

PHYTOCHEMICAL AND BIOACTIVITIES OF MALAYSIAN *ARTOCARPUS LOWII*
KING, *A. SCORTECHINII* KING AND *A. TEYSMANII* MIQ.

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Dedicated to...

My parents

My beloved husband, Mr Kamaruddin bin Mohammed

My sons, Uzair bin Kamaruddin, Zaid bin Kamaruddin and

Adam bin Kamaruddin

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ABSTRACT

Phytochemical studies of *Artocarpus lowii* King, *A. scortechinii* King and *A. teysmanii* Miq. have resulted in the isolation of four new compounds and eight known compounds. Three new compounds have been successfully isolated from *A. lowii* King, i.e. 2',4'-dihydroxy-4-methoxy-3'-prenyldihydrochalcone, 2',3,4',4-tetrahydroxy-3'-prenylchalcone and 2-hydroxyparatocarpin C. Three known compounds were identified as cycloheterophyllin, 2',4',4-trihydroxy-3'-prenylchalcone and 4-hydroxylonchocarpin. Methylation of 2',4'-dihydroxy-4-methoxy-3'-prenyldihydrochalcone and 2',4',4-trihydroxy-3'-prenylchalcone gave 2',4',4-trimethoxy-3'-prenyldihydrochalcone and 2'-hydroxy-4',4-dimethoxy-3'-prenylchalcone, respectively while methylation of cycloheterophyllin gave mixtures of dimethoxy and trimethoxy derivatives. Acetylation of cycloheterophyllin afforded cycloheterophyllin diacetate. A new compound was isolated from *A. scortechinii* King and was identified as 2',4',5',5-tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6H-pyrano)-6-prenylflavone together with three known compounds, i.e. artonin E, artobiloxanthone and lupeol 3-acetate. Methylation of artonin E gave artonin E trimethyl ether while acetylation of artonin E afforded artonin E tetraacetate. Four known compounds were isolated from *A. teysmanii* Miq., which were identified as artonin E, artobiloxanthone, artonol B and cycloartobiloxanthone. The structures of all compounds were established based on spectral studies using nuclear magnetic resonance spectroscopy, mass spectrometry, infrared spectroscopy and ultraviolet spectroscopy. The biological studies on the crude extracts and pure compounds of these three species showed that several pure compounds have significant biological activity especially in the antioxidant, platelet aggregation and cytotoxicity assays. Cycloheterophyllin and artonin E showed high ability to act as free radical scavengers with scavenging concentration values of 51.6 µg/mL and 48.3 µg/mL, respectively. Cycloheterophyllin, artonin E, isobavachalcone and 2',4'-dihydroxy-4-methoxy-3'-prenyldihydrochalcone totally inhibited adenosine diphosphate-induced platelet aggregation compared to standard aspirin which suppressed only 31.6% of the platelet aggregation. Cycloheterophyllin and artonin E were found to be active against breast cancer cell line, MCF7 comparable to the standard tamoxifen citrate. Finally, 2'-hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone was synthesized through a three step synthesis. The steps involved Friedel-Crafts prenylation followed by methylation and Claisen-Schmidt condensation. Oxidative cyclization of this chalcone yielded an aldehyde-type chalcone derivative. 2'-Hydroxy-4-methoxy-4'-O-prenylchalcone was synthesized through a two steps synthesis involving Friedel-Crafts prenylation followed by Claisen-Schmidt condensation. Attempted Claisen rearrangement produced 4'-hydroxy-4-methoxy-2'-O-prenylchalcone as the major product.

ABSTRAK

Kajian fitokimia ke atas *Artocarpus lowii* King, *A. scortechinii* King dan *A. teysmanii* Miq. berjaya menemukan empat sebatian baru dan lapan sebatian yang diketahui. Tiga sebatian baru ditemui daripada *A. lowii* King dikenalpasti sebagai 2',4'-dihidroksi-4-metoksi-3'-prenildihidrokkalkon, 2',3,4',4-tetrahidroksi-3'-prenil-kalkon dan 2-hidroksiparatokarpin C. Tiga sebatian lain dikenalpasti sebagai sikloheterofilin, 2',4',4-trihidroksi-3'-prenilkalkon dan 4-hidroksilonchokarpin. Pemetilan 2',4'-dihidroksi-4-metoksi-3'-prenildihidrokkalkon dan 2',4',4-trihidroksi-3'-prenilkalkon masing-masing berjaya menghasilkan 2',4',4-trimetoksi-3'-prenildihidrokkalkon dan 2'-hidroksi-4',4-dimetoksi-3'-prenilkalkon, sementara pemetilan sikloheterofilin menghasilkan campuran terbitan dimetoksi dan trimetoksi. Pengasetilan terhadap sikloheterofilin berjaya menghasilkan sikloheterofilin diasetat. Satu sebatian baru telah berjaya diasingkan daripada *A. scortechinii* King dan dikenalpasti sebagai 2',4',5',5-tetrahidroksi-3-geranil-7,8-(2,2-dimetil-6*H*-pirano)-6-prenilflavon bersama-sama tiga sebatian diketahui iaitu artonin E, artobiloxanton dan lupeol 3-asetat. Pemetilan artonin E berjaya menghasilkan artonin E trimetil eter, sementara pengasetilan artonin E berjaya menghasilkan artonin E tetraasetat. Empat sebatian berjaya diasingkan daripada *A. teysmanii* Miq. dan dikenalpasti sebagai artonin E, artobiloxanton, artonol B dan sikloartobiloxanton. Struktur kesemua sebatian dikenalpasti berdasarkan kepada kajian spektrum dengan menggunakan spektroskopi resonan magnet nukleus, spektrometri jisim, spektroskopi inframerah dan spektroskopi ultralembayung. Kajian aktiviti biologi ke atas ekstrak mentah dan sebatian tulen daripada ketiga-tiga spesies *Artocarpus* ini mendapati beberapa sebatian menunjukkan aktiviti biologi yang signifikan terutamanya di dalam cerakin antioksidan, agregasi platelet dan sitoketoksikan. Sikloheterofilin dan artonin E menunjukkan keupayaan yang tinggi sebagai perencat radikal bebas dengan nilai perencatan masing-masing 51.6 µg/mL dan 48.3 µg/mL. Sikloheterofilin, artonin E, isobavakalkon dan 2',4'-dihidroksi-4-metoksi-3'-prenildihidrokkalkon merencat 100% agregasi platelet yang dirangsang oleh adenosin difosfat, berbanding dengan aspirin yang hanya mampu merencat sebanyak 31.6% sahaja. Sikloheterofilin dan artonin E juga didapati aktif terhadap MCF7 iaitu sel kanser payu dara, setanding dengan tamoksifen sitrat iaitu dadah anti kanser piawai. 2'-Hidroksi-4,4',6'-trimetoksi-3'-prenilkalkon telah berjaya disintesis melalui tiga langkah tindak balas. Tindak balas yang terlibat ialah pempenilan Friedel-Crafts, tindak balas pemetilan dan kondensasi Claisen-Schmidt. Pensiklikan oksidatif terhadap kalkon ini menghasilkan satu terbitan kalkon jenis aldehid. 2'-Hidroksi-4-metoksi-4'-*O*-prenilkalkon pula telah berjaya disintesis melalui dua langkah tindak balas iaitu tindak balas pempenilan Friedel-Crafts, diikuti dengan kondensasi Claisen-Schmidt. Percubaan melakukan penyusunan semula Claisen menghasilkan 4'-hidroksi-4-metoksi-2'-*O*-prenilkalkon sebagai hasil utama.

TABLE OF CONTENTS

CHAPTER	TITLE	PAGE
	DECLARATION OF THE STATUS OF THESIS	
	SUPERVISOR'S DECLARATION	
	CERTIFICATION OF EXAMINATION	
	TITLE PAGE	i
	DECLARATION OF ORIGINALITY AND EXCLUSIVENESS	ii
	DEDICATION	iii
	ACKNOWLEDGEMENTS	iv
	ABSTRACT	v
	ABSTRAK	vi
	TABLE OF CONTENTS	vii
	LIST OF TABLES	xii
	LIST OF SCHEMES	xiii
	LIST OF FIGURES	xv
	LIST OF ABBREVIATIONS	xix
	LIST OF APPENDICES	xxii
1	INTRODUCTION	
	1.1 General Introduction	1
	1.2 Family Moraceae	2
	1.3 Genus <i>Artocarpus</i>	3
	1.4 A Review of Phytochemicals and Biological Properties of <i>Artocarpus</i> Species	4

1.4.1	Isoprenylflavonoids	5
1.4.2	Stilbenoid and 2-Arylbenzofuran Derivatives	19
1.4.3	Phenolic Compounds with Oxepine Ring	22
1.4.4	Diels-Alder Type Adducts	23
1.5	Biosynthesis of Flavonoids	24
1.6	Synthesis of Flavonoids	27
1.7	Research Objectives	32
2	PHYTOCHEMICALS OF <i>ARTOCARPUS LOWII</i> KING, <i>A. SCORTECHINII</i> KING AND <i>A. TEYSMANII</i> MIQ. FROM MALAYSIA	
2.1	Phytochemicals of <i>Artocarpus lowii</i> King	33
2.1.1	2',4',4'-Trihydroxy-3'-prenyl-chalcone (Isobavachalcone) (115)	34
2.1.2	2',3,4',4'-Tetrahydroxy-3'-prenyl-chalcone (117)	37
2.1.3	2',4'-Dihydroxy-3',4'-(2,2-dimethylchromeno)chalcone (4-hydroxylonchocarpin) (118)	48
2.1.4	2-Hydroxyparatocarpin C (119)	49
2.1.5	2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (120)	62
2.1.6	Cycloheterophyllin (3)	81
2.2	Phytochemicals of <i>Artocarpus scortechinii</i> King	84
2.2.1	Lupeol 3-acetate (125)	85
2.2.2	Artonin E (14)	86
2.2.3	Artobiloxanthone (15)	89
2.2.4	2',4',5',5'-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128)	91
2.3	Phytochemicals of <i>Artocarpus teysmanii</i> Miq.	102
2.3.1	Artonol B (19)	102
2.3.2	Cycloartobiloxanthone (17)	103

3	BIOLOGICAL ACTIVITIES OF <i>ARTOCARPUS LOWII</i> KING, <i>A. SCORTECHINII</i> KING, AND <i>A. TEYSMANII</i> MIQ.	
3.1	Biological Activities of <i>Artocarpus</i> Species	106
3.2	Antibacterial Activity of <i>Artocarpus lowii</i> King, <i>A. scortechinii</i> King and <i>A. teysmanii</i> Miq.	108
3.3	Antioxidant Activity of <i>Artocarpus lowii</i> King, <i>A. scortechinii</i> King and <i>A. teysmanii</i> Miq.	111
3.3.1	Inhibition of Lipid Peroxidation	113
3.3.2	Scavenging Activity on 2,2-diphenyl-1-picrylhydrazine (DPPH) Radical	117
3.4	Platelet Activating Factor (PAF) Receptor Binding Activity	122
3.5	Platelet Aggregation Activity	123
3.6	Cytotoxic Activity	125
4	SYNTHESIS OF FLAVONOIDS	
4.1	Introduction	126
4.2	Attempted Synthesis of 2'-Hydroxy-4',4-dimethoxy-3'-prenylchalcone (116)	130
4.3	Attempted Claisen Rearrangement of 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151) to Form 2',4'-Dihydroxy-4-methoxy-3'-(1'',1''-dimethylallyl)chalcone (152)	139
4.4	Attempted Synthesis of 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154)	146
4.5	Attempted Synthesis of 2',4',4-Trimethoxy-5'-prenylflavone (162)	150
5	EXPERIMENTAL	
5.1	General	158
5.2	Chromatographic Methods	159
5.3	Plant Materials	159
5.4	Extraction and Isolation of Leaves and Bark of <i>Artocarpus lowii</i> King	159

5.4.1	2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (120)	160
5.4.2	2-Hydroxyparatocarpin C (119)	161
5.4.3	2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115)	161
5.4.4	4-HydroxyLonchocarpin (118)	163
5.4.5	2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	164
5.4.6	Cycloheterophyllin (3)	165
5.5	Extraction and Isolation of The Bark of <i>Artocarpus scortechinii</i> King	167
5.5.1	Lupeol 3-acetate (125)	168
5.5.2	Artonin E (14)	168
5.5.3	Artobiloxanthone (15)	170
5.5.4	2',4',5',5-Tetrahydroxy-3-geranyl 7,8-(2,2-dimethyl-6H-pyrano)-6-prenylflavone (128)	171
5.6	Extraction and Isolation of The Bark of <i>Artocarpus teysmanii</i> Miq.	172
5.6.1	Artonol B (19)	172
5.6.2	Cycloartobiloxanthone (17)	173
5.7	Antibacterial Assay	173
5.7.1	Microorganisms	174
5.7.2	Disc Diffusion Method	174
5.7.3	Minimum Inhibitory Concentration (MIC)	174
5.7.4	Minimum Bactericidal Concentration (MBC)	175
5.8	Antioxidant Assay	176
5.8.1	Ferric Thiocyanate (FTC) Method	176
5.8.2	Free Radical Scavenging Activity (DPPH) Assays	176
	5.8.2.1 UV Spectrophotometric Assay	176
	5.8.2.2 DPPH Electron Spin Resonance (ESR) Assay	177
5.9	Platelet-Activating Factor (PAF) Receptor Binding Assay	177

5.10	Platelet Aggregation Assay	179
5.11	Cytotoxic Assay	179
5.12	Attempted Synthesis of 2'-Hydroxy-4',4'-dimethoxy-3'-prenylchalcone (116)	180
5.12.1	Formation of 2'-Hydroxy-4'- <i>O</i> -prenylacetophenone (150)	180
5.12.2	Synthesis of 2'-hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151)	181
5.12.3	Attempted Synthesis of 2',4'-dihydroxy-4-methoxy-3'-(1'',1''-dimethylallyl)chalcone (152)	181
5.13	Attempted Synthesis of 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154)	182
5.13.1	Formation of 2',4',6'-Trihydroxy-3'-prenylacetophenone (157)	182
5.13.2	Formation of 2'-hydroxy-4',6'-dimethoxy-3'-prenylacetophenone (155)	183
5.13.3	Formation of 2'-hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154)	184
5.14	Attempted Synthesis of 2',4',4'-trimethoxy-5'-prenylflavone (162)	184
6	CONCLUSION	186
	REFERENCES	189
	APPENDICES	202

LIST OF TABLES

TABLE	TITLE	PAGE
2.1	Yield of Crude Extracts of <i>A. lowii</i> King	34
2.2	¹ H NMR and ¹³ C NMR Data of Compound (118) and 4-Hydroxylonchocarpin	50
2.3	¹ H NMR and ¹³ C NMR Data of 2-Hydroxyparatocarpin C (119) and Paratocarpin C (67)	59
2.4	¹ H NMR and ¹³ C NMR Data of 2',4',5',5'-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128) and Artocommunol CB (33)	100
3.1	Antibacterial Activity of The Extracts of <i>A. lowii</i> , <i>A. scortechinii</i> and <i>A. teysmanii</i>	109
3.2	Antibacterial Activity of Several Pure Compounds	110
3.3	Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) of Several Pure Compounds	111
3.4	Scavenging Capacity of DPPH by Several Pure Compounds Isolated From <i>A. lowii</i> , <i>A. scortechinii</i> and <i>A. teysmanii</i> Measured by UV Spectrophotometric Assay	118
3.5	Scavenging of DPPH Free Radical by Pure Compounds Isolated From <i>A. lowii</i> , <i>A. scortechinii</i> and <i>A. teysmanii</i> Measured by ESR Spectrometric Assay	120
3.6	Inhibitory Effects of Compounds of <i>Artocarpus</i> Species on PAF Receptor Binding to Rabbit Platelets ^a	123
3.7	Percentage Inhibition of Compounds (100 µg/mL) From <i>Artocarpus lowii</i> , <i>A. scortechinii</i> and <i>A. teysmanii</i> , Two Derivatives and Aspirin (25 µg/mL) on Platelet Aggregation of Human Whole Blood	124
3.8	IC ₅₀ Values of Flavonoids From <i>Artocarpus lowii</i> , <i>A. scortechinii</i> and <i>A. teysmanii</i> Against Breast Carcinoma Cell Line, MCF7	125

LIST OF SCHEMES

SCHEME	TITLE	PAGE
2.1	EIMS Fragmentation Pattern of 2',4',4-Trihydroxy-3'-prenylchalcone (115)	35
2.2	EIMS Fragmentation Pattern of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	39
2.3	EIMS Fragmentation Pattern of 2-Hydroxyparatocarpin C (119)	61
2.4	EIMS Fragmentation Pattern of 2',4',4-Trimethoxy-3'-prenyldihydrochalcone (121)	81
2.5	EIMS Fragmentation Pattern of Artonin E (14)	87
2.6	EIMS Fragmentation Pattern of Artobiloxanthone (15)	90
2.7	EIMS Fragmentation Pattern of 2',4',5',5-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128)	101
4.1	Synthetic Route to 2,4,5,7-Tetrahydroxy-8-geranylflavanone or Sophoraflavanone C (143)	127
4.2	Retrosynthesis of 2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (120) and Isobavachalcone (115)	129
4.3	Synthetic Route to 2'-Hydroxy-4',4-dimethoxy-3'-prenylchalcone (116)	130
4.4	Mechanism for the Formation of 2'-Hydroxy-4'- <i>O</i> -prenylacetophenone (150) from 2',4'-Dihydroxyacetophenone (144)	131
4.5	Synthetic Route to 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151) and 2',4'-Dihydroxy-4-methoxy-3'-(1'',1''-dimethylallyl)chalcone (152) from 2'-Hydroxy-4'- <i>O</i> -prenylacetophenone (150)	132
4.6	Proposed Mechanism of Claisen Rearrangement of 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151) to form 4'-Hydroxy-4-methoxy-2'- <i>O</i> -prenylchalcone (153)	140

4.7	Synthetic Route to 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenyl-chalcone (154)	147
4.8	Attempted Synthesis of 2',4',4'-Trimethoxy-5'-prenylflavone (162) from 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154)	150
4.9	Possible Mechanism of SeO ₂ Promoted Oxidative Cyclization of (154) to form an Aldehyde-type Chalcone (163)	157

