OCTADECYLSILANE AND MULTIWALL CARBON NANOTUBES BASED CELLULOSE TRIACETATE MIXED MATRIX MEMBRANE FOR NEW MICROEXTRACTION APPROACHES

THIPASHINI A/P GANESAN

A thesis submitted in fulfilment of the requirements for the award of the degree of Master of Philosophy

> Faculty of Science Universiti Teknologi Malaysia

> > JULY 2020

DEDICATION

This thesis is dedicated specifically to my mom for her endless support and encouragement to complete my master.

ACKNOWLEDGEMENT

First and foremost, I would like to thank God Almighty for giving me the strength, knowledge, and opportunity to undertake this journey and complete it satisfactorily.

I would like to express my heart-felt thanks and profound gratitude to my main supervisor, Assoc. Prof. Dr. See Hong Heng for his continuous guidance, inspiration, support, great advice and for sparing his time throughout my research as well as in the completion of this thesis. And special thanks, to my co-supervisor Assoc. Prof. Dr. Lim Hong Ngee.

Furthermore, I have a great pleasure in acknowledging my gratitude to my colleagues for the tremendous help and contribution who have aided at various occasions. I would like to extend my overwhelming gratitude to entire University Teknologi Malaysia staffs for their support and assistance.

I would like to convey warmest appreciation and special thanks to my mom for being the biggest strength, supporting me spiritually and continuous encouragement throughout this research. Finally, to my friends, relatives, and everyone known and unknown that contributed directly or indirectly to the successful completion of my study.

ABSTRACT

Widespread use of pharmaceuticals and herbicides leads to accumulation of their residues in surface water with detrimental effects and they have been recognized as emerging environmental pollutants. Therefore, it is important to develop a suitable sample pre-treatment method for the determination of these compounds in water. In this study, the fabrication of a mixed matrix membrane (MMM) using carbonaceous material immobilized in cellulose triacetate polymer matrix for microextraction is reported. The applicability and reliability of utilizing carbonaceous-based MMM in two different miniaturised microextraction designs, namely dynamic MMM tip extraction and automated flow-through MMM microextraction, are comprehensively demonstrated. To evaluate the performance of dynamic MMM tip extraction, the developed approach was applied for determination of five selected nonsteroidal anti-inflammatory drugs (NSAIDs) namely indoprofen, sulindac, naproxen, diclofenac, and ibuprofen in sewage water samples prior to ultraperformance liquid chromatography-tandem mass spectrometry (UPLC-MS/MS) analysis. The established method showed good linear responses in the concentration range of 0.25–500 pg/mL with correlation coefficients (r) from 0.9988 to 0.9992. The limits of detections (LODs) were 0.08–0.40 pg/mL. The relative recoveries of NSAIDs from spiked water samples were in the range of 92-99% and exhibited excellent precision (relative standard deviations, RSDs \leq 4.9%). The proposed analytical methodology allowed pre-concentration factors up to 250. Automated flow-through MMM microextraction approach was developed for the analysis of six chlorophenoxy acid herbicides, namely 2-(4-chlorophenoxy)acetic acid, 2-(3,4dichlorophenoxy)acetic acid, 2-(2,4-dichlorophenoxy)acetic acid, 2-(4-chloro-2methylphenoxy)acetic acid, 2-(2,4,5-trichlorophenoxy)acetic acid, and 4-(2,4dichlorophenoxy)butanoic acid in the spiked sewage water sample prior to high performance liquid chromatography-ultraviolet detection (HPLC-UV) analysis. Under optimum conditions, the linearity of this method ranged from 50 - 1000 ng/mL with correlation coefficients (r) ≥ 0.9939 , while LODs ranged from 15 - 20 ng/mL. The recoveries of the compounds in spiked sewage water samples were from 95% to 99% with RSDs \leq 7.5% and enrichment factors of 19 to 55. The developed analytical methodology has the advantage of requiring less adsorbent (only 62.5 µg) and less organic reagent (60 µL of desorption solvent).

ABSTRAK

Penggunaan farmaseutikal dan herbisida secara meluas membawa kepada pengumpulan residu di dalam air yang membawa kesan memudaratkan dan ia dikenali sebagai bahan pencemar alam sekitar muncul. Oleh itu, pembangunan kaedah pra-rawatan sampel yang sesuai adalah penting untuk penentuan sebatian ini di dalam air. Dalam kajian ini, fabrikasi membran matriks bercampur (MMM) menggunakan bahan berkarbon yang tidak bergerak di dalam matrik polimer selulosa pengekstrakan dilaporkan. triasetat untuk mikro Kebolehlaksanaan dan kebolehpercayaan penggunaan MMM berasaskan karbon dalam dua reka bentuk pengekstrakan miniatur yang berbeza iaitu pengekstrakan muncung MMM dinamik dan pengekstrakan mikro MMM mengalir-lalu automatik diperlihatkan secara komprehensif. Untuk menilai prestasi pengekstrakan muncung MMM dinamik, pendekatan yang dibangunkan telah digunakan bagi penentuan lima jenis ubat antiradang bukan steroid (NSAIDs) terpilih iaitu indoprofen, sulindac, naproxen, diclofenac, dan ibuprofen di dalam sampel air kumbahan sebelum analisis kromatografi cecair berprestasi ultra-spektromertri jisim seiring (UPLC-MS/MS). Kaedah yang dibangunkan telah menunjukkan gerak balas linear yang baik dalam julat kepekatan 0.25-500 pg/mL dengan pekali korelasi (r) dari 0.9988 hingga 0.9992. Had pengesanan (LOD) adalah 0.08 -0.40 pg/mL. Perolehan semula relatif NSAIDs daripada sampel air terpaku adalah dalam julat 92-99% dan menunjukkan kepersisan yang cemerlang (sisihan piawaian relatif, $RSDs \le 4.9\%$). Kaedah analisis yang dicadangkan membolehkan faktor pra-pemekatan sehingga 250. Pendekatan pengekstrakan mikro MMM aliran automatik telah dibangunkan untuk menganalisis enam racun rumpai asid klorofenoksi, iaitu asid 2-4(-klorofenoksi)asetik, asid 2-(3,4diklorofenoksi)asetik, asid 2-(2,4-diklorofenoksi)asetik, asid 2-(4-kloro-2metilfenoksi)asetik, asid 2-(2,4,5-triklorofenoksi)asetik, dan asid 4-(2,4diklorofenoksi)butanoik di dalam sampel air kumbahan terpaku sebelum analisis kromatografi cecair berprestasi tinggi dengan pengesanan ultra ungu (HPLC-UV). Dalam keadaan optimum, kelinearan kaedah ini berjulat dari 50 - 1000 ng/mL dengan pekali kolerasi (r) \ge 0.9939, sementara LODs berjulat dari 15 - 20 ng/mL. Perolehan semula sebatian daripada sampel air kumbahan terpaku ialah dari 95% hingga 99% dengan RSDs \leq 7.5% dan faktor pengkayaan sebanyak 19 hingga 55. Kaedah analisis yang dibangunkan mempunyai kelebihan iaitu memerlukan kurang bahan penjerap (62.5 µg sahaja) dan kurang reagen organik (60 µL pelarut penyaherapan).

TABLE OF CONTENTS

TITLE

	DECLARATION						
	DEDICATION						
	ACKNOWLEDGEMENT						
	ABSTRACT						
	ABST	RAK	vii				
1	TABL	E OF CONTENTS	viii				
	LIST	OF TABLES	xii				
	LIST	OF FIGURES	xiii				
	LIST	OF ABBREVIATIONS	xvii				
	LIST	OF SYMBOLS	XX				
CHAPTER	1	INTRODUCTION	1				
	1.1	Background Study	1				
	1.2	Problem Statement					
	1.3	Objectives of Study					
	1.4	Scope of Study					
	1.5	Significance of Research					
	1.6	Flowchart/Scheme of the Research					
CHAPTER	2	LITERATURE REVIEW	7				
	2.1	Introduction	7				
	2.2	Sample Preparation in Chemical Analysis	7				
	2.3	Conventional Sample Preparation Techniques in Chemical Analysis	8				
		2.3.1 Liquid-Liquid Extraction	8				
		2.3.2 Solid Phase Extraction	9				
:	2.4	Miniaturisation of Sample Preparation Techniques in Chemical Analysis	10				
		2.4.1 Solid Phase Microextraction	10				

	2.4.2 Liquid Phase Microextraction	12		
2.5	Membrane-Based Microextractions	13		
	2.5.1 Hollow Fibre Liquid Phase Microextraction	13		
	2.5.2 Micro Solid Phase Extraction	14		
	2.5.3 Electromembrane microextraction	15		
	2.5.4 Solid Phase Membrane Tip Extraction	16		
	2.5.5 Thin film microextraction (TFME)	18		
2.6	Mixed Matrix Membrane	18		
	2.6.1 Fabrication of MMM	20		
	2.6.2 Factors influencing MMM fabrication	21		
	2.6.3 Application of the MMM in Gaseous and Liquid Separation	23		
	2.6.4 Previous Studies on MMM Microextraction	32		
2.7	Nanomaterials			
	2.7.1 Carbon nanotubes	33		
	2.7.2 Polymeric Octadecylsilane Particles	34		
2.8	Nonsteroidal anti-inflammatory drugs	35		
2.9	Chlorophenoxy acid herbicides	36		
CHAPTER 3	MIXED MATRIX MEMBRANE MICROEXTRACTION COUPLED WITH UPLC-MS/MS FOR THE MONITORING OF NON-STEROIDAL ANTI-INFLAMMATORY DRUGS IN WATER SAMPLES	37		
3.1	Introduction	37		
3.2	Experimental Section			
	3.2.1 Chemicals/Reagents	37		
	3.2.2 Standard and sample preparation	38		
	3.2.3 Mixed matrix membrane tip preparation	39		
	3.2.4 Extraction Procedure	40		
	3.2.5 HPLC-UV analysis and operating conditions	42		
	3.2.6 UPLC–MS/MS analysis and operating conditions	43		
	3.2.7 Method Validation	43		
3.3	Results and Discussion	44		

