

**OPTIMIZATION OF PROCESSING PARAMETERS ON THE YIELD OF  
OLEORESIN FROM *ZINGIBER ZERUMBET* AND ITS ANTIBACTERIAL  
ACTIVITY**

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OLEORESIN FROM *ZINGIBER ZERUMBET* AND ITS ANTIBACTERIAL  
ACTIVITY

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*Dedicated to my beloved Husband, Mak, Abah, Ariff, Iman and Affiq*

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## ABSTRACT

The effect of processing parameters on the yield of oleoresin from *Zingiber zerumbet* and its antibacterial activity was investigated. The preliminary experiments were carried out to determine the main parameters that affect the yield of oleoresin, zerumbone content and antibacterial activity on the *Escherichia coli* and *Staphylococcus aureus* of *Z. zerumbet* oleoresin. The main parameters were found to be extraction time, type of solvent used and blanching treatment. The range of those parameters that was investigated to get the highest of all yield were blanching treatments (untreated, boiled, steamed), solvent (acetone, ethanol and methanol) and extraction time (6 hours to 18 hours). A D-optimal design was employed to study the effect of different range parameter of *Z. zerumbet* oleoresin on four response variables; the yield of oleoresin, the zerumbone content, the antibacterial activity on the *E. coli* and *S. aureus*. Analysis of variance and response surface methodology were applied to identify the optimal processing parameter. The optimal processing parameters that fulfilled the requirement for yield of oleoresin, zerumbone content, antibacterial activity on the *E. coli* and *S. aureus* with high desirability were found to be 18 hours of extraction time, methanol as the solvent used and steaming as the blanching treatment. The desirability value was 84 %. Meanwhile, the optimal yield of *Z. zerumbet* oleoresin was 31.50 % (w/w), the zerumbone content was 0.56 % (w/w) and the antibacterial activities (inhibition zone) on the *E. coli* and *S. aureus* were 14.58 mm and 13.51 mm respectively. Processing optimization has resulted in an increase of overall antibacterial activity on the *E. coli* and *S. aureus* of *Z. zerumbet* oleoresin as well as the increment yield of oleoresin and zerumbone content.

## ABSTRAK

Kesan parameter proses terhadap hasil oleoresin lempoyang (*Zingiber zerumbet*) dan aktiviti antibakterianya telah dikaji. Ujian awal dilakukan untuk menentukan parameter-parameter utama yang mempengaruhi hasil oleoresin, kandungan zerumbon serta aktiviti antibakteria terhadap *Escherichia coli* dan *Staphylococcus aureus* dalam oleoresin lempoyang. Parameter-parameter utama yang bersesuaian adalah rawatan penceluran, pelarut dan masa pengekstrakan. Julat parameter-parameter tersebut yang dikaji bagi mendapatkan keseluruhan nilai hasil yang tertinggi terdiri daripada rawatan penceluran (stim, rebus dan segar), pelarut (aseton, etanol dan metanol) dan masa pengekstrakan (6 jam hingga 18 jam). Satu kaedah rekabentuk *D-optimal* telah digunakan untuk mengkaji kesan perbezaan parameter proses oleoresin lempoyang terhadap empat pembolehubah respon; hasil oleoresin, kandungan zerumbon, aktiviti antibakteria terhadap *E. coli* dan *S. aureus*. Analisis varians dan kaedah tindak balas permukaan telah diaplikasikan untuk mengenalpasti parameter yang optimal. Parameter-parameter proses optimal yang dikenalpasti memenuhi kecenderungan ideal yang tinggi bagi hasil oleoresin, kandungan zerumbon dan aktiviti antibakteria terhadap *E. coli* dan *S. aureus* adalah masa pengekstrakan selama 18 jam, metanol sebagai pelarut dan stim sebagai rawatan penceluran. Keadaan ideal yang dinilai adalah sebanyak 84 %. Sementara itu, hasil kajian mendapati hasil optimal oleoresin ialah 31.50 % (w/w), kandungan zerumbon ialah 0.56 % (w/w), aktiviti antibakteria (zon perencatan) terhadap *E. coli* ialah 14.58 mm dan *S. aureus* ialah 13.51 mm. Parameter-parameter proses yang meningkatkan aktiviti antibakteria terhadap *E. coli* dan *S. aureus* dalam oleoresin lempoyang telah meningkatkan hasil oleoresin dan kandungan zerumbon.

