

**CHEMICAL CONSTITUENTS FROM THE LEAVES OF *GARCINIA*
*PARVIFOLIA***

ATIKAH BINTI SALLEH

UNIVERSITI TEKNOLOGI MALAYSIA

**CHEMICAL CONSTITUENTS FROM THE LEAVES OF *GARCINIA
PARVIFOLIA***

ATIKAH BINTI SALLEH

A dissertation in partial fulfillment of the
requirements for the award of the degree of
Master of Science (Chemistry)

**Faculty of Science
Universiti Teknologi Malaysia**

2013

Dedicated to my beloved family...

ACKNOWLEDGEMENT

First and foremost, I show my gratitude to The Almighty God for giving me the strength to complete this thesis.

A million of thanks to my supervisor, Dr. Norazah Basar for her guidance, patience, encouragement and continuous support to me in order to complete this work. Without her guidance and patience, this thesis will difficult to complete.

I wish to extend my warmest thanks to Mr. Azmi and Mr. Rasyidi for their assistance and co-operation in carrying for the NMR facility. I also appreciated to all Natural Product Research member; Amira, Atiqah, Iryani, Siti Maryam, Fatin Fasihah, Erniyanti, Syazwani and Awanis for the invaluable assistance and make me comfortable during my labwork. A million thanks to all Master students especially to Farhana, Mardiana and Fatimah for their support and friendship.

Last but not least, I owe the most special thanks to my parent, Hj Salleh bin Sujak and Hj Amerahwati Abd. Rahman and siblings for their unflagging love, support and pray for my success.

ABSTRACT

The chemical constituents of the leaves of *G. parvifolia* (known as kandis) have been investigated. Extraction of dried leaves was successfully done by cold extraction method using ethanol as solvent, followed by partitioning using *n*-hexane, chloroform and ethyl acetate. Fractionation and purification on the *n*-hexane crude extract by using Vacuum Liquid Chromatography (VLC) and Column Chromatography (CC) have resulted four compounds. The elucidation of the structures were carried out by spectroscopic techniques using IR, ¹H, ¹³C, DEPT and GC-MS. Analysis by spectroscopic data showed the isolated compounds are β-sitosterol, squalene, friedelin-3β-ol, and mixture of friedelin and friedelin-3β-ol. The antibacterial activity of crude extracts was carried out using disc diffusion methods with eight strains of bacteria, *Enterococcus faecalis*, *Staphylococcus aureus*, *Bacillus subtilis*, *Bacillus cereus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia* and *Pseudomonas putida*. Antibacterial screening showed that the chloroform crude extract gave a strong towards *Klebsiella pneumonia*, *Pseudomonas aeruginosa* and *Pseudomonas putida* with minimum inhibition concentration of 450 µg/mL.

ABSTRAK

Sebatian kimia daripada daun *G. parvifolia* (dikenali sebagai kandis) telah dikaji. Pengekstrakan daun kering telah berjaya dilakukan menggunakan kaedah pengekstrakan sejuk yang menggunakan pelarut etanol, diikuti dengan pembahagian menggunakan *n*-heksana, kloroform and etil asetat. Pemeringkatan dan penulenan ekstrak mentah menggunakan Kromatografi Cecair Vakum dan Kromatografi Turus telah menghasilkan empat sebatian. Pencirian struktur sebatian telah dilakukan dengan teknik spektroskopi menggunakan IM, ¹H, ¹³C, DEPT RMN dan KG-SJ. Analisa terhadap data spektroskopi menunjukkan sebatian yang telah diasingkan ialah β -sitosterol, skualena, friedelin-3 β -ol dan campuran friedelin dan friedelin-3 β -ol. Ujian bakteria ke atas ekstrak mentah dan β -sitosterol telah dilakukan dengan menggunakan teknik pembauran cakera terhadap lapan jenis bakteria, *Enterococcus faecalis*, *Staphylococcus aureus*, *Bacillus subtilis*, *Bacillus cereus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia* and *Pseudomonas putida*. Penyaringan antibakteria menunjukkan ekstrak mentah kloroform memberikan perencatan yang kuat terhadap bakteria *Klebsiella pneumonia*, *Pseudomonas aeruginosa* dan *Pseudomonas putida* dengan kepekatan perencatan minimum 450 μ g/mL.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION	ii
	DEDICATION	iv
	ACKNOWLEDGEMENT	v
	ABSTRACT	vi
	ABSTRAK	vii
	TABLE OF CONTENTS	viii
	LIST OF TABLES	xi
	LIST OF FIGURES	xii
	LIST OF ABBREVIATIONS	xiii
	LIST OF APPENDICES	xv
1	INTRODUCTION	
	1.1 General Introduction	1
	1.2 The Guttiferae Family	2
	1.3 Genus <i>Garcinia</i>	3
	1.4 <i>Garcinia parvifolia</i>	4
	1.5 Problem Statement	5
	1.6 Objectives	5
	1.7 Scope of Research	6
2	LITERATURE REVIEW	
	2.1 Phytochemicals Study of <i>Garcinia parvifolia</i>	7

