Effect of the squeezing  
process and the mean particles size to the oleoresin  
yield percentage, % (w/w). Poster Presented at  
UMS Biotechnology Symposium IV, 1-3 December  
2010, Universiti Malaysia Sabah

## CHAPTER 1

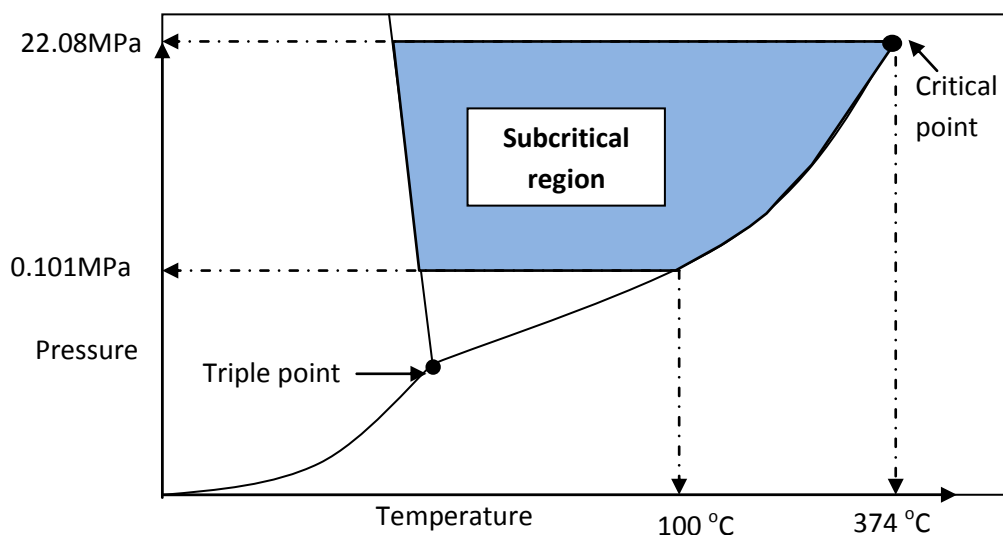
### INTRODUCTION

#### 1.1 Introduction

Water is known as a universal solvent and it is cheap, abundant, highly pure, non-toxic and easily handled. Normally, water was regarded as the poor solvent for the most organic compounds because of its highly polar or high dielectric constant (Smith, 2006). These make water as an ineffective solvent in an extraction process. Water utilization in the extraction process only can be effective at high temperature because the dielectric constants of water would decrease as the temperature increases and make water mimic the organic solvent dielectric constant. By that, the solubility of organic compounds in water is also increased which consequently increase the extraction efficiency.

Furthermore, other physicochemical properties of water such as viscosity and surface tension also decrease with the temperature increased and enhanced the extraction efficiency through the promoting mass transfer properties and matrix particle penetration (Wiboonsirikul & Adachi, 2008). However, the extraction above the critical point is not favourable because it can promote degradation of the extract (Eikani *et al.*, 2006).

Smith (2006) stated that the interest region in the utilization of water in the separation technology is between the boiling point (100°C) and critical point (374°C) (Smith, 2006). In this region, water already vaporized and modest pressure was needed to force it in the liquid phase as referred to the water phase diagram as shown in Figure 1.1 to avoid the possible degradation of the interest organic compound. The pressure is required to ensure the water in the liquid phase depends on the operating temperature. This region is described and named differently by the researchers such as subcritical water (Amashukeli *et al.*, 2007; Ayala & Castro, 2001; Budrat & Shotipruk, 2008; Ghoreishi *et al.*, 2008), superheated water (Ammann *et al.*, 1999; Eikani *et al.*, 2007; Smith, 2006), pressurized water (Cacace & Mazza, 2006; Markom *et al.*, 2010), pressurized hot water (Teo *et al.*, 2010), pressurized low polarity water (Cacace & Mazza, 2006) and compressed hot water (Liu & Wyman, 2005). In this study, water in this state is referred as subcritical water.



**Figure 1.1:** Water phase diagram: Temperature versus pressure

Since 1980s, the subcritical water extraction (SWE) have been applied in the numerous organic compound extraction such as Gallic acid (Markom *et al.*, 2010), polyphenolic compound (Antuono, 2009; Kim *et al.*, 2010; Turner *et al.*, 2006), monoterpenoid phenol (Ozel *et al.*, 2002), ellagic acid (Rangsrivong *et al.*, 2008), asiatic acid (Kim *et al.*, 2009) and magiferin (Kim, *et al.*, 2010). In 1994, Hawthorne



demonstrated that water can be used to extract polar, moderately polar and non-polar compound by varying the temperature from 50°C to 400°C (Hawthorne *et al.*, 1994). Hawthorne also reported on his extended work that SWE can be utilized to extract largely polar compound at the lower temperature and less polar compound at the higher temperature (Hawthorne *et al.*, 2000). This work gives the idea of the selective extraction and will be the most important features of SWE. Besides that, SWE implementation when compared with the traditional method provides a lot advantages such as low extraction time, high quality extract, inexpensive solvent and environmentally benign (Basile *et al.*, 1998; Herrero *et al.*, 2006; Saim & Osman, 2008). In order to evaluate the efficiency of the SWE process, a well-known natural herb, common ginger is used in this study.