LIST OF FIGURES

FIGURE	TITLE	PAGE
1.1	Biosynthetic Pathway of Flavonoids [48-49]	26
1.2	The Claisen-Schmidt Reaction	28
1.3	Conversion of Chalcone to Various Types of Flavonoids	28
1.4	Baker-Venkataraman Rearrangement	29
1.5	Modified Baker-Venkataraman Rearrangement	30
1.6	Allan-Robinson Synthesis	30
1.7	Algar-Flynn-Oyamada (AFO) Reaction	31
2.1	IR Spectrum of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	40
2.2	UV Spectrum of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	41
2.3	EIMS Spectrum of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	42
2.4	The 300 MHz ¹ H NMR Spectrum of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	44
2.5	The 300 MHz ¹ H NMR Spectrum of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117) (expansion)	45
2.6	COSY Spectrum of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	46
2.7	¹³ C NMR and DEPT Spectra of 2',3,4',4-Tetrahydroxy-3'-prenylchalcone (117)	47
2.8	IR Spectrum of 2-Hydroxyparatocarpin C (119)	51
2.9	UV Spectrum of 2-Hydroxyparatocarpin C (119)	52
2.10	The 300 MHz ¹ H NMR Spectrum of 2-Hydroxyparatocarpin C (119)	54
2.11	The 300 MHz ¹ H NMR Spectrum of 2-Hydroxyparatocarpin C (119) (expansion)	55
2.12	COSY Spectrum of 2-Hydroxyparatocarpin C (119)	56
2.13	¹³ C NMR and DEPT Spectra of 2-Hydroxyparatocarpin C (119)	57

2.14	EIMS Spectrum of 2-Hydroxyparatocarpin C (119)	60
2.15	FABMS Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (120)	65
2.16	IR Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (120)	66
2.17	UV Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (120)	67
2.18	The 500 MHz ¹ H NMR Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120)	68
2.19	The 500 MHz ¹ H NMR Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120) (expansion 1)	69
2.20	COSY Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120)	70
2.21	¹³ C NMR Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120)	71
2.22	DEPT Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120)	72
2.23	HMQC Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120)	73
2.24	HMBC Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120)	74
2.25	HMBC Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyldihydrochalcone (120) (expansion 1)	75
2.26	HMBC Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120) (expansion 2)	76
2.27	HMBC Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120) (expansion 3)	77
2.28	HMBC Spectrum of 2',4'-Dihydroxy-4-methoxy-3'-prenyl-dihydrochalcone (120) (expansion 4)	78
2.29	The 300 MHz ¹ H NMR Spectrum of 2',4',4'-Trimethoxy-3'-prenyldihydrochalcone (121)	79
2.30	EIMS Spectrum of 2',4',4'-Trimethoxy-3'-prenyldihydrochalcone (121)	80
2.31	EIMS Spectrum of 2',4',5',5'-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128)	92
2.32	¹³ C NMR Spectrum of 2',4',5',5'-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128) (expansion 1)	93
2.33	¹³ C NMR Spectrum of 2',4',5',5'-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128) (expansion 2)	94

2.34	IR Spectrum of 2',4',5',5-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128)	95
2.35	UV Spectrum of 2',4',5',5-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128)	96
2.36	The 500 MHz ¹ H NMR Spectrum of 2',4',5',5-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128)	98
2.37	DEPT Spectrum of 2',4',5',5-Tetrahydroxy-3-geranyl-7,8-(2,2-dimethyl-6 <i>H</i> -pyrano)-6-prenylflavone (128)	99
3.1	Graph of Absorbance versus Time Illustrating Antioxidant Activity of <i>A. lowii</i> Extracts Measured by FTC Method	113
3.2	Graph of Absorbance versus Time Illustrating Antioxidant Activity <i>A.scortechinii</i> Extracts Measured by FTC Method	114
3.3	Graph of Absorbance versus Time Illustrating Antioxidant Activity of <i>A. teysmanii</i> Extracts Measured by FTC Method	114
3.4	Graph of Absorbance versus Time Illustrating Antioxidant Activity of Pure Compounds of <i>A. lowii</i> Measured by FTC Method	115
3.5	Graph of Absorbance versus Time Illustrating Antioxidant Activity of Pure Compounds of <i>A. scortechinii</i> and <i>A. teysmanii</i> Measured by FTC Method	116
3.6	Reduction of DPPH (132) by Antioxidant Compound	117
3.7	ESR Spectrum of 25 mM ethanolic DPPH solution	119
3.8	ESR Spectra of Scavenging Effects of (a) Vitamin E and (b) Vitamin C on 25 mM of DPPH Radical at Various Concentrations	120
3.9	ESR Spectra of Scavenging Effects of (a) Cycloheterphyllin and (b) Artonin E on 25 mM of DPPH Radical at Various Concentrations	121
3.10	ESR Spectra of Scavenging Effects of (a) 4-Hydroxyl-onchocarpin and (b) Cycloartobiloxanthone on 25 mM of DPPH Radical at Various Concentrations	121
4.1	EIMS Spectrum of 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151)	134
4.2	IR Spectrum of 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151)	135
4.3	¹³ C NMR and DEPT Spectra of 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151)	136
4.4	The 300 MHz ¹ H NMR Spectrum of 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151)	137
4.5	The 300 MHz ¹ H NMR Spectrum of 2'-Hydroxy-4-methoxy-4'- <i>O</i> -prenylchalcone (151) (expansion)	138

4.6	The 300 MHz ^1H NMR Spectrum of 4'-Hydroxy-4-methoxy-2'- <i>O</i> -prenylchalcone (153)	141
4.7	The 300 MHz ^1H NMR Spectrum of 4'-Hydroxy-4-methoxy-2'- <i>O</i> -prenylchalcone (153) (expansion)	142
4.8	EIMS Spectrum of 4'-Hydroxy-4-methoxy-2'- <i>O</i> -prenylchalcone (153)	143
4.9	^{13}C NMR and DEPT Spectra of 4'-Hydroxy-4-methoxy-2'- <i>O</i> -prenylchalcone (153)	144
4.10	IR Spectrum of 4'-Hydroxy-4-methoxy-2'- <i>O</i> -prenylchalcone (153)	145
4.11	The 300 MHz ^1H NMR Spectrum of Chalcone (163)	153
4.12	The 300 MHz ^1H NMR Spectrum of Chalcone (163) (expansion)	154
4.13	^{13}C NMR and DEPT Spectra of Chalcone (163)	155
4.14	EIMS Spectrum of Chalcone (163)	156

LIST OF ABBREVIATIONS

AA	arachidonic acid
Ac ₂ O	acetic anhydride
ADP	Adenosine diphosphate
AlCl ₃	Aluminium trichloride
b.p	boiling point
br	Broad
¹³ C	Carbon-13
cm ⁻¹	Per centimetre
cm	Centimetre
CDCl ₃	deuterated chloroform
CD ₃ OD	deuterated methanol
CHCl ₃	Chloroform
CH ₂ Cl ₂	Dichloromethane
COSY	Correlation Spectroscopy
CO ₂	Carbon dioxide
d	Doublet
dd	doublet of doublets
DEPT	Distortionless Enhancement of Polarisation Transfer
DMSO	dimethyl sulphoxide
DMAP	4-(<i>N,N</i> -dimethylamino)pyridine
DMEM	Dulbecco's Modified Eagle Medium
<i>E</i>	<i>Entgegen</i>
Et ₂ O	Diethyl ether
EtOH	Ethanol
EIMS	Electron Impact Mass Spectrometry

ESR	Electron Spin Resonance
GHz	Gigahertz
¹ H	Proton
³ H-PAF	Radiolabelled platelet activating factor
HEPES	N-[2-hydroxyethyl]piperazine-N'-[2-ethanesulphonic acid]
HMBC	Heteronuclear Multiple Bond Correlation
HMQC	Heteronuclear Multiple Quantum Coherence
HRMS	High Resolution Mass Spectroscopy
HCl	Hydrochloric Acid
Hz	Hertz
IC	inhibition concentration
IR	Infrared
<i>J</i>	coupling constant
K ₂ HPO ₄	Potassium hydrogen phosphate
lit.	Literature
m	Multiplet
M	Molar
mg	Milligram
mM	Millimolar
MBC	Minimum bactericidal concentration
MeOH	Methanol
MHz	Megahertz
MIC	minimum inhibition concentration
m.p	melting point
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
<i>m/z</i>	mass-to-charge ratio
nM	Nanomolar
nm	Nanometer
NMR	Nuclear Magnetic Resonance
pet. ether	Petroleum ether
ppm	Part per million
R _f	retention factor

rt	room temperature
s	Singlet
t	Triplet
TLC	Thin-layer chromatography
VLC	vacuum liquid chromatography
δ	chemical shift
UV	Ultraviolet
μM	Micromolar
γ	Gamma
λ	Lamda

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
1	IR Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115)	202
2	UV Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115)	203
3	EIMS Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115)	204
4	The 500 MHz ¹ H NMR Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115)	205
5	The 500 MHz ¹ H NMR Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115) (expansion)	206
6	COSY Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115) (expansion 1)	207
7	COSY Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115) (expansion 2)	208
8	HMBC Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115) (expansion 1)	209
9	HMBC Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115) (expansion 2)	210
10	¹³ C NMR Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115)	211
11	DEPT Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115)	212
12	HMQC Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115) (expansion 1)	213
13	HMQC Spectrum of 2',4',4-Trihydroxy-3'-prenylchalcone (Isobavachalcone) (115) (expansion 2)	214
14	The 300 MHz ¹ H NMR Spectrum of 2'-Hydroxy-4',4-dimethoxy-3'-prenylchalcone (116)	215

15	EIMS Spectrum of 2'-Hydroxy-4',4-dimethoxy-3'-prenyl-chalcone (116)	216
16	IR Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	217
17	UV Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	218
18	FABMS Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	219
19	The 500 MHz ¹ H NMR Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	220
20	The 500 MHz ¹ H NMR Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118) (expansion)	221
21	COSY Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	222
22	¹³ C NMR Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	223
23	¹³ C NMR Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118) (expansion)	224
24	HMQC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	225
25	HMQC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118) (expansion 1)	226
26	HMQC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118) (expansion 2)	227
27	HMBC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	228
28	HMBC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118) (expansion 1)	229
29	HMBC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118) (expansion 2)	230
30	HMBC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118) (expansion 3)	231
31	HMBC Spectrum of 2',4-Dihydroxy-3',4'-(2,2-dimethyl-chromeno)chalcone (4-HydroxyLonchocarpin) (118)	232

	(expansion 4)	
32	EIMS Spectrum of Cycloheterophyllin (3)	233
33	IR Spectrum of Cycloheterophyllin (3)	234
34	UV Spectrum of Cycloheterophyllin (3)	235
35	The 500 MHz ^1H NMR Spectrum of Cycloheterophyllin (3)	236
36	The 500 MHz ^1H NMR Spectrum of Cycloheterophyllin (3) (expansion)	237
37	COSY Spectrum of Cycloheterophyllin (3)	238
38	^{13}C NMR Spectrum of Cycloheterophyllin (3)	239
39	^{13}C NMR Spectrum of Cycloheterophyllin (3) (expansion)	240
40	DEPT Spectrum of Cycloheterophyllin (3)	241
41	HMQC Spectrum of Cycloheterophyllin (3)	242
42	HMBC Spectrum of Cycloheterophyllin (3)	243
43	The 300 MHz ^1H NMR Spectrum of Cycloheterophyllin dimethyl ether (122)	244
44	EIMS spectrum of Cycloheterophyllin dimethyl ether (122)	245
45	The 300 MHz ^1H NMR Spectrum of Cycloheterophyllin trimethyl ether (123)	246
46	EIMS spectrum of Cycloheterophyllin trimethyl ether (123)	247
47	The 300 MHz ^1H NMR Spectrum of Cycloheterophyllin diacetate (124)	248
48	The 300 MHz ^1H NMR Spectrum of Cycloheterophyllin diacetate (124) (expansion)	249
49	IR Spectrum of Cycloheterophyllin diacetate (124)	250
50	EIMS Spectrum of Cycloheterophyllin diacetate (124)	251
51	IR Spectrum of Lupeol 3-acetate (125)	252
52	EIMS Spectrum of Lupeol 3-acetate (125)	253
53	The 300 MHz ^1H NMR Spectrum of Lupeol 3-acetate (125)	254
54	^{13}C NMR and DEPT Spectra of Lupeol 3-acetate (125)	255
55	EIMS Spectrum of Artonin E (14)	256
56	UV Spectrum of Artonin E (14)	257
57	The 500 MHz ^1H NMR Spectrum of Artonin E (14)	258
58	The 500 MHz ^1H NMR Spectrum of Artonin E (14) (expansion)	259

59	¹³ C NMR Spectrum of Artonin E (14)	260
60	¹³ C NMR Spectrum of Artonin E (14) (expansion)	261
61	IR Spectrum of Artonin E (14)	262
62	HMBC Spectrum of Artonin E (14)	263
63	HMBC Spectrum of Artonin E (14) (expansion 1)	264
64	HMBC Spectrum of Artonin E (14) (expansion 2)	265
65	HMBC Spectrum of Artonin E (14) (expansion 3)	266
66	HMBC Spectrum of Artonin E (14) (expansion 4)	267
67	DEPT Spectrum of Artonin E (14)	268
68	COSY Spectrum of Artonin E (14)	269
69	HMQC Spectrum of Artonin E (14)	270
70	The 300 MHz ¹ H NMR Spectrum of Artonin E trimethyl ether (126)	271
71	EIMS Spectrum of Artonin E trimethyl ether (126)	272
72	The 300 MHz ¹ H NMR Spectrum of Artonin E tetraacetate (127)	273
73	EIMS Spectrum of Artonin E tetraacetate (127)	274
74	IR Spectrum of Artonin E tetraacetate (127)	275
75	FABMS Spectrum of Artobiloxanthone (15)	276
76	EIMS Spectrum of Artobiloxanthone (15)	277
77	The 600 MHz ¹ H NMR Spectrum of Artobiloxanthone (15)	278
78	The 600 MHz ¹ H NMR Spectrum of Artobiloxanthone (15) (expansion 1)	279
79	The 600 MHz ¹ H NMR Spectrum of Artobiloxanthone (15) (expansion 2)	280
80	DEPT Spectrum of Artobiloxanthone (15)	281
81	COSY Spectrum of Artobiloxanthone (15)	282
82	HMBC Spectrum of Artobiloxanthone (15) (expansion 1)	283
83	HMBC Spectrum of Artobiloxanthone (15) (expansion 2)	284
84	IR Spectrum of Artobiloxanthone (15)	285
85	UV Spectrum of Artobiloxanthone (15)	286
86	¹³ C NMR Spectrum of Artobiloxanthone (15)	287
87	HMQC Spectrum of Artobiloxanthone (15)	288
88	EIMS Spectrum of Artonol B (19)	289
89	IR Spectrum of Artonol B (19)	290
90	UV Spectrum of Artonol B (19)	291

91	The 500 MHz ¹ H NMR Spectrum of Artonol B (19)	292
92	¹³ C NMR Spectrum of Artonol B (19)	293
93	¹³ C NMR Spectrum of Artonol B (19) (expansion)	294
94	DEPT Spectrum of Artonol B (19)	295
95	HMQC Spectrum of Artonol B (19)	296
96	HMBC Spectrum of Artonol B (19)	297
97	EIMS Spectrum of Cycloartobiloxanthone (17)	298
98	IR Spectrum of Cycloartobiloxanthone (17)	299
99	UV Spectrum of Cycloartobiloxanthone (17)	300
100	The 600 MHz ¹ H NMR Spectrum of Cycloartobiloxanthone (17)	301
101	COSY Spectrum of Cycloartobiloxanthone (17)	302
102	¹³ C NMR Spectrum of Cycloartobiloxanthone (17)	303
103	¹³ C NMR Spectrum of Cycloartobiloxanthone (17) (expansion 1)	304
104	¹³ C NMR Spectrum of Cycloartobiloxanthone (17) (expansion 2)	305
105	DEPT Spectrum of Cycloartobiloxanthone (17)	306
106	HMQC Spectrum of Cycloartobiloxanthone (17)	307
107	HMQC Spectrum of Cycloartobiloxanthone (17) (expansion)	308
108	HMBC Spectrum of Cycloartobiloxanthone (17)	309
109	EIMS Spectrum of 2'-Hydroxy-4'- <i>O</i> -prenylacetophenone (150)	310
110	The 300 MHz ¹ H NMR Spectrum of 2'-Hydroxy-4'- <i>O</i> -prenyl-acetophenone (150)	311
111	EIMS Spectrum of 2',4',6'-Trihydroxy-3'-prenyl-acetophenone (157)	312
112	IR Spectrum of 2',4',6'-Trihydroxy-3'-prenylacetophenone (157)	313
113	The 300 MHz ¹ H NMR Spectrum of 2',4',6'-Trihydroxy-3'-prenylacetophenone (157)	314
114	The 300 MHz ¹ H NMR Spectrum of 2',6'-Dihydroxy-4'- <i>O</i> -prenylacetophenone (158) and 2',4'-Dihydroxy-6'- <i>O</i> -prenylacetophenone (159)	315
115	The 300 MHz ¹ H NMR Spectrum of 2',6'-Dihydroxy-4'- <i>O</i> -prenylacetophenone (158) and 2',4'-Dihydroxy-6'- <i>O</i> -prenylacetophenone (159) (expansion)	316

116	EIMS Spectrum of 2'-Hydroxy-4',6'-dimethoxy-3'-prenylacetophenone (155)	317
117	IR Spectrum of 2'-Hydroxy-4',6'-dimethoxy-3'-prenylacetophenone (155)	318
118	The 300 MHz ¹ H NMR Spectrum of 2'-Hydroxy-4',6'-dimethoxy-3'-prenylacetophenone (155)	319
119	EIMS Spectrum of 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154)	320
120	IR Spectrum of 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154)	321
121	The 300 MHz ¹ H NMR Spectrum of 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154)	322
122	The 300 MHz ¹ H NMR Spectrum of 2'-Hydroxy-4,4',6'-trimethoxy-3'-prenylchalcone (154) (expansion)	323
A	Flavones From <i>Artocarpus scortechinii</i> King	
B	Flavonoids From <i>Artocarpus teysmanii</i> Miq.	

