CHLOROPLAST DNA DIVERSIFICATION OF MALAYSIAN PINEAPPLE

NORFADILAH BINTI HAMDAN

A dissertation is submitted in partial fulfillment of the requirements for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Medical Engineering Universiti Teknologi Malaysia

JANUARY 2013

To my Parent,

Sisters,

and Friends,

Thanks for your pray, attention and spiritual support...

ACKNOWLEDGEMENT

Alhamdulillah, I praise to the almighty Allah for giving me the strength and patience to complete my research. This project would not have been possible without the support of many people and I deeply indebted to many people who helped in the completion of this thesis. I would like to begin by thanking my sole supervisor Dr. Azman Abd. Samad and co- supervisor Dr. Topik Hidayat for their guidance, advice and support. My deep appreciation to both of them for giving me the advice, guidance and suggestions I needed to complete my research. I would also like to express my sincere gratitude to Dr. Faezah Salleh for her tips and consultation in completion and improvement of this thesis. Last but not least, words are inadequate to express my thanks to my parents, family members, lab mate Ms Shahkila Mohd Arif, and numerous friends who endured this long process with me, always offering valuable advice, support and love.

ABSTRACT

There are several genes are normally used for the phylogenetic study such as Mitochondria DNA (mtDNA), Chloroplast DNA (cpDNA) and Nuclear DNA (nDNA). Among these three genomes, cpDNA is commonly being used in phylogenetic study because it is easy to amplify via PCR, evolves at a conservation rate and it has appropriate length and base substitution rate for inferring phylogeny at higher levels. In this study, phylogenetic tree of eight Malaysian pineapple (Ananas comosus) cultivars were constructed using sequences of large subunit of the ribulosebisphosphate carboxylase (rbcL) gene. A rbcL gene was isolated from genomic DNA, amplified and sequenced. Phylogenetic analysis was carried out using maximum parsimony method. Results revealed that rbcL gene of Ananas comosus is about 1250 bps. Based on the tree, eight Malaysian pineapple cultivars were classified into two groups. The first group consisted of Yankee and Gandul cultivars while second group consisted of Moris, Moris Bentanggur, Moris Gajah, N36, Josaphine and Sarawak. Bootstrap value in some branches were low this is reflected by the small number of informative characters (981 were conserved and 85 were potentially informative) to build the tree. Formation of several group or subclades is due to its similar genetic pattern, thus support this system classification. The study suggested that rbcL gene could be used to determine the phylogenetic relationship distinguish the pineapple cultivars.

ABSTRAK

Terdapat beberapa jenis gen yang biasanya digunakan untuk kajian filogenetik seperti DNA Mitokondria (mtDNA), DNA Kloroplas (cpDNA) dan DNA Nuklear (nDNA). Antara ketiga-tiga genom ini, cpDNA banyak digunakan dalam kajian filogenetik kerana ianya mudah diamplifikasi melalui PCR, dan ia mempunyai saiz dan kadar pengantian base yang sesuai untuk membuat kesimpulan filogeni pada peringkat taksonomi yang tinggi. Dalam kajian ini, rajah pokok filogenetik dari lapan kultivar nanas (Ananas comosus) Malaysia telah dibina menggunakan gen ribulose*bisphosphate carboxylase* (rbcL). Gen rbcL telah diekstrak dari genomic DNA nanas. Kemudian, ia diamplikasi dan diklon sebelum di hantar untuk penjujukan. Jujukan DNA ini seterusnya melalui proses analisis filogenetik dan ianya dijalankan menggunakan kaedah Maximum Parsimony. Hasil analisa menunjukkan bahawa saiz gen rbcL bagi Ananas comosus adalah ±1250 bp. Berdasarkan filogeni ini, lapan kultivar nanas Malaysia boleh dikelaskan kepada dua kumpulan. Kumpulan pertama terdiri daripada kultivar Yan Kee dan Gandul manakala kumpulan kedua terdiri daripada kultivar Moris, Bentanggur Moris, Moris Gajah, N36, Josaphine dan Sarawak. Nilai Bootstrap di beberapa kumpulan adalah rendah dan ini disebabkan oleh bilangan karakter yang konservatif (981 konservatif dan 85 adalah karakter berpotensi) untuk membina pokok. Pembentukan kumpulan atau beberapa subkumpulan adalah disebabkan oleh persaman ciri genetik, seterusnya menyokong sistem klasifikasi ini. Kajian ini membuktikan bahawa jujukan dari gen rbcL boleh digunakan untuk menentukan hubungan filogeni bagi pembezaan antara kultivar nanas.