Ginger is an everyday term for members of the *Zingiberaceae* family (Grieve, 1992). Most *Zingiberaceae* family is native to tropical and subtropical locations. Ginger is classified in the division Magnoliophyta, class Liliopsida, and order Zingiberales, family Zingiberaceae. There are 18 known genera with more than 160 species of *Zingiberaceae* in Peninsula Malaysia. Among these myriad of species, the common ginger is known as *Zingiber officinale*. Ginger originates from Southern China and widely believed to possess many medicinal benefits in the ancient Orient, and Indian Peninsula. The name “ginger” itself is derived from the Gingi district in India, where the upsets stomach is treated with ginger tea. Nowadays, ginger rhizomes are actively cultivated in the warmer parts of the globe such as India, the Middle East, and the Far East. The useful part of ginger was its tuberous rhizome that yields oleoresin or oil depends on the type of extraction. Ginger oil normally used in the fragrances and beverage industry for its unique flavour properties. Meanwhile, ginger oleoresins are used in the health food and pharmaceutical industry for its healing properties. Ginger oleoresin composed of non-volatiles pungent and non-pungent constituent including the bioactive compounds such as 6-gingerol, 6-shogaol, 8-gingerol and 10-gingerol. Other compounds such as 6-paradol, zingerone, zingiberene, gingerdiols and gingerdiones were the minor components. As the well-known herb, ginger oleoresin are scientifically proven for its anti-cancer (Abdullah *et al.*, 2010; Harliansyah *et al.*, 2007), anti-oxidants (Harliansyah *et al.*, 2007) and anti-tumour properties (Vimala *et al.*, 1999). This

pharmacological study is not limited on the ginger oleoresin but selectively tested on the individual compound such as 6-gingerol, 6-shogaol and 10-gingerol. 6-gingerol is proven to medicated cardiac contractile (Antipenko *et al.*, 1999) and gives the positive response on the anti-oxidant, anti-proliverative and apoptosis properties (Harliansyah *et al.*, 2007). Meanwhile, 6-shogaol is proven to medicate the spinal cord injury (Kyung *et al.*, 2006) and lessens the human oral cancer (Chen *et al.*, 2010). 10-gingerol, another series of gingerol is proven to give the positive response on the anti-bacterial (Park *et al.*, 2008) and anti-microbial activities (Nagoshi *et al.*, 2006).

## 1.2 Problem Statement

There are several methods that can be used in the natural herb extraction process such as solvent extraction, hydro distillation, supercritical fluid extraction (SFE), cold pressing and vapour cracking. Among that, the most common ones are solvent extraction and hydro distillation due to their simplicity and economical characteristics. Solvent extraction refers to separation method using organic solvents such as hexane, ethanol, iso-propanol, acetone or methanol to extract oleoresin from plants. Nevertheless, due to demands on 'green' technology, this method is not preferred as the use of organic solvents may lead to the formation of toxic residue in the extract which will eventually be marketed for consumption by consumers (Basile *et al.*, 1998). The purification or solvent removal process may not promise the total toxic elimination. Complete removal of the solvent residues may be possible, if the high sensitive equipment is implemented but it would increase the production cost. Furthermore, the tedious handling procedure may cause the pollution in the workplace and effect to the worker in a long term effect. The current climate of environmental awareness has given rise to various organizations such as Greenpeace, Environmental Protection Society of Malaysia (EPSM), and Environmental Protection Agencies to help educate the public to matters that can cause harm to the

environment. Thus, with the availability of various knowledge sources, people are also, now more than ever, aware about the environmental impacts of non-eco-friendly processes. Other than that, the solvent extraction require a long extraction time from 5 to 10 hours which consumed a lot of energy.

Another extractions process known as supercritical fluid extraction (SFE) was a new 'green' extraction process with the unique features. The process involve the utilization of the common gases normally CO<sub>2</sub> at its supercritical phase. It was recognized as the most eco-friendly extraction method. However, the polarity or dielectric constants of supercritical CO<sub>2</sub> are too low to obtain the efficient extraction (Hawthorne *et al.*, 1994; Herrero *et al.*, 2009). Thus, it is only efficient for the non-polar organic compound (Hawthorne *et al.*, 2000). In fact, many researchers have tried to increase the polarity by adding the organic modifier and this process cannot be considered as completely 'green' anymore. Furthermore, the need of dried sample in the SFE process to avoid the reaction between carbon dioxide and water will increased the pre-treatment cost. Besides that, the possible cuticular wax and lipid extraction in this process make it not preferable for natural herb extraction (Castro *et al.*, 1999). In addition, the extremely high pressure process requirement for the SFE equipment from its critical pressure (7.38 MPa) up to 20 MPa and if carbon dioxide need to be recycle the high end compressor system is required, adds up to expensive operating costs (Castro *et al.*, 1999; Smith, 2002). Indeed, the involvement of high pressure process also needs additional safety consideration and experienced operator.

Therefore, with the general consensus of water as a "green" solvent, it seems to be the best available solution to reduce health and environmental problems in industries. The conventional water extraction or so called hydro distillation is not suitable and uneconomic due to the long extraction time. The energy cost of hydro distillation was estimated to be 20 times more compared with the SWE process (Wiboonsirikul & Adachi, 2008). For example, the extraction of marjoram leaves using SWE for 15 minutes yield five times more oil compare with the three hours of hydro distillation process (Jimenez-Carmona *et al.*, 1999). This would lead to the

waste of energy and contribute to further global warming. So, this SWE process is seen to be an alternative way for the herbal product extraction which offers the 'green' and economic process. Furthermore, the possibilities for the selective extraction using SWE may contribute to local herb added value not only for the ginger (Galkin & Lunin, 2005; Hawthorne, *et al.*, 1994). Besides that, the different benefit and medicinal properties of ginger bioactive compounds namely 6-gingerol, 6-shogaol and 10-gingerol creating the increase on the ginger selective extraction. Thus, the aim of this study is to investigate the feasibility of the subcritical water extraction to extract ginger bioactive compounds namely 6-gingerol, 6-shogaol and 10-gingerol. Various operating conditions will be experimented to find an optimum operating condition. The study will also explore on the selective extraction on that bioactive compounds using SWE. Furthermore, two type of SWE equipment will utilized in this research that is CLEAR SWE prototype and accelerated solvent extractor, ASE 200.