3.3 **Results and Discussion**

	3.3.1	Optimization of extraction condition				
		3.3.1.1	Effect of sample pH	44		
		3.3.1.2	Effect of salting-out	45		
		3.3.1.3	Effect of dynamic extraction cycle	46		
		3.3.1.4	Effect of type of solvent	47		
		3.3.1.5	Effect of desorption time	48		
	3.3.2	Analytic method	Analytical performance of dynamic MMMTE method			
		3.3.2.1	Method Validation	49		
		3.3.2.2	Analysis of real samples	51		
		3.3.2.3	Comparison of dynamic MMMTE with other reported method	52		
CHAPTER 4	MICI CHR DETI	ROEXTR OMATO ERMINA	D MIXED MATRIX MEMBRANE ACTION PRIOR TO LIQUID GRAPHY FOR THE TION OF CHLOROPHENOXY CIDES IN WATER SAMPLES	55		
4.1	Introd	uction	ction			
4.2	Experimental Section			55		
	4.2.1	Chemica	55			
	4.2.2	Standard	56			
	4.2.3	Preparat flow cel	ion of mixed matrix membrane in a l	58		
	4.2.4	System 1	Design and Operation	58		
	4.2.5	HPLC-U	JV analysis and operating conditions	60		
4.3	Resul	ts and Discussion				
	4.3.1	Optimis	ation of extraction conditions	61		
		4.3.1.1	Effect of adsorbent loading	61		
		4.3.1.2	Effect of sample pH	62		
		4.3.1.3	Effect of ionic strength	63		
		4.3.1.4	Effect of sample flow rate	63		
		4.3.1.5	Effects of sample volume	64		
		4.3.1.6	Effects of type of solvent	65		

		4.3.1.7	Effects of desorption method	66
	4.3.2	Analytic	al performance	67
		4.3.2.1	Method Validation	67
		4.3.2.2	Analysis of real samples	68
		4.3.2.3	microextraction with other reported	60
			method	69
CHAPTER 5	CON	CLUSIO	N AND RECOMMENDATION	71
5.1	Concl	usion		71
5.2	Recor	nmendatio	on	72
REFERENCES				73
LIST OF PUBLICATIONS			91	

LIST OF TABLES

TABLE NO.	TITLE	PAGE
Table 2.1	Applications of MMM in gas phase	24
Table 2.2	Applications of MMM in liquid phase	27
Table 3.1	Name, chemical structure, chemical formula, molecular weight, pKa, and log P for NSAIDs from Chemicalize [153].	39
Table 3.2	Finalized MRM transition by UPLC-MS/MS ESI polarity switch	43
Table 3.3	Linear range, regression data (r), limits of detection (LOD), limits of quantification (LOQ), and enrichment factors (EF) of the extraction of NSAIDs in spiked effluent water samples in combination with UPLC-MS/MS	50
Table 3.4	Comparison of the current work with other previous methods for analysis of NSAIDs	53
Table 4.1	Name, molecular structure, molecular formula, molecular weight, p Ka value and log P of selected CPAHs from Chemicalize [153].	57
Table 4.2	Outline of Operation Sequence	60
Table 4.3	Validation data of developed automated MMM microextraction method for the extraction and determination of CPAHs in water samples in combination with HPLC-UV	68
Table 4.4	Comparison of the current work with other previous methods for analysis of CPAHs	70

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE			
Figure 1.1	Flowchart of research work for MMM microextraction	6			
Figure 2.1	Basic procedures in SPE [51]				
Figure 2.2	Mode of fibre SPME operation; (a) direct extraction, (b) headspace SPME and (c) membrane protected SPME [4]	11			
Figure 2.3	Schematic of static LPME [66]	13			
Figure 2.4	Microextraction modes used in HF-LPME: (A) two- phase system; and (B) three-phase system [68]	14			
Figure 2.5	Schematic diagram of µ-SPE [75]	15			
Figure 2.6	Schematic diagram of EME [80]	16			
Figure 2.7	Schematic diagram of SPMTE [6].	17			
Figure 2.8	Schematic diagram of an ideal MMM [89]	19			
Figure 2.9	Different methods for mixed matrix dope preparation [89]	21			
Figure 2.10	Schematic illustration of the C18-MMM microextraction setup [31]	32			
Figure 2.11	Different types of CNT in grey are models of either single-walled or multi-walled CNTs and in green are the various forms of SWCNTs [134]	34			
Figure 3.1	Schematic diagram of the dynamic MMMTE experimental setup	41			
Figure 3.2	Expanded view of dynamic MMMTE (a) The sample solution (SS) is continuously withdrawn into the membrane tip for analyte adsorption (b) The withdrawn SS is released from the tip back into sample vial with analytes retained in membrane phase (MP) (c) The desorption solvent (DS) is continuously flushed in the membrane tip for ten times, and the analytes is transferred rapidly from MP to DS	42			
Figure 3.3	Effect of sample pH on the peak area of NSAIDs using dynamic MMMTE. Initial extraction parameters: 500 ng/mL of spiked solution; adsorbent loading, 25%; no addition of salt; sample volume, 10 mL; dynamic				

extraction cycle, 2; desorption solvent, 40 μ L methanol; desorption time, 5 min. HPLC-UV conditions: Purospher® STAR RP-18 (5 μ m, 150 mm length \times 4.6 mm id). isocratic mode using a mobile phase of acetonitrile – 25 mM phosphate buffer (pH 3.5) (55:45, v/v), flow rate 1 mL/min, wavelength 210 nm

- Figure 3.4 Effect of salting-out on the peak area of NSAIDs using dynamic MMMTE. Initial extraction parameters: 500 ng/mL of spiked solution; adsorbent loading, 25%; sample pH 3.5; sample volume, 10 mL; dynamic extraction cycle, 2; desorption solvent, 40 µL methanol; desorption time, 5 min. HPLC-UV conditions: Purospher® STAR RP-18 (5 µm, 150 mm length \times 4.6 mm id). isocratic mode using a mobile phase of acetonitrile – 25 mM phosphate buffer (pH (55:45, v/v), flow rate 1 mL/min, wavelength 210 nm
- Figure 3.5 Effect of dynamic extraction cycle on the peak area of NSAIDs using dynamic MMMTE. Initial extraction parameters: 500 ng/mL of spiked solution; adsorbent loading, 25%; sample pH 3.5; sample volume, 10 mL; addition of salt, 2.0 %; desorption solvent, 40 μ L methanol; desorption time, 5 min. HPLC-UV conditions: Purospher® STAR RP-18 (5 μ m, 150 mm length × 4.6 mm id). isocratic mode using a mobile phase of acetonitrile 25 mM phosphate buffer (pH 3.5) (55:45, v/v), flow rate 1 mL/min, wavelength 210 nm
- Figure 3.6 Effect of type of solvent on the peak area of NSAIDs using dynamic MMMTE. Initial extraction parameters: 500 ng/mL of spiked solution; adsorbent loading, 25%; sample pH 3.5; sample volume, 10 mL; addition of salt, 2.0 %; dynamic extraction cycle, 10; desorption time, 5 min. HPLC-UV conditions: Purospher® STAR RP-18 (5 μ m, 150 mm length × 4.6 mm id). isocratic mode using a mobile phase of acetonitrile 25 mM phosphate buffer (pH 3.5) (55:45, v/v), flow rate 1 mL/min, wavelength 210 nm
- Figure 3.7 Effect of desorption time on the peak area of NSAIDs using dynamic MMMTE. Initial extraction parameters: 500 ng/mL of spiked solution; adsorbent loading, 25%; sample pH 3.5; sample volume, 10 mL; addition of salt, 2.0 %; dynamic extraction cycle, 10; desorption solvent, 40 μ L methanol:ACN; desorption time, 5 min. HPLC-UV conditions: Purospher® STAR RP-18 (5 μ m, 150 mm length × 4.6 mm id). isocratic mode using

47

46

48

45

xiv

a mobile phase of acetonitrile -25 mM phosphate buffer (pH 3.5) (55:45, v/v), flow rate 1 mL/min, wavelength 210 nm

