

# **THE EFFECT OF LAURIC ACID AS ANTIMICROBIAL AGENT IN CHITOSAN- BASED PACKAGING FILM**

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**BY :**

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**BACKGROUND**

# OBJECTIVE

To study the effect of Lauric acid as antimicrobial agent in Chitosan -based packaging film.

**Investigate The  
Effective Of The Food  
Preservative To Food  
Surface**

**Estimate The  
Antimicrobial Activities  
Using Agar Diffusion  
Test**

**Scope**

**Study The  
Antimicrobial Activities  
Using Liquid Culture  
Test**

**Select The  
Optimum Concentration Of  
Lauric Acid To Minimize  
The Migration Of Lauric Acid  
To Food Surface**

# INTRODUCTION

# Active Packaging

- As a barrier between the product and the outside environment in that interactions between the food and package occur that ultimately improve product quality and safety (Alvarez, 2000)
- It can control, and even react to, events taking place inside the package

# Active Packaging System

# Application

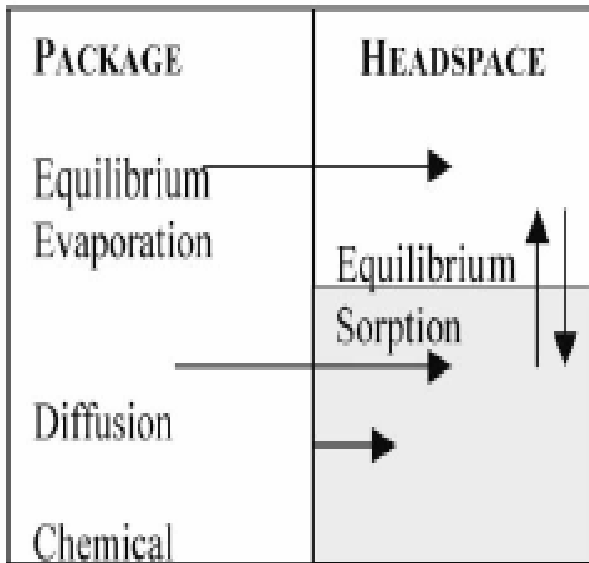
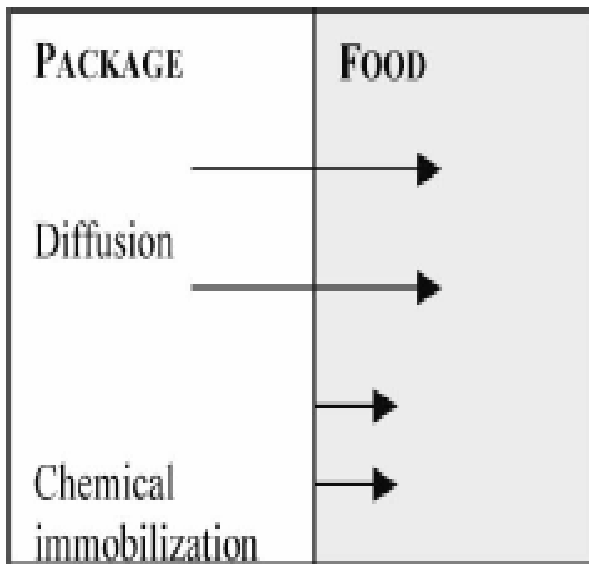
- ✿ Oxygen scavenging
  - ~Most food classes
- ✿ Carbon dioxide production
  - ~Most food affected by moulds
- ✿ Water vapour removal
  - ~Dried and mold-sensitive foods
- ✿ Ethylene removal
  - ~Horticultural produce
- ✿ Ethanol release
  - ~Baked foods (where permitted)
- ✿ Antimicrobial Activity
  - ~Most food classes

# Antimicrobial Packaging

- ✓ To extend the lag period and reduce the growth rate of microorganisms to prolong shelf life and maintain food safety
- ✓ To reduce microbial growth of nonsterile foods or maintain the stability of pasteurized foods without post-contamination



# **Antimicrobial Packaging System**



Ⓢ If the void volume of solid food products is assumed as a kind of headspace, most food packaging systems represent either a package/food system or a package/headspace/ food system

Ⓢ Diffusion between the packaging material and the food and partitioning at the interface are the main migration phenomena involved in this system.

Ⓢ Antimicrobial agents may be incorporated into the packaging materials initially and migrate into the food through diffusion and partitioning.