### **1.3 Objective of the Study**

- A. To study the effect of temperature, time and ratio of solvent to ginger bioactive compounds (6-gingerol, 6-shogaol and 10-gingerol) using CLEAR SWE prototype.
- B. To evaluate the competence of CLEAR SWE prototype by comparing with ASE 200 equipment.
- C. To explain the SWE mechanism using overall mass transfer coefficient model for 6-gingerol and 6-shogaol at different temperature.

## 1.4 Scope of the Study

In order to achieve the objectives of the study, the following tasks will be carried out:

- A. The sample pre-treatment process for the mean particles size optimization using ethanol extraction was done prior to the SWE process. There are three mean particles sizes (MPS) used in this process that are 0.75 mm, 1.5 mm and 2.68 mm. The optimum MPS will be standardized throughout SWE study.
- B. The establishment of quantitative analysis method for ginger bioactive compounds (6-gingerol, 6-shogaol and 10-gingerol) using High Performance Liquid Chromatography, HPLC (Water, USA) for water based ginger extract.
- C. The optimization of SWE process using CLEAR SWE prototype for the extraction of ginger bioactive compounds that is 6-gingerol, 6-shogaol and 10-gingerol. The parameter involve in this optimization process was temperature, extraction times and solvent to sample ratio.
- D. The performance evaluation for CLEAR SWE prototype is done by the comparison with ASE 200 equipment data on the effect of temperature to the ginger bioactive compounds that is 6-gingerol, 6-shogaol and 10-gingerol. An ASE 200 was the commercial accelerated solvent extractor which commonly used by the researcher for SWE implementation. The temperature range is from 130°C to 200°C.
- E. The measurement of the overall mass transfer coefficient for 6-gingerol and 6-shagoal extraction using SWE process from the temperature of 110°C to 170 °C.

## **1.5 Thesis Summary**

This thesis is divided into five chapters. Chapter one is the introduction of the study which consists of research objective, scope of the study and the research problem. Chapter two gives the review on the raw material used, the extraction process for the natural herb product and the relationship between the physicochemical properties of water with extraction process. Chapter three describes the research methodology of the study. This chapter discuss the detail experimental procedures of pre-treatment process, the analytical method, SWE optimization and the modelling for the SWE process. The result finding is discussed in the chapter four. The study has been concluded in the chapter five together with the some recommendation on the future work.

## **1.6 Significance Contribution**

The significant contribution of this study can be divided into two aspects; scientific and industry. The scientific data provide in this research would enhance the understanding of the SWE application from the view of physicochemical properties of water specifically the dielectric constants. Furthermore, the non-existence of research on the SWE application to the ginger bioactive extraction would make this research finding useful to support the product development of this commodity. Not limited to that, the modelling on the two different compounds mechanism of extraction namely 6-gingerol and 6-shogaol in the SWE process would provide the useful insight on the extraction rate of the ginger bioactive compound extraction. The data can be utilized for the prediction of other local herbs in Malaysia and for the purpose of scaling up elsewhere.

Form the industrial point of view, this newly improved method of herbal extraction will reduce the usage of organic solvents and hence support the recent awareness to create 'greener' environment by the use of natural solvent. This process is also believed to increase the value of herbal products through selective extraction that would benefit the manufacture. Recommendation on optimum operational condition also will help the industries to scale up the process for commercialization. This technology is also predicted to be beneficial not only to industry based on ginger, but to other herbal industries, as it is proven to be effective for the extraction of essential oils from black pepper, cumin, wheat bran, turmeric and coriander seeds.

## REFERENCES

- Abdullah, S., Abidin, S. A. Z., Murad, N. A., Makpol, S., Ngah, W. Z. W., & Yusof, Y. A. M. (2010). Ginger extract (*Zingiber officinale*) triggers apoptosis and G0/G1 cells arrest in HCT 116 and HT 29 colon cancer cell lines. *African Journal of Biochemistry Research*, 4(4), 134-142.
- Ali, B. H., Blunden, G., Tanira, M. O., & Nemmar, A. (2008). Some phytochemical, pharmacological and toxicological properties of ginger (*Zingiber officinale* Roscoe): A review of recent research. *Food and Chemical Toxicology*, 46, 409-420.
- Alqasoumi, S. I. (2009). Quantification of 6-gingerol in *Zingiber officinale* extract, ginger-containing dietary supplements, teas and commercial creams by validated HPTLC densitometry. *Journal of Pharmaceutical Sciences*, 34, 33–42.
- Amashukeli, X., Pelletier, C. C., Kirby, J. P., & Grunthaner, F. J. (2007). Subcritical Water Extraction of Amino Acids from Atacama Desert Soils. *Journal of geophysical research* 112, 1 -10.
- Ammann, A., Hinz, D. C., Addleman, R. S., Wai, C. M., & Wenclawiak, B. W. (1999). Superheated Water Extraction, Steam Distillation and SFE of Peppermint Oil. *Fresenius Journal of Analytical Chemistr*, 364 650–653.
- Antipenko, A. Y., Spielman, A. I., & Kirchberger, M. A. (1999). Interactions of 6-Gingerol and Ellagic Acid with the Cardiac Sarcoplasmic Reticulum Ca<sup>2+</sup>-ATPase1. *The journal of pharmacology and experimental therapeutics*, 290, 227-234.