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## NOMENCLATURE

%	-	Percentage
$\mu\text{g/ml}$	-	Microgram per milliliter
$\mu\text{L}$	-	Microlitre
$^{\circ}\text{C}$	-	Degree Celsius
ANOVA	-	Analysis of Variance
CFU/ml	-	Colony Forming Unit/milliliter
DOE	-	Design of Experiment
<i>E. coli</i>	-	<i>Escherichia coli</i>
FID	-	Flame ionization detector
g	-	Gram
g/L	-	Gram per litre
GC	-	Gas Chromatography
$\text{gmol}^{-1}$	-	Gram per mole
hr	-	Hour
I	-	Intermediate
kg	-	Kilogram
mg/g	-	Milligram per gram
ml	-	Millilitre
mm	-	Millimeter
MS		Spectrophotometer and Mass Spectra
ppm	-	Part per million
psi	-	Pounds per square inch
R	-	Resistance
$R^2$	-	Coefficient of multiple determinations

RSM	-	Response surface methodology
S	-	Susceptible
<i>S. aureus</i>	-	<i>Staphylococcus aureus</i>
SFE	-	Supercritical fluid extraction
TLC	-	Thin Layer Chromatography
UV	-	Ultraviolet
VPC	-	Vapor phase chromatography
w/w	-	Weight per weight
<i>Z. zerumbet</i>	-	<i>Zingiber zerumbet</i>
ZER	-	Zerumbone

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## CHAPTER 1

### INTRODUCTION

#### 1.1 Introduction of Research

The emergence of new infectious diseases, the resurgence of several infections that used to be under controlled and the increase in bacterial resistance have created the necessity for studies directed towards the development of new antimicrobials. Considering the failure to acquire new molecules with antimicrobial properties from microorganisms, the optimization for screening methods used for identification of antimicrobials from other natural sources is of great importance (Valgas *et al.*, 2007). Moreover, natural products are widely recognised in the pharmaceutical industry for their broad structural diversity as well as their wide range of pharmacological activities (William, 2000).

Currently, one of the natural sources that receive consumer attention is the use of phytochemical extract as antibacterial agent. *Zingiberaceae* species are among the most prolific plants in the tropical rainforests. Somchit and Nur Shukriyah, 2003 reported that plants from *Zingiberaceae* family have anti-inflammatory, antiulcer, antioxidant and antimicrobial properties. Among these

species, *Z. zerumbet* is one of the most commonly used ingredient in Indo-Malaysian traditional medicines, health supplements and tonics (Ruslay *et al.*, 2007). In view of its popularity among the locals, it is important that the chemical profiles of this species be established as the marker compounds, as well as to validate the rationale for their efficacies (Ruslay *et al.*, 2007).

*Z. zerumbet* which is also known as Lempoyang contain active ingredient that act as anti-inflammatory, antitumor and antimicrobial (Somchit and Nur Shukriyah., 2003). According to Sanagi (1994), there are at least 10 groups of chemicals constituents present in *Z. zerumbet*. Zerumbone (ZER) was identified as the active ingredient in *Z. zerumbet*. It is a monocyclic sesquiterpene in *Z. zerumbet* which recently been found to suppress tumor promoter 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced Epstein-Barr virus activation in a potent manner (Murakami *et al.*, 2002). Murakami *et al.*, 2002, also indicate that zerumbone is a food phytochemical that has distinct potentials for use in anti-inflammation, chemoprevention and chemotherapy strategies. In addition, *Z. zerumbet* reported to have the properties of antibacterial towards gram positive strains such as *S. aureus* and *Micrococcus luteus* (Azmi Muda *et al.*, 2001). Takashi *et al.*, 2001 also found that certain products derived from zerumbone selectively inhibited the growth of Gram-positive bacteria.