- Figure 3.8 UPLC-MS/MS chromatogram of the separation of the NSAIDs spiked in water sample after treatment with MMMTE under optimum conditions. Dynamic MMMTE conditions: 10 mL sample volume, sample pH 3.5, 2.0 w/v % of salt addition, 40 µL mixture of methanol: acetonitrile (1:1) as desorption solvent and desorption time of 10 min. UPLC-MS/MS Conditions: Waters UPLC BEH C₁₈ (1.7 μ m, 100 mm length \times 2.1 mm I.D.), gradient elution with 10 mM ammonium acetate (A)- 100 % MeOH (B), flow rate 0.3 mL/min, column temperature at 40 °C and injection volume of 5 μL.
- Figure 4.1 Schematic representation of the automated flow through MMM microextraction system
- Figure 4.2 Effect of adsorbent loading on peak areas of CPAHs using automated MMM microextraction- HPLC-UV analysis. Extraction parameters: Initial extraction parameters: 1 μ g/mL of spiked solution; pH, 3; sample flow rate, 0.2 mL/min; sample volume, 2 mL; conditioning solvent, methanol; desorption solvent, 60 μ L of methanol; desorption time, 10 min (static). Peak areas calculated based on average values of peak areas of CPAs, *n* =3. HPLC-UV conditions: Purospher® STAR RP-18 (5 μ m, 150 mm length × 4.6 mm id), gradient mode using a mobile phase acetonitrile (A) and 25 mM phosphate buffer (pH 3) (B), flow rate 1 mL/min, wavelength 230 nm
- Figure 4.3 Effect of sample flowrate on the peak areas of CPAHs using automated MMM microextraction- HPLC-UV analysis. Extraction parameters: Initial extraction parameters: 1 µg/mL of spiked solution; adsorbent loading, 0.0625 mg; pH, 3; addition of salt, 7.5% (w/v); sample volume, 2 mL; conditioning solvent, methanol; desorption solvent, 60 µL of methanol; desorption time, 10 min (static); analyte concentration, 1 µg/mL of spiked solution. Peak areas calculated based on average values of peak areas of CPAs, n = 3. HPLC-UV conditions: Purospher® STAR RP-18 (5 µm, 150 mm length × 4.6 mm id), gradient mode using a mobile phase acetonitrile (A) and 25 mM phosphate buffer (pH 3) (B), flow rate 1 mL/min, wavelength 230 nm
- Figure 4.2 Effect of membrane conditioning/desorption solvent on the peak areas of CPAHs using automated MMM microextraction- HPLC-UV analysis. Extraction

49

51

59

64

conditions: membrane CTA amount, 0.25 mg; MWCNT amount, 0.0625 mg; sample pH, 3; addition of salt, 7.5% (w/v); sample volume, 6 mL; desorption time, 10 min (static); analyte concentration, 1 µg/mL of spiked solution. Peak areas calculated based on average values of peak areas of CPAs, n = 3. HPLC-UV conditions: Purospher® STAR RP-18 (5 µm, 150 mm length × 4.6 mm id), gradient mode using a mobile phase acetonitrile (A) and 25 mM phosphate buffer (pH 3) (B), flow rate 1 mL/min, wavelength 230 nm

Figure 4.2 HPLC/UV chromatograms of CPAHs in (a) blank sewage water samples (b) spiked samples, with CPAHs at concentration of 70 ng/mL each (c) enriched extracts using the automated microextraction method 66

69

LIST OF ABBREVIATIONS

2,4,5-T	-	2-(2,4,5-trichlorophenoxy)acetic acid
2,4-D	-	2,4-dichlorophenoxyacetic acid
2,4-DB	-	4-(2,4-dichlorophenoxy)butanoic acid
2-PrOH	-	2-propanol
3,4-DA	-	3,4-dichlorophenoxyacetic acid
4-CPA	-	(4-chlorophenoxy)acetic acid
C ₁₈	-	Polymeric octadecylsilane particles
CE	-	Capillary electrophoresis
CMS	-	Carbon molecular sieves
CNTs	-	Carbon nanotubes
COF	-	Covalent organic framework
CPAHs	-	Chlorophenoxy acid herbicides
СТА	-	Cellulose triacetate
DCM	-	Dichloromethane
DI	-	Ultrapure deionized
DI-SPME	-	Direct-immersion solid phase microextraction
DLLME	-	Dispersive liquid–liquid microextraction
DS	-	Desorption solution
DSPE	-	Dispersive solid phase extraction
E2	-	Beta-estradiol
EE2	-	Alpha-ethinylestradiol
EF	-	Enrichment factor
EME	-	Electromembrane microextractions
f- Mt	-	Modified montmorillonite
GC	-	Gas chromatography
GO	-	Graphene oxide
HCl	-	Hydrochloric acid
HF-LPME	-	Hollow fibre liquid phase microextraction
HLB	-	Hydrophilic lipophilic balance
HPLC-UV	-	High performance liquid chromatography ultraviolet

HS-SPME	-	Headspace solid phase microextraction
KH ₂ PO ₄	-	Potassium dihydrogen phosphate
LDH/GO	-	Hydroxide-graphene oxide
LLE	-	Liquid-liquid extraction
LOD	-	Limit of detection
LOD	-	Limit of quantification
LPME	-	Liquid-phase microextraction
MCPA	-	2-(4-chloro-2-methylphenoxy)acetic acid
MeCN	-	Acetonitrile
MeOH	-	Methanol
MMM	-	Mixed matrix membrane
MMMTE	-	Mixed matrix membrane tip extraction
MOF	-	Metal organic framework
MP	-	Membrane phase
MRM	-	Multiple reaction monitoring
MWCNTs	-	Multiwall carbon nanotubes
NaCl	-	Sodium chloride
NaOH	-	Sodium hydroxide
NSAIDs	-	Nonsteroidal anti-inflammatory drugs
PAHs	-	Polyaromatic hydrocarbons
PDMS	-	Polymer polydimethyl siloxane
PEBAX	-	Poly ether-block-amide
PEI	-	Polyetherimide
PES	-	Polyethersulfone
PFS	-	Polysulfone
PFTE	-	Polytetrafluoroethylene
PI	-	Polyimide
PMMA	-	Poly(methyl methacrylate)
SDME	-	Single-drop microextraction
SIA	-	Sequential injection analysis
SLG	-	Single layer graphene
SLM	-	Supported liquid membrane
SPE	-	Solid phase extraction

SPME	-	Solid phase microextraction			
SPMTE	-	Solid phase membrane tip extraction			
SS	-	Sample solution			
SWCNTs	-	Single-walled carbon nanotubes			
TFME	-	Thin film microextraction			
UPLC-MS/MS	-	Ultra-performance liquid chromatography-tandem			
		mass			
ZIF	-	Zeolitic imidazolate framework			
β-CD	-	β-cyclodextrin			
μ-SPE	-	Micro Solid Phase Extraction			

LIST OF SYMBOLS

g/L	-	gram per liter
I.D	-	Internal diameter
m ² /g	-	Meter square per gram
mg	-	Miligram
min	-	Minutes
mL	-	Mililiter
mL/min	-	Mililiter per minute
mm	-	Milimeter
mol/L	-	Molarity
ng/L	-	nanogram per liter
ng/mL	-	nanogram per mililiter
nm	-	nanometer
pg/mL	-	picogram per mililiter
рКа	-	Acid dissociation constant
v/v	-	volume per volume
w/v	-	Weight per volume
w/w	-	weight per weight
wt %	-	weight percentage
μg	-	Microgram
μg/L	-	Microgram per liter
µg/mL	-	Microgram per mililiter
μL	-	Microliter
μL/min	-	Microliter per minute

CHAPTER 1

INTRODUCTION

1.1 Background Study

The development of fast and sensitive determination of trace levels of contaminants in environmental complex matrices remains challenging in analytical method [1]. Sample pre-treatment is a necessary step to quantify analytes in complex matrices and enrichment of the target analytes to final acceptor phase. However, sample preparation step is regarded time-consuming step and prone to significant experimental errors in analysis.

Liquid-liquid extraction (LLE) and solid phase extraction (SPE) are wellknown classical sample pre-treatment techniques for the extraction of target compounds that are generally preferred; however, limitations often stated are time-consuming, large organic waste is produced and the need for multistep procedures with high risk of analyte loss [2]. Recent developments in sample pretreatment have largely been aimed at the need for very small volumes of the extracting phase or the possibility to become solventless as well as the ability to incorporate several processing steps such as sampling, detection, separation, preconcentration, and even derivatization [3].

Over the past decade, membrane technologies have grown rapidly by developing materials that offer improvements in flux and selectivity. Various techniques of membrane-based microextraction [4] have been employed in the separation and detection of organic pollutants from different environmental matrices. For instance, the use of hollow fibre liquid phase microextraction (HF-LPME) [5], solid phase membrane tip extraction (SPMTE) [6], micro–solid phase extraction (μ -SPE) [7] and thin film microextraction (TFME) [8] are proven to be simplified in experimental setup and low cost as well as reduce organic solvent consumption. In

these published techniques, membrane-based extraction techniques is based on the use of a membrane as a selective thin barrier between miscible fluids for sample pre-treatment [9].

A mixed matrix membrane (MMM) is a type of membrane formed by the incorporation of inorganic materials in polymeric membranes that has attracted much attention recently. Several inorganic nanoparticles such as zeolites [10, 11], silica [12], carbon nanotubes (CNTs) [13, 14], metal organic frameworks (MOFs) [15, 16] and graphene [17, 18] have been introduced as fillers into various polymeric matrices to enhance the performance of MMM. This combination of MMM has resulted in high efficiency in extraction of pollutants due to better flexibility, sensitivity, selectivity, mechanical, thermal, and chemical stability being achieved. The use of MMM has been reported in gas separation applications [19-24] and liquid phase separations [25-33].

Herein, we demonstrated two MMM microextraction methods, namely dynamic MMM tip extraction (MMMTE) and automated flow-through MMM microextraction. Fabrication of MMM using carbonaceous material immobilized in cellulose triacetate polymer matrix for microextraction was developed. The applicability and reliability of utilizing carbonaceous-based MMM in two different miniaturised microextraction designs were comprehensively investigated, optimised and validated by determining the presence of the selected nonsteroidal antiinflammatory drugs (NSAIDs) and chlorophenoxy acid herbicides (CPAHs) in real environmental water samples.

1.2 Problem Statement

The contamination of pharmaceuticals and herbicides in environment has now become critical issues due to their continuous release into the environment. Diclofenac, together with 17 alpha-ethinylestradiol (EE2) and 17 beta-estradiol (E2) is listed as new priority restricted substance to be scrutinized in Europe in surface water. Adding to that, several CPAHs have been included in the European list of priority pollutants. Moreover, the U.S. Environmental Protection Agency (USEPA) has set a maximum tolerated contaminant level for drinking water of 2,4-dichlorophenoxyacetic acid (2,4-D) at 70 μ g/L. These organic pollutants pose significant toxicological risk to humans and marine life. However, direct analysis of samples of these pollutants is difficult because they are in complex matrices and in very low concentrations [34]. Moreover, sample extraction and clean-up procedures greatly influence the accuracy and precision of the analysis. For all these reasons, sample preparation technique is crucial for an efficient, reliable, and accurate data analysis.

