

PRODUCTION OF POLYHYDROXYALKANATE (PHA) FROM WASTE  
COOKING OIL USING *PSEUDOMONAS OLEOVORANS*

FARZANEH SABBAGH MOJAVERYAZDI

A dissertation report submitted in partial fulfilment of the  
requirements for the award of the degree of  
Master of Science (Biotechnology)

Faculty of Biosciences and Bioengineering  
Universiti Teknologi Malaysia

FEBRUARY 2013

*To my beloved husband, father and mother  
For their love, support, sacrifices and blessings  
And to all other beloved ones*

*God bless them all!*

## ACKNOWLEDGEMENT

First and foremost, I would like to express my appreciation and sincere gratitude to my supervisor, Dr. Nor Azimah Mohd Zain, who always giving me lots of guidance, advice and invaluable support as well as giving lots of patient throughout the trials and tribulations in completing this project.

Besides, I would like to grab this opportunity to convey my gratitude to my good husband Hamidreza who, cares, support and helped me, as well as to all lecturers, academic staff and all of my classmates who have helped and guided during the research process in the Project Lab. Special thanks to those who had given their support, helped and advice me in completing my project directly or indirectly.

Finally, I would like to thank my mother, and father for their support, cares, advice and loves in completing this master project.

Thank you everyone.

## ABSTRACT

Polyhydroxyalkanoates (PHAs) are biodegradable polyesters which are stored in bacterial cell cytoplasm as reserve materials for carbon and energy. One of the main problems of plastics is that they have resistance to biological breakdown that result in accumulation in the environment. Bacteria synthesize and accumulate polyhydroxyalkanoate (PHA) as carbon source under limiting conditions of nutrients. In this study, the bacteria, *P. oleovorans* was studied for its ability to produce PHA in the minimal basal medium supplemented with glucose as carbon source and  $(\text{NH}_4)_2\text{SO}_3$  as nitrogen source that was grown at 25°C. The functional groups of the extracted PHA granules were identified as a C=O group by Fourier Transform Infrared (FTIR) spectroscopy analysis. The drastic absorption band at approximately 1720  $\text{cm}^{-1}$  indicated the stretching vibration of the C=O groups in the PHA polyester. The influence of different carbon sources, nitrogen sources, pH and inoculums on PHA production was investigated. The production optimization of PHA was done by RSM (Response Surface Method) through various growth parameters. After optimization obtained the best condition of productivity in range are pH 5.9, carbon source 93.4419 g/l, inoculums size 2.07% and nitrogen source 101.691 g/l. Also the highest PHA production after optimization is 2.28236 g/L with a desirability of 0.986 g/l, meanwhile the highest amount of PHA produced from *P. oleovorans* was 2.30 g/l.

## ABSTRAK

Polyhydroxyalkanoates (PHA) adalah polyesters terbiodegradasi yang mana telah disimpan di dalam sel cytoplasm sebagai bahan bakteria tersimpan untuk karbon dan tenaga. Satu daripada masalah utama adalah plastik yang mana ianya mempunyai rintangan terhadap penguraian biologikal maka mengakibatkan pengumpulan terhadap alam sekitar. Bacteria sintesis dan PHA terkumpul sebagai sumber karbon di bawah had keadaan nutrient. Di dalam kajian ini bakteria *P.Oleovorans* dikaji keupayaannya untuk menghasilkan PHA di dalam medium asas tambahan bersama glukosa sebagai sumber karbon dan  $(\text{NH}_4)_2\text{SO}_3$  sebagai sumber nitrogen yang mana hidup pada suhu 25°C. Fungsi kumpulan-kumpulan granular PHA yang diekstrakkan telah dikenalpasti sebagai satu kumpulan C=O oleh analisis Fourrer Transform Infrared (FTIR). Kumpulan serapan drastic terletak kira kira 1720  $\text{cm}^{-1}$  adalah menunjukkan getaran penegangan kumpulan C=O didalam polyester PHA. Pengaruh sumber karbon yang berbeza, sumber nitrogen yang berbeza, pH yang berbeza dan inoculum yang berbeza keatas penghasilan PHA dikaji. Analisis statistikal telah dibuat untuk menganalisis data menggunakan kaedah tindak balas permukaan (RSM). Daripada data analisis yang diperolehi, nilai pH terbaik untuk penghasilan PHA adalah 5.98201, sumber karbon adalah 93.4419, sumber nitrogen adalah 101.691 dan saiz inokulum adalah 2.07. Jumlah penghasilan PHA yang tertinggi selepas proses pengoptimuman adalah 2.28236 g/L dengan nilai kejituan sebanyak 0.986. Manakala jumlah penghasilan tertinggi PHA daripada *P.oleovorans* adalah 2.30 g/l.