TABLE OF CONTENTS

CHAPTER

TITLE

PAGE

DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGMENT	iv
ABSTRACT	v
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	Х
LIST OF FIGURES	xi
LIST OF ABBREVIATIONS/	xiv
SYMBOLS	

CHAPTER 1 INTRODUCTION

1.1	Study background	1
1.2	Problem statement	3
1.3	Objectives of the study	4
1.4	Scope of the study	4
1.5	Significance of the study	5

CHAPTER 2 LITERATURE REVIEW

2.1	History and distribution of pineapple	6
	(Ananas comosus)	
2.2	Ananas comosus taxonomy	7
2.3	Plant morphology	9
2.4	Pineapple cultivars	12
2.5	Importances of pineapple	15
2.6	Molecular phylogenetic study on Ananas	16
2.7	Application of rbcL gene in molecular	17
	phylogenetic analysis	
2.8	Method of analysis in phylogenetic	19
	studies	

CHAPTER 3 MATERIALS AND METHODS

3.1	Exper	imental design and procedure	21
3.2	Plant	materials	24
3.3	Genor	mic DNA extraction	24
3.4	DNA	quantification	
	3.4.1	Spectrophotometer	25
	3.4.2	Agarose gel electrophoresis	25
3.5	Cloni	ng of rbc <i>L</i> gene	
	3.5.1	Amplification of rbcL gene by	26
PCR			
	3.5.2	Purufication of PCR product	28
	3.5.3	Ligation	28
	3.5.4	Transformation	29
	3.5.5	Blue white screening	29

3.6	Plasm	id isolation	
	3.6.1	Plasmid extraction and glycerol	30
		stock preparation	
	3.6.2	Restriction Enzyme (RE)	31
		Digestion	
3.7	Bioinf	formatics analysis	32

CHAPTER 4 RESULTS AND DISCUSSION

4.1	Genomic DNA extraction	33
4.2	Amplification of rbcL gene by PCR	36
4.3	Selection for transformant	38
4.4	Plasmid isolation	39
4.5	Reconfirmation by Restriction enzyme	42
	(RE)	44
4.6	DNA sequencing, homology and	
	similarity search for rbcL gene of Ananas	44
	comosus	
4.7	Complete sequences alignment and	
	sequences editing using ClustalX and	
	Bioedit software	
4.8	Phylogenetic tree construction and	54
	analysis	

CHAPTER 5 CONCLUSION AND FUTURE WORK

5.1	Conclusion	59
5.2	Future work	60

REFERENCES		6	1
APPENDIX	A-Q	7	1

LIST OF TABLES

TABLE NO.	TITLE	PAGE
2.1	Morphology of Malaysian pineapple cultivars adapted from Hidayat et al., (unpublished)	10
3.1	Primers used in this study	27
3.2	Reagent for rbcL amplification	27
3.3	PCR amplification programmes	27
3.4	Ligation mixture preparation	28
3.5	Digestion mixture preparation	31
4.1	Purity of DNA extracted from eight Malaysian pineapple cultivars	34
4.2	Concentration of DNA extracted from eight Malaysian pineapple cultivars	35

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
2.1	Structure of pineapple plant (Ananas comosus var. comosus) adapted from Coppens d'Eeckenbrugge et al., 2011	11
2.2	Matured pineapple leaf (Malaysian Pineapple Industry Board, 2012)	11
2.3 (a)	Moris Gajah cultivar	14
2.3(b)	Josaphine cultivar	14
2.3 (c)	MD2 cultivar	14
2.3(d)	N36 cultivar	14
2.3(e)	MD2/T cultivar	14
2.3 (f)	Sarawak Green Local cultivar	14
2.3 (g)	Moris Bentanggur cultivar	14
2.3(h)	Yan Kee cultivar	14
2.3 (i)	Gandul cultivar	14
3.1 (a)	Flow chart of experimental design for the amplification of the rbc <i>L</i> gene from chloroplast genome of pineapple leaf	22
3.1(b)	Flow chart of experimental design for the cloning of rbc <i>L</i> gene	23
3.1 (c)	Flow chart of experimental design for the construction of rbc <i>L</i> phylogenetic tree	23

4.1	DNA extracted from Malaysian pineapple cultivars. M: 1kb DNA ladder (New England BioLabs), 1: Sarawak, 2: Josaphine, 3: Moris Gajah, 4: Moris Bentanggur, 5: Moris, 6: Gandul, 7: Yan Kee, 8: Josaphine	34
4.2	PCR products for rbc<i>L</i> gene amplification M: 1kbDNA ladder (New England BioLabs), 1: Sarawak,2: Josaphine, 3: N36, 4: Gandul	37
4.3	PCR products for rbc <i>L</i> gene amplification M : 1kb DNA ladder (New England BioLabs), 1 : Moris Bentanggur, 2 : Moris, 3 : Moris Gajah, 4 : Yan Kee	38
4.4	Result for plasmid extraction, M: 1 kb DNA ladder (New England BioLabs), 1: Blue colony (negative control), 2: pGEM-T Easy::rbcLJ1 (with inserted gene), 3: pGEM-T Easy::rbcLJ2 (negative- no inserted gene), 3: pGEM-T Easy::rbcLS1 (with inserted gene), 4: pGEM-T Easy::rbcLS2 (with inserted gene), 5: pGEM-T Easy::rbcLN1 (with inserted gene), 6: pGEM-T Easy::rbcLN2 (with inserted gene), 7: pGEM-T Easy::rbcLG1 (with inserted gene), 8: pGEM-T Easy::rbcLG2 (with inserted gene)	40
4.5	Result for plasmid extraction, M : 1kb DNA ladder (New England BioLabs), 1 : pGEM-T Easy::rbc <i>L</i> M1 (with inserted gene), 2 : pGEM-T Easy::rbc <i>L</i> M2 (with inserted gene), 3 : pGEM-T Easy::rbc <i>L</i> MG1 (with inserted gene), 4 : pGEM-T Easy::rbc <i>L</i> MG2 (with inserted gene), 5 : pGEM-T Easy::rbc <i>L</i> Y1 (with inserted gene), 5 : pGEM-T Easy::rbc <i>L</i> Y2 (with inserted gene), 7 : pGEM-T Easy::rbc <i>L</i> MB1 (with inserted gene), 8 : pGEM-T Easy::rbc <i>L</i> MB2 (with inserted gene), 9 : Blue colony (negative control)	41
4.6	Results for reconfirmation by Restriction Enzyme (RE) using EcoRI, M: 1 kb DNA ladder (New England BioLabs), 1: Moris Gajah, 2: Moris Bentanggur, 3: Yan Kee.	43
4.7	Results for reconfirmation by Restriction Enzyme	43