Although classical extraction methods such as liquid-liquid extraction (LLE) and solid phase extraction (SPE) are generally favoured, however these extraction techniques are time-consuming and require large amounts of toxic and expensive solvents. Nowadays, miniaturized techniques have been utilized to improve extraction selectivity, less consumption of organic solvent, low-cost, and minimize the sample pre-treatment steps. However, these existing procedures, have several disadvantages in terms of cost of analysis, lower enrichment power and possible analyte carryover effects. In this present study, new miniaturised sample pre-treatment techniques based on MMM are demonstrated for a smaller sample size, less consumption of organic solvent and more cost-effective sample pre-treatment method.

1.3 Objectives of Study

The aim of this research is to develop miniaturised microextraction techniques using the MMM microextraction technique for the analysis of NSAIDs and CPAHs in environmental water matrices. To achieve this aim, the following objectives have been planned:

• To develop and validate the dynamic MMMTE coupled with ultraperformance liquid chromatography-tandem mass spectrometry (UPLC-MS/MS) for the determination of NSAIDs in environmental water matrices.

3

• To develop and validate the automated flow-through MMM microextraction method coupled with high performance liquid chromatography ultraviolet detection (HPLC-UV) for the determination of CPAHs in environmental water matrices.

1.4 Scope of Study

In this study, an advanced method to fabricate MMM in a commercially available pipette tip is demonstrated. This new sample preparation method designated as dynamic MMMTE was established for the analysis of NSAIDs in water samples using polymeric octadecylsilane (C_{18}) particles. Operational parameters of MMMTE such as effect of sample pH, salting-out, dynamic extraction cycle, type of solvent and desorption time were investigated. The developed dynamic MMMTE approach was coupled with UPLC-MS/MS for trace level detection of pollutants present in the environmental water samples.

On the other hand, an automated analyte pre-concentration system was developed for the analysis of CPAHs in sewage water samples using a multiwall carbon nanotubes (MWCNTs) immobilized mixed matrix membrane. The analyte adsorption and desorption processes were performed in a fully automated mode by making use of a sequential injection analysis (SIA) manifold based on a reversible syringe pump fitted with a selection valve to perform liquid handling. Several extraction conditions were comprehensively optimized including sample pH, ionic strength, sample volume, extraction time, desorption solvent and desorption time. The enriched extracts obtained from the automated approach were subsequently analysed by the HPLC-UV technique for separation and quantification.

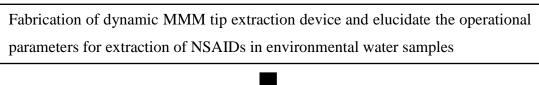
1.5 Significance of Research

This research is significant to the development of the sample pre-treatment step in the MMM microextraction technique. This procedure has been developed to speed up and simplify the microextraction method, smaller initial sample volumes is required, high sensitivity and selectivity is achieved. It also minimizes organic solvent consumption and improve the efficiency of extraction of organic pollutants. This study has focused on the wider range of nanomaterial adsorbents based on MMM application in various sizes and shapes to tailor different experimental designs and sample size requirements for the analysis of NSAIDs and CPAHs in environmental analysis. Besides that, through this work determination of various organic pollutants in complex, matrices can be regarded as green analytical chemistry method due to consumption of microliters organic solvent, simplicity of the microextraction setup and cost effectiveness.

1.6 Flowchart/Scheme of the Research

In Chapter 1, a detailed account of the research background, problem statement, objective, scope, and significance of the study. In Chapter 2 provides the literature review on conventional extraction and microextraction techniques, mixed matrix membrane and model analytes in this study.

In Chapter 3, the experimental methodology and application of the dynamic mixed matrix membrane tip extraction for the determination of five selected NSAIDs in environmental water samples.





Validate and apply the dynamic MMM microextraction coupled with UPLC-MS/MS for monitoring of NSAIDs in environmental water samples In Chapter 4, the experimental methodology and application of automated flow through cell MMM microextraction for the determination of six selected CPAHs in environmental water samples.

Fabrication of flow cell device immobilised with MMM and elucidate the automated parameters for extraction of CPAHs



Validate and apply the automated flow-through MMM microextraction coupled with HPLC-UV for monitoring of CPAHs in environmental water samples

Figure 1.1 Flowchart of research work for MMM microextraction

REFERENCES

- Ribeiro, C., Ribeiro, A. R., Maia, A. S., Gonçalves, V. M. F., Tiritan, M. E. New Trends in Sample Preparation Techniques for Environmental Analysis. *Critical Reviews in Analytical Chemistry*. 2014. 44(2): 142-85.
- Sarah Montesdeoca-Esponda, M Esther Torres-Padrón, Zoraida Sosa-Ferrera, Santana-Rodríguez, J. J. Environmental Applications of Solid Phase Microextraction Techniques. *Analytical Separation Science*. 1897-927.
- Dimpe, K. M., Nomngongo, P. N. Current sample preparation methodologies for analysis of emerging pollutants in different environmental matrices. *TrAC Trends in Analytical Chemistry*. 2016. 82: 199-207.
- 4. Carasek, E., Merib, J. Membrane-based microextraction techniques in analytical chemistry: A review. *Analytica Chimica Acta*. 2015. 880: 8-25.
- Al Azzam, K. M., Makahleah, A., Saad, B., Mansor, S. M. Hollow fiber liquid-phase microextraction for the determination of trace amounts of rosiglitazone (anti-diabetic drug) in biological fluids using capillary electrophoresis and high performance liquid chromatographic methods. *Journal of Chromatography A*. 2010. 1217(23): 3654-9.
- See, H. H., Marsin Sanagi, M., Ibrahim, W. A. W., Naim, A. A. Determination of triazine herbicides using membrane-protected carbon nanotubes solid phase membrane tip extraction prior to micro-liquid chromatography. *Journal of Chromatography A*. 2010. 1217(11): 1767-72.
- Lashgari, M., Lee, H. K. Micro-solid phase extraction of perfluorinated carboxylic acids from human plasma. *Journal of Chromatography A*. 2016. 1432: 7-16.
- 8. Olcer, Y. A., Tascon, M., Eroglu, A. E., Boyacı, E. Thin film microextraction: Towards faster and more sensitive microextraction. *TrAC Trends in Analytical Chemistry*. 2019. 113: 93-101.
- Goh, P. S., Ismail, A. F., Sanip, S. M., Ng, B. C., Aziz, M. Recent advances of inorganic fillers in mixed matrix membrane for gas separation. *Separation and Purification Technology*. 2011. 81(3): 243-64.

- Zhan, X., Lu, J., Tan, T., Li, J. Mixed matrix membranes with HF acid etched ZSM-5 for ethanol/water separation: Preparation and pervaporation performance. *Applied Surface Science*. 2012. 259: 547-56.
- Gong, H., Lee, S. S., Bae, T.-H. Mixed-matrix membranes containing inorganically surface-modified 5A zeolite for enhanced CO2/CH4 separation. *Microporous and Mesoporous Materials*. 2017. 237: 82-9.
- Waqas Anjum, M., de Clippel, F., Didden, J., Laeeq Khan, A., Couck, S., Baron, G. V., et al. Polyimide mixed matrix membranes for CO2 separations using carbon-silica nanocomposite fillers. *Journal of Membrane Science*. 2015. 495: 121-9.
- Wang, L., Song, X., Wang, T., Wang, S., Wang, Z., Gao, C. Fabrication and characterization of polyethersulfone/carbon nanotubes (PES/CNTs) based mixed matrix membranes (MMMs) for nanofiltration application. *Applied Surface Science*. 2015. 330: 118-25.
- Azimi, H., Tezel, F. H., Thibault, J. Effect of embedded activated carbon nanoparticles on the performance of polydimethylsiloxane (PDMS) membrane for pervaporation separation of butanol. *Journal of Chemical Technology & Biotechnology*. 2017. 92(12): 2901-11.
- 15. Jia, Z., Wu, G. Metal-organic frameworks based mixed matrix membranes for pervaporation. *Microporous and Mesoporous Materials*. 2016. 235: 151-9.
- de la Iglesia, Ó., Sorribas, S., Almendro, E., Zornoza, B., Téllez, C., Coronas, J. Metal-organic framework MIL-101(Cr) based mixed matrix membranes for esterification of ethanol and acetic acid in a membrane reactor. *Renewable Energy*. 2016. 88: 12-9.
- Qian, X., Li, N., Wang, Q., Ji, S. Chitosan/graphene oxide mixed matrix membrane with enhanced water permeability for high-salinity water desalination by pervaporation. *Desalination*. 2018. 438: 83-96.
- Wang, L., Wang, N., Yang, H., An, Q., Li, B., Ji, S. Facile fabrication of mixed matrix membranes from simultaneously polymerized hyperbranched polymer/modified graphene oxide for MTBE/MeOH separation. *Journal of Membrane Science*. 2018. 559: 8-18.
- Molki, B., Aframehr, W. M., Bagheri, R., Salimi, J. Mixed matrix membranes of polyurethane with nickel oxide nanoparticles for CO2 gas separation. *Journal of Membrane Science*. 2018. 549: 588-601.