CHAPTER 1

INTRODUCTION

1.1 General Introduction

Malaysia is blessed with an abundance of untapped variety of medicinal plants as she is among the world's 12 mega biodiversity-rich countries. Our diverse ethnic communities meant that Malaysia has inherited a unique confluence of deep-rooted traditional medicine systems, including those practiced by our indigenous people. These elements have put Malaysia in an exceptional position to tap into the herbal medicine potential. Researchers from all over the world are racing against time to find cures for diseases and ailments such as HIV, cancer, Alzheimer's, Parkinsons and meningitis. Phytochemical research has provided stimulation in organic synthesis of potentially superior agents and in providing more compounds to be used as tools to understand the biochemical mechanisms involved in the occurrence of certain diseases. We believe the elixir is locked in the secrets of the rainforest plants. Due to this reason, initiation is taken to further explore higher plants for biologically active compounds.

Malaysia has about 12,000 species of flowering plants of which about 1300 are said to be medicinal [1]. The Taman Negara Reserve Rainforest in Pahang is considered the grandmother to the rest of the world's rainforests. It is considered to be one of the richest natural environments on earth with 10,000 species of plants, 350 species of birds,

100 types of snakes, 1000 varieties of butterflies, 150,000 kinds of insects and 140 types of animals. The huge diversity of Malaysian flora means that we can expect well diverse chemical structures from their secondary metabolites. However, there is still more effort to be made locally to establish and to develop the available plants into useful and valuable pharmaceutical products [2].

Natural products isolated from higher plants have been providing novel, clinically active drugs. The development of medicinal plants into therapeutic drugs takes several years and a substantial amount of money is needed. The process is very capital-intensive, high risk and the success rate is not very good. Despite all these, natural products drug discovery programmes in Malaysia are still existing, mainly because the potential of high chemical diversity from natural products is largely unknown and the large number of our terrestrial species have yet to be investigated. It is believed that there is still a lot more waiting for discovery as what have been studied to date is just a small fraction [3]. Bioassay-guided research and multidisciplinary concepts were introduced so that the research carried out can be more meaningful. The Government of Malaysia has taken steps to increase the scientific knowledge of our rich source of medicinal plants by making available funds for research beginning in 1985 with the Intensification of Research in Priority Areas (IRPA) programme. One of the areas that have been identified as a priority is the commercialization of biotechnology, which also takes into account the development and production of biopharmaceuticals from plant genetic resources.

1.2 Family Moraceae

Moraceae is a large family comprising of about 60 genera and approximately 1400 species that form a significant element in the flora of the tropical region of Southeast Asia [4]. The most important genera are *Morus*, *Ficus* and *Artocarpus*. Only nine genera and 137 species could be found in Malaysia, distributed from lowlands to mountain forests. *Ficus* and *Artocarpus* plants are quite abundant in Malaysian forests. Several members of this family produce valuable timbers and edible fruits. Another important economic plant is the mulberry tree or *Morus*.

Morus is a small genus found primarily in temperate and subtropical regions of the Northern Hemisphere and has been widely cultivated in China and Japan for its leaves, which are fed to silkworms. *Morus nigra* (black mulberry) can grow up to 35 feet high and forms a compact crown. The heart-shaped leaves are usually whole, except when younger, they often separate into several lobes. The berries are dark red when ripe and can be eaten fresh or used to make jams. *M. rubra* (red mulberry) is the largest of all the Mulberries and can grow from 60 to 70 feet high. The leaves are oval or oblong heart-shaped with a pointed tip and serrated edges. The edible berries are dark red while the wood can be used for light carpentry. *M. alba* (white mulberry) can grow up to 50 feet high and produces extremely sweet, pinkish, white, or purplish berries.

Ficus is a large genus with about 600 species that is commonly found in the tropical regions. *Ficus* in Malaysia is a big and ubiquitous genus, consists of 101 species. The trees grow easily in all types of forest where the seeds are spread by small mammals and birds. Some of the species are deciduous while others like *Ficus benjamina* and *F. microcarpa* are evergreen. *Ficus* is popular as a landscape tree because of its deep green leaves and long red stipules. The trees are suitable for 'bonsai' as it can grow fast, tolerant to most soil and light condition. Only several species of *Ficus* produce edible fruits [4].

1.3 Genus *Artocarpus*

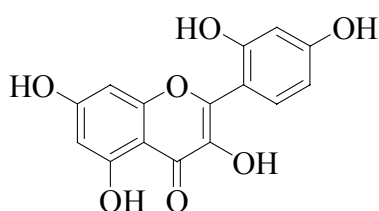
Artocarpus is the most commonly encountered genus, representatives of the Moraceae in the lowland forest of the tropical Southeast Asia, apart from *Ficus*. There are 47 species of *Artocarpus* in which only 20 species including the cultivated plants could be found in Malaysia. This genus is known world wide for its edible fruits like the jackfruit, *A. heterophyllus* locally known as 'nangka', bread fruit, *A. communis* ('sukun') and 'cempedak', *A. integer*. These species are widely cultivated in Malaysia as villagers and traders commercially sell their fruits in local market. The lightwood known locally as 'terap' and the medium hardwood known as 'keledang' constitute valuable timber resources [4-5]. Some of Malaysian *Artocarpus* species are rare. Most of these species

have never been chemically investigated including *A. anisophyllus*, *A. bracteata*, *A. fulvicortex*, *A. hispidus*, *A. kemando*, *A. lowii*, *A. nitidus*, and *A. odoratissima* [4].

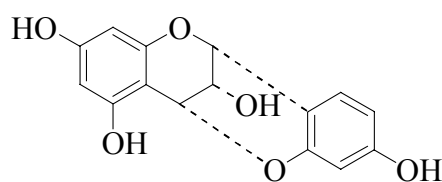
1.4 A Review of Phytochemicals and Biological Properties of *Artocarpus* Species

Artocarpus species have been studied quite thoroughly, chemically or biologically by few groups of researchers from Indonesia, Japan, and Taiwan. Most of the plants studied were collected from the rain forest of Indonesia. Some studies cover *Artocarpus* species of Taiwan, Caribbean and Thailand. Several new and interesting compounds have been isolated, characterized and evaluated for their biological activities. *Artocarpus* species are noted as an abundant source of phenolic constituents. These constituents can be classified into isoprenylflavonoids, stilbenoid and 2-arylbenzofuran derivatives, phenolic compounds with oxepine ring and natural Diels-Alder type adducts.

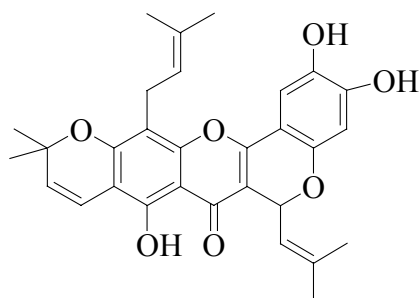
The earlier work on the phytochemical investigation of *Artocarpus* species started long ago in 1895 where morin (**1**) and cyanomaclurin (**2**) were isolated from *A. heterophyllus*. It was only in 1963 that a study of the NMR spectrum of the acetate of cyanomaclurin trimethyl ether led to the structure of cyanomaclurin (**2**) [6]. Two more prenylflavonoids were isolated in very minute quantities from the same species and identified as cycloheterophyllin (**3**) and heterophyllin (**4**) [7]. Since then, many types of new isoprenoid-substituted phenolic compounds were isolated from *Artocarpus* species.



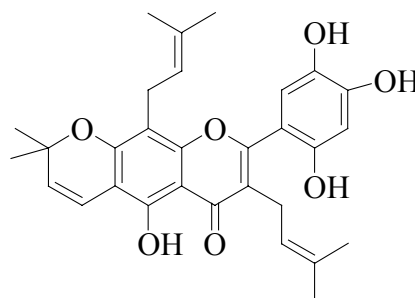
(1)



(2)



(3)

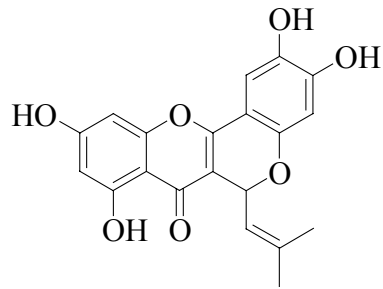


(4)

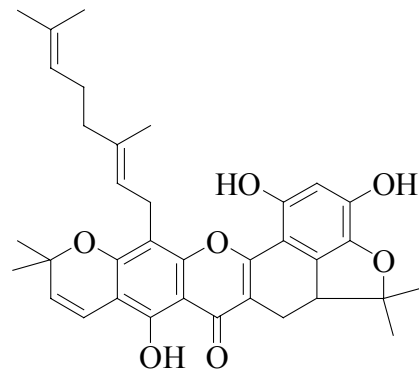
1.4.1 Isoprenylflavonoids

Most of the isolated isoprenylflavonoids have common features of hydroxyl groups in the 5,7,2',4'- positions and C- γ , γ -dimethylallyl or isoprenyl substituents in the 6-, 3,6- or 3,6,8-positions of the flavone skeleton [6]. These remarkable structural features of the flavonoids correlate with their biological activities very significantly.

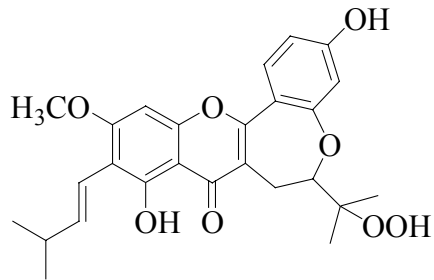
Nine new isoprenylflavonoids were isolated from the tree bark, root and heartwood of *Artocarpus champeden*, Spreng. This species is an endemic species found throughout the Indonesian archipelago. The fruits, locally known as 'cempedak' are eaten as staple food and its wood is used commercially as timber. In Malaysia, this species is known as *A. integer* [4]. The compounds isolated were characterized spectroscopically as cyclochampedol (5), artoindonesianins A-B (6-7), artoindonesianins Q-T (8-11), and artoindonesianins U-V (12-13) [8-11]. Cyclochampedol (5) was shown to be toxic to brine shrimps (*Artemia salina*) [8], whereas artoindonesianin A-B (6-7) and artoindonesianins U-V (12-13) exhibited cytotoxic effects against murine P388 leukemia cells [9, 11].



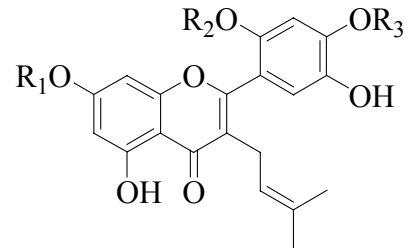
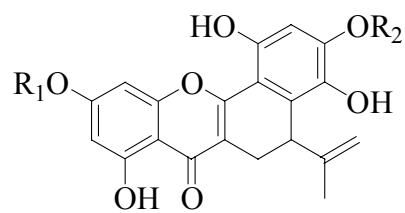
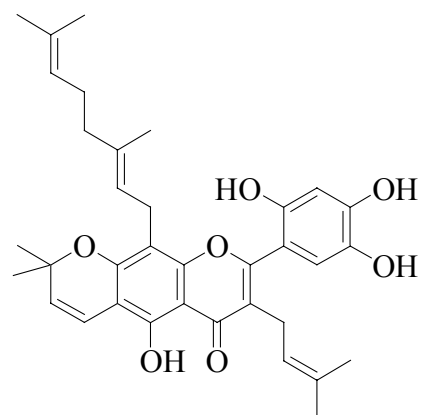
(5)



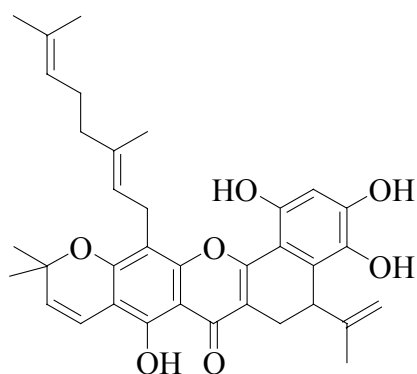
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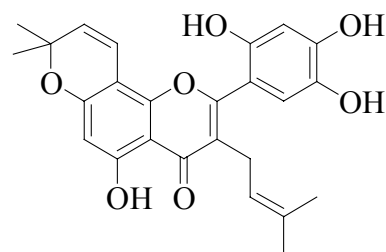
(7)

(8) $R_1 = R_3 = \text{CH}_3$, $R_2 = \text{H}$ (9) $R_1 = \text{H}$, $R_2 = R_3 = \text{CH}_3$ (10) $R_1 = R_2 = \text{CH}_3$ (11) $R_1 = \text{H}$, $R_2 = \text{CH}_3$ 

(12)

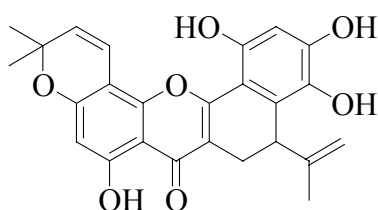


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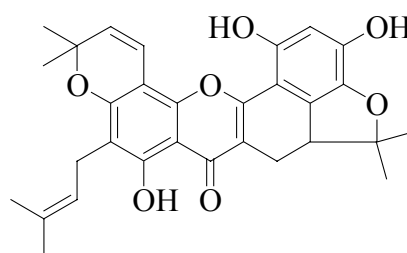


(14)

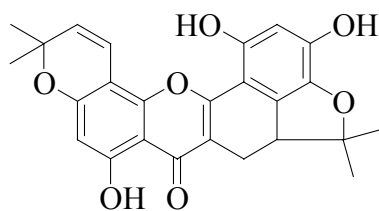
The dried bark of *Artocarpus communis* Forst. (synonym *A. altilis*) contained isoprenylflavonoids identified as artonin E (**14**), artobiloxanthone (**15**), artonin F (**16**), cycloartobiloxanthone (**17**), and artonols A-E (**18-22**) [12-14]. This species is widely distributed throughout the tropical area in Southeast Asia, especially in Malaysia and Indonesia. In Malaysia, this species is known as ‘sukun’ while in Indonesia, there are two varieties; namely ‘kukur’ (breadfruit tree) and the other which produce edible fruits called ‘sukun’ (seedless breadfruit tree) [12]. The leaves are used for hepatomegalis and febris, while the flowers are used against parulis and adontalgia. Furthermore, the dried flower of the plant has been used as a mosquito repellent [13].



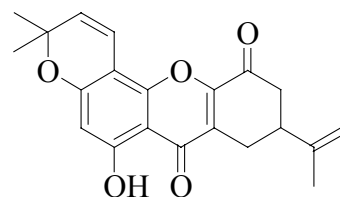
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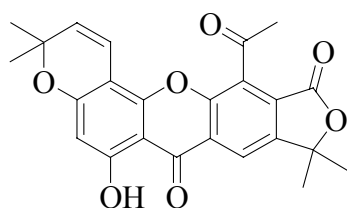
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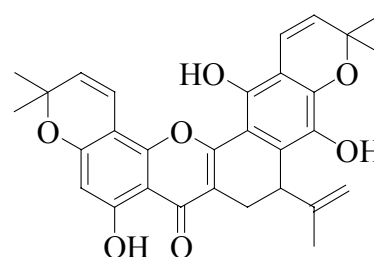
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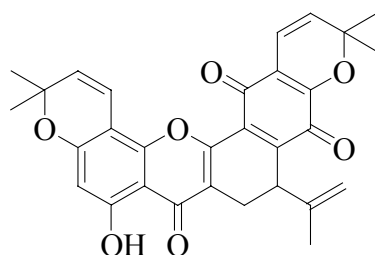
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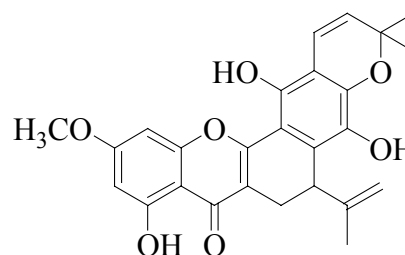
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(20)

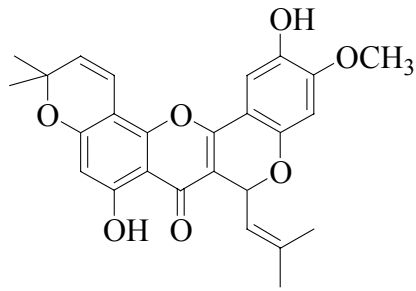


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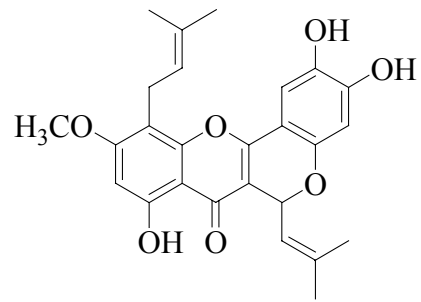


(22)

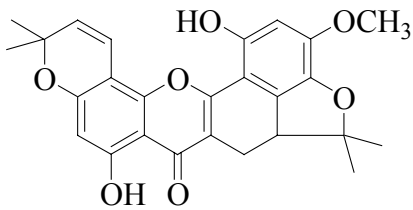
Nine new isoprenylflavonoids were successfully isolated from the root bark of *Artocarpus communis* of Taiwan [15-18]. The compounds were characterized as cycloartomunin (**23**), dihydrocycloartomunin (**24**), cycloartomunoxanthone (**25**), artomunoxanthone (**26**), artomunoxanthentrione (**27**), artomunoxanthotrione epoxide (**28**), cyclocommunol (**29**), cyclocommunin (**30**) and dihydroisocycloartomunin (**31**) [15-18]. Another five prenylflavonoids were isolated from the cortex of the roots of *A. communis* of Taiwan. These compounds were characterized as artocommunols CA (**32**), CB (**33**), CC (**34**), CD (**35**) and CE (**36**) [19]. However, these studies did not include any work on their biological activities.



