## SCREENING OF SELECTED POLYPHENOL COMPOUNDS IN PALM OIL MILL EFFLUENT (POME)

### MOHD FAEZ BIN SHARIF

A dissertation submitted in partial fulfillment of the requirements for the award of the degree of Master of Science (Biotechnology)

Faculty of Biosciences and Bioengineering Universiti Teknologi Malaysia

NOVEMBER 2009

To my beloved father and mother, Mr. Sharif and Mrs. Fouziah Ali, sister, brother, friends and not to be forgotten Dr. Razauden Mohamed Zulkifli, thanks for all supports and motivations given...

#### ACKNOWLEDGEMENT

Thanks to God for granting me the chance and ability to successfully complete this project. In preparing this dissertation, many people have contributed their advices and knowledge towards my understanding and thoughts. This page is specially designed to express my appreciation to those peoples.

I wish to express my deepest gratitude to my dearest supervisor, Dr. Razauden Mohamed Zulkifli for all his efforts and valuable advice given throughout this project. Special thanks also to my co-supervisor, Dr. Norazah Basar who had guided me in conducting the thin layer chromatography and high performance liquid chromatography.

Not to be forgotten, my dearest parents; Mr. Sharif and Mrs. Fouziah Ali, brother and sister whom had support and pray for my success in completing this project. I am also indebted to all lectures of Bioscience and Bioengineering Faculty's, lab assistants and fellow friends for their guidance, advices and motivation. Without their continued support and interest, this thesis would not have been the same as presented here.

#### ABSTRACT

Total antioxidant activities and phenolic contents of raw and treated POME extracts (rPOME and tPOME respectively) were investigated. The total antioxidant activity was estimated using DPPH (2,2-diphenyl-1-picrylhydrazyl) assay. The total phenolic contents were measured using Folin-Ciocalteu assay. Thin layer chromatography and reversed-phase high-performance liquid chromatographic (RP-HPLC) separation methods with UV spectrophotometric detection were also used for the determination of compound of interest in POME extracts. Identification was based on retention times and UV spectra by comparison with commercial standards. The standards used were Gallic acid, Resveratrol, Quercetin and Kaempferol. The extracts were analyzed after extraction with ethanol. Resveratrol were identified in the rPOME while no detection of any four of the standards was showed in the tPOME. From the study, rPOME was clearly shown to have higher antioxidant activities and total phenolic contents than the tPOME. The highest percentage inhibition of the DPPH by rPOME was 89.5% and the total phenolic content measured was 6.5 mg of gallic acid equivalents (GAE). In conclusion, palm oil mill effluent could potentially become an alternative source of antioxidant.

#### ABSTRAK

Kajian mengenai aktiviti antioksidan dan kandungan bahan phenolik di dalam ekstrak bahan buangan kelapa sawit yang tidak dirawat dan dirawat (rPOME dan tPOME) telah dijalankan. Analisis DPPH (2,2-diphenyl-1-picrylhydrazyl) telah digunakan untuk menentukan aktiviti antioksidan yang terdapat di dalam ekstrak. Kandungan bahan phenolik di dalam ekstrak pula diukur menggunakan analisis Folin-Ciocalteu. Teknik Thin Layer Chromatography (TLC) dan teknik pemisahan Reversed-phase High Performance Liquid Chromatography (RP-HPLC) dengan pengesan spektrofotometri juga telah digunakan untuk mengenal pasti bahan-bahan yang terkandung di dalam ekstrak POME. Bahan-bahan yang terkandung di dalam ekstrak dikenalpasti berdasarkan perbandingan terhadap waktu tahanan dan spektrum UV bahan piawai. Bahan piawai yang digunakan sebagai perbandingan adalah Gallic acid, Resveratrol, Quercetin dan Kaempferol. Semua analisis dijalankan setelah sampel bahan buangan sawit diekstrak dengan pelarut ethanol. Resveratrol telah dikanalpasti terkandung di dalam rPOME manakala tPOME tidak mengandungi keempat-empat bahan piawai. Daripada kajian ini, nilai aktiviti antioksidan dan kandungan bahan phenolik didapati lebih tinggi di dalam rPOME berbanding tPOME. rPOME menunjukkan peratusan penghalangan aktiviti DPPH tertinggi oleh ekstrak sebanyak 89.5% dan jumlah keseluruhan kandungan bahan phenolik di dalam rPOME adalah 6.5 mg bersamaan piawai Gallic acid (GAE). Sebagai kesimpulan, bahan buangan kelapa sawit berpotensi sebagai sumber alternatif antioksidan.