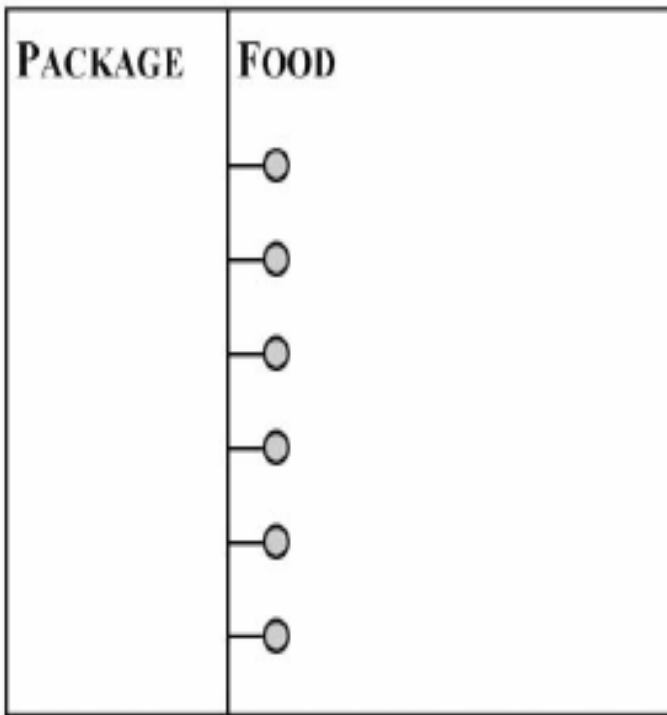
# **Type Of Antimicrobial Packaging**

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graph TD; A[Type Of Antimicrobial Packaging] --> B[Immobilization of antimicrobials to polymers by ion or covalent linkages]; A --> C[Coating or adsorbing antimicrobials onto polymer surfaces]; A --> D[Incorporation antimicrobial agents directly into polymers];
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**Immobilization of antimicrobials to polymers by ion or covalent linkages**

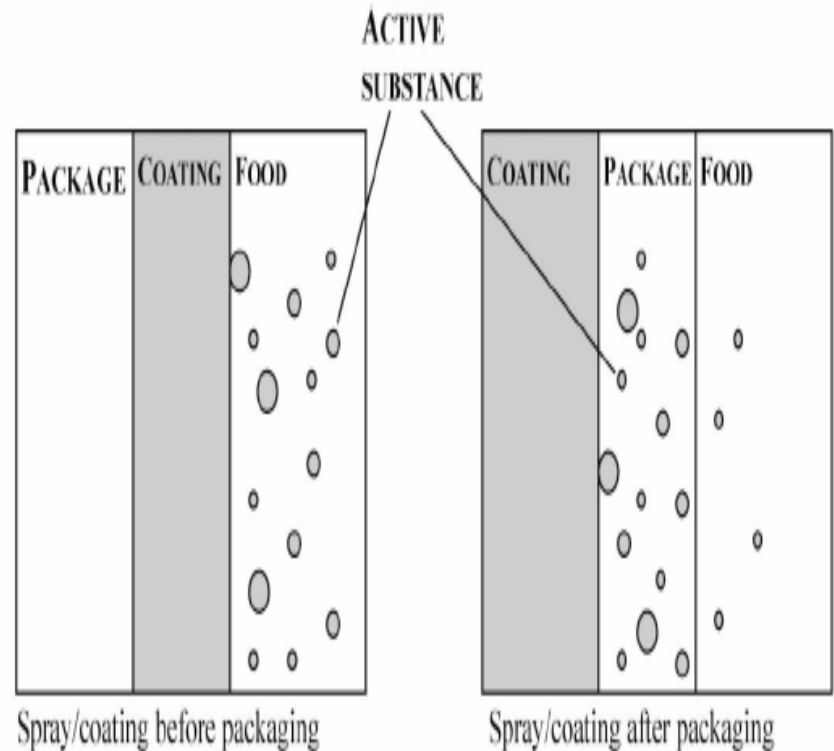
**Coating or adsorbing antimicrobials onto polymer surfaces**

**Incorporation antimicrobial agents directly into polymers**

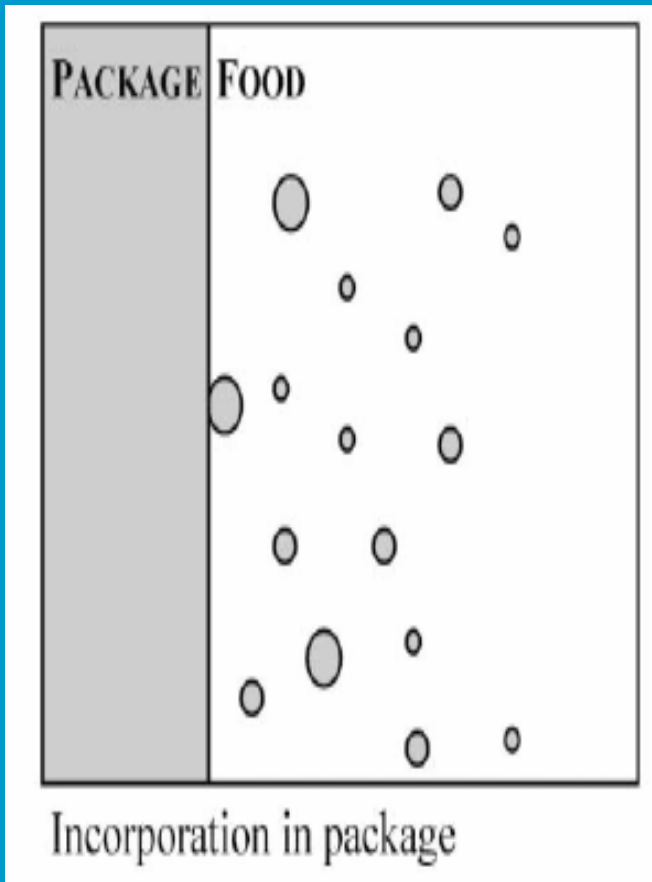


Chemical immobilization in package

**Immobilization of antimicrobial agent in package**



**Coating antimicrobial agent in package**



- ✓ In the mechanism incorporate antimicrobial agent, antimicrobial films might allow for migration of the antimicrobial to the film surface , therefore a continued antimicrobial effect at the food surface during extended exposure.
- ✓ Direct addition of antimicrobials to food will result in an immediate reduction of bacterial population.