With increasing awareness on the role of antibacterials in phytochemical extract, their prevention during processing has become increasingly important. Processing is known to affect content, activity and bioavailability of bioactive compounds (Hashim, 2002). The choice of processing methods and parameters is important in minimizing the nutrient losses during processing (Ang *et al.*, 1982). As a result, a lot of developments in phytochemical processing have been directed toward the preservation of nutritional contents such as micronutrients, antioxidant and bioactive constituents (Nicoli *et al.*, 1999). Plant materials with retained antimicrobials are more functional towards improving the shelf life of the product

and providing the health promotion compared to extracts whose antimicrobials have been removed or destroyed during processing (Smith and Paton, 2002).

Previous researchers have been applying the strategy of ‘reconstitution’, achieved by the addition of natural antibacterials to the product in order to minimize the eventual processing damages (Zivanovic *et al.*, 2005 and Burt, 2004). However, the strategy has succeeded in explaining the effects of the addition of natural antibacterials on the product rather than the processing effects on the natural antibacterials presents in the phytochemical extracts.

Based on these observations, the aim of the present study was to investigate the effect of processing parameters on the yield of *Z. zerumbet* oleoresin, zerumbone content and antibacterial activity towards *E. coli* and *S. aureus*.

## **1.2 Research Background**

Oleoresins are solvent extractives of spice or herbs. They are concentrated liquid products that contain both volatile and non-volatile flavour components. The quality of oleoresins is typically evaluated based on the presence of active ingredients at the desired level (Hashim, 2002). Therefore, the choice of processing parameters is important in minimizing the compounds losses during processing (Ang *et al.*, 1982).

The main processing parameters of *Z. zerumbet* oleoresin are solvents, extraction time and blanching treatments. There are many choices of solvents that can be used in the extraction, such as n-hexane, short chain alcohol (methanol,

ethanol), ketone (acetone), hydrocarbons (methylene dichloride) and liquid carbon dioxide (Tiwari, 1995). The two most common and widely used blanching methods are steam blanchers and hot-water blanchers (Hashim, 2002), while the extraction time differs according to the phases of experiments involve (Hashim, 2002).

At present, the processing parameters of oleoresin are as follows (Hashim, 2002):

- i. Blanching treatment – steam treated, boil treated and untreated
- ii. Solvent extraction – methanol, ethanol, acetone and hexane
- iii. Extraction time – 6 to 38 hours

### **1.3 Research Overview**

In this project, yield of oleoresin, zerumbone content, antibacterial activity on the *E. coli* and *S. aureus* are the most important properties to be considered in the extraction of a *Z. zerumbet* oleoresin. In order to obtain the most appropriate parameter, statistical experimental designs were used. Design Expert Version 6.0.8 software was used to design and analyze the experiments (Stat-ease Inc., 2002). To establish the possible range of processing parameters, several preliminary experiments were carried out. These experiments were to determine the most suitable range of the parameters and identify the significant factors that affect the oleoresin quality. Statistical software of D-optimal Design Techniques was used to design the experiment.



The D-optimal design techniques were used to obtain the mathematical and polynomial relationship between the factors and the response variables to create a mathematical model from the data (Cornell, 1990). These factors are the significant ingredients that had been identified during the initial experiments. D-optimal design was used to determine the combination of the significant ingredients to obtain the best parameters in term of yield of oleoresin, zerumbone content, antibacterial activity on the *E. coli* and *S. aureus* of oleoresin.

There are four responses that were considered in this research; the yield of oleoresin, zerumbone content, antibacterial activity on the *E. coli* and *S. aureus*. All properties were determined and the result was used for further calculation using D-optimal Design Techniques. The optimization step was carried out in order to determine the optimal processing parameter of *Z. zerumbet* oleoresin. The result of optimization work was analyzed using ANOVA (analysis of variances) table as well as representation by graph or contour graph (Stat-ease Inc., 2002).