**TABLE OF CONTENTS**

<b>CHAPTER</b>	<b>TITLE</b>	<b>PAGE</b>
	<b>TITLE</b>	ii
	<b>DECLARATION</b>	iii
	<b>DEDICATION</b>	iv
	<b>ACKNOWLEDGEMENT</b>	v
	<b>ABSTRACT</b>	vi
	<b>ABSTRAK</b>	vii
	<b>TABLE OF CONTENTS</b>	viii
	<b>LIST OF TABLES</b>	xii
	<b>LIST OF FIGURES</b>	xiii
<b>1</b>	<b>INTRODUCTION</b>	1
	1.1 Literature Review	1
	1.2 Statement of Problem	3
	1.3 Objectives of Study	5
	1.4 Scope of Study	5
	1.5 Significance of Study	5
	1.6 Pathway of PHA Production	6
<b>2</b>	<b>LITERATURE REVIEW</b>	8

2.1	General Overview of Polyhydroxyalkanoates (PHA)	8
2.1.1	Scl-PHA	9
2.1.2	Mcl-PHA	10
2.2	Chemistry of PHA	11
2.3	Physical Properties of PHA	13
2.4	Advantages Characteristics of PHAs	14
2.4.1	Biodegradable	14
2.4.2	Bio-Based Nature of PHA	15
2.4.3	Carbon dioxide Release	15
2.4.4	Biocompatibility	15
2.5	The Biology of PHA	16
2.6	Bacteria Strain	17
2.7	Recovery of PHA	20
2.8	Pathways for Mcl-PHA Synthesis	21
2.8.1	$\beta$ -oxidation	21
2.8.2	Fatty acid Biosynthesis	23
2.9	Carbon Substrate and Yield	24
2.10	Economics of PHA Production	26
2.11	Other Types of PHA and Application	27
2.11.1	The Possible Applications of PHA	29
2.12	Industrial Production	30
<b>3</b>	<b>Materials and Methods</b>	<b>31</b>
3.1	Materials	31
3.2	Microorganism	31
3.3	Characterization of Waste Cooking Oil	32
3.3.1	Determination of Saponification Value	32

3.3.2	Determination of Acid Value	33
3.3.3	Determination of Moisture Content	33
3.3.4	Determination of Fatty acid Composition of Oils and Fats by GC	34
3.4	Preliminary Study to Determine the Best Nitrogen Source and Carbon Source	35
3.4.1	Effect of Different Nitrogen Sources on PHA Production	35
3.4.2	Effect of Different Carbon Sources on PHA Production	35
3.5	Experimental Design	35
3.6	Determination of Bacteria Density and Cell Dry Weight	36
3.6.1	Growth Experiments with Selected Bacteria	36
3.7	Preparation of Inoculums	37
3.7.1	Preparation of Stock Culture	37
3.8	Media Preparation	37
3.8.1	Nutrient Broth	37
3.8.2	Nutrient Agar	38
3.8.3	Minimal Basal Medium (MBM)	39
3.9	Optimization by Response Surface Methodology (RSM)	39
3.10	PHA Production	41
3.10.1	Extraction of PHA	41
3.10.2	Staining Procedure	42
3.11	Characterization of PHA	42



3.11.1 PHA Identification by UV Spectrophotometer	42
3.11.2 Estimation of Phase by Crotonic Acid Assay	43
3.11.3 Fourier Transform-Infrared Spectroscopy (FT- IR Analysis)	44
<b>4 RESULTS AND DISCUSSION</b>	<b>45</b>
4.1 Preliminary Study of <i>Pseudomonas oleovorans</i> as Potential PHA Production	45
4.1.1 Effect of Different Carbon Sources on Yield of PHA (g/l) Produced By <i>Pseudomonas oleovorans</i>	45
4.1.2 Effect of Different Nitrogen Sources on PHA Yield by the <i>Pseudomonas oleovorans</i>	46
4.2 Characterization of Waste Cooking Oil	47
4.3 Optimum Production of PHA using CCD	50
4.4 Diagnostic Plots for Response (PHA)	52
4.5 Confirmation Runs	55
4.6 Analysis of Variance (ANOVA) Using CCD	56
4.7 Monitoring the Crystallization of PHA Granules Using Electron Microscope	59
4.8 Isolation and Characterization of PHA	60
4.9 FTIR Analysis	61
<b>5 CONCLUSION</b>	<b>62</b>
5.1 Conclusion	62
5.2 Future Work	63
<b>6 REFERENCES</b>	<b>64</b>

**LIST OF TABLES**

<b>TABLE NO.</b>	<b>TITLE</b>	<b>PAGE</b>
2.1	Classification of microbial bio-polymers (PHA) according to different scales	13
3.2	Complete design layout corresponding runs Overview of all experiments designed by central composite	40
4.1	Properties of waste cooking oil from palm oil	48
4.2	Overview of all experiments designed by central composite design (CCD) with 4 sets of parameters (pH, carbon source, nitrogen source, inoculum)	51
4.3	Conformation runs results	56
4.4	Analysis of variance (ANOVA)	58

## LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
1.1	MCL-PHA biosynthesis pathway of <i>P.oleovorans</i>	7
2.1	Scl-PHA biosynthetic pathway 1- $\beta$ -Ketoacyl-CoA thiolase (PhbA).	10
2.2	General structural formula of polyhydroxyalkanoate (PHA)	12
2.3	PHA granules inside cytoplasm of the cell	16
2.4	De novo Mcl-PHA production	19
2.5	Pathway for the biosynthesis of mcl-PHA from fatty acids in <i>Pseudomonas oleovorans</i>	22
2.6	Pathway for the biosynthesis of mcl-PHA from carbohydrates in <i>pseudomonas</i>	23
4.1	Effect of different carbon sources on PHA production (w/v%)	46
4.2	Effect of Different Nitrogen Sources on PHA Yield by the <i>Pseudomonas oleovorans</i>	47
4.3	Saponification of fatty acids	49
4.4	Normal plot of residuals	52
4.5	Outlier T plot	53

4.6	Cook's distance	54
4.7	Predicted vs. actual plot Leverage vs. run plot	54
4.8	Predicted value for optimum PHA production as	54
4.9	generated by the design	55
4.10	Electron microscope image of PHA produced by <i>P.oleovorans</i>	59
4.11	UV spectrophotometer of PHA (red line) and crotonic acid (blue line)	60
4.12	FT-IR Spectrum of PHA produced by <i>Pseudomonas</i> <i>oleovorans</i>	61

## LIST OF ABBREVIATIONS

ASTM	-	American Society for Testing and Materials
CCD	-	Central composite design
FT-IR	-	Fourier Transform-Infrared Spectroscopy
FLP	-	FLiPpase
GC	-	Gas Chromatography
g	-	Gram
g/L	-	Gram per Liter
GLP	-	Glycerol liquid phase
HA	-	hydroxyalkanoate
mL	-	Mililiter
MBM	-	Minimal Basal Medium
w/v	-	Weight/ Volume
v/v	-	Volume/ Volume
°C	-	Degree Celcius
PHA	-	Polyhydroxyalkanoate
PHA <sub>SCL</sub>	-	Short chain length PHA
PHA <sub>MCL</sub>	-	Medium chain length PHA
PHA <sub>LC</sub>	-	Long chain length PHA
P (3HB)	-	poly-3-hydroxybutyrate
P (3HV)	-	poly-3-hydroxyvalerate
P (3HB-3HV)	-	poly-3-hydroxybutyrate-co-3-hydroxyvalerate
P (4HB)	-	poly-4-hydroxybutyrate
USDA		United States Department of Agriculture
WCO		Waste cooking oil
RSM		Response Surface Methodology

## CHAPTER 1

### INTRODUCTION

#### 1.1 Literature review

Polyhydroxyalkanoates (PHAs) are polymers of hydroxy fatty acids and family of synthesized biopolyesters that agglomerated by some of bacteria strains as carbon storage materials. These polymers are natural synthesized biopolymers and decomposed by some microbial metabolisms, although these biopolymers are able to be melted and molded similar to chemical and unnatural thermoplastics (Koller *et al*, 2005).

Because of potential using PHAs as biodegradable thermoplastics these biopolymers have been more attended at academic and industrial fields. Several academic studies have been repeated on the physiology of PHA, many processing procedures, molecular biology, biochemistry of these biopolymers, many kinds of material properties and biodegradation of PHA. They can be polymerized from degradable carbon sources which are produced by agricultural and industrial wastes and products (Lee, 1996).

The wastes are generated by many industries including wood-processing industry, slaughterhouse industry, etc. These wastes sometimes containing carbon-rich substrates for example polysaccharides, disaccharides, monosaccharides, lipids which can be useful for PHA production. In general, molass wastes, starch wastes, whey as dairy wastes, glycerol liquid phase (GLP) as the biodiesel production waste, xylose, lipids and waste water obtained from oil production occurring in large quantities (Koller *et al.*, 2007). Production of PHA from waste contains double benefits, one of them is that polluting waste derived from environment can be converted into biodegradable polymer or environment friendly pollutants. Economically, upon using waste products as a substrate, the cost of the carbon source, as the most important factor significantly can be decreased to the overall production cost (Choi, 1997). Its benefit is that, it makes the PHA production more economical and to handle waste without further disposal cost (Lee, 1999).

Additionally, PHAs have various properties that can be produced and commercially used in different fields such as packaging films and containers (Reddy *et al.*, 2003). Furthermore biodegradable carriers which are used for drugs carrying, hormones carrying, insecticides or herbicides carrying is also recommended (Reddy *et al.*, 2003; Zinn *et al.*, 2001). Production of PHA in large scale, using microbial fermentation, has so far been done. However the improvement of transgenic PHA agglomerating plants has found considerable progress and its production in industrial scale is also possible (Valentin *et al.*, 1999).

