

VITAMIN D LOADED ELECTROSPUN CELLULOSE
ACETATE/POLYCAPROLACTONE NANOFIBERS FOR DRUG
DELIVERY APPLICATIONS

MOHAMMED AHMAD WSOO

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy

Bioscience Department
Faculty of Science
Universiti Teknologi Malaysia

AUGUST 2021

DEDICATION

This thesis is dedicated to my father, who taught me that the best kind of knowledge to have is that which is learned for its own sake. It is also dedicated to my mother, who taught me that even the largest task can be accomplished if it is done one step at a time.

ACKNOWLEDGEMENT

In the name of Allah, the most gracious and the most merciful, all praise to Allah the Almighty for giving me the strength, guidance and patience to complete this thesis.

I would like to express my genuine gratitude to my main thesis supervisor, Dr. Saiful Izwan Bin Dato Abd Razak, for his patience, guidance, encouragement, and support throughout my research. I am also very thankful to my co-supervisors, Associate Professor Dr. Shafinaz Binti Shahir and Dr. Siti Paulienah Binti Mohd Bohari, for their support.

I am deeply grateful to all my colleagues and others who have provided assistance and infinite knowledge; Mr. Hafedh, Mr. Rabiu, and Ms. Khalida. I would also like to thank the laboratory technicians who have assisted me greatly in my research.

I would also like to express my heartfelt gratitude to my close friend Mr. Omer Ahmad Pirot, for the continuous support me during my study.

I would like to express my special appreciation to my lovely family for their love and support during these years. Words cannot express how thankful to my beloved wife, Vinos Aziz Hamad, and my children (Varya, Varin, and Bahand) for their patience and help.

ABSTRACT

Vitamin D is a steroid hormone that plays a crucial role in regulating physiological functions in the human body. Its supplements have been used to prevent and treat vitamin D deficiency. Although many approaches are currently available for delivering vitamin D, the low bioavailability and loss of bioactivity of vitamin D remains a challenging task. Therefore, this study aims to introduce a new implantable drug delivery system (IDDS) for delivering vitamin D. An IDDS offers many advantages over other routes of drug administration due to direct delivery into the body. The IDDS was developed from the electrospun cellulose acetate (CA) and polycaprolactone (PCL) nanofibrous membrane, in which the core of the IDDS consisted of vitamin D₃-loaded CA nanofiber (CAVD) and was enclosed in a thin layer of the sintered PCL membrane (CAVD/PCL). The morphological surface and physicochemical properties of the produced electrospun nanofiber of the vitamin D₃-loaded CA and PCL membranes were characterized using a scanning electron microscope (SEM), attenuated total reflection Fourier transform infrared spectroscopy (ATR-FTIR), X-ray diffraction (XRD), and differential scanning calorimetry (DSC). The vitamin D loading efficiency and vitamin D stability were characterized by high-performance liquid chromatography (HPLC) and UV-Visible spectroscopy. Mechanical properties, drug release studies and *in vitro* cytotoxicity studies were also performed in this study. Vitamin D₃ in three different concentrations, including 6, 12, and 20% (w/w) based on the weight of the CA polymer, was efficiently loaded into the CA nanofibrous membrane using electrospinning. The surface morphologies of CA nanofiber and vitamin D₃-loaded CA nanofiber were smooth and bead-free, while their average diameters increased from 324 nm to 428 nm when the weight ratios of vitamin D₃ were increased. The results from HPLC and UV spectra showed that the vitamin D₃ compound in CA nanofiber has a stable structure and did not degrade during electrospinning incorporation. The drug release study and tensile testing showed that the PCL membrane as the outer layer around the core's implants plays a crucial role in the improved mechanical properties and kinetic drug release. The Young modulus and tensile strength of CAVD/PCL were significantly increased as compared to CAVD. The kinetic drug release of CAVD followed the first-order model and converted to the zero-order model in the CAVD/PCL at the first stage of the drug release. The CA and PCL nanofibers are non-cytotoxic based on the results of *in vitro* cytotoxicity studies. In conclusion, based on the outcomes and methods outlined in the present study, the vitamin D-loaded CA nanofibrous membrane and the PCL nanofibrous membrane are suitable for developing an IDDS for delivery of vitamin D.

ABSTRAK

Vitamin D adalah hormon steroid yang memainkan peranan penting dalam mengatur fungsi fisiologi dalam tubuh manusia. Pengambilan vitamin D sebagai sumber makanan tambahan boleh mencegah kekurangan vitamin D. Walaupun banyak pendekatan yang ada setakat ini untuk menyampaikan vitamin D, bioketersediaan yang rendah dan kehilangan bioaktiviti vitamin D tetap menjadi isu utama. Oleh itu, kajian ini bertujuan untuk memperkenalkan sistem penyampaian ubat implan yang baharu (IDDS) untuk penyampaian vitamin D. Kaedah baru (IDDS) mempunyai banyak kelebihan berbanding kaedah penyampaian ubatan yang lain kerana penyampaiannya adalah secara langsung ke dalam badan. IDDS dihasilkan daripada membran serat nano elektrospun selulosa asetat (CA) dan polikaprolakton (PCL), di mana teras IDDS terdiri daripada serat nano CA (CAVD) yang dimuatkan vitamin D₃ dan dilapisi lapisan nipis membran PCL yang disinter (CAVD/PCL). Membran serat nano PCL dijadikan kapsul pada teras melalui proses sintering termal. Pencirian morfologi permukaan dan sifat fisikokimia membran CA dan PCL yang dimuatkan vitamin D₃ telah di analisa menggunakan mikroskop elektron imbasan (SEM), spektroskopi inframerah Fourier transformasi (ATR-FTIR), difraksi sinar-X (XRD) dan kalorimetri pengimbasan pembezaan (DSC). Tahap keefisienan dan kestabilan pemuatan ubat telah dianalisis menggunakan Kromatografi Cecair Prestasi Tinggi (HPLC) dan spektroskopi UV. Analisis mekanikal, kajian pelepasan ubat, dan kajian kesitotoksikan *in vitro* juga dilakukan. Vitamin D₃ dalam tiga kepekatan yang berbeza merangkumi 6, 12, dan 20% (b/b) berdasarkan berat polimer CA, telah dimasukkan ke dalam membran nanofiber CA melalui proses elektrospinning. Morfologi permukaan dari pempentukan CA dan serat nano CA yang dimuatkan dengan Vitamin D₃ adalah halus dan bebas manik, sementara purata diameter nanofiber meningkat dari 324 nm kepada 428 nm apabila nisbah berat vitamin D₃ meningkat. Keputusan spektrum HPLC dan UV menunjukkan sebatian vitamin D₃ dalam CA mempunyai struktur yang stabil dan tidak berkurangan semasa proses penggabungan. Kajian pelepasan ubat dan kajian tegangan menunjukkan bahawa membran PCL sebagai lapisan luar implan teras memainkan peranan penting dalam meningkatkan sifat mekanikal dan pelepasan kinetik. Modulus Young dan kekuatan tegangan CAVD/PCL didapati meningkat dengan ketara berbanding dengan CAVD. Pelepasan kinetik CAVD mengikuti model susunan pertama dan ditukarkan ke model susunan sifar bagi CAVD / PCL pada peringkat pertama pelepasan ubat. Serat nano CA dan PCL bersifat tidak toksik berdasarkan hasil kajian kesitotoksikan secara *in vitro*. Kesimpulannya, berdasarkan hasil dan kaedah yang digariskan dalam kajian ini, membran serat nano CA yang dimuatkan dengan vitamin D dan membran serat nano PCL sesuai untuk menghasilkan IDDS untuk penyampaian vitamin D.

TABLE OF CONTENTS

	TITLE	PAGE
DECLARATION		iii
DEDICATION		iv
ACKNOWLEDGEMENT		v
ABSTRACT		vi
ABSTRAK		vii
TABLE OF CONTENTS		viii
LIST OF TABLES		xiv
LIST OF FIGURES		xvi
LIST OF ABBREVIATIONS		xx
LIST OF SYMBOLS		xxii
LIST OF APPENDICES		xxiv
CHAPTER 1 INTRODUCTION		1
1.1 Background of the Study		1
1.2 Problem Statement		3
1.3 Research Objectives		5
1.4 Scope of Research		6
1.5 Significance of the Study		7
1.6 Thesis Outline		7
CHAPTER 2 LITERATURE REVIEW		9
2.1 Drug Delivery System		9
2.1.1 Routes of Drug Administration		9
2.1.2 Implantable Drug Delivery System		12
2.1.3 Mechanism of Drug Release From IDDS		14
2.1.4 The Kinetic Models of Drug Release		15
2.2 Vitamin D		17
2.2.1 Synthesis and Metabolism of Vitamin D		19
2.2.2 Vitamin D Functions and Its Deficiency Factors		20

2.2.3 Drug Delivery System for Vitamin D	21
2.3 Nanofiber Production Using Electrospinning	25
2.4 Electrospinning	26
2.4.1 Types of Electrospinning	27
2.4.2 The Effect of Electrospinning Processing Parameters on Nanofiber production	28
2.4.2.1 Applied Voltage	29
2.4.2.2 Flow Rate	30
2.4.2.3 Working Distance	30
2.4.3 The Effect of Solution Parameters on Nanofiber Production	31
2.4.3.1 Concentration of Polymer Solution	31
2.4.3.2 Conductivity of Polymer Solution	32
2.4.3.3 Solvent System	32
2.4.3.4 Surface Tension	33
2.5 Cellulose Acetate	33
2.5.1 Synthesis and Structure of CA	34
2.5.2 Thermal Properties of CA Nanofiber	35
2.5.3 Tensile Properties of CA Nanofiber	37
2.5.4 <i>In-Vitro</i> Biodegradability Study of CA Nanofibers	37
2.5.5 <i>In-Vitro</i> Biocompatibility Study of CA Nanofiber	38
2.5.6 Application of Electrospun CA Nanofiber in the Drug Delivery System	39
2.6 Polycaprolactone	43
2.6.1 Biodegradability Properties of PCL	44
2.6.2 <i>In-Vitro</i> Biocompatibility Study of PCL Nanofiber	45
2.6.3 Thermal Properties of PCL Nanofiber	45
2.6.4 Tensile Properties of PCL Nanofiber	46
2.6.5 Application of Electrospun PCL Nanofiber in Drug Delivery System	47
2.7 Co-Electrospinning of CA/PCL Fibrous Membrane	49
2.8 Multi-Layered Electrospun Nanofibrous Structure	50
2.9 Implanted Electrospun Nanofiber as Drug Carrier	51

2.10	Summary of Literature Review	52
CHAPTER 3	MATERIALS AND METHODS	53
3.1	Materials	53
3.2	Optimization of Electrospinning Process Parameters & Polymer Solution Properties for Producing CA and PCL Nanofiber	55
3.2.1	Preparation of Cellulose Acetate Solutions	55
3.2.1.1	Electrospinning of Cellulose Acetate Solutions	56
3.2.2	Preparation of Polycaprolactone Solutions	57
3.2.2.1	Electrospinning of Polycaprolactone Solutions	57
3.3	Loading Vitamin D ₃ into Cellulose Acetate Fibrous Membrane	57
3.3.1	Preparation of Blended Cellulose Acetate/Vitamin D ₃ Solutions	57
3.3.2	Fabrication of CA and Blended CA/Vitamin D ₃ Solutions	58
3.4	Preparation of PCL Spinning Solution	59
3.4.1	Fabrication of PCL Nanofiber Membrane by Electrospinning	59
3.5	Design Implantable Drug Delivery System	60
3.5.1	Formulation Rod-Shaped Vitamin D ₃ -Loaded CA Nanofiber (CAVD)	60
3.5.2	Formulation Rod-Shaped Vitamin D ₃ -Loaded Cellulose Acetate/Polycaprolactone (CAVD/PCL)	61
3.6	Characterization	63
3.6.1	Viscosity and Conductivity Measurement	63
3.6.2	Morphological Characterization	63
3.6.3	Fourier Transform Infrared (FT-IR) Spectroscopy	63
3.6.4	X-Ray Diffractometer	64
3.6.5	Thermal Characterization	64
3.6.6	Tensile Properties	65

3.6.7 Surface Wettability Characterization	66
3.6.8 Degree of Swelling and Weight Loss	66
3.6.9 Stability of Vitamin D ₃ after Loaded CA Nanofiber	67
3.7 <i>In-Vitro</i> Drug Release Study	68
3.7.1 Actual Vitamin D ₃ Content	68
3.7.2 Preparation of Releasing Medium	69
3.7.3 <i>In-Vitro</i> Vitamin D ₃ Release Study	69
3.7.4 Determination of Vitamin D ₃	70
3.7.5 Kinetic of Drug Release Study	71
3.8 <i>In-Vitro</i> Biocompatibility Study	72
3.8.1 Cell Culture	72
3.8.2 Sterilization of Electrospun Nanofibers	73
3.8.3 Optimization of Cell Number	73
3.8.4 MTS Assay	73
3.8.5 Dye Exclusion Method (Trypan Blue)	74
3.8.6 Cell Adhesion Study	74
3.9 Statistical Analysis	75
3.10 Flow Chart	75
CHAPTER 4 OPTIMIZATION OF SOLVENTS AND PROCESS PARAMETERS	77
4.1 Introduction	77
4.2 Results and Discussion	77
4.2.1 Optimization of CA Nanofiber Production Using Electrospinning	77
4.2.1.1 Morphological Analysis of Cellulose Acetate Nanofiber Using Acetone/DMAc	80
4.2.1.2 Morphological Analysis of Cellulose Acetate Nanofiber Using Acetone/Ethanol/DMAc (3:1:2)	85
4.2.2 Optimization of Electrospun PCL Nanofiber Production Using Electrospinning	88
4.3 Summary of Optimization	91

