# THE EFFECTS OF MEDIUM MODIFICATION ON IN VITRO SEED GERMINATION AND CALLUS INDUCTION IN TOMATO

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This dissertation is dedicated to my family

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#### ABSTRACT

Tomato is one of the most important vegetable and it is part of human daily food. High world population growth rate demands to increase the tomato productivity. Tomato is chosen among the researchers due to its high anti-oxidant and mineral properties. In the present study, in vitro response of tomato c.v. Rio Grande was investigated. It was found that when sucrose was used as the source of carbon, the highest germination frequency of 85% was achieved in modified  $N_6$  whereas the highest germination rate in modified 2N6 medium was 15.6. In medium supplemented with 1.5 mg/l GA<sub>3</sub>, 1.5 mg/l BAP and 0.25 mg/l, 0.5 mg/l and 0.75 mg/l of 2, 4-D, the highest callus induction frequency of 39% was observed. In the same time, the highest induction rate of 1.3 was observed in medium supplemented with 1.5 mg/l GA<sub>3</sub>, 1.5 mg/l BAP and 0.25 and 0.5 mg/l 2, 4-D. Analysis of variance showed significant effects of medium concentration on in vitro seed germination and plant growth regulators on callus induction in different tomato explants. Cotyledons showed better response compared to hypocotyls. These results suggested that sucrose would be a better carbon source either for in vitro germination or callogenesis and cotyledon would be better explants for tissue culture studies.

#### ABSTRAK

Tomato merupakan salah satu sayur-sayuran dan merupakan antara sayuran utama dalam pemakanan seharian manusia. Kadar pertumbuhan penduduk dunia yang tinggi menuntut untuk meningkatkan produktiviti tomato. Ciri-ciri tomato yang mempunyai komponen anti oksida dan kandungan mineral yang tinggi menyebabkan menjadi pilihan bagi pengkaji-pengkaji. Kajian ini mengkaji kesan ia pengubahsuaian media kultur, pengawalan tumbesaran tumbuhan dan penggunaan sumber karbon ke atas percambahan benih dan induksi kalus secara in vitro. Sukrosa telah digunakan sebagai sumber karbon dalam kajian ini. Dalam media N6 frekuensi percambahan tertinggi ialah 85% manakala kadar percambahan bagi media terubahsuai  $2N_6$  ialah 15.98. Dalam medium yang dibekalkan dengan 1.5 mg/l GA<sub>3</sub>, 1.5 mg/l BAP dan 0.25 mg/l, 0.5 mg/l dan 0.75 mg/l 2, 4-D, frekuensi induksi kalus tertinggi ialah 39%. Padamasa yang sama, kadar percambahan tertinggi adalah sebanyak 1.3. telah didapati dalam medium yang dibekalkan dangan 1.5 mg/l GA<sub>3</sub>, 1.5 mg/l BAP, dan 0.25mg/l dan 0.5mg/l 2, 4-D. Analisis varian menunjukkan kesan ketara kepekatan medium dalam percambahan benih in vitro dengan pengawalan tumbesaran tumbuhan dalan induski kalus dalam eksplan tomato yang berbeza. Kotiledon didapati lebih responsif daripada hipokotil. Kajian membuktikan bahawa sukrosa merupakan sumber karbon yang lebih baik sama ada untuk percambahan in vitro atau kalus genesis dan kotiledon adalah explan yang lebih baik untuk kajian kultur tisu.

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### LIST OF ABBREVIATIONS / SYMBOLS

N6	-	Chu (1975)
MS	-	Murashige & Skoog (1962)
%	-	percentage
W/V	-	Weight per volume
V/V	-	Volume per volume
g	-	Gram
g/l	-	Gram per liter
°C	-	Degree celcius
et al.,	-	and friends
UV	-	Ultra violet
Var	-	Variety
cv.	-	Cultivar
mg/l	-	Mg per liter
NaOH	-	Sodium hydroxide
HCl	-	Hydrochloric acid
BAP	-	6-Benzylaminopurine
2, 4-D	-	2, 4-Dichlorophenoxyacetic acid
GA <sub>3</sub>	-	Gibberellic acid
IAA	-	Indole-3-acetic acid