Due to the restriction of a nutrient, electron donor and electron acceptor, under stress conditions, bioplastics or biopolymers are stored as granules inside the cytoplasm of bacteria. Generally, PHA accumulation is favored by sufficient carbon sources and their availability also limiting supplies nitrogen and phosphate as macro components and

or micro components for example magnesium (Mg), sulphate (S), iron (Fe), potassium (K), manganese (Mn), copper (Cu), calcium (Ca), tin and sodium cobalt (Sc) (Kim *et al.*,2001; Helm *et al.*,2008). For microbial production of PHA, PHAs feed as store materials as carbon and energy sources. In starvation conditions, these store matters can be mobilized, therefore the cell provides with the superiority to maintenance (Goh, 2008).

## 1.2 Statement of Problem

Currently, there are a lot of environmental problems of plastics, used in packaging, thus making biodegradable plastics from bacteria is needed (Ahleum *et al.*,2009). In this case the first material is waste cooking oil (palm oil), which can help to dominate to this problem. Many organizations have established that they have facilitated the development in the production of bioplastics (Davisa, 2006). Whereas considerable advancement has been achieved in recycling of metals and also packaging with glass (Oakley-Hill, 1999). Few successful in this area have been achieved by decreasing the fuel polymer packaging wastes in landfill. Fuel packaging materials consist of large amount different types of polymers that each of these polymers may contain different chemical additives for example fillers, colorants or plasticizers.

By incinerating the plastic waste, the chemical energy stored therein recovers as thermal energy. Degradation of PHA will be occurred in two-steps. First, based on the type of depolymerases, PHA will be degraded into its monomers (Jendrossek,1993), and dimmers (Schrimmer,1993) or a mixture of oligomers. Second, an enzyme called oligomer hydrolase cleaves the PHA oligomers into simple monomers (Shirakura,1983). Nowadays, increasing industrial interest is due to the bio-producing the PHA from



recyclable carbon sources to make a bio-material and biodegradable polymers that can act as intermittent for current fuel plastics (Braunegg *et al.*, 1998).

If products made of PHAs are cycled in the nature, the final products of oxidative breakdown they will be naturally degrade to produce water and CO<sub>2</sub>. Carbon dioxide and water as the ultimate products of oxidation are the first substrates for the production of carbohydrates produced by plants. This matter indicates that, compare with fuel plastics, polyhydroxyalkanoates are encompassing into the natural carbon cycle. The PHA application not only apply for packaging industry, but also embraces commodity stuffs for some agricultural purposes also for some applications such as pharmaceutical and medical. A resistant solution procedure can be identified as consumption of a spread span of waste materials which can be explained as the role of substrate for the biomediat generation of favorable final products. These natural materials are mostly generated in agricultural and industrial products (Braunegg *et al.*, 1998).

In waste oils, plant oils, or in waste water produced from oil factories in the industry, the triacylglycerides immediately, or during hydrolysatation of oils to glycerol and free fatty acids, and also after transesterification to the glycerols could be utilized as carbon sources in order to PHA production ( Archana *et al.*, 2012).

### 1.3 Objectives of Study

The objectives of the study are

- To culture *Pseudomonas* strain in a suitable media.
- To characterize waste cooking oil (WCO)
- To produce mcl-PHA by *Pseudomonas oleovorans* from WCO.
- To study the important parameters in producing PHA

### 1.4 Scope of Study

This study is limited to the production of polyhydroxyalkanoates from waste cooking oil using *Pseudomonas oleovorans*. *Pseudomonas oleovorans* is a gram negative bacteria. Its optimal temperature is 35°C and has poor growth at 41°C. These bacteria will be grown under optimized conditions with suitable growth factors. The procedure which should be done for recovery of PHA including pre-treatment and extraction. Centrifugation and washing the biomass with distilled water is the pre-treatment step and after that extraction will be done using sodium hypochlorite and chloroform. Different factors affecting on PHA production by *Pseudomonas oleovorans* will be optimized, including carbon source, nitrogen source, pH and inoculumns.

## 1.5 Significance of Study

The world we are living today is definitely covered with fossil fuel plastics. From historical point of view, the production of fossil fuel plastics has been increasing rapidly since the 20th century (Cervenkova, 2007). Nowadays, the overall production volume of fuel plastics reached over 250 million tons at the end of 2010 (Marsalek, 2011).

