DECOLORIZATION OF PALM OIL MILL EFFLUENT USING SELECTED INDIGENOUS BACTERIA

NURUL ASMAK BINTI MD LAZIM

A dissertation submitted in partial fulfillment of the requirement for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

JANUARY 2013

To Allah S.W.T

...Your Mercy has been always surpasses all understanding...

To my parents and family

...Your hopes and wishes has been the greatest poem in my life...

To my lecturer

...Your guardian has made all these become possible...

To my friends

...Thanks a lot...

ACKNOWLEDGEMENTS

Special thanks to my supervisor, Prof.Madya Dr. Zaharah Ibrahim for her guidance within stimulating ideas and encouragement all the time during final year project.

Besides, I would like to express my gratitude to all laboratory staff and fellow course mate as well as postgraduate seniors for their help.

Last but not least, I would like to show my thankfulness to my beloved parents and family for their loving support along the way in finishing my study.

Thank you very much.

ABSTRACT

In this study, the selection of suitable bacteria that is best to treat POME from final pond is the main objective. There are 20 bacteria that had been isolated from final pond of POME but only 11 species has the ability to reduce color of POME. However, only five strains out of 11 species were capable to reduce the color after 6 days of incubation compared to others which took longer time. Bacteria D showed the best results in reducing the color of POME which was 45% (from 4437 to 2448 ADMI) after 6 days of treatment. Partial identification of bacteria D using biochemical test showed that it belonged to *Proteus inconstans*. Results showed that POME wastewater treated with indigenous bacteria D successfully reduced the COD level up to 61 % (from 34170 to 13464 mg/L), pH value from 9.25 to 9.11 at day 2, 19% of lignin contents (from 310 to 250 mg/L), 17% of phenolics compound (from 271 to 225 mg/L), 47% (from 5202 to 2754 ADMI) of color intensity (ADMI) respectively. The highest lignin peroxidase activity was 1.94 U/mL implying the optimum bacterial activity occurred during the exponential growth phase. The study suggested that isolated indigenous bacteria could decolorize POME wastewater.

ABSTRAK

Dalam kajian ini, pemilihan bakteria yang sesuai yang terbaik untuk merawat POME dari kolam akhir adalah objektif utama. Terdapat 20 bakteria yang telah diasingkan daripada kolam akhir POME tetapi hanya 11 spesis yang berupaya untuk mengurangkan warna POME. Walau bagaimanapun, hanya lima jenis daripada 11 spesis mampu mengurangkan warna selepas 6 hari perawatan berbanding dengan lainlain yang mengambil masa yang panjang. Bakteria D menunjukkan hasil yang terbaik dalam mengurangkan warna POME yang merupakan 45% (4437-2448 ADMI) selepas 6 hari rawatan. Separa pengenalan bakteria D menggunakan ujian biokimia menunjukkan bahawa ia kepunyaan Proteus inconstans. Keputusan menunjukkan bahawa air sisa POME dirawat dengan bakteria D berjaya mengurangkan tahap COD sehingga kepada 61% (dari 34170 ke 13464 mg / L), nilai pH dari 9.25 ke 9.11 pada hari ke2, 19% kandungan lignin (dari 310 ke 250 mg / L), 17% komponen fenol (dari 271 ke 225 mg / L), 47% (dari 5202 ke 2754 ADMI) kepekatan warna (ADMI). Lignin peroxidas aktiviti yang paling tinggi adalah 1.94 U / mL yang memantulkan peringkat optimum bakteria aktiviti yang merupakan fasa eksponen pertumbuhan bakteria. Hasil kajian mencadangkan bahawa bakteria asli boleh mengurangkan warna air sisa POME.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xi
	LIST OF FIGURES	xii
	LIST OF ABBREVIATIONS	xiii
	LIST OF SYMBOLS	xiv
	LIST OF APPENDICS	XV
1	INTRODUCTION	1
	1.1 Research background	1
	1.2 Problem statement	3
	1.3 Objectives of the research	4
	1.4 Research significance	4
	1.5 Scope of the research	5

