

Complete genome sequence of *Methylomonas* sp. UP202 isolated from an urban waterway sediment

Beng-Soon Teh,¹ Yiik-Siang Hii,² Jamie Hinks,² Mohd Firdaus Abdul-Wahab,³ Sanjay Swarup^{1,4,5}

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT We report the complete genome sequence of *Methylomonas* sp. UP202 isolated from an urban waterway sediment in Singapore. The genome contains genes involved in methane, methanol, formaldehyde, and formate oxidation. It also contains genes utilizing various nitrogen sources such as nitrogen, nitrate, nitrite, urea, and ammonium.

KEYWORDS *Methylomonas*, methanotroph, whole genome sequencing, canal sediment

Methanotrophs offer various industrial applications (1–5). Here, we sequenced the genome of *Methylomonas* sp. UP202 to uncover its metabolic potential. An aerobic methanotroph *Methylomonas* sp. UP202 was isolated from the canal sediment at the Ulu Pandan Waterway in Singapore (1.3188° N; 103.7708° E). Sediment was added to ammonium mineral salt (AMS) (6) broth in a serum bottle supplemented with methane and air (20%:80%, vol/vol) and incubated at 30°C for a week. After several subcultures, colonies were selected on AMS agar with methane in Oxoid AnaeroJar. The purity of isolates was confirmed through phase-contrast microscopy and streaking on 10% TSA, with no growth indicating purity (7). Genomic DNA was extracted from 2 mL pure cultures grown in AMS broth using the Wizard Genomic DNA Purification Kit (Promega) following the manufacturer's protocol for Gram-negative bacteria.

Library preparation for Illumina sequencing was performed according to Illumina's TruSeq Nano DNA Sample Preparation protocol. The samples were sheared on a Covaris E220 to ~550 bp, following the manufacturer's recommendation, and uniquely tagged with Illumina's TruSeq HT DNA dual barcodes to enable sample pooling for sequencing. The libraries were sequenced on the Illumina MiSeq platform at a read length of 300 bp paired-end (Illumina, Inc.). A total of 4,354,004 paired-end reads were obtained for the MiSeq data. Oxford Nanopore (ONT) libraries were prepared following the manufacturer's instructions for the SQK-LSK110 Genomic DNA by ligation library prep kit (Oxford Nanopore Technologies), together with the NEBNext Companion Module for ONT Ligation Sequencing (New England Biolabs). The library was loaded on a Flongle R9.4.1 flowcell using the Flongle adapter attached to a GridION Mk1 device and sequenced for 24 h on MinKNOW (v.22.08.9). The raw ONT data were basecalled using Guppy (v.6.2.1) and resulted in 134,811 ONT reads with an N_{50} value of 7,939 bp. Adapter sequences of short reads were removed using Cutadapt (v.3.5) (8), and low-quality reads were removed using BBDuk function in BBTools (v.37.99) (<https://sourceforge.net/projects/bbmap/>). The adapters of ONT long reads were trimmed using Porechop (v.0.2.3) (9), and the reads were quality filtered and trimmed using Filtlong (v.0.2.0) (<https://github.com/rrwick/Filtlong>). Short- and long-read data were hybrid assembled using Unicycler (v.0.4.8) (10). Platon (v.1.6) (11) was used for plasmid detection in the assembled genome. Average nucleotide identity (ANI) was determined using Pyani (v.0.2.11) (12). The assembled

Editor Julia A. Maresca, University of Delaware College of Engineering, Newark, Delaware, USA

Address correspondence to Beng-Soon Teh, bengsoon@nus.edu.sg, Yiik-Siang Hii, yiiksiang.hii@ntu.edu.sg, or Sanjay Swarup, sanjay@nus.edu.sg.

Jamie Hinks Deceased 19 March 2023

The authors declare no conflict of interest.

See the funding table on p. 2.

Received 20 July 2023

Accepted 14 October 2023

Published 20 November 2023

Copyright © 2023 Teh et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

genome was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (v.6.5) (13). The quality of hybrid assembly was evaluated using QCAST (v.5.2.0) (14).

The assembled reads resulted in two circular contigs, corresponding to a chromosome of 5,526,923 bp (GC content, 55.84%) and a plasmid of 148,958 bp (GC content, 52.27%). The genome of *Methylomonas* sp. UP202 contains 4,924 protein-coding genes, 9 rRNA operons (5S, 16S, and 23S), and 47 tRNAs. The ANI value between *Methylomonas* sp. UP202 and *Methylomonas* sp. LWB was ~97%. Enzymes such as methane monooxygenases (pMMO and sMMO), methanol dehydrogenases, and formate dehydrogenases were found in the genome. Gene clusters involved in nitrogen fixation (*nif*), urease, urea transporters, ammonium, nitrate, and nitrite reductases were present.

ACKNOWLEDGMENTS

We would like to acknowledge PUB, Singapore's National Water Agency, for providing permit for field sampling at the Ulu Pandan Waterway. We would like to thank the NUS Metabolite Biology Lab members consisting of Andrew Elohim Laloo, Ooi Qi En, and Koh Yi Zi for their assistance in the sediment collection and former member Hitesh Tikariha for his invaluable input on the genome assembly pipeline. We thankfully acknowledge that the computational work involved in this work is fully supported by NUS IT's Research Computing group and the Sequencing Core Facility at SCELSE NTU.

This study was supported by the A*STAR Singapore Food Story (SFS) R&D Programme 1st Alternative Protein Seed Challenge Grant (Grant no. W20W2D0018).

This paper is dedicated to Dr. Jamie Hinks, without whose direction it would not have been possible.