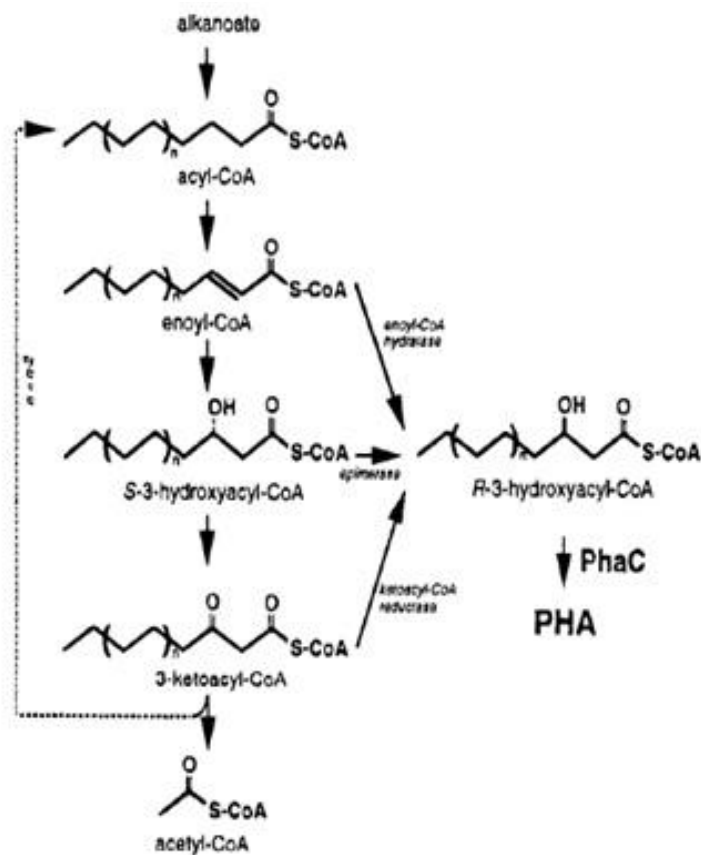
The yearly world production of fuel polymers amounts is more than 140 million tons. A large amount of this volume is composed of chemically stable polymers that are not easily degraded to the nature (Shimao, 2001). These are mostly synthetic polymers that are produced essentially by chemical addition or condensation reactions in a large number of monomers and are joined sequentially. The strong using of stabilizers and its presence in widely used plastics such as nylon, polyethylene (PE), polyethylene terephthalate (PET) or polyvinyl chloride (PVC) is partially responsible for their poor biodegradability. Therefore, interest in the degradation process of several polymers and in the use of environmental-friendly alternatives has increased (Birgit Kessler *et al.*, 2001).

## 1.6 Pathway of PHA Production

*Pseudomonas oleovorans* has a vast broad substrate range (Huisman *et al.*, 1989). It exhibits a high flexibility in incorporating monomers into PHA producing (Witholt and Kessler 1999) and accumulates PHA polymers up to a cellular content of 65 % (Hazenberg and Witholt 1997). These aspects make *P. oleovorans* an excellent candidate for the production of a different variety of PHAs. To produce bio polymers

(PHA), the strain produces two PHA polymerases, PhaCl and PhaCz (Huisman *et al.*, 1991).

These two polymerases have the same function, i.e. condense R-3-OH-acyl-CoA moieties through an ester bond. The acyl-CoA activated precursors of the polymerases are derived from either fatty acids, alkanals, alkanols or alkanes, which are channeled through the P-oxidation cycle via 3-ketoacyl-CoA to R-3-OH-acyl-CoA. Once polymerized, PHA accumulates at insoluble gathered, so-called granules, which are formed in the cytoplasm of the bacteria. An accepted model of the general structure of PHA granules was presented by Steinbtichel *et al.* (1995).



**Figure 1.1** MCL-PHA biosynthesis pathway of *P. Oleovorans* (Madison *et- al.* 1999)

## REFERENCES

- Ahleum Chung , Q. L., Shao-Ping Ouyang , Qiong Wu , Guo-Qiang Chen (2009). Microbial production of 3-hydroxydodecanoic acid by PHA operon and fadBA knockout mutant of *Pseudomonas putida* KT2442 harboring tesB gene. *Appl Microbiol Biotechnol*.
- Amutha Santhanam, S. S. (2010). Microbial production of polyhydroxy alkanotes (PHA) from *Alcaligenes spp.* and *Pseudomonas oleovorans* using different carbon sources. *African Journal of Biotechnology Vol. 9(21)*, 3144-3150.
- Anderson, A. J., and E. A. Dawes. (1990). Occurrence, metabolism, metabolic role, and industrial uses of bacterial polyhydroxyalkanoates. 450-472.
- Ankamong, E. (2010). *The Influence of Physicochemical Characteristics of Vegetable Oils on the Quality of Biodiesel Produced from Palm Oil, Palm Kernel Oil, Refined Soyabean Oil, Unrefined Soyabean Oil and Jatropha Curcas Oil*. Kumasi.
- Anwar Munir, S. K., Anna Villar Piqué, Hans Leemhuis, Marc J. E. C. Van Der Maarel And Lubbert Dijkhuizen.(2010). Inulin and levan synthesis by probiotic *Lactobacillus gasseri* strains: characterization of three novel fructansucrase enzymes and their fructan products.
- Archana Tiwari, A. M. R., Roopesh Jain, Anushri Saxena. (2012). Green Chemistry for the Production of Biodegradable Polymers as Solid Substrate and the Formation of Sustainable Biofilm. *Key Engineering Materials Vol. 517* 755-762.
- Azmalisa, T., Wan Asma, I., Zulkafli, H. and Norazwina, Z.(2010). *Optimization of Glucose Production from Oil Palm Trunk via Enzymatic Hydrolysis*. Paper presented at the The 13th Asia Pacific Confederation of Chemical Engineering Congress (APCChE'2012).