2.2	Phytochemicals Study on other <i>Garcinia</i> species	13
2.3	Bioactivities of Phytochemicals from <i>Garcinia parvifolia</i>	23
2.4	Bioactivities of Phytochemicals from other <i>Garcinia</i> species	25
3	RESEARCH METHODOLOGY	
3.1	General Experimental Procedures	28
3.2	Chemicals and solvents	29
3.3	Plant Material	29
3.4	Phytochemicals Study of the leaves of <i>Garcinia parvifolia</i>	29
3.4.1	β -Sitosterol (35)	30
3.4.2	Squalene (103)	31
3.4.3	Friedelin-3 β -ol (104)	31
3.4.4	Mixture of Friedelin (87) and Friedelin-3 β -ol (104)	32
3.5	Antibacterial Assay	33
3.5.1	Chemicals and Microorganism	33
3.5.2	Microorganisms and Culture Media	34
3.5.3	Disc Diffusion Method	34
3.5.4	Minimum Inhibition Concentration (MIC)	35
3.5.5	Minimum Bactericidal Concentration (MBC)	36
4	RESULTS AND DISCUSSION	
4.1	Phytochemicals of the leaves of <i>Garcinia parvifolia</i>	37
4.1.1	β -sitosterol (35)	38

4.1.2	Squalene (103)	39
4.1.3	Friedelin-3 β -ol (104)	40
4.1.4	Mixture of Friedelin (87) and Friedelin-3 β -ol (104)	42
4.1.5	Derivative of xanthone	47
4.2	Antibacterial Activity	49
5	CONCLUSION AND RECOMMENDATIONS	
5.1	Phytochemicals Study of the leaves of <i>Garcinia parvifolia</i>	53 53
5.2	Antibacterial Activity	54
5.3	Recommendations	
	REFERENCES	55
	APPENDICES	63 – 80

LIST OF TABLES

TABLE NO.	TITLE	PAGE
4.1	Yield of crude extract of leaves of <i>G. parvifolia</i>	38
4.2	The NMR spectral data of compound (104)	42
4.3	The overlapping of carbons in mixture of compound (87) and compound (104)	43
4.4	The NMR spectral data of compound (87) and compound (104)	45 - 46
4.5	Inhibition zone of the crude extracts and isolated compounds for Gram positive bacteria	49
4.6	Inhibition zone of the crude extracts and isolated compounds for Gram negative bacteria	50
4.7	MIC of the crude extract and pure compounds	51
4.8	MBC of the crude extract and pure compounds	51

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
1.1	Leaves of <i>G. parvifolia</i>	5
3.1	Arrangement of the discs in petri dish	35
4.1	IR spectrum of xanthone derivative	48
4.2	¹ H NMR spectrum of xanthone derivative	48
4.3	Skeleton of xanthone	49

LIST OF ABBREVIATIONS

B	-	Beta
Δ	-	Chemical shift
^{13}C NMR	-	Carbon- 13 Nuclear Magnetic Resonance
^1H NMR	-	Proton Nuclear Magnetic Resonance
CC	-	Column Chromatography
$(\text{CD}_3)_2\text{CO}$	-	Deuterated acetone
CDCl_3	-	Deuterated Chloroform
CHCl_3	-	Chloroform
D	-	Doublet
Dd	-	doublet of doublet
DDM	-	Disc Diffusion Method
DEPT	-	Distortionless Enhancement by Polarization Transfer
EtOAc	-	Ethyl acetate
Et_2O	-	Diethyl ether
GC	-	Gas Chromatography
GC-MS	-	Gas Chromatography-Mass Spectrometry
HMBC	-	Heteronuclear multiple bond correlation
HMQC	-	Heteronuclear multiple quantum correlation
IR	-	Infrared
J	-	Coupling constant
Lit.	-	Literature
M	-	Multiplet
MeOH	-	Methanol
Mg	-	Milligram
mL	-	Mililiter
MBC	-	Minimum Bactericidal Concentration