CHAPTER 5	VITAMIN D₃-LOADED CA-NANOFIBER AND PCL-NANOFIBER PRODUCTION AND THEIR CHARACTERIZATIONS	93
5.1	Introduction	93
5.2	Results and Discussion	93
5.2.1	Vitamin D ₃ -Loaded CA Fibrous Membrane	93
5.2.1.1	Morphological Surface Characterization	94
5.2.1.2	Drug Loading Efficiency	97
5.2.1.3	ATR-FTIR Characterization	98
5.2.1.4	X-Ray Diffraction Characterization	100
5.2.1.5	Thermal Properties	101
5.2.1.6	Stability of Vitamin D ₃ after Loaded CA Fibrous Membrane	105
5.2.1.7	Surface-Wetting Characteristic	107
5.2.2	PCL Nanofibrous Mat and Sintered PCL Membrane	109
5.2.2.1	Morphological Surface Characterization	110
5.2.2.2	ATR-FTIR Characterization	111
5.2.2.3	X-Ray Diffraction Characterization	112
5.2.2.4	Thermal Properties	113
5.2.2.5	Surface-Wetting Characteristic	115
5.3	Summary	117
CHAPTER 6	STUDY OF IMPLANTABLE DRUG DELIVERY SYSTEM	119
6.1	Introduction	119
6.2	Results and Discussion	119
6.2.1	Rod-shaped Vitamin D ₃ -Loaded Cellulose Acetate/Polycaprolactone (CAVD/PCL)	119
6.2.1.1	Characterization of the CAVD/PCL	121
6.2.2	Rod-shaped Vitamin D ₃ -Loaded Cellulose Acetate (CAVD)	123
6.2.3	Tensile Properties	125
6.2.4	Degree of Swelling	129

6.2.5 Degree of Weight Loss	130
6.2.6 <i>In-Vitro</i> Drug Release Study	132
6.2.6.1 Drug Release Study of CAVD	132
6.2.6.2 Kinetic Drug Release of the CAVD	134
6.2.6.3 Drug Release Study of CAVD/PCL	135
6.2.6.4 Kinetic Drug Release of the CAVD/PCL	138
6.2.7 <i>In-Vitro</i> Cytotoxicity Study	139
6.2.7.1 Cell optimization	139
6.2.7.2 MTS Assay	140
6.2.7.3 Cell Viability Testing Using Trypan Blue Dye Exclusion Method	142
6.2.7.4 Cell Adhesion Study	143
6.3 Summary of Study IDDS	146
CHAPTER 7 CONCLUSIONS AND RECOMMENDATIONS	149
7.1 Conclusion	149
7.2 Recommendations for Future Work	150
REFERENCES	153
LIST OF PUBLICATIONS	183

LIST OF TABLES

TABLE NO.	TITLE	PAGE
Table 2.1	Route of administration: advantages and disadvantages (Jain, 2020)	10
Table 2.2	Various vitamin D delivery systems	22
Table 2.3	Glass transition temperature, melting temperature, and decomposition temperature of CA at different degrees of substitution (Bao, 2015)	36
Table 2.4	Drugs loaded in electrospun cellulose acetate nanofibers and its application in drug delivery systems	41
Table 2.5	Application of electrospun PCL nanofibers in drug delivery systems	48
Table 3.1	List of materials and reagents used in the present research	54
Table 3.2	Cellulose acetate polymer solutions are prepared in different solvent systems, and spun under various electrospinning process parameters	56
Table 3.3	CA and blended CA/vitamin D ₃ solutions and their electrospinning process parameters	59
Table 4.1	Viscosity and conductivity of CA1 solution and its electrospun process parameters	80
Table 4.2	Viscosity and conductivity of CA2 solution and its electrospun process parameters	82
Table 4.3	Viscosity and conductivity of CA ₃ solution and its electrospun process parameters	85
Table 4.4	Conductivity and viscosity properties of the PCL solutions and the process parameters used to spin the PCL polymer solution	88
Table 5.1	Viscosity and conductivity of pure CA and blended CA/vitamin D ₃ solutions at different shear rate (n=3)	96
Table 5.2	DSC analysis data of the CANF7 and vitamin D ₃ -loaded CANF	104
Table 5.3	Wettability of the surface of CANF7 and vitamin D ₃ -loaded CA nanofibrous mats	109

Table 6.1	Detail information on the rod-shaped CAVD/PCL	120
Table 6.2	Detail information on the rod-shaped CAVD	123
Table 6.3	Tensile properties of CANF, CAVD, CAVD/PCL under dry and wet conditions	126
Table 6.4	Results of fitting the experimental data with four kinetic models	135
Table 6.5	Results of fitting the experimental data with four kinetic models	139

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
Figure 2.1	Structure of skin layers and adipocytes	13
Figure 2.2	Structure of the 7-dehydrocholesterol, Ergocalciferol (vitamin D ₂), Cholecalciferol (vitamin D ₃), and Calciferol (active form)	18
Figure 2.3	Synthesis and Metabolism of vitamin Ds	19
Figure 2.4	Risk factors of vitamin D deficiency (Amrein et al., 2020)	20
Figure 2.5	Basic electrospinning setup	26
Figure 2.6	Types of electrospinning: (A) blend electrospinning, (B) emulsion electrospinning, and (C) coaxial electrospinning	28
Figure 2.7	Average electrospun nanofiber diameters (nm) at different applied voltages (kV) (Wei et al., 2019)	29
Figure 2.8	SEM images of evolution of the fiber production with different concentrations from low to high using electrospinning. Reprinted from (Fong et al., 1999), with permission from Elsevier.	31
Figure 2.9	Schematic preparation of cellulose acetate from cellulose by acetylation	35
Figure 2.10	Synthesis of PCL from caprolactone	43
Figure 2.11	Biodegradation of PCL in the human body	44
Figure 3.1	Schematic of the formulation implantable drug delivery systems (A) rod-shaped CAVD, (B) rod-shaped CAVD/PCL	62
Figure 3.2	Standard Curve of vitamin D ₃	70
Figure 3.3	Flow chart of the working study	76
Figure 4.1	17% of CA1, CA2, and CA3 polymer solutions	78

Figure 4.2	SEM images and their nanofiber diameter distribution of (A) CANF1, (B) CANF2, and (C) CANF3	81
Figure 4.3	SEM images and their nanofiber diameter distribution of (A) CANF4, (B) CANF5, and (C) CANF6	83
Figure 4.4	SEM images and their nanofiber diameter distribution of (A) CANF7, (B) CANF8, (C) CANF9, (D) CANF10	86
Figure 4.5	SEM images and fiber diameter distribution (A) PCLNF1 (B) PCLNF2, and (C) PCLNF3	89
Figure 5.1	FESEM images and fiber diameter distributions of (A) Pure CANF7, (B) 6% of vitamin D ₃ -loaded CA nanofiber, (C) 12% of vitamin D ₃ -loaded CA nanofiber, and (D) 20% of vitamin D ₃ -loaded CA nanofiber	95
Figure 5.2	FTIR spectra of the (A) CANF7, (B) CAVD20, (C) CAVD12, (D) CAVD6, and (E) Vitamin D ₃	98
Figure 5.3	XRD diffractograms of the (A) Pure vitamin D ₃ powder, (B) CANF7, and (C) vitamin D ₃ -loaded CA nanofibrous membrane	100
Figure 5.4	DSC thermal curves of (A) heating-cooling-heating scans of the vitamin D ₃ ; (B) first heating cycle, (C) cooling cycle, and (D) second heating cycle of the CANF7, and vitamin D ₃ -loaded CA nanofibers	103
Figure 5.5	HPLC-PAD chromatograms of (A) vitamin D ₃ powder dissolved in methanol (1µg/mL), vitamin D ₃ extracted from (B) CAVD20, (C) CAVD12 by methanol	105
Figure 5.6	UV spectra of CANF7, vitamin D ₃ powder (2mg/mL), CAVD12, and CAVD20 dissolved in acetone	106
Figure 5.7	Water contact angle of (A) CANF7, (B) CAVD6, (C) CAVD12, and (D) CAVD20 at two different times	108
Figure 5.8	SEM images and fiber diameter distributions of (A) PCLNF1 at a magnification of 5000x and (B, C) sintered PCL membrane at a magnification of 200x and 15000x, respectively.	110
Figure 5.9	FTIR spectra of the PCLNF1 and sintered PCL membrane	112

Figure 5.10	XRD diffraction patterns of PCLNF1 and sintered PCL membrane	113
Figure 5.11	DSC thermal curves of (A) first heating cycle, (B) cooling cycle, and (C) second heating cycle of the PCLNF1, and sintered PCL	114
Figure 5.12	Water contact angle of PCLNF1 and sintering PCL membrane	116
Figure 6.1	An image showing the size and shape of (A) CAVD6/PCL (1), CAVD12/PCL (2), and CAVD20/PCL (3); (B) schematic representation of reservoir drug delivery system; (C) their cross-sectional SEM image	121
Figure 6.2	FTIR spectra of CANF alone and CANF at the cross-section area of CA/PCL layers	122
Figure 6.3	FESEM images at the cross-section area of CA/PCL layers	123
Figure 6.4	(A) shape and the morphological surface of the CAVD6, CAVD12, and CAVD20, and (B) schematic representation of matrix drug delivery system	124
Figure 6.5	Stress-strain curves of the rod-shaped pure CANF, vitamin D ₃ loaded CA (CAVD), vitamin D ₃ loaded CA/PCL (CAVD/PCL) under dry and wet conditions	125
Figure 6.6	Process of inserting CAVD/PCL into sub-dermal pig's skin and withdrawn implants after immersed in PBS for 30 days.	128
Figure 6.7	Swelling behavior of the CANF, CAVD12, CAVD20, CAVD12/PCL, and CAVD20/PCL in the phosphate buffer at 37 °C.	129
Figure 6.8	Weight loss (%) of the CANF, CAVD12, CAVD20, CAVD12/PCL, and CAVD20/PCL in the phosphate buffer at 37°C, * p < 0.05, ** p < 0.01.	131
Figure 6.9	Cumulative vitamin D ₃ release profile of (A) CAVD6, CAVD12, and CAVD20; and (B) its linear regression of fitting experimental data to the first-order model at 24h	133
Figure 6.10	Fitting curves of drug release from the CAVD with (A) Zero-order model, (B) First-order model, (C) Higuchi model, and (D) Korsmeyer-Peppa model	134

Figure 6.11	Cumulative vitamin D ₃ release profile of (A) CAVD20/PCL, CAVD12/PCL, and CAVD6/PCL, (B) its linear regression of fitting experimental data to the zero-order model at five days	137
Figure 6.12	Fitting curves of drug release from the CAVD/PCL with (A) Zero-order model, (B) First-order model, (C) Higuchi model, and (D) Korsmeyer-Peppa model	138
Figure 6.13	HDF cell confluence for culture time of 7 days at the density of (A) 1000 cells/well, (B) 2500 cells/well, (C) 5000 cells/well, and (D) 7500 cells/well	140
Figure 6.14	MTS assay results: histogram comparing the proliferation of HDF cells on the CANF, PCLNF, and sintered PCL membrane for cell culture periods of 3, 5, and 7 days. The data was provided as mean ± SD (n=3). ** p < 0.01 (obtained by one-way ANOVA, Tukey HSD test).	141
Figure 6.15	Cell viability percentage of the HDF cell study by cell counting using trypan blue dye exclusion method (data represented as the mean ± SD of n = 4).	143
Figure 6.16	FESEM images of HDF adhered and proliferated on the surfaces of sintered PCL membranes at cultured times of (A) 3-days, (B) 5-days, and (C) 7-days; with the magnification value of 1000 x and 30,000 x	144
Figure 6.17	FESEM images of HDF adhered and proliferated on the surfaces of CA nanofibrous membranes at cultured times of (A) 3-days, (B) 5-days, and (C) 7-days; with the magnification value of 1000 x and 10,000 x	145