NAA	-	Naphthalene acetic acid
min	-	Minute
h	-	Hour
PGR	-	Plant growth regulator

**CHAPTER 1** 

#### **INTRODUCTION**

#### **1.1** Back ground of study

Tomato belongs to the family Solanaceae by its nature of a perennial plant but is commercially cultivated as an annual crop. Tomato (*Lycopersicon esculentum* Mill.) is the second most popular vegetable crop next to potato in the world (Bhatia *et al.*, 2005). Tomato is planted in almost 4 million hectar worldwide. Lycopene is part of the pigments known as carotenoids which are natural substances that generate colors of fresh fruits and vegetables. Lycopene defends people from free radicals that degrade many parts of the body it is the most powerful anti-oxidant in the carotenoid family and it; moreover, lycopene is also known as an anti cancer (Rao and Agarwal, 2000). At present, tomato vegetables are used at a higher rate in developed countries compared to developing countries, thus, it may be referred as a luxury crop. It is grown in tropical, sub-tropical and temperate areas. It is one of the most important protective

foods as it possesses significant quantities of minerals and vitamins and sometime termed as poor person's orange (Devi *et al.*, 2008).

Plant tissue culture techniques are identified as valuable instrument in crop improvement. In tomato, *in vitro* culture is used in tomato in several biotechnological applications such as development of virus free plants (Moghaib *et al.*, 1999) and genetic transformation (Park *et al.*, 2003). Moreover, Plant tissue culture method is used for highest callus induction and improved plantlet regeneration (Amini and Ehsan pour, 2006). Plant regeneration is a key facilitator component in genetic transformations, using *Agrobacterium tumefaciens*, electroporation and particle bombardment (Amini and Ehsan pour, 2006).

*In vitro* regeneration of cultivated tomato has been a topic of research because of the commercial value of tomato plants and its potential for additional improvement via genetic manipulation (Evans, 1989). Other quality components e.g. color, acidity and flavors can be enhanced by introgression of genes (Chaudary *et al.*, 2001).

The successful application of plant tissue culture presupposes the establishment of an efficient culture system, consisting of a competent genotype, explant source as well as optimal culture conditions (Chaudary *et al.*, 2001). Different explant sources can be used for callogenesis and regeneration. Studies about the effect of variety and plant growth regulators on callus proliferation and regeneration of three tomato cultivars has been reported (Chaudhry *et al.*, 2007). Shoot apex, nodal segments and root segments were successfully used for callus induction and regeneration (Jatoi *et al.*, 2001). Various hormonal combinations are

used to induce callus and regeneration like BAP and IAA, IAA and Kin (Chen *et al.*, 1999).

### **1.2.** Problem statement

Tomato (*Lycopersicon esculentum* Mill.) is a major vegetable crop that has achieved tremendous popularity over the last century and it is grown in almost every country of the world (Abu-El-Heba *et al.*, 2008). However, Tomato production is adversely affected by wide ranges of biotic and abiotic stresses (Osman *et al.*, 2010) such as disease, high temperature, draught, salinity and its vulnerability to frequent insect and pest attacks. Diseases infestations are notorious factors that reduce crop yields and inflate production costs (Ishag *et al.*, 2009). These diseases include viral, bacterial and fungal (Chaudhry *et al.*, 2010).

To attain sustainable tomato productions, such above mentioned constraints have been addressed by conventional breeding and enhanced management but it has resulted in limited commercial success (Osman *et al.*, 2010). Plus, the improvement of plant through conventional breeding methods is slow, time-consuming and labor-intensive (Moghaieb *et al.*, 1999). The integration of modern biotechnology like tissue culture into breeding programs may provide powerful tools to overcome these limitations.

#### 1.3. Objectives of study

The objectives of this study are:

- 1. To investigate the effects of different strength of modified  $N_6$  medium namely  $1/2 N_6$ ,  $N_6$  and  $2N_6$  on *in vitro* seed germination.
- 2. To assess the effects of carbon source and plant growth regulators on callus induction from cotyledon in tomato.
- 3. To study the effects of plant growth regulator and carbon source on callus induction from hypocotyl in tomato.