This method is an alternative to the conventional method of one-factor at a time production optimization which is time-consuming, expensive and inaccurate as the existence of interactions between various factors occurs. Statistical experimental designs allow simultaneous, systematic and efficient variation of all components. The use of user-friendly software packages has made this technique increasingly popular for production optimization (Indrani and Rao, 2001).

#### **1.4 Research Objectives**

The objective of this research was to study the effect of processing parameters on the yield of oleoresin from *Z. zerumbet* and its antibacterial activity.

## 1.5 Research Scopes

To achieve the objectives, four scopes have been identified in this research:

- (i) To study the effect of processing parameters on the yield of oleoresin.
- (ii) To study the effect of processing parameters on the zerumbone content.
- (iii) To study the effect of processing parameters on the antibacterial activity on *E. coli* and *S. aureus*.
- (iv) To optimize the yield of oleoresin, the zerumbone content and the antibacterial activity on *E. coli* and *S. aureus* using D-optimal Design Techniques.

The processing parameters studied were the blanching treatments, solvent and extraction time. Experiments were conducted according to an experimental design generated by Design Expert Version 6.0.8. Each sample obtained from each run of experiments was evaluated for the yield of oleoresin, zerumbone content and antibacterial activity on *E. coli* and *S. aureus*.

## 1.6 Contribution of the Study

The study of natural antibacterial nowadays becoming increasingly important in the developed countries due to the most available antibacterial was failure to treat infectious diseases. In addition, the study of *Z. zerumbet* oleoresin as antibacterial is an important contribution as it is one of the most important materials in Malaysian traditional medicine. Zerumbone is a major component of *Z. zerumbet* that has distinct potentials for use in anti-inflammation,

chemoprevention and chemotherapy strategies. This study would help to understand the effect of processing parameter towards yield of oleoresin, zerumbone content and antibacterial activity on *E. coli* and *S. aureus* of *Z. zerumbet* oleoresin. The understanding of the processing effect is useful in designing a better processing technology to retain the antibacterial activity in *Z. zerumbet* extracts.

## REFERENCES

- Abdul Rahman, R. (2004). Response Surface Methods: Using Design Expert Software. *Two Day Course Note on Design Expert*. 6-7 February. Skudai: Chemical Engineering Pilot Plant, Universiti Teknologi Malaysia.
- Al-Dabbas, M. M., Hashinaga, F., Abdelgale, S.A.M., Suganuma, T., Akiyama, K., and Hayashi, H. (2005). Antibacterial Activity of an Eudesmane Sesquiterpene Isolated from Common Varthemia, *Varthemia iphionoides*. *Journal of Ethnopharmacology*. 97 (2), 237-240.
- Ang, H. G., Theng, C. Y., and Lim, K. K. (1982). The Role of Food Technology in Community Nutrition. In Foo, L. C., and Chong, Y. H. *ASEAN Subcommittee on Protein*. (pp.33-34). Kuala Lumpur: Ministry of Science, Technology and Environment.
- Azmi Muda, M., Ibrahim, H. and Khalid, N. (2001). *Micropropagation Study and Genetic Analysis with RAPD Markers on Selected Medicinal Gingers*. Kuala Lumpur: University of Malaya.
- Balladin, D. A., Headly, O., Chang-Yen, I., Duncan, E. J. and Mc Gaw, D. R. (1999). Comparison of the Histology of Fresh, Solar Dried and Steam Distilled Ginger Rhizome Tissue Prior to the Extraction of Its Pungent Principles. *Journal of Renewable Energy*. 17(2), 207-211.
- Berry, P. B. and Rodriguez, E. (2003). *Benchtop Bioassays: Methods in Chemical Prospecting*. New York: Cornell University.
- Bhuvaneswari, K., Poongothai, S. G., Kuruvilla, A. and Raju, B. A. (2002). Inhibitory Concentration of *Lawsonia inermis* Dry Powder for Urinary Pathogens. *Indian Journal of Pharmacology*. 34, 260-263.