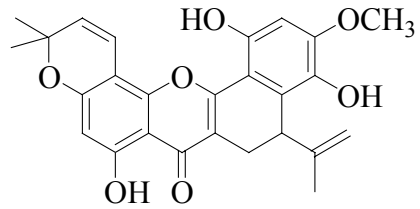
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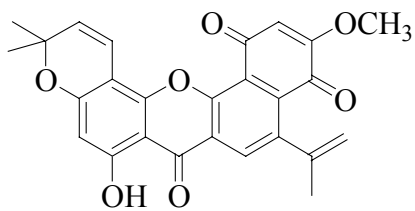
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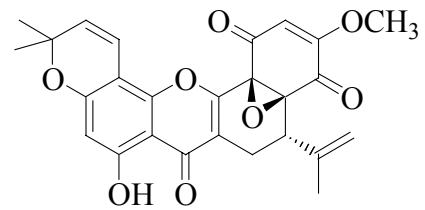
(25)



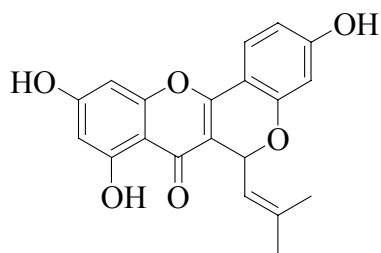
(26)



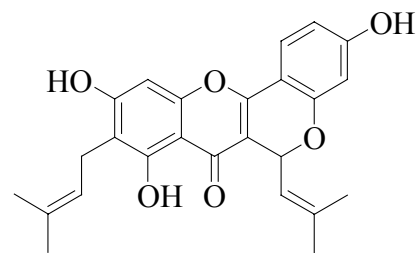
(27)



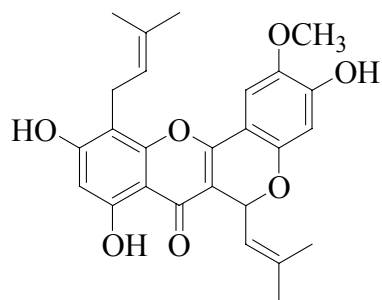
(28)



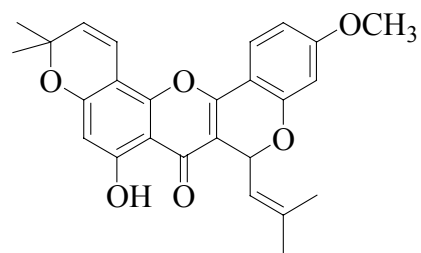
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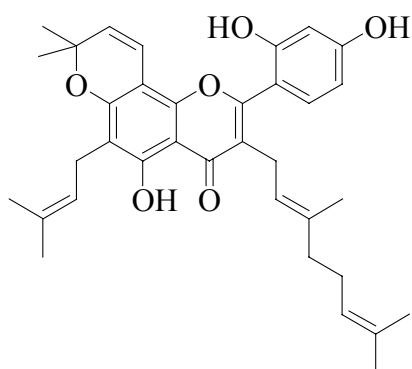
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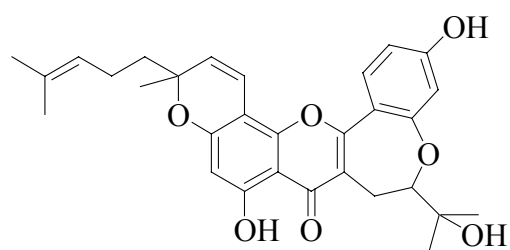
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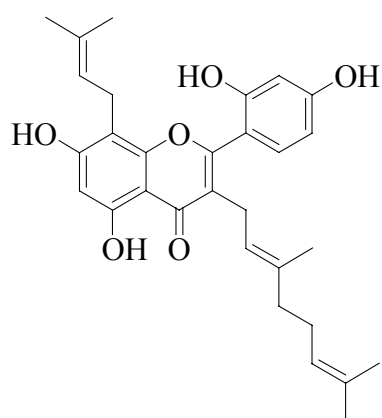
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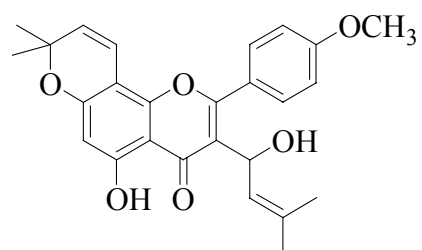
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(34)



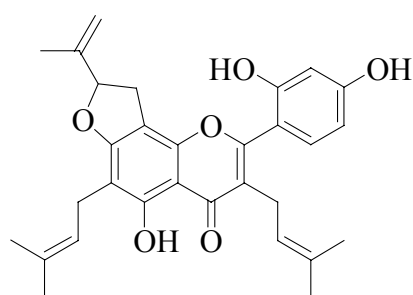
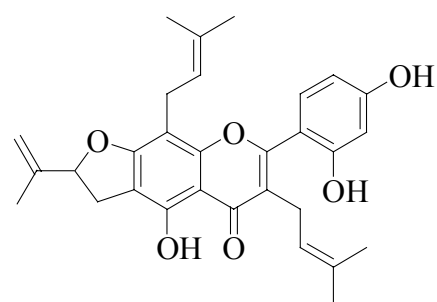
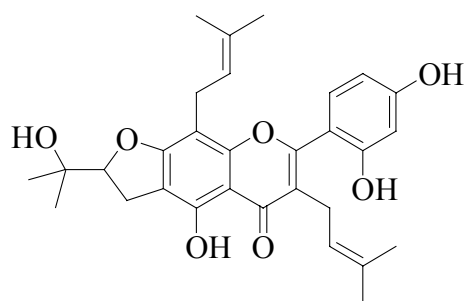
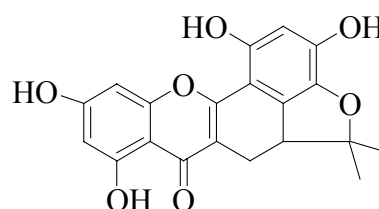
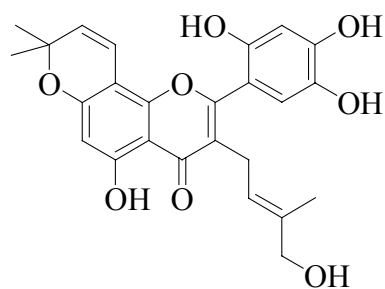
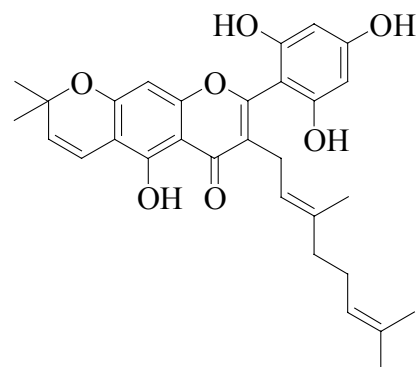
(35)



(36)

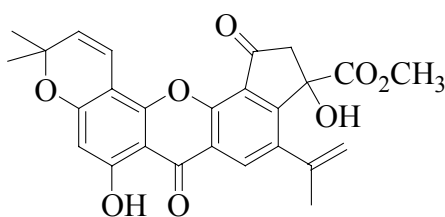
Artocarpus lanceifolius, Roxb. is a rare species endemic to lowlands and hill forests in Malaysia and the province of West Sumatra, Indonesia. It is locally known as 'keledang'. The wood is used for making coffins and for heavy construction [4].

Phytochemical and biological studies have been carried out on the heartwood and the tree bark of *A. lanceifolius* of Indonesia. Five new prenylated flavones were isolated. These flavones were identified as artoindonesianins G-I (**37-39**), artoindonesianin P (**40**) and 14-hydroartonin E (**41**) [20-23]. Biological evaluation of artoindonesianins G-I (**37-39**) and artoindonesianin P (**40**) showed that these compounds exhibited cytotoxicity effect against murine P388 leukemia cells [21-22].

**(37)****(38)****(39)****(40)****(41)****(42)**

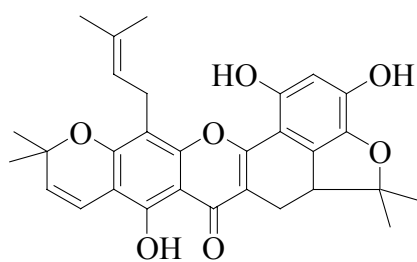
Artocarpus rotunda (Hout) Panzer is a wild tree growing in Southern Sumatera, Indonesia. Studies on the root bark of this species have resulted in the isolation of a new prenylated flavone, named artoindonesianin L (**42**). This flavone showed significant cytotoxicity against murine P388 leukemia cells [24].

Artocarpus teysmanii Miq. is a rare species found in swampy areas of West Coast of Peninsular Malaysia, Sumatera, South Sulawesi and Western New Guinea [4]. A new xanthone derivative, artoindonesianin C (**43**) together with known cycloartobiloxanthone (**17**) and artonol B (**19**) were isolated from the root bark of this species. The two known compounds were found to be active in the *Artemia salina* bioassay, while artoindonesianin C (**43**) was shown to be inactive [25-26].

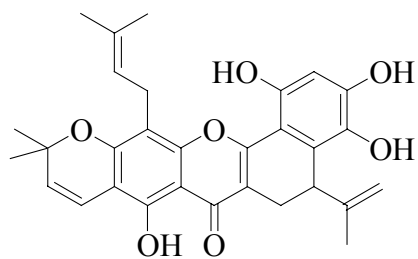


(43)

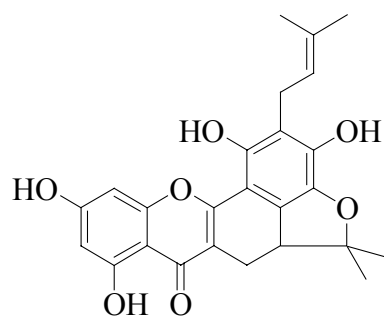
Artocarpus heterophyllus Lamk. or jackfruit is a laticiferous fruit tree probably native to Southern India and cultivated throughout the tropical world including Malaysia. *A. rigida* Bl. or *A. rigidus* Bl. is locally known as ‘temponok’. There are four varieties of *A. rigidus* Bl. in Malaysia viz. “hispidus”, “asperulus”, “tomentosa”, and “glabra”. These species can be found throughout the lowland and hill forests of Malaysia, Thailand and Indonesia [4]. These two species had been intensively investigated and contained a substantial amount of prenylflavonoids. Among the new prenylflavonoids isolated were artonins A-B (**44-45**), artonins J-L (**46-48**), artonins Q-U (**49-53**) from the root bark and tree bark of *A. heterophyllus* Lamk. [27-29] and artonins G-H (**54-55**), artonins M-P (**56-59**) from the tree bark of *A. rigida* Bl. [30-31].



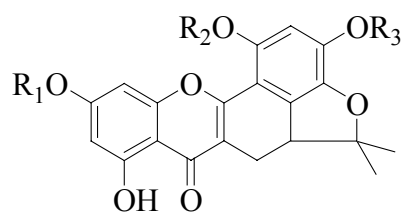
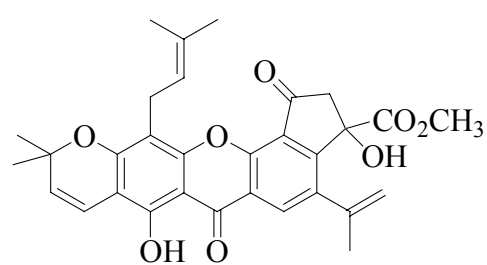
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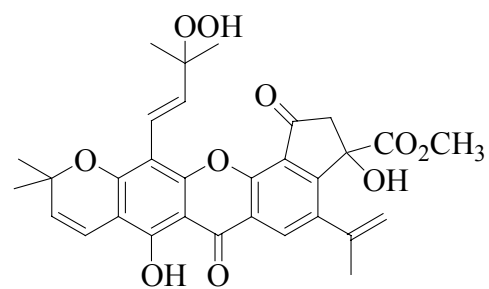
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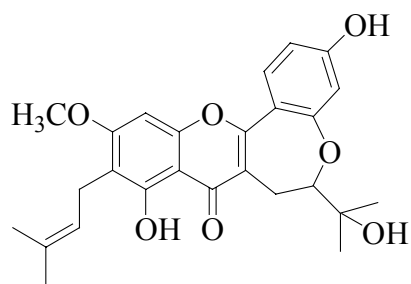
(46)

(47) $R_1 = \text{CH}_3, R_2 = R_3 = \text{H}$ (48) $R_1 = R_2 = \text{CH}_3, R_3 = \text{H}$ 

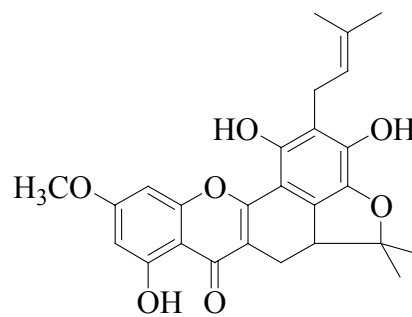
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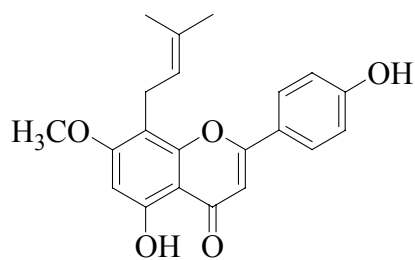
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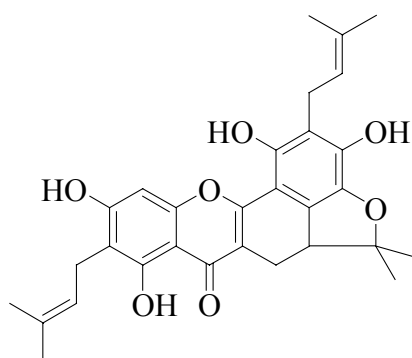
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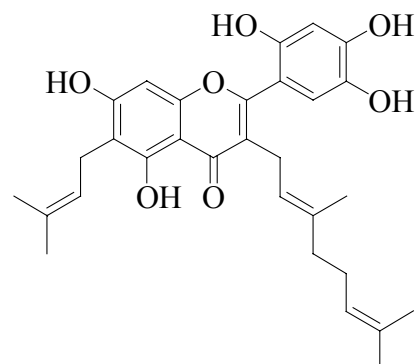
(52)



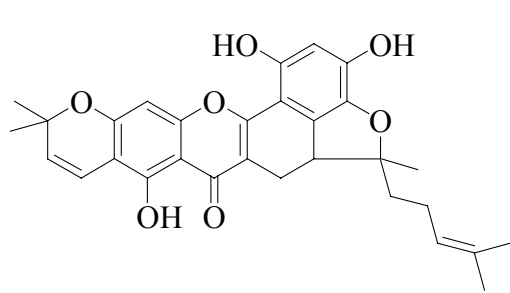
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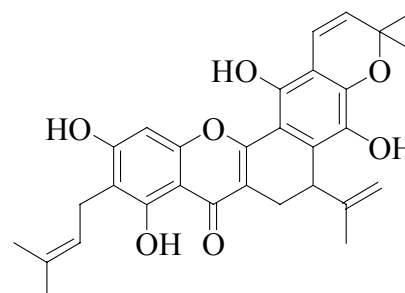
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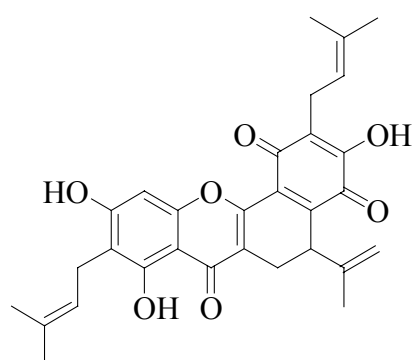
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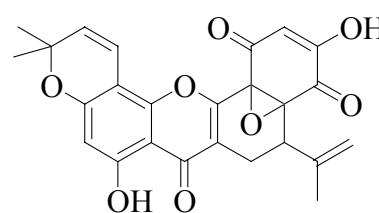
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(57)

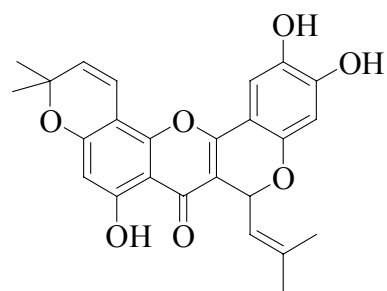


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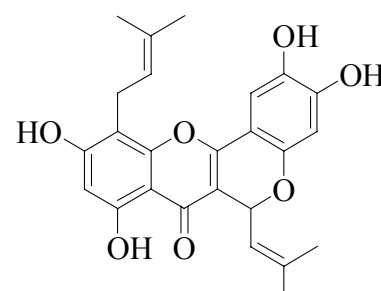


(59)

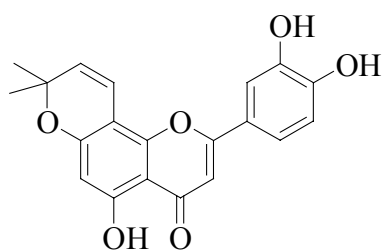
Artocarpus chama Buch.-Ham can be found growing in Yunnan, China. Investigation on the roots of this species has led to a report of five new isoprenylated flavones. These flavones were elucidated spectroscopically and identified as artochamins A-E (**60-64**). Artochamin C (**62**) was found to be potent against human lung carcinoma (A549) and breast adenocarcinoma (MCF7) [32].