- Safak Boroglu, M., Ugur, M., Boz, I. Enhanced gas transport properties of mixed matrix membranes consisting of Matrimid and RHO type ZIF-12 particles. *Chemical Engineering Research and Design*. 2017. 123: 201-13.
- Zarshenas, K., Raisi, A., Aroujalian, A. Mixed matrix membrane of nanozeolite NaX/poly (ether-block-amide) for gas separation applications. *Journal* of Membrane Science. 2016. 510: 270-83.
- Martin-Gil, V., López, A., Hrabanek, P., Mallada, R., Vankelecom, I. F. J., Fila, V. Study of different titanosilicate (TS-1 and ETS-10) as fillers for Mixed Matrix Membranes for CO2/CH4 gas separation applications. *Journal* of Membrane Science. 2017. 523: 24-35.
- Zornoza, B., Téllez, C., Coronas, J. Mixed matrix membranes comprising glassy polymers and dispersed mesoporous silica spheres for gas separation. *Journal of Membrane Science*. 2011. 368(1): 100-9.
- Le, N. L., Wang, Y., Chung, T.-S. Pebax/POSS mixed matrix membranes for ethanol recovery from aqueous solutions via pervaporation. *Journal of Membrane Science*. 2011. 379(1): 174-83.
- Qadir, D., Mukhtar, H., Keong, L. K. Synthesis and Characterization of Polyethersulfone/Carbon Molecular Sieve Based Mixed Matrix Membranes for Water Treatment Applications. *Procedia Engineering*. 2016. 148: 588-93.
- 26. Rozaini, M. N. H., Semail, N.-f., Saad, B., Kamaruzaman, S., Abdullah, W. N., Rahim, N. A., et al. Molecularly imprinted silica gel incorporated with agarose polymer matrix as mixed matrix membrane for separation and preconcentration of sulfonamide antibiotics in water samples. *Talanta*. 2019. 199: 522-31.
- 27. Rajeswari, A., Jackcina Stobel Christy, E., Ida Celine Mary, G., Jayaraj, K., Pius, A. Cellulose acetate based biopolymeric mixed matrix membranes with various nanoparticles for environmental remediation-A comparative study. *Journal of Environmental Chemical Engineering*. 2019. 7(4): 103278.
- Zhang, G., Zhou, M., Xu, Z., Jiang, C., Shen, C., Meng, Q. Guanidylfunctionalized graphene/polysulfone mixed matrix ultrafiltration membrane with superior permselective, antifouling and antibacterial properties for water treatment. *Journal of Colloid and Interface Science*. 2019. 540: 295-305.

- 29. Tang, W., Lou, H., Li, Y., Kong, X., Wu, Y., Gu, X. Ionic liquid modified graphene oxide-PEBA mixed matrix membrane for pervaporation of butanol aqueous solutions. *Journal of Membrane Science*. 2019. 581: 93-104.
- Mukhtar, N. H., See, H. H. Carbonaceous nanomaterials immobilised mixed matrix membrane microextraction for the determination of polycyclic aromatic hydrocarbons in sewage pond water samples. *Analytica Chimica Acta*. 2016. 931: 57-63.
- Kamaruzaman, S., Hauser, P. C., Sanagi, M. M., Ibrahim, W. A. W., Endud, S., See, H. H. A simple microextraction and preconcentration approach based on a mixed matrix membrane. *Analytica Chimica Acta*. 2013. 783: 24-30.
- 32. Mao, X., He, M., Chen, B., Hu, B. Membrane protected C18 coated stir bar sorptive extraction combined with high performance liquid chromatographyultraviolet detection for the determination of non-steroidal anti-inflammatory drugs in water samples. *Journal of Chromatography A*. 2016. 1472: 27-34.
- 33. Ghani, M., Ghoreishi, S. M., Azamati, M. Magnesium-aluminum-layered double hydroxide-graphene oxide composite mixed-matrix membrane for the thin-film microextraction of diclofenac in biological fluids. *Journal of Chromatography A*. 2018. 1575: 11-7.
- Picó, Y., Fernández, M., Ruiz, M. J., Font, G. Current trends in solid-phasebased extraction techniques for the determination of pesticides in food and environment. *Journal of Biochemical and Biophysical Methods*. 2007. 70(2): 117-31.
- 35. Wen, Y., Chen, L., Li, J., Liu, D., Chen, L. Recent advances in solid-phase sorbents for sample preparation prior to chromatographic analysis. *TrAC Trends in Analytical Chemistry*. 2014. 59: 26-41.
- Otles, S., Ozyurt, V. H. Sampling and Sample Preparation. In: Cheung, P. C. K., Mehta, B. M., editors. Handbook of Food Chemistry. Berlin, Heidelberg: Springer Berlin Heidelberg; 2015. p. 151-64.
- Ridgway, K., Lalljie, S. P. D., Smith, R. M. Sample preparation techniques for the determination of trace residues and contaminants in foods. *Journal of Chromatography A*. 2007. 1153(1): 36-53.
- Tankiewicz, M., Fenik, J., Biziuk, M. Solventless and solvent-minimized sample preparation techniques for determining currently used pesticides in water samples: A review. *Talanta*. 2011. 86: 8-22.

- Chormey, D. S., Bakırdere, S. Chapter Seven Principles and Recent Advancements in Microextraction Techniques. In: Chormey, D. S., Bakırdere, S., Turan, N. B., Engin, G. Ö., editors. Comprehensive Analytical Chemistry. 81: Elsevier; 2018. p. 257-94.
- Clement, R. E., Hao, C. 2.03 Liquid–Liquid Extraction: Basic Principles and Automation. In: Pawliszyn, J., editor. Comprehensive Sampling and Sample Preparation. Oxford: Academic Press; 2012. p. 51-63.
- Moldoveanu, S., David, V. Chapter 6 Solvent Extraction. In: Moldoveanu,
 S., David, V., editors. Modern Sample Preparation for Chromatography. Amsterdam: Elsevier; 2015. p. 131-89.
- 42. Nerín, C., Salafranca, J., Aznar, M., Batlle, R. Critical review on recent developments in solventless techniques for extraction of analytes. *Analytical and Bioanalytical Chemistry*. 2008. 393(3): 809.
- 43. Tran, N. H., Chen, H., Do, T. V., Reinhard, M., Ngo, H. H., He, Y., et al. Simultaneous analysis of multiple classes of antimicrobials in environmental water samples using SPE coupled with UHPLC-ESI-MS/MS and isotope dilution. *Talanta*. 2016. 159: 163-73.
- 44. Dai, X., Jia, X., Zhao, P., Wang, T., Wang, J., Huang, P., et al. A combined experimental/computational study on metal-organic framework MIL-101(Cr) as a SPE sorbent for the determination of sulphonamides in environmental water samples coupling with UPLC-MS/MS. *Talanta*. 2016. 154: 581-8.
- 45. Gurke, R., Rossmann, J., Schubert, S., Sandmann, T., Rößler, M., Oertel, R., et al. Development of a SPE-HPLC–MS/MS method for the determination of most prescribed pharmaceuticals and related metabolites in urban sewage samples. *Journal of Chromatography B*. 2015. 990: 23-30.
- 46. Paíga, P., Santos, L. H. M. L. M., Delerue-Matos, C. Development of a multiresidue method for the determination of human and veterinary pharmaceuticals and some of their metabolites in aqueous environmental matrices by SPE-UHPLC–MS/MS. *Journal of Pharmaceutical and Biomedical Analysis*. 2017. 135: 75-86.
- 47. Ji, W.-H., Guo, Y.-S., Wang, X., Lu, X.-F., Guo, D.-S. Amino-modified covalent organic framework as solid phase extraction absorbent for determination of carboxylic acid pesticides in environmental water samples. *Journal of Chromatography A*. 2019. 1595: 11-8.

- Yang, H.-H., Zhou, W.-H., Guo, X.-C., Chen, F.-R., Zhao, H.-Q., Lin, L.-M., et al. Molecularly imprinted polymer as SPE sorbent for selective extraction of melamine in dairy products. *Talanta*. 2009. 80(2): 821-5.
- 49. Zhou, N.-Z., Liu, P., Su, X.-C., Liao, Y.-H., Lei, N.-S., Liang, Y.-H., et al. Low-cost humic acid-bonded silica as an effective solid-phase extraction sorbent for convenient determination of aflatoxins in edible oils. *Analytica Chimica Acta*. 2017. 970: 38-46.
- Płotka-Wasylka, J., Szczepańska, N., de la Guardia, M., Namieśnik, J. Miniaturized solid-phase extraction techniques. *TrAC Trends in Analytical Chemistry*. 2015. 73: 19-38.
- Andrade-Eiroa, A., Canle, M., Leroy-Cancellieri, V., Cerdà, V. Solid-phase extraction of organic compounds: A critical review (Part I). *TrAC Trends in Analytical Chemistry*. 2016. 80: 641-54.
- Arthur, C. L., Pawliszyn, J. Solid phase microextraction with thermal desorption using fused silica optical fibers. *Analytical Chemistry*. 1990. 62(19): 2145-8.
- Kataoka, H. Current Developments and Future Trends in Solid-phase Microextraction Techniques for Pharmaceutical and Biomedical Analyses. *Analytical Sciences*. 2011. 27(9): 893-.
- Kataoka, H., Lord, H. L., Pawliszyn, J. Applications of solid-phase microextraction in food analysis. *Journal of Chromatography A*. 2000. 880(1): 35-62.
- 55. Souza-Silva, É. A., Jiang, R., Rodríguez-Lafuente, A., Gionfriddo, E., Pawliszyn, J. A critical review of the state of the art of solid-phase microextraction of complex matrices I. Environmental analysis. *TrAC Trends in Analytical Chemistry*. 2015. 71: 224-35.
- 56. Razmi, H., Khosrowshahi, E. M., Farrokhzadeh, S. Introduction of coiled solid phase microextraction fiber coated by mesoporous silica/cetyltrimethylammonium bromide for ultra-trace environmental analysis. *Journal of Chromatography A*. 2017. 1506: 1-8.
- 57. Lan, H., Rönkkö, T., Parshintsev, J., Hartonen, K., Gan, N., Sakeye, M., et al. Modified zeolitic imidazolate framework-8 as solid-phase microextraction Arrow coating for sampling of amines in wastewater and food samples

followed by gas chromatography-mass spectrometry. *Journal of Chromatography A*. 2017. 1486: 76-85.

