MICROBIAL COMMUNITY PROFILING OF ARSENIC-RICH MINE TAILING AND ARSENIC BIOADSORPTION BY INDIGENOUS BACTERIA

WAHID ALI HAMOOD AL-TOWAYTI

UNIVERSITI TEKNOLOGI MALAYSIA

MICROBIAL COMMUNITY PROFILING OF ARSENIC-RICH MINE TAILING AND ARSENIC BIOADSORPTION BY INDIGENOUS BACTERIA

WAHID ALI HAMOOD AL-TOWAYTI

A thesis submitted in fulfilment of the requirements for the award of the degree of Doctor of Philosophy

> Faculty of Science Universiti Teknologi Malaysia

> > MARCH 2020

ACKNOWLEDGEMENT

First and foremost, all praise be to Allah the Almighty, thanks to Him for giving me the opportunity and will to finish this research and to complete this thesis. I would like to express my sincere appreciation and gratitude to my research supervisor, Associate Professor Dr. Shafinaz Shahir for her knowledge, acquaintance, guidance, supervision, critics, evaluation, encouragement, and for supporting me throughout the undertaking of this thesis. I am also very thankful to my senior friend, Hassan Amr Algaifi for giving me the motivation, assistance, information, opinion and for his kind contribution for helping me to finish my thesis.

A thousand thanks also to all of the staff at bioscience department, faculty of Sciences for helping me during this research, particularly, the lab assistants. Also, I will not forget to thank Associate Professor Dr. Sallehuddin Ibrahim and his family for the motivation, support, encouragement, friendship, advice and understanding. I greatly appreciate it.

I want to extend my utmost gratitude and appreciation to my parents, family, fellow friends and those who provided assistance in this research either intentionally or unintentionally at various conditions and occasions during the progress of this research and the completion of this thesis. All of their support towards the completion of this thesis will be reciprocated by Allah the Almighty. Without their support and contributions, this thesis would not have been the same as presented here. Thank you very much.

ABSTRACT

Arsenic is a common contaminant in gold mine soil and tailings. Moreover, the contamination of water with arsenic is a serious health issue. Microbes present an opportunity to remove arsenic from wastewater via adsorption process, which is distinguished by its low cost and easy technique in comparison with conventional techniques include oxidation, coagulation-flocculation, and membrane techniques. However, the development of existing bio-treatment approaches depends on isolation of arsenic-resistant microbes from arsenic contaminated samples. In this study, a culture-independent approach using Illumina sequencing technology was used to profile the microbial community *in situ*. This was coupled with a culture-dependent technique to analyse the microbial population in arsenic-laden tailing dam sludge based on the culture-independent sequencing approach. Based on the cultureindependent sequencing approach, 4 phyla and 8 genera were identified in a sample from the arsenic-rich goldmine. Firmicutes (92.23%) was the dominant phylum, followed by Proteobacteria (3.21%), Actinobacteria (2.41%), and Bacteroidetes (1.49%). The identified genera included *Staphylococcus* (89%), *Pseudomonas* (1.25%), *Corynebacterium* (0.82%), *Prevotella* (0.54%), *Pseudonocardia* (0.39%), *Megamonas* (0.38%) and S*phingomonas* (0.36%). The culture dependent method exposed significant similarities with culture independent methods at the phylum level with Firmicutes, Proteobacteria and Actinobacteria, being common, and Firmicutes was the dominant phylum whereas, at the genus level, only *Pseudomonas* was presented by both methods. Considering the advantage of the different structures of these bacterial cell walls in adsorption, attempts were made to use individual dried biomass of *Bacillus thuringiensis* strain WS3 (IDB) and mixed dried biomass of three species *B. thuringiensis* strain WS3, *Pseudomonas stutzeri* strain WS9 and *Micrococcus yunnanensis* strain WS11 (MDB) to achieve highest As (III) and As (V) removal under different conditions. Successively, MDB were found to be efficient in the removal of As (III) and As (V) up to 95 % and 98 %, respectively. The maximum adsorption capacity of As (III) and As (V) increased from 95 mg/g and 145 mg/g for IDB to 217 mg/g and 333 mg/g for MDB as obtained from the Langmuir isotherm. The pattern of adsorption fitted well with the Langmuir isotherm model and kinetic data followed a pseudo-second-order model for both IDB and MDB. The thermodynamic parameters *∆G°*, *∆H°* and *∆S°* revealed that the adsorptions of both As (III) and As (V) were spontaneous, feasible and endothermic in nature. FESEM-EDX analysis established diverse cell morphological changes with significant amounts of arsenic adsorbed onto biomass compared to original biomass. Results from FTIR have shown the involvement of mainly hydroxyl, thiol, amide and amino functional groups in the arsenic adsorption. Batch experimental data were taken into account to create an artificial neural network (ANN) model that mimicked the human brain function. 5-7-1 neurons were in the input, hidden and output layers respectively. The batch data was reserved for training (75%), testing (10%) and validation process (15%). The predicted output of the proposed model showed a good agreement with the batch experiments with reasonable accuracy. This study has demonstrated the potential for using mixed dried non-living biomass as a new biosorbent for arsenic removal.