### TABLE OF CONTENTS

CHAPTER

TITLE

PAGE

DECLARATION	ii
TITLE	iii
DEDICATION	v
ACKNOWLEDGEMENTS	vi
ABSTRACT	vii
ABSTRAK	viii
TABLE OF CONTENTS	ix
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xiv
LIST OF APPENDICES	xvii

### 1 INTRODUCTION

1.1 Background	1
1.2 Justification Rational of the Research	3
1.3 Objectives	3

# 2 LITERATURE REVIEW

2.1 Potentiality of POME as Reusable Products		
2.2 Polyphenolic Compounds	5	
2.2.1 Phenolic acids	6	
2.2.2 Medicinal Value of Resveratol		

and it's Commercial Potentials		
2.2.3 Neuroprotective Function of Flavonoids		
2.3 Antioxidant, Free Radical and DPPH Assay		
2.3.1 Mechanism Involved in DPPH		
(2,2-diphenyl-1-picrylhydrazyl) Assay	11	
2.4 Folin-Ciocalteu assay		
2.5 Extraction and Analysis of Polyphenol Compounds		

### 3 MATERIALS AND METHODS

3.1 POME Sampling	15
3.1.1 Determination of POME pH	15
3.1.2 Determination of Chemical Oxygen	
Demand (COD)	16
3.1.2.1COD dichromate and COD acid reagent	
Preparation	16
3.1.2.2 Determination of COD using	
Hach DR 4000 Spectrophotometer	17
3.1.3 Determination of Biochemical	
Oxygen Demand (BOD)	17
3.2 Sample Preparation for Polyphenol	
Measurement by Solvent Extraction	18
3.3 Free Radical Scavenging Activity (DPPH) Assay	19
3.3.1 Sample Preparation for DPPH Assay	19
3.3.2 Determination of Antioxidant Activity	
using DPPH Assay	19
3.4 Total Phenolic Content	20
3.4.1 Sample Preparation for Folin-ciocalteu	
Assay	20
3.4.2 Gallic Acid Stock and Sodium Carbonate	
Preparation for Folin-ciocalteu Assay.	21
3.4.3 Determination of Total Phenolic Content	
using Folin-ciocalteu Assay	21
3.5 Thin Layer Chromatography (TLC)	22

3.5.1 Preparation of TLC Plate		
3.5.2 Sample Preparation for Thin Layer		
Chromatography	22	
3.5.3 Identification of Polyphenol by		
Thin Layer Chromatography	23	
3.6 RP-HPLC		
3.6.1 Reagents, solvents and standard phenolics	23	
3.6.2 Sample preparation for RP-HPLC	24	
3.6.3 Screening on Selected Polyphenol		
Compounds from POME extracts		
by RP-HPLC	24	
3.7 Statistical analysis		

### 4 **RESULTS AND DISCUSSIONS**

4.1 Basic Parameter	26
4.2 Antioxidant activity	27
4.3 Total Phenolic Content	28
4.4 Thin Layer Chromatography (TLC)	30
4.5 RP-HPLC	31
4.5.1 Standards	31
4.5.2 Treated POME Extract	33
4.5.3 Raw POME Extract	34
4.5.3.1 rPOME Spike With Resveratrol	35
4.5.3.2 rPOME Spike With Gallic Acid	37

# 5 CONCLUSION

5.1	Conclusion	39
5.1	Conclusion	39

## REFERENCES

46

40

### LIST OF TABLES

TABLE NO.	TITLE	PAGE
4.1	Basic parameters of the rPOME and tPOME	26
4.2	DPPH scavenging activity of the POME extract and Vitamin C	28
4.3	Total phenolic content of the POME extracts	29
4.4	Color and Rf value of the spot detected by TLC	30