MIC	-	Minimum Inhibitory Concentration
NA	-	Nutrient Agar
NB	-	Nutrient Broth
$\mu\text{g/mL}$	-	Microgram per millilitre
Ppm	-	part per million
R_f	-	Retention factor
T	-	Triplet
TLC	-	Thin layer chromatography
VLC	-	Vacuum liquid chromatography

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
1	Mass Spectrum of β -sitosterol (35)	63
2	IR spectrum of β -sitosterol (35)	64
3	^1H NMR spectrum of β -sitosterol (35)	65
4	^{13}C NMR spectrum of β -sitosterol (35)	66
5	DEPT NMR spectrum of β -sitosterol (35)	67
6	Mass spectrum of squalene (103)	68
7	IR spectrum of squalene (107)	69
8	^1H NMR spectrum of squalene (107)	70
9	^{13}C NMR spectrum of squalene (107)	71
10	IR spectrum of friedelin-3 β -ol (108)	72
11	^1H NMR spectrum of friedelin-3 β -ol (108)	73
12	^{13}C NMR spectrum of friedelin-3 β -ol (108)	74
13	IR spectrum of mixture of friedelin (87) and friedelin-3 β -ol (108)	75
14	^1H NMR spectrum of mixture of friedelin (87) and friedelin-3 β -ol (108)	76
15	^{13}C NMR spectrum of mixture of friedelin (87) and friedelin-3 β -ol (108)	77
16	DEPT NMR spectrum of mixture of friedelin (87) and friedelin-3 β -ol (108)	78
17	^{13}C NMR spectrum of (A) mixture of friedelin (87) and friedelin-3 β -ol (108) and (B) friedelan-3 β -ol (108)	79

18	¹³ C NMR spectrum of (A) mixture of friedelin (87) and friedelin-3 β -ol (108) and (B) friedelin-3 β -ol (108) (expansion)	80
----	--	----

CHAPTER 1

INTRODUCTION

1.1 General Introduction

Natural products are derived from natural sources such as plants, animals or microorganisms. There are 250,000 to 500,000 species of plants were estimated on earth. Unfortunately, among the species, there is only 1-10% that used as food for human and animals and more species of plant more used for medicinal purposes [1]. Plant is always important role in traditional systems of medicine to prevent and treat the disease worldwide. The traditional medicines in all countries were employed naturally occurring plant medicines for thousands of years. People preferred in natural medicines rather than synthetic medicines because of its safe and effective [2].

Plants can be used as medicine because of the richest bio-resources of drugs of traditional medicinal systems, modern medicines, nutraceuticals, food supplements, folk medicines, pharmaceuticals, intermediate and chemical entitled for synthetic drugs. Plants have their intrinsic ability to resist pathogenic microorganisms [3]. Some plant can be used in traditional medicine because they contain a vast array of substances and active constituents that can be treat chronic and infectious diseases. Medicinal plants are considerably useful and economically essential [4].

According to World Health Organization, medicinal plants would be the best source to variety of drugs. Approximately 20 % of the plants found in the world have

been submitted to pharmaceutical and biological test. Nowadays, many researchers were screened of antibacterial plant extracts to find new compounds with the potential to act against multi resistant bacteria. The development of medicine was started with the Ayurveda in India. Ayurveda system of medicine has its long history of therapeutic potential. After Ayurveda, current advancements in drug discovery technology and search for novel chemical diversity have intensified the efforts for exploring leads. For a long period of time, plants have been valuable source of natural products for maintaining human health [5].

Plants can be divided into their main classes of bioactive compounds based on their similar characteristics. The main classes included flavonoids, terpenes, alkaloids, saponins, coumarins, steroid, lactones, phenols, cyclic ketones, stilbenes and acidamides [6, 7]. The drugs are derived from the whole plant or from different organs like leaves, stem, bark, root, flower, seeds and twig. Some drugs also can be prepared from excretory plant product such as gum, resins and latex [8]. The different parts of plant have different potential as medicinal properties.

Over the last two decades, intensive effort has been made to discover chemically useful antibacterial or antifungal drugs of plant origin. Medicinal plant based antimicrobials represent a vast unstapped source of pharmaceuticals. Further exploration of plant antimicrobials need to occur for treatment of infectious diseases both in plants and humans while simultaneously for mitigating many of the side effects that are often associated with synthetic antimicrobials. However, only a small portion of the several hundred thousand medicinal plant species has been investigated both phytochemically and pharmacologically [3].