2	LITE	ERATUI	RE REVIEW	6	
	2.1	Produ	ction of palm oil wastes	6	
	2.2	Micro	Microorganisms in POME treatment		
	2.3	Identi	fication of microbes	9	
		2.3.1	Morphological characteristic	10	
			2.3.1.1 Gram staining	10	
			2.3.1.2 Biochemical test	11	
	2.4	Analy	tical review for POME treatment	14	
		2.4.1	Water quality parameter	14	
			2.4.1.1 pH	14	
			2.4.1.2 Chemical Oxygen Demand (COD)	15	
		2.4.2	Total phenolics compounds	15	
		2.4.3	Determination of lignin content	15	
		2.4.4	Color intensity (ADMI)	17	
		2.4.5	Enzyme activity (lignin peroxidase)	18	
3	RES	EARCH	I METHODOLOGY	20	
	3.1	Sample collection			
	3.2	Prepar	ration of medium	20	
		3.2.1	Kraft lignin medium	20	
		3.2.2	POME medium	21	
	3.3	Cultur	re and cultivation	21	
		3.3.1	Isolation of selected bacteria from		
			POME wastewater	21	

viii

		3.3.1.1 Spread plate	22
		3.3.1.2 Streak plate	22
3.4	Stock	culture	22
3.5	Screen	ning for the best microbes	23
3.6	Micro	bial partial identification	23
	3.6.1	Colony morphology characteristic	23
		3.6.1.1 Gram staining	23
		3.6.1.2 Biochemical test	24
3.7	Analy	tical method for POME treatment	24
	3.7.1	Preliminary screening for isolation of selected	
		bacteria	24
	3.7.2	рН	25
	3.7.3	Chemical oxygen demand (COD)	25
	3.7.4	Determination of total phenolic	
		compounds	26
	3.7.5	Determination of lignin compounds	26
	3.7.6	Determination of color intensity	
		(ADMI)	27
	3.7.7	Enzyme activity (lignin peroxidase)	27
3.8	Overv	iew experiment	28
RESU	JLTS A	ND DISCUSSION	29
4.1	Isolati	on of the best microbes for POME treatment	29
4.2	Growt	th profile of selected bacteria	31

4

ix

	4.3	Partial	l identification of bacteria	34
		4.3.1	Morphological characteristics and	
			Gram staining	34
		4.3.2	Biochemical test	34
		4.3.3	Partial identification of bacteria D	35
	4.4	Analy	ses of pH, Chemical oxygen demand (COD),	
		total p	henolic compounds, lignin content, enzyme	
		activit	y (lignin peroxidase)and color intensity	37
		4.4.1	pH changes	37
		4.4.2	Chemical oxygen demand (COD)	38
		4.4.3	Determination of total phenolics	
			compound	39
		4.4.4	Determination of lignin content during POME	
			treatment	40
		4.4.5	The enzyme activity (lignin peroxidase) of	
			bacteria D	41
		4.4.6	Profile of color intensity (ADMI) after six	
			days treatment	43
	CON	CLUSI	ON	45
	5.1	Concl	usion	45
	5.2	Recon	nmendation	46
REFERENCES		47		
	Appendices A-C 5		51	

5

х

LIST OF TABLES

TABLE NO.	TITLE	PAGE	
2.1	2.1 Characterization of raw POME and the regulatory		
	discharge limits (Parveen et al., 2010)		
3.1	3.1 COD reagents preparation		
4.1	The summary of color removal for preliminary screening for the best bacteria in treatment of POME	30	
4.2	Colony morphology and Gram staining of bacteria D	34	
4.3	Results of biochemical tests	35	
4.4	The result analysis after POME treatment	44	

LIST OF FIGURES

FIGURE NO	. TITLE	PAGE
2.1	The model structure of lignin (Wershaw et al., 2005)	16
2.2	The model structure of biodegradation of lignin pathway (Fuhrmann <i>et al.</i> , 2005)	17
3.1	The overall experiment	28
4.1	Growth profiles of bacteria D	32
4.2	Results for bacteria D growth curve	33
4.3	OD reading versus Cell dry weight	33
4.4	Partial identification of bacteria D	36
4.5	pH changes after applying pure culture of bacteria D	37
4.6	COD reduction of POME during wastewater treatment	38
4.7	Profile of total phenolics compound during treatment	39
4.8	Profile of lignin concentration versus LiP activity during	41
	treatment	41
4.9	Profile of LiP activity versus OD reading	42
4.10	The color reduction during 6 days treatment	43