- 58. Wang, R., Li, W., Chen, Z. Solid phase microextraction with poly(deep eutectic solvent) monolithic column online coupled to HPLC for determination of non-steroidal anti-inflammatory drugs. *Analytica Chimica Acta*. 2018. 1018: 111-8.
- Saraji, M., Bidgoli, A. A. H. Dispersive liquid–liquid microextraction using a surfactant as disperser agent. *Analytical and Bioanalytical Chemistry*. 2010. 397(7): 3107-15.
- Rutkowska, M., Płotka-Wasylka, J., Sajid, M., Andruch, V. Liquid–phase microextraction: A review of reviews. *Microchemical Journal*. 2019. 149: 103989.
- 61. Sarafraz-Yazdi, A., Amiri, A. Liquid-phase microextraction. *TrAC Trends in Analytical Chemistry*. 2010. 29(1): 1-14.
- Zhao, L., Lee, H. K. Application of static liquid-phase microextraction to the analysis of organochlorine pesticides in water. *Journal of Chromatography A*. 2001. 919(2): 381-8.
- Huang, S.-P., Huang, S.-D. Determination of organochlorine pesticides in water using solvent cooling assisted dynamic hollow-fiber-supported headspace liquid-phase microextraction. *Journal of Chromatography A*. 2007. 1176(1): 19-25.
- 64. Xu, J., Liang, P., Zhang, T. Dynamic liquid-phase microextraction of three phthalate esters from water samples and determination by gas chromatography. *Analytica Chimica Acta*. 2007. 597(1): 1-5.
- Salvatierra-stamp, V., Muñiz-Valencia, R., Jurado, J. M., Ceballos-Magaña,
 S. G. Hollow fiber liquid phase microextraction combined with liquid chromatography-tandem mass spectrometry for the analysis of emerging contaminants in water samples. *Microchemical Journal*. 2018. 140: 87-95.
- Zhao, L., Lee, H. Application of Static Liquid-Phase Microextraction to the Analysis of Organochlorine Pesticides in Water. *Journal of Chromatography* A. 2001. 919: 381-8.
- Rasmussen, K. E., Pedersen-Bjergaard, S. Developments in hollow fibrebased, liquid-phase microextraction. *TrAC Trends in Analytical Chemistry*. 2004. 23(1): 1-10.

- 68. Esrafili, A., Baharfar, M., Tajik, M., Yamini, Y., Ghambarian, M. Two-phase hollow fiber liquid-phase microextraction. *TrAC Trends in Analytical Chemistry*. 2018. 108: 314-22.
- 69. Tajabadi, F., Ghambarian, M., Yamini, Y., Yazdanfar, N. Combination of hollow fiber liquid phase microextraction followed by HPLC-DAD and multivariate curve resolution to determine antibacterial residues in foods of animal origin. *Talanta*. 2016. 160: 400-9.
- 70. Cai, J. a., Chen, G., Qiu, J., Jiang, R., Zeng, F., Zhu, F., et al. Hollow fiber based liquid phase microextraction for the determination of organochlorine pesticides in ecological textiles by gas chromatography–mass spectrometry. *Talanta*. 2016. 146: 375-80.
- Zhang, J., Su, T., Lee, H. K. Development and application of microporous hollow fiber protected liquid-phase microextraction via gaseous diffusion to the determination of phenols in water. *Journal of Chromatography A*. 2006. 1121(1): 10-5.
- 72. Ramos Payán, M., Bello López, M. Á., Fernández-Torres, R., Callejón Mochón, M., Gómez Ariza, J. L. Application of hollow fiber-based liquidphase microextraction (HF-LPME) for the determination of acidic pharmaceuticals in wastewaters. *Talanta*. 2010. 82(2): 854-8.
- Asensio-Ramos, M., Ravelo-Pérez, L. M., González-Curbelo, M. Á., Hernández-Borges, J. Liquid phase microextraction applications in food analysis. *Journal of Chromatography A*. 2011. 1218(42): 7415-37.
- 74. Pedersen-Bjergaard, S., Rasmussen, K. Liquid-phase microextraction and capillary electrophoresis of acidic drugs. *Electrophoresis*. 2000. 21: 579-85.
- 75. Basheer, C., Alnedhary, A., Rao, B., Valliyaveettil, S., Lee, H. Development and Application of Porous Membrane-Protected Carbon Nanotube Micro-Solid-Phase Extraction Combined with Gas Chromatography/Mass Spectrometry. *Analytical chemistry*. 2006. 78: 2853-8.
- 76. Manaf, N. A., Saad, B., Mohamed, M. H., Wilson, L. D., Latiff, A. A. Cyclodextrin based polymer sorbents for micro-solid phase extraction followed by liquid chromatography tandem mass spectrometry in determination of endogenous steroids. *Journal of Chromatography A*. 2018. 1543: 23-33.

- Nojavan, S., Yazdanpanah, M. Micro-solid phase extraction of benzene, toluene, ethylbenzene and xylenes from aqueous solutions using water-insoluble β-cyclodextrin polymer as sorbent. *Journal of Chromatography A*. 2017. 1525: 51-9.
- Pedersen-Bjergaard, S., Rasmussen, K. E. Electrokinetic migration across artificial liquid membranes: New concept for rapid sample preparation of biological fluids. *Journal of Chromatography A*. 2006. 1109(2): 183-90.
- Gjelstad, A., Rasmussen, K. E., Pedersen-Bjergaard, S. Electrokinetic migration across artificial liquid membranes: Tuning the membrane chemistry to different types of drug substances. *Journal of Chromatography* A. 2006. 1124(1): 29-34.
- 80. Gjelstad, A., Pedersen-Bjergaard, S. Recent developments in electromembrane extraction. *Analytical Methods*. 2013. 5(18): 4549-57.
- Payán, M. R., López, M. Á. B., Torres, R. F., Navarro, M. V., Mochón, M. C. Electromembrane extraction (EME) and HPLC determination of non-steroidal anti-inflammatory drugs (NSAIDs) in wastewater samples. *Talanta*. 2011. 85(1): 394-9.
- 82. Chong, M. H., Sanagi, M. M., Endud, S., Ibrahim, W. A. W., Lau, S. C., Alharbi, O. M. L., et al. Determination of N-nitrosamines in water by nano iron-porphyrinated poly(amidoamine) dendrimer MCM-41 generation-3 through solid phase membrane tip extraction and HPLC. *Environmental Technology & Innovation*. 2018. 10: 102-10.
- Kamaruzaman, S., Sanagi, M. M., Endud, S., Wan Ibrahim, W. A., Yahaya, N. MCM-41 solid phase membrane tip extraction combined with liquid chromatography for the determination of non-steroidal anti-inflammatory drugs in human urine. *Journal of Chromatography B*. 2013. 940: 59-65.
- Jiang, R., Pawliszyn, J. Thin-film microextraction offers another geometry for solid-phase microextraction. *TrAC Trends in Analytical Chemistry*. 2012. 39: 245-53.
- 85. Bruheim, I., Liu, X., Pawliszyn, J. Thin-Film Microextraction. *Analytical Chemistry*. 2003. 75(4): 1002-10.
- Togunde, O., Cudjoe, E., Oakes, K., Mirnaghi, F., Servos, M., Pawliszyn, J.
 Determination of selected pharmaceutical residues in wastewater using an

automated open bed solid phase microextraction system. *Journal of Chromatography A*. 2012. 1262: 34–42.

- Paul, D., Kemp, D. The Diffusion Time Lag in Polymer Membranes Containing Adsorptive Fillers. *Journal of Polymer Science: Polymer Symposia*. 2007. 41: 79-93.
- Chung, T.-S., Jiang, L. Y., Li, Y., Kulprathipanja, S. Mixed matrix membranes (MMMs) comprising organic polymers with dispersed inorganic fillers for gas separation. *Progress in Polymer Science*. 2007. 32(4): 483-507.
- Aroon, M. A., Ismail, A. F., Matsuura, T., Montazer-Rahmati, M. M. Performance studies of mixed matrix membranes for gas separation: A review. *Separation and Purification Technology*. 2010. 75(3): 229-42.
- 90. Siddique, H., Rundquist, E., Bhole, Y., Peeva, L. G., Livingston, A. G. Mixed matrix membranes for organic solvent nanofiltration. *Journal of Membrane Science*. 2014. 452: 354-66.
- 91. Tetala, K. K. R., Stamatialis, D. F. Mixed matrix membranes for efficient adsorption of copper ions from aqueous solutions. *Separation and Purification Technology*. 2013. 104: 214-20.
- Kim, S., Marand, E. High permeability nano-composite membranes based on mesoporous MCM-41 nanoparticles in a polysulfone matrix. *Microporous* and Mesoporous Materials. 2008. 114(1): 129-36.
- 93. Pechar, T. W., Kim, S., Vaughan, B., Marand, E., Tsapatsis, M., Jeong, H. K., et al. Fabrication and characterization of polyimide–zeolite L mixed matrix membranes for gas separations. *Journal of Membrane Science*. 2006. 277(1): 195-202.
- 94. Şen, D., Kalıpçılar, H., Yilmaz, L. Development of polycarbonate based zeolite 4A filled mixed matrix gas separation membranes. *Journal of Membrane Science*. 2007. 303(1): 194-203.
- 95. Li, Y., Chung, T.-S., Huang, Z., Kulprathipanja, S. Dual-layer polyethersulfone (PES)/BTDA-TDI/MDI co-polyimide (P84) hollow fiber membranes with a submicron PES–zeolite beta mixed matrix dense-selective layer for gas separation. *Journal of Membrane Science*. 2006. 277(1): 28-37.
- Cong, H., Radosz, M., Towler, B. F., Shen, Y. Polymer–inorganic nanocomposite membranes for gas separation. *Separation and Purification Technology*. 2007. 55(3): 281-91.