1.2 The Guttiferae Family

Guttiferae (or known as Clusiaceae) is one of the higher plant families of the plant kingdom. Guttiferae is comprised into six subfamilies, Kielmeyeroideae, Calophylloideae, Clusiodeae, Moronoboideae, Lorostemonodeae, Hypericoideae.

Every subfamily is divided into tribes. Subfamily of Clusiodeae is divided into two tribes: Clusieae and Garcinieae. Garcinieae is further divided into four genera: *Allanblackia*, *Garcinia*, *Pentapthalangium* and *Rheedia* while the tribe Clusieae consists of two genera (*Clustu* and *Tovomita*) [9].

Guttiferae family contains more than 1000 species and widely distributed in tropical region but majority of *Hypericum* genus occurs widely in temperate region. Oxygenated xanthenes and benzophenones have been found widely in all genera of the Guttiferae [9]. Whitmore, T.C. reported there are 4 genera and 121 species in Malaysia. The genera that can be found in Malaysia are *Garcinia*, *Calophyllum*, *Mesua* and *Mammea*. *Mammea* can be found only on lowlands whereas *Garcinia*, *Calophyllum* and *Mesua* are found from sea level to the tops of the highest mountain. Meanwhile *Garcinia* and *Mesua* are found in dry land forests and only *Calophyllum* is common in swampy forests [10].

1.3 Genus *Garcinia*

Garcinia belongs to the family Guttiferae (Clusiaceae), which is large genus of polygamus trees in the world tropics. It is widely distributed in tropical Africa, Asia, New Caledonia and Polynesia. In Indonesia, these plants were reported to be rich in natural chemical substances. Mostly, *garcinia* was grown and distributed in all over Indonesia [11].

Garcinia contains 35 genera and divided into 800 species and approximately 28 species that can be found in Malaysia. Genus *Garcinia* in Malaysia can be also called as fruit trees because it can provide palatable fresh fruit pulp and fruit peel. The best known fruit in Malaysia is the mangosteen (*Garcinia mangostana*). Asam gelugor (*Garcinia atroviridis*) used as tamarind slices in local dishes. *Garcinia* is very popular as high nutritional values but still underutilized due to lack of popularity among the local citizen. Most of these species have too acidic fruit pulp,

lack information on nutritional compositions and physical qualities and lack of promotional activities for these species [11, 12].

1.4 *Garcinia parvifolia*

Garcinia parvifolia is one of the species in genus *Garcinia*. It is synonyms with *G. dioica* and *G. globulosa* and also locally known as 'kandis' or cherry mangosteen. It is widely distributed in Thailand, Peninsular Malaysia, Sumatra, Java, Borneo, Celebes, Moluccas and New Guinea. It is a tropical species which can be found wild in various area like peat swamp forests, primary or secondary undisturbed mixed dipterocarp forests in the low-lands and humid submontane and up to an attitude of 1000 m. It also can be lived on hillsides, ridges, on well-drained, alluvial sites and along rivers [13, 14].

The species of *G. parvifolia* can be described as an evergreen, small to medium-sized, sub-canopy, perennial tree, 5-25 m high with a bole of 23 cm and with black widely patent branches and exuding yellow latex when bruised. While their leaves are opposite, shortly petiolate, oblong or lanceolate-oblong, entire, simple, penniveined, glabrous, 3.5-17 cm long, 2-7 cm wide, petiole 0.5-1.5 cm long with a shallow furrow on the anterior side. Besides that, the flowers are unisexual-monoecious, bisexual or polygamous, pale yellow or orange coloured, 0.5-1 cm across, pedicellate, solitary or in axillary fascicles of 2-12. Other part is fruit which can be described as depressed globose, pale-green turning yellow or orange-yellow when ripe 2.5-3.5 by 3.5-4 cm across, crowned by sunken remains of the small stigma with thin skin [14].

In Indonesia, the young leaf with sour taste can be eaten as a vegetable by their residences [13]. The ripe fruits can be used as flavouring in rice and curries since the arils have a pleasant and agreeable sub-acid taste [14].