### 1.4. Aim of study

The aim of this work was to produce information regarding the effects of different strength of modified  $N_6$  medium on *in vitro* seed germination in tomato. Moreover, the present study was conducted to explore the callogenic potential of hypocotyls and cotyledon segments so as to establish a reproducible protocol for callus induction from tomato (*Lycopersicon esculentum* Mill.) cultivar *Rio Grande* by using different sources of carbon and different combination and concentration of plant growth regulators.

#### REFERENCES

- Abdel-Raheem, A.T., Ragab, A.R., Kasem, Z.A., Omar, F.D., and Samera, AM. (2007). *In vitro* selection for tomato plants for drought tolerance via callus culture under polyethylene glycol (PEG) and mannitol treatments. *African Crop Science Conference*. El-Minia, Egypt, 2027-2032.
- Abu-El-Heba, G.A., Hussein, G.M., and Abdalla, N.A. (2008). A rapid and efficient tomato regeneration and transformation system. *Agriculture and Forestry Research*, 58(1/2), 103-110.
- Afrasiab, H., and Jafar, R. (2011). Effect of different media and solidifying agent on callogenesis and plant regeneration from different explants of rice (*Oryza sativa L*) varieties *Super basmati* and *IRRI-6. Pakistan Journal of Botany*, 43(1), 487-501.
- Afroz, A., Chaudhry, Z., Khan, R., Rashid, H., and Khan, S. (2009). Effect of GA<sub>3</sub> on regeneration response of three tomato cultivars (*Lycopersicon esculentum*). Pakistan Journal of Botany, 41(1), 143-151.
- Ahmed, E. E., Bisztray, G., and Velich, I. (2002). Plant regeneration from seedling explants of common bean (*Phaseolus vulgaris L.*). Acta Biologica Szegediensis, 46(3-4), 27-28.
- Amini, F., and Ehsanpour, A. (2006). Response of tomato (Lycopersicon esculentum Mill.) cultivars to MS, water agar and salt stress in in

vitro culture. Pakistan Journal of Biological Sciences, 9(1), 170-175.

- Arditti, J. (1984). Physiology of germinating orchid seeds. Orchid Biology: Reviews and Perspectives III (pp. 177 – 222). New York: Cornell University.
- Ashok Kumar, H.G., and Murthy, H.N. (2004). Effect of sugars and amino acids on androgenesis of *Cucumis sativus*. *Plant Cell, Tissue and Organ Culture*, 78(3), 201-208.
- Azria, D., and Bhalla, P. L. (2000). Plant regeneration from mature embryo derived callus of Australian rice (*Oryza sativa L.*) varieties. *Crop* and Pasture Science, 51(2), 305-312.
- Bhatia, P., Ashwath, N., and Midmore, D. J. (2005). Effects of genotype, explant orientation, and wounding on shoot regeneration in tomato. *In Vitro Cellular and Developmental Biology Plant*, 41(4), 457-464.
- Bhatia, P., Ashwath, N., Senaratna, T., and Midmore, D. (2004). Tissue culture studies of tomato (*Lycopersicon esculentum*). *Plant Cell*, *Tissue and Organ Culture*, 78(1), 1-21.
- Bhattacharya, J., and Khuspe, SS. (2001). *In vitro* and *in vivo* germination of papaya (*Carica papaya L.*) *seeds. Scientia Horticulturae*, *91*(1), 39-49.
- Bewley, J.D. (1997). Seed germination and dormancy. *Plant Cell*, 9(7), 1055–1066.
- Bewley, J.D., and Black, M. (1994). *Seeds: Physiology of Development and Germination*. (2<sup>nd</sup> ed) New York, Plenum Press.