- Zhang, Y., Li, H., Lin, J., Li, R., Liang, X. Preparation and characterization of zirconium oxide particles filled acrylonitrile-methyl acrylate-sodium sulfonate acrylate copolymer hybrid membranes. *Desalination*. 2006. 192(1): 198-206.
- 98. Husain, S., Koros, W. J. Mixed matrix hollow fiber membranes made with modified HSSZ-13 zeolite in polyetherimide polymer matrix for gas separation. *Journal of Membrane Science*. 2007. 288(1): 195-207.
- 99. Rafizah, W. A. W., Ismail, A. F. Effect of carbon molecular sieve sizing with poly(vinyl pyrrolidone) K-15 on carbon molecular sieve–polysulfone mixed matrix membrane. *Journal of Membrane Science*. 2008. 307(1): 53-61.
- Chung, T.-S., Jiang, L. Y., Li, Y., Kulprathipanja, S. Mixed matrix membranes (MMMs) comprising organic polymers with dispersed inorganic fillers for gas separation. 2007.
- Tantekin-Ersolmaz, Ş. B., Atalay-Oral, Ç., Tatlıer, M., Erdem-Şenatalar, A., Schoeman, B., Sterte, J. Effect of zeolite particle size on the performance of polymer–zeolite mixed matrix membranes. *Journal of Membrane Science*. 2000. 175(2): 285-8.
- 102. He, Z., Pinnau, I., Morisato, A. Nanostructured poly(4-methyl-2pentyne)/silica hybrid membranes for gas separation. *Desalination*. 2002. 146(1): 11-5.
- 103. Murugiah, P. S., Oh, P. C., Lau, K. K. Collegial effect of carbonaceous hybrid fillers in mixed matrix membrane development. *Reactive and Functional Polymers*. 2019. 135: 8-15.
- 104. Dai, Z., Ansaloni, L., Deng, L. Recent advances in multi-layer composite polymeric membranes for CO2 separation: A review. *Green Energy & Environment*. 2016. 1(2): 102-28.
- 105. Dunleavy, M. Polymeric membranes. A review of applications. *Medical device technology*. 1996. 7(4): 14-6, 8-21.
- 106. Li, H., Haas-Santo, K., Schygulla, U., Dittmeyer, R. Inorganic microporous membranes for H2 and CO2 separation—Review of experimental and modeling progress. *Chemical Engineering Science*. 2015. 127: 401-17.
- 107. Wan Ikhsan, S. N., Yusof, N., Aziz, F., Misdan, N., Ismail, A. F., Lau, W.-J., et al. Efficient separation of oily wastewater using polyethersulfone mixed

matrix membrane incorporated with halloysite nanotube-hydrous ferric oxide nanoparticle. *Separation and Purification Technology*. 2018. 199: 161-9.

- 108. Dong, G., Hou, J., Wang, J., Zhang, Y., Chen, V., Liu, J. Enhanced CO2/N2 separation by porous reduced graphene oxide/Pebax mixed matrix membranes. *Journal of Membrane Science*. 2016. 520: 860-8.
- 109. Dorosti, F., Alizadehdakhel, A. Fabrication and investigation of PEBAX/Fe-BTC, a high permeable and CO2 selective mixed matrix membrane. *Chemical Engineering Research and Design*. 2018. 136: 119-28.
- Etxeberria-Benavides, M., David, O., Johnson, T., Łozińska, M. M., Orsi, A., Wright, P. A., et al. High performance mixed matrix membranes (MMMs) composed of ZIF-94 filler and 6FDA-DAM polymer. *Journal of Membrane Science*. 2018. 550: 198-207.
- 111. Yazdi, M. N., Yamini, Y., Asiabi, H. Multiwall carbon nanotube- zirconium oxide nanocomposite hollow fiber solid phase microextraction for determination of polyaromatic hydrocarbons in water, coffee and tea samples. *Journal of Chromatography A*. 2018. 1554: 8-15.
- 112. Jia, M., Feng, Y., Qiu, J., Zhang, X.-F., Yao, J. Amine-functionalized MOFs@GO as filler in mixed matrix membrane for selective CO2 separation. *Separation and Purification Technology*. 2019. 213: 63-9.
- 113. Lin, R., Villacorta Hernandez, B., Ge, L., Zhu, Z. Metal organic framework based mixed matrix membranes: an overview on filler/polymer interfaces. *Journal of Materials Chemistry A*. 2018. 6(2): 293-312.
- 114. Cao, X., Qiao, Z., Wang, Z., Zhao, S., Li, P., Wang, J., et al. Enhanced performance of mixed matrix membrane by incorporating a highly compatible covalent organic framework into poly(vinylamine) for hydrogen purification. *International Journal of Hydrogen Energy*. 2016. 41(21): 9167-74.
- 115. Jamil, A., Ching, O. P., Shariff, A. M. Mixed matrix hollow fibre membrane comprising polyetherimide and modified montmorillonite with improved filler dispersion and CO2/CH4 separation performance. *Applied Clay Science*. 2017. 143: 115-24.
- 116. Nasir, R., Ahmad, N. N. R., Mukhtar, H., Mohshim, D. F. Effect of ionic liquid inclusion and amino-functionalized SAPO-34 on the performance of

mixed matrix membranes for CO2/CH4 separation. *Journal of Environmental Chemical Engineering*. 2018. 6(2): 2363-8.

- 117. Mukherjee, R., Bhunia, P., De, S. Impact of graphene oxide on removal of heavy metals using mixed matrix membrane. *Chemical Engineering Journal*. 2016. 292: 284-97.
- 118. Wang, X., Chen, J., Fang, M., Wang, T., Yu, L., Li, J. ZIF-7/PDMS mixed matrix membranes for pervaporation recovery of butanol from aqueous solution. *Separation and Purification Technology*. 2016. 163: 39-47.
- Wang, N., Zhang, G., Wang, L., Li, J., An, Q., Ji, S. Pervaporation dehydration of acetic acid using NH2-UiO-66/PEI mixed matrix membranes. *Separation and Purification Technology*. 2017. 186: 20-7.
- 120. Chatterjee, S., De, S. Adsorptive removal of arsenic from groundwater using chemically treated iron ore slime incorporated mixed matrix hollow fiber membrane. *Separation and Purification Technology*. 2017. 179: 357-68.
- Lawrence Arockiasamy, D., Alhoshan, M., Alam, J., Muthumareeswaran, M. R., Figoli, A., Arun Kumar, S. Separation of proteins and antifouling properties of polyphenylsulfone based mixed matrix hollow fiber membranes. *Separation and Purification Technology*. 2017. 174: 529-43.
- 122. Mukhtar, N. H., Mamat, N. A., See, H. H. Monitoring of tobramycin in human plasma via mixed matrix membrane extraction prior to capillary electrophoresis with contactless conductivity detection. *Journal of Pharmaceutical and Biomedical Analysis*. 2018. 158: 184-8.
- 123. Ahmadiannamini, P., Eswaranandam, S., Wickramasinghe, R., Qian, X. Mixed-matrix membranes for efficient ammonium removal from wastewaters. *Journal of Membrane Science*. 2017. 526: 147-55.
- 124. Xu, S., Zhang, H., Yu, F., Zhao, X., Wang, Y. Enhanced ethanol recovery of PDMS mixed matrix membranes with hydrophobically modified ZIF-90. *Separation and Purification Technology*. 2018. 206: 80-9.
- 125. Davood Abadi Farahani, M. H., Hua, D., Chung, T.-S. Cross-linked mixed matrix membranes (MMMs) consisting of amine-functionalized multi-walled carbon nanotubes and P84 polyimide for organic solvent nanofiltration (OSN) with enhanced flux. *Journal of Membrane Science*. 2018. 548: 319-31.