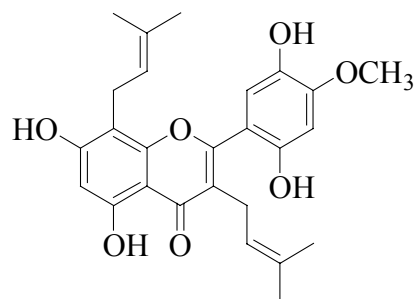
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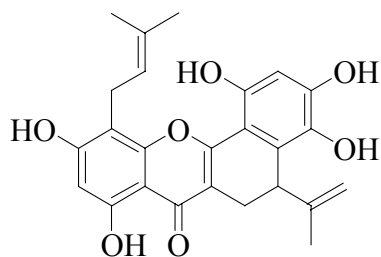
(61)



(62)

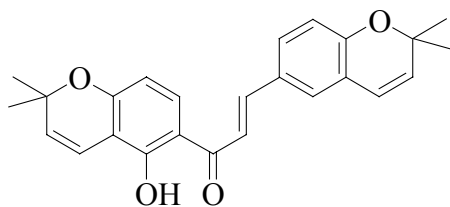


(63)

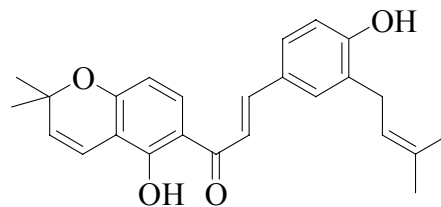


(64)

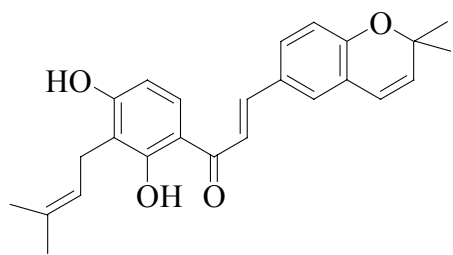
Besides isoprenylflavones, there are a few reports on isoprenylchalcones, dihydrochalcones and isoprenylflavanones from *Artocarpus* species. Among the species that contained these constituents are *Paratocarpus venenosa* Zoll., *A. altilis*, and *A. nobilis* Thw.. Seven new isoprenoid-substituted chalcones and five isoprenoid-substituted flavanones were isolated from the tree bark of *Paratocarpus venenosa* Zoll. collected in Bogor, Indonesia. These compounds were characterized as paratocarpins A-G (65-71) and paratocarpins H-L (72-76) [33-34].



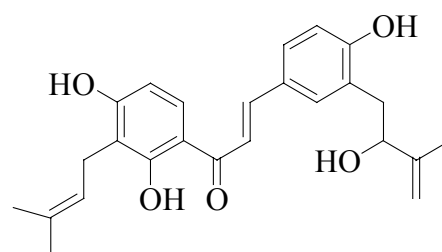
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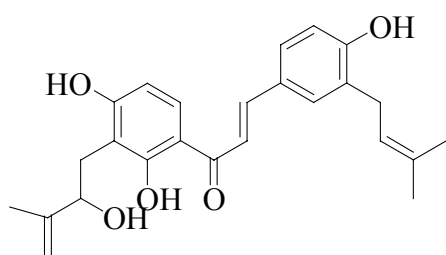
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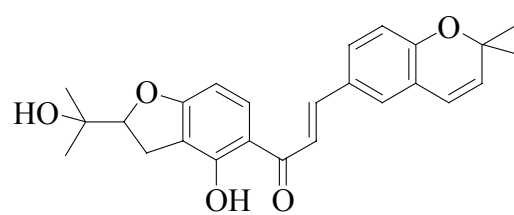
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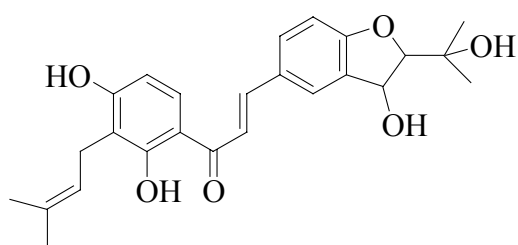
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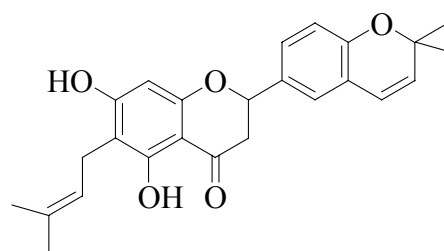
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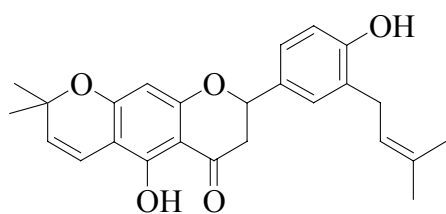
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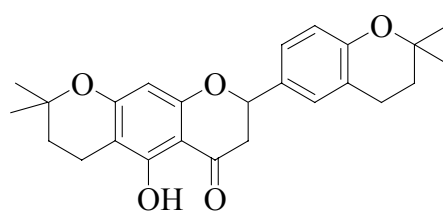
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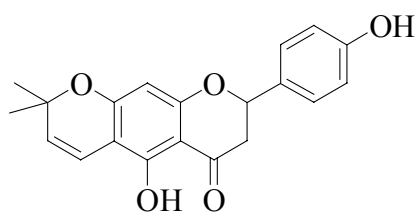
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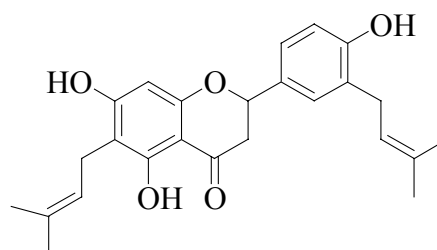
(73)



(74)

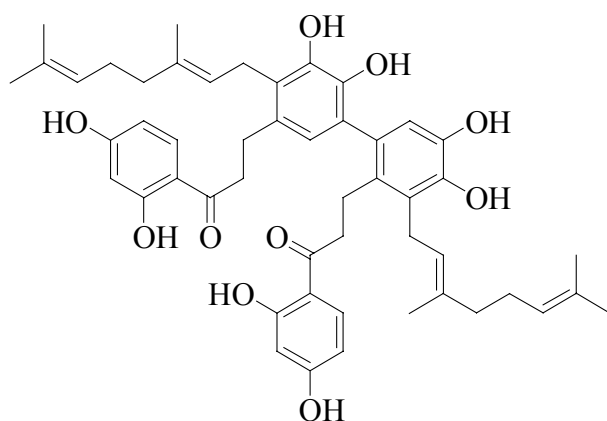


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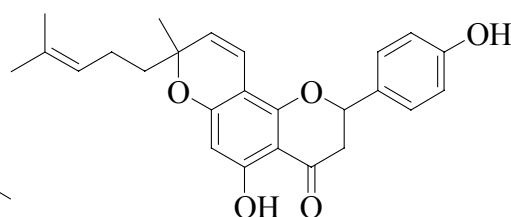


(76)

The buds of *Artocarpus altilis* have been used traditionally in Taiwan for the treatment of liver cirrhosis and hypertension and have been reported to possess anti-inflammatory and detoxifying effects. Investigation on the bud covers of *A. altilis* has led to the isolation of a new dimeric dihydrochalcone, cycloaltisin 6 (**77**) and an isoprenylflavanone, cycloaltisin 7 (**78**). Both compounds showed activity in a cathepsin K inhibition assay with IC_{50} values of 98 and 840 nM, respectively. Cathepsin K is a novel cysteine protease that has been implicated in osteoporosis. It has been established that cysteine protease inhibitors are very effective in preventing bone resorption [35].

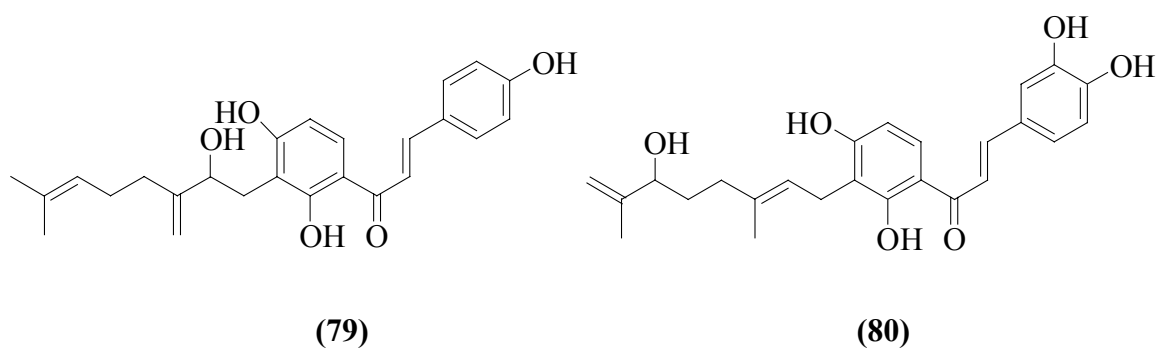


(77)



(78)

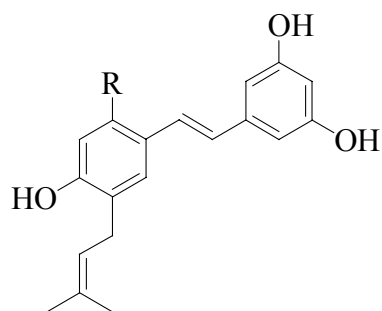
Artocarpus nobilis is a tree of moderate size and the only endemic species of the genus *Artocarpus* found in Sri Lanka. Two new chalcones were isolated from the leaves of this species and identified as 2', 4', 4'-trihydroxy-3'-(2-hydroxy-7-methyl-3-methylene-6-octaenyl)chalcone (**79**) and 2', 3, 4, 4'-tetrahydroxy-3'-(6-hydroxy-3,7-dimethyl-2*E*,7-octadienyl)chalcone (**80**). These compounds showed significant fungicidal activity against *Cladosporium cladosporioides* and high radical scavenging activity towards DPPH radical in TLC bio-autography method [36].



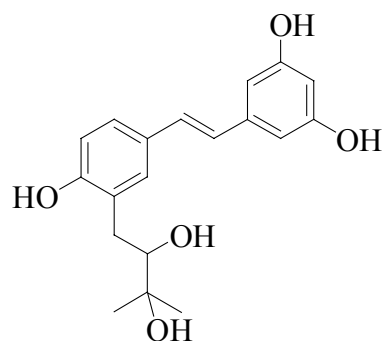
1.4.2 Stilbenoid and 2-Arylbenzofuran Derivatives

Artocarpus species also contain the biosynthetically related stilbene and 2-arylbenzofuran derivatives although their distributions are more limited. Stilbenoids are bibenzyl compounds produced *via* the mixed phenylpropanoid or polyketide biosynthetic pathway. Among the species reported to have these types of compounds are *A. dadah*, *A. fretessi*, *A. gomezianus*, *A. heterophyllus*, *A. integer*, *A. incisus*, and *A. tonkinensis* [37-43].

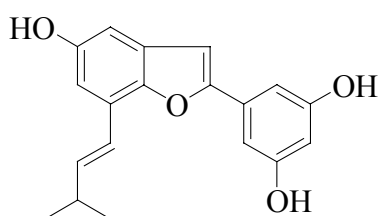
A. dadah Miq. is known as ‘tampang’ in Malaysia and Kalimantan, Indonesia and its bark has been used as an ingredient in the betel nut chewing mixture. Three new prenylated stilbenoid derivatives, 3-(3, 3-dimethylallyl)resveratrol (**81**), 5-(3, 3-dimethylallyl)oxyresveratrol (**82**) and 3-(2,3-dihydroxy-3-methylbutyl)resveratrol (**83**), and a new benzofuran derivatives, 3-(3, 3-dimethylpropenyl)morusin M (**84**) were isolated from the bark and twigs of *A. dadah* Miq. [37].



(81) R = H
 (82) R = OH

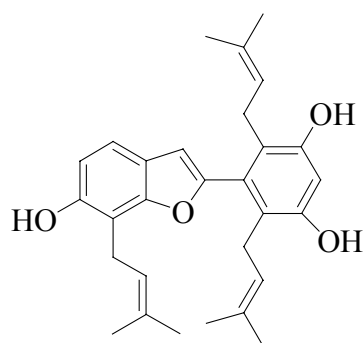


(83)

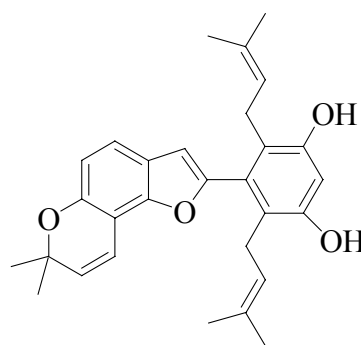


(84)

Two new isoprenylated arylbenzofurans, namely artoindonesianins X (**85**) and Y (**86**) were isolated from the root bark of *Artocarpus fretessi* Hassk. These compounds showed moderate activity against the brine shrimp, *Artemia salina* [38].

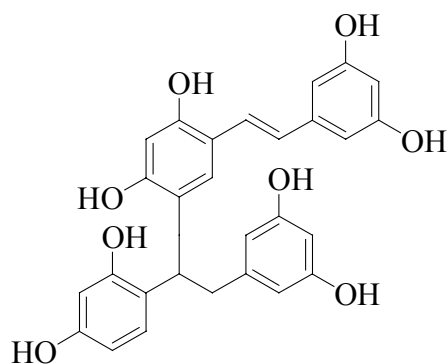
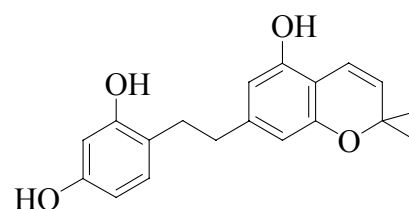


(85)

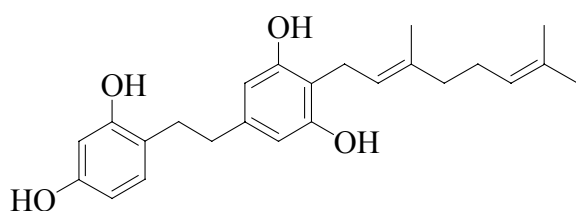
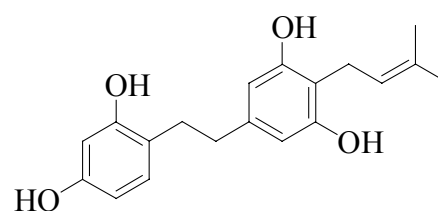


(86)

Study on the roots of *Artocarpus gomezianus*, Wall ex Tre'c revealed the presence of a new dimeric stilbene, artogomezianus (**87**) which displayed potent tyrosinase inhibitory activity [39]. Investigation of tyrosinase inhibitors may provide important clues for developing new insects control agents [40]. In plants, this enzyme is responsible for the browning of some fruits and vegetables; therefore its inhibitors may have potential uses as food preservatives. In man, potent tyrosinase inhibitors, such as kojic acid have been used as whitening agents in cosmetic products, due to their ability to suppress dermal melanin production [39-40].

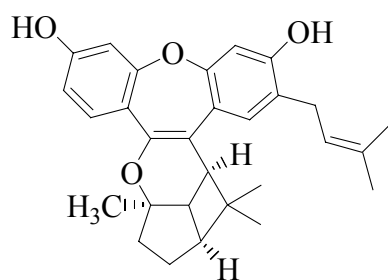
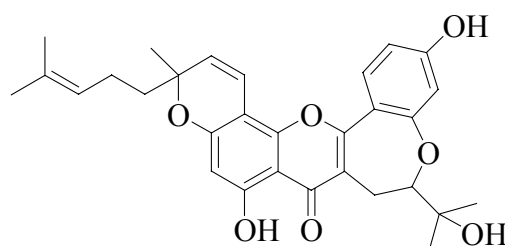
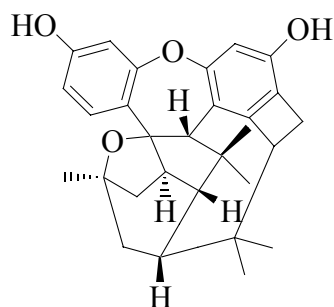
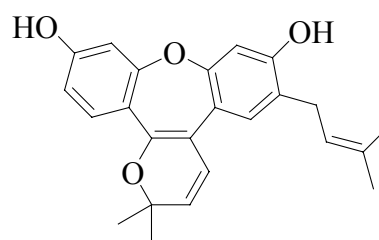
**(87)****(88)**

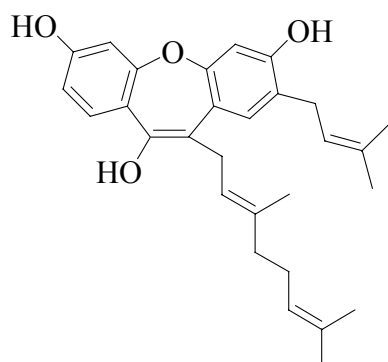
Investigation on the heartwood of *Artocarpus incisus* also revealed the presence of stilbenoid derivatives with tyrosinase inhibitory properties. These compounds were identified as artocarbene (**88**), chlorophorin (**89**) and 4-prenyloxyresveratrol (**90**) [41-42].

**(89)****(90)**

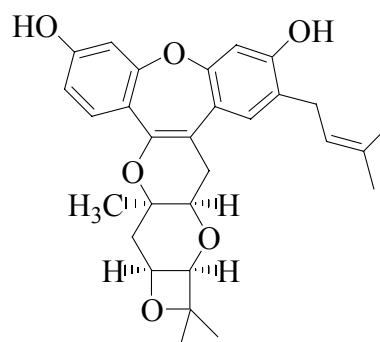
1.4.3 Phenolic Compounds with Oxepine Ring

The root barks of *Artocarpus rigida* of Taiwan were found to contain some novel compounds with an oxepine ring. These compounds were named as artocarpols A-F (**91-96**). Artocarpol A (**91**) strongly inhibited superoxide formation in phorbol 12-myristate 13-acetate (PMA) stimulated rat neutophils in a concentration-dependent manner with an IC_{50} value of $13.7 \pm 0.7 \mu\text{M}$ and also showed a significant inhibitory effect on tumour necrosis factor- α (TNF- α) formation in lipopolysaccharide (LPS)-stimulated RAW 264.7 cells. Artocarpols A (**91**), C (**93**), D (**94**) and E (**95**) are the first natural products containing an oxepine ring with a novel skeleton [43-45].