ABSTRAK

Arsenik adalah pencemar biasa di tanah dan amang lombong emas. Selain itu, pencemaran air dengan arsenik merupakan masalah kesihatan yang serius. Mikrob menyediakan peluang menyingkirkan arsenik dari air sisa melalui proses penjerapan, yang dibezakan oleh kos rendah dan teknik mudahnya berbanding dengan teknik konvensional termasuk teknik pengoksidaan, koagulasi-flokulasi, dan membran. Walau bagaimanapun, perkembangan pendekatan bio-rawatan yang sedia ada bergantung kepada pemencilan mikrob rintang arsenik daripada sampel yang tercemar arsenik. Dalam kajian ini, pendekatan bebas kultur menggunakan teknologi penjujukan Illumina digunakan untuk memprofilkan komuniti mikrob in situ. Pendekatan ini digandingkan dengan teknik bergantung kultur, iaitu pemencilan menggunakan dua media pertumbuhan berbeza, LB dan CDM untuk menganalisis populasi mikrob dalam enap cemar empangan amang muatan arsenik berdasarkan pendekatan penjujukan bebas kultur. Berdasarkan pendekatan penjujukan bebas kultur, 4 fila dan 8 genus dikenal pasti dalam sampel dari lombong emas yang kaya dengan arsenik. Firmicutes (92.23%) merupakan filum dominan, diikuti oleh Proteobakteria (3.21%), Aktinobakteria (2.41%), dan Bakteroidetes (1.49%). Genus yang dikenal pasti termasuk *Staphylococcus* (89%), *Pseudomonas* (1.25%), *Corynebacterium* (0.82%), *Prevotella* (0.54%), *Pseudonocardia* (0.39%), *Megamonas* (0.38%) dan *Sphingomonas* (0.36%). Kaedah bergantung kultur mendedahkan bahawa terdapat persamaan yang signifikan dengan kaedah bebas kultur pada tahap filum dengan Firmikutes, Proteobakteria dan Aktinobakteria sebagai yang biasa, dan Firmikutes sebagai filum dominan manakala pada tahap genus, hanya *Pseudomonas* yang ditunjukkan oleh kedua-dua kaedah. Mempertimbangkan kelebihan struktur yang berbeza pada dinding sel bakteria-bakteria ini dalam penjerapan, percubaan dibuat untuk menggunakan biojisim kering *Bacillus thuringiensis* strain WS3 (IDB) dan campuran biojisim kering tiga spesies iaitu *Bacillus thuringiensis* strain WS3, *Pseudomonas stutzeri* strain WS9 dan *Micrococcus yunnanensis* strain WS11 (MDB) untuk mencapai penyingkiran tertinggi As (III) dan As (V) pada keadaan yang berbeza. Seterusnya, MDB didapati berkesan dalam penyingkiran As (III) dan As (V) masingmasing sebanyak 95% dan 98%. Kapasiti penjerapan maksimum As (III) dan As (V) meningkat daripada 95 mg/g dan 145 mg/g bagi IDB kepada 217 mg/g dan 333 mg/g untuk MDB seperti yang diperoleh daripada isoterm Langmuir. Corak penjerapan menepati dengan baik model isoterm Langmuir dan data kinetik mengikut model tertib pseudo-kedua bagi IDB dan MDB. Parameter termodinamik *ΔG°*, *ΔH°* dan *ΔS°* mendedahkan bahawa penjerapan kedua-dua As (III) dan As (V) adalah spontan, boleh dilaksanakan dan endotermik. Analisis FESEM-EDX menunjukkan perubahan morfologi sel pelbagai dengan jumlah arsenik yang signifikan diserap ke biojisim berbanding dengan biojisim asal. Keputusan FTIR menunjukkan perubahan spektral utama pada kawasan jalur 400 cm⁻¹ hingga 4000 cm⁻¹ menunjukkan penglibatan kumpulan-kumpulan berfungsi terutamanya hidroksil, tiol, amida dan amino dalam penjerapan arsenik. Data eksperimen berkelompok diambil kira untuk membina model rangkaian neural buatan (ANN) yang meniru fungsi otak manusia. Neuron 5-7-1 masing-masing berada dalam lapisan input, lapisan tersembunyi dan lapisan output. Data berkelompok disimpan untuk latihan (75%), ujian (10%) dan proses pengesahan (15%). Output ramalan model yang dicadangkan menunjukkan persetujuan yang baik dengan eksperimen berkelompok dengan ketepatan yang munasabah. Kajian ini menunjukkan potensi untuk menggunakan biojisim kering tidak hidup sebagai biopanjerap baharu untuk penyingkiran arsenik.