(RE) using EcoRI, M: 1 kb DNA ladder (New
England BioLabs), 1: Josaphine, 2: Sarawak, 3:
N36, 4: Gandul, 5: Moris

4.8(a) rbcL gene full sequence of Moris Bentanggur 45 cultivar **4.8(b)** rbcL gene full sequence of Moris Gajah cultivar 46 **4.8(c)** rbc*L* gene full sequence of N36 cultivar 47 **4.8(d)** rbcL gene full sequence of Sarawak cultivar 48 **4.8(e)** rbcL gene full sequence of Gandul cultivar 49 **4.8(f)** rbcL gene full sequence of Yan Kee cultivar 50 **4.8(g)** rbcL gene full sequence of Josaphine cultivar 51 **4.8(h)** rbc*L* gene full sequence of Moris cultivar 52 4.9 Complete alignment of the eight sequences of 53 Malaysian pineapple cultivars. The black colour regions in the alignment shows the consensus sequences of the eight pineapple cultivars 4.10 Strict consensus tree from the Maximum 54 Parsimony analysis of the rbcL gene

LIST OF ABBREVIATIONS/ SYMBOLS

±	-	plus minus
%	-	percentage
° C	-	degree Celcius
μL	-	microliter
bp	-	basepair
BS	-	bootstrapt support
CI	-	consistency index
cm	-	centimetre
cpDNA	-	chloroplast DNA
g	-	gram
h	-	hour
IPTG	-	isopropyl β - D- 1- thiogalactopyranoside
ITS	-	internal transcribed spacer
kb	-	kilobase
kg	-	kilogram
LAIX	-	LB, ampicillin, IPTG, X- gal
LB	-	Luria- Bertani Broth
m	-	meter
min	-	minute
mL	-	mililiter
mM	-	micromolar
MP	-	Maximum Parsimony
MPIB	-	Malaysian Pineapple Industry Board

mtDNA	-	mitochondria DNA
nDNA	-	nuclear DNA
nm	-	nanometer
PCR	-	Polymerase chain reaction
rbcL	-	large subunit of ribulose- bisphosphate carboxylase gene
RI	-	retention index
rpm	-	revolutions per minute
sec	-	second
SOC	-	Super Optimal Broth with Catabolite repression
T _m	-	annealing temperature
U	-	unit
X- gal	-	5- bromo-4- chloro- indolyl- β - D- galactopyranoside

CHAPTER 1

INTRODUCTION

1.1 Study background

Chloroplasts are organelles which present abundantly in the leaf cells which play important role in metabolic activity known as photosynthesis process. The other important roles plays by the chloroplast are producing the starch, give the colour pigments to the flowers, certain amino acids, lipids and vitamins. Chloroplasts have their own genome whereby it possess a full complement of transcriptional and translation machinery in order to express their genetic information.

On the other hand, Pineapple (*Ananas comosus*) is the third most important tropical fruit crop in the world after bananas and mangoes (Botella and Smith 2008; Carlier *et al.*, 2007). It also has become one of the leading commercial fruit crops of the tropics over the past of 100 years. Other than other than Thailand, Philippines, Indonesia, Hawaii, Ivory Coast, Kenya, Brazil, Taiwan, Australia, India and South Africa countries, it was reported that Malaysia has become one of the world producer of pineapple fruit. Development of pineapple industry in Malaysia is monitored by Malaysian Pineapple Industry Board (MPIB). MPIB was established since the year of 1957. The role of MPIB is to manage and develop Malaysian Pineapple Industry.