AUTHOR AFFILIATIONS

¹NUS Environmental Research Institute, National University of Singapore, Singapore, Singapore

²Singapore Centre for Environmental Life Sciences Engineering, Nanyang Technological University, Singapore, Singapore

³Department of Biosciences, Faculty of Science, Universiti Teknologi Malaysia, Johor Bahru, Johor, Malaysia

⁴Singapore Centre for Environmental Life Sciences Engineering, National University of Singapore, Singapore, Singapore

⁵Department of Biological Sciences, National University of Singapore, Singapore, Singapore

AUTHOR ORCIDs

Beng-Soon Teh  <http://orcid.org/0000-0002-2846-1197>

Yiik-Siang Hii  <http://orcid.org/0000-0003-0972-2993>

Jamie Hinks  <http://orcid.org/0000-0002-9254-0041>

Sanjay Swarup  <http://orcid.org/0000-0001-6391-0624>

FUNDING

Funder	Grant(s)	Author(s)
Agency for Science, Technology and Research (A*STAR)	W20W2D0018	Jamie Hinks Sanjay Swarup Mohd Firdaus Abdul-Wahab

AUTHOR CONTRIBUTIONS

Beng-Soon Teh, Investigation, Methodology, Writing – original draft, Writing – review and editing | Yiik-Siang Hii, Investigation, Methodology, Writing – review and editing

| Jamie Hinks, Funding acquisition, Project administration, Supervision | Mohd Firdaus Abdul-Wahab, Funding acquisition, Investigation, Project administration, Supervision, Writing – review and editing | Sanjay Swarup, Funding acquisition, Project administration, Supervision, Writing – review and editing

DATA AVAILABILITY

The complete genome sequence of *Methylomonas* sp. UP202 has been deposited in GenBank under the accession numbers [CP123897](https://doi.org/10.1093/ncbi/CP123897) and [CP123898](https://doi.org/10.1093/ncbi/CP123898). Raw reads were submitted to the NCBI SRA under the accession numbers [SRR24223047](https://doi.org/10.1093/ncbi/SRR24223047) and [SRR24223048](https://doi.org/10.1093/ncbi/SRR24223048). The BioProject accession number is [PRJNA954998](https://doi.org/10.1093/ncbi/PRJNA954998) and the BioSample accession number is [SAMN34162754](https://doi.org/10.1093/ncbi/SAMN34162754).

REFERENCES

1. Cantera S, Sánchez-Andrea I, Lebrero R, García-Encina PA, Stams AJM, Muñoz R. 2018. Multi-production of high added market value metabolites from diluted methane emissions via methanotrophic extremophiles. *Bioresour Technol* 267:401–407. <https://doi.org/10.1016/j.biortech.2018.07.057>
2. Rasouli Z, Valverde-Pérez B, D'Este M, De Francisci D, Angelidaki I. 2018. Nutrient recovery from industrial wastewater as single cell protein by a co-culture of green microalgae and methanotrophs. *Biochemical Engineering Journal* 134:129–135. <https://doi.org/10.1016/j.bej.2018.03.010>
3. Rostkowski KH, Pfluger AR, Criddle CS. 2013. Stoichiometry and kinetics of the PHB-producing type II methanotrophs *Methylosinus trichosporium* OB3b and *Methylocystis parvus* OBBP. *Bioresour Technol* 132:71–77. <https://doi.org/10.1016/j.biortech.2012.12.129>
4. Tsapekos P, Zhu X, Pallis E, Angelidaki I. 2020. Proteinaceous methanotrophs for feed additive using biowaste as carbon and nutrients source. *Bioresour Technol* 313:123646. <https://doi.org/10.1016/j.biortech.2020.123646>
5. Xin J-Y, Zhang Y-X, Zhang S, Xia C-G, Li S-B. 2007. Methanol production from CO₂ by resting cells of the methanotrophic bacterium *Methylosinus trichosporium* IMV 3011. *J Basic Microbiol* 47:426–435. <https://doi.org/10.1002/jobm.200710313>
6. Nyerges G, Han S-K, Stein LY. 2010. Effects of ammonium and nitrite on growth and competitive fitness of cultivated methanotrophic bacteria. *Appl Environ Microbiol* 76:5648–5651. <https://doi.org/10.1128/AEM.00747-10>
7. Rahalkar MC, Khatri K, Pandit P, Bahulikar RA, Mohite JA. 2021. Cultivation of important methanotrophs from Indian rice fields. *Front Microbiol* 12:669244. <https://doi.org/10.3389/fmicb.2021.669244>
8. Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet j* 17:10. <https://doi.org/10.14806/ej.17.1.200>
9. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. *Microb Genom* 3:e000132. <https://doi.org/10.1099/mgen.0.000132>
10. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
11. Schwengers O, Barth P, Falgenhauer L, Hain T, Chakraborty T, Goesmann A. 2020. Platon: identification and characterization of bacterial plasmid contigs in short-read draft assemblies exploiting protein sequence-based replicon distribution scores. *Microb Genom* 6:mgen000398. <https://doi.org/10.1099/mgen.0.000398>
12. Pritchard L, Glover RH, Humphris S, Elphinstone JG, Toth IK. 2016. Genomics and taxonomy in diagnostics for food security: soft-rotting enterobacterial plant pathogens. *Anal Methods* 8:12–24. <https://doi.org/10.1039/C5AY02550H>
13. Tatusova T, DiCuccio M, Badretdin A, Chetvermin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
14. Mikheenko A, Pribelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUASt-LG. *Bioinformatics* 34:i142–i150. <https://doi.org/10.1093/bioinformatics/bty266>