Figure 1.1: Leaves of *G. parvifolia*

1.5 Problem Statement

Garcinia is one of the genera in Guttiferae family. Many medicinal plants from genus of *Garcinia* used as herbal medicine. Phytochemical research of *Garcinia parvifolia* was greatly enhanced by the search for bioactive compound since there are not been many phytochemicals and bioactivity studies conducted on this species. The purpose of this study is to identify the chemical constituent from the leaves of *G. parvifolia*. The isolation and evaluation of chemical compounds in the *G. parvifolia* is essential to be carried out to determine the chemical compounds. It is also important to study the antimicrobial activity of the chemical constituents to determine the medicinal value of the plant.

1.6 Objectives

The objectives of this research are:-

- 1) To extract and isolate chemical compounds from the leaves of *G. parvifolia*.
- 2) To characterize the structure of the chemical compounds by spectroscopic method.
- 3) To screen the antibacterial activity of the crude extract and pure isolated compounds.

REFERENCES

1. Varalakshmi, K.N., Sangeetha, C.G., Shabeena, A.N., Sunitha, S.R. and Vapika, J. (2010). Antimicrobial and Cytotoxic Effects of *Garcinia indica* Fruit Rind Extract. *American-Eurasian J. Agric. Environ. Sci.* **7 (6)**, 652-656.
2. Mahady, G.B., Huang, Y., Doyle, B.J. and Locklear, T. (2008). Natural products as antibacterial agents. *Atta-ur-Rahman (Ed.) ST. Nat. Pro. Chem.* **35**, 423-444.
3. Dasgupta, T., Rao, A.R. and Yadava, P.K. (2003). Modulatory effect of Henna leaf (*Lawsonia inermis*) on drug metabolizing phase I and phase II enzymes, antioxidant enzymes, lipid peroxidation and chemically induced skin and forestomach papillomagenesis in mice. *Mol. Cell. Biochem.* **245**, 11-22.
4. Sumathi, P. and Parvathi, A. (2010). Antimicrobial activity of some traditional medicinal plants. *J. Med. Plants. Res.* **4 (4)**, 316-321.
5. Sukanya, S.L., Sudisha, J., Hariprasad, P., Niranjana, S.R., Prakash, H.S. and Fathima, S.K. (2009). Antimicrobial activity of leaf extracts of Indian medicinal plants against clinical and phytophogenic bacteria. *Afr. J. Biotechnol.* **8(23)**, 6677-6682.
6. Yang, Y., Xi-Qiang, L. and Chun-Ping, T. (2011). Natural Products Chemistry Research 2009's Progress in China. *Chinese. J. Nat. Med.* **9 (1)**, 0007-0016.

7. Mcrae, J., Yang, Q., Crawford, R. and Palombo, E. (2007). Review of the methods used for isolating pharmaceutical lead compounds from traditional medicinal plants. *Environmentalist*. **27**, 165-174.
8. Joy, P.P., Thomas, J., Mathew, S., and Skaria, B.P. (2001). Medicinal Plants. Tropical Horticulture Vol. 2. (eds. Bose, T.K., Kabir, J., Das, P. and joy, P.P.). Naya Prokash, Calcutta. 449-632.
9. Bennett, G.J. and Lee, H. (1989). Xanthones from Guttiferae. *Phytochemistry*. **28 (4)**, 967-998.
10. Whitmore, T.C., (1973). Guttiferae. In: Tree Flora of Malaya, Whitmore, T.C. (Es.). Kuala Lumpur: Longman Malaysia. 162-236.
11. Joseph, G.S., Jayaprakasha, G.K., Selvi, A.T., Jena, B.S. and Sakariah, K.K. (2005). Antiaflatoxic and antioxidant activities of *Garcinia* extracts. *Int. J. Food Microbiol.* **101**, 153-160.
12. Hamid, M. A., Wen, P.K., Mamat, H. and Ibrahim, S. (2012). Determination of Nutritional Composition and effect of Various Storage Conditions on the Vitamin C Content in *Garcinia dulcis*, in Fresh and Dry Form. *Int. J. Adv. Sci. Eng. Info. Technol.* **2 (4)**, 42-47.
13. Syamsudin, Kumala, S. and Sutaryo. (2007). Screening of some extracts from *Garcinia parvifolia* Miq. (Guttiferae) for antiplasmodial, antioxidant, cytotoxic and antibacterial activities. *Asian J. Plant. Sci.* **6**, 972-976.
14. Lim T.K. (2012). Edible Medicinal and Non-Medicinal Plants: Fruits. Vol. 2. Springer Science + Business Media. 115-119.
15. Iinuma, M., Tosa, H., Tanaka, T. and Riswan, S. (1996). Three New Xanthones from the Bark of *Garcinia dioica*. *Chem. Pharm. Bull.* **44 (1)**, 232-234.