TABLE OF CONTENTS

TITLE PAGE

CHAPTER 2 [LITERATURE REVIEW](#page--1-0) 9 2.1 [Arsenic Sources](#page--1-1) 9 2.2 [Arsenic Contamination](#page--1-2) 9

- 7.3 [The Biomass Adsorption Capacity](#page--1-82) 113
- 7.4 [Isotherm Studies](#page--1-40) 114

LIST OF TABLES

LIST OF FIGURES

LIST OF ABBREVIATIONS

LIST OF SYMBOLS

LIST OF APPENDICES

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Metagenomics (culture-independent) is a relatively new, yet a rapidly developing technology to analyse and characterize microbial communities in the environment. Microorganisms are an important aspect of ecological processes; helping for biogeochemical cycling for important elements such as sulfur, nitrogen, phosphorous, and carbon, decay of organic substance and xenobiotics and formation of soil structure. Thus, bacteria play a significant role in regulating the biogeochemical cycles and affect life on Earth (McHardy and Rigoutsos, 2007; Smith *et al.*, 2015). Studies on bacterial dynamics and their interaction with the abiotic and biotic elements are essential to understanding their involvement in energy generation, mining, bio-treatment, and biotechnology (Rastogi and Sani, 2011; Santoyo *et al.*, 2017). A well-ordered microbial community with a firm level of diversity is stable (Wu *et al.*, 2018; Yannarell and Triplett, 2005). However, when presented with some kind of stress, the diversity of the community might change, thus collapsing its stability. Therefore, microbial diversity used to study the effect of perturbations in the environment. In the regular environment, microorganisms occur in great numbers despite the fact that there are millions of bacterial species that have not been identified. Therefore, pure seawater might contain 10⁶ bacteria per millilitre and one gram of soil or sediment might contain approximately 10^{10} bacteria as calculated via fluorescence microscopy after staining with a dye (Fakruddin and Mannan, 2013; Torsvik *et al.*, 1990).

Recently, new non-cultural approaches have been developed that can be used extensively in a microbial consortium for comprehensive analysis of various communities (Lugli *et al.*, 2019; Mahajan *et al.*, 2018). Metagenomics or genomic microorganism studies refer to a non-cultural approach in which the genomes of a mixed microbe population are collectively studied. Population genomics, environmental genomics and Community genomics are frequently used as metagenomics synonyms (Neelakanta and Sultana, 2013). Since the use of the metagenomics method in this research, several other studies have used this technique to study microbial communities in different environments such as acid-mine drainage (Tyson *et al.*, 2004), marine water and sediments (DeLong *et al.*, 2006; Yooseph *et al.*, 2010) and arsenic-contaminated soils (Layton *et al.*, 2014; Luo *et al.*, 2014).