### LIST OF FIGURES

FIGURE NO	. TITLE	PAGE
2.1	Chemical structure of Gallic acid	7
2.2	Chemical structure of Resveratrol	7
2.3	Chemical structure of kaempferol (a) and quercetin (b)	9
2.4	Reduction of DPPH by antioxidant compound	12
4.1	Standards chromatograms of a) Gallic acid, b) Resveratrol, c) Quercetin and d) Kaempferol	32
4.2	Chromatogram of the treated POME extract	33
4.3	Chromatogram of the raw POME extract	34
4.3.1	Chromatogram of the a) resveratrol, b) rPOME extract before spiking with resveratrol and c) rPOME extract after spiking with resveratrol.	36
4.3.2	Chromatogram of the a) gallic acid, b) rPOME extract before spiking with gallic acid and c) rPOME extract after spiking with gallic acid.	38

### LIST OF ABBREVIATIONS

AgSO <sub>4</sub>	-	Silver sulphate
ApcI	-	Atmospheric pressure chemical ionization
BOD	-	Biochemical oxygen demand
°C	-	Degree celcius
CID	-	Collision-induced dissociation
Cm	-	Centimeters
COD	-	Chemical oxygen demand
DAD	-	Diod array detector
DNA	-	Deoxyribonucleic acid
DO	-	Dissolved oxygen
DPPH	-	2,2-diphenyl-1-picrylhydrazyl
ESI	-	Electrospray ionization
g	-	Gram
GAE	-	Gallic acid equivalent
g/kg	-	Gram per kilogram
GSH	-	Glutathione
GSSG	-	Glutathione disulfide
$H_2O_2$	-	Hydrogen peroxide
$H_2SO_4$	-	Sulphuric acid
HgSO <sub>4</sub>	-	Mercury (II) sulphate
$K_2Cr_2O_7$	-	Potassium dichromate

L	-	Liter	
LC/MS	-	Liquid chromatography coupled to mass spectrometry	
m <sup>3</sup>	-	Meter cubic	
mg	-	Miligram	
ml	-	Mililitre	
mg/kg	-	Miligram per kilogram	
mg/l	-	Miligram per liter	
MS	-	Mass spectrometry	
Na <sub>2</sub> CO <sub>3</sub>	-	Sodium carbonate	
NADPH	-	Nicotinamide adenine dinucleotide phosphate	
nm	-	Nanometer	
NMR	-	Nuclear magnetic resonance	
O <sub>2</sub> -•	-	Superoxide anion radicals	
•ОН	-	Hydroxyl radical species	
POME	-	Palm oil mill effluent	
$R_{\rm f}$	-	Retention factor	
ROS	-	Reactive oxygen species	
rpm	-	Rotary per minutes	
RP-HPLC	-	Reversed-phase high performance liquid chromatography	
rPOME	-	Raw palm oil mill effluent	
S.E.M	-	Standard error mean	
tPOME	-	Treated palm oil mill effluent	
TLC	-	Thin layer chromatography	
UV	-	Ultraviolet	
UV-vis	-	Ultraviolet-visible	
μl	-	Microliter	

μΜ	-	Micromolar
µg/ml	-	Microgram per milliliter
v/v	-	Volume per volume
wt	-	Weight

### LIST OF APPENDICES

## APPENDIX NO.

TITLE

### PAGE

46

1

Standard Gallic acid calibration curve

### **CHAPTER 1**

#### INTRODUCTION

#### 1.1 Background

The Malaysian palm oil industry is growing rapidly and is becoming a very important agriculture-based industry, where the country today is the world's leading producer and exporter of palm oil, replacing Nigeria as the chief producer since 1971 (Yusoff, 2006). However, wet process of palm oil milling consumes a large amount of process water. It is estimated that for 1 tonne of crude palm oil produced, 5-7.5 tonnes of water are required, and more than 50% of the water will end up as palm oil mill effluent (POME) (Ahmad *et al.*, 2003). Raw POME is a colloidal suspension containing 95-96% water, 0.6-0.7% oil and 4-5% total solids including 2-4% suspended solids that are mainly consisted of debris from palm fruit mesocarp generated from three main sources, namely sterilizer condensate, separator sludge and hydrocyclone wastewater (Borja and Banks, 1994). For a well-controlled conventional mill, about 0.9, 1.5 and 0.1m<sup>3</sup> wastewater are generated from sterilizer condensate, separator sludge and hydrocyclone wastewater, respectively, for each tonne of crude palm oil produced (Khalid and Wan, 1992).