16. Xu, Y., Cao, S., Wu, X. H., Lai, Y.H., Tan, B.H.K., Pereira, J.T., Goh, S.H., Venkatraman, G. and Harrison, L.J. (1998). Griffipavixanthone, a Novel Cytotoxic Bixanthone from *Garcinia griffithii* and *Garcinia parvifolia*. *Tetrahedron Lett.* **39**, 9103-9106.
17. Xu, Y., Chiang, P., Lai, Y., Vittal, J.J., Wu, X., Tan, B.K.H., Imiyabir, Z. and Goh, S. (2000). Cytotoxic Prenylated Depsidones from *Garcinia parvifolia*. *J. Nat. Prod.* **63**, 1361-1363.
18. Rukachaisirikul, V., Trisuwan, K., Sukpondma, Y. and Phongpaichit, S. (2008). A New Benzoquinone Derivative from the Leaves of *Garcinia parvifolia*. *Arch Pharm Res.* **31 (1)**, 17-20.
19. Rukachaisirikul, V., Naklue, W., Phongpaichit, S., Towatana, N.H. and Maneenoon, K. (2006). Phloroglucinols, depsidones and xanthenes from the twigs of *Garcinia parvifolia*. *Tetrahedron.* **62**, 8578-8585
20. Xu, Y., Lai, Y., Imiyabir, Z. and Goh, S. (2001). Xanthenes from *Garcinia parvifolia*. *J. Nat. Prod.* **64**, 1191-1195.
21. Ee, G.L., Ng, S.H., Goh, J.K., Sukari, M.A. and Rahman, M. (2009). Chemical constituents of *Garcinia parvifolia* (Guttiferae). *Malaysian J. Sci.* **28(1)**, 105-110.
22. Lathifah, U.Z., Rahim, R.A., Sudrajat, H. and Khairi, S. (2010). Antiplasmodial activity of alkaloids from *Garcinia parvifolia* Miq. Stem Bark. *J. Iran. Chem. Res.* **3**, 59-63.
23. Nguyen, H.D., Trinh, B.T.D., Tran, Q.N., Nguyen, H.D., Pham, H.D., Hansen, P.E., Duus, F., Connolly, J.D. and Nguyen, L.D. (2011). Friedolanostane, friedocycloartane and benzophenone constituents of the bark and leaves of *Garcinia benthami*. *Phytochemistry.* **72**, 290-295.

24. Santa-Cecilia F.V., Freitas, L.A.S., Vilela, F.C., Veloso, C., Rocha, C.Q., Moreira, M.E.C., Dias, D.F., Giusti-Paiva, A. and dos Santos, M.H. (2011). Antinociceptive and anti-inflammatory properties of 7-epiclusianone, a prenylated benzophenone from *Garcinia brasiliensis*. *Eur. J. Pharmacol.* **670**, 280-285.
25. Gontijo, V. S., Souza, T.C., Rosa, I.A., Soares, M.G., da Silva, M.A., Vilegas, W., Junior, C.V. and dos Santos, M.H. (2012). Isolation and evaluation of the antioxidant activity of phenolic constituents of the *Garcinia brasiliensis* epicarp. *Food Chem.* **132(3)**, 1230-1235.
26. Shadid, K.A., Shaari, K., Abas, F., Israfi, D.A., Hamzah, A.S., Syakroni, N., Saha, K. and Lajis, N. (2007). Cytotoxic caged-polyprenylated xanthonoids and a xanthone from *Garcinia cantleyana*. *Phytochemistry.* **68**, 2537-2544.
27. Nguyen, H. D., Trinh, B. T. D. and Nguyen, L.D. (2011). "Guttiferones Q-S, cytotoxic polyisoprenylated benzophenones from the pericarp of *Garcinia cochinchinensis*. *Phytochem. Lett.* **4**, 129-133.
28. Mahabusarakam, W., Chairerk, P. and Taylor, W.C. (2005). Xanthones from *Garcinia cowa* Roxb. Latex. *Phytochemistry.* **66**, 1148-1153.
29. Deachathai, S., Mahabusarakam, W., Phongpaichit, S. and Taylor, W.C. (2005). Phenolic compounds from the fruit of *Garcinia dulcis*. *Phytochemistry.* **66**, 2368-2375.
30. Wu, J., Xu, Y., Cheng, X., Harrison, L.J., Sim, K., Goh, S.H. (2001) A highly rearranged tetraprenylxanthonoid from *Garcinia gaudichaudii* (Guttiferae). *Tetrahedron Lett.* **42**, 727-729.
31. Rukachaisirikul, V., Adair, A., Dampawan, P., Taylor, W.C. and Turner, P.C. (2000). Lanostanes and friedolanostanes from the pericarp of *Garcinia hombroniana*. *Phytochemistry.* **55**, 183-188.