Detailed insight into microbial communities in arsenic contaminated water in the natural environment is challenging owing to their extreme conditions and uncultivated status (Das *et al.*, 2017). However, a large variety of metagenomies of microbial communities in arsenic contaminated water have been reported include: *Helicobacter pylori* (0.01%), *Campylobacter jejuni* (0.01%), *Staphylococcus aureus* (0.02%), *Shigella flexneri* and *Shigella dysenteriae* (0.03%), *Cronobacter sakazakii* (0.03%), *Clostridium difficile* (0.03%), *Salmonella enterica* (0.07%), *Vibrio cholerae* (0.08%) and *Vibrio parahaemolyticus* (0.08%) (Layton*, et al.*, 2014). Furthermore, a great diversity of arsenic-resistant microbes have been stated, including *Bacillus* sp. and *Aneurinibacillus aneurinilyticus* (Dey *et al.*, 2016); *Acinetobacter calcoaceticus*, *A. baumannii*, *A. junii*, *A. venetianus*, *A. soli*, and *Microbacterium oleivorans* (Goswami *et al.*, 2015); *Enterobacter* sp. and *Klebsiella pneumoniae* (Abbas *et al.*, 2014); *Bacillus smithii*, *B. cereus*, *Pseudomonas maltophilia*, *Vibrio parahaemolyticus*, *Pseudomonas* sp, *Micrococcus varians*, *M. luteus*, and *M. roseus* (Shakya *et al.*, 2012); *Geobacillus kaustophilus* (Cuebas *et al.*, 2011), *Bacillus* sp., *Enterobacter* sp., *Stenotrophomonas* sp., and *Rhizobium* (Tiwari *et al.*, 2016); *P.* strain As-11(Jebelli *et al.*, 2017); *B. cereus* strain SZ2 (Bahari *et al.*, 2013), and *Microbacterium* sp. strain SZ (Bahari *et al.*, 2017). Furthermore, there are microbes that can adapt to diverse environmental conditions on earth and decay chemical components produced by living things (Fakruddin and Mannan, 2013).

Arsenic is generally distributed in Earth's crust. It is leaked by common phenomena such as mineral weathering or volcanic ash and human activities such as gold mining and various resources (Cullen and Reimer, 1989; Smedley and Kinniburgh, 2002; Tamaki and Frankenberger, 1992). Arsenic can be found in the

environment as arsenious acids $(H_3AsO_3^{2+}, H_3AsO_4, H_3AsO_3)$. Moreover, As (V) is like a soft acid and can form a complex with sulfides. On the other hand, As (III) is a firm acid which makes a compound with nitrogen and oxides (Mohan and Pittman, 2007). However, there are large numbers of arsenic contaminated areas with high concentrations of arsenic around the world, especially in Argentina, Chile, Mexico, China, Hungary, West Bengal (India), Bangladesh, Vietnam, and the USA (Herath *et al.*, 2016). There are generally 15 gold mines in Malaysia with large gold mines in Pahang (Penjom, Raub and Selinsing gold mines), Kelantan and Terengganu. Therefore, it is very likely that arsenic concentrations in these areas are high.

Microbes have co-habited with different metals from initial history. Thus, microorganisms have been effectively used to remove heavy metal such as Arsenic (As) from wastewater in a variety of patterns. Consequently, from a functional concept, metals divided into three groups: (i) non-toxic and essential such as Mg and Ca, (ii) harmful at high concentrations and essential in low concentration such as Zn, Mo, Cu, Ni, Co, Fe, and Mn, and (iii) toxic even in low concentration such as Cd, Hg and As. In addition, interaction with metals relies on specific metal and its chemical speciation (Valls and De Lorenzo, 2002). The basic mechanism of adsorption by biomass can be described as passive metal ions immobilization. Briefly, it essentially relies on the physicochemical interaction between metals and different functional groups of the cell wall. Microorganisms have been effectively used to remove arsenic from wastewater (Bahari*, et al.*, 2013; Haris *et al.*, 2018; Kao *et al.*, 2013; Prasad *et al.*, 2013). Likewise, the cell wall of bacteria generally comprises proteins, lipids and polysaccharides, which contain functional groups, such as amine groups, phosphate, hydroxyl and carboxylate, and these functional groups offer binding sites for metals (Mohan and Pittman, 2007).

Artificial Neural Network (ANN) is classified as an artificial intelligence modelling technique because of its ability to recognize patterns and relationships in historical data and then to deduce new data (Aleboyeh *et al.*, 2008). The ANN uses a specified algorithm to analyse data cases or similarity patterns and then divides them into a defined class number. In addition, the ANN learns to accurately predict the output parameter value when data with adequate input parameters are given (Yetilmezsoy and Demirel, 2008). Process models and model-based process monitoring are the main applications of ANN in the water treatment industry (Shetty and Chellam, 2003). Therefore, adsorption results can be predicted using the artificial neural network (ANN), as ANN can efficiently map inputs and outputs in complex situations (Aleboyeh*, et al.*, 2008; Annadurai *et al.*, 2007; Chu, 2003; Saha *et al.*, 2010; Texier *et al.*, 2002; Yetilmezsoy and Demirel, 2008).