- Antuono, I. D. (2009). Use of sub-critical water for the extraction of natural antioxidants from byproducts and wastes of the food industry. *Proceeding of the 14th Workshop on the Developments in the Italian PhD Research on Food Science Technology and Biotechnology*. 16 – 18<sup>th</sup> September. University of Sassari Oristano, Italy.
- Anderson, G. S., & Miller, R. C. (2000). Static Dielectric Constants for Liquid Water from 300 K to 350 K at Pressures to 13 MPa Using a New Radio-Frequency Resonator. *Journal of Chemical Engineering Data*, 45, 549-554.
- Aneupankul, T., Goto, M., Sasaki, M., Pavasant, P., & Shotipruk, A. (2007). Extraction of Anti-cancer Damnacanthol from Roots of *Morinda Citrifolia* by Subcritical Water. *Separation and Purification Technology*, 55 343–349.
- Ayala, R. S., & Castro, M. D. L. d. (2001). Continuous subcritical water extraction as a useful tool for isolation of edible essential oils. *Food Chemistry*, 75(2001), 109-113.
- Basile, A., M.M., J.-C., & Clifford, A. A. (1998). Extraction of Rosemary by Superheated Water. *Journal of Agriculture and Food Chemistry*, 46, 5205-5209.
- Bart, H.-J. (2011). *Extraction of natural products from plants – An introduction*. In S. Pilz (Ed.), *Industrial scale natural products extraction* (First Edition ed.). Weinheim: Wiley-VCH Verlag GmbH & Co KGaA.
- Bartle, K. D., Clifford, A. A., Hawthorne, S. B., Langenfeld, J. J., Miller, D. J., & Robinson, R. (1990). A model for dynamic extraction using a supercritical fluid. *The Journal of Supercritical Fluids*, 3(3), 143-149.
- Bartle, K. D., Boddington, T., Clifford, A. A., & Hawthorne, S. B. (1992). The Effect of solubility on the kinetics of dynamic supercritical-fluid extraction. *The Journal of Supercritical Fluids*, 5(3), 207-212.
- Bai, Y.-X., & Li, Y.-F. (2006). Preparation and characterization of crosslinked porous cellulose beads. *Carbohydrate Polymers*, 64, 402–407.
- Balachandran, S., Kentish, S. E., & Mawsonb, R. (2006). The effects of both preparation method and season on the supercritical extraction of ginger. *Separation and Purification Technology*, 48, 94–105.
- Bhattacharai, S., Tran, V. H., & Duke, C. C. (2001). The Stability of Gingerol and Shogaol in Aqueous Solutions. *Journal of Pharmaceutical Sciences*, 90(10), 1658-1664.

- Bertolini, M., Cassettari, & Salvetti, G. (1982). The dielectric relaxation time of supercooled water. *Journal of Chemical Physics*, 76(6), 3285-3290.
- Brunner, G. (2005). Supercritical fluids: technology and application to food processing. *Journal of Food Engineering*, 67, 21-33.
- Budrat, P., & Shotipruk, A. (2008). Extraction of Phenolic Compound from Fruits of Bitter Melon (*Momordica Charantia*) with Subcritical Water Extraction and Antioxidant Activities of These Extracts. *Chiang Mai Journal of Science*, 35, 123-130.
- Budrat, P., & Shotipruk, A. (2009). Enhanced recovery of phenolic compounds from bitter melon (*Momordica charantia*) by subcritical water extraction. *Separation and Purification Technology* 66, 125–129.
- Cacace, J. E., & Mazza, G. (2006). Pressurized low polarity water extraction of lignans from whole flaxseed. *Journal of Food Engineering*, 77, 1087–1095
- Castro, M. D. L. d., Jimeñez-Carmona, M. M., & Fernàndez-Peèrez, V. (1999). Towards more rational techniques for the isolation of valuable essential oils from plants. *trends in analytical chemistry*, 18(11), 708-716.
- Cebovic, T., Spasic, S., & Popovic, M. (2008). Cytotoxic Effects of the *Viscum album L.* Extract on Ehrlich Tumour Cells In Vivo. *Phytotherapy Research*, 22, 1097–1103.
- Chen, C.-C., Rosen, R. T., & Ho, C.-T. (1986 ). Chromatographic Analyses of Isomeric Shogaol Compounds Derived from Isolated Gingerol Compound of Ginger ( *Zingiber Officinale Roscoe*) *Journal of Chromatography, A*, 360, 175-184.
- Chen, C.-Y., Yang, Y.-H., & Kuo, S.-Y. (2010). Effect of [6]-Shogaol on Cytosolic Ca<sup>2+</sup> Levels and Proliferation in Human Oral Cancer Cells (OC2). *Journal of Natural Products*, 73, 1370–1374.
- Chassagnez-Méndez, A. L., Corrêa, N. C. F., França, L. F., Machado, N. T., & Araújo, M. E. (2000). A mass transfer model applied to the supercritical extraction with CO<sub>2</sub> of curcumins from turmeric rhizomes (*Curcume Longa L.*). *Brazilian Journal of Chemical Engineering*, 17(3), 21.
- Cooper, J. R. (1994). *IAPWS Releas on Surface Tension of Ordinary Water Substance*. Palo Alto: International Association for the Properties of Water and Steam.