In the year 2004, more than 40 million tonnes of POME was generated from 372 mills in Malaysia (Yacob *et al.*, 2006). If the effluent is discharged untreated, it can certainly cause considerable environmental problems (Davis and Reilly, 1980) due to its high biochemical oxygen demand (25,000 mg/l), chemical oxygen demand (53,630 mg/l), oil and grease (8370 mg/l), total solids (43,635 mg/l) as well as

suspended solids (19,020 mg/l) (Ma, 1995). Therefore, the palm oil mill industry in Malaysia is identified as the one that produces the largest pollution load into the rivers throughout the country (Hwang *et al.*, 1978).

The discharge of untreated POME creates adverse impact to the environment. However the idea of nurturing POME and its derivatives as valuable resources should not be dismissed. This is because POME contains high concentrations of protein, carbohydrate, nitrogenous compounds, lipids and minerals (Habib *et al.*, 1997) that may be converted into useful materials (Agamuthu and Tan, 1985). Several studies have been reported on the exploitation of POME and its derivative as fermentation media to produce antibiotic and bioinsecticide (Suwandi, 1991), solvents (Somrutai *et al*, 1996), polyhydroxyalkanoates (Hassan *et al*, 1996), organic acids (Yee *et al*, 2003) as well as enzymes. Furthermore it is also believed that POME contains polyphenolic compounds that play important role as antioxidant (Prasertsan *et al*, 1997).

Polyphenol is a group of chemical substances found in plant that can be characterized based on more than one phenol groups in the compound itself. Recently, polyphenol has been given a great attention due to huge market for food supplement but most importantly their ability to act as an effective antioxidant, anticancer properties, and beneficial cardiovascular effect and promotes long life (Cho *et al.*, 2006, Pearson et al., 2008, Howitz *et al.*, 2003, Valenzano *et al.*, 2006). Example of polyphenol derived from plant are Syringic acid, Coumaric acid, Ferulic acid, Resveratrol, Quercetin, Kaempferol and Isorhamnetin (Li *et al.*, 2009). Some of these polyphenol for instance already on the market for sale as a supplement such as Resveratrol. Resveratrol is found in red grape, red wines and onions in small quantity (Soleas *et al.*, 2000). However, the studies of the polyphenol content in POME have yet to be discovered.

#### **1.2 Justification Rational of the Research**

Waste substances generated from industries have becoming a major problem to the environment. Some wastes are hazardous and need to be removed immediately to avoid it from entering the food chain and polluting the ecosystem. Biorecycling or the potential usage of industrial waste in particular Palm Oil Mill Effluents (POME) is one of the solution other than treated the waste directly. It is estimated that an average about ten percent of raw POME is generated for every fresh fruit bunch processed. POME consists of water soluble component as well as suspended materials like palm fiber and oil. It is believed that this waste carrying secondary metabolite that can be extract and are useful for commercialization in particular polyphenol and flavanoids. If the potential polyphenols, were found and characterized, the waste can be turned into supplement to human and farm animal hence increased our health, capital and at the same time reducing the waste.

#### **1.3 Objectives**

The objectives of the research are:

- a) To measure the antioxidant activities of the POME extracts by DPPH (2,2 diphenyl-1-picrylhydrazyl) assay.
- b) To measure the amount of total polyphenol extracted from the POME by Folin-ciocalteu assay.
- c) To extract and identify different types of polyphenol present in the POME extracts by Thin Layer Chromatography (TLC) and Reversed-phase High Performance Liquid Chromatography (RP-HPLC).