1.2 Problem Statement

Globally, arsenic contamination in groundwater is presently a major problem, particularly in areas where people depend on groundwater. Poisonous arsenic has resulted in health disasters for over 100 million people universally, mainly in China, India, Bangladesh, Taiwan, Thailand, Chili and Romania (Miyatake and Hayashi, 2009; Shahid *et al.*, 2018; Singh *et al.*, 2007; Tabassum *et al.*, 2019). The Environmental Protection Agency of the United States declared that all forms of arsenic pose a serious health risk (Sarkar *et al.*, 2007). Accordingly, the recommended concentration of arsenic in drinking water has since been modified from 50 ppb (0.05 mg/L) to 10 ppb (0.01 mg/L) by the World Health Organization (WHO), while the standard concentration of arsenic in industrial effluents is restricted to 0.1 mg/L (0.1 ppm) (Wu *et al.*, 2010). Whereas, the Malaysian Environment Department reported in 1985 that drinking water quality standard for maximum arsenic concentration is 0.01 mg /L (Huang *et al.*, 2015).

Chronic arsenic poisoning causes skin lesions with hyperkeratosis, depigmentation, and hyperpigmentation (Sun, 2004; Yoshida *et al.*, 2004), vascular diseases, such as cardiovascular, arteriosclerosis and hypertension (Rahman *et al.*, 1999; Wang *et al.*, 2002; Yu *et al.*, 2002) and non-specific signs of the effect on the digestive system, such as dyspepsia, diarrhoea and abdominalgia (Sun *et al.*, 2001) and also has extensive and complex effects on developing infants, such as poor memory, mental slowing, cognitive delays and reduced intelligent quotient (IQ) (Chattopadhyay *et al.*, 2002). Therefore, arsenic contamination of water has become a serious problem for the community (Hao *et al.*, 2018; Nidheesh and Singh, 2017;

Zhang *et al.*, 2018). Currently, there is no medical treatment for arsenicosis and the only guaranteed way of preventing chronic arsenic poisoning is to stop the ingestion of arsenic (Sun *et al.*, 2006).

Several researchers identified biotreatment of arsenic ions from contaminated water by living organisms as a viable solution for the removal of these contaminants (Ike *et al.*, 2008; Lu *et al.*, 2018; Pandey and Bhatt, 2015), whereas the effective biotreatment depends on our ability to study microbes that are indigenous to polluted sites regardless of the approach taken (Stefani *et al.*, 2015). Therefore, in this study, culture independent was combined with culture-dependent methods to isolate indigenous microbes using soil samples harvested from tailing dam sludge because it contains the highest concentration of arsenic in the gold mining environment. Consequently, the removing of arsenic from contaminated water is necessary for confirming the safety of drinking water and protect public health (Nickson *et al.*, 2000; Zaini *et al.*, 2011). Some of the conventional solutions to remove As (III) and As (V) from wastewater are filtration, flotation, flocculation with sulfide or ferric hydroxide and ion exchange. However, these techniques require pre-treatment, the oxidation of As (III) to As (V) and involve a high cost (Valls and De Lorenzo, 2002).

1.3 Contributions to Knowledge

This is the first study on biodiversity of microbes for Malaysian gold mining environment that use independent and dependent approaches. Moreover, employment of culture based approach with metagenomics analysis helps to isolate indigenous arsenic resistant microbes and their potential use in bioremediation of arsenic contaminated sites. Furthermore, individual and mixed culture dried biomass of indigenous arsenic resistant microbial (WS3, WS9, WS11) have been used after regeneration by acid washing to enhance the removing of As (III) and As (V). Mixed dried biomass of WS3, WS9 and WS11 was found to be efficient in removing As (III) and As (V) due to the benefits of the distinct structures of these bacterial cell walls in adsorption.

The experimental data was then taken into account to develop an artificial neural network (ANN) model; the ANN model mimicked the function of the human brain, to predict the removal of As (III) and As (V) from aqueous solution by adsorption process. Moreover, mixed dried biomass of three indigenous arsenic resistant bacteria (WS3, WS9, and WS11) is distinguished by its low cost and high capacities for bio-treatment of arsenic from wastewater. Consequently, the adsorptions of As (III) and As (V) ions using the above microbial mix have not been reported elsewhere. Hence it was considered the best choice to use these microbes for adsorption of these two ions. The novelty of this work is to use new indigenous arsenic resistant microbes for the removal of As (III) and As (V) and the results are compared with a model to find the validity of the experimental results.

1.4 Objectives

1) To investigate the microbial communities' in soil contaminated by arsenic using culture independent strategy (metagenomics approach) and isolate and characterize the indigenous arsenic resistant bacteria (culture dependent approach), according to the metagenomics profile obtained and assess the reusability of the bacterial biomass by acid washing (regeneration study).