LIST OF ABBREVIATIONS

AC	-	Acetone
CA	-	Cellulose Acetate
CANF	-	Cellulose acetate nanofiber
CAVD	-	Cellulose acetate loaded with vitamin D
CAVD/PCL	-	Cellulose acetate loaded with vitamin D/ Polycaprolactone
DDS	-	Drug delivery system
DCM	-	Dichloromethane
DMAc	-	N,N-dimethylacetamide
DMEM		Dulbecco's Modified Eagle Medium
DMF	-	N,N-dimethylformamide
EtOH	-	Ethanol
FDA	-	Food and Drug Administration
FESEM	-	Field emission scanning electron microscop
HDF	-	Human dermal fibroblast
M.W	-	Molecular weight
MTS	-	(3-(4, 5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium) assay
NF	-	Nanofiber
PBS	-	Phosphate buffered saline
PCL	-	Polycaprolactone
SEM	-	Scanning electron microscope
Vit.D ₃	-	Vitamin D ₃
VD	-	Vitamin D ₃
DS	-	Degree of substitution
¹³ C NMR	-	Carbon-13 (C13) nuclear magnetic resonance
XRD	-	X-Ray Diffractometer
FT-IR	-	Fourier Transform Infrared Spectroscopy
UV	-	Ultraviolet

LIST OF SYMBOLS

$^{\circ}\text{C}$	-	Degree Celsius
2Θ	-	Bragg angle
%	-	Percentage
T_v	-	Temperature of water evaporation
T_g	-	Glass transition temperature
T_d	-	Thermal decomposition
T_m	-	Melting point
E_t	-	Young's modulus
σ_m	-	Tensile strength
ϵ_b	-	Elongation at break
η	-	Viscosity
ΔH_m	-	Melting enthalpy
ΔH_v	-	Enthalpy of water vaporization
λ	-	Wave length
nm	-	Nanometre
X_m	-	Degree of crystallinity
kV	-	Kilovolt
h	-	Hour
cP	-	Centipoise
μS	-	Micro Siemens
MPa	-	Mega Pascal
Sec	-	Second (s)
g	-	Gram
mg	-	Milligram
mL	-	Millilitre
L	-	Litter
μm	-	Micrometre
S^{-1}	-	1/second
cm	-	Centimetre (s)
d	-	Diameter

Min	-	Minute (s)
t	-	Time
v/v	-	Volume per volume
w/v	-	Weight per volume
w/w	-	Weight per weight
wt %	-	Weight percentage
log	-	logarithm
J/g	-	Joule per gram

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
A	A review on the properties of electrospun cellulose acetate and its application in drug delivery systems: A new perspective	184
B	Vitamin D ₃ -loaded electrospun cellulose acetate/polycaprolactone nanofibers: Characterization, <i>in-vitro</i> drug release and cytotoxicity studies	185
C	Development of prolonged drug delivery system using electrospun cellulose acetate/polycaprolactone nanofibers: Future subcutaneous implantation	186

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Drug delivery systems (DDSs) are defined as formulations or devices that have been used to introduce pharmaceutical agents into the human or animal body to achieve a desirable therapeutic effect (Jain, 2020). Moreover, drug delivery is a drug carrier for effective therapeutic delivery of drugs into the body via various administration routes, such as the implantable drug delivery system (IDDS), oral drug delivery, injectable drug delivery, pulmonary drug delivery, and nasal drug delivery (Kohrs et al., 2019). IDDS offer many advantages over other routes of drug delivery. Most significantly, it reduces the dosage of medication needed for treatment, and enhances the medication's bioavailability due to the drug being directly delivered to the site of action, avoiding first-pass metabolism (Stewart et al., 2020). It can also improve the efficacy and safety of an administered drug by controlling the release rate, protecting the drug level, and reaching the target site selectively while avoiding non-target cells (Pons-Faudoa et al., 2019).

A relatively new system known as the nano-based delivery system (NDDS) uses nanoscale biomaterials to deliver therapeutic agents to specific sites in a controlled manner (Patra et al., 2018). Nanofibers are nanoscale biomaterials that are generally electrospun from a variety of polymers via one of the most flexible techniques known as electrospinning (Shahriar et al., 2019). Electrospinning has become an exciting approach to developing NDDSs due to its increased surface-to-mass ratio, ease of operation, and cost-effectiveness (Iacob et al., 2020; Torres-Martínez et al., 2018). Also, drug-loaded electrospun-polymer nanofibers could overcome several key challenges, such as low solubility of drugs, loading efficiency, and irregular drug-release behavior (Wsoo et al., 2020).

Vitamin D₃ (cholecalciferol) is a secosteroid hormone that is crucial in regulating physiological functions in the human body (Grant, 2020). It plays a meaningful role in reducing the risk of Coronavirus disease 2019 (COVID-19) and other acute respiratory tract infections (Ali, 2020; Ilie et al., 2020). Vitamin D supplements are widely used as an exogenous candidate for treatment or prevention of vitamin D deficiency via either oral administration or injectable drug delivery (Glowka et al., 2019). Vitamin D deficiency is a worldwide health problem in all age groups, especially in those living in countries with inadequate exposure to sunlight (Palacios & Gonzalez, 2014). Despite the fact that many approaches are currently available for delivering vitamin D, the low bioavailability and loss of bioactivity of vitamin D due to taken via oral administration remains a challenging task. Little is known about using electrospun-polymer nanofibers for the treatment of vitamin D deficiency. Therefore, this study used electrospun-polymer nanofibers in formulating IDDS or long-term delivery of vitamin D₃ for the treatment of vitamin D deficiency.

Cellulose acetate (CA) and polycaprolactone (PCL) were used as carrier polymers to produce electrospun-polymer nanofibers. In the current study, CA was chosen owing to its high solubility in organic solvents, renewability, affordability, and ease of mass production (Mehrabi et al., 2017). It is also biocompatible, semi-biodegradable, non-irritating, and non-toxic (Dos Santos et al., 2021; Wsoo et al., 2020). Meanwhile, PCL is more applicable for long-term IDDS due to its slow biodegradation, and the US Food and Drug Administration has approved PCL's biocompatibility (Rezk et al., 2019). PCL could also be fabricated into nanofibers using electrospinning (Sattary et al., 2019), and modified to become a capsule by thermally sintering to improve its long-term tensile properties. This attribute is particularly advantageous for controlling drug releases due to the reduced electrospun pore size and porosity (Chaparro et al., 2019; Nelson et al., 2014). In this study, CA nanofiber loaded with vitamin D₃ was used to develop the core of a rod-shaped IDDS, while PCL polymer was used to produce a rate-limiting membrane around the core. Up to date, no studies have been conducted to investigate the possibility of electrospun polymer nanofiber to produce an IDDS. Furthermore, the use of IDDS for delivery of vitamin D via implanted in subcutaneous tissue is expected to overcome low bioavailability and bioactivity of vitamin D due to vitamin D directly delivered into the site of action.

1.2 Problem Statement

Vitamin D supplements are widely used as an exogenous candidate for treatment or prevention of vitamin D deficiency. However, the low bioavailability and loss of bioactivity of vitamin D remains a challenging task (Glowka et al., 2019). Drug bioavailability is the fraction of active drugs that reach the site of action. The absolute availability of a drug may be determined by comparing the respective areas under the plasma concentration curves after oral and intravenous administration (Bhosle et al., 2017).

Today, vitamin D is mostly taken via either oral administration (McCullough et al., 2019) or subcutaneous injection (Gupta et al., 2017). The oral route of vitamin D administration is problematic for some reasons. First, vitamin D reduces some bioactivity when it reaches the stomach because it is sensitive to gastric acid (Hasanvand et al., 2018). Second, it also has low intestinal drug absorption because vitamin D is a hydrophobic molecule with limited solubility in the aqueous environment (Glowka et al., 2019). The third one related to the oral administration of vitamin D is first-pass metabolism, which is the phenomenon that occurs when a medication is taken orally, reaches the liver, and undergoes substantial biotransformation, resulting in a significant reduction in bioavailability, and it is linked to low vitamin D bioavailability in the body (Maurya & Aggarwal, 2017). As a result, low bioavailability and loss of bioactivity of vitamin D are critical challenges for oral administration (Glowka et al., 2019).

Subcutaneous injections (SC) may avoid some of the problems related to oral administration, such as first-pass metabolism, gastric acid degradation, and low GIT absorption. This route, however, is not without its drawbacks. The problem with SC revolves around the drug's dosage and frequency of administration. Vitamin D toxicity would become a major concern if it is administered in high doses at a low frequency, whereas needle phobia and a reduction in the patient's compliance would be the main limitations if it is administered in low doses but at a high frequency. It has been reported that levels of vitamin D₃ fluctuated in serum after a high single dose injection (Einarsdóttir et al., 2010). Another danger of high-dose vitamin D injections is

intoxication, which occurs when levels of vitamin D₃ plasma exceed 100 ng/ml, or 250 nmol/L (Jones, 2008).

Therefore, this study aims to improve the bioavailability and bioactivity of vitamin D in the body by introducing a new IDDS. IDDS is an alternative system that can achieve a therapeutic effect with lower concentrations of drugs. As a result, it minimizes side-effects whilst increasing patient compliance (Stewart et al., 2018). On the other hand, IDDS can overcome the issues associated with oral and SC delivery by site-specific implantation. The sites of specific implantation can bypass first-pass metabolism; stomach degradation; or poor solubility and distribution phase of oral administration, resulting in higher drug concentrations in targeted areas (Pons-Faudoa et al., 2019).

Drug-loaded electrospun-polymer nanofibers could overcome several key challenges as well, such as low solubility of drugs, loading efficiency, and irregular drug-release behaviour (Wsoo et al., 2020). Based on the aforementioned information, a rod-shaped IDDS is an ideal idea to deliver vitamin D via subcutaneous tissue. It was produced from CA nanofiber loaded with vitamin D₃ and an electrospun-PCL-nanofibrous membrane. In which, the core implants consist of vitamin D-loaded CA nanofibrous membrane, while PCL polymer was used to produce a rate-limiting membrane around the core. In addition, the IDDS was formulated as rod-shaped because implantable biomaterials with rounded edges are easier to implant and more biocompatible than those with sharp edges when they react with host cells at the implant site. According to Ratner (2015), when the sharp edges of an implant come into contact with tissue, undesirable reactions may occur. A mechanical mismatch between a hard biomaterial and soft tissue may damage or irritate the soft tissue (Ratner, 2015).

Regulating long-term drug release, long-term biodegradation and biocompatibility, and suitable tensile properties of IDDS are all important aspects to consider when designing an IDDS. In this respect, both CA (Wsoo et al., 2020) and PCL (Chaparro et al., 2019) would be suitable biopolymers for producing electrospun

nanofiber. However, before moving forward to mass production, the following questions will first need to be answered.

1. What are suitable solvent systems and process parameters to produce electrospun-CA and electrospun-PCL nanofibers?
2. Is electrospinning a good technique for loading vitamin D into electrospun-polymer nanofibers?
3. To what extent can the release of vitamin D₃ from the IDDS be regulated?
4. Do CA and PCL biopolymers have good biocompatibility to formulate an IDDS?

1.3 Research Objectives

This study aimed to develop a rod-shaped IDDS from the electrospun-CA-and electrospun-PCL-nanofibrous membrane, and the core of the IDDS would consist of CA nanofiber loaded with vitamin D (CAVD) and enclosed in a thin layer of the PCL membrane (CAVD/PCL). Therefore, this study aimed to achieve the following objectives:

1. To optimize the solvent and process parameters for the fabrication process of electrospun-CA and electrospun-PCL nanofibers using electrospinning method.
2. To characterize morphological, structural, stability, and physicochemical properties of electrospun-CA nanofiber, vitamin-loaded CA nanofiber, and electrospun-PCL nanofibers.
3. To study the influence of the outer layer PCL on drug release kinetics.
4. To study *in-vitro* cell cytotoxicity and cell viability of electrospun polymer nanofibers against human dermal fibroblast cell lines.

1.4 Scope of Research

The scope of the first objective was to optimize electrospinning conditions by studying the effects of solvent systems and process parameters on the production of smooth and beadless CA-and PCL-nanofibers for effectively loading vitamin D into the electrospun-CA-nanofiber. The optimization included using mixing two solvents as binary solvent systems, such as acetone/dimethylacetamide (DMAc) and dichloromethane (DCM)/dimethylformamide (DMF) at different ratios, as well as a ternary solvent system consisting of acetone, ethanol, and DMAc. Likewise, the optimization of process parameters included the applied voltage, flow rate, and working distance. The morphological surface of the produced CA-and PCL-nanofibrous membrane was characterized by Scanning Electron Microscopy (SEM).

The second objective was defined with the following scope: loading vitamin D₃ into the CA nanofiber. Then, Fourier Transform Infrared Spectroscopy (FT-IR), Differential Scanning Calorimetry (DSC), X-ray diffraction (XRD), and Field Emission Scanning Electron Microscopy (FESEM) were used to characterize the produced vitamin D-loaded CA nanofibrous membrane and PCL nanofibrous membrane. Formulation of two samples of rod-shaped IDDS such as CAVD and CAVD/PCL from vitamin D-loaded CA nanofiber membrane and sintered PCL membrane.