- Beaulieu, J.P.e.a. (1995). Spectroscopic studies of the two EROS candidate microlensed stars. *Astronomy and Astrophysics*.
- Birgit Kessler, B. W. (2001). Factors involved in the regulatory network of polyhydroxyalkanoate metabolism. *Journal of Biotechnology* 86, 97–104.
- Burdon, K. L. (1946). Fatty Material in Bacteria and Fungi Revealed by Staining Dried, Fixed Slide Preparations. *Journal of Bacteriology*, 52(6), 665–678.
- C.S.K Reddy, R. G., Rashmi, V.C Kalia. (2003). Polyhydroxyalkanoates: an overview. *Bioresource Technology*, 87(2), 137-146.
- Catalina voaides, D. g., Matilda ciuca, Irina lupescu, Aneta pop, Calina petruta cornea (2010). PHAs accumulation in *pseudomonas putida* p5 (wild type and mutants) in lipid containing media. *Romanian biotechnological letters*, 15.
- Cynthia Nkolika Ibeto, C.O. B. O., and Akuzuo Uwaoma Ofoefule. (2012). Comparative Study of the Physicochemical Characterization of Some Oils as Potential Feedstock for Biodiesel Production. *ISRN Renewable Energy*.
- Escapa J.L.García, B.B., L. M.Blank and M.A. Prieto.(2012). The polyhydroxyalkanoate metabolism controls carbon and energy spillage in *Pseudomonas putida*. *Environmental Microbiology* 14, 1049–1063.
- Hanna D. olvan, L. F., Knud V. Christensen, Birgir Norddahl. (2008). A Review of the Current State of Biodiesel Production Using Enzymatic Transesterification. *Biotechnology and Bioengineering*, 102(5), 1298-1315.
- Hanna, F.M.a.M.A. (1999). Biodiesel production: a review. *Bioresource Technology*, 70, 15.
- Harrington Aa, K. R. (1960). Oxidation of methanol and formaldehyde by *Pseudomonas methanica*. *Canadian Journal of Microbiology*, 6, 7.
- Isabel F. Escapa , V. M., Verónica P. Martino , Eric Pollet , Luc Avérous , José L. García, MaríaA. Prieto. (2011). Disruption of  $\beta$ -oxidation pathway in *Pseudomonas putida* KT2442 to produce new functionalized PHAs with thioester groups. *Applied Microbiology and Biotechnology*, 89(5), 1583-1598.
- Jantima Teeka, T. I., Xuehang Cheng, Alissara Reungsang, Takaya Higuchi, Koichi Yamamoto, Masahiko Sekine (2010). Screening of PHA-Producing Bacteria

- Using Biodiesel- Derived Waste Glycerol as a Sole Carbon Source. *Journal of Water and Environment Technology*, 8 (2010) No. 4 373-381.
- Jawahar Nisha, N.M., Senthilkumar P., Narendrakumar and Antony V.Samrot (2012). Influence of substrate concentration in accumulation pattern of poly(R) hydroxyalkanoate in *Pseudomonas putida* SU-8. *African Journal of Microbiology Research*,
- Jendrossek.D and Gebauer, B. (2005). Assay of poly(3-hydroxybutyrate) depolymerase activity and product determination. *Applied and Environmental Microbiology*, 72 No. 9, 6094-6100.
- Joaõ M.B.T. Cavaleiro, M. C. M. D. d. A., Christian Grandfils, M.M.R. da Fonseca. (2009). Poly (3-hydroxybutyrate) production by *Cupriavidus necator* using waste glycerol, *Process Biochemistry. Process Biochemistry*, Volume 44(5), 509–515.
- Joanne M. Curley, B. H., and Robert W. Lenz. (1996). Production of Poly(3-hydroxyalkanoates) Containing Aromatic Substituents by *Pseudomonas oleovorans*. *Macromolecules*, 29(5), 1762–1766.
- Jong-il Choi,(1998).Cloning of the *Alcaligenes latus* Polyhydroxyalkanoate Biosynthesis Genes and Use of These Genes for Enhanced Production of Poly(3-hydroxybutyrate) in *Escherichia coli*. *Applied and Environmental Microbiology*, 64 No.12, 4897-4903.
- Jun Xu, X. Q., Jiayin Dai, Hong Cao, Min Yang, Jing Zhang & Muqi Xu. (2006). Isolation and characterization of a *Pseudomonas oleovorans* degrading the chloroacetamide herbicide acetochlor. *Biodegradation*, 17 219-225.
- Kac.A. (2001). The foolproof way to make biodiesel: free fatty acid to ester conversion. *Journey to forever- Handmade projects*.
- Lin,C.S. K., Sabirova, Julia, Soetaert, Wim, Du Chenyu. (2012). Polyhydroxyalkanoates Production From Low-cost Sustainable Raw Materials. *Current Chemical Biology*, 14-22.
- Luengo, J. M. G., B.; Sandoval, A.; Naharro, G. and Oliver, E.R. (2003). Bioplastics from microorganisms *Current Opinion in Microbiology*, 6(3), 251-260.
- M. Beaulieu, Y. B., J. Melinard, S. Pandian, and J.Goulet. (1995). Influence of ammonium salts and cane molasses on growth of *Alcaligenes eutrophus* and