- Bilal, N. E. and Gedebo, M. (2000). *Staphylococcus aureus* as a Paradigm of a Persistent Problem of Bacterial Multiple Antibiotic Resistance in Abha, Saudi Arabia. *Eastern Mediterranean Health Journal*. 6, 948-954.
- Boa, A. N. (2001). *The Bacterial Cell Wall*. Hull: Department of Chemistry, University of Hull.
- Burkill, I. H. (1935). *A Dictionary of the Economic Products of the Malay Peninsula*. London: Crown Agents for the colonies.
- Burt, S. (2004). Essential Oils: Their Antibacterial Properties and Potential Applications in Foods - a Review. *International Journal of Food Microbiology*. 94(3), 223-253
- Camp Micro, Inc. (2002). *MICRO METHODS #3: Preparation and Use of McFarland Standard Test Suspension*. United State of America: Trade Brochure.
- Chhabra, B. R., Dhillon, R. S., Wadia, M. S. and Kalsi, P. S. (1975). Structure of Zerumbone Oxide, a New Sesquiterpene Epoxy Ketone from *Zingiber zerumbet* (White Ginger Oil). *Indian Journal of Chemistry*. 13, 222-224.
- Charles, E. B. J., Taber, D. F. and Clark, A. K. (2001). *Organic Chemistry Laboratory with Qualitative Analysis Standard and Microscale Experiments*. (3<sup>rd</sup> ed.) United State of America: Harcourt College Publishers.
- Cornell, J. A. (1990). *How to Apply Response Surface Methodology (RSM)*. (2<sup>nd</sup> ed.) Wisconsin, United State of America: ASQC.
- Dai, J. R., Cardellina, J. H. II, McMahon, J. B. and Boyd, M. R. (1997). Zerumbone, an HIV-Inhibitory and Cytotoxic Sesquiterpene of *Zingiber aromaticum* and *Z. zerumbet*. *Natural Product Letters*. 10, 115-118.
- Damodaran, N. P. and Dev. S. 1963. Some New Humulene-based Sesquiterpenoids. *Tetrahedron Letters*. 28, 1941-1948.
- Devore, J.L and Farnum, N. R. (1999). *Applied Statistic for Engineers and Scientist*. Pasific Grove: Duxbury Press.
- Fennema, O., (1985). Chemical Changes in Food During Processing – An Overview. In Richardson, T and Finley, J. W. *Chemical Changes in Food During Processing* (pp. 1-16). Connecticut: Avi Publishing Company, Inc.

- Gaikar, V. G. and Dandekar, D. V., (2001). *U. S. Patent No. 6,224,877*. Washington DC: U.S. Patent and Trademark Office.
- Geankoplis, C. J. (1993). *Transport Processes and Unit Operation*. (3<sup>rd</sup> ed.) New Jersey, United State of America: Prentice Hall.
- Govindarajan, V. S. (1980). Turmeric – Chemistry, Technology and Quality. *Critical Reviews in Food Science*. 12(3). 199-301
- Guerrero, L. Ma. (1921). Medicinal uses of Philippine Plants. *Philippine Bureau of Forestry Bulletin*. 22. 149-246.
- Haaland, P. D. (1989). *Experimental Design in Biotechnology*. New York: Marcel Dekker.
- Harborne, J. B. (1973). *Phytochemical Methods – A Guide to Modern Technique of Plant Analysis*. London: Chapman and Hall. .
- Harborne, J.B. (1998). *Phytochemical Methods – A Guide to Modern Technique of Plant Analysis*. (2<sup>nd</sup> Ed). In Che Pa, N. F. *Extraction of Zingiber zerumbet : A Study on Yield of Oleoresin, Zerumbone and Curcumin Content*. B.Sc. Thesis. Universiti Teknologi Malaysia, Skudai.
- Harvey, P. H. and Mark, D. P. (1991). *The Comparative Method in Evolutionary Biology*. Oxford: Oxford University Press.
- Hashim, Y. Z. H. (2002). *Effect of Processing Parameters on the Turmeric Oleoresin Antioxidant Activity*. M.Sc. Thesis. Universiti Teknologi Malaysia, Skudai.
- Houghton, P. J. and Raman, A. (1998). *Laboratory Handbook for the Fractionation of Natural Extracts*. London: Chapman & Hall.
- Hughes, I. (2002). *Herbs in Africa: Part 3- Extraction in Herbal Material*. Africa: Africa's First On-Line Science Magazine.
- Indrani, D. and Rao, G. V. (2001). Optimization of the Quality of South Indian Parotta by Modelling the Ingredient Composition Using the Response Surface Methodology. *International Journal Food Science Technology*. 36(2), 189-167.
- Ibrahim, D. and Osman, H. (1995), Antimicrobial Activity of *Cassia alata* from Malaysia. *Journal of Ethnopharmacology* , 45(3), 151–156.