The third scope was conducted to investigate the effect of the outer layer sintered PCL membrane on the *in-vitro* kinetic drug release from CAVD/PCL compared to CAVD. The Franz diffusion cell was used for drug release studies.

The final scope was conducted to investigate electrospun-CA and electrospun-PCL nanofiber biological properties using an *in-vitro* cytotoxicity and cell viability assay against HDF cells using the MTS assay and Trypan Blue Exclusion Test.

1.5 Significance of the Study

In this study, the IDDS was designed as a rod-shaped entity for subcutaneous administration by using an applicator implant device that had already been used for Nexplanon implantation (Palomba et al., 2012; Pons-Faudoa et al., 2019). Subcutaneous tissue is an ideal location for implanting vitamin D-depot devices due to the high-fat content that promotes the release and storage of drugs in the body's adipose (fat) tissue for a longer time, and the vitamin would slowly spread through the body by the systemic circulation.

The subcutaneous tissue has already been determined as a lower reactive site for insertion of foreign materials and the probability of localizing inflammation (Kumar & Pillai, 2018). This system offers many advantages over other routes of drug delivery. In particular, it would efficiently carry vitamin D at lower doses while potentially reducing side effects and increasing the patient's compliance. Direct site-localized drug delivery would bypass gastrointestinal tract (GIT) absorption barriers, first-pass metabolism, and prevent gastrointestinal degradation, thereby improving the bioavailability of vitamin D. The level of vitamin D could be regulated in the therapeutic window by managing the drug release and sustained release, resulting in a reduced dosage therapeutic effect and avoiding a high concentration of drugs from being exposed to the systemic environment. It is expected the IDDS will treat vitamin D deficiency more effectively. It might also be used to carry other hydrophobic pharmaceutical substances in the future.

1.6 Thesis Outline

This thesis consists of 7 chapters. Chapter 1 outlines the research context, problem statement, objectives, scope, and significance of the study. Chapter 2 reviews the available literature on the drug delivery system, electrospinning technique and its process parameters, vitamin D, cellulose acetate, and PCL that are relevant to this study. The experimental procedure and characterization techniques are discussed in Chapter 3. Chapter 4-6 results and discusses the data generated from various tests.

Chapter 4 reports on the effect of process parameters and solvent systems on the production of CA nanofiber and PCL nanofiber membranes and their morphological surfaces. Chapter 5 reports on loading vitamin D into the CA fibrous membrane and characterization of vitamin D-loaded CA nanofibrous membrane and PCL fibrous membrane using various techniques. Chapter 6 studies *in-vitro* biocompatibility, *in-vitro* drug release, tensile properties, *in-vitro* swelling and degradation of the CAVD and CAVD/PCL. The final chapter concludes the study while providing suggestions for future work.

REFERENCE

- Abid, S., Hussain, T., Nazir, A., Zahir, A., & Khenoussi, N. (2019). A novel double-layered polymeric nanofiber-based dressing with controlled drug delivery for pain management in burn wounds. *Polymer Bulletin*, 1-25.
- Aboamera, N. M., Mohamed, A., Salama, A., Osman, T., & Khattab, A. (2019). Characterization and mechanical properties of electrospun cellulose acetate/graphene oxide composite nanofibers. *Mechanics of Advanced Materials and Structures*, 26(9), 765-769.
- Agarwal, A., Jeengar, A., Bhowmick, M., Samanta, K., Satyamurthy, P., D'Souza, C., & Vigneshwaran, N. (2016). Performance characteristics of electrospun cellulose acetate nanofiber mat embedded with Nano-ZnO/vitamins. *International Journal of Nanotechnology and Application*, 2277-4777.
- Agrahari, V., Agrahari, V., Meng, J., & Mitra, A. K. (2017). Chapter 9 - Electrospun Nanofibers in Drug Delivery: Fabrication, Advances, and Biomedical Applications. In A. K. Mitra, K. Cholkar, & A. Mandal (Eds.), *Emerging Nanotechnologies for Diagnostics, Drug Delivery and Medical Devices* (pp. 189-215). Elsevier.
- Ahmed, M. K., Menazea, A. A., & Abdelghany, A. M. (2020). Blend biopolymeric nanofibrous scaffolds of cellulose acetate/ ϵ -polycaprolactone containing metallic nanoparticles prepared by laser ablation for wound disinfection applications. *International Journal of Biological Macromolecules*, 155, 636-644.
- Al-Rajabi, M. M., & Haan, T. Y. (2021). Influence of Vertical Diffusion Cell Set-Up on *In Vitro* Silver Sulfadiazine Drug Release from Thermo-Responsive Cellulose Hydrogel. *Materials Science Forum*, 1030, 19-26. Trans Tech Publications Ltd.

- Alam, A., Ewaldz, E., Xiang, C., Qu, W., & Bai, X. (2020). Tunable wettability of biodegradable multilayer sandwich-structured electrospun nanofibrous membranes. *Polymers*, 12(9), 2092.
- Alghoraibi, I., & Alomari, S. (2018). Different Methods for Nanofiber Design and Fabrication. *Handbook of Nanofibers*, 1-46.
- Ali, A. S., El-Aassar, M., Hashem, F., & Moussa, N. (2019). Surface Modified of Cellulose Acetate Electrospun Nanofibers by Polyaniline/β-cyclodextrin Composite for Removal of Cationic Dye from Aqueous Medium. *Fibers and Polymers*, 20(10), 2057-2069.
- Ali, N. (2020). Role of vitamin D in preventing of COVID-19 infection, progression and severity. *Journal of infection and public health*, 13(10), 1373-1380.
- Alves, J. A. A., dos Santos, M. D. L., Morais, C. C., Ascheri, J. L. R., Signini, R., dos Santos, D. M., Bastos, S. M. C., & Ascheri, D. P. R. (2019). Sorghum straw: Pulping and bleaching process optimization and synthesis of cellulose acetate. *International Journal of Biological Macromolecules*, 135, 877-886.
- Amaral, H. R., Cipriano, D. F., Santos, M. S., Schettino, M. A., Ferreti, J. V. T., Meirelles, C. S., Pereira, V. S., Cunha, A. G., Emmerich, F. G., & Freitas, J. C. C. (2019). Production of high-purity cellulose, cellulose acetate and cellulose-silica composite from babassu coconut shells. *Carbohydrate Polymers*, 210, 127-134.
- Amrein, K., Scherkl, M., Hoffmann, M., Neuwersch-Sommeregger, S., Köstenberger, M., Berisha, A. T., Martucci, G., Pilz, S., & Malle, O. (2020). Vitamin D deficiency 2.0: an update on the current status worldwide. *European journal of clinical nutrition*, 74(11), 1498-1513.
- Ardekani, N. T., Khorram, M., Zomorodian, K., Yazdanpanah, S., Veisi, H., & Veisi, H. (2019). Evaluation of electrospun poly (vinyl alcohol)-based nanofiber mats

incorporated with Zataria multiflora essential oil as potential wound dressing.
International Journal of Biological Macromolecules, 125, 743-750.

Ashraf, R., Sofi, H. S., Akram, T., Rather, H. A., Abdal-hay, A., Shabir, N., Vasita, R., Alrokayan, S. H., Khan, H. A., & Sheikh, F. A. (2020). Fabrication of Multifunctional Cellulose/TiO₂/AgComposite Nanofibers Scaffold with Antibacterial and Bioactivity Properties for Future Tissue Engineering Applications. *Journal of Biomedical Materials Research Part A*, 108(4), 947-962.

Ashraf, R., Sofi, H. S., Malik, A., Beigh, M. A., Hamid, R., & Sheikh, F. A. (2019). Recent Trends in the Fabrication of Starch Nanofibers: Electrospinning and Non-electrospinning Routes and Their Applications in Biotechnology. *Applied Biochemistry and Biotechnology*, 187(1), 47-74.

Aslantürk, Ö. S. (2018). *In vitro* cytotoxicity and cell viability assays: principles, advantages, and disadvantages. *Genotoxicity-A predictable risk to our actual world*, 2, 64-80.

Bagheri, M., & Mahmoodzadeh, A. (2020). Polycaprolactone/Graphene Nanocomposites: Synthesis, Characterization and Mechanical Properties of Electrospun Nanofibers. *Journal of Inorganic and Organometallic Polymers and Materials*, 30(5), 1566-1577.

Bao, C. (2015). Cellulose acetate/plasticizer systems: structure, morphology and dynamics. Doctoral dissertation, Universite Claude Bernard-Lyon I.

Barhoum, A., Rasouli, R., Yousefzadeh, M., Rahier, H., & Bechelany, M. (2018). Nanofiber Technology: History and Developments. *Handbook of Nanofibers*, 1-42.

Bate, T. S., Forbes, S. J., & Callanan, A. (2020). Controlling electrospun polymer morphology for tissue engineering demonstrated using hepG2 cell line. *JoVE (Journal of Visualized Experiments)*(159), e61043.

Bhosle, V. K., Altit, G., Autmizguine, J., & Chemtob, S. (2017). 18 - Basic Pharmacologic Principles. In R. A. Polin, S. H. Abman, D. H. Rowitch, W. E. Benitz, & W. W. Fox (Eds.), *Fetal and Neonatal Physiology (Fifth Edition)* (pp. 187-201.e183). Elsevier.

Bikle, D. D., Adams, J. S., & Christakos, S. (2018). Vitamin D: production, metabolism, action, and clinical requirements. In *Primer on the Metabolic Bone Diseases and Disorders of Mineral Metabolism* (Vol. 25, pp. 230-240). John Wiley & Sons.

Branton, A., & Jana, S. (2017). A Study on the Effect of the Energy of Consciousness Healing Treatment on Physicochemical and Thermal Properties of Vitamin D₃ (Cholecalciferol). *Science Journal of Analytical Chemistry*, 5(4), 46-55.

Bruschi, M. L. (2015). Mathematical models of drug release. Strategies to modify the drug release from pharmaceutical systems, 63.

Candido, R. G., Godoy, G. G., & Gonçalves, A. R. (2017). Characterization and application of cellulose acetate synthesized from sugarcane bagasse. *Carbohydrate Polymers*, 167, 280-289.

Castoldi, A., Herr, C., Niederstraßer, J., Labouta, H. I., Melero, A., Gordon, S., Schneider-Daum, N., Bals, R., & Lehr, C.-M. (2017). Calcifediol-loaded liposomes for local treatment of pulmonary bacterial infections. *European Journal of Pharmaceutics and Biopharmaceutics*, 118, 62-67.

Chaparro, F. J., Presley, K. F., Coutinho da Silva, M. A., & Lannutti, J. J. (2019). Sintered electrospun polycaprolactone for controlled model drug delivery. *Materials Science and Engineering: C*, 99, 112-120.

Chen, J., Xu, J., Wang, K., Cao, X., & Sun, R. (2016). Cellulose acetate fibers prepared from different raw materials with rapid synthesis method. *Carbohydrate Polymers*, 137, 685-692.

- Chen, S., Li, R., Li, X., & Xie, J. (2018). Electrospinning: An enabling nanotechnology platform for drug delivery and regenerative medicine. *Advanced Drug Delivery Reviews*, 132, 188-213.
- Chen, Y., Qiu, Y., Chen, W., & Wei, Q. (2020). Electrospun thymol-loaded porous cellulose acetate fibers with potential biomedical applications. *Materials Science and Engineering: C*, 109, 110536.
- Chou, S.-F., Carson, D., & Woodrow, K. A. (2015). Current strategies for sustaining drug release from electrospun nanofibers. *Journal of controlled release*, 220, 584-591.
- Chou, S.-F., & Woodrow, K. A. (2017). Relationships between mechanical properties and drug release from electrospun fibers of PCL and PLGA blends. *Journal of the Mechanical Behavior of Biomedical Materials*, 65, 724-733.
- Christensen, G., Younes, H., Hong, H., & Smith, P. (2015). Effects of solvent hydrogen bonding, viscosity, and polarity on the dispersion and alignment of nanofluids containing Fe₂O₃ nanoparticles. *Journal of Applied Physics*, 118(21), 214302.
- Chung, J., & Kwak, S. Y. (2019). Effect of nanoscale confinement on molecular mobility and drug release properties of cellulose acetate/sulindac nanofibers. *Journal of applied polymer science*, 136(33), 47863.
- Costa, G. M. D. A., de Oliveira Pinto, C. A. S., Leite-Silva, V. R., Baby, A. R., & Velasco, M. V. R. (2018). Is Vitamin D 3 Transdermal Formulation Feasible? An Ex Vivo Skin Retention and Permeation. *AAPSpharmscitech*, 19(5), 2418-2425.
- Da Silva, G. R., Lima, T. H., Fernandes-Cunha, G. M., Oréfice, R. L., Da Silva-Cunha, A., Zhao, M., & Behar-Cohen, F. (2019). Ocular biocompatibility of dexamethasone acetate loaded poly (ϵ -caprolactone) nanofibers. *European Journal of Pharmaceutics and Biopharmaceutics*, 142, 20-30.