# Incorporation Antimicrobial Agents

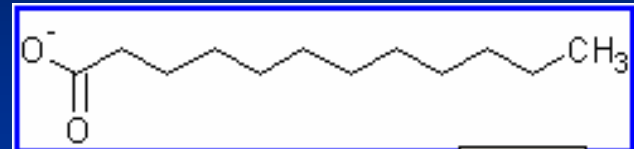
- © Films with low diffusion rates were desirable since they maintained higher surface concentrations for longer periods
- © The gradual release of an antimicrobial from a packaging film to the food surface may have an advantage over dipping and spraying
- © Antimicrobial activity may be rapidly lost due to inactivation of the antimicrobials by food components or dilution below active concentration due to migration into the bulk food matrix.

# Factors To Be Considered in the Manufacturing of Antimicrobial Films

- Casting process conditions and residual antimicrobial activity
- Characteristics of antimicrobial substances and foods.
- Storage temperature
- Chemical interaction of additives with film matrix
- Physical properties of packaging materials
- Mass transfer coefficients and modeling

# Antimicrobial Agent

## Lauric acid



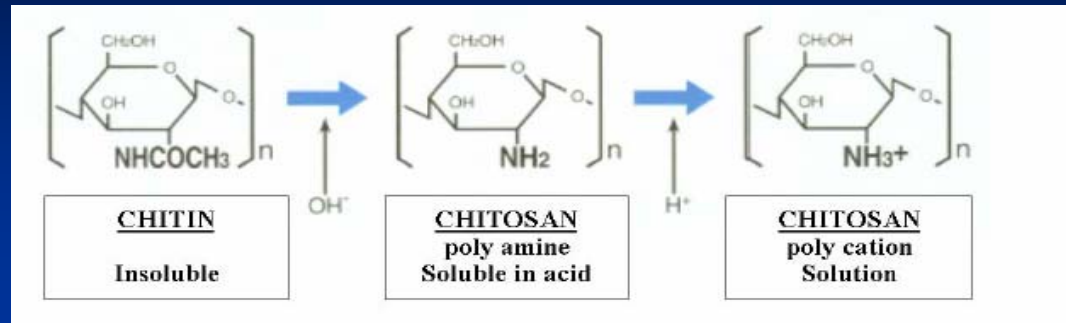
- ✓ Extract of coconut oils
- ✓ Is a medium-chain fatty acid
- ✓ Function : Antiviral, antibacterial and antiprotozoal
- ✓ Additional Beneficial : Formed into monolaurin in the human or animal body

According to the published research, Lauric Acid is one of the best "inactivating" fatty acids, and its monoglyceride is even more effective than the fatty acid alone (Kabara, 1978; Sands et al., 1978; Fletcher et al., 1985; Kabara, 1985).



# Film Based

## Chitosan



- Insoluble in water
  - Soluble in acidic solvent below pH 6.
  - Organic acid are used for dissolving chitosan
  - Chitosan solution's stability is poor above about pH 7
- Ⓢ Most commercially available chitosan preparations are more than 85 % deacetylated of chitin, and have molecular weights between 100kDa and 1000kDa.
- Ⓢ They are usually complexed with acids, such as acetic or lactic acids.
- Ⓢ An important property of chitosan is its positive charge in acidic solution.
- Ⓢ This is due to the presence of primary amines on the molecule that bind protons according to the equation.