- Jitoe, A., Masuda, T., Tengah, I. G. P., Suprpta, D. N., Gaea, I. W. And Nakatani, N. (1992). Antioxidant Activity of Tropical Ginger Extracts and Analysis of the Contained Curcuminoids. *Journal of Agricultural and Food Chemistry*. 40(8), 1337–1340.
- Kai, L., Paul-Georges, R., Bernard, F., Liliane, B., Joseph, C., Fe'lix, T. (2007). Composition, Irregular Terpenoids, Chemical Variability and Antibacterial Activity of the Essential Oil from *Santolina corsica jordan et fourr.* *Phytochemistry*. 68(12), 1698-1705.
- Kaiser, G. E. (2002). *Microbiology Laboratory Manual*. Maryland: Copyright@ Gary E. Kaiser.
- Kerr, T. J. and McHale, B. B. (2001). *Applications in General Microbiology: A Laboratory Manual*. (6<sup>th</sup> ed.). Winston-Salem: Hunter Textbooks Inc.
- Kikuzaki, H. and Nakatani, N. (1993). Antioxidant Effects of Some Ginger Constituents. *Journal of Food Science*. 58(12), 1407–1410
- Lee, A.P. and Ke Ola 0 Hawaii (1997). *Hawaiian Medicine*. Hawaii: School of Medicine, University of Hawaii,
- Lewis, R. J. Sr. (1997). *Hawley's Condensed Chemical Dictionary*. (13<sup>th</sup> ed.). New York: John Wiley and Sons
- Masuda, T., Jitoe, A., Kato, S. and Nakatani, N. (1991). Acetylated flavonol glycosides from *Zingiber zerumbet*. *Phytochemistry*. 30(7), 2391-2392.
- Matthes , H.W.D., Luu, B. and Ourisson, G. (1980). Cytotoxic Components of *Zingiber zerumbet*, *Curcuma zedoaria*, and *Curcuma domestica*. *Phytochemistry*, 19(12), 2643-2650.
- Mohd Salleh, L. (2001). *Production of Soy Sauce: An Aroma Retention Study*. M.Sc. Thesis. Universiti Teknologi Malaysia, Skudai.
- Mohd Sirat, H., Lim, G.N., Saat, R., N. Leh, N.H. and Lee, L.M. (2000). Komposisi Kimia bagi Minyak Pati Lima Spesies Zingiber. In Zainal Abidin, M.Z. *Kajian Kesan Parameter-parameter Proses Pengekstrakan Zerumbone daripada Zingiber zerumbet*. B.Sc. Thesis. Universiti Teknologi Malaysia, Skudai.