LIST OF ABBREVIATIONS

ADMI	American Dye Manufacturers Institute
BOD	Biochemical Oxygen Demand
COD	Chemical Oxygen Demand
NB	Nutrient Broth
WQI	Water Quality Analysis Index
TSI	Triple Sugar Iron
MR-VP	Methyl Red- Voges Proskauer
O-F	Oxidation- Fermentation
H_2O_2	Hydrogen Peroxide
$K_2Cr_2O_7$	Potassium dichromate
CaCI ₂	Calcium chloride
MgSO ₄	Magnesium sulphate
KH ₂ PO ₄	Potassium dihydrogen phosphate
NaOH	Sodium hydroxide
(NH ₄) ₃ SO ₄	Ammonium sulphate
AgSO ₄	Silver sulphate
HgSO ₄	Mercury sulphate
H_2SO_4	Sulphuric acid

LIST OF SYMBOLS

L	Liter
mL	Milliliter
g/L	Gram per liter
et al.	And friends
sp.	Species
v/v	volume over volume

LIST OF APPENDICES

APPENDICES	TITLE	PAGE
Α	Biochemical test	51
В	Follin Ciocalteau Gallic Acid Standard	
	Curve	56
С	Lignin Standard Curve	57

CHAPTER 1

INTRODUCTION

1.1 Research background

Palm oil is one of the most important vegetable oil in the world (Hartley, 1988). Neoh *et al* (2012) has reported that the second world largest palm oil producer after Indonesia was Malaysia; with the amount of 10.6 million tonnes of palm oil production as far back as in year 1999 and this amount has becomes increasing to 17.7 million tonnes in year 2008 and expected to rise every year. The increasing of the palm oil production generally contributed to increase the different kind of waste such as solid waste; in example is empty fruit bunch and liquid waste like POME (Igwe and Onyegbado, 2007).

Palm Oil Mill Effluent also known as POME is an effluent that is large in quantity resulting from the process of extracting the crude palm oil from the fresh fruit bunch (Ahmad *et al.*, 2003). It consists of colloidal suspension of 95-96% water, 0.6-0.7% oil, and yet, 4-5% total solids including 2-4% suspended solids originating from the mixture of a sterilizer condensate, separator sludge and hydrocyclone wastewater (Ma, 2000; Ahmad *et al.*, 2003)

Studies have reported that POME also consists of soluble materials such as CH_4 , SO_2 , NH_3 , halogens or soluble liquids or solids which contain ions of either organic or inorganic with their concentration were over the standard limit of threshold value (Igwe and Onyegbado, 2007). Consequently, the discharged of POME directly to nature will harm aquatic life, also the environment.

Even though it is not highly polluted, the wastewater is harmful if directly released to the environment due to its high chemical oxygen demand (COD), biochemical oxygen demand (BOD), phenols and color concentration. as well as unpleasant odors (Poh *et al.*, 2009). There is no chemical addition during palm oil mill processing. However, the appeared color during discharged of POME was derived from plant constituents such as lignin and phenolic compounds.

Recent study showed that phenolics (tannic and humic acids) from the feedstock and melanoidins from Maillard reaction of sugars (carbohydrates) with proteins (amino groups) might contribute to the color of the sugar processing effluent. Consequently, this sugar processing effluent was then mixed together with other wastes and discharged through POME (Zahrim *et al.*, 2009).

The discharge of color effluents directly to water may affect aquatic life as it will reduce the penetration of sunlight that will inhibit the photosynthetic activity inside water (Mohan and Karthikeyan, 1997; Neoh *et al.*, 2012). Yet, the colored compounds from POME may chelate with metals ions inside water and result in toxicity to aquatic organisms as reported by Mohan and Karthikeyan (1997) and Neoh *et al* (2012). Indeed, the presence of humic compounds will react with chlorine may produce carcinogenic byproducts like trihalomethanes (Vukovic *et al.*, 2008; Neoh *et al.*, 2012). Other than that, studies have reported that the compounds that were derived from lignin might inhibit the development of embryo of aquatic organisms (Pillai *et al.*, 1997; Neoh *et al.*, 2012).