- production of polyhydroxybutyrate. *Applied and Environmental Microbiology*, 61 No.1, 165-169.
- M. Venkateswar Reddy, S.V.M. (2012). Effect of substrate load and nutrients concentration on the polyhydroxyalkanoates (PHA) production using mixed consortia through wastewater treatment. *Bioresource Technology*, 114, 573-582.
- Macedonia, K. M. a. R. (2010). Experimental investigations of submerged fermentation and synthesis of pectinolytic enzymes by *Aspergillus Niger*: Effect of inoculum size and age of spores. 2(2), 40-46.
- Majid, H. U. B. A. (2007). *Screening and characterization of phaproducing bacteria from activated sludge*. Faculty of Science University Technology Malaysia.
- Marsalek, L. (2011). *Production of polyhydroxyalkanoates by Pseudomonas putida strains using alternative substrates*.
- Martin Koller, R. B., Gerhart Braunegg, Carmen Hermann, Predrag Horvat, Markus Kroutil, Julia Martinz, Jose Neto, Luis Pereira, and Paula Varila. (2005). Production of Polyhydroxyalkanoates from Agricultural Waste and Surplus Materials. *Macromolecules*, 6 No.2, 561–565.
- Mior Ahmad Khushairi Mohd Zahari, H. A., Mohd Noriznan Mokhtar, Jailani Salihon, Yoshihito Shirai, and Mohd Ali Hassan. (2012). Factors Affecting Poly(3-hydroxybutyrate) Production from Oil Palm Frond Juice by *Cupriavidus necator* *Biomedical and Biotechnology*, 8.
- Mohd Fadhil Md. Din, P. M., Zaini Ujang, Mark van Loosdrecht, Salmiati Muhd Yunus, Shreeshivadasan Chelliapan, Vasudeo Zambare, Gustaf Olsson (2012). Development of Bio-PORec\_system for polyhydroxyalkanoates (PHA) production and its storage in mixed cultures of palm oil mill effluent (POME). *Bioresource Technology*, 124, 208-216.
- Mohd Rafein Zakaria, S. A.-A., Hidayah Ariffin, Nor `Aini Abdul Rahman, Phang Lai Yee and Mohd Ali Hassan (2008). *Comamonas sp.* EB172 isolated from digester treating palm oil mill effluent as potential polyhydroxyalkanoate (PHA) producer. *African Journal of Biotechnology*, 7, 4118-4121.
- Noor Azman Mohd Johar, M. A. H., Mohd Rafein Zakaria, Phang Lai Yee, Yoshihito Shirai and Hidayah Ariffin (2012). Evaluation of Factors Affecting Polyhydroxyalkanoates Production by *Comamonas Sp.* EB172



- Using Central Composite Design. *Malaysian Journal of Microbiology*, 8, 184-190.
- Ojumu, T. V., Yu, J. and Solomon, B.O. (2004). Production of Polyhydroxyalkanoates, a bacterial biodegradable polymer. *African Journal of Biotechnology*, 3, 18-24.
- Oluwaniyi, O.O. D., O.O. (2009). Preliminary studies on the effect of processing methods on the quality of three commonly consumed marine fishes in Nigeria. *African Journal of Biotechnology*, 2, 14-20.
- Omorogbe, S. O., Ikhuoria E.U. and Igbozurike, C. C. (1999). Production of Biodiesel from Partially Refined Palm Oil *Bioresource Technology*, 70(1), 1-15.
- P.S,C.(2005). *Isolation, screening and selection of efficient poly--hydroxybutyrate (PHB) synthesizing bacteria*. Harvard university of agricultural sciences, Harvard.
- Palleroni, N. J. P. a. A. V. (1978). *Alcaligenes latus*, a new species of hydrogen-utilizing bacteria. *International Journal of Systematic and Evolutionary Microbiology*, 28 No.3, 416-424.
- Parihar, D. K. (2012). Production of lipase utilizing linseed oilcake as fermentation substrate. *International Journal of Science, Environment and Technology*, Vol.1, 135 – 143.
- Peng, Y.S. (2007) *.Studies on the Starch-Like Granules Co-Accumulated with Polyhydroxybutyrate in Spirulina platensis* University Sains Malaysia.
- Pötter, M., M. H. Madkour, F. Mayer, and A. Steinbüchel (2002). Regulation of phasin expression and polyhydroxyalkanoate (PHA) granule formation in *Ralstonia eutropha* H16. *Microbiology* 148 No.8, 2413-2426.
- Pötter, M. a. A. S. (2006). Biogenesis and structure of polyhydroxyalkanoate granules. *Microbiology Monographs*, 1, 109-136.
- Preethi.R, S. P., Aravind.J. (2012). Microbial production of polyhydroxyalkanoate (PHA) utilizing fruit waste as a substrate. *Research in Biotechnology*, 3 No.1, 61-69.
- Punrattanasin., W. (2001). *The Utilization of Activated Sludge Polyhydroxyalkanoates for the Production of Biodegradable Plastics*. Virginia Polytechnic Institute and State University.