- Communities, E. (2002). *Performance Criteria, Other Requirement and Procedures for Analytical Methods* Brussel
- Cussler, E. L. (1984). *Diffusion: Mass transfer in fluid systems*. New York: Cambridge University Press.
- Deul, R. (1984). *Dielektrizitätskonstante und Dichte von Wasser-Benzol - Mischungen bis 400°C und 3000 bar*. Doctor Philosophy, Karlsruhe University, Karlsruhe.
- Dugasan, S., Pichika, M. R., Vishna Devi Nadarajah, Balijepalli, M. K., Tandra, S., & Korlakunta, J. N. (2010). Comparative antioxidant and anti-inflammatory effects of [6]-gingerol, [8]-gingerol, [10]-gingerol and [6]-shogaol. *Journal of Ethnopharmacology* 127, 515-520.
- Eikani, M. H., Golmohammad, F., Mirza, M., & Rowshanzamir, S. (2007). Extraction of Volatile Oil from Cumin (*Cuminum Cyminum* L.) with Superheated Water. *Journal of Food Process Engineering* 30 255-266.
- Eikani, M. H., Golmohammad, F., & Rowshanzamir, S. (2007). Subcritical water extraction of essential oils from coriander seeds (*Coriandrum sativum* L.). *Journal of Food Engineering*, 80, 735–740.
- EURACHEM. (1998). *The Fitness for Purpose of Analytical Methods, A Laboratory Guide to Method Validation and Related Topics*. Middlesex, UK.
- Florianio, W. B., & Nascimento, M. A. C. (2004). Dielectric constant and density of water as a function of pressure at constant temperature. *Brazilian Journal of Physics*, 34(1), 38-41.
- Galkin, A. A., & Lunin, V. V. (2005). Subcritical and Supercritical Water: A Universal Medium for Chemical Reactions. *Russian Chemical Reviews*, 74(1), 21-35.
- Geankoplis, C. J. (2003). *Transport processes and separation process principles (Includes unit operation)*. (Fourth ed). Upper Saddle River, New Jersey: Pearson Education, Inc.
- Ghoreishi, S. M., & Shahrestani, R. G. (2009). Subcritical water extraction of mannitol from olive leaves. *Journal of Food Engineering*, 93, 474–481.
- Goto, M., Roy, B. C., & Hirose, T. (1996). Shrinking-core leaching model for supercritical-fluid extraction. *The Journal of Supercritical Fluids*, 9(2), 128-133.

- Grieve, M., & Lyle, C. F. (1992). *A modern herbal: The medicinal, culinary, cosmetic and economic properties, cultivation and folklore of herbs, grasses, fungi, shrubs and trees with all their modern scientific uses* (Revised ed.). London: Tiger Book International.
- Harliansyah, Murad, N. A., Ngah, W. Z. W., & Yusof, Y. A. M. (2007). Antiproliferative, Antioxidant and Apoptosis Effects of Zingiber Officinale and 6-Gingerol on HepG2 Cells. *Asian Journal of Biochemistry* 2(6), 421-426.
- Hawthorne, S. B., Grabanski, C. B., Martin, E., & Miller, D. J. (2000). Comparisons of Soxhlet extraction, pressurized liquid extraction, supercritical fluid extraction and subcritical water extraction for environmental solids: recovery, selectivity and effects on sample matrix. *Journal of Chromatography A*, 892, 421-433.
- Hawthorne, S. B., Yang, Y., & Mille, D. J. (1994). Extraction of Organic Pollutants from Environmental Solids with Sub- and Supercritical Water. *Analytical Chemistry*, 66(18), 2912-2920.
- Harvey, D. J. (1981). Gas Chromatographic and Mass Spectrometric Studies of Ginger Constituents Identification of Gingerdiones and New Hexahydrocurcumin Analogues *Journal of Chromatography*, 212, 75-84.
- Hao, W. W., Min, W. Z., Zhen, X. L., & Lin, Y. S. (2002). HPLC determination of 6-gingerol in Rhizoma Zingiberis Recens. *China journal of Chinese materia medica* 27(5), 348-349.
- Hayashi, Y., Matsuda, R., & Poe, R. B. (1995). Prediction of precision from signal and noise measurement in liquid chromatography: Limit of detection. *Chromatographia*, 41(1/2), 66-74.
- He, X.-g., Bernart, M. W., Lian, L.-z., & Lin, L.-z. (1998). High-performance liquid chromatography–electrospray mass spectrometric analysis of pungent constituents of ginger. *Journal of Chromatography A*, 796 327-334.
- Herrero, M. C., Prados-Rosales, R. C., Luque-Garcia, J. L., & Castro, M. D. L. d. (2002). Static–dynamic pressurized hot water extraction coupled to on-line filtration–solid-phase extraction–high-performance liquid chromatography–post-column derivatization–fluorescence detection for the analysis of N-methylcarbamates in foods. *Analytica Chimica Acta*, 463, 189-197.