Pineapple is a type of tropical plant which is believed to originate from East Area of South America. It was introduced in Malaya in the 16th century by the Portuguese. During a year of 1921, pineapple started to be planted in Singapore, Johor and Selangor as cash crop. Now, pineapple plantation is continued to be expended in peat soil area especially in Johor (Malaysian Pineapple Industry Board, 2012). There are many cultivars and 'Smooth Cayenne' is the most commonly grown worldwide. In Malaysia, there are about three common cultivars widely planted in Malaysian which are Spanish (also known as Maspine, Josapine and Hybrid pineapple), Smooth Cayenne (Sarawak pineapple) and Queen (Morris pineapple) (Malaysian Pineapple Board, 2012).

Recently, molecular systematics in plants has progressed rapidly with DNA amplification or known as polymerase chain reaction, PCR is mediated by the direct sequencing methods (Schulte *et al.*, 2008; Sheng-Guo *et al.*, 2008; Osaloo and Kawano, 1999). Molecular approach has offered effective method in addressing many phylogenetic questions which cannot be solved using morphology characters. Previous studies (Chase *et al.*, 1993; Spreitzer *et al.*, 2002) claimed that large subunit of the ribulose- bisphosphate carboxylase (rbcL) gene is suitable for inference phylogenetic relationship at higher taxonomic levels. The rbcL gene is usually up to 1250 bp in size and the used of this gene in phylogenetic analysis has been reviewed in many studies (Chase *et al.*, 1993; Clegg, 1993; Spreitzer *et al.*, 2002). This is due to its advantages where it is easy to amplify using PCR, have appropriate length and base substitution rate for inferring phylogeny at higher taxonomy levels and it evolves at a conservation rate which make it suitable to study plant phylogeny (Clegg, 1993).

Pineapple is well known among user and it is highly consumed due to its pleasant taste and medical values. Though it is has become popular, very little is known about the molecular genetics of pineapple. It is due to limitation on available data of *Ananas* genetic diversity and most of it is based on morphology character only. They apparently arose due to spontaneous mutation, followed by natural selection and cross hybrid with unknown ancestor (Ruas *et al.*, 1996). Hence, in this study, phylogenetic analysis of Malaysian pineapple or *Ananas comosus* was

conducted by using sequence data from the chloroplast gene which is known as large subunit of the ribulose- bisphosphate carboxylase (rbc*L*). The information about genetic variability at the molecular level is said to be useful to identify and characterize the unique germplasm that compliments the existing cultivars.

1.2 Problem statement

Pineapple is the third most important tropical fruit in the world after banana and mangoes (Botella and Smith, 2008). However, there is limitation on available data at molecular level for Malaysian pineapple. Previously, most of the classification of pineapple cultivars is based on morphology character. The drawback of classification based on morphological characteristics is inconsistency that arose due to disagreements among morphologist who applied different classification for interpretation of the characteristics.

Hence, in this study, phylogenetic analysis of Malaysian pineapple or *Ananas comosus* was conducted using sequence data from the chloroplast gene, *rbcL* to reconstruct a more detailed phylogenetic frame work of Malaysian pineapple and investigate the evolutionary relationships among these cultivars. The information about genetic variability at the molecular level is said to be useful to identify and characterize the Malaysian pineapple cultivars.

1.3 Objectives of the study

The objectives of the study are:

- 1. To amplify rbcL gene from eight Malaysian pineapple cultivars.
- To clone the rbcL into pGEM-T Easy Vector System (Promega) and sequence the rbcL gene from eight Malaysian pineapple cultivars.
- 3. To construct a phylogenetic tree from isolated sequence of rbc*L*.

1.4 Scope of study

The scope of the study covered the construction of phylogenetic tree from eight commercial Malaysian pineapple cultivars based on rbc*L* gene. In order to achieve this, genomic DNA was isolated from leaf samples of eight commercial Malaysian pineapple cultivars; Moris, Moris Gajah, Moris Bentanggur, Yan Kee, Sarawak, Gandul and N36. PCR was carried out using rbc*L* primers and purified PCR product was subjected to cloning procedure comprising ligation of rbc*L* gene into the pGEM- T Easy Vector System (Promega) followed by transformation into NEB-5 α competent cell (New England BioLabs). Positive transformants were sent for sequencing and sequences obtained were aligned using ClustalX software. Phylogenetic tree were constructed using MEGA 5 software and analysis were carried out using Maximum Parsimony method.

1.5 Significance of the study

Chloroplast DNA (cpDNA) has been used widely to infer plant systematic at different taxonomy levels (Clegg, 1993; Gielly and Taberlet, 1994). In this study, gene of the large subunit of ribulose –bisphosphate carboxylase (rbc*L*) was used as an alternative approach for morphology identification and to study the evolutionary status and relationship among the eight cultivars of Malaysian pineapple. The important of this study was to provide additional information on the relationship pattern among the cultivars which in future can be used as source of knowledge and information for successful interbreeding on creating new cultivars.