#### REFERENCES

- Agamuthu, P., and Tan, E.L. (1985) Digestion of dried palm oil mill effluent by *Cellulomonas* sp. Microbiol. Lett. 30: 109–113.
- Aggarwal, B.B., Bhardway, A., Aggarwal, R.S., Seeram, N.P., Shishodia, S., and Takada, Y. (2004). Role of resveratrol in prevention and therapy of cancer: preclinical and clinical studies. Anticancer Res. 24, 2783–2840.
- Ahmad, A.L., Ismail, S., Bhatia, S. (2003) Water recycling from palm oil mill effluent (POME) using membrane technology. Desalination 157: 87–95.
- Antolovich, M., Prenzler, P.D., Patsalides, E., McDonald, S., and Robards, K. (2002). Methods for testing antioxidant activity. Analyst 127: 183–198.
- Barker, T.W and Worgan, J.T. (1981) The utilization of palm oil processing effluents as substrates for microbial protein production by the fungus Aspergillus oryzae. *European J Appl Microbiol Biotechnol*;11: 234–40.
- Baur, J.A. and Sinclair, D.A. (2006). Therapeutical potential of resveratrol: the in vivo evidence. Nature Rev. Drug Discov. 5: 493–506.
- Baur, J.A., Pearson, K.J., Price, N.L., Jamieson, H.A., Lerin, C., Kalra, A., Prabhu, V.V., Allard, J.S., Lopez-Lluch, G., Lewis, K., Pistell, P.J., Poosala, S., Becker, K.G., Boss, O., Gwinn, D., Wang, M., Ramaswamy, S., Fishbein, K.W., Spencer, R.G., Lakatta, E.G., Le Couteur, D., Shaw, R.J., Navas, P., Puigserver, P., Ingram, D.K., de Cabo, R., Sinclair, D.A. (2006). Resveratrol improves health and survival of mice on a high-calorie diet. Nature 444: 337–342.
- Bhat, K.P and Pezzuto, J.M. (2002) Cancer chemopreventive activity of resveratrol. Ann N Y Acad Sci. 957: 210–29.
- Borja, R., and Banks, C.J. (1994) Anaerobic digestion of palm oil mill effluent using up-flow anaerobic sludge blanket reactor. Biomass Bioenergy 6: 381–389.
- Bradamante, S., Barenghi, L., and Villa, A. (2004). Cardiovascular protective effects of resveratrol. Cardiovasc. Drug Rev. 22: 169–188.

- Cai, Y., Luo, Q., Sun, M. and Corke, H. (2004) Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. Life Sciences 74 pp. 2157–2184.
- Cho, Y.J., Hong J.Y., et al. (2006). Ultrasonication-assisted extraction of resveratrol from grapes. *Journal of Food Engineering* 77(3): 725-730.
- Clifford, MN. (1999) Chlorogenic acids and other cinnamates-nature, occurrence and dietary burden. *J Sci Food Agric*;79: 362-372.
- Clifford, M.N. and Scalbert, A. (2000) Ellagitannins-occurrence in food, bioavailability and cancer prevention. *J Food Sci Agric*; 80: 1118–25.
- Cuyckens, F. and Claeys, M. (2004). Mass spectrometry in the structural analysis of flavonoids. Journal of mass spectrometry : JMS 2004;39(1): 1-15.
- Czech-Kozbowska, M., Z. K. (1984). Phenolic Compounds and the Polyphenoloxidase and Peroxidase Activity in Callus Tissue Culture-Pathogen Combination of Red Raspberry and Didymella applanata (Niessl.) Sacc. *Journal of Phytopathology* 109(2): 176-182.
- Datla, K.P., Christidou, M., Widmer, W.W., Rooprai, H.K., and Dexter, D.T. (2001) Tissue distribution and neuroprotective effects of citrus flavonoid tangeretin in a rat model of Parkinson's disease. Neuroreport 12: 3871–3875.
- Davis, J.B., Reilly, and P.J.A. (1980) Palm oil mill effluent-a summary of treatment methods. Oleagineux 35: 323–330.
- Galli, R.L, Shukitt-Hale, B., Youdim, K.A., and Joseph, J.A. (2002). Fruit polyphenolics and brain aging: nutritional interventions targeting age-related neuronal and behavioral deficits. Ann N Y Acad Sci 959: 128–132.
- Habib, M.A.B., Yusoff, F.M., Phang, S.M., Ang, K.J., Mohamed, S. (1997). Nutritional values of chironomid larvae grown in palm oil mill effluent and algal culture. Aquaculture 158: 95–105.
- Hassan, M.A., Shirai, Y., Kusubayashi, N., Abdul Karim, M.I., Nakanishi, K., Hashimoto, K. (1996). Effect of organic acid profiles during anaerobic treatment of palm oil mill effluent on the production of polyhydroxyalkanoates by *Rhodobacter sphaeroides*. J. Ferment. Bioeng. 82: 151–156.
- Halliwell, B., (1994). Free radicals, antioxidants and human diseases: curiosity, cause and consequences. Lancet 344, 721–724.
- Herrmann, K. (1976). Flavonols and flavones in food plants: a review. J Food Technol;11:433–48.
- Howitz, K. T., Bitterman, K. J., et al. (2003). Small molecule activators of sirtuins extend Saccharomyces cerevisiae lifespan. Nature 425(6954): 191-196.