- Dahe, G. J., Singh, R. P., Dudeck, K. W., Yang, D., & Berchtold, K. A. (2019). Influence of non-solvent chemistry on polybenzimidazole hollow fiber membrane preparation. *Journal of Membrane Science*, 577, 91-103.
- Damodaran, V. B., Bhatnagar, D., & Murthy, N. S. (2016). Biomedical Polymers: Processing. In Biomedical Polymers (pp. 55-71). Springer, Cham.
- Dash, S., Murthy, P. N., Nath, L., & Chowdhury, P. (2010). Kinetic modeling on drug release from controlled drug delivery systems. *Acta Pol Pharm*, 67(3), 217-223.
- Davoodi, P., Lee, L. Y., Xu, Q., Sunil, V., Sun, Y., Soh, S., & Wang, C.-H. (2018). Drug delivery systems for programmed and on-demand release. *Advanced Drug Delivery Reviews*, 132, 104-138.
- De Freitas, R. R. M., Senna, A. M., & Botaro, V. R. (2017). Influence of degree of substitution on thermal dynamic mechanical and physicochemical properties of cellulose acetate. *Industrial Crops and Products*, 109, 452-458.
- de Melo Brites, M., Cerón, A. A., Costa, S. M., Oliveira, R. C., Ferraz, H. G., Catalani, L. H., & Costa, S. A. (2020). Bromelain immobilization in cellulose triacetate nanofiber membranes from sugarcane bagasse by electrospinning technique. *Enzyme and microbial technology*, 132, 109384.
- Debeer, S., Le Luduec, J.-B., Kaiserlian, D., Laurent, P., Nicolas, J.-F., Dubois, B., & Kanitakis, J. (2013). Comparative histology and immunohistochemistry of porcine versus human skin. *European journal of dermatology*, 23(4), 456-466.
- Deepak, A., Goyal, A. K., & Rath, G. (2018). Nanofiber in transmucosal drug delivery. *Journal of Drug Delivery Science and Technology*, 43, 379-387.
- Deshmukh, K., Basheer Ahamed, M., Deshmukh, R. R., Khadheer Pasha, S. K., Bhagat, P. R., & Chidambaram, K. (2017). 3 - Biopolymer Composites With High Dielectric Performance: Interface Engineering. In K. K. Sadasivuni, D.

Ponnamma, J. Kim, J. J. Cabibihan, & M. A. AlMaadeed (Eds.), *Biopolymer Composites in Electronics* (pp. 27-128). Elsevier.

Doostan, M., Maleki, H., Doostan, M., Khoshnevisan, K., Faridi-Majidi, R., & Arkan, E. (2020). Effective antibacterial electrospun cellulose acetate nanofibrous patches containing chitosan/erythromycin nanoparticles. *International Journal of Biological Macromolecules*, 168, 464-473.

Dos Santos, A. E. A., dos Santos, F. V., Freitas, K. M., Pimenta, L. P. S., de Oliveira Andrade, L., Marinho, T. A., de Avelar, G. F., da Silva, A. B., & Ferreira, R. V. (2021). Cellulose acetate nanofibers loaded with crude annatto extract: Preparation, characterization, and in vivo evaluation for potential wound healing applications. *Materials Science and Engineering: C*, 118, 111322.

Drosou, C., Krokida, M., & Biliaderis, C. G. (2018). Composite pullulan-whey protein nanofibers made by electrospinning: Impact of process parameters on fiber morphology and physical properties. *Food Hydrocolloids*, 77, 726-735.

Einarsdóttir, K., Preen, D. B., Clay, T. D., Kiely, L., Holman, C. A. J., & Cohen, L. D. (2010). Effect of a single ‘megadose’intramuscular vitamin D (600,000 IU) injection on vitamin D concentrations and bone mineral density following biliopancreatic diversion surgery. *Obesity surgery*, 20(6), 732-737.

Erdmann, R., Kabascı, S., & Heim, H.-P. (2021). Thermal Properties of Plasticized Cellulose Acetate and Its β -Relaxation Phenomenon. *Polymers*, 13(9), 1356.

Eskitoros-Togay, Ş. M., Bulbul, Y. E., Tort, S., Demirtaş Korkmaz, F., Acartürk, F., & Dilsiz, N. (2019). Fabrication of doxycycline-loaded electrospun PCL/PEO membranes for a potential drug delivery system. *International Journal of Pharmaceutics*, 565, 83-94.

Fadaie, M., Mirzaei, E., Geramizadeh, B., & Asvar, Z. (2018). Incorporation of nanofibrillated chitosan into electrospun PCL nanofibers makes scaffolds with

enhanced mechanical and biological properties. *Carbohydrate Polymers*, 199, 628-640.

Fan, F., Coutinho da Silva, M. A., Moraes, C. R., Dunham, A. D., HogenEsch, H., Turner, J. W., & Lannutti, J. J. (2020). Self-reinforcing nanoscalar polycaprolactone-polyethylene terephthalate electrospun fiber blends. *Polymer*, 202, 122573.

Fong, H., Chun, I., & Reneker, D. H. (1999, 1999/07/01/). Beaded nanofibers formed during electrospinning. *Polymer*, 40(16), 4585-4592.

Fu, Y., & Kao, W. J. (2010). Drug release kinetics and transport mechanisms of non-degradable and degradable polymeric delivery systems. *Expert Opinion on Drug Delivery*, 7(4), 429-444.

Garg, K., & Bowlin, G. L. (2011). Electrospinning jets and nanofibrous structures. *Biomicrofluidics*, 5(1), 013403.

Ghaffarian, V., Mousavi, S. M., Bahreini, M., & Parchin, N. S. (2017). Biodegradation of cellulose acetate/poly (butylene succinate) membrane. *International journal of environmental science and technology*, 14(6), 1197-1208.

Ghafoor, B., Aleem, A., Najabat Ali, M., & Mir, M. (2018). Review of the fabrication techniques and applications of polymeric electrospun nanofibers for drug delivery systems. *Journal of Drug Delivery Science and Technology*, 48, 82-87.

Ghorani, B., Goswami, P., Blackburn, R. S., & Russell, S. J. (2018). Enrichment of cellulose acetate nanofiber assemblies for therapeutic delivery of l-tryptophan. *International Journal of Biological Macromolecules*, 108, 1-8.

Glowka, E., Stasiak, J., & Lulek, J. (2019). Drug Delivery Systems for Vitamin D Supplementation and Therapy. *Pharmaceutics*, 11(7), 347.

- Golizadeh, M., Karimi, A., Gandomi-Ravandi, S., Vossoughi, M., Khafaji, M., Joghataei, M. T., & Faghihi, F. (2019). Evaluation of cellular attachment and proliferation on different surface charged functional cellulose electrospun nanofibers. *Carbohydrate Polymers*, 207, 796-805.
- Gong, Z., Du, Y., He, Y., Yang, A., Yang, Y., & Yu, D. (2019). *The Influence of DMAc Ratio in Sheath Fluid on the Diameters of Medicated Cellulose Acetate Nanofibers* International Conference on Biology, Chemistry and Medical Engineering, Hefei, China,.
- Grant, W. B. (2020). Review of recent advances in understanding the role of vitamin D in reducing cancer risk: breast, colorectal, prostate, and overall cancer. *Anticancer Research*, 40(1), 491-499.
- Greiner, A., & Wendorff, J. H. (2007). Electrospinning: a fascinating method for the preparation of ultrathin fibers. *Angewandte Chemie International Edition*, 46(30), 5670-5703.
- Gupta, N., Farooqui, K. J., Batra, C. M., Marwaha, R. K., & Mithal, A. (2017, Jan-Feb). Effect of oral versus intramuscular Vitamin D replacement in apparently healthy adults with Vitamin D deficiency. *Indian journal of endocrinology and metabolism*, 21(1), 131-136.
- Gupta, R., Behera, C., Paudwal, G., Rawat, N., Baldi, A., & Gupta, P. N. (2019). Recent advances in formulation strategies for efficient delivery of vitamin D. *AAPS pharmscitech*, 20(1), 1-12.
- Haas, D., Heinrich, S., & Greil, P. (2010). Solvent control of cellulose acetate nanofiber felt structure produced by electrospinning. *Journal of Materials Science*, 45(5), 1299-1306.
- Haider, A., Haider, S., & Kang, I.-K. (2018). A comprehensive review summarizing the effect of electrospinning parameters and potential applications of

nanofibers in biomedical and biotechnology. *Arabian Journal of Chemistry*, 11(8), 1165-1188.

Hanel, A., & Carlberg, C. (2020). Vitamin D and evolution: Pharmacologic implications. *Biochemical Pharmacology*, 173, 113595.

Hasanvand, E., Fathi, M., & Bassiri, A. (2018). Production and characterization of vitamin D 3 loaded starch nanoparticles: effect of amylose to amylopectin ratio and sonication parameters. *Journal of food science and technology*, 55(4), 1314-1324.

Haske-Cornelius, O., Pellis, A., Tegl, G., Wurz, S., Saake, B., Ludwig, R., Sebastian, A., Nyanhongo, G. S., & Guebitz, G. M. (2017). Enzymatic systems for cellulose acetate degradation. *Catalysts*, 7(10).

Hassan, M. I., & Sultana, N. (2017). Characterization, drug loading and antibacterial activity of nanohydroxyapatite/polycaprolactone (nHA/PCL) electrospun membrane. *3 Biotech*, 7(4), 1-9.

Heinze, T., & Liebert, T. (2004). 4.2 Chemical characteristics of cellulose acetate. *Macromolecular symposia*, 208(1), 167-238.

Hivechi, A., Bahrami, S. H., & Siegel, R. A. (2019). Drug release and biodegradability of electrospun cellulose nanocrystal reinforced polycaprolactone. *Materials Science and Engineering: C*, 94, 929-937.

Holick, M. F. (2018). Chapter 4 - Photobiology of Vitamin D. In D. Feldman (Ed.), *Vitamin D (Fourth Edition)* (pp. 45-55). Academic Press.

Hwang, T. I., Kim, J. I., Joshi, M. K., Park, C. H., & Kim, C. S. (2019). Simultaneous regeneration of calcium lactate and cellulose into PCL nanofiber for biomedical application. *Carbohydrate Polymers*, 212, 21-29.

- Iacob, A.-T., Drăgan, M., Ionescu, O.-M., Profire, L., Ficai, A., Andronescu, E., Confederat, L. G., & Lupașcu, D. (2020). An Overview of Biopolymeric Electrospun Nanofibers Based on Polysaccharides for Wound Healing Management. *Pharmaceutics*, 12(10), 983.
- Ilie, P. C., Stefanescu, S., & Smith, L. (2020). The role of vitamin D in the prevention of coronavirus disease 2019 infection and mortality. *Aging clinical and experimental research*, 32(7), 1195-1198.
- Jain, K. K. (2020). An overview of drug delivery systems. *Drug Delivery Systems*, 2059, 1-54.
- Jatoi, A. W., Kim, I. S., & Ni, Q.-Q. (2019). Cellulose acetate nanofibers embedded with AgNPs anchored TiO₂ nanoparticles for long term excellent antibacterial applications. *Carbohydrate Polymers*, 207, 640-649.
- Jiang, J., Chen, G., Shuler, F. D., Wang, C.-H., & Xie, J. (2015). Local sustained delivery of 25-hydroxyvitamin D₃ for production of antimicrobial peptides. *Pharmaceutical research*, 32(9), 2851-2862.
- Johnson, A. R., Forster, S. P., White, D., Terife, G., Lowinger, M., Teller, R. S., & Barrett, S. E. (2020). Drug eluting implants in pharmaceutical development and clinical practice. *Expert Opinion on Drug Delivery*, 1-17.
- Jones, G. (2008). Pharmacokinetics of vitamin D toxicity. *The American journal of clinical nutrition*, 88(2), 582S-586S.
- Joshi, D. R., & Adhikari, N. (2019). An overview on common organic solvents and their toxicity. *Journal of Pharmaceutical Research International*, 1-18.
- Junaid, K., & Rehman, A. (2019). Impact of vitamin D on infectious disease-tuberculosis-A review. *Clinical Nutrition Experimental*, 25, 1-10.