**MATERIAL**  
**&**  
**METHODOLOGY**

# Methodology

```
graph TD; Methodology --> Film_solution[Film solution]; Methodology --> Broth_medium[Broth medium]; Methodology --> Food_simulant[Food Simulant]; Film_solution --> Film[Film]; Broth_medium --> Luria_Agar[Luria Agar]; Broth_medium --> Luria_Broth[Luria Broth]; Luria_Agar --> Agar_Diffusion_Test[Agar Diffusion Test]; Luria_Broth --> Liquid_Culture_test[Liquid Culture test];
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The diagram is a hierarchical flowchart. At the top is a box labeled 'Methodology'. A horizontal line with three downward-pointing arrows branches from 'Methodology' to three boxes: 'Film solution', 'Broth medium', and 'Food Simulant'. From 'Film solution', a vertical arrow points down to a box labeled 'Film'. From 'Broth medium', a horizontal line with two downward-pointing arrows branches to two boxes: 'Luria Agar' and 'Luria Broth'. From 'Luria Agar', a vertical arrow points down to a box labeled 'Agar Diffusion Test'. From 'Luria Broth', a vertical arrow points down to a box labeled 'Liquid Culture test'. All boxes are rounded rectangles with a light blue gradient and dark blue text.

**Film solution**

**Broth medium**

**Food Simulant**

**Film**

**Luria Agar**

**Luria Broth**

**Agar  
Diffusion  
Test**

**Liquid  
Culture test**