- Ho, C.C. and Tan, Y.K. (1983). Centrifugal fractionation studies on the particulates of palm oil mill effluent. Water Res; 17:613–8.
- Ho, C.C, Tan, Y.K and Wang, C.W. (1984). The distribution of chemical constituents between the soluble and the particulate fractions of palm oil mill effluent and its significance on its utilization/treatment. Agric Wastes;11: 61–71.
- Hsu, C.Y. (2006). Antioxidant activity of extract from Polygonum aviculare L. Biol. Res. 39, 281–288.
- Hwang, T.K., Ong, S.M., Seow, C.C., and Tan H.K. (1978). Chemical composition of palm oil mill effluents. Planter 54: 749–756.
- Ikawa, M., Schaper, T.D., Dollard, C.A., and Sasner, J.J. (2003). Utilization of Folin-Ciocalteu phenol reagent for the detection of certain nitrogen compounds. J. Agric. Food Chem. 51 (7): 1811–5.
- Inanami, O., Watanabe, Y., Syuto, B., Nakano, M., Tsuji, M. and Kuwabara, M. (1998). Oral administration of catechin protects against ischemia-reperfusioninduced neuronal death in the gerbil. Free Radic Res 29; 359–365.
- Jang, M., Cai, L., Udeani, G.O., Slowing, K.V., Thomas, C.F., Beecher, C.W.W., Fong, H.H.S., Farnsworth, N.R., Kinghorn, A.D., Mehta, R.G., Moon, R.C., and Pezzuto, J.M. (1997). Cancer chemopreventative activity of resveratrol, a natural product derived from grapes. Science 275, 218–220.
- Jaeschke, H. (1995). Mechanisms of oxidant stress-induced acute tissue injury. Proc. Soc. Exp. Biol. Med. 209, 104–111.
- Khalid, A.R., Wan, W.A., Mustafa. (1992). External benifits of environmental regulation: resource recovery and the utilization of effluents. The Environmentalist 12: 277–285.
- Li, G. Beutel, M. Rhein, J. Meyer, C. Koenecke, T. Neumann, M. Yang, J. Krauter, N. von Neuhoff, M. and Heuser. (2009). Murine Leukemia Induced by Retroviral Gene Marking. Science magazine, 12: 27–28.
- Lopez, M., Martinez, F., Valle, C.D., Ferrit, M., and Luque, R. (2003). Study of phenolic compounds as natural antioxidants by a fluorescence method. Talanta 60, 609–616.
- Luo, Y., Smith, J.V., Paramasivam, V., Burdick, A., Curry, K.J., Buford, J.P., Khan, I., Netzer, W.J., Xu, H. and Butko, P. (2002). Inhibition of amyloid-beta aggregation and caspase-3 activation by the Ginkgo biloba extract EGb761. Proc Natl Acad Sci U S A 99, 12197–12202.
- Ma A.N. (1995). A novel treatment for palm oil mill effluent. Palm Oil Res. Instit. Malaysia (PORIM) 29: 201–212.