- Herrero, M., Cifuentes, A., & Ibanez, E. (2006). Sub-and supercritical fluid extraction of functional ingredients from different natural sources: Plants, food-by-products, algae and microalgae : A review. *Food Chemistry*, 98, 136-148.
- Herrero, M., Mendiola, J. A., Cifuentes, A., & Ibáñez, E. (2009). Supercritical fluid extraction: Recent advances and applications. *Journal of Chromatography A*, 1217(16), 1-17.
- Heinerman, J. (1988). *Heinermans Encyclopedia of Fruits, Vegetables and Herbs*. West Nyack, New York: Parker Publishing Company.
- Huang, Z., Shi, X.-h., & Jiang, W.-j. (2012). Theoretical models for supercritical fluid extraction. *Journal of Chromatography A*, 1250(0), 2-26.
- Geankoplis, C. J. (2003). *Transport processes and separation process principles (Includes unit operation)* (Fourth ed.). Upper Saddle River, New Jersey: Pearson Education, Inc.
- Iheozor-Ejiofor, P., & Dey, E. S. (2009). Extraction of rosavin from *Rhodiola rosea* root using supercritical carbon dioxide with water. *The Journal of Supercritical Fluids*, 50, 29–32.
- IUPAC. (1997). *Compendium of Chemical Terminology* 2nd Ed. Oxford: Blackwell Scientific Publications.
- Jensen, W. B. (2007). The Origin of the Soxhlet Extractor. *Journal of Chemistry Education*, 84, 1913-1914.
- Jimenez-Carmona, M. M., Ubera, J. L., & Castro, M. D. L. d. (1999). Comparison of continuous subcritical water extraction and hydrodistillation of marjoram essential oil. *Journal of Chromatography A*, 855, 625-632.
- Khajenoori, M., Asl, A. H., Hormozi, F., Eikani, M. H., & Bidgoli, H. N. (2009). Subcritical water extraction of essential oils from *Zataria multiflora* Boiss. *Journal of Food Process Engineering* 32, 804-816.
- Kim, W. J., Kim, J., Veriansyah, B., Kim, J. D., Lee, Y. W., Oh, S. G., et al. (2009). Extraction of Bioactive Components from *Centella Asiatica* using Subcritical Water. *Journal of Supercritical Fluids* 48, 211-216.
- Kim, W.-J., Veriansyah, B., Lee, Y.-W., Kim, J., & Kim, J.-D. (2010). Extraction of Mangiferin from Mahkota Dewa (*Phaleria macrocarpa*) using Subcritical Water *Journal of Industrial and Engineering Chemistry*, 16 (425–430).

- King, J. W., Jr, H. H. H., & Lee, M. L. (1993). Analytical supercritical fluid chromatography and extraction. In B. W. Rossiter & R. C. Baetzold (Eds.), *Physical Methods of Chemistry Series* (2 ed., Vol. 10). Peoria, IL: John Wiley & Sons, Inc.
- King, J. W., Grabiell, R. D., & J.D.Wightman. (2003, April 28-30). *Subcritical Water Extracion of Anthocyanins from Fruit Berry Substrates*. Paper presented at the 6th International Symposium on Supercritical Fluids, Versailles, France.
- Kubatova, A., Jansen, B., Vaudoisot, J.-F., & Hawthorne, S. B. (2002). Thermodynamic and Kinetic Models for the Extraction of Essential Oil from Savory and Polycyclic Aromatic Hydrocarbons from Soil with Hot (Subcritical) Water and Supercritical CO<sub>2</sub>. *Journal of Chromatography A*, 975, 175-188.
- Kuhn, E. R. (2002). Water Injections in GC : How Wet Can You Get? *LCGC Asia Pacific*, 5(3), 30-32.
- Kyung, K. S., Gon, J. H., Geun, K. Y., Sup, J. J., Suk, W. J., & Ho, K. J. (2006). 6-Shogaol, a Natural Product, Reduces Cell Death and Restores Motor Function in Rat Spinal Cord Injury. *European Journal of Neuroscience* 24, 1042 - 1052.
- Lack, E., & Seidlitz, H. (2001). Economics of high pressure process. In A. Bertucco & G. Fetter (Eds.), *High Pressure Process Technology: Fundamentals and Applications* (pp. 437-452). Amsterdam, Netherlands: Elsevier Science B. V.
- Lagadec, A. J. M., Miller, D. J., Lilke, A. V., & Hawthorne, S. B. (2000). Pilot-scale subcritical water remediation of polycyclic aromatic hydrocarbon and pesticide contaminated Soil. *Enviromental Science & Technology*, 34(8), 1542-1548.
- Lee, S., Khoo, C., Halstead, C. W., Huynh, T., & Bensoussan, A. (2007). Liquid chromatographic determination of 6-, 8-, 10-gingerol, and 6-shogaol in ginger (*Zingiber officinale*) as the raw herb and dried aqueous extract. *Journal of AOAC International*, 90( 5), 1219-1226.
- Liu, C., & Wyman, C. E. (2005). Partial flow of compressed-hot water through corn stover to enhance hemicellulose sugar recovery and enzymatic digestibility of cellulose. *Bioresource Technology*, 96(18), 1978-1985.
- Li, B., Yang, Y., Gan, Y., Eaton, C. D., He, P., & Jones, A. D. (2000). On-line coupling of subcritical water extraction with high-performance liquid