Since conducting an experiment based on water contamination with palm oil mill effluent (POME) has recently attracted much attention, decolorization of palm oil mill effluent using selected indigenous bacteria had been carried out in this study. The dominant bacteria for POME color removal were more focused and partially identified after Gram staining and biochemical test were performed and these bacteria were then applied to POME waste to assess their effectiveness in treating POME wastewater.

1.2 Problem statement

As mentioned earlier, huge quantities of waste such as empty fruit bunch and POME are produced in the palm oil mill industry. There was chemical and physical methods that were applied to treat POME wastewater included adsorption, coagulation-flocculation, oxidation and electrochemical method. However, these methods are highly expensive, formation of hazardous byproducts or intensive energy requirements (Couto, 2009; Chanida and Poonsuk, 2011).

Another alternative way is by using biological treatment. The biological method is more effective in treatment and it sufficiently needs to apply due to cost-effective and environmental friendly (Zahrim *et al.*, 2009).

Previous study had shown that the use of white-rot fungi were capable to decolorize and phenol removal from wastewaters (Kissi *et al.*, 2001). But, Akin and Benner have reported that the anaerobe rumen bacteria could solubilize lignin better than the rumen fungi (Hiroshi *et al.*, 2000). In addition, there is biological treatment that utilized aerobic and anaerobic; results showed that the aerobic treatment was more efficient compared to anaerobic (Anggelis *et al.*, 2001). Due to ability to degrade organic material as reported by Epstein (1997) and Ipek *et al* (2002), aerobic microorganisms were applied in this study.

1.3 Objectives of the research

The objectives of this study were:

- i. To isolate and partial characterize the selected indigenous bacteria present in palm oil mill effluent that had roles in POME treatment.
- ii. To investigate the effectiveness of isolated microorganisms in treating POME such as chemical oxygen demand (COD), pH, total phenolic compounds and determining lignin content.
- iii. To examine POME color removal (ADMI unit) after selected bacteria were applied

1.4 Research significance

The importance of this study was to isolate and characterize the selected bacteria from POME that had roles in POME treatment. This is very important to isolate indigenous microorganism with specific enzyme that can be used for the treatment. The effectiveness of isolated microorganisms in treating POME such as chemical oxygen demand (COD), pH, total phenolic compounds and lignin were determined. These tests were used to compare between treated and untreated sample after the selected bacteria were applied. This is very important to analyze the effectiveness of isolated bacteria in treating POME. The study was then followed by analyzing the color reduction of POME in order to solve unwanted color present by using effective microorganisms.

REFERENCES

- Agamuthu, P. (1995). *Palm oil mill effluent and utilization*. Waste treatment plant. New Delhi: Narosa Publishing House. 338-360
- Aggelis, G.G. Gavala, H. N. and Lyberatos, G. (2001). Combined and separate aerobic and anaerobic bio-treatment of green olive debittering wastewater. *Agricultural Engineering Research* (3). 283–292
- Ahmad, A. L Ismail, S. and Bathia, S. (2003). Water recycling from palm oil mill effluent (pome) using membrane technology. *Desalination*. (157). 87-95
- Akin, D. E. and Benner, R. (1988). Degradation of polysaccharides and lignin by ruminal bacteria and fungi. *Application Environment Microbiology*. (54). 1117-1125
- Chandra, R. Raj, A. Purohit, H. J. and Kapley, A. (2007). Characterisation and optimisation of three potential aerobic bacterial strains for kraft lignin degradation from pulp paper waste. *Chemosphere*. (67). 839–846
- Couto, S. R. (2009). Dye removal by immobilized fungi. *Biotechnology advances*. (27). 227-235
- del Río, J.C. Martínez, Á. T. and Gutiérrez, A. (2007). Presence of 5-hydroxyguaiacyl units as native lignin constituents in plants as seen by py-GC/MS. *Analytical Application Pyrolysis.* (79). 33–38