- Kajdič, S., Planinšek, O., Gašperlin, M., & Kocbek, P. (2019). Electrospun nanofibers for customized drug-delivery systems. *Journal of Drug Delivery Science and Technology*, 51, 672-681.
- Kamble, P., Sadarani, B., Majumdar, A., & Bhullar, S. (2017, 2017/10/01/). Nanofiber based drug delivery systems for skin: A promising therapeutic approach. *Journal of Drug Delivery Science and Technology*, 41, 124-133.
- Kaur, G., Grewal, J., Jyoti, K., Jain, U. K., Chandra, R., & Madan, J. (2018). Oral controlled and sustained drug delivery systems: Concepts, advances, preclinical, and clinical status. In *Drug Targeting and Stimuli Sensitive Drug Delivery Systems* (pp. 567-626). Elsevier.
- Khalid, N., Kobayashi, I., Neves, M. A., Uemura, K., Nakajima, M., & Nabetani, H. (2017). Encapsulation of cholecalciferol and ergocalciferol in oil-in-water emulsions by different homogenization techniques. *European Journal of Lipid Science and Technology*, 119(6), 1600247.
- Khan, M. Q., Kharaghani, D., Shahzad, A., Saito, Y., Yamamoto, T., Ogasawara, H., & Kim, I. S. (2019). Fabrication of antibacterial electrospun cellulose acetate/silver-sulfadiazine nanofibers composites for wound dressings applications. *Polymer Testing*, 74, 39-44.
- Khan, W. S., Ceylan, M., Jabarrania, A., Saeednia, L., & Asmatulu, R. (2017). Chemical and thermal investigations of electrospun polyacrylonitrile nanofibers incorporated with various nanoscale inclusions. *Journal of Thermal Engineering*, 3(4), 1375-1390.
- Khoshnevisan, K., Maleki, H., Samadian, H., Doostan, M., & Khorramizadeh, M. R. (2019). Antibacterial and antioxidant assessment of cellulose acetate/polycaprolactone nanofibrous mats impregnated with propolis. *International Journal of Biological Macromolecules*, 140, 1260-1268.

- Khoshnevisan, K., Maleki, H., Samadian, H., Shahsavari, S., Sarrafzadeh, M. H., Larijani, B., Dorkoosh, F. A., Haghpanah, V., & Khorramizadeh, M. R. (2018). Cellulose acetate electrospun nanofibers for drug delivery systems: Applications and recent advances. *Carbohydrate Polymers*, 198, 131-141.
- Kim, H.-G., Gater, D. L., & Kim, Y.-C. (2018). Development of transdermal vitamin D₃ (VD3) delivery system using combinations of PLGA nanoparticles and microneedles. *Drug delivery and translational research*, 8(1), 281-290.
- Kinnunen, H. M., & Mrsny, R. J. (2014). Improving the outcomes of biopharmaceutical delivery via the subcutaneous route by understanding the chemical, physical and physiological properties of the subcutaneous injection site. *Journal of controlled release*, 182, 22-32.
- Kohrs, N. J., Liyanage, T., Venkatesan, N., Najazadeh, A., & Puleo, D. A. (2019). Drug Delivery Systems and Controlled Release. In R. Narayan (Ed.), *Encyclopedia of Biomedical Engineering* (pp. 316-329). Elsevier.
- Kumar, A., & Pillai, J. (2018). Chapter 13 - Implantable drug delivery systems: An overview. In A. M. Grumezescu (Ed.), *Nanostructures for the Engineering of Cells, Tissues and Organs* (pp. 473-511). William Andrew Publishing.
- Kurečič, M., Maver, T., Virant, N., Ojstršek, A., Gradišnik, L., Hribernik, S., Kolar, M., Maver, U., & Kleinschek, K. S. (2018). A multifunctional electrospun and dual nano-carrier biobased system for simultaneous detection of pH in the wound bed and controlled release of benzocaine. *Cellulose*, 25(12), 7277-7297.
- Lalloz, A., Bolzinger, M.-A., Faivre, J., Latreille, P.-L., Ac, A. G., Rakotovao, C., Rabanel, J.-M., Hildgen, P., Banquy, X., & Briançon, S. (2018). Effect of surface chemistry of polymeric nanoparticles on cutaneous penetration of cholecalciferol. *International Journal of Pharmaceutics*, 553(1-2), 120-131.

- Li, B., & Yang, X. (2020). Rutin-loaded cellulose acetate/poly(ethylene oxide) fiber membrane fabricated by electrospinning: A bioactive material. *Materials Science and Engineering: C*, 109, 110601.
- Li, S., Huang, J., Chen, Z., Chen, G., & Lai, Y. (2017). A review on special wettability textiles: theoretical models, fabrication technologies and multifunctional applications. *Journal of Materials Chemistry A*, 5(1), 31-55.
- Li, Z., & Wang, C. (2013). Effects of working parameters on electrospinning. In *One-dimensional nanostructures* (pp. 15-28). Springer, Berlin, Heidelberg.
- Liakos, I., Holban, A., Carzino, R., Lauciello, S., & Grumezescu, A. (2017). Electrospun fiber pads of cellulose acetate and essential oils with antimicrobial activity. *Nanomaterials*, 7(4), 84.
- Liang, R., Bao, Z., Su, B., Xing, H., & Ren, Q. (2012). Solubility of vitamin D₃ in six organic solvents at temperatures from (248.2 to 273.2) K. *Journal of Chemical & Engineering Data*, 57(8), 2328-2331.
- Liang, W., Hou, J., Fang, X., Bai, F., Zhu, T., Gao, F., Wei, C., Mo, X., & Lang, M. (2018). Synthesis of cellulose diacetate based copolymer electrospun nanofibers for tissues scaffold. *Applied Surface Science*, 443, 374-381.
- Lim, M. M., Sun, T., & Sultana, N. (2015). *In vitro* biological evaluation of electrospun polycaprolactone/gelatine nanofibrous scaffold for tissue engineering. *Journal of Nanomaterials*, 16(1), 416.
- Liu, F., Li, X., Wang, L., Yan, X., Ma, D., Liu, Z., & Liu, X. (2020). Sesamol incorporated cellulose acetate-zein composite nanofiber membrane: An efficient strategy to accelerate diabetic wound healing. *International Journal of Biological Macromolecules*, 149, 627-638.

- Liu, H., & Hsieh, Y. L. (2002). Ultrafine fibrous cellulose membranes from electrospinning of cellulose acetate. *Journal of Polymer Science Part B: Polymer Physics*, 40(18), 2119-2129.
- Liu, H., & Tang, C. (2007). Electrospinning of cellulose acetate in solvent mixture N, N-dimethylacetamide (DMAc)/acetone. *Polymer Journal*, 39(1), 65-72.
- Luraghi, A., Peri, F., & Moroni, L. (2021, 2021/06/10/). Electrospinning for drug delivery applications: A review. *Journal of controlled release*, 334, 463-484.
- Lysik, D., Mystkowska, J., Markiewicz, G., Deptuła, P., & Bucki, R. (2019). The Influence of Mucin-Based Artificial Saliva on Properties of Polycaprolactone and Polylactide. *Polymers*, 11(11), 1880.
- Lyu, J. S., Lee, J.-S., & Han, J. (2019, 2019/12/27). Development of a biodegradable polycaprolactone film incorporated with an antimicrobial agent via an extrusion process. *Scientific Reports*, 9(1), 20236.
- Ma, H., & Hsiao, B. S. (2019). Chapter 4 - Electrospun Nanofibrous Membranes for Desalination. In A. Basile, E. Curcio, & Inamuddin (Eds.), *Current Trends and Future Developments on (Bio-) Membranes* (pp. 81-104). Elsevier.
- Mahmoodani, F., Perera, C. O., Abernethy, G., Fedrizzi, B., & Chen, H. (2018, 2018/09/30/). Lipid oxidation and vitamin D₃ degradation in simulated whole milk powder as influenced by processing and storage. *Food Chemistry*, 261, 149-156.
- Mahmoodani, F., Perera, C. O., Fedrizzi, B., Abernethy, G., & Chen, H. (2017). Degradation studies of cholecalciferol (vitamin D₃) using HPLC-DAD, UHPLC-MS/MS and chemical derivatization. *Food Chemistry*, 219, 373-381.
- Malikmammadov, E., Tanir, T. E., Kiziltay, A., Hasirci, V., & Hasirci, N. (2018). PCL and PCL-based materials in biomedical applications. *Journal of Biomaterials Science, Polymer Edition*, 29(7-9), 863-893.

Mandic, M., Spasic, J., Ponjovic, M., Nikolic, M. S., Cosovic, V. R., O'Connor, K. E., Nikodinovic-Runic, J., Djokic, L., & Jeremic, S. (2019). Biodegradation of poly(ϵ -caprolactone) (PCL) and medium chain length polyhydroxyalkanoate (mcl-PHA) using whole cells and cell free protein preparations of *Pseudomonas* and *Streptomyces* strains grown on waste cooking oil. *Polymer Degradation and Stability*, 162, 160-168.

Manoukian, O. S., Sardashti, N., Stedman, T., Gailiunas, K., Ojha, A., Penalosa, A., Mancuso, C., Hobert, M., & Kumbar, S. G. (2019). Biomaterials for Tissue Engineering and Regenerative Medicine. In R. Narayan (Ed.), *Encyclopedia of Biomedical Engineering* (pp. 462-482). Elsevier.

Marginot, A. J., Calderón, F. J., Goyne, K. W., Mukome, F. N. D., & Parikh, S. J. (2017). IR Spectroscopy, Soil Analysis Applications. In J. C. Lindon, G. E. Tranter, & D. W. Koppenaal (Eds.), *Encyclopedia of Spectroscopy and Spectrometry (Third Edition)* (pp. 448-454). Academic Press.

Märtson, M., Viljanto, J., Hurme, T., Laippala, P., & Saukko, P. (1999). Is cellulose sponge degradable or stable as implantation material? An in vivo subcutaneous study in the rat. *Biomaterials*, 20(21), 1989-1995.

Maurya, V. K., & Aggarwal, M. (2017). Enhancing bio-availability of vitamin D by Nano-engineered based delivery systems-An overview. *IIInternational Journal of Current Microbiology and Applied Sciences*, 6(7), 340-353.

McCullough, P. J., Lehrer, D. S., & Amend, J. (2019, 2019/05/01/). Daily oral dosing of vitamin D₃ using 5000 TO 50,000 international units a day in long-term hospitalized patients: Insights from a seven year experience. *The Journal of Steroid Biochemistry and Molecular Biology*, 189, 228-239.

McKeen, L. W. (2017). 13 - Environmentally Friendly Polymers. In L. W. McKeen (Ed.), *Permeability Properties of Plastics and Elastomers (Fourth Edition)* (pp. 305-323). William Andrew Publishing.

- Mehrabi, F., Shamspur, T., Mostafavi, A., Saljooqi, A., & Fathirad, F. (2017). Synthesis of cellulose acetate nanofibers and its application in the release of some drugs. *Nanomedicine Research Journal*, 2(3), 199-207.
- Mehta, P. P., & Pawar, V. S. (2018). 22 - Electrospun nanofiber scaffolds: Technology and applications. In Inamuddin, A. M. Asiri, & A. Mohammad (Eds.), *Applications of Nanocomposite Materials in Drug Delivery* (pp. 509-573). Woodhead Publishing.
- Metwally, S., Ferraris, S., Spriano, S., Krysiak, Z. J., Kaniuk, L., Marzec, M. M., Kim, S. K., Szewczyk, P. K., Gruszczynski, A., & Wytrwal-Sarna, M. (2020). Surface potential and roughness controlled cell adhesion and collagen formation in electrospun PCL fibers for bone regeneration. *Materials & Design*, 194, 108915.
- Mikaeili, F., & Gouma, P. I. (2018). Super water-repellent cellulose acetate mats. *Scientific Reports*, 8(1), 1-8.
- Miranda, C. S., Ribeiro, A. R., Homem, N. C., & Felgueiras, H. P. (2020). Spun Bioteextiles in Tissue Engineering and Biomolecules Delivery Systems. *Antibiotics*, 9(4), 174.
- Mochane, M. J., Motsoeneng, T. S., Sadiku, E. R., Mokhena, T. C., & Sefadi, J. S. (2019). Morphology and Properties of Electrospun PCL and Its Composites for Medical Applications: A Mini Review. *Applied Sciences*, 9(11), 2205.
- Mohammadi, M., Pezeshki, A., Abbasi, M. M., Ghanbarzadeh, B., & Hamishehkar, H. (2017). Vitamin D₃-loaded nanostructured lipid carriers as a potential approach for fortifying food beverages; *in vitro* and *in vivo* evaluation. *Advanced pharmaceutical bulletin*, 7(1), 61.
- Mohapatra, S., Samanta, S., Kothari, K., Mistry, P., & Suryanarayanan, R. (2017). Effect of polymer molecular weight on the crystallization behavior of

indomethacin amorphous solid dispersions. *Crystal Growth & Design*, 17(6), 3142-3150.

Molaveisi, M., Shahidi-Noghabi, M., & Naji-Tabasi, S. (2020). Vitamin D₃-loaded nanophytosomes for enrichment purposes: Formulation, structure optimization, and controlled release. *Journal of Food Process Engineering*, 43(12), e13560.

Moreno Raja, M., Lim, P. Q., Wong, Y. S., Xiong, G. M., Zhang, Y., Venkatraman, S., & Huang, Y. (2019). Chapter 18 - Polymeric Nanomaterials: Methods of Preparation and Characterization. In S. S. Mohapatra, S. Ranjan, N. Dasgupta, R. K. Mishra, & S. Thomas (Eds.), *Nanocarriers for Drug Delivery* (pp. 557-653). Elsevier.