- Rao, K. J. K., Chul-Ho and Rhee, Sang-Ki. (2000). Statistical optimization of medium for the production of recombinant hirudin from *Saccharomyces cerevisiae* using response surface methodology. *Process Biochemistry*, 35(7), Pages 639–647.
- Rehm, Q. Q. A. S. c. B. H. A. (2000). In vitro synthesis of poly (3-hydroxydecanoate): purification and enzymatic characterization of type II polyhydroxyalkanoate synthases PhaC1 and PhaC2 from *Pseudomonas aeruginosa*. *Applied Microbiology and Biotechnology*, 54(1), 37-43.
- Rob AJ Verlinden, D. J. H., Melvin A Kenward, Craig D Williams, Zofia Piotrowska Seget and Iza K Radecka (2011). Production of polyhydroxyalkanoates from waste frying oil by *Cupriavidus necator*, AMB Express. *AMB Express a Springer Open Journal*.
- Sidik Marsudi , H. U., Katsutoshi Hori (2008). Palm oil utilization for the simultaneous production of polyhydroxyalkanoates and rhamnolipids by *Pseudomonas aeruginosa*. *Applied Microbiology and Biotechnology*, 78(6), 955-961.
- Steinbüchel, A., K. Aerts, W. Babel, C. Follner, M. Liebergesell, M. H. Madkour, F. Mayer, U. Pieper-Fürst, A. Pries, H. E. Valentin, et al.,. (1995). Considerations on the structure and biochemistry of bacterial polyhydroxyalkanoic acid inclusions. *Canadian Journal of Microbiology*, 41 No.13, 94-105.
- Sudesh, K. (2010). *Polyhydroxyalkanoates from Palm Oil: Biodegradable Plastics*.
- Tabassum Mumtaz, S. A.-A., Nor'Aini Abdul Rahman, Phang Lai Yee, Yoshihito Shirai, Mohd Ali Hassan (2009). Fed-batch Production of P(3HB-co-3HV) Copolymer by *Comamonas Sp* EB 172 Using Mixed Organic Acids Under Dual Nutrient Limitation. *European Journal of Scientific Research*, 33 No.3, 374-384.
- Thi Hang Pham, J. S. W. a. B. H. A. R. (2004). The role of polyhydroxyalkanoate biosynthesis by *Pseudomonas aeruginosa* in rhamnolipid and alginate production as well as stress tolerance and biofilm formation. *Microbiology*, 150, 3405-3413.
- Venkateswar Reddy M., V. M. S. (2012). Effect of substrate load and nutrients concentration on the polyhydroxyalkanoates (PHA) production using mixed

- consortia through wastewater treatment. *Bioresource Technology*, 114, 573-582.
- Y. K. Huangetal. (2011). Screening and evaluation of polyhydroxybutyrate-producing strains from indigenous isolate *Cupriavidus taiwanensis* strains. *International Journal of Molecular Sciences*, 12 No.1, 252–265.
- Yang, Y. H.-B., C. J. - Budde, C. F. - Boccazzi, P. - Willia, L. B. - Hassan, M. A-Yusof, Z. A. M. - Rha, C. - Sinskey, A. J.(2010). Optimization of growth media components for polyhydroxyalkonoate (PHA) production from organic acids by *Ralstonia eutropha*. *Applied Microbiology and Biotechnology*, 87, 237-204.
- York, G. M., J. Lupberger, J. Tian, A. G. Lawrence, J. Stubbe, and A. J. Sinskey. (2003). *Ralstonia eutropha* H16 encodes two and possibly three intracellular Poly[d(-)-3-hydroxybutyrate] depolymerase genes. *Journal of Bacteriology*, 12, 134-142.
- Zakaria, M. R., Ariffin, H., Johar, N. A. M., Aziz, S. A., Nishida, H., Shirai, Y. and Hassan, M.A. (2010). Biosynthesis and characterization of poly (3-hydroxybutyrate-co-3-hydroxybutyrate) copolymer from wild type *Comamonas Sp.* EB172. . *Journal of Biotechnology*, 95, 1382-1386.