- Epstein, E. (1997). The science of compositing. Technomic Publishing Co. Inc., Lancaster
- Frederick, S. A. (1992). A New Assay for Lignin-Type Peroxidases Employing the Dye Azure B. *Applied and environmental microbiology*. Sept. 1992. 3110-3116
- Igberaharha, L. O. (1998). Zero discharge layout of palm oil mill effluent (POME). Research project. Department of chemical engineering University of Port Harcourt. 1-5
- Igwe, J. C. and Onyegbado, C.C. (2007). A review of palm oil mill effluent (POME) water treatment. *Global Journal of Environmental Research*. (2). 54-62
- Ipek, U. Obek, E. Akca, L. Arslan, E. I. Hasar, H. Dogru, M. and Baykara, O. (2002). Determination of degradation on radioactivity and its kinetics in aerobic compositing. *Bioresource technology*. (84). 283-286
- James, G. C. and Natalie, S. (2011). *Microbiology a laboratory manual*. Ninth edition. Pearson education. Inc.
- Kissi, M. Mountadar, M. Gargiulo, E. Palmieri, G. Giardina, P. and Sannia, G. (2001).
 Roles of two white-rot basidiomycete fungi in decolorisation and detoxification of olive mill wastewater. *Applied microbiology and biotechnology*. (57). 221-226
- Ko, J. J. Shimizu, Y. Ikeda, K. Kim, S. K. Park, C. H. and Matsui, S. (2009). Biodegradation of high molecular weight lignin under sulfate reducing conditions: Lignin degradability and degradation by-products. *Bioresource Technology*. (100). 1622–1627
- Ma, A. N. (2000). Environmental Management for the Oil Palm Industry Palm Oil Development. (30). 1-10

- Mazidatul Ashiqeen Balqiah Binti Mohamad Lazim. Treatment of Final Discharge Palm Oil Mill Effluent (POME) using Biofilm-Coated Macrocomposite. Master of Science (Biotechnology). Universiti Technologi Malaysia. 2012
- Neoh, C. H. Yahya, A. Adnan, R. Majid, A. Z. and Ibrahim, Z. (2012). Optimization of decolorization of palm oil mill effluent (POME) by growing cultures of Aspergillus fumigates using response surface methodology. *Environ Sci Pollut Res* DOI 10.1007/s11356-012-1193-5
- Poh, P. E. and Chong, M. F. (2009). Development of anaerobic digestion methods for palm oil mill effluent (POME) treatment. *Bioresource technology*. (100). 1-9
- Rakamthong, C. and Prasertsan, P. Decolorization and phenol removal of anaerobic palm oil mill effluent by Phanerochaete chrysosporium ATCC 24725, *TIChE International Conference 2011*, 10-11 November 2011, Hatyai, Songkhla, Thailand
- Rupani, P. F. Singh, R. P. Ibrahim, M. H. and Esa, N. (2010). Review of current palm oil mill effluent (POME) treatment methods: Vermicomposting as a sustainable practice. *World Applied Sciences Journal*. (1). 70-81
- Singh, D. and Maurys, S. (2010). Quantitative analysis of total phenolic content on adhatoda vasica nees extracts. (4). 2403- 2406
- Sjostrom, E. (1981). Wood Chemistry, Fundamentals and Applications. Academic Press. New York
- Suriati Binti Mat Ghani, Treatment of anaerobic palm oil mill effluent (POME) using single bacteria, Bachelor of Science, Universiti Teknologi Malaysia, 2011
- Sylvia, D. M. Fuhrmann, J. J. Hartel, P. G. and Zuberer, D. A. (2005). *Principles and Applications of Soil Microbiology*. 2nd edition. 304
- Tariq, M. (2002). On the mechanism of nitriles toxicity. (110). 246-248

- Tortora, G. J. Funke, B. R. and Case, C. L (2007). *Microbiology: An introduction*.9th *Ed., Pearson Education*
- Vairappan, C. S. and Yen, A. M. (2008). Palm oil mill effluent (POME) cultured marine microalgae as supplementary diet for rotifer culture. *Application Physics*. (5). 153-158
- Wershaw, R. L. Rutherford, D. W. Leenheer, J. A. Kennedy, K. R. Larry, G. C. and Koci, D. R. (2005). *Biogeochemical processes that produce dissolved organic matter from wheat straw*. <u>http://pubs.usgs.gov/wri034213/report.html</u>
- Zahrim, A. Y. Rachel, F. M. Menaka, S. Su, S. Y. Melvin, F. and Chan, E. S. (2009). Decolourisation of anaerobic palm oil mill effluent via activated sludge-granular activated carbon. *World applied sciences journal*. (5). 126-129
- Zimbro, M. J. and Power, D. A. (2003). Difco & BBL manual: manual of microbiological culture media. Becton, Dickinson and Company, Sparks, Maryland.