**(91)****(92)****(93)****(94)**



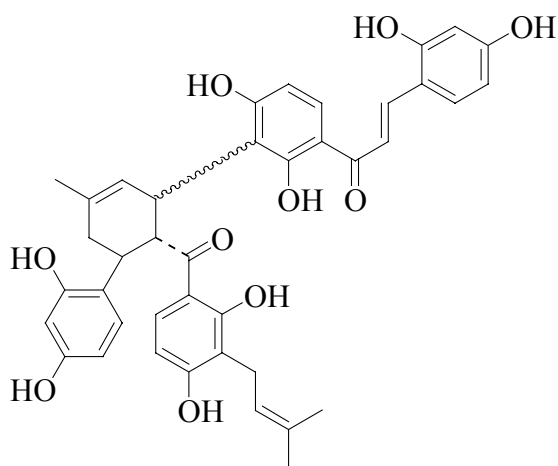
(95)



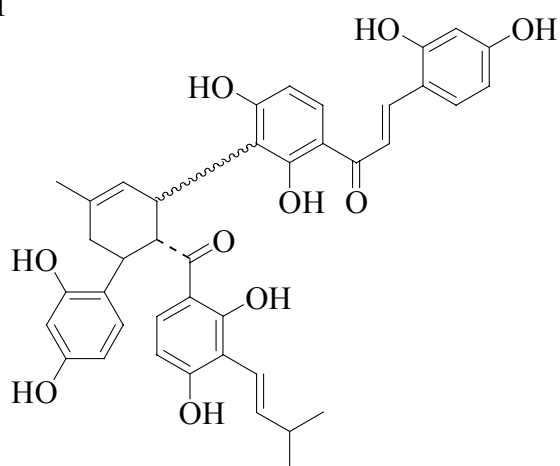
(96)

1.4.4 Diels-Alder Type Adducts

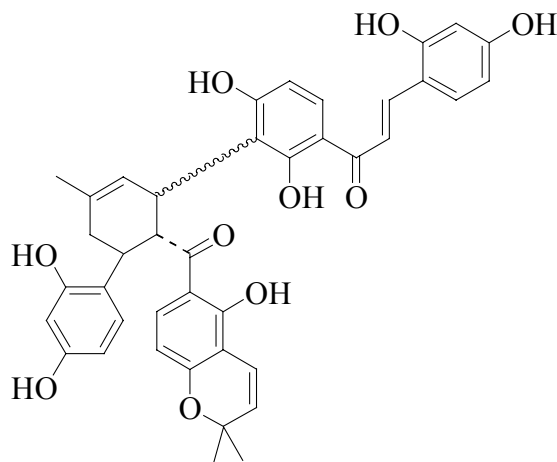
Diels-Alder type adducts are regarded as an intermolecular [4+2] cycloaddition products from the isoprenyl portion of a dehydroprenylphenols, as a diene, and the α,β -unsaturated carbon double bond of a chalcone skeleton, as a dienophile. Artonins C-D (97-98) and artonin X (99) are examples of this type of compounds, isolated from the root bark of *Artocarpus heterophyllus* [46-47].



(97)



(98)



(99)

1.5 Biosynthesis of Flavonoids

Over the past 30 years, there have been increasing reports of isoprenylated compounds belonging to different classes of flavonoids especially the isoprenylated flavones, flavanones and chalcones. In spite of the rich variety and structural diversity of isoprenylated flavonoids, these compounds have been isolated from a rather limited number of unrelated plant families especially the Leguminosae, Moraceae and Asteraceae. Prenylated flavonoids are most frequently found in roots, barks and heartwoods, but also occur in the aerial parts, buds and seeds [48].

All flavonoids derive their carbon skeletons from two basic compounds, malonyl CoA that is synthesized from the glycolysis intermediate acetyl-CoA and carbon dioxide, and the CoA ester of a hydrocinnamic acid (**Figure 1.1**). The aromatic ring B and its adjacent 3-carbon side chain are derived from L-phenylalanine *via* the shikimate pathway, whereas ring A is formed by the head-to-tail condensation of three acetate units *via* the polyketide pathway leading to the formation of the C₁₅ chalcone intermediate [48]. Flavonoids, aurones and other diphenylpropanoids are derived from the C₁₅ chalcone intermediate and the first flavonoid, flavanone is formed by stereospecific

action of chalcone isomerase on this compound. Oxidative rearrangement of this flavanone yields an isoflavone. Introduction of a double bond between C-2 and C-3 of the flavanone leads to the abundant class of flavone. Dihydroflavonol is formed by direct hydroxylation of flavanone at C-3, which is catalysed by flavanone 3-hydroxylase. Dihydroflavonol is biosynthetic intermediate in the formation of flavonol, catechin, proanthocyanidin and anthocyanidin. The large class of flavonol is formed by introduction of a double bond between C-2 and C-3 of the dihydroflavonol [48-49]. They are summarized in **Figure 1.1**.

Generally, most flavonoids are *C*-prenylated, whereas *O*-prenylation is quite rare. *C*-prenylation takes place more frequently on ring A at C-6 or C-8, as well as C-3' or C-5' especially in flavanones and flavones. From the biosynthetic point of view, it is agreed that the basic skeleton of the different flavonoid classes, including isoflavonoids, is constructed before any isoprenoid substituents are added. A structural analysis of prenylated flavonoids from Leguminosae suggests that all modifications of ring A occur at the chalcone stage including isoprenylation, β -hydroxylation and elimination of the C-6' hydroxyl group or 2'-*O*-methylation. In addition, cyclization of the prenyl substituent is determined by the *O*-methylation pattern of C-2' and/or C-6' hydroxyl groups. *C*-prenylation usually occurs *ortho* to a phenolic hydroxyl group e.g. at positions C-6, C-8, C-3', C-4' or C-5', except for C-3 prenylation [49].

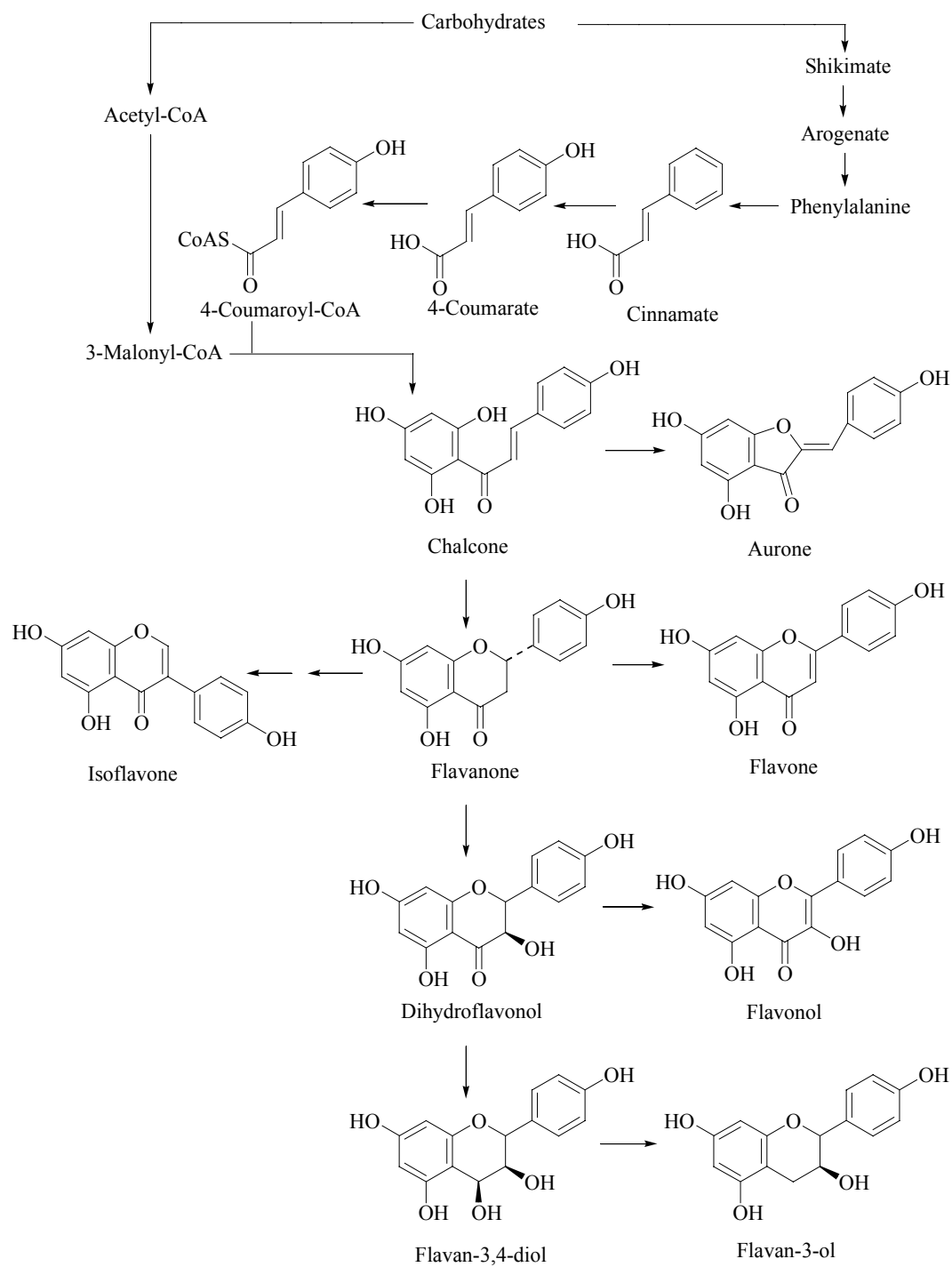


Figure 1.1: Biosynthetic Pathway of Flavonoids [48-49]

1.6 Synthesis of Flavonoids

The flavonoids are a very well known family of natural products found almost exclusively in the plant kingdom, most of them are highly coloured and, as a consequence, play a vital role in the ecology of plants by making flowers and fruits attractive to bees and birds. Many naturally occurring flavonoids are known to have significant biological activities. They were reported to be useful as antioxidants, anti-inflammatories, pulmonary carcinogenesis inhibitors, antimalarials, and antileishmanials [50]. With their vast biological potential, synthesis of flavonoids and flavonoid precursors have become great interests to many researchers nowadays. The classical flavonoid synthetic routes are known by the name of their developers, viz., Claisen-Schmidt, Baker-Venkataraman, Allan-Robinson, and Algar-Flynn-Oyamada [51].

The Claisen-Schmidt reaction (**Figure 1.2**) is the most frequently used means of establishing the C₆-C₃-C₆ flavonoid nucleus owing to the availability of starting materials and comparative ease with which the reaction can be run. This reaction involves condensation of a C₆-C₂ unit, substituted 2-hydroxyacetophenone (**100**) and a C₆-C₁ unit, benzaldehyde derivatives (**101**) to obtain a 2'-hydroxychalcone (**102**) or the isomeric flavanone, bearing A-ring substituents provided by the acetophenone (indicated as R₁) and B-ring substituents provided by the benzaldehyde (indicated as R₂). The classical Claisen-Schmidt reaction is routinely run using aqueous sodium or potassium hydroxide or ethanolic sodium ethoxide at about 50°C over a period of several hours. The benzaldehyde is often used in slightly more than equivalent amounts. Besides the Claisen-Schmidt reaction, chalcones can also be synthesized by the direct Friedel-Crafts acylation of a phenol. In this approach the phenol becomes the A-ring while the acylating agent provides both the B-ring carbons and the three-carbon bridge to form the C₆-C₃-C₆ unit [51]. Chalcones became the key precursors in the synthesis of various flavonoids as they can be transformed easily to other classes of flavonoids by using different reagents and conditions (**Figure 1.3**) [51].

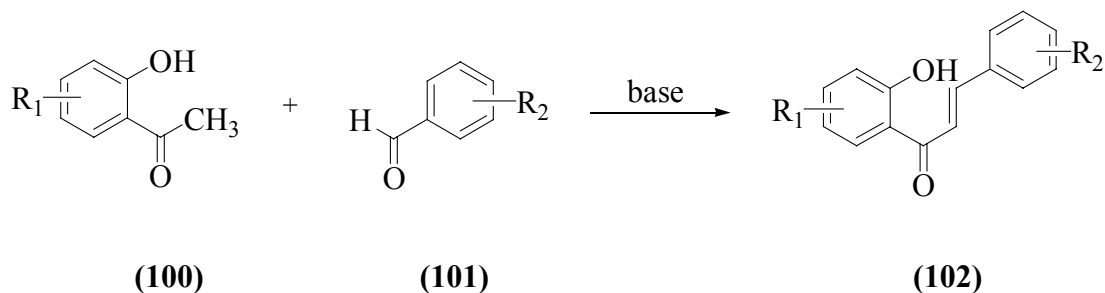


Figure 1.2: The Claisen-Schmidt Reaction

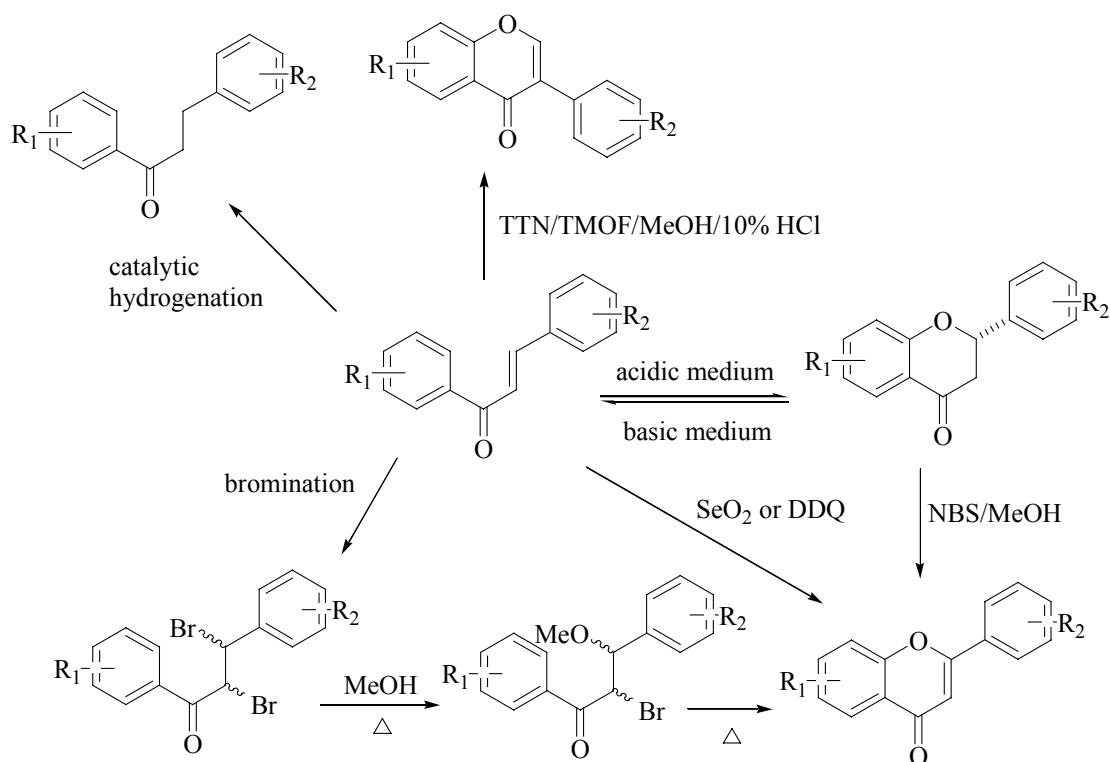


Figure 1.3: Conversion of Chalcone to Various Types of Flavonoids

The Baker-Venkataraman rearrangement involves acylation of a 2'-hydroxyacetophenone with an aromatic acid chloride at oil bath temperature in the presence of a base such as potassium carbonate or pyridine. The resulting esters are converted into β -diketone with a strong base, potassium hydroxide in pyridine or with sodium hydride. Treatment of the β -diketone with ethanol-sulphuric acid, in glacial acetic acid and

anhydrous sodium acetate, results in recyclisation to the hemiketal followed by elimination of water to form the flavone. A typical example of this method involves the conversion of the 3-methoxy-4-benzyloxybenzoyl ester of 2,5-dihydroxy-4,6-dimethoxyacetophenone (**103**) to 4',5,6-trihydroxy-3',7-dimethoxyflavone (**104**). Removal of the benzyl group from the 4'- position and the methyl group from the 5-position afforded the desired compound (**Figure 1.4**) [51-52].

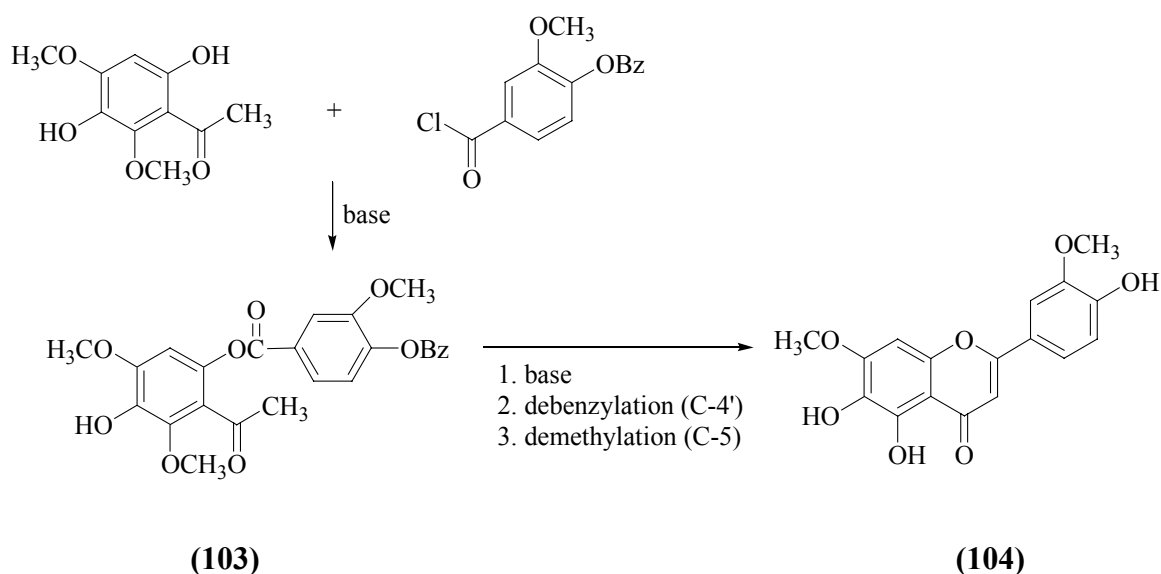


Figure 1.4: Baker-Venkataraman Rearrangement

In 1993, Ares and co-workers described a simple and convenient large-scale synthesis of 5-methoxyflavone (**108**), which employs potassium *tert*-butoxide (*t*-BuOK) in a modified Baker-Venkataraman process (**Figure 1.5**). The starting material, 2-hydroxy-6-methoxyacetophenone (**106**) was readily prepared from commercially available 2,6-dihydroxyacetophenone (**105**) using methyl iodide and potassium carbonate in acetone. Transformation of (**106**) into its potassium phenoxide anion with 1.1 equivalent of *t*-BuOK was followed by treatment with benzoyl chloride to form the benzoyl ester. A second 1.1 equivalent of *t*-BuOK was directly added to this reaction mixture, which after refluxing overnight, provided crystalline diketone (**107**) in 64-68% isolated yield. Treatment of (**107**) with sulphuric acid in refluxing acetic acid afforded 5-methoxyflavone (**108**) in 70-75% yield [53].