- Macheix, J.J, Fleuriet A. and Billot, J. (1990). Fruit phenolics. Boca Raton, FL: CRC Press.
- Mansouri A., Embarek G., Kokkalou E. and Kefalas P. (2005). Phenolic profile and antioxidant activity of the Algerian ripe date palm fruit (Phoenix dactylifera), Food Chemistry 89 pp. 411–420.
- Markham, K.R. (1982) Techniques of Flavonoid Identification. Academic Press, London.
- Middleton, E., Kandaswami, C. and Theoharides, T.C. (2000). The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. Pharmacol Rev; 52:673–751.
- Neuzil, J., Gebicki, J.M., and Stocker, R. (1993). Radical-induced chain oxidation of proteins and its inhibition by chain-breaking antioxidants. *Biochem. J.* 293, 601–606.
- Oshima, Y., Namao, K., Kamijou, A., Matsuoka, S., Nakano, M., Terao, K., and Ohizumi, Y. (1995). Powerful hepatoprotective and hepatotoxic plant oligostilbenes, isolated from the Oriental medicinal plant Vitis coignetiae (Vitaceae). Experientia 51, 63–66.
- Pearson, K. J., Baur, J. A., et al. (2008). Resveratrol Delays Age-Related Deterioration and Mimics Transcriptional Aspects of Dietary Restriction without Extending Life Span. Cell Metabolism 8(2): 157-168.
- Price, S.F., Breen, P.J., Valladao, M. and Watson, B.T. (1995). Cluster sun exposure and quercetin in Pinot noir grapes and wine. *Am J Enol Vitic*; 46: 187–94.
- Piver, B., Berthou, F., Dreano, Y., and Lucas, D. (2003). Differential inhibition of human cytochrome P450 enzymes by viniferin, the dimer of resveratrol: comparison with resveratrol and polyphenols from alcoholized beverages. Life Sci. 73, 1199–1213.
- Prasertsan, P., Kittikul, A. H, Kunghae, A., Maneesri, J., Oi S. (1997). Optimization for xylanase and cellulase production from *Aspergillus niger* ATCC 6275 palm oil mill wastes and its application. *World J. Microbiol. Biotechnol.* 13: 555–559.
- Privat, C., Telo, J.P., Bernardes-Genisson, V., Vieira, A., Souchard, J.P., and Nepveu, F. (2002). Antioxidant properties of trans-ε-viniferin as compared to stilbene derivatives in aqueous and nonaqueous media. J. Agric. Food Chem. 50, 1213–1217.
- Quiney, C., Dauzonne, D., Kern, C., Fourneron, J.D., Izard, J.C., Mohammad, R.M., Kolb, J.P., and Billard, C. (2004). Flavones and polyphenols inhibit the NO pathway during apoptosis of leukemia B-cells. Leuk. Res. 28, 851–861.