Nagarajan, S., Bechelany, M., Kalkura, N. S., Miele, P., Bohatier, C. P., & Balme, S. (2019). Chapter 20 - Electrospun Nanofibers for Drug Delivery in Regenerative Medicine. In S. S. Mohapatra, S. Ranjan, N. Dasgupta, R. K. Mishra, & S. Thomas (Eds.), *Applications of Targeted Nano Drugs and Delivery Systems* (pp. 595-625). Elsevier.

Naragund, V. S., & Panda, P. (2020). Electrospinning of cellulose acetate nanofiber membrane using methyl ethyl ketone and N, N-Dimethylacetamide as solvents. *Materials Chemistry and Physics*, 240, 122147.

Nazeer, M. A., Yilgor, E., & Yilgor, I. (2019). Electrospun polycaprolactone/silk fibroin nanofibrous bioactive scaffolds for tissue engineering applications. *Polymer*, 168, 86-94.

Nelson, M. T., Keith, J. P., Li, B.-B., Stocum, D. L., & Li, J. (2012). Electrospun composite polycaprolactone scaffolds for optimized tissue regeneration. *Proceedings of the Institution of Mechanical Engineers, Part N: Journal of Nanoengineering and Nanosystems*, 226(3), 111-121.

- Nelson, M. T., Pattanaik, L., Allen, M., Gerbich, M., Hux, K., Allen, M., & Lannutti, J. J. (2014). Recrystallization improves the mechanical properties of sintered electrospun polycaprolactone. *Journal of the Mechanical Behavior of Biomedical Materials*, 30, 150-158.
- Ng, K., Azari, P., Nam, H. Y., Xu, F., & Pingguan-Murphy, B. (2019). Electrospin-Coating of Paper: A Natural Extracellular Matrix Inspired Design of Scaffold. *Polymers*, 11(4), 650.
- Olson-Kennedy, J., Streeter, L. H., Garofalo, R., Chan, Y.-M., & Rosenthal, S. M. (2021). Histrelin implants for suppression of puberty in youth with gender dysphoria: a comparison of 50 mcg/day (Vantas) and 65 mcg/day (SupprelinLA). *Transgender Health*, 6(1), 36-42.
- Palacios, C., & Gonzalez, L. (2014). Is vitamin D deficiency a major global public health problem? *The Journal of Steroid Biochemistry and Molecular Biology*, 144, 138-145.
- Palomba, S., Falbo, A., Di Cello, A., Materazzo, C., & Zullo, F. (2012). Nexplanon: the new implant for long-term contraception. A comprehensive descriptive review. *Gynecol Endocrinol*, 28(9), 710-721.
- Pant, B., Park, M., & Park, S.-J. (2019). Drug Delivery Applications of Core-Sheath Nanofibers Prepared by Coaxial Electrospinning: A Review. *Pharmaceutics*, 11(7).
- Park, H.-M., Liang, X., Mohanty, A. K., Misra, M., & Drzal, L. T. (2004). Effect of compatibilizer on nanostructure of the biodegradable cellulose acetate/organoclay nanocomposites. *Macromolecules*, 37(24), 9076-9082.
- Park, S. J., Garcia, C. V., Shin, G. H., & Kim, J. T. (2017). Development of nanostructured lipid carriers for the encapsulation and controlled release of vitamin D₃. *Food Chemistry*, 225, 213-219.

Parrott, M., & Dunn, S. (2018). Design of Biomedical Polymers. *Functional Biopolymers*, 1-48.

Patra, J. K., Das, G., Fraceto, L. F., Campos, E. V. R., Rodriguez-Torres, M. d. P., Acosta-Torres, L. S., Diaz-Torres, L. A., Grillo, R., Swamy, M. K., Sharma, S., Habtemariam, S., & Shin, H.-S. (2018). Nano based drug delivery systems: recent developments and future prospects. *Journal of Nanobiotechnology*, 16(1), 71.

Phiriyawirut, M., & Phaechamud, T. (2012a). Cellulose acetate electrospun fiber mats for controlled release of silymarin. *Journal of nanoscience and nanotechnology*, 12(1), 793-799.

Phiriyawirut, M., & Phaechamud, T. (2012b). Gallic acid-loaded cellulose acetate electrospun nanofibers: thermal properties, mechanical properties, and drug release behavior. *Open Journal of Polymer Chemistry*(2), 21-29.

Pillay, V., Dott, C., Choonara, Y. E., Tyagi, C., Tomar, L., Kumar, P., du Toit, L. C., & Ndesendo, V. M. (2013). A review of the effect of processing variables on the fabrication of electrospun nanofibers for drug delivery applications. *Journal of Nanomaterials*, 2013.

Pondi, S. B. (2017). Controlled-Release Of Curcumin From Poly (Lactide-Co-Glycolide) Acid/Albumin/Curcumin And Silica/Albumin/Curcumin Drug-Delivery Systems. Ph.D. Thesis. Universiti Teknologi Malaysia.

Pons-Faudoa, F. P., Ballerini, A., Sakamoto, J., & Grattoni, A. (2019). Advanced implantable drug delivery technologies: transforming the clinical landscape of therapeutics for chronic diseases. *Biomedical microdevices*, 21(2), 47.

Qin, X., & Wu, D. (2012). Effect of different solvents on poly (caprolactone)(PCL) electrospun nonwoven membranes. *Journal of Thermal Analysis and Calorimetry*, 107(3), 1007-1013.

Qin, Y. (2016). 14 - Biocompatibility testing for medical textile products. In Y. Qin (Ed.), *Medical Textile Materials* (pp. 191-201). Woodhead Publishing.

Radisavljevic, A., Stojanovic, D. B., Perisic, S., Djokic, V., Radojevic, V., Rajilic-Stojanovic, M., & Uskokovic, P. S. (2018). Cefazolin-loaded polycaprolactone fibers produced via different electrospinning methods: Characterization, drug release and antibacterial effect. *European Journal of Pharmaceutical Sciences*, 124, 26-36.

Ratner, B. D. (2015). Chapter 3 - The Biocompatibility of Implant Materials. In S. F. Badylak (Ed.), *Host Response to Biomaterials* (pp. 37-51). Academic Press.

Ravikumar, R., Ganesh, M., Ubaidulla, U., Young Choi, E., & Tae Jang, H. (2017). Preparation, characterization, and *in vitro* diffusion study of nonwoven electrospun nanofiber of curcumin-loaded cellulose acetate phthalate polymer. *Saudi Pharmaceutical Journal*, 25(6), 921-926.

Rezaie, H. R., Esnaashary, M., & Öchsner, A. (2018). Classification of Drug Delivery Systems. In *A Review of Biomaterials and Their Applications in Drug Delivery* (pp. 9-25). Springer.

Rezk, A. I., Mousa, H. M., Lee, J., Park, C. H., & Kim, C. S. (2019). Composite PCL/HA/simvastatin electrospun nanofiber coating on biodegradable Mg alloy for orthopedic implant application. *Journal of Coatings Technology and Research*, 16(2), 477-489.

Rizzoli, R. (2021, 2021/01/01). Vitamin D supplementation: upper limit for safety revisited? *Aging clinical and experimental research*, 33(1), 19-24.

Rochel, N., & Molnár, F. (2017). Structural aspects of Vitamin D endocrinology. *Molecular and Cellular Endocrinology*, 453, 22-35.

Rojo, J., Sousa-Herves, A., & Mascaraque, A. (2017). 1.24 - Perspectives of Carbohydrates in Drug Discovery. In S. Chackalamannil, D. Rotella, & S. E. Ward (Eds.), *Comprehensive Medicinal Chemistry III* (pp. 577-610). Elsevier.

Rychter, M., Baranowska-Korczyc, A., Milanowski, B., Jarek, M., Maciejewska, B. M., Coy, E. L., & Lulek, J. (2018). Cilostazol-Loaded Poly(ϵ -Caprolactone) Electrospun Drug Delivery System for Cardiovascular Applications. *Pharmaceutical research*, 35(2), 32.

Salvia-Trujillo, L., Fumiaki, B., Park, Y., & McClements, D. (2017). The influence of lipid droplet size on the oral bioavailability of vitamin D₂ encapsulated in emulsions: an *in vitro* and *in vivo* study. *Food & function*, 8(2), 767-777.

Samadian, H., Salehi, M., Farzamfar, S., Vaez, A., Ehterami, A., Sahrapeyma, H., Goodarzi, A., & Ghorbani, S. (2018). *In vitro* and *in vivo* evaluation of electrospun cellulose acetate/gelatin/hydroxyapatite nanocomposite mats for wound dressing applications. *Artificial cells, nanomedicine, and biotechnology*, 1-11.

Samadian, H., Zamiri, S., Ehterami, A., Farzamfar, S., Vaez, A., Khastar, H., Alam, M., Ai, A., Derakhshankhah, H., & Allahyari, Z. (2020). Electrospun cellulose acetate/gelatin nanofibrous wound dressing containing berberine for diabetic foot ulcer healing: *in vitro* and *in vivo* studies. *Scientific Reports*, 10(1), 1-12.

Sattary, M., Rafienia, M., Kazemi, M., Salehi, H., & Mahmoudzadeh, M. (2019). Promoting effect of nano hydroxyapatite and vitamin D₃ on the osteogenic differentiation of human adipose-derived stem cells in polycaprolactone/gelatin scaffold for bone tissue engineering. *Materials Science and Engineering: C*, 97, 141-155.

Schoolaert, E., Steyaert, I., Vancoillie, G., Geltmeyer, J., Lava, K., Hoogenboom, R., & De Clerck, K. (2016). Blend electrospinning of dye-functionalized chitosan and poly (ϵ -caprolactone): towards biocompatible pH-sensors. *Journal of Materials Chemistry B*, 4(26), 4507-4516.

Selatile, M. K., Ray, S. S., Ojijo, V., & Sadiku, R. (2019). Correlations between fiber diameter, physical parameters, and the mechanical properties of randomly oriented biobased polylactide nanofibers. *Fibers and Polymers*, 20(1), 100-112.

Semnani, D., Nasari, M., & Fakhrali, A. (2018). PCL nanofibers loaded with beta-carotene: a novel treatment for eczema. *Polymer Bulletin*, 1-12.

Shahriar, S., Mondal, J., Hasan, M. N., Revuri, V., Lee, D. Y., & Lee, Y.-K. (2019). Electrospinning Nanofibers for Therapeutics Delivery. *Nanomaterials*, 9(4), 532.

Sharifi, L., Assa, F., Ajamein, H., & Mirhosseini, S. H. (2017). Effect of Voltage and Distance on Synthesis of Boehmite Nanofibers with PVP by the Electrospinning Method. *International Journal of Advanced Science and Technology*, 98, 63-74.

Sharma, S., Madhyastha, H., Swetha, K. L., Maravajjala, K. S., Singh, A., Madhyastha, R., Nakajima, Y., & Roy, A. (2021). Development of an in-situ forming, self-healing scaffold for dermal wound healing: in-vivo and in-vivo studies. *Materials Science and Engineering: C*, 112263.

Şimşek, M. (2020). Tuning surface texture of electrospun polycaprolactone fibers: Effects of solvent systems and relative humidity. *Journal of Materials Research*, 35(3), 332-342.

Sirc, J., Kubinova, S., Hobzova, R., Stranska, D., Kozlik, P., Bosakova, Z., Marekova, D., Holan, V., Sykova, E., & Michalek, J. (2012). Controlled gentamicin release from multi-layered electrospun nanofibrous structures of various thicknesses. *International journal of nanomedicine*, 7, 5315.

Somsap, J., Kanjanapongkul, K., & Tepsorn, R. (2018). Effect of parameters on the morphology and fiber diameters of edible electrospun chitosan-cellulose

acetate-gelatin hybrid nanofibers. MATEC Web of Conferences, (Vol. 192, p. 03038). EDP Sciences.

Sridhar, R., Lakshminarayanan, R., Madhaiyan, K., Barathi, V. A., Lim, K. H. C., & Ramakrishna, S. (2015). Electrosprayed nanoparticles and electrospun nanofibers based on natural materials: applications in tissue regeneration, drug delivery and pharmaceuticals. *Chemical Society Reviews*, 44(3), 790-814.

Stewart, S., Domínguez-Robles, J., Donnelly, R., & Larrañeta, E. (2018). Implantable Polymeric Drug Delivery Devices: Classification, Manufacture, Materials, and Clinical Applications. *Polymers*, 10(12), 1379.

Stewart, S. A., Domínguez-Robles, J., McIlorum, V. J., Mancuso, E., Lamprou, D. A., Donnelly, R. F., & Larrañeta, E. (2020). Development of a biodegradable subcutaneous implant for prolonged drug delivery using 3D printing. *Pharmaceutics*, 12(2), 105.

Sultanova, Z., Kaleli, G., Kabay, G., & Mutlu, M. (2016). Controlled release of a hydrophilic drug from coaxially electrospun polycaprolactone nanofibers. *International Journal of Pharmaceutics*, 505(1), 133-138.

Suwantong, O., & Supaphol, P. (2015). Applications of cellulose acetate nanofiber mats. In *Handbook of Polymer Nanocomposites. Processing, Performance and Application* (pp. 355-368). Springer.