REFERENCES

- Aradhya, M., Zee, F., and Manshardt, R. M. (1994). Isoenzyme Variation in Cultivated and Wild Pineapple. *Euphytica*, *79*, 87-99.
- Araujo, E. F., Queiroz, L. P., and Machado, M. A. (2003). What is Citrus? Taxonomic Implications from a Study of cpDNA Evolution in the Tribe Citreae (Rutaceae Subfamily Aurantioideae). Org. Divers. Evol., 3, 55-62.
- Assuncao, P., Jaen-Molina, R., Caujape-Castells, J., de la Jara, A., Carmona, L., Freijanes, K., and Mendoza, H. (2011). Phylogenetic position of Dunaliella acidophila (Chlorophyceae) based on ITS and rbcL sequences. *J Appl Phycol*, 24, 635-639.
- Bartholomew, D. P., Paull, R. E., and Rohrback, K. G. (2003). The pineapple: Botany, Production and Uses. . Wallingford, UK: CABI Publishing.
- Berry, V., and O. Gascuel. (1996). On the Interpretation of Bootstrap Trees: Appropriate Threshold of Clade Selection and Induced Gain. *Mol. Biol. Evol*, 13, 999-1011.
- Boesenberg-Smith, K. A., Pessarakli, M. M., & Wolk, D. M. (2012). Assessment of DNA Yield and Purity: an Overlooked Detail of PCR Troubleshooting. *Clinical Microbiology Newsletter*, 34(1), 1-6.
- Botella, J. R., and Smith, M. (2008). Genomics of Pineapple, Crowning The King of Tropical Fruits. In P. H. Moore, and Ming, R. (Ed.), *Genomics of Tropical Crop Plants* (Vol. 1, pp. 441-451). New York: Springer.
- Brown, G. K., Palaci, C. A., and Luther, H. E. (1997). Chromosome Numbers in Bromeliaceae. *Selbyanna*, 18, 85-88.

- Brown, T. A. (2002). *Molecular Phylogenetics*. [Electronic Version]. From:http://www.ncbi.nlm.nih.gov/books/NBK21122/.
- Cabral, J. R. S., de Matos, A. P., and Coppens d'Eeckenbrugge, G. (1997). Segregation for Resistance to Fusariose, Leaf Margin Type and Leaf Colour from the EMBRAPA Pineapple Hybridization Programme. *Acta. Hort.*, 425, 23-28.
- Carlier, J. D., Coppens d'Eeckenbrugge, G., and Leitao, J. M. (2007). Pineapple. In Kole. C. (Ed.), Genome Mapping and Molecular Breeding in Plants, Fruits and Nuts (Vol. 4). Berlin, Heideberg: Springer- Verlag.
- Chan, Y. K., Coppens d'eeckenbrugge, G., and Sanewski, G. M. (2003). Chapter 3: Breeding and Variety Improvement In D. P. Bartholomew, Paull, R. E., and Rohrbach, K. G. (Ed.), *The Pineapple: Botany, Production and Uses* (pp. 33-51). Wallingford, UK: CABI Publishing.
- Chan, Y. K., and Lee, H. K. (1996). Josaphine: A New Pineapple Hybrid Developed at Mardi. *Proceeding of 2nd National Congress on Genetics, Kuala Lumpur.*
- Chase, M. W., Soltis, D. E., Olmstead, R. G., Morgan, D., Les, D. H., Mishler, B. D., Duvall, M. R., Price, R., Hills, H. G., Qui, Y. L., Kron, K. A., Rettig, J. H., Conti, E., Palmer, J. D., Manhart, J. R., Sytsma, K. J., Michaels, H. J., Kress, W. J., Karol, K. G., Clark, W. D., Hedren, M., Gaut, B. S., Jansen, R. K., Kim, K. J., Wimpee, C. F., Smith, J. F., Furnier, G. R., Strauss, S. H., Xiang, Q. Y., Plunkett, G. M., Soltis, P. S., Williams, S. E., Gadek, P. A., Quinn, C. J., Eguiarte, L. E., Golenberg, E., Learn, G. H., Graham, S., Barrett, S. C. H., Dayanandan, S., and Albert, V. A. (1993). Phylogenetics of Seed Plants: An Analysis of Nucleotide Sequences from the Plastid Gene rbcL. *Ann. Mo. Bot. Gard.*, 80, 528- 580.
- Chen, F. C., Chen, C. J., Li, W. H., and Chuang, T. J. (2010). Gene Family Size Conservation Is a Good Indicator of Evolutionary Rates. *Molecular Biology and Evolutionary*, 27(8), 1750-1758.