32. Kaikabo, A.A., Samuel, B.B. and Eloff, J.N. (2009). Isolation and activity of two antibacterial biflavonoids from leaf extracts of *Garcinia livingstonei* (Clusiaceae). *Nat. Prod. Commun.* **4**(10), 1363-6
33. Nguyen, L.D., Vo, H.T., Pham, H.D., Connolly, J.D. and Harrison, L.J. (2003). Xanthones from the bark of *Garcinia merguensis*. *Phytochemistry.* **63**, 467-470.
34. Chien, S., Chyu, C., Chang, I., Chiu, H. and Kuo, Y. (2008). A novel polyprenylated phloroglucinol, garcinialone, from the roots of *Garcinia multiflora*. *Tetrahedron Lett.* **48**, 5276-5278.
35. Mawa, S. and Said, I.M. (2012). Chemical Constituents of *Garcinia prainiana*. *Sains Malays.* **41** (5), 585-590.
36. Lin, K., Huang, A., Yang, S., Weng, J., Hour, T., Pu, Y. and Lin, C. (2012). Cytotoxic and antioxidant constituents from *Garcinia subelliptica*. *Food Chem.* **135**, 851-859.
37. Nguyen, L.H.D. and Harrison, L. J. (2000). Xanthones and triterpenoids from the bark of *Garcinia vilersiana*. *Phytochemistry.* **53**, 111-114.
38. Chanmahasathien, W., Li, Y., Satake, M., Oshima, Y., Ruangrunsi, N. and Ohizumi, Y. (2003). Prenylated xanthones with NGF-potentiating activity from *Garcinia xanthochymus*. *Phytochemistry.* **64**, 981-986.
39. Syamsudin, Kumala, S. and Sutaryo. (2007). Screening of some extracts from *Garcinia parvifolia* Miq. (Guttiferae) for antiplasmodial, antioxidant, cytotoxic and antibacterial activities. *Asian J. Plant. Sci.* **6**, 972-976.
40. Kardono, L.B.S., Hanafi, M., Sherley, G., Kosela, S. and Harrison, L.J. (2006). Bioactive Constituents of *Garcinia porrecta* and *Garcinia parvifolia* Grown in Indonesia. *Pak J. Bio. Sci.* **9**, 483-486.

41. Jantan, I., Pizar, M.M., Idris, M.S., Taher, M. and Ali, R.M. (2012). In vitro inhibitory effect of rubraxanthone isolated from *Garcinia parvifolia* on Platelet-Activating Factor Receptor Binding. *Lett. Planta. Med.* **68**, 1133-1134.
42. Mackeen, M.M., Ali, A.M., Lajis, N.H., Kawazu, K., Hassan, Z., Amran, M., Habsah, M., Mooi, L.Y. and Mohamed, S.M. (2000). Antimicrobial, antioxidant, antitumour-promoting and cytotoxic activities of different plant part extracts of *Garcinia atroviridis* Griff. Ex T. Anders. *J. Ethnopharmacol.* **72**, 395-402.
43. Castardo J.C., Prudente, A.S., Ferreira, J., Guimaraes, C.L., Monache, F.D., Filho, V.C., Otuki, M.F. and Cabrini, D.A. (2008). Anti-inflammatory effects of hydroalcoholic extract and two biflavonoids from *Garcinia gardneriana* leaves in mouse paw oedema. *J. Ethnopharmacol.* **118**, 405-411.
44. Panthong, A., Norkaew, P., Kanjanapothi, D., Taesotikul, T., Anantachoke, N. and Reutrakul, V. (2007). Anti-inflammatory, analgesic and antipyretic activities of the extract of gamboges from *Garcinia hanburyi* Hook f. *J. Ethnopharmacol.* **111**, 335-340.
45. Baliga, M.S., Bhat, H.P., Pai, R.J. Bloor, R and Palatty, P.L. (2011). The chemistry and medicinal uses of the underutilized Indian fruit tree *Garcinia indica* Choisy (kokum): A review. *Food Res. Int.* **44**, 1790-1799.
46. Kaikabo, A.A. and Eloff, J.N. (2011). Antibacterial activity of two biflavonoids from *Garcinia livingstonei* leaves against *Mycobacterium smegmatis*. *J. Ethnopharmacol.* **138**, 253-255.
47. Negi, P.S., Jayaprakasha, G.K. and Jena, B.S. (2008). Antibacterial activity of the extracts from the fruit rinds of *Garcinia cowa* and *Garcinia pedunculata* against food borne pathogens and spoilage bacteria. *LWT-Food Sci. Technol.* **41**, 1857-1861.