- Reich, E. and Schibli, A. (2006). High-Performance Thin-layer Chromatography for the Analysis of Medicinal Plants, Thieme, New York.
- Rincon, B., Raposo, F., Dominguez, J.R., Millan, F., Jimenez, A.M., Martin, A. and Borja, R. (2006). Kinetic models of an anaerobic bioreactor for restoring wastewater generated by industrial chickpea protein production. Biodeterioration and Biodegradation 57, 114–120.
- Romero-Perez, A. I., Lamuela-Ravento's, R. M., Andre's-Lacueva, C., and Carmen de la Torre-Boronat, M. (2001). Method for the quantitative extraction of resveratrol and piceid isomers in grape berry skins: effect of powdery mildew on the stilbene content. *Journal of Agricultural and Food Chemistry*, 49, 210–215.
- Satoh, M.S., Jones, C.J., Wood, R.D., and Lindahl, T. (1993). DNA excision repair defect of xeroderma pigmentosum prevents removal of a class of oxygen free radical-induced base lesions. Proc. Natl. Acad. Sci. USA 90, 6335–6339.
- Shahidi, F., and Naczk, M. (2004). Food and Nutraceuticals: Sources, Applications and Health Effects. CRC Press, Boca Raton.
- Shahidi, F. and Naczk, M. (1995). Food phenolics, sources, chemistry, effects, applications. Lancaster, PA: Technomic Publishing Co Inc.
- Singleton, V.L., Orthofer, R., and Lamuela-Raventos, R.M. (1999). Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Meth Enzymol; 299: 152-178.
- Soleas, G., Grass L., et al. (2000). Resveratrol, a red wine constituent, has anticarcinogenic properties. Clinical Biochemistry 33(3): 240-240.
- Somrutai, W., Takagi, M., and Yoshida, T. (1996). Acetone-butanol fermentation by *Clostridium aurantibutyricum*ATCC17777 from a model medium for palm oil mill effluent. *J. Ferment. Bioeng.* 81: 543–547.
- Stohs, S.J., and Bagchi, D., (1995). Oxidative mechanisms in the toxicity of metal ions. J. Free Radic. Biol. Med. 18 (2), 321–336.
- Suwandi, M.S. (1991). POME, from waste to antibiotic and bioinsecticide. Jurutera Kimia Malaysia 1: 79–99.
- Tomas, F.A. and Clifford, M.N. (2000). Dietary hydroxybenzoic acid derivatives and their possible role in health protection. *J Sci Food Agric*; 80:1024–32.
- Tseng, Y.M., Lin, S.K., Hsiao, J.K., Chen, I.J., Lee, J.H., Wu, S.H., and Tsai, L.Y. (2006). Whey protein concentrate promotes the production of glutathione (GSH) by GSH reductase in the PC12 cell line after acute ethanol exposure. Food Chem. Toxicol. 44, 574–578.

- Valenzano, D. R., Terzibasi E., et al. (2006). Resveratrol Prolongs Lifespan and Retards the Onset of Age-Related Markers in a Short-Lived Vertebrate. Current Biology 16(3): 296-300.
- Vinson, J., Zubik, L., Bose, P., Samman, N., and Proch, J. (2005). Dried fruits: excellent in vitro and in vivo antioxidants. *J Am Coll Nutr* 24 (1): 44–50.
- Wang, C.W, Chong, C.N. and Rahim, B. (1981). Growth of SCP on hydrolysate of palm oil sludge. Proceedings of the First ASEAN Workshop on the Technology of Animal Feed Production Utilizing Food Waste Materials. Bandung; p. 1-13.
- Yacob, S., Shirai, Y., Hassan, M.A., Wakisaka, M., and Subash, S. (2006). Start-up operation of semi commercial closed anaerobic digester for palm oil mill effluent treatment. Process Biochem. 41: 962–964.
- Yanez, M., Fraiz, N., Cano, E., and Orallo, F. (2006). Trans-ε-viniferin, a polyphenol present in wines, is an inhibitor of noradrenaline and 5-hydroxytryptamine uptake and of monoamine oxidase activity. *Eur. J. Pharmacol.* 542, 54–60.
- Yean-Yean, S. and Philip, J. B. (2004). Antioxidant activity and phenolic content of selected fruit seeds. Food Chemistry. 88 (3): 411-417.
- Yee, P.L., Hassan, M.A., Shirai, Y., Inagaki, M., Nakanishi, K., and Hasimoto, K. (2003). Continuous production of organic acids from palm oil mill effluent with sludge recycle by the freezing thawing method. J. Chem. Eng. Japan 36: 707–710.
- Yusoff. (2006). Renewable energy from palm oil innovation on effective utilization of waste. *J. Cleaner Product* 14: 87–93.
- Zhao, F. P., Strack, D., Baumert, A., Ramanathan, S., Ngoh, K. G., Tet, F. C. and Swee, N. T. (2003). Antioxidant Flavonoids from Leaves of Polygonum hydropiper L. Phytochemistry. 62: 219-228.
- Zheng W. and Wang S.Y. (2001). Antioxidant activity and phenolic compounds in selected herbs. *Journal of the Agricultural and Food Chemistry* 49, pp. 5165– 5170.
- Zhou, K. and Yu, L. (2006). Total phenolic contents and antioxidant properties of commonly consumed vegetables grown in Colorado. LWT 39: 1155-1162.