- chromatography via solid-phase trapping. *Journal of Chromatography A*, 873, 175-184.
- Liang, M. H. (1992). From America: cookbook medicine or food for thought: practice guidelines development in USA. *Annals of the Rheumatic Diseases*, 51, 1257-1258.
- Lynden-Bell, R. M., Morris, S. C., Barrow, J. D., Finney, J. L., & Charles L. Harper, J. (2010). Water's hydrogen bond strength *Water and life: The unique properties of H<sub>2</sub>O* (1st ed.). Boca Raton, FL: CRC Press.
- Markom, M., Hasan, M., & Daud, W. R. W. (2010). Pressurized Water Extraction of Hydrolysable Tannins from *Phyllanthus Niruri* Linn. *Separation Science and Technology*, 45, 548-553.
- McNaught, A. D., & Wilkinson, A. (1997). *Compendium of chemical terminology* (2nd ed.). Oxford: Blackwell Science.
- Melianita, F., Witha, J., Arifin, S., Kartinasari, W. F., & Indrayanto, G. (2009). Simultaneous densitometric determination of 6-Gingerol, 8-Gingerol, 10-Gingerol, and 6-Shogaol in some commercial gingers. *Journal of Liquid Chromatography & Related Technologies*, 32, 567-577.
- Mitra, P., Barman, P. C., & Chang, K. S. (2011). Coumarin Extraction from *Cuscuta reflexa* using Supercritical Fluid Carbon Dioxide and Development of an Artificial Neural Network Model to Predict the Coumarin Yield. *Food and Bioprocess Technology*, 4, 737-744.
- Miller, J. N. (1991). Basic Statistical Methods for Analytical Chemistry Part 2. Calibration and Regression Methods. *Analyst*, 116, 3-14.
- Mohammad H. Eikani, F. G., Mehdi Mirza and Soosan Rowshanzamir. (2006). Extraction of Volatile Oil from Cumin (*Cuminum Cyminum* L.) with Superheated Water. *Journal of Food Process Engineering* 30 (2007), 255-266.
- Mohamed, N. A. (2005). *Study on important parameters affecting the hydro-distillation for ginger oil production*. Master Degree, Universiti Teknologi Malaysia, Skudai.
- Morad, N. A., Mulop, N., & Darus, A. N. (2011). *Introduction to thermodynamics for engineering students*. Kuala Lumpur, Malaysia: Pearson Custom Publishing.

- Nagoshi, C., Shiota, S., Kuroda, T., Hatano, T., Yoshida, T., Kariyama, R., et al. (2006). Synergistic Effect of [10]-Gingerol and Aminoglycosides against Vancomycin-Resistant Enterococci (VRE). *Biological & Pharmaceutical Bulletin*, 29(3), 443-447.
- Norde, W. (2003). *Water Colloids and interfaces in life sciences*. Dresden, Germany: Marcel Dekker, Inc.
- Ozel, M. Z., Gogus, F., & Lewis, A. C. (2002). Subcritical water extraction of essential oils from *Thymbra Spicata*. *Food Chemistry*, 82, 381-386.
- Park, M., Bae, J., & Lee, D.-S. (2008). Antibacterial Activity of [10]-Gingerol and [12]-Gingerol Isolated from Ginger Rhizome Against Periodontal Bacteria. *Phytotherapy research* 22, 1446-1449.
- Perry, R. H. (1997). *Perry chemical engineers' handbook* (Seven ed.). New York: McGraw-Hill.
- Pronyk, C., & Mazza, G. (2009). Design and scale-up of pressurized fluid extractors for food and bioproducts. *Journal of Food Engineering*, 95, 215-226.
- Rangsriwong, P., Rangkadilok, N., & Shotipruk, A. (2008). Subcritical Water Extraction of Polyphenolic Compounds from *Terminalia chebula* Fruits. *Chiang Mai Journal of Science*, 35(1), 103-108.
- Richter, B. E., Jones, B. A., Ezzell, J. L., & Porter, N. L. (1996). Accelerated Solvent Extraction: A Technique for Sample Preparation. *Analytical Chemistry*, 68, 1033-1039.
- Rodriguez-Meizoso, I., Marin, F. R., Herrero, M., Senorans, F. J., Reglero, G., Cifuentes, A., et al. (2006). Subcritical water extraction of nutraceuticals with antioxidant activity from oregano. Chemical and functional characterization. *Journal of Pharmaceutical and Biomedical Analysis*, 41, 1560–1565.
- Rouatbi, M., Duquenoy, A., & Giampaoli, P. (2007). Extraction of the essential oil of thyme and black pepper by superheated steam. *Journal of food engineering* 78, 708-714.
- Rovio, S., Hartonen, K., Holm, Y., Hiltunen, R., & Riekkola, M. L. (1999). Extraction of clove using pressurized hot water. *Flavour and fragrance journal* 14, 399-404.
- Saim, N., & Osman, R. (2008). Subcritical Water Extraction of Essential Oil from Coriander (*Coriandrum Sativum* L.) Seeds. *The Malaysian Journal of Analytical Science*, 12, 22-24.