- Chouhan, U., and Pardasani, K. R. (2007). A Maximum Parsimony Model to Reconstruct Phylogenetic Network in Honey Bee Evolution. *International Journal of Biological and Life Sciences*, 3(3), 220-224.
- Clegg, M. T. (1993). Review: Chloroplast Gene Sequences and the Study of Plant Evolution. *Proc. Natl. Acad. Sci. USA*, *90*, 363-367.
- Coppens d'Eeckenbrugge, G., Leal, F. and Duval, M. F. (1997). Germplasm Resources of Pineapple. In J. Jules (Ed.), *Hortriculture Review* (Vol. 22, pp. 133-137). Oxford, UK: John Wiley & Sons, Inc.
- Coppens d'Eeckenbrugge, G., and Leal, F. (2003). Morphology, Anatomy and Taxanomy. In D. P. Bartholomew, Paull, R. E., and Rohrbach, K. G. (Ed.), *The Pineapple: Botany, Production and Uses* (pp. 13-32). Wallingford, UK: CABI International.
- Coppens d'Eeckenbruggee, G., and Sanewski, G. M. (2011, July 2011). Leaf Margin in Pineapple. *Newsletter of the Pineapple Working Group, International Society for HortScience. July 2011*, pp. 32-37.
- De La Cruz, M. J., and Garcia, H. S. (2005). *Pineapple: Post- Harvest Operation*. Veracruz, Ver, MEXICO: Instituto Tecnologico de Veracruz.

[Electronic Version] From: http://www.fao.org/inpho/inpho-post-harvestcompendium/fruits-vegetables/en/

- DeWald, M. G., Moore, G. A., and Sherman, W. B. (1992). Isozymes in Ananas (Pineapple): Genetics and Usefulness in Taxonomy. J. Amer. Soc. Hort. Sci., 117(3), 491-496.
- Duval, M. F., Buso, G. S. C., Ferreira, F. R., Noyer, J. L., Coppens d'Eeckenbrugge, G., Hamon, P., and Ferreira, M. E. (2003). Relationships in Ananas and Other Related Genera Using Chloroplast DNA Restriction Site Variation. *Genome* NRC Research Press, 46, 990-1004.

- Duval, M. F., Noyer, J. L., Perrier, X., Coppens d'Eeckenbrugge, G., and Hamon, P. (2001). Molecular Diversity in Pineapple Assessed by RFLP markers. *Theoritical Applied Genetics*, 102, 83-90.
- Graybeal, A. (1998). Is It Better to Add Taxa or Characters to a Difficult Phylogenetic Problem? *Systematic Biology*, 47, 9-17.
- Gielly, L., and Taberlet, P. (1994). The Use of Chloroplast DNA to Resolve Plant Phylogenies: Noncoding versus rbcL Sequences. *Mol. Biol. Evol.*, 11(5), 769-777.
- Gitai, J., Horres, R., and Benko-Iseppon, I. M. (2005). Chromosomal Features and Evolution of Bromeliaceae. *Plant Systematics Evolution*, 253, 65-80.
- Hale, L. P., Greer, P. K., Trinh, C. T., and Gottfried, M. R. (2005). Treatment with Oral Bromelain Decreases Colonic Inflammation in the IL-10 Deficient Murine Model of Inflammatory Bowel Disease. *Clinical Immunology*, 116, 135-142.
- Hamid, M. J. A., and Ali, A. K. (2005). An Assessment of the Impact of Technology on Josaphine Pineapple Grown in Malaysia. Unpublished Research Report. Economics and Technology Management Research Centre, MARDI.
- Hamry, R. K., and Zimmer, E. A. (1992). Ribosoman RNA as Phylogenetic Tool in Plant Systematic. In P. S. Soltis, Soltis, D. E. and Doyle, J.J. (Ed.), *Molecular Systematic of Plants* (pp. 50-91). New York: International Thompson Publishing.
- Harrison, C. J., and Langdale, J. A. (2006). A Step by Step Guide to Phylogeny Reconstruction. *The Plant Journal, 46*, 561-572.
- Hidayat, T. (2012). Morphological Characteristic and Phylogeny of Malaysian Pineapple Cultivars. Unpublished Research Report. Universiti Teknologi Malaysia.
- Hidayat, T., and Pancoro, A. (2006). Short Comunication: DNA Technology and Studies in Phylogenetic Relationship of tropical Plant: Prospect in Indonesia.

Paper presented at the International Conference on Mathematics and Natural Sciences (ICMNS) on 29- 30 November 2006.

- Hidayat, T., Pancoro, A., Kusumawaty, D., and Eiadthong, W. (2011). *Molecular Diversification and Phylogeny of Mangifera (Anacardiaceae) in Indonesia and Thailand*. Paper presented at the Proceeding of the International Conference on Advance Science, Engineering and Information Technology 2011, Hotel Equatorial Bangi- Putrajaya, Malaysia on 14-15 January 2011.
- Hillis, D. M. (1996). Inferring Complex Phylogenetic. Nature, 383, 130-131.
- Horres, R., Schulte, K., Weising, K., and Zizka, G. (2007). Systematics of Bromelioideae (Bromeliaceae): Evidence from Molecular and Anatomial Studies. *Aliso*, 23, 27-43.
- Horres, R., Zizka, G., and Weising, K. (2000). Molecular Phylogenetics of Bromeliaceae: Evidence from trnL(UAA) Intron Sequences of the Chloroplast Genome. *Plant Biology*, 2, 306-315.
- Huang, J., Giannasi, D. E., Price, R. A. (2003). Phylogenetic Relationships in Ephedra (Ephedraceae) Inferred from Chloroplast and Nuclear DNA Sequences. *Molecular Phyogenetics and Evolution*, 35, 48-59.
- Ji, R. M. T., and Li, J.Q. (2011). The Comparative Study of Different Methods for Total DNA Extraction in Tibet Planteau Potentilla anserine. Northern Horticulture, 6, 152-154.
- Kato, C. Y., Nagai, C., Moore, P. H., Zee, F., Kim, M. S., Steiger, D. L., and Ming, R. (2005). Intra- specific DNA Polymorphism in Pineapple (Ananas comosus (L.) Merr.) Assessed by AFLP Markers. *Genetic Resources and Crop Evolution*, 52(8), 815-825.
- Koichiro, T., Peterson, D., Peterson, N., Trecher, G., Nei, M., and Kumar, S. (2011).
 MEGA 5: Molecular Evolutionary Genetics Analysis Using Maximum Likelihood, Evolutionary Distance and Maximum Parsimony Methods. *Molecular Biology and Evolution*, 28(10), 2731-2739.