- Muntada, V., Gerschenson, L. N., Alzamro, S. M. and Lustro, M. A. (1998), Solute Infusion Effect on Texture of Minimally Processed Kiwi Fruit. *Journal Food Science*. 63(4), 616-620.
- Murakami, A., Takahashi, D., Kinoshita, T., Koshimizu, K., Kim, H. W., Yoshihiro, A., Nakamura, Y., Jiwajinda, S., Terao, J. and Ohigashi, H. (2002). Zerumbone, A Southeast Asian Ginger Sesquiterpene, Markedly Suppresses Free Radical Generation, Proinflammatory Protein Production, and Cancer Cell Proliferation Accompanied by Apoptosis: The Alpha,Beta-Unsaturated Carbonyl Group is a Prerequisite. *Journal of National Center for Biotechnology Information*. 23(5), 795-802.
- Nakatani, N., Jitoe, A., Masuda, T. and Yonemori, S. (1991). Flavonoid Constituents of *Zingiber zerumbet* Smith. *Agricultural Biology Chemistry Journal*. 55(2), 455-460.
- National Committee for Clinical Laboratory Standard (1999). *M100-S9*. Wayne, Pennsylvania: National Committee for Clinical Laboratory Standard.
- Nicoli, M. C., Anese, M. and Parpinel, M. (1999). Influence of Processing on the Antioxidant Properties in Fruit and Vegetables. *Trends in Food Science and Technology*. 10(3), 94-100.
- Nielsen, S. (1994). *Introduction to the Chemical Analysis of Foods*. Boston: Jones and Bartlett.
- Nigam, I. C., and Levi, L. (1963). Column and Gas Chromatographic Analysis of Oil of Wild Ginger. *Canadian Journal of Chemistry*. 44(7), 1728-1735.
- Noor Azian, M., Mustafa Kamal, A. A., Nurul Azlina, M. (2004). Changes of Cell Structure in Ginger During Processing. *Journal of Food Engineering*. 62(4), 359-364.
- Oliveros, M. B., and Cantoria, M. C. (1993). *Pharmacognostical Studies On Zingiber zerumbet (L.) smith and Its Proposed Variety (Family Zingiberaceae)*. Quenzon City, Philippine: University of the Philippines
- Philips, R. J. (1996). *Blanching and Dehydration Technology*. USA: Food Manufacturing Coalition (FMC) Need Statement.



- Pokorny, J. and Schmidt, S. (2001). Natural Antioxidant Functionality During Food Processing. In Pokorny, J., Yanishlieva, N. and Gordon, M. (Ed.). *Antioxidants in Food: Practical Application*. (331-351). Cambridge: Woodhead Publishing Limited.
- Quisumbing, E. (1978). *Medicinal Plants of the Philippines*. Quezon City, Philippines: Katha Publishing Co. Inc.
- Ramaswami, S. K. and Bhattacharyya S. C. (1962). Terpenoids – XXXI. Isolation of Humelene Monoxide and Humelene Dioxide. *Tetrahedron*. 18, 575-579.
- Rao, H. N., Damodaran, N. P. and Dev, S. (1967). Photochemistry of Zerumbone. *Tetrahedron Letters*. 3, 227-233.
- Raskin, I., Ribnicky, D. M., Komarnytsky, S., Ilic, N., Poulev, A., Borisjuk, N., Brinker, A., Diego A. M., Ripoll, C, Yakoby, N., O'Neal, J. M., Cornwell, T., Pastor, I., and Fridlender B. (2002). Plants and Human Health in the Twenty-first Century. *Trends in Biotechnology*. 20(12), 522–531
- Rojas, I, Valenzuela. O, Anguita. M, Prieto. A (1998), Analysis of the Operators Involved in the Definition of the Implication Functions and in the Fuzzy Inference Process. *International Journal of Approximate Reasoning*. 19(3-4), 367-389.
- Ruslay, S., Abas, F., Shaari, K., Zainal, Z., Maulidiani, Sirat, H., Israf, D. A. and Lajis, N. H. (2007). Characterization of the Components Present in the Active Fractions of Health Gingers (*C. xanthorrhiza* and *Z. zerumbet*) by HPLC – DAD – ESIMS. *Food Chemistry*. 104(3), 1183-1191.
- Russell, F. B. (2001). *Microbiology Laboratory Manual*. United States of America: Wadsworth Group. Brooks / Cole.
- Sachais, B. (1997). *University of Pennsylvania Medical Center Guidelines for Antimicrobial Therapy: Antimicrobial Susceptibility Testing*. Philadelphia: Copyright@ University of Pennsylvania.
- Sanagi, M. M.. (1994). Analysis of *Zingiber zerumbet* Smith using Superficial Fluid Extraction. In: Che Pa, N. F. *Extraction of Zingiber zerumbet: A Study on Yield of Oleoresin, Zerumbone and Curcumin Content*. B.Sc. Thesis. Universiti Teknologi Malaysia, Skudai.