- Biswas, M. K., Roy, U. K., Islam, R., and Hossain, M. (2010). Callus culture from leaf blade, nodal, and runner segments of three strawberry (*Fragaria sp.*) clones. *Turk Journal Biology*, 34, 75-80.
- Bürün, B., and Şahin, O. (2009). In vitro and in vivo germination of Cyclamen alpinum seeds. Turk Journal of Botany, 33, 277-283.
- Capote Rodríguez, A., Fundora Mayor, Z., and Pérez Díaz, O. (2000). Effect of different factors on the *in vitro* plant regeneration from leaflets of five genotypes of tomato (*Lycopersicon esculentum* Mill.). *Revista del Jardín Botánico Nacional*, 21(1), 71-76.
- Chandel, G., and Katiyar, S. (2000). Organogenesis and somatic embryogenesis in tomato (*Lycopersicon esculantum* Mill.). *Advances in Plant Sciences*, 13(1), 11-17.
- Chaudary, Z., Feroz, I., Ahmed, W., Rashid, H., Mirza, B., and Quraishi, A. (2001). Varietal response of *Lycopersicon esculentum* L. to callogenesis and regeneration. *Journal of Biological Sciences*, *1*(12), 1138-1140.
- Chaudhry, Z., Afroz, A., and Rashid, H. (2007). Effect of variety and plant growth regulators on callus proliferation and regeneration response of three tomato cultivars (*Lycopersicon esculentum* Mill.). *Pakistan Journal of Botany*, 39(3), 857-869.
- Chaudhry, Z., Abbas, S., Yasmin, A, Rashid, H, Ahmed, H., and Anjum, M.A. (2010). Tissue culture studies in tomato (*Lycopersicon* esculentum Mill.) Var. Money Maker. Pakistan Journal of Botany, 42(1), 155-163.
- Chen, H., Zhang, J., and Zhuang, T. (1999). Studies on optimum hormone levels for tomato plant regeneration from hypocotyl explants cultured *in vitro*. *Shanghai Nongye Xuebao*, 15(2), 26-29.

- Chu, C.C. (1978). The N<sub>6</sub> medium and its applications to anther culture of cereal crops. Symposium on Plant Tissue Culture. May 25-30, Beijing, 43-50.
- Compton, M. E., and Veilleux, R. E. (1991). Shoot, root and flower morphogenesis on tomato inflorescence explants. *Plant Cell, Tissue and Organ Culture, 24*(3), 223-231.
- Conger, B., Novak, F., Afza, R., and Erdelsky, K. (1987). Somatic embryogenesis from cultured leaf segments of Zea mays. Plant Cell Reports, 6(5), 345-347.
- De Klerk, G. J., Hanecakova, J., and Jásik, J. (2008). Effect of medium pH and MES on adventitious root formation from stem disks of apple. *Plant Cell, Tissue and Organ Culture, 95*(3), 285-292.
- Devi, R., Dhaliwal, M., Kaur, A. and Gosal, S. (2008). Effect of growth regulators on *in vitro* morphogenic response of tomato. *Indian Journal of Biotechnology*, 7, 526-530.
- Ehsanpour, A., and Amini. F. (2005). Callus production and plant regeneration of two tomato (*Lycopersicon esculentum* Mill.) cultivars. 4th National Biotechnology Congress of Iran, Kerman. 23-27.
- Evans, D. A. (1989). Somaclonal variation genetic basis and breeding applications. *Trends in Genetics*, 7, 46-50.
- George, E. F., Hall, M. A., and De Klerk, G. J. (1986) *Plant Propagation by Tissue Culture* (3<sup>rd</sup> ed). Institute of Biological Sciences: University of Wales, UK.
- George, E. F., Hall, M. A., and Klerk, G. J. D. (2008a). The components of plant tissue culture media I: macro-and micro-nutrients. *Plant Propagation by Tissue Culture* (pp. 65-113). Netherland: Springer.