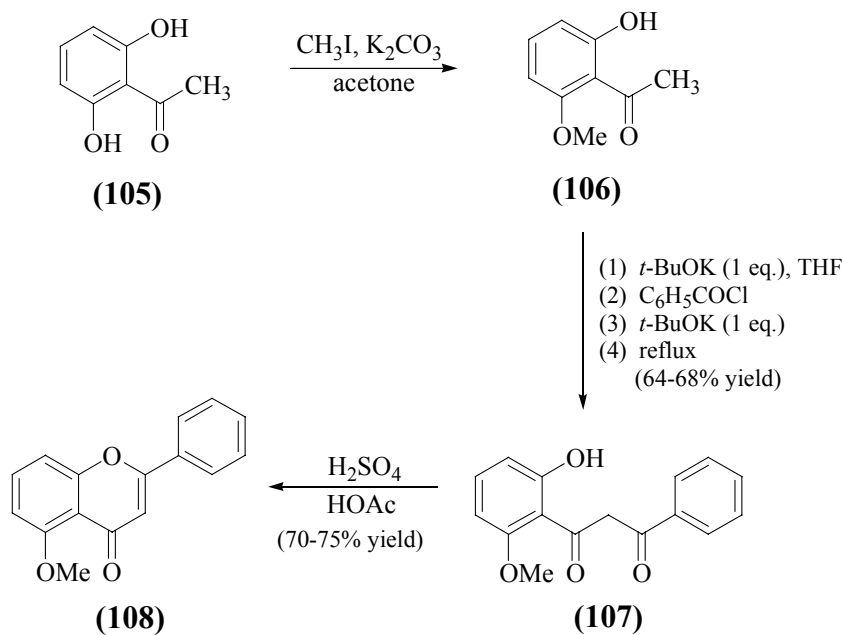


Figure 1.5: Modified Baker-Venkataraman Rearrangement

The Allan-Robinson synthesis is a variation of the Baker-Venkataraman route in which a 2'-hydroxyacetophenone derivative is heated at oil bath temperature with the anhydride of an aromatic acid and the sodium salt of that acid, or in the presence of pyridine or triethylamine as catalyst. This method has been used for the preparation of corymbosin (**109**) isolated from *Webera corymbosa* Willd. (**Figure 1.6**) [52].

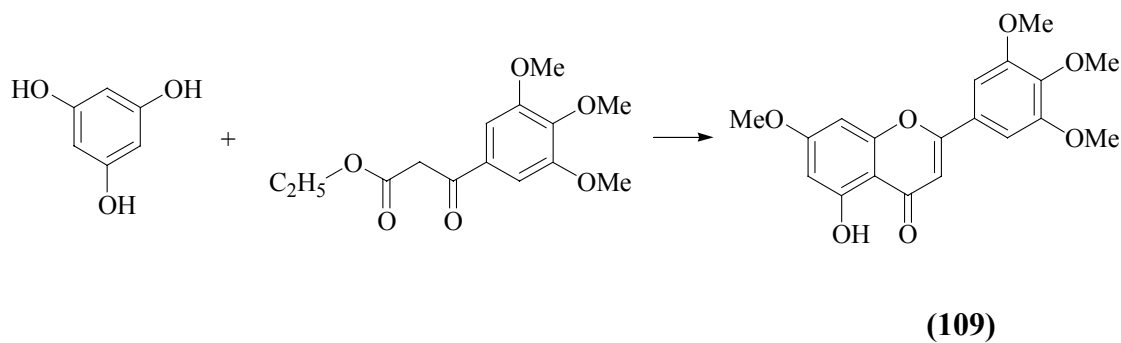


Figure 1.6: Allan-Robinson Synthesis

Algar-Flynn-Oyamada described a method for converting chalcones to flavonols or flavonol 3-methyl ethers in a single step. The method which came to be known as the AFO reaction involves oxidizing a chalcone with hydrogen peroxide in an alkaline medium (**Figure 1.7**). Epoxide (**110**) was the key intermediate in these reactions, which proceeded by the intramolecular displacement of the oxirane oxygen by the phenoxide at the β -position to give dihydroflavonol (**111**), and subsequently flavonol (**112**), or at the α -position to give aurone (**113**) [54].

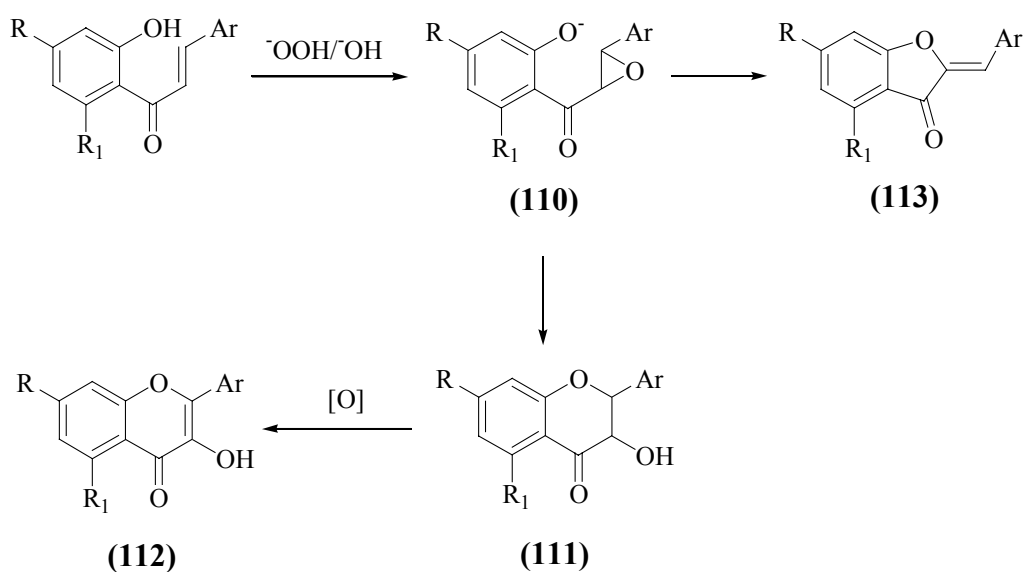


Figure 1.7: Algar-Flynn-Oyamada (AFO) Reaction

1.7 Research Objectives

The phytochemical investigations reported in the review are studies mostly on the *Artocarpus* species of Indonesia and Taiwan. A thorough literature search did not reveal any report on the chemical constituents or biological activity of Malaysian *Artocarpus* except for a report on the volatile flavour constituents of *A. polyphema* Pers. and *A. heterophyllus* Lamk. [55]. Therefore, this research will focus on the phytochemical and biological activity studies of three Malaysian *Artocarpus* i.e. *A. lowii* King ('miku'), *A. teysmanii* Miq. and *A. scortechinii* King ('terap hitam').

The objectives of this research are to extract the plants samples using different polarity of organic solvents at room temperature. The crude extracts obtained after removal of solvents will be fractionated into several fractions based on polarity using vacuum column chromatographic technique. The natural compounds of each fraction will be analyzed using thin layer chromatography and then, will be isolated by using various chromatographic techniques on silica gel or Sephadex LH20. The structures of the isolated pure compounds will be spectroscopically identified by using high field NMR, 2D NMR (COSY, HMQC, HMBC), high resolution MS, FTIR and UV. Synthesis of prenylated chalcone and flavone will also be attempted by using 2,4,6-trihydroxyacetophenone and 2,4-dihydroxyacetophenone as starting materials. Finally, evaluation on the biological activities of the crude extracts and pure compounds will be carried out by using several bioassays including antioxidant (FTC method and free radical scavenging on 2,2-diphenyl-1-picrylhydrazine (DPPH) method), antibacterial, platelet activating factor (PAF) receptor binding, platelet aggregation and cytotoxic assays.

REFERENCES

1. Burkill, I. H., A Dictionary of the Economic Products of the Malay Peninsula, Ministry of Agriculture and Cooperatives, Kuala Lumpur, Malaysia, 1935.
2. Rao, A. N., Diversity of Medicinal Plant Species in Certain Asian Countries, Their Conservation and Uses, *J. Trop. Med. Plants*, **1**, 82-108, 2000.
3. Ali, M. A. R., in Ali, A. M., Shaari, K., and Zakaria, Z., Phytochemicals and Biopharmaceutins from the Malaysian Rainforest, Forest Research Institute Malaysia, Kepong, Kuala Lumpur, v, 1999.
4. Whitmore, T. C, Tree Flora of Malaya, Longman Malaysia Sdn. Bhd., Kuala Lumpur, Vol. **3**, 119-167, 1978.
5. Ng, F. S. P., Manual of Forest Fruits, Seeds and Seedlings, Forest Research Institute Malaysia, Kepong, Kuala Lumpur, Vol. **2**, 451-454, 1992.
6. Venkataraman, K., Wood Phenolics in the Chemotaxonomy of the Moraceae, *Phytochemistry*, **11**, 1571-1586, 1972.
7. Rao, A. V. R., Mala, V., and Venkataraman, K., Colouring Matters of the Wood of *A. heterophyllus*: Part VI - Cycloheterophyllin, a Flavone Linked to Three Isoprenoid Groups, *Ind. J. Chem.*, **9**, 7-13, 1971.
8. Achmad, S. A., Hakim, E. H., Juliawaty, L. D., Makmur, L., and Suyatno, A New Prenylated Flavone from *Artocarpus champeden*, *J. Nat. Prod.*, **59**, 878-879, 1996.
9. Hakim, E. H., Fahriyati, A., Kau, M. S., Achmad, S. A., Makmur, L.,

- Ghisalberti, E. L., and Nomura, T., Artoindonesianins A and B, Two New Prenylated Flavones from the Root of *Artocarpus champeden*, *J. Nat. Prod.*, **62**, 613-615, 1999.
10. Syah, Y. M., Achmad, S. A., Ghisalberti, E. L., Hakim, E. H., Makmur, L., and Mujahidin, D., Artoindonesianins Q-T, Four Isoprenylated Flavones from *Artocarpus champeden* Spreng. (Moraceae), *Phytochemistry*, **61**, 949-953, 2002.
 11. Syah, Y. M., Achmad, S. A., Ghisalberti, E. L., Hakim, E. H., and Mujahidin, D., Two New Cytotoxic Isoprenylated Flavones, Artoindonesianins U and V, from the Heartwood of *Artocarpus champeden*, *Fitoterapia*, **75**, 134-140, 2004.
 12. Fujimoto, Y., Zhang, X.-X., Kirisawa, M., Uzawa, J., and Sumatra, M., New Flavones from *Artocarpus communis* Forst., *Chem. Pharm. Bull.*, **38**, 1787-1789, 1990.
 13. Hano, Y., Yamagami, Y., Kobayashi, M., Isohata, R., and Nomura, T., Artonins E and F, Two New Prenylflavones from the Bark of *Artocarpus communis* Forst., *Heterocycles*, **31**, 877-882, 1990.
 14. Aida, M., Yamaguchi, N., Hano, Y., and Nomura, T., Artonols A, B, C, D, and E, Five New Isoprenylated Phenols from the Bark of *Artocarpus communis* Forst., *Heterocycles*, **45**, 163-175, 1997.
 15. Lin, C. -N., and Shieh, W. -L., Prenylflavonoids and A Pyranodihydrobenzoxanthone from *Artocarpus communis*, *Phytochemistry*, **30**, 1669-1671, 1991.
 16. Shieh, W. -L., and Lin, C. -N., A Quinonoid Pyranobenzoxanthone and Pyranodihydrobenzoxanthone from *Artocarpus communis*, *Phytochemistry*, **31**, 364-367, 1992.
 17. Lin, C. -N., Shieh, W. -L., and Jong, T. -T., A Pyranodihydroxanthone Epoxide from *Artocarpus communis*, *Phytochemistry*, **31**, 2563-2564, 1992.

18. Lin, C. -N., and Shieh, W. -L., Pyranoflavonoids from *Artocarpus communis*, *Phytochemistry*, **31**, 2922-2924, 1992.
19. Chan, S. -C., Ko, H. -H., and Lin, C. -N., New Prenylflavonoids from *Artocarpus communis*, *J. Nat. Prod.*, **66**, 427-430, 2003.
20. Mujahidin, D., Achmad, S. A., Syah, Y. M., Aimi, N., Hakim, E. H., Kitajima, M., Makmur, L., Takayama, H., dan Tamin, R., Artelastokromen Suatu Diprenilpiranoflavon dan β -resorsilaldehid dari Kayu Batang *Artocarpus lanceifolius*, *Proc. ITB*, **32**, 41-49, 2000.
21. Syah, Y. M., Achmad, S. A., Ghisalberti, E. L., Hakim, E. H., Makmur, L., and Mujahidin, D., Artoindonesianins G-I, Three New Isoprenylated Flavones from *Artocarpus lanceifolius*, *Fitoterapia*, **72**, 765-773, 2001.
22. Hakim, E. H, Asnizar, Yurnawilis, Aimi, N., Kitajima, M., and Takayama, H., Artoindonesianin P, A New Prenylated Flavone with Cytotoxic Activity from *Artocarpus lanceifolius*, *Fitoterapia*, **73**, 668-673, 2002.
23. Shugeng, C., Mark, B. S., and Antony, B. D., Flavonoids from *Artocarpus lanceifolius*, *Nat. Prod. Res.*, **17**, 79-81, 2003.
24. Suhartati, T., Achmad, S. A., Aimi, N., Hakim, E. H., Kitakima, M., Takayama, H., and Takeya, K., Artoindonesianin L, A New Prenylated Flavone with Cytotoxic Activity from *A. rotunda*, *Fitoterapia*, **72**, 912-918, 2001.
25. Makmur, L., Syamsurizal, Tukiran, Achmad, S. A., Aimi, N., Hakim, E. H., Kitajima, M., and Takayama, H., Artoindonesianin C, A New Xanthone Derivative from *Artocarpus teysmanii*, *J. Nat. Prod.*, **63**, 243-244, 2000.
26. Makmur, L., Syamsurizal, Tukiran, Syamsu, Y., Achmad, S. A., Aimi, N., Hakim, E. H., Kitajima, M., Mujahidin, D., dan Takayama, H., Artonol B dan Sikloartobilosanton dari Tumbuhan *Artocarpus teysmanii* Miq., *Proc. ITB*, **31**, 63, 1999.

27. Hano, Y., Aida, S., Shiina, M., Nomura, T., Kawaii, T., Ohe, H., and Kagei, K., Artonins A and B, Two New Prenylflavones from The Root Bark of *Artocarpus heterophyllus* Lamk., *Heterocycles*, **29**, 1447-1453, 1989.
28. Aida, M., Shinomiya, K., Hano, Y., and Nomura, T., Artonins J, K and L, Three New Isoprenylated Flavones from The Root Bark of *Artocarpus heterophyllus* Lamk., *Heterocycles*, **36**, 575-582, 1993.
29. Aida, M., Shinomiya, K., Matsuzawa, K., Hano, Y., and Nomura, T., Artonins Q, R, S, T and U, Five New Isoprenylated Phenols from the Bark of *Artocarpus heterophyllus* Lamk., *Heterocycles*, **39**, 847-857, 1994.
30. Hano, Y., Inami, R., and Nomura, T., Components of the Bark of *Artocarpus rigida* Bl. 1. Structures of the Two New Isoprenylated Flavones, Artonins G and H, *Heterocycles*, **31**, 2173-2178, 1990.
31. Hano, Y., Inami, R., and Nomura, T., Components of the Bark of *Artocarpus rigida* Bl. 2. Structures of Four New Isoprenylated Flavone Derivatives Artonins M, N, O, and P, *Heterocycles*, **35**, 1993.
32. Wang, Y. -H., Hou, Ai. -J., Chen, L., Chen, D. -F., Sun, H. -D., Zhao, Q. -S., Bastow, K. F., Nakanish, Y., Wang, X. -H., and Lee, K. -H., New Isoprenylated Flavones, Artochamins A-E, and Cytotoxic Principles from *Artocarpus chama*, *J. Nat. Prod.*, **67**, 757-761, 2004.
33. Hano, H., Itoh, N., Hanaoka, A., Itoh, Y., and Nomura, T., Paratocarpins A-E, Five New Isoprenoid-Substituted Chalcones from *Paratocarpus venenosa* Zoll., *Heterocycles*, **41**, 191-198, 1995.
34. Hano, H., Itoh, N., Hanaoka, A., and Nomura, T., Paratocarpins F-L, Seven New Isoprenoid-Substituted Flavonoids from *Paratocarpus venenosa* Zoll., *Heterocycles*, **41**, 2313-2326, 1995.
35. Patil, A. D., Freyer, A. J., Killmer, L., Offen, P., Taylor, P. B., Votta, B. J., and Johnson, R. K., A New Dimeric Dihydrochalcone and a New Prenylated Flavone from the Bud Covers of *Artocarpus altilis*: Potent Inhibitors of