## Raw material

- ◆ Chitosan powder
- ◆ Lauric acid
- ◆ Glycerin
- ◆ Acetic Acid 1 %
- ◆ Luria Broth
- ◆ Luria Agar
- ◆ Food Simulant

## Equipment

- ◆ Petri disc
- ◆ Homogenizer
- ◆ Universal Bottles
- ◆ Incubator Shaker
- ◆ Spectrophotometer
- ◆ Incubator
- ◆ HPLC

# Media- LB Broth

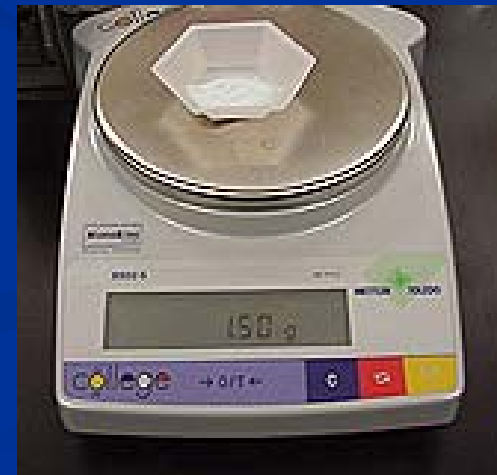
- In a bottle with a cap, dissolve 2.5 grams of Luria broth powder in 100 ml of distilled water.
- Cap the bottle and autoclave for 30 minutes.
- Refrigerate to prevent contamination.



# Media-LB Agar Plates

## Step 1

- To make 4 to 5 plates of Luria broth agar:
- Place 2.5 grams of LB powder and 1.5 grams of agar in 100 ml of distilled water.



# Media-LB Agar Plates

## Step 2

- ⓐ Autoclave until the solution is clear.



# Media-LB Agar Plates

## Step 3

- ⊗ Allow the LB agar solution to cool until you can just hold the bottle with your bare hand.
- ⊗ Pour the LB agar solution into 2 petri dishes and cover with the laminar flow.





# Media-LB Agar Plates

## Step 4

- Ⓢ Let stand 30 minutes to dry and then keep it into refrigerator.



# Food Simulant

Food Type	Food Simulant	Abbreviation
Aqueous Foods ( pH > 5 )	Distilled water	Simulant A
Acidic foods (pH <4,5)	Acetic acid 3 %	Simulant B
Alcoholic foods	Ethanol 10 % (v/v)	Simulant C
Fatty foods	olive oil	Simulant D