- George, E. F., Hall, M. A. and Klerk, G. J. D. (2008b). The components of plant tissue culture media II: organic additions, osmotic and pH effects, and support systems. *Plant Propagation by Tissue Culture* (pp. 115-173). Netherland: Springer.
- George, F. E. and Sherrington, P. D. (1984). Plant Propagation by Tissue Culture: Handbook and Directory of Commercial Laborations: Exegetics Ltd: England.
- Gubis, J., Lajchova, Z., Faragoand, J., Jurekova, Z. (2003). Effect of genotype and explant type on shoot regeneration in tomato (*Lycopersicon esculentum* Mill.) in vitro. Czech Journal of Genetics and Plant Breeding, 39(1): 9-14.
- Gubis, J., Lajchova, Z., Klcova, L., and Jurekova, Z. (2005). Influence of growth regulators on plant regeneration in tomato. *Hort Science*, 32(3): 118–122.
- Guillermo, P., L.N. Canepa, R., Zorzoli and Gulshan, T., and Sharma, D. (1981). Studies on anther cultures of tomato (*Lycopersicon esculentum* Mill.) *Biologia Plantarum*, 23(6), 414-420.
- Harish, M.C., Rajeevkumar, S., and Sathishkumar, R. (2010). Efficient *in vitro* callus induction and regeneration of different tomato cultivars of India. *Asian Journal of Biotechnology*, 2(3), 178-184.
- Hille, J., Koornneef, M., Ramanna, M., and Zabel, P. (1989). Tomato: a crop species amenable to improvement by cellular and molecular methods. *Euphytica*, 42(1), 1-23.
- Ishag, S., Osman, M.G., and Khalafalla, M.M. (2009). Effects of growth regulators, explant and genotype on shoot regeneration in tomato (Lycopersicon esculentum c.v. Omdurman). International Journal of Sustainable Crop Production, 4(6), 7-13.

- Jabeen, N., Chaudhry, Z., Rashid, H., and Mirza, B. (2005). Effect of genotype and explant type on *in vitro* shoot regeneration of tomato (*Lycopersicon esculentum* Mill.). *Pakistan Journal of Botany*, 37(4), 899.
- JaeBok, P., B.Y., Yi and C.K., Lee. (2001). Effects of plant growth regulators, bud length, donor plant age, low temperature treatment and glucose concentration on callus induction and plant regeneration in anther culture of cherry tomato 'Mini-carol'. *Journal of the Korean Society for Horticulture Science*, 4 (1), 32-37.
- Jatoi, S. A., Sajid, G. M., Sappal, H. U., Baloch, M. S., Quraishi, A., and Anwar, R. (2001). Differential *in vitro* response of tomato hybrids against a multitude of hormonal regimes. *On Line Journal of Biological Sciences*, 1(12), 1141-1144.
- Javed, M. A, Misoo, S., T Mahmood, M.S.H., Shah, Ah, Rahid, V. N., and Iqbal, J. (2007). Effectiveness of alternate culture temperatures and maltose in the anther culture of salt tolerant indica rice cultivars. *African Crop Science Society Conference*, October 2007. El-Minia, Egypt, 27-31.
- Johnson, T.R., Kane, M.E., and Pérez, H.E. (2011). Examining the interaction of light, nutrients and carbohydrates on seed germination and early seedling development of *Bletia purpurea* (Orchidaceae). *Plant Growth Regulation*, 63(1), 89-99.
- Kamo, K.K., Chang, K.L., Lynn, M.E., and Hodges, T.K. (1987). Embryogenic callus formation from maize protoplasts. *Planta*, *172*(2), 245-251.
- Karsai, I., Bedo, Z., and Hayes, PM. (1994). Effect of induction medium pH and maltose concentration on *in vitro* androgenesis of hexaploid

winter triticale and wheat. *Plant Cell, Tissue and Organ Culture,* 39(1), 49-53.

- Khan, M.S., Usman, M., and Lilla, M.I. (2006). Facile plant regeneration from tomato leaves induced with spectinomycin. *Pakistan Journal* of Botany, 38(4), 947-952.
- Kumaria, S., and Tandon, p., (2010). Asymbiotic germination of Dendrobium fimbriatuin var. Oculatum Hk. f. seeds on different media. Indian National Science Academy, 57(3/4), 227-229.
- Kunitake, H., Imamizo, H., and Mii, M. (1993). Somatic embryogenesis and plant regeneration from immature seed-derived calli of *Rugosa rose* (*Rosa Rugosa Thunb*). *Plant Science*, *90*(2), 187-194.
- Last, D., and Brettell, R. I. (1990). Embryo yield in anther culture is influenced by the choice of sugar in the culture medium. *Plant Cell Reports*, 9:14 16.
- Laxminarasu, M., Swamy, N.R., Godishala, V., Kairamkonda M., and Srikanth. K. (2012). Zeatin induced direct multiple shoots development and plant regeneration from cotyledon explants of cultivated tomato (*Solanum lycopersicum L.*). *Australian Journal* of Crop Science, 6(1), 31-35.
- Leifert, C., Murphy, K.P., and Lumsden, P.J. (1995). Mineral and carbohydrate nutrition of plant cell and tissue cultures. *Critical Reviews in Plant Sciences*, 14(2), 83-109.
- Locy, R. D. (1995). Selection of tomato tissue cultures able to grow on ribose as the sole carbon source. *Plant Cell Reports*, *14*(12), 777-780.
- Mamidala, P., and Nanna, R.S. (2009). Influence of antibiotics on regeneration efficiency in tomato. *Plant Omics*, 2(4), 135-139.