- Krizman. M., J. J., Baricevic. D., Javornik. B., and Prosek. M. (2006). Robust CTAB-activated Charcoal Protocol for Plant DNA Extraction. Acta Agric. Slovenica., 87, 427-433.
- Kumar, S., and Filipski, A. J. (Ed.) (2001) Encyclopedia of Life Sciences. Macmillan Publishers Ltd, Nature Publishing Group.
- Malaysian Pineapple Industry Board. Kultivar Nanas. Retrived October 19, 2012 from http://www.mpib.gov.my/web/guest/asal_usul_nanas
- Mateljan, G. (2007). The Worlds Healthiest Foods. Seattle WA, United States.
- Mondragon- Jacobo, C., Doudareva, N., Bordelon, B.P. (2000). DNA Extraction from Several Cacti. *HortScience*, *35*(6), 1124-1126.
- Moyo, M., Amoo, S.O., Bairu, M.W., Finnie, J.F., Van Staden, J. (2008). Optimising DNA Isolation for Medicinal Plants. South African Journal of Botany, 74, 771-775.
- Mynott, T. L., Ladhams, A., Scarmato, P., and Engwerda, C. R. (1999). Bromelain, from Pineapple Stems, Proteolytically Blocks Activation of Extracellular Regulated Kinase- 2 in T Cells. *The Journal of Immunology*, 163, 2568-2575.
- Nei, M., and Kumar, S. (2000). Chapter 7: Phylogenetic Inference- Maximum Parsimony Method. In M. Nei, and Kumar, S. (Ed.), *Molecular Evolution and Phylogenetics* (pp. 115-143). New York: University Press Incorporation.
- Osaloo, S. K., and Kawano, S. (1999). Molecular systematics of Trilliaceae II. Phylogenetic Analyses of Trillium and its Allies Using Sequences of rbcL and matK Genes of cpDNA and Internal Transcribed Spacers of 18S–26S nrDNA. *Plant Species Biology*, *14*, 75-94.
- Pandey, R. N., Adams, R. P., and Flournoy, L. E. (1996). Inhibition of Random Amplified Polymorphic DNAs (RAPDs) by Plant Polysaccharides. *Plant Molecular Biology Reporter*, 14(1), 17-22.
- Paz, E. Y., Gil, K., Rebolledo, L., Rebolledo, A., Uriza, D., Martines, O., Isidron,M., and Simpson, J. (2005). AFLP Characterization of the Mexican Pineapple

Germplasm Collection. Journal of the American Society for Horticultural Science, 130(4), 575-579.

- Penjor, T., Toyoaki, A., Yukio, N., Ryoji, M. and Masashi, Y. (2010). Phylogenetic Relationships of Citrus and its Relatives Based on rbcL Gene Sequences. *Tree Genetics & Genomes.*, 6, 931-939.
- Purseglove, J. W. (1972). *Tropical Crops: Monocotyledons*. London, England: Longman Group Ltd London.
- Py, C., Lacoeuilhe, J. J., and Teisson, C. (1987). Part 1: The Plant and the Environment. In C. Py, Lacoeuilhe, J. J., and Teisson, C. (Ed.), *The Pineapple: Cultivation and Uses* (pp. 29-72). Paris: Maisonneuve et Larose.
- Ruas, C. F., Ruas, P. M., and Cabral, J. R. (2001). Assessment of Genetic Relatedness of the Genera Ananas and Pseudonanas Confirmed by RAPD Markers. *Euphytica*, 119, 245-252.
- Ruas, P. M., Ruas, C. F., Fairbanks, D. J., Andersen, W. R., and Cabral, J. R. S. (1996). Genetic Relationship Among Four Varieties of Pineapple, Ananas comosus, Revealed by Random Amplified Polymorphic DNA (RAPD) Analysis. *Brazilian Journal of Genetics*, 18(3), 413-416.
- Samuels, G. (1970). Pineapple Cultivars. Proceeding of Tropical Region America Society Horticultural Science, 14, 13-24.
- Schulte, K., Barfuss, M. H. J., and Zizka, G. (2008). Phylogeny of Bromelioideae (Bromeliaceae) Inferred from Nuclear and Plastid DNA Loci Reveals the Evolution of the Tank Habit Within the Subfamily. *Molecular Phylogenetics* and Evolution, 51, 327-339.
- Shankar, K., L. Chavan, Shinde, S., and Patil, B. (2011). An Improved DNA Extraction Protocol from Four *in vitro* Banana Cultivars. *Asian Journal of Biotechnology*, 3(1), 84-90.