**RESULT**

**&**

**DISCUSSION**

# Film Formulation

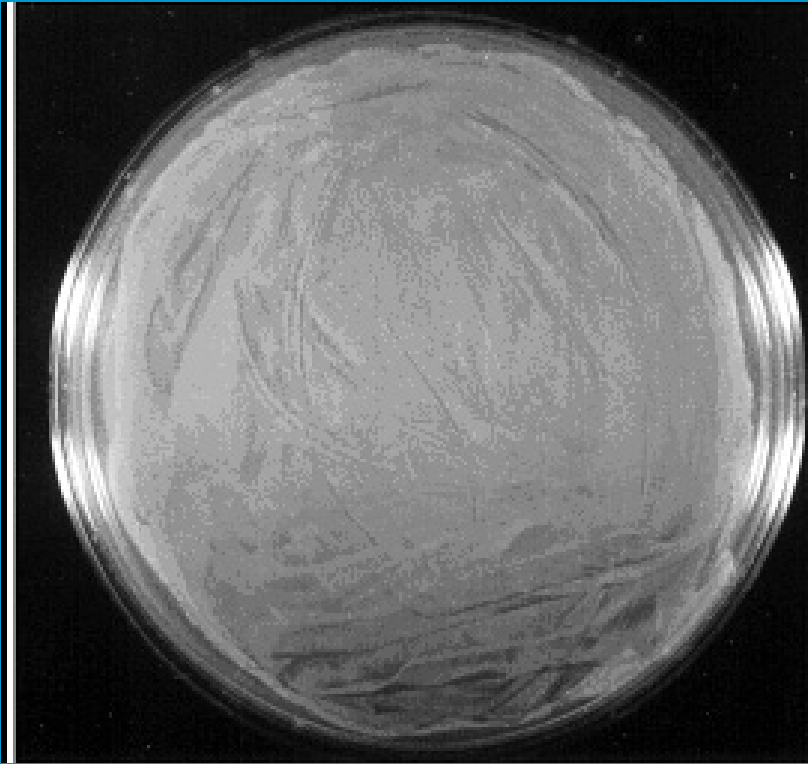
- Best formulation :
- Chitosan powder = 2g/100ml water
- Adding plasticizer Glycerin  
= 2 mL
- Acetic acid 1% as dilution for chitosan
- Casting volume = 20ml / Petri dish

# Film Appearance

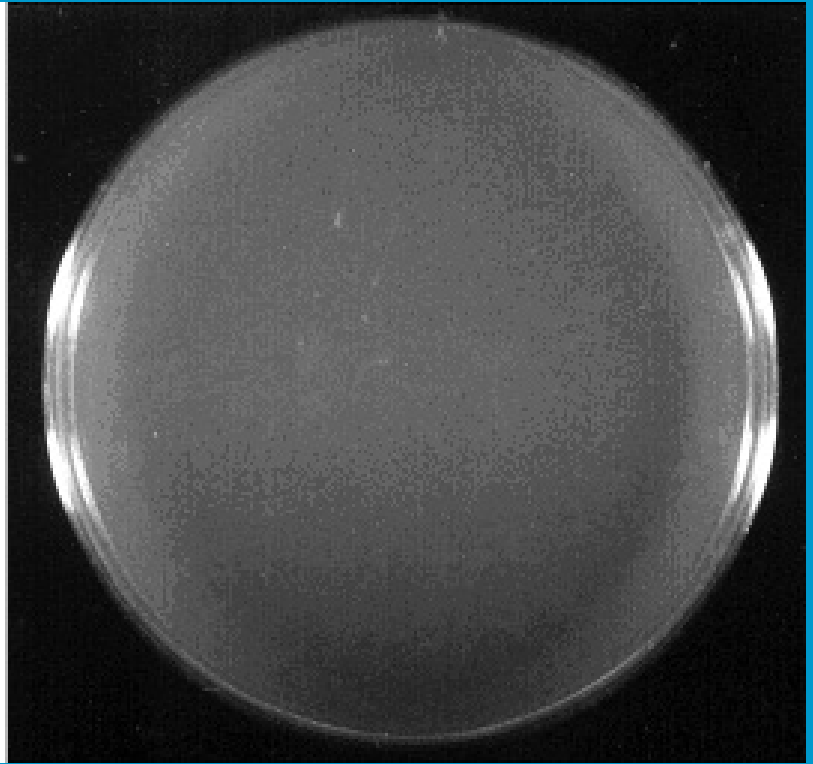
- ⊙ Yellow Transparent, homogenous, strength, less elastics
- ⊙ Concentration Lauric acid increase, film quality decrease



# Lawns of *E. coli* incubated at 37°C and 4°C



37°C



4°C

# **AGAR DIFFUSION TEST**



**B. subtilis 0.000 M**



**B. subtilis 0.625 x 10<sup>-2</sup> M**



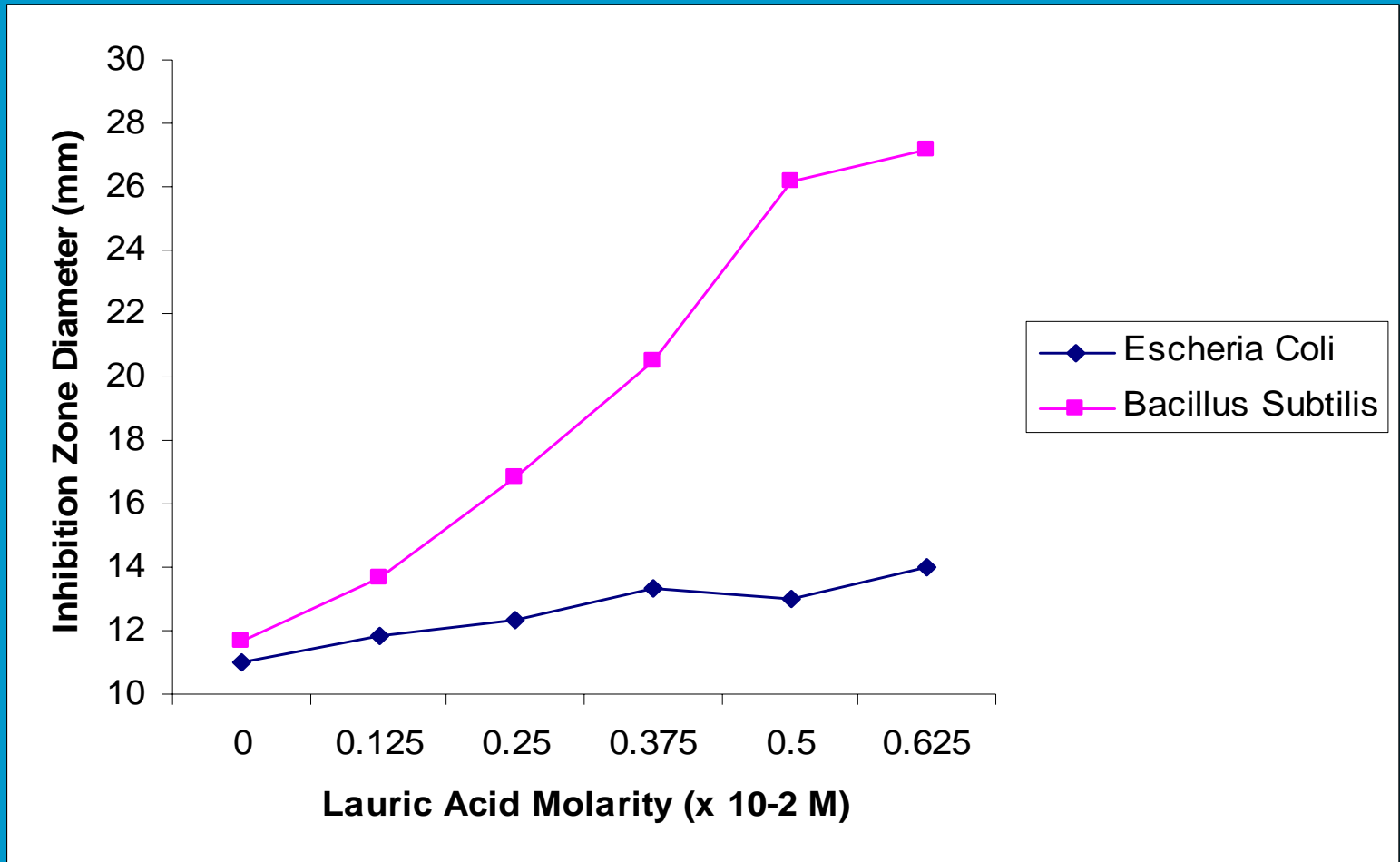
**E. coli 0.000 M**



**E. coli 0.375 x 10<sup>-2</sup> M**



# Comparison Inhibition Zone Between E.Coli and B.Subtilis



# Relation Between Inhibition Diameter with Concentration Of Lauric Acid

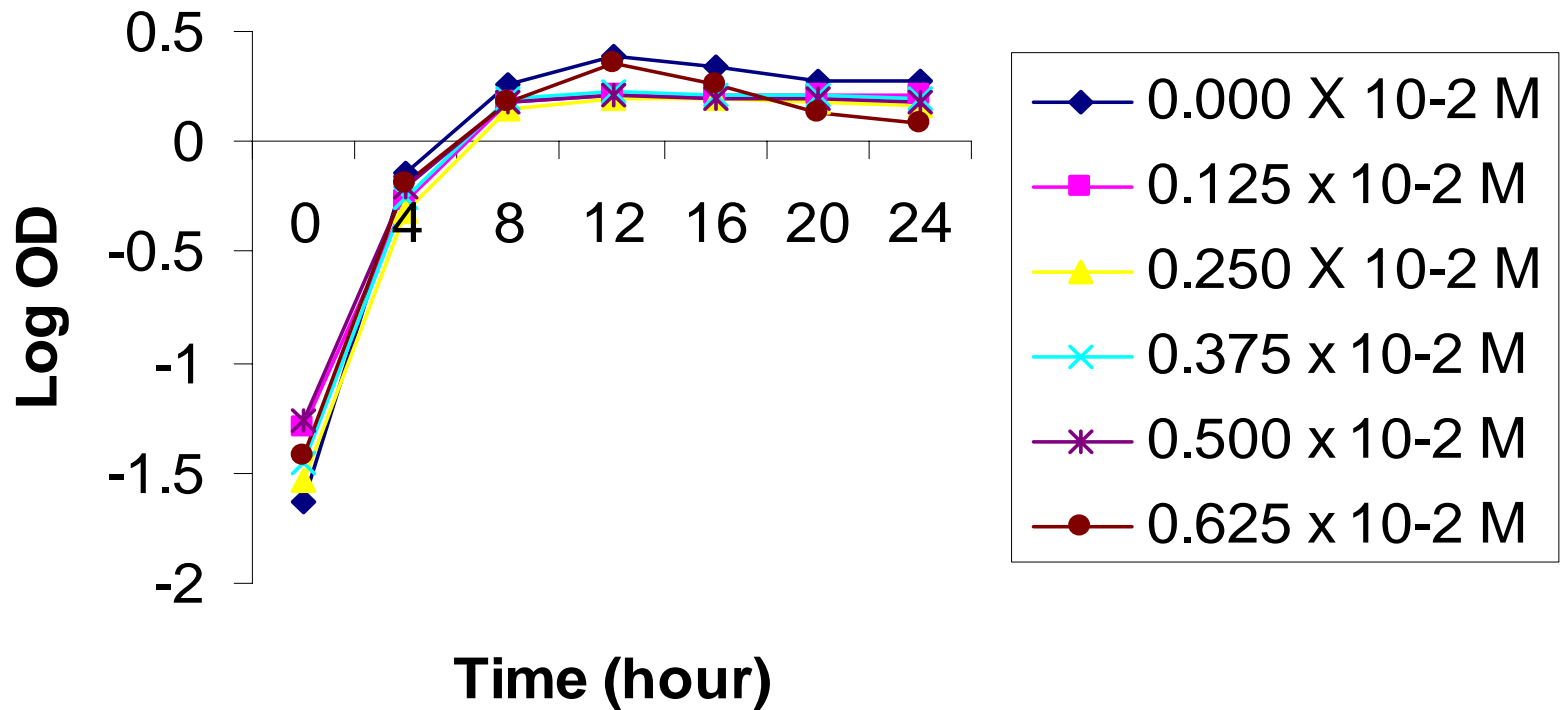
- Lauric acid had been used as the antimicrobial compound
- For E.Coli, it shows small impact on the inhibition growth after 24 hour of incubation
- For B.Subtilis, it shows that the concentration of Lauric acid in film increased, the inhibition area also increased . At highest concentration Lauric acid, the diameter become almost 3 times from