- Mamidala, P., and Nanna, R.S. (2011). Effect of genotype, explant source and medium on *in vitro* regeneration of tomato. *International Journal of Genetics and Molecular Biology*, *3*(3), 45-50.
- Mandal, N., and Gupta, S. (1997). Anther culture of an interspecific rice hybrid and selection of fine grain type with submergence tolerance. *Plant Cell, Tissue and Organ Culture, 51*(1), 79-82.
- Marschner, H. (1995). *Mineral Nutrition of Higher Plants*. (2<sup>nd</sup> ed). Academic Press: London.
- Mendoza, M.G., and Kaeppler, H.F. (2002). Auxin and sugar effects on callus induction and plant regeneration frequencies from mature embryos of wheat (*Triticum aestivum L.*). In Vitro Cellular and Developmental Biology Plant, 38(1), 39-45.
- Moghaieb, R. E. A., Saneoka, H., and Fujita, K. (1999). Plant regeneration from hypocotyl and cotyledon explant of tomato (*Lycopersicon esculentum* Mill.). *Soil Science and Plant Nutrition*, 45(3), 639-646.
- Mohamed, A.N., Ismail, M.R., and Rahman, M.H. (2010). In vitro response from cotyledon and hypocotyls explants in tomato by inducing 6benzylaminopurine. African Journal of Biotechnology, 9(30), 4802-4807.
- Molnar, S.J. (1988). Nutrient modifications for improved growth of Brassica nigra cell suspension cultures. Plant cell, Tissue and Organ Culture, 15(3), 257-267.
- Murthy, B.N.S., Murch, S.J., and Saxena, P.K. (1998). Thidiazuron: A potent regulator of *in vitro* plant morphogenesis. In Vitro Cellular and Developmental Biology Plant, 34(4), 267-275.

- Nikam, T.D., and Shitole, M.G. (1998). *In vitro* culture of *Safflower L.* c.v. *Bhima*: initiation, growth optimization and organogenesis. *Plant Cell, Tissue and Organ Culture,* 55(1), 15-22.
- Negi R.S., and Sharma, K.C. (2012). Seed germination in medicinally important plant *Cassai Auriculatia*. *Online International Journal*, *1* (1), 12-13.
- Orsinky, B. L., McGregor, G. I., Johnson, G. I., Kartha, K. K. (1990). Improved embryoid induction and green shoot regeneration from wheat anther cultures with medium with maltose. *Plant Cell Reports*. 9:365 – 369.
- Osman, M. G., Elhadi, E. A., and Khalafalla, M. M. (2010). Callus formation and organogenesis of tomato (*Lycopersicon esculentum* Mill, c.v. *Omdurman*) induced by thidiazuron. *African Journal of Biotechnology*, 9(28), 4407-4413.
- Park, S. H., Morris, J. L., Park, J. E., Hirschi, K. D., and Smith, R. H. (2003). Efficient and genotype independent Agrobacterium mediated tomato transformation. Journal of Plant Physiology, 160(10), 1253-1257.
- Paul S., Kumaria S., and Tandon P., (2010). An effective nutrient medium for asymbiotic seed germination and large-scale *in vitro* regeneration of *Dendrobium hookerianum*, a threatened orchid of northeast India. *Aob Plants. Oxford Journals*, 1-7.
- Ramage, C. M., and Williams, R. R. (2002). Mineral nutrition and plant morphogenesis. In vitro Cellular and Developmental Biology Plant, 38(2), 116-124.
- Raj, S.K., R. Singh, S.K., Pandey and B.P., Singh. (2005). Agrobacterium mediated tomato transformation and regeneration of transgenic lines expressing tomato leaf curl virus coat protein gene for resistance against TLCV infection. *Current Science*, 88(10): 1674-1679.