Syed, U.-I.-Z., Li, H., & Zhu, L. (2017). Effect of Different Parameters on the Fabrication of Sustained Release Cellulose Acetate and Ethyl Cellulose Polymer Blends. *Cellulose Chemistry and Technology*, 51 (9-10), 899-909.

Taepaiboon, P., Rungsardthong, U., & Supaphol, P. (2007). Vitamin-loaded electrospun cellulose acetate nanofiber mats as transdermal and dermal therapeutic agents of vitamin A acid and vitamin E. *European Journal of Pharmaceutics and Biopharmaceutics*, 67(2), 387-397.

- Tarus, B., Fadel, N., Al-Oufy, A., & El-Messiry, M. (2016). Effect of polymer concentration on the morphology and mechanical characteristics of electrospun cellulose acetate and poly (vinyl chloride) nanofiber mats. *Alexandria Engineering Journal*, 55(3), 2975-2984.
- Tawwab, M. Y. A., Abdel-Hady, B. M., Rizk, R. A. E.-M., & Shafaa, M. W. (2019). Effect of electrospinning parameters on the versatile production of polycaprolactone/gelatin nanofiber mats. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, 10(2), 025009.
- Temova, Ž., & Roškar, R. (2016). Stability-indicating HPLC–UV method for vitamin D₃ determination in solutions, nutritional supplements and pharmaceuticals. *Journal of chromatographic science*, 54(7), 1180-1186.
- Topuz, F., Satilmis, B., & Uyar, T. (2019). Electrospinning of uniform nanofibers of Polymers of Intrinsic Microporosity (PIM-1): The influence of solution conductivity and relative humidity. *Polymer*, 178, 121610.
- Torres-Martínez, E. J., Bravo, J. M. C., Medina, A. S., González, G. L. P., & Gómez, L. J. V. (2018). A Summary of Electrospun Nanofibers as Drug Delivery System: Drugs Loaded and Biopolymers Used as Matrices. *Current drug delivery*, 15(10), 1360-1374.
- Tsai, S.-Y., Lin, H.-Y., Hong, W.-P., & Lin, C.-P. (2017). Evaluation of preliminary causes for vitamin D series degradation via DSC and HPLC analyses. *Journal of Thermal Analysis and Calorimetry*, 130(3), 1357-1369.
- Tuburan, C., Rosa, L. D., & Reyes, L. (2017). Silver-Loaded Cellulose Acetate-g-Poly (ε-caprolactone) Composites. IOP Conference Series: Materials Science and Engineering, (Vol. 205, No. 1, p. 012008). IOP Publishing.
- Tungprapa, S., Jangchud, I., & Supaphol, P. (2007). Release characteristics of four model drugs from drug-loaded electrospun cellulose acetate fiber mats. *Polymer*, 48(17), 5030-5041.

- Ullah, A., Saito, Y., Ullah, S., Haider, M. K., Nawaz, H., Duy-Nam, P., Kharaghani, D., & Kim, I. S. (2021). Bioactive Sambong oil-loaded electrospun cellulose acetate nanofibers: Preparation, characterization, and *in-vitro* biocompatibility. *International Journal of Biological Macromolecules*, 166, 1009-1021.
- Ullah, A., Ullah, S., Khan, M. Q., Hashmi, M., Nam, P. D., Kato, Y., Tamada, Y., & Kim, I. S. (2020). Manuka honey incorporated cellulose acetate nanofibrous mats: Fabrication and *in vitro* evaluation as a potential wound dressing. *International Journal of Biological Macromolecules*, 155, 479-489.
- Vatankhah, E. (2018). Rosmarinic acid-loaded electrospun nanofibers: *In vitro* release kinetic study and bioactivity assessment. *Engineering in Life Sciences*, 18(10), 732-742.
- Vatankhah, E., Prabhakaran, M. P., Jin, G., Mobarakeh, L. G., & Ramakrishna, S. (2014). Development of nanofibrous cellulose acetate/gelatin skin substitutes for variety wound treatment applications. *Journal of biomaterials applications*, 28(6), 909-921.
- Vlachou, M., Kikionis, S., Siamidi, A., Tragou, K., Ioannou, E., Roussis, V., & Tsotinis, A. (2019). Modified *In Vitro* Release of Melatonin Loaded in Nanofibrous Electrospun Mats Incorporated Into Monolayered and Three-Layered Tablets. *Journal of pharmaceutical sciences*, 108(2), 970-976.
- Wahab, J. A., Ogasawara, H., Kim, I. S., & Ni, Q.-Q. (2020). Cellulose acetate/multi-wall carbon nanotube/Ag nanofiber composite for antibacterial applications. *Materials Science and Engineering: C*, 110679.
- Walia, N., Dasgupta, N., Ranjan, S., Chen, L., & Ramalingam, C. (2017). Fish oil based vitamin D nanoencapsulation by ultrasonication and bioaccessibility analysis in simulated gastro-intestinal tract. *Ultrasonics sonochemistry*, 39, 623-635.

Wan, Y., Lin, Z., Gan, D., Cui, T., Wan, M., Yao, F., Zhang, Q., & Luo, H. (2019, 2019/08/01). Effect of Graphene Oxide Incorporation into Electrospun Cellulose Acetate Scaffolds on Breast Cancer Cell Culture. *Fibers and Polymers*, 20(8).

Wang, B., Lv, X., Chen, S., Li, Z., Sun, X., Feng, C., Wang, H., & Xu, Y. (2016). *In vitro* biodegradability of bacterial cellulose by cellulase in simulated body fluid and compatibility in vivo. *Cellulose*, 23(5), 3187-3198.

Wang, D., Yue, Y., Wang, Q., Cheng, W., & Han, G. (2020). Preparation of cellulose acetate-polyacrylonitrile composite nanofibers by multi-fluid mixing electrospinning method: morphology, wettability, and mechanical properties. *Applied Surface Science*, 145462.

Wei, L., Sun, R., Liu, C., Xiong, J., & Qin, X. (2019). Mass production of nanofibers from needleless electrospinning by a novel annular spinneret. *Materials & Design*, 179, 107885.

Wimalawansa, S. J. (2018). Vitamin D and cardiovascular diseases: Causality. *The Journal of Steroid Biochemistry and Molecular Biology*, 175, 29-43.

Woodruff, M. A., & Hutmacher, D. W. (2010). The return of a forgotten polymer—Polycaprolactone in the 21st century. *Progress in Polymer Science*, 35(10), 1217-1256.

Wsoo, M. A., Abd Razak, S. I., Bohari, S. P. M., Shahir, S., Salihu, R., Kadir, M. R. A., & Nayan, N. H. M. (2021). Vitamin D₃-loaded electrospun cellulose acetate/polycaprolactone nanofibers: Characterization, *in-vitro* drug release and cytotoxicity studies. *International Journal of Biological Macromolecules*, 181, 82-98.

Wsoo, M. A., Shahir, S., Bohari, S. P. M., Nayan, N. H. M., & Abd Razak, S. I. (2020). A review on the properties of electrospun cellulose acetate and its application

in drug delivery systems: A new perspective. *Carbohydrate Research*, 491, 107978.

Wutticharoenmongkol, P., Hannirojram, P., & Nuthong, P. (2019). Gallic acid-loaded electrospun cellulose acetate nanofibers as potential wound dressing materials. *Polymers for advanced technologies*, 30(4), 1135-1147.

Xue, J., He, M., Liang, Y., Crawford, A., Coates, P., Chen, D., Shi, R., & Zhang, L. (2014). Fabrication and evaluation of electrospun PCL-gelatin micro/nanofiber membranes for anti-infective GTR implants. *Journal of Materials Chemistry B*, 2(39), 6867-6877.

Yang, G., Li, X., He, Y., Ma, J., Ni, G., & Zhou, S. (2018). From nano to micro to macro: Electrospun hierarchically structured polymeric fibers for biomedical applications. *Progress in Polymer Science*, 81, 80-113.

Yang, Q., Li, Z., Hong, Y., Zhao, Y., Qiu, S., Wang, C., & Wei, Y. (2004). Influence of solvents on the formation of ultrathin uniform poly (vinyl pyrrolidone) nanofibers with electrospinning. *Journal of Polymer Science Part B: Polymer Physics*, 42(20), 3721-3726.

Yang, Y., Chang, S., Bai, Y., Du, Y., & Yu, D.-G. (2020). Electrospun triaxial nanofibers with middle blank cellulose acetate layers for accurate dual-stage drug release. *Carbohydrate Polymers*, 243, 116477.

Yang, Y., Li, W., Yu, D.-G., Wang, G., Williams, G. R., & Zhang, Z. (2019). Tunable drug release from nanofibers coated with blank cellulose acetate layers fabricated using tri-axial electrospinning. *Carbohydrate Polymers*, 203, 228-237.

Yu, D.-G., Li, X.-Y., Wang, X., Chian, W., Liao, Y.-Z., & Li, Y. (2013). Zero-order drug release cellulose acetate nanofibers prepared using coaxial electrospinning. *Cellulose*, 20(1), 379-389.

- Zamani, M., Morshed, M., Varshosaz, J., & Jannesari, M. (2010). Controlled release of metronidazole benzoate from poly ϵ -caprolactone electrospun nanofibers for periodontal diseases. *European Journal of Pharmaceutics and Biopharmaceutics*, 75(2), 179-185.
- Zareie M, A. A., Faghah S. (2019, sep, 2019). Thermal Stability and Kinetic Study on Thermal Degradation of Vitamin D₃ in Fortified Canola Oil. *Journal of food science and technology*, 84(9), 2475-2481.
- Zareie, M., Abbasi, A., & Faghah, S. (2019). Thermal Stability and Kinetic Study on Thermal Degradation of Vitamin D₃ in Fortified Canola Oil. *Journal of food science*, 84(9), 2475-2481.
- Zavan, B., Gardin, C., Guarino, V., Rocca, T., Cruz Maya, I., Zanotti, F., Ferroni, L., Brunello, G., Chachques, J.-C., Ambrosio, L., & Gasbarro, V. (2021). Electrospun PCL-Based Vascular Grafts: *In Vitro* Tests. *Nanomaterials*, 11(3).
- Zhan, S., Wang, J., Wang, W., Cui, L., & Zhao, Q. (2019). Preparation and *in vitro* release kinetics of nitrendipine-loaded PLLA–PEG–PLLA microparticles by supercritical solution impregnation process. *RSC Advances*, 9(28), 16167-16175.
- Zhang, S., Campagne, C., & Salaün, F. (2019). Preparation of electrosprayed poly (caprolactone) microparticles based on green solvents and related investigations on the effects of solution properties as well as operating parameters. *Coatings*, 9(2), 84.
- Zhang, Y., Deng, C., Qu, B., Zhan, Q., & Jin, X. (2017). A study on wet and dry tensile properties of wood pulp/lyocell wetlace nonwovens. IOP Conference Series: Materials Science and Engineering, (Vol. 241, No. 1, p. 012013). IOP Publishing.

Zhang, Y., Yin, C., Cheng, Y., Huang, X., Liu, K., Cheng, G., & Li, Z. (2020). Electrospinning Nanofiber-Reinforced Aerogels for the Treatment of Bone Defects. *Advances in Wound Care*, 9(8), 441-452.

Zhijiang, C., Yi, X., Haizheng, Y., Jia, J., & Liu, Y. (2016). Poly(hydroxybutyrate)/cellulose acetate blend nanofiber scaffolds: Preparation, characterization and cytocompatibility. *Materials Science and Engineering: C*, 58, 757-767.

Zukowski, B., dos Santos Mendonça, Y. G., de Andrade Silva, F., & Toledo Filho, R. D. (2020). Effect of moisture movement on the tensile stress-strain behavior of SHCC with alkali treated curauá fiber. *Materials and Structures*, 53, 1-11.

LIST OF PUBLICATIONS

1. Wsoo, M. A., Shahir, S., Bohari, S. P. M., Nayan, N. H. M., & Abd Razak, S. I. (2020). A review on the properties of electrospun cellulose acetate and its application in drug delivery systems: A new perspective. *Carbohydrate Research*, 491, 107978. (Q2, IF: 1.87).
2. Wsoo, M. A., Abd Razak, S. I., Bohari, S. P. M., Shahir, S., Salihu, R., Kadir, M. R. A., & Nayan, N. H. M. (2021). Vitamin D₃-loaded electrospun cellulose acetate/polycaprolactone nanofibers: Characterization, *in-vitro* drug release and cytotoxicity studies. *International Journal of Biological Macromolecules*, 81, 82-98 (Q1, IF: 6.95).
3. Wsoo, M. A., Abd Razak, S. I., Shahir, S., Ahmed Abdullah Al-Moalemi, H., Rafiq Abdul Kadir, M., & Hasraf Mat Nayan, N. (2021). Development of prolonged drug delivery system using electrospun cellulose acetate/polycaprolactone nanofibers: Future subcutaneous implantation. *Polymers for Advanced Technologies*, 32(9), 3664-3678. (Q2, IF 3.66).