48. Jayaprakasha, G.K., Negi, P.S. and Jena, B.S. (2006). Antioxidative and antimutagenic activities of the extracts from the rinds of *Garcinia pedunculata*. *Innov. Food Sci. Emerg. Technol.* **7**, 246-250.
49. Hill, R.A., Kirk, D.N., Makin, H.L.J. and Murphy, C.M. (1991). Dictionary of Steroids. Great Britain: Chapman and Hall.
50. Minh, C. V., Kiem, P. V., Huong, H.T., Dat, N. T., Nam, N.H., Lee, J. J. and Kim, Y.H. (2005). Chemical investigations and biological studies of *Mallotus apelta*. *J. Chem.* **43**, 235-239.
51. Mbaveng, A. T., Ngameni, et. al. (2008). Antimicrobial Activity of the Crude Extracts and Five Flavanoids from the Twigs of *Dorstenia barteri* (Moraceae). *J. Ethnopharmacol.* **116**, 489-489.
52. Aligians, N., Kalpoutzakis, E., Mitaku, S. and Chinou, I.B. (2001). Composition and Antimicrobial Activity of the Essential Oil from *Origanum* Species. *J. Agric Food Chem.* **49**, 4168-4170.
53. Pateh, U.U., Haruna, A.K., Garba, M., Iliya, I., Sule, I.M., Abubakar, M.S. and Ambi, A.A. (2008). Isolation of stigmasterol, β -sitosterol and 2-hydroxyhexadecanoic acid methyl ester from the rhizomes of *Stylochiton Lancifolius* Pyer and Kotchy (Araceae). *Niger J. Pharma. Sci.* **7(1)**, 19-25.
54. Ragasa, C.Y., Espineli, D.L., Mandia, E.H., Don, M. and Shen, C. (2012). A new triterpene from *Glinus oppositifolius*. *Chin. J. Nat. Med.* **10(4)**, 284-286.
55. Pie, H., Cai, Y., Sun, M. and Corke, H. (2002) Extraction and purification of squalene from *Amaranthus Grain*. *J. Argic. Food Chem.* **50**, 368-372.
56. Salazar, G.C.M., Silva, G.D.F., Duarte, L.P., Filho, S.A.V. and Lula, I.S. (2000). Two epimeric friedelane triterpenes isolated from *Maytenus truncate*

Reiss: ^1H and ^{13}C chemical shift assignments. *Magn. Reson. Chem.* **38**, 977-980.

57. Gunatilaka, A.A.L., de Silva, A.M.Y.J., Sotheeswaran, S., Balasubramaniam, S. and Wazeer, M.I.M. (1984). Terpenoid and biflavonoid constituents of *Calophyllum Calaba* and *Garcinia Spicata* from Sri Lanka. *Phytochemistry*. **23(2)**, 323-328.
58. Queiroga, C.L., Silva, G.F., Dias, P.C., Possenti, A. and de Carvalho, J.E. (2000). Evaluation of the antiulcerogenic activity of friedelan-3 β -ol and friedelin isolated from *Maytenus ilicifolia* (Celastraceae). *J. Ethnopharmacol.* **72**, 465-468.
59. Thanakijcharoenpath, W. and Theanphong, O. (2007). Triterpenoids from the stem of *Diospyros glandulosa*. *Thai J. Pharm. Sci.* **31**, 1-8.
60. Acar, J.F. and Goldstein, F.W. (1991). Disk Test. In: Antibiotic in Laboratory Medicine. Lorian (Ed), 3rd Edn.
61. Hess, S.C., Brum, R.L., Honda, N.K., Cruz, A.B., Moretto, E., Cruz, R.B., Mesanna, I., Ferrari, F., Filho, V.C. and Yunes, R.A. (1995). Antibacterial activity and phytochemical analysis of *Vochysia divergens* (Vochysiaceae). *J. Ethnopharmacol.* **47**, 97-100.