# Liquid Culture Test

The image shows the interior of a laboratory incubator. A metal tray is positioned inside, holding several glass bottles and petri dishes. The bottles have yellow caps, and the petri dishes appear to contain a liquid medium. The text 'Liquid Culture Test' is prominently displayed in the center-left of the image in a large, bold, pink font with a black outline. A blue wavy graphic element is on the right side of the image.

# Growth curve for *E.coli*

## Log OD Versus Time For E.Coli

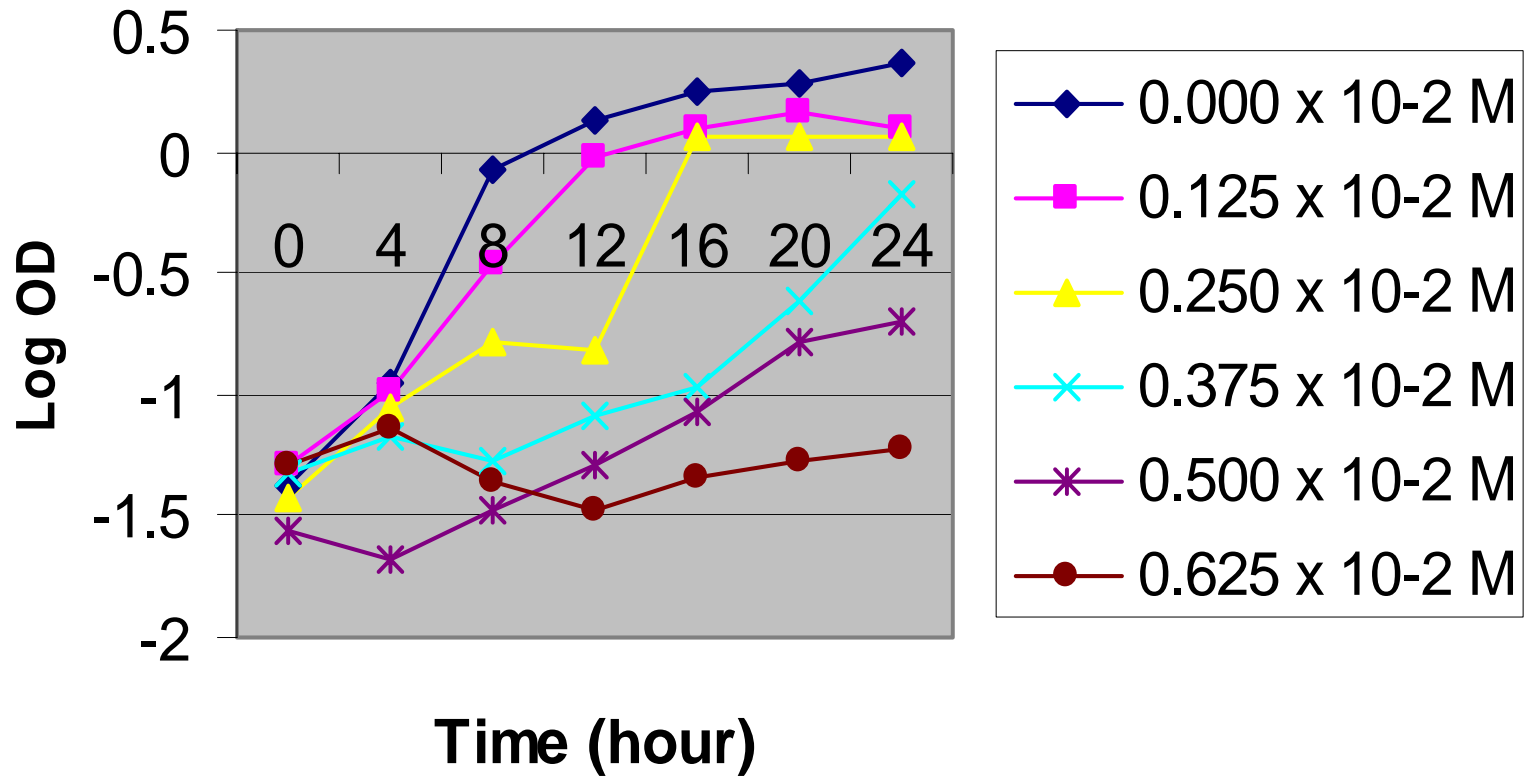


# Liquid Culture Test For E.Coli

- The growth curve are similar pattern for all different concentration of lauric acid used
- Growth of E.Coli unaffected by present of Lauric acid
- Deceleration phase at 10hours
- E.Coli has an outer membrane supported by a thin peptidoglycan layer

# Growth Curve for *B. Subtilis*

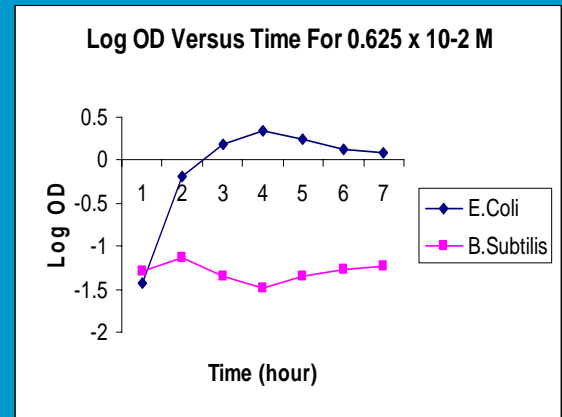
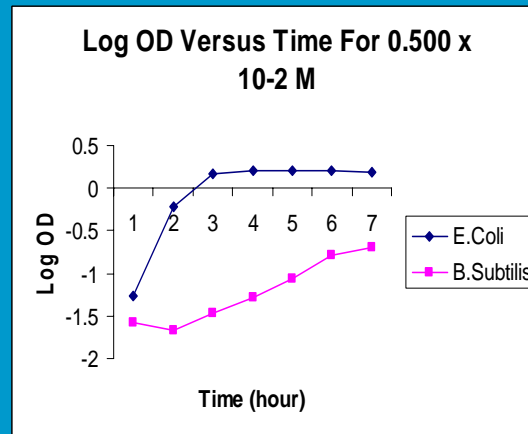
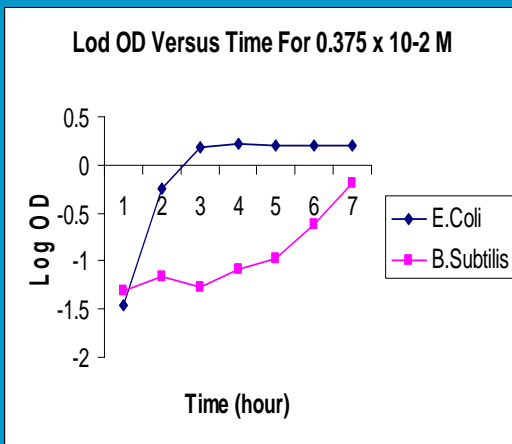
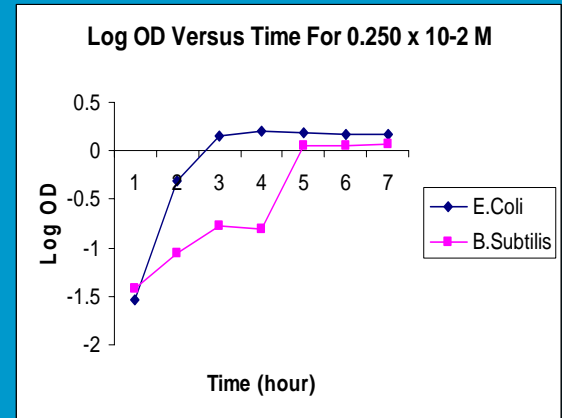
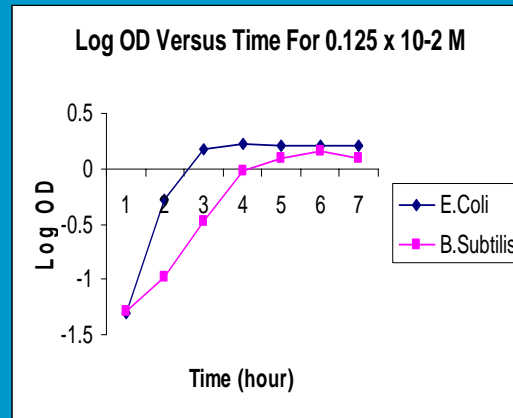
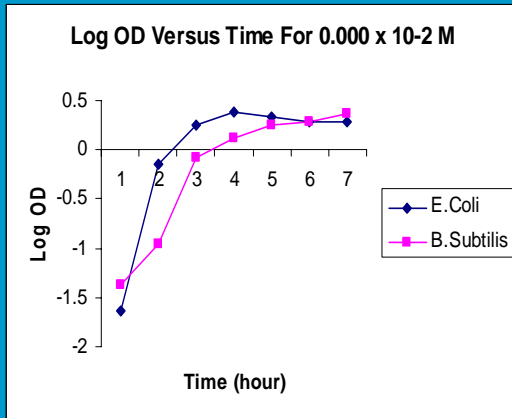
## Log OD Versus Time For B.Subtilis



# Liquid Culture Test For B.Subtilis

- Selected  $0.625 \times 10^{-2} \text{M}$  Lauric acid because it delay the lag phase for 12 hours
- Gram positive cell do not have outer membrane
- The structure of B.Subtilis easily to be destroyed or affected by lauric acid