- Rao, A. V., and Agarwal, S. (2000). Role of antioxidant lycopene in cancer and heart disease. *Journal of the American College of Nutrition*, 19(5), 563-569.
- Ramesh, M., Murugiah, V., and Gupta, A.K. (2009). Efficient *in vitro* plant regeneration via leaf base segments of indica rice (*Oryza sativa* L.). Indian Journal of Experimental Biology, 47(1):68-74.
- Rashid, R., and Bal, S.S. (2010). Effect of hormones on direct shoot regeneration in hypocotyl explants of tomato. *Notulae Scientia Biologicae*, 2(1), 70-73.
- Rzepka-Plevneš, D., Kulpa, D., Grabiec, M., Kowalczys, K., and Kurek, J. (2006). The effect of growth regulators and culture conditions on the callus induction in tomato *Lycopersicon sp. Acta Scientiarum Polonorum. Hortorum Cultus*, 5(2), 23-34.
- Shadang, R., Dwivedi, P., Hegde, SN, and Ahmed, N. (2007). Effects of different culture media on seed germination and subsequent *in vitro* development of protocorms of *Hygrochilus parishii* (*Veith and Rchb. f.*) *Pfitz* (Orchidaceae). *Indian Journal of Biotechnology*, 6(2), 256.
- Shakti Prosad, P., Alam, I., Anisuzzaman, M., Sarker, K.K., Sharmin, S.A., and Alam, M.F. (2007). Indirect organogenesis in summer squash (*Cucurbita pepo L.*). *Turk Journal of Agriculture*, 31, 63-70.
- Sharma, DK., Chaudhary, DR., and Verma, TS. (2001). Growth and seed yield of tomato (*Lycopersicon esculentum* Mill.) c.v. *Roma* as influenced by levels of nitrogen and plant spacing. *Haryana Journal of Horticultural Sciences*, 30(1/2), 95-96.
- Sheeja, T.E., and Mandal, A.B. (2003). In vitro flowering and fruiting in tomato (Lycopersicon esculentum). Asia Pacific Journal of Molecular Biology and Biotechnology, 11(1): 37-42.

- Slater, A., Scott, N., and Fowler, M. (2003). *Plant Biotechnology*. (2<sup>nd</sup> ed) Oxford University Press: UK.
- Takahashi, N. (1986). Chemistry of Plant Hormones CRC Press: Florida: USA.
- Thodsaporn, P., Sumontip, B., Piyada, T., and Manit, K. (2004). Transformation of indica rice (*Oryza sativa L.*) c.v. *RD*<sub>6</sub> mediated by *Agrobacterium Tumefaciens*. Songklanakarin Journal Science of Technology, 26(1), 1-13.
- Uddin, M. F. (2004). Effect of variety and plant growth regulators in MS medium on callus proliferation from virus infected tomato plant. *Journal of Science Nature*, 2(1), 1-6.
- Venkatachalam, P., Geetha, N., Priya, P., Rajaseger, G., and Jayabalan, N. (1998). High frequency plantlet regeneration from hypocotyl explants of tomato (*Lycopersicon esculentum* Mill.) via organogenesis. *Plant Cell Biotechnology and Molecular Biology*, 1(3/4), 95-100.
- Williams, RR. (1995). Towards a Model of Mineral Nutrition In Vitro. Transplant Production Systems (pp. 213-229). Springer: Netherlands.
- Xie, J., Gao, Cai, Q., Cheng, X., Shen, Y., and Liang, Z. (1995). Improved isolated microspore culture efficiency in medium with maltose and optimized growth regulator combination in japonica rice (*Oryza* sativa). Plant Cell, Tissue and Organ Culture, 42(3), 245-250.
- Zelcer, A., Soferman, O., and Izhar, S. (1984). An *in vitro* screening for tomato genotypes exhibiting efficient shoot regeneration. *Journal* of Plant Physiology, 115(3), 211-215.

Zhang, W., Hou, L., Zhao, H., and Li, M. (2012). Factors affecting regeneration of tomato cotyledons. *Bioscience Methods*, 8(5), 913-919.