- Sanagi, M. M., Mohd Sirat, H. and Hasan, K. (1991). The Characterization of *Zingiber zerumbet*, Smith Using High Performance Liquid Chromatography with Diode Array Detection. *Proceedings of the Conference on Medicinal products from Tropical Rain Forest*. 13-15 May. FRIM, Kuala Lumpur, 353-358.
- Sharma, S. K., Mulvaney, S. J. and Rizvi, S. S. H. (2000). Introducing Natural Antioxidants. In Pokorny, J. Yanishlieva, N. and Gordon, M (Eds.). *Antioxidant in Food: Practical Application*. (pp. 147-158). Cambridge: Woodhead Publishing Limited.
- Singleton, P. (1999). *Bacteria in Biology, Biotechnology and Medicine*. (5th ed). West Sussex, England: John Wiley and Sons.
- Skinner, D. (2003). *Floridata*. Florida: Bulb Mall.
- Smith, M. F. and Paton, J. E., (2002). *Innovative Products from Australian Native Foods*. Australia: RIRDC Publication.
- Somchit M.N. and Nur Shukriyah M. H. (2003). Anti Inflammatory Property of Ethanol and Water Extracts of *Zingiber zerumbet*. *Indian Journal of Pharmacology*. 35, 181-182.
- Soto-Cordoba, S. M., Baeza, J. and Freer, J. (2001). Soxhlet Extraction of Pentachlorophenol from Soil with *In Situ* Derivatization. *Journal of the Chilean Chemical Society*. 46, 126-128.
- Stat-Ease, Inc. (2002). Design Expert. Version 6.0.8. Software.
- Takahashi (1988). *Jamu*. Tokyo: Hirakawa Shuppansha.
- Takashi, K., Kaneyoshi, Y., Ryutaro, U., Masahiro, T., Richard, K. H., Yasishi, K., Seiji, S. and Tadashi, O. (2001), Chemistry of Zerumbone. 2. Regularion of Ring Bond Cleavage and Unique Antibacterial Activities of Zerumbone Derivatives. *Journal of Bioscience, Biotechnology and Biochemistry*. 65(10), 2193-2199.
- Tiwari, K. K., (1995). Extraction Technologies Related to Food Processing. In Gaonkar, A.G. (Ed). *Food Processing: Recent Development*. (pp. 269-301). USA: Elsevier Science.
- Todar, K. (2003). *Bacteriology: Staphylococcus*. Madison: University of Wisconsin-Madison

- Umi K. Ahmad, Hasnah M. Sirat, M. Marsin Sanagi and Roger M. Smith. (1994). Supercritical Fluid Extraction and Capillary Gas Chromatography of the Rhizomes of *Z. zerumbet*. *Journal of Microcolumn Separations*. 6(1), 27–32
- Valgas, C., Sauza, M. D. S., Smania, E. F. A. and Smania, J. A. (2007). Screening Methods to Determine Antibacterial Activity of Natural Products. *Brazilian Journal of Microbiology*. 38(2), 369-380.
- Valls, J. S., Nacente, R. B. and Coll, M. S. (2001). *Handbook of Microbiological Culture Media*. Spain: Scharlau Chemie.
- Varier, N. S. (1944). Chemical Examination of Rhizomes of *Zingiber zerumbet* Smith. *Proc. Indian Academy of Science*. 20, 257-266.
- William, R. S. (2000). The Role of Natural Products in a Modern Drug Discovery Programme. *Drug Discovery Today*. 5(2), 39-41.
- Yaakob, H. (2006). *The Effect of Fermentation Process on the Diadzin And Diadzein Content in Fermented Soybean*. M.Sc. Thesis. Universiti Teknologi Malaysia, Skudai.
- Zivanovic, S., Chi, S. and Droughon, F.A. (2005). Antimicrobial Activity of Essential Oils Incorporated in Chitosan Films. *Journal of Food Science*. 70(1), 44-49.