# Comparison Growth Curve Of Both Bacteria





**CONCLUSION**

- ✓ Lauric acid has a good potential as antimicrobial agent
- ✓ Increasing of lauric acid concentration give small impact on E.Coli inhibition zone
- ✓ For B.Subtilis, clear zone diameter expand by increasing lauric acid concentration
- ✓ Lauric acid inhibit against gram positive microorganism
- ✓ Lauric acid did not change the growth curve pattern of E.Coli
- ✓ Based on B.Subtilis longest lag phase time, the most optimum concentration of Lauric acid is  $0.625 \times 10^{-2}$  M.

**RECOMMENDATION**

- For Lauric acid, a more suitable formulation is needed to get a smoother and higher quality film
- Further study can be carried out the physical and mechanical properties of the film such as thickness, strength, tensile and melting point of film
- Varies the use of bacteria and extend the time of incubation

# REFERENCE

- ✓ Alvarez, M. F. Review: Active food packaging, *Food Science & Technology International* 6:97-108
- ✓ Kabara JJ. Fatty acids and derivatives as antimicrobial agents: A review. In *The Pharmacological Effect of Lipids* (JJ Kabara, ed), *American Oil Chemists' Society*, Champaign IL, 1978
- ✓ Vojdani, F. et J.A. Toms. 1990, Potassium sorbate permeability of methylcellulose and hydroxypropyl methylcellulose coatings : effect of fatty acids. *J. Food Sci.* 55841-846

*I THANK*

*you*