- 126. White, R. J., Luque, R., Budarin, V. L., Clark, J. H., Macquarrie, D. J. Supported metal nanoparticles on porous materials. Methods and applications. *Chemical Society reviews*. 2009. 38(2): 481-94.
- 127. Iijima, S. Helical microtubules of graphitic carbon. *Nature*. 1991. 354(6348): 56-8.
- 128. Pan, C., Xu, S., Zou, H., Guo, Z., Zhang, Y., Guo, B. Carbon nanotubes as adsorbent of solid-phase extraction and matrix for laser desorption/ionization mass spectrometry. *Journal of the American Society for Mass Spectrometry*. 2005. 16(2): 263-70.
- 129. Song, X.-Y., Chen, J., Shi, Y.-P. Different configurations of carbon nanotubes reinforced solid-phase microextraction techniques and their applications in the environmental analysis. *TrAC Trends in Analytical Chemistry*. 2017. 86: 263-75.
- Tian, J., Xu, J., Zhu, F., Lu, T., Su, C., Ouyang, G. Application of nanomaterials in sample preparation. *Journal of Chromatography A*. 2013. 1300: 2-16.
- 131. Wang, N., Xin, H., Zhang, Q., Jiang, Y., Wang, X., Shou, D., et al. Carbon nanotube-polymer composite for effervescent pipette tip solid phase microextraction of alkaloids and flavonoids from Epimedii herba in biological samples. *Talanta*. 2017. 162: 10-8.
- 132. Wang, S.-L., Pang, X.-Q., Cao, J., Cao, W., Xu, J.-J., Zhu, Q.-Y., et al. Effervescence and graphitized multi-walled carbon nanotubes assisted microextraction for natural antioxidants by ultra high performance liquid chromatography with electrochemical detection and quadrupole time-of-flight tandem mass spectrometry. *Journal of Chromatography A*. 2015. 1418: 12-20.
- 133. Abujaber, F., Ahmad, S. M., Neng, N. R., Rodríguez Martín-Doimeadios, R. C., Guzmán Bernardo, F. J., Nogueira, J. M. F. Bar adsorptive microextraction coated with multi-walled carbon nanotube phases Application for trace analysis of pharmaceuticals in environmental waters. *Journal of Chromatography A*. 2019. 1600: 17-22.
- Madani, S. Y., Mandel, A., Seifalian, A. M. A concise review of carbon nanotube's toxicology. *Nano Rev.* 2013. 4.

- 135. Stafiej, A., Biesaga, M. Sorption behavior of acidic herbicides on carbon nanotubes. *Microchimica Acta*. 2007. 159: 293-8.
- 136. Rissato, S. R., Galhiane, M. S., Apon, B. M., Arruda, M. S. P. Multiresidue Analysis of Pesticides in Soil by Supercritical Fluid Extraction/Gas Chromatography with Electron-Capture Detection and Confirmation by Gas Chromatography–Mass Spectrometry. *Journal of Agricultural and Food Chemistry*. 2005. 53(1): 62-9.
- 137. Xu, L., Feng, J., Liang, X., Li, J., Jiang, S. C18 functionalized graphene oxide as a novel coating for solid-phase microextraction. *Journal of Separation Science*. 2012. 35(12): 1531-7.
- Li, S., Lu, C., Zhu, F., Jiang, R., Ouyang, G. Preparation of C18 composite solid-phase microextraction fiber and its application to the determination of organochlorine pesticides in water samples. *Analytica Chimica Acta*. 2015. 873: 57-62.
- Gentili, A. Determination of non-steroidal anti-inflammatory drugs in environmental samples by chromatographic and electrophoretic techniques. *Analytical and bioanalytical chemistry*. 2007. 387(4): 1185-202.
- Manzo, V., Honda, L., Navarro, O., Ascar, L., Richter, P. Microextraction of non-steroidal anti-inflammatory drugs from waste water samples by rotatingdisk sorptive extraction. *Talanta*. 2014. 128: 486-92.
- Richardson, S. D., Ternes, T. A. Water Analysis: Emerging Contaminants and Current Issues. *Analytical Chemistry*. 2018. 90(1): 398-428.
- 142. Grossmann, K. Mode of action of auxin herbicides: a new ending to a long, drawn out story. *Trends in Plant Science*. 2000. 5(12): 506-8.
- 143. Melwanki, M. B., Fuh, M.-R. Partitioned dispersive liquid–liquid microextraction: An approach for polar organic compounds extraction from aqueous samples. *Journal of Chromatography A*. 2008. 1207(1): 24-8.
- 144. Ranz, A., Korpecka, J., Lankmayr, E. Optimized derivatization of acidic herbicides with trimethylsilyldiazomethane for GC analysis. *Journal of Separation Science*. 2008. 31(4): 746-52.
- Söderholm, S. L., Damm, M., Kappe, C. O. Microwave-assisted derivatization procedures for gas chromatography/mass spectrometry analysis. *Molecular Diversity*. 2010. 14(4): 869-88.

- 146. Catalina, M. I., Dallüge, J., Vreuls, R. J., Brinkman, U. A. Determination of chlorophenoxy acid herbicides in water by in situ esterification followed by in-vial liquid-liquid extraction combined with large-volume on-column injection and gas chromatography-mass spectrometry. *J Chromatogr A*. 2000. 877(1-2): 153-66.
- 147. Ji, Z., Cheng, J., Song, C., Hu, N., Zhou, W., Suo, Y., et al. A highly sensitive and selective method for determination of phenoxy carboxylic acids from environmental water samples by dispersive solid-phase extraction coupled with ultra high performance liquid chromatography-tandem mass spectrometry. *Talanta*. 2019. 191: 313-23.
- Pei, M., Shi, X., Wu, J., Huang, X. Graphene reinforced multiple monolithic fiber solid-phase microextraction of phenoxyacetic acid herbicides in complex samples. *Talanta*. 2019. 191: 257-64.
- 149. Mamat, N. A., See, H. H. Simultaneous electromembrane extraction of cationic and anionic herbicides across hollow polymer inclusion membranes with a bubbleless electrode. *Journal of Chromatography A*. 2017. 1504: 9-16.
- 150. Valimaña-Traverso, J., Morante-Zarcero, S., Pérez-Quintanilla, D., García, M. Á., Sierra, I., Marina, M. L. Cationic amine-bridged periodic mesoporous organosilica materials for off-line solid-phase extraction of phenoxy acid herbicides from water samples prior to their simultaneous enantiomeric determination by capillary electrophoresis. *Journal of Chromatography A*. 2018. 1566: 146-57.
- Barbosa, M. O., Moreira, N. F. F., Ribeiro, A. R., Pereira, M. F. R., Silva, A. M. T. Occurrence and removal of organic micropollutants: An overview of the watch list of EU Decision 2015/495. *Water Research*. 2016. 94: 257-79.
- Sousa, J. C. G., Ribeiro, A. R., Barbosa, M. O., Pereira, M. F. R., Silva, A. M. T. A review on environmental monitoring of water organic pollutants identified by EU guidelines. *Journal of Hazardous Materials*. 2018. 344: 146-62.
- 153. Swain, M. chemicalize.org. *Journal of Chemical Information and Modeling*. 2012. 52(2): 613-5.
- 154. Gardner, J. S., Walker, J. O., Lamb, J. D. Permeability and durability effects of cellulose polymer variation in polymer inclusion membranes. *Journal of Membrane Science*. 2004. 229(1): 87-93.

- 155. Li, N., Chen, J., Shi, Y.-P. Magnetic polyethyleneimine functionalized reduced graphene oxide as a novel magnetic sorbent for the separation of polar non-steroidal anti-inflammatory drugs in waters. *Talanta*. 2019. 191: 526-34.
- 156. Ye, Z., Gao, H. Evaluation of sample extraction methods for minimizing hematocrit effect on whole blood analysis with volumetric absorptive microsampling. *Bioanalysis*. 2017. 9(4): 349-57.
- 157. Ferrone, V., Carlucci, M., Ettorre, V., Cotellese, R., Palumbo, P., Fontana, A., et al. Dispersive magnetic solid phase extraction exploiting magnetic graphene nanocomposite coupled with UHPLC-PDA for simultaneous determination of NSAIDs in human plasma and urine. *Journal of Pharmaceutical and Biomedical Analysis*. 2018. 161: 280-8.
- 158. Abd Wahib, S. M., Wan Ibrahim, W. A., Sanagi, M. M., Kamboh, M. A., Abdul Keyon, A. S. Magnetic sporopollenin-cyanopropyltriethoxysilanedispersive micro-solid phase extraction coupled with high performance liquid chromatography for the determination of selected non-steroidal antiinflammatory drugs in water samples. *Journal of Chromatography A*. 2018. 1532: 50-7.
- 159. Racamonde, I., Rodil, R., Quintana, J. B., Sieira, B. J., Kabir, A., Furton, K. G., et al. Fabric phase sorptive extraction: A new sorptive microextraction technique for the determination of non-steroidal anti-inflammatory drugs from environmental water samples. *Analytica Chimica Acta*. 2015. 865: 22-30.
- 160. Loh, S. H., Sanagi, M. M., Wan Ibrahim, W. A., Hasan, M. N. Solventimpregnated agarose gel liquid phase microextraction of polycyclic aromatic hydrocarbons in water. *Journal of Chromatography A*. 2013. 1302: 14-9.
- Balchen, M., Gjelstad, A., Rasmussen, K. E., Pedersen-Bjergaard, S. Electrokinetic migration of acidic drugs across a supported liquid membrane. *Journal of Chromatography A*. 2007. 1152(1): 220-5.
- 162. Tabani, H., Fakhari, A. R., Shahsavani, A., Behbahani, M., Salarian, M., Bagheri, A., et al. Combination of graphene oxide-based solid phase extraction and electro membrane extraction for the preconcentration of chlorophenoxy acid herbicides in environmental samples. *Journal of Chromatography A*. 2013. 1300: 227-35.

163. See, H. H., Hauser, P. C. Automated Electric-Field-Driven Membrane Extraction System Coupled to Liquid Chromatography–Mass Spectrometry. *Analytical Chemistry*. 2014. 86(17): 8665-70.

LIST OF PUBLICATIONS

- [1] Ganesan, T., Lim, H. N., See, H. H. Automated Mixed Matrix Membrane Microextraction Prior to Liquid Chromatography for the Determination of Chlorophenoxy Acid Herbicides in Sewage Water Samples. *Chromatographia*. 2020. 83(4): 497-505.
- [2] Ganesan, T., Mukhtar, N., Lim, H., See, H. H. Mixed Matrix Membrane Tip Extraction Coupled with UPLC–MS/MS for the Monitoring of Nonsteroidal Anti-Inflammatory Drugs in Water Samples. *Separations*. 2020. 7: 19.