- Sheng-Guo, J., Ke-Ke, H., Jun, W., and Sheng-Li, P. (2008). A Molecular Phylogenetic Study of Huperziaceae Based on Chloroplast rbcL and psbAtrnH Sequences. *Journal of Systematics and Evolution*, 46(2), 213-219.
- Smith, L. B., and Downs, R. J. (1979). Monograph 14, Pt 3, Flora Neotropica. In L.B. Smith, and Downs, R. J. (Ed.), Bromelioideae (Bromeliaceae) (pp. 2142).NYBG, New York, USA.
- Soltis, D. E., Soltis, P. S., Mort, M. E., Chase, M. W., Savolainen, V., Hoot, S. B., and Morton, C. M. (1998). Inferring Complex Phylogenies Using Parsimony: An Empirical Approach Using Three Large DNA Data Sets for Angiosperms. *Syst. Biol.*, 47, 32-42.
- Sounders, G. W., and Kucera, H. (2010). An Evaliation of rbcL, tufA, UPA, LSU and ITS as DNA Barcode Markers for the Marine Green Macroalgae. *Cryptogamie, Algologie, 31*(4), 487-528.
- Spreitzer, R. J., and Salvucci, M. E. (2002). RUBISCO: Structure, Regulatory Interactions, and Possibilities for a Better Enzyme. *Anual Review of Plant Biology*, 53, 449-475.
- Steege, D. A. (1977). 5'-Terminal Nucleotide Sequence of E. coli Lactose Repressor mRNA: Features of Translational Initiation and Reinitiation Sites. *Proc. Natl. Acad. Sci. U.S.A.*, 74, 4163-4167.
- The Biology of Ananas comosus var. comosus (Pineapple). (2007). Australian Government, Department of Health and Ageing, Office of the Gene Technology Regulator.
- Thomas, M. G., Hagelberg, E., Jone, H. B., Yang, Z., and Lister, A. M. (2000). Molecular and Morphological Evidence on the Phylogeny of the Elephantidae. *Proceedings of Biological Sciences*, 22(267), 2493–2500.
- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F., and Higgins, D. G. (1997). The CLUSTAL_X Windowa Interface: Flexible Strategies for Multiple Sequence Alignment Aided by Quality Analysis Tools. *Nucleic* Acids Research, 25(24), 4876-4882.

- Vandamme, A. M. (2003). Basic Concepts of Molecular Evolution. In M. Salemi, and Vandamme, A. M. (Ed.), *The Phylogenetic Handbook: A Practical Approach to DNA and Protein Phylogeny* (pp. 10-14). United Kingdom: Cambridge University Press.
- Van de Vlies, P. and van Diemen, C. C. (2012). Guidelines Analysis of DNA Quantity and Quality for Infinium and GoldenGate Projects. Version 2. UMCG Genetica- Genome Analysis Facility.
- Varma, A. H., and Padh, N. (2007). Plant Genomic DNA Isolation: An Art or A Science. *Biotechnology Journal*, 2, 386-392.
- Wang, X., Xiao, H., Zhao, X, Li, C., Ren, J., Wang, F., and Pang, L. (2012).
 Isolation of High-Quality DNA from a Desert Plant Reaumuria soongorica.
 Çalişkan, M. (Ed.), *Genetic Diversity in Plants*, ISBN: 978-953-51-0185-7,
 InTech, DOI: 10.5772/38367. [Electronic Version]
 From:http://www.intechopen.com/books/genetic-diversity-in-plants/isolationof-high-quality-dna-from-a-desert-plant-reaumuria-soongorica
- Wee, Y. C. (1970). Some Common pineapple Cultivars of West Malaysia. Malays Pineapple, pp. 7-13.
- Williams, T. L., and Moret, B. M. E. (2003). An Investigation of Phylogenetic Likelihood Methods. Proceedings of 3rd IEEE Symposium on Bioinformatics and Engineering (BIBE '03).
- Wolfe, K. H., Li, W. H., and Sharp, P. M. (1987). Rates of Nucleotide Substitution Vary Greatly Among Plant Mitochondrial, Chloroplast and Nuclear DNAs. *Proc. Natl. Acad. Sci.*, 84, 9054- 9058.
- Yang, Z. (1995). Phylogenetic Analysis Using Parsimony and Likelihood Methods. Journal of Molecular Evolution, 42, 294-307.
- Zhang, K. (2011). Comparison of DNA Extration Methods from *Herba cistanche*. Journal Anhui. Agri. Sci., 39, 3957-3959.