- Samuri, S. M. (2005). *Optimisation of operating condition parameters for removal of ethanol from zingiber officinale roscoe (Ginger) oleoresin using short-part distillation*. Master Thesis, Universiti Teknologi Malaysia, Skudai.
- Schwertner, H. A., & Rios, D. C. (2007). High-Performance Liquid Chromatographic Analysis of 6-gingerol, 8-gingerol, 10-gingerol, and 6-shogaol in Ginger-Containing Dietary Supplements, Spices, Teas, and Beverages. *Journal of Chromatography B*, 856, 41-47.
- Schwartzberg, H. G., & Chao, R. Y. (1982). Solute diffusivities in leaching processes. *Food Technology* 36, 73–86.
- Smith, R. M. (1982). Analysis of the pungent principles of ginger and grains of paradise by high-performance liquid chromatography using electrochemical detection. *Chromatographia*, 16, 155-157.
- Smith, R. M. (2002). Review: Extractions with superheated water. *Journal of Chromatography A*, 975, 31-46.
- Smith, R. M. (2006). Superheated water: The Ultimate Green Solvent for Separation Science. *Anal Bioanal Chem* 385, 419-421.
- Shukla, Y., & Singh, M. (2007). Cancer preventive properties of ginger: A brief review. *Food and Chemical Toxicology* 45, 683 - 690.
- Sovová, H. (1994). Rate of the vegetable oil extraction with supercritical CO<sub>2</sub>-I. Modelling of extraction curves. *Chemical Engineering Science*, 49(3), 409-414.
- Spellman, F. R. (2010). *Water The Science of Environmental Pollution* (Second ed.). Boca Raton, FL: CRC Press.
- Sun, Y., Liu, Z., Wang, J., Tian, W., Zhou, H., Zhu, L., et al. (2008). Supercritical fluid extraction of paeonol from *Cynanchum paniculatum* (Bge.) Kitag. and subsequent isolation by high-speed counter-current chromatography coupled with high-performance liquid chromatography-photodiode array detector. *Separation and Purification Technology* 64, 221-226.
- Tajuddin, R., & Smith, R. (2005). On-line coupled extraction and separation using superheated water for the analysis of triazine herbicides in spiked compost samples. *Journal of Chromatography A*, 1084, 194-200.
- Teo, C. C., Tan, S. N., Yong, J. W. H., Hew, C. S., & Ong, E. S. (2010). Pressurized Hot Water Extraction (PHWE). *Journal of Chromatography A*, 1217, 2484-2494.

- Timmermans, J. (1950). *Physico-chemical constants of pure organic compounds*. Brussel: Elsevier Publishing Company, Inc
- Turner, C., Turner, P., Jacobson, G., Almgren, K., Waldeba, M., Sjöberg, P., et al. (2006). Subcritical Water Extraction and  $\beta$ -glucosidase-catalyzed Hydrolysis of Quercetin Glycosides in Onion Waste. *The Royal Society of Chemistry*, 8, 949–959.
- Uematsu, M., & Frank, E. U. (1980). Static Dielectric Constant of Water and Steam. *Journal of Physical and Chemistry Reference Data*, 9(4), 1291-1306.
- Vimala, S., Norhanom, A., & Yadav, M. (1999). Anti-tumour Promoter Activity in Malaysian Ginger Rhizobia Used in Traditional Medicine. *British Journal of Cancer*, 80(1/2), 110 - 116.
- Wagner, W., & Pruß, A. (2002). The IAPWS Formulation 1995 for the Thermodynamic Properties of Ordinary Water Substance for General and Scientific Use. *Journal of Physical and Chemistry Reference Data*, 31(2). 387-512.
- Watanabe, K. (2003). *Revised Release on the IAPS Formulation 1985 for the Viscosity of Ordinary Water Substance*. Vejle, Denmark: The International Association for the Properties of Water and Steam.
- WADA. (2010). Identification Criteria for Qualitative Assays Incorporating Column Chromatography and Mass Spectrometry WADA Executive Committee
- Weidner, M. S., & Sigwart, K. (2000). The safety of a ginger extract in the rat. *Journal of Ethnopharmacology*, 73, 513–520.
- Weingärtner, H., Franck, E. U., Wiegand, G., Dahmen, N., Schwedt, G., Frimmel, F. H., et al. (2002). Water : Properties, analysis and hydrological cycle *Ullmann's Encyclopedia of Industrial Chemistry*. Weinheim, Germany: Wiley-VCH Verlag GmbH & Co. KGaA.
- Wiboonsirikul, J., & Adachi, S. (2008). Extraction of Functional Substances from Agricultural Products or By-products by Subcritical Water Treatment. *Food Science Technology Research*, 14 (4), 319-328.
- Wohlmuth, H., Leach, D. n., Smith, M. K., & Myers, S. P. (2005). Gingerol Content of Diploid and Tetraploid Clones of Ginger (*Zingiber officinale* Roscoe). *Journal of Agricultural and Food Chemistry*, 53, 5772-5778.
- Wohlmuth, H., Smith, M. K., Brooks, L. O., Myers, S. P., & Leach, D. N. (2006). Essential Oil Composition of Diploid and Tetraploid Clones of Ginger

(*Zingiber officinale* Roscoe) Grown in Australia. *Journal of Agricultural and food Chemistry* 54, 1414-1419.

Yunus, M. A. C. (2007). *Extraction Identification and Separation of Vitamin E and Djenkolic Acid from Pithecellobium Jiringan (Jack) Prain Seeds using Supercritical Carbon Dioxide*. Doctor Philosophy, Universiti Sains Malaysia, Penang.

Zachariah, T. J. (2008). Ginger. In V. A. Parthasarathy, B. Chempakam & T. J. Zachariah (Eds.), *Chemistry of Spices* (pp. 70-95). India: CAB International.

Zick, S. M., Turgeon, D. K., Vareed, S. K., Ruffin, M. T., Litzinger, A. J., Wright, B.D., Sara, A, Normolle, D.P., Djuric, D., Brenner, D.E. (2011). Phase II Study of the Effects of Ginger Root Extract on Eicosanoids in Colon Mucosa in People at Normal Risk for Colorectal Cancer. *Cancer Prevention Research* 4(11), 1-9.