- Cathepsin K, *J. Nat. Prod.*, **65**, 624-627, 2002.
36. Jayasinghe, L., Balasooriya, B. A. I. S., Padmini, W. C., Hara, N., and Fujimoto, Y., Geranyl Chalcone Derivatives with Antifungal and Radical Scavenging Properties from the Leaves of *Artocarpus nobilis*, *Phytochemistry*, **65**, 1287-1290, 2004.
 37. Su, B. -N., Cuendet, M., Hawthorne, M. E., Kardono, L. B. S., Riswan, S., Fong, H. H. S., Mehta, R. G., Pezzuto, J. M., and Kinghorn, A. D., Constituents of the Bark and Twigs of *Artocarpus dadah* with Cyclooxygenase Inhibitory Activity, *J. Nat. Prod.*, **65**, 163-169, 2002.
 38. Soekarto, N. H., Achmad, S. A., Ghisalberti, E. L., Hakim, E. H., and Syah, Y. M., Artoindonesianins X and Y, Two Isoprenylated 2-Arylbenzofurans from *Artocarpus fretessi* (Moraceae), *Phytochemistry*, **64**, 831-834, 2003.
 39. Likhitwitayawuid, K., and Sritularak, B., A New Dimeric Stilbene with Tyrosinase Inhibitory Activity from *Artocarpus gomezianus*, *J. Nat. Prod.*, **64**, 1457-1459, 2001.
 40. Likhitwitayawuid, K., Sritularak, B., and De-Eknamkul, W., Tyrosinase Inhibitors from *Artocarpus gomezianus*, *Planta Med.*, **66**, 275-277, 2000.
 41. Shimizu, K., Kondo, R., and Sakai, K., A Stilbene Derivative from *Artocarpus incisus*, *Phytochemistry*, **45**, 1297-1298, 1997.
 42. Shimizu, K., Kondo, R., Sakai, K., Lee, S. -L, and Sato, H., The Inhibitory Components from *Artocarpus incisus* on Melanin Biosynthesis, *Planta Med.*, **64**, 408-412, 1998.
 43. Chung, M. -I., Ko, H. -H., Yen, M. -H, Lin, C. -N., Yang, S. -Z., Tsao, L. -T., and Wang, J. -P., Artocarpol A, a Novel Constituents with Potent Anti-inflammatory Effect, Isolated from *Artocarpus rigida*, *Helv. Chim. Acta*, **83**, 1200-1204, 2000.
 44. Ko, H. -H., Lin, C. -N., and Yang, S. -Z., New Constituents of *Artocarpus*

- rigida*, *Helv. Chim. Acta*, **83**, 3000-3005, 2000.
45. Ko, H. -H., Yang, S. -Z., and Lin, C. -N., Artocarpols F, A Phenolic Compound with a Novel Skeleton, Isolated from *Artocarpus rigida*, *Tet. Lett.*, **42**, 5269-5270, 2001.
 46. Hano, Y., Aida, M., and Nomura, T., Two New Natural Diels-Alder Type Adducts from The Root Bark of *Artocarpus heterophyllus*, *J. Nat. Prod.*, **53**, 391-395, 1990.
 47. Shinomiya, K., Aida, M., Hano, Y., and Nomura, T., A Diels-Alder Type Adduct from *Artocarpus heterophyllus*, *Phytochemistry*, **40**, 1317-1319, 1995.
 48. Heller, W., and Forkmann, G., Biosynthesis. In *The Flavonoids: Advances in Research since 1980*, Harborne, J. B. (ed.), Chapman and Hall, London, 399-425, 1988.
 49. Barron, D., and Ibrahim, R. K., Isoprenylated Flavonoids - A Survey, *Phytochemistry*, **43**, 921-982, 1996.
 50. Harborne, J. B., and Williams, C.A., Advances in Flavonoid Research Since 1992, *Phytochemistry*, **55**, 481-504, 2000.
 51. Bohm, A. B., Introduction to Flavonoids, Harwood Academic Pub., London, 243-284, 1998.
 52. Harborne, J. B., Mabry, T. J., and Mabry, H., The Flavonoids, Chapman and Hall, London, 127-213, 1975.
 53. Ares, J. J., Outt, P. E., Kakodkar, S. V., Buss, R. C., and Geiger, J. C., A Convenient Large-Scale Synthesis of 5-Methoxyflavone and Its Application to Analog Preparation, *J. Org. Chem.*, **58**, 7903-7905, 1993.
 54. Bennett, M., Burke, A. J., and O'Sullivan, W. I., Aspects of the Algar-Flynn-Oyamada (AFO) Reaction, *Tetrahedron*, **52**, 7163-7178, 1996.
 55. Wong, K. C., Lim, C. L., and Wong, L. L., Volatile Flavour Constituents of

- Chempedak (*Artocarpus polyphema* Pers.) Fruit and Jackfruit (*Artocarpus heterophyllus* Lam.) from Malaysia, *Flav. Fragr. J.*, **7**, 307-311, 1992.
56. Markham, K. R., Techniques of Flavonoid Identification, Academic Press, London, 41-45, 1982.
 57. Abegaz, B. M., Ngadjui, B. T., Dongo, E., and Tamboue, H., Prenylated Chalcones and Flavones From The Leaves of *Dorstenia kameruniana*, *Phytochemistry*, **49**, 1147-1150, 1998.
 58. Stevens, J. F., Ivancic, M., Hsu, V. L., and Deinzer, M. L., Prenylflavonoids From *Humulus lupulus*, *Phytochemistry*, **44**, 1575-1585, 1997.
 59. Pistelli, L., Spera, K., Flamini, G., Mele, S., and Morelli, I., Isoflavonoids and Chalcones from *Anthyllis hermanniae*, *Phytochemistry*, **42**, 1455-1458, 1996.
 60. Krishnaswamy, N. R., T. R. Seshadri's Contributions to the Chemistry of Natural Products, Some Illustrative Examples, *Resonance*, **9**, 26-38, 2004.
 61. Asada, Y., Li, W., and Yoshikawa, T., Isoprenylated Flavonoids From Hairy Root Cultures of *Glycyrrhiza glabra*, *Phytochemistry*, **47**, 389-392, 1998.
 62. Tsopmo, A., Tene, M., Kamnaing, P., Ngnokam, D., Ayafor, J. F., and Sterner, O., Geranylated Flavonoids From *Dorstenia poinsettifolia*, *Phytochemistry*, **48**, 345-348, 1998.
 63. Abegaz, B. M., Ngadjui, B. T., Dongo, E., Ngameni, B., Nindi, M. N., and Bezabih, M., Chalcones and other Constituents of *Dorstenia prorepens* and *Dorstenia zenkeri*, *Phytochemistry*, **59**, 877-883, 2002.
 64. Yenesew, A., Midiwo, J. O., Miessner, M., Heydenreich, M., and Peter, M. G., Two Prenylated Flavanones From The Stem Bark of *Erythrina burttii*, *Phytochemistry*, **48**, 1439-1443, 1998.
 65. AlSohly, H. N., Joshi, A. S., Nimrod, A. C., Walker, L. A., and Clark, A. M., Antifungal Chalcones From *Maclura tinctoria*, *Planta Med.*, **67**, 87-89, 2001.

66. Haraguchi, H., Inoue, J., Tamura, Y., and Mizutani, K., Antioxidative Components of *Psoralea corylifolia* (Leguminosae), *Phytotherapy Res.*, **16**, 539-544, 2002.
67. Akihisa, T., Tokuda, H., Ukiya, M., Iizuka, M., Schneider, S., Ogasawara, K., Mukainaka, T., Iwatsuki, K., Suzuki, T., and Nishino, H., Chalcones, Coumarins, and Flavanones from the Exudate of *Angelica keiskei* and Their Chemopreventive Effects, *Cancer Lett.*, **201**, 133-137, 2003.
68. Ngadjui, B. T., Abegaz, B. M., Dongo, E., Tamboue, H., and Fogue, K., Geranylated and Prenylated Flavonoids From The Twigs of *Dorstenia mannii*, *Phytochemistry*, **48**, 349-354, 1998.
69. Shieh, W. -L., and Lin, C. -N., A Quinonoid Pyranobenzoxanthone and Pyranodihydrobenzoxanthone from *Artocarpus communis*, *Phytochemistry*, **31**, 364-367, 1992.
70. Sultanbawa, M. U. S., and Surendrakumar, Two Pyranodihydrobenzoxanthenes From *Artocarpus nobilis*, *Phytochemistry*, **28**, 599-605, 1989.
71. Nomura, T., Hano, Y., and Aida, M., Isoprenoid-Substituted Flavonoids From *Artocarpus* Plants (Moraceae), *Heterocycles*, **47**, 1179-1205, 1998.
72. Nomura, T., The Chemistry and Biosynthesis of Isoprenylated Flavonoids From Moraceous Plants, *Pure Appl. Chem.*, **71**, 1115-1118, 1999.
73. Nomura, T., Fukai, T., Yamada, S., and Katayanagi, K., Studies on the Constituents of the Cultivated Mulberry Tree.I. Three New Prenylflavone From The Root Bark of *Morus alba* L., *Chem. Pharm. Bull.*, **26**, 1394-1402, 1978.
74. Yoshizawa, S., Suganuma, M., Fujiki, H., Nomura, T., and Sugimura, T., Morusin, Isolated From Root Bark of *Morus alba* L., Inhibits Tumor Promotion by Teleocidin, *Phytotherapy Res.*, **3**, 193-195, 1989.

75. Fujiki, H., and Suganuma, M., Tumor Promotion by Inhibitors of Protein Phosphatases 1 and 2A: The Okadaic Acid Class of Compounds, *Adv. Cancer Res.*, **61**, 143-193, 1993.
76. Komori, A., Yatsunami, J., Suganuma, M., Okabe, S., Abe, S., Sasaki, K., and Fujiki, H., Tumor Necrosis Factor Acts as A Tumor Promoter in BALB/3T3 Cell Transformation, *Cancer Res.*, **53**, 1982-1985, 1993.
77. Liou, S. -S., Shieh, W. -L., Chen, T. -H., Won, S. -J., and Lin, C. -N., γ -Pyrone Compounds As Potential Anti Cancer Drugs, *J. Pharm. Pharmacol.*, **45**, 791-794, 1993.
78. Lin, C. -N., Shieh, W. -L., Ko, F. -N., and Teng, C. -M., Antiplatelet Activity of Some Prenylflavonoids, *Biochem. Pharmacol.*, **45**, 509-512, 1993.
79. Lin, C. -N., Lu, C. -M., Lin, H. -C., Fang, S. -C., Shieh, B. -J., Hsu, M. -F., Wang, J. -P., Ko, F. -N., and Teng, C. -M., Novel Antiplatelet Constituents from Formosan Moraceous Plants, *J. Nat. Prod.*, **59**, 834-838, 1996.
80. Khan, M. R., Omoloso, A. D., and Kihara, M., Antibacterial Activity of *Artocarpus heterophyllus*, *Fitoterapia*, **74**, 501-505, 2003.
81. Sato, M., Fujiwara, S., Tsuchiya, H., Fujii, T., Iimuna, M., Tosa, H., and Ohkawa, Y., Flavones with Antibacterial Activity Against Cariogenic Bacteria, *J. Ethnopharmacol.*, **54**, 171-176, 1996.
82. Mackeen, M.M., Ali, A. M., El-Sharkawy, S. H., Manap, M. Y., Salleh, K. M., Lajis, N. H., Kawazu, K., Antimicrobial and Cytotoxic Properties of Some Malaysian Traditional Vegetables, *Int. J. Pharmacog.*, **35**, 174-178, 1997.
83. Arias, M. E., Gomez, J. D., Cudmani, N. M., Vattuone, M. A., and Isla, M. I., Antibacterial Activity of Ethanolic and Aqueous Extracts of *Acacia aroma* Gill. Ex Hook et Arn, *Life Sciences*, **75**, 191-202, 2004.

84. Nester, E. W., Anderson, D. G., Robert, J. C. E., Pearsall, N. N., Nester, M. T., and Hurley, D., *Microbiology: A Human Perspective*, McGraw Hill, New York, 4th Ed., 2004.
85. Pokorny, J., Yanishlieva, N., and Gordon, M., *Antioxidants in Food: Practical Applications*, CRC Press, Woodhead Publishing Ltd., England, 7-368, 2001.
86. Ferrari, R., Ceconi, C., Curello, S., Cargnoni, A., Alfieri, O., Pardini, A., Marzollo, P., and Visioli, O., Oxygen Free Radicals and Myocardial Damage: Protective Role of Thiol-containing Agents, *Amer. J. Med.*, **91**, 95, 1991.
87. Bondet, V., Williams, W. -B., and Berset, C., Kinetics and Mechanisms of Antioxidant Activity using the DPPH[•] Free Radical Method, *Lebensm.-Wiss. U.-Technol.*, **30**, 609-615, 1997.
88. Choi, C. W., Kim, S. C., Hwang, S. S., Choi, B. K., Ahn, H. J., Lee, M. Y., Park, S. H., and Kim, S. K., Antioxidant Activity and Free Radical Scavenging Capacity Between Korean Medicinal Plants and Flavonoids by Assay-Guided Comparison, *Plant Sci.*, **163**, 1161-1168, 2002.
89. Ahn, C. -B., Jeon, Y. -J., Kang, D. -S., Shin, T. -S., and Jung, B. -M., Free Radical Scavenging Activity of Enzymatic Extracts From A Brown Seaweed *Scytosiphon lomentaria* by Electron Spin Resonance Spectrometry, *Food Research International*, **37**, 253-258, 2004.
90. Larson, R. A., The Antioxidant of Higher Plants, *Phytochemistry*, **4**, 969-978, 1988.
91. Kikuzaki, H., and Nakatani, N., Antioxidant Effects of Some Ginger Constituents, *J. Food Science*, **58**, 1407-1410, 1993.
92. Sanchez, C. -M., Larrauri, J. A., and Saura, F. -C., Free Radical Scavenging Capacity and Inhibition of Lipid Oxidation of Wines, Grape Juices and Related Polyphenolic Constituents, *Food Research International*, **32**, 407-412, 1999.

93. Tagashira, M., and Ohtake, Y., A New Antioxidative 1,3-benzodioxole from *Melissa officinalis*, *Planta Med.*, **64**, 555-558, 1998.
94. Ohtani, I. I., Gotoh, N., Tanaka, J., Higa, T., Gyamfi, A., and Aniya, Y., Thonningianins A and B, New Antioxidants from the African Medicinal Herb *Thonningia sanguinea*, *J. Nat. Prod.*, **63**, 676-679, 2000.
95. Valone, F. H., Coles, E., Reinhold, V. R., and Goetzl, E. J., Specific Binding of Phospholipid Platelet-activating Factor by Human Platelets, *J. Immunol.*, **129**, 1637-1641, 1982.
96. Min, J. -H., Jain, M. K., Wilder, C., Paul, L., Rafael, A. -C., Aspleaf, D. C., and Gelb, M. H., Membrane-Bound Plasma Platelet Activating Factor Acetylhydrolase Acts on Substrate in the Aqueous Phase, *Biochemistry*, **38**, 12935-12942, 1999.
97. Kim, K. A., Moon, T. C., Lee, S. W., Chung, K. C., Han, B. H., and Chang, H. W., Pinusolide from the Leaves of *Biota orientalis* as Potent Platelet Activating Factor Antagonist, *Planta Med.*, **65**, 39-42, 1999.
98. Beumont, G. H., and Egido, J., PAF, A Potent Proinflammatory Mediator, Looking For Its Role In The Pathogenesis of The Joint Damage, *Annals of the Rheumatic Diseases*, **56**, 211-213, 1997.
99. Maclennan, K. M., Smith, P. F., and Darlington, C. L., Platelet-Activating Factor In The CNS, *Progress in Neurobiology*, **50**, 585-596, 1996.
100. Braquet, P., Tougui, L., Shen, T. Y., Vargaftig, B. B., Perspectives in Platelet Activating Factor Research, *Pharmacol. Rev.*, **39**, 97-145, 1987.
101. Jantan, I., Juriyati, J., and Warif, N. A., Inhibitory Effects of Xanthenes on Platelet Activating Factor Receptor Binding *In Vitro*, *J. Ethnopharm.*, **75**, 287-290, 2001.
102. Dong, H., and Chen, S. -Xing, A New Antiplatelet Diarylheptanoid from *Alpinia blepharocalyx*, *J. Nat. Prod.*, **61**, 142-144, 1998.

103. Pulcinelli, F. M., Pignatelli, P., Celestini, A., Riondino, S., Gazzaniga, P. P., and Violi, F., Inhibition of Platelet Aggregation by Aspirin Progressively Decreases in Long-Term Treated Patients, *J. Am. Cardiology*, **43**, 979-984, 2004.
104. Suzuki, Y., Kondo, K., Ikeda, Y., and Umemura, K., Antithrombotic Effect of Geniposide and Genipin in The Mouse Thrombosis Model, *Planta Med.*, **67**, 807-810, 2001.
105. Chen, J.-J., Chang, Y.-L., Teng, C.-M., Su, C.-C., and Chen, I.-S., Quinoline Alkaloids and Anti-Platelet Aggregation Constituents From The Leaves of *Melicope semecarpifolia*, *Planta Med.*, **68**, 790-793, 2002.
106. Mosmann, T., Rapid Colorimetric Assay for Cellular Growth and Survival: Application to Proliferation and Cytotoxicity Assays, *J. Immunological*, **65**, 55-63, 1983.
107. Doyle, A. and Griffiths, J. B., Cell and Tissue Culture for Medical Research, John Wiley and Sons, LTD., Singapore, 49-409, 2000.
108. Huang, C., Zhang, Z., and Li, Y., Total Synthesis of (*R,S*)-Sophoraflavanone C, *J. Nat. Prod.*, **61**, 1283-1285, 1998.
109. Daskiewicz, J.-B., Bayet, C., and Barron, D., Rearrangement of 5-*O*-prenyl Flavones: A Regioselective Access to 6-*C*-(1,1-dimethylallyl)- and 8-*C*-(3,3-dimethylallyl)flavones, *Tet. Lett.*, **42**, 7241-7244, 2001.
110. Barron, D., and Mariotte, A. M., Synthesis of 8-*C*-(1,1-dimethylallyl)flavones and 3-methyl flavonols, *Nat. Prod. Lett.*, **4**, 21-28, 1994.
111. Stevens, J. F., Taylor, A. W., Nickerson, G. B., Ivancic, M., Henning, J., Haunold, A., and Deinzer, M. L., Prenylflavonoid Variation in *Humulus lupulus*: Distribution and Taxonomic Significance of Xanthogalenol and 4-*O*-Methylxanthohumol, *Phytochemistry*, **53**, 759-775, 2000.

112. Subrahmanyam, K., Madhusudhana, R. J., Vemuri, V. S. S., Sivaram, B. S., Roy, C. P., Jagannadha, R. K. V. and Merlini, L, New Chalcones from Leaves of *Flemingia stricta* Roxb. (Leguminosae), *Indian J. Chem.* **21B**, 895-897, 1982.
113. Macias, F. A., Velasco, R. F., Alvarez, J. A., Castello, D., and Galindo, C. G., Synthesis of Melampolides and *Cis,cis*-germacranolides as Natural Herbicide Models, *Tetrahedron*, **60**, 8477-8488, 2004.