2) To study the adsorption of As (III) and As (V) using individual and mix dried bacterial biomass, by varying the various process parameters e.g initial contact time, arsenic concentration, pH, temperature and adsorbent dose.

3) To evaluate the mechanism of As (III) and As (V) adsorption using various adsorption isotherms, kinetic and thermodynamic models and characterized the indigenous arsenic resistant bacteria biomass before and after adsorption.

4) To predict and compare the removal efficiency of As (III) and As (V) from aqueous solution by individual and mixed dried biomass of indigenous arsenic resistant bacteria using artificial neural network (ANN) model.

6

1.5 Scope of Study

In this study, we combined metagenomics with culture-dependent methods to isolate indigenous microbes from high arsenic contaminated soil samples harvested from a tailing dam sludge in one gold mine in Pahang (Selinsing Gold Mine) using two different culturing media (LB and CDM with 2mM As (III) or 5 mM As (V). Furthermore, this study was proposed a suitable conventional pre-treatment technology of indigenous arsenic resistant bacterial biomass to increase arsenic removal efficiency from wastewater and regenerate the biomass. Moreover, this study is determined the efficacy of individual and mixed dried biomass of three strains WS3, WS9 and WS11 in the removal of As (III) and As (V). The present study aimed to evaluate the isolated indigenous biomass in order to remove As (III) and As (V) through experimental and theoretical (mathematical modelling) studies.

In the present study, the ANN model mimicked the function of the human brain, which has billions of neurons. These neurons are connected to each other through pathways that transmit electronic signals. These connections enable the neurons to send or receive electrical impulses, which in turn are responsible for the brain function. Likewise, ANN has the capability of mapping inputs and outputs professionally. The ANN consists of an input layer, a hidden middle layer, and an output layer. The hidden and output layers are composed of computational nodes called neurons, and one-layer neurons are connected to the neurons of the preceding layer by means of weights, which regulate the connection between two neurons. The neurons use differentiated activation functions to generate output by transferring weighted input from the previous layer (Prasenjit *et al.*, 2012). The inputs included contact time, arsenic concentration, pH, temperature and adsorbent dosage. On the contrary, the output of the model was the predicted removal of arsenic (ppm). MATLAB2017b function was utilized to create the model. The process of the model was categorized into three steps, which were training, testing, and validation. In addition, the model was also verified with experimental data to evaluate the outcome vector using statistics indicators such as mean square error and correlation coefficient. In addition, Isotherm, kinetic and thermodynamic studies were applied to analyse the mechanisms of arsenic adsorption. The biomass of indigenous arsenic

resistant bacteria was characterized before and after arsenic adsorption by using FESEM–EDAX and FTIR analyses.

1.6 Significance of the Study

Currently, the metagenomics method is regarded as the most efficient, reliable, rapid and accurate way to reveal the entire microbial composition of a community under complex environment conditions. Also, culture dependent method is used to complement the microbial biodiversity and this extends our knowledge of microbial diversity in a gold mining environment. A deep and direct insight into the soil biodiversity and microbial community and its functions can be investigated by using culture independent and dependent methods. In addition, employment of culture based approach with metagenomics analysis helps to know the biodiversity of microbes from Malaysian gold mining environment and isolate indigenous arsenic resistant microbes and their potential use in bioremediation of arsenic contaminated sites.

Until now a few reports have been put forward studying water, sediment, contaminated environments with relatively different concentrations of arsenic by both methods (Luo*, et al.*, 2014). Therefore, an alternative technique to remove arsenic from wastewater is in demand. Considerable efforts have been devoted to overcoming this serious issue by using individual and mixed dried biomass of indigenous arsenic resistant microbial as an adsorbent to remove arsenic. The biomass is distinguished by its low cost and high capacities. Furthermore, a majority of previous research focused primarily on laboratory and experimental works and they suffer from a lack of modeling in order to accurately predict the experimental behaviour of As (III) and As (V) removal by biomass. Moreover, the experimental results were compared with a model to find the validity of the experimental adsorption results.

REFERENCES

- Abbas, S. Z., Riaz, M., Ramzan, N., Zahid, M. T., Shakoori, F. R. and Rafatullah, M. (2014). Isolation and characterization of arsenic resistant bacteria from wastewater. *Brazilian Journal of Microbiology*, 45(4), 1309-1315.
- Abdul, K. S. M., Jayasinghe, S. S., Chandana, E. P., Jayasumana, C. and De Silva, P. M. C. (2015). Arsenic and human health effects: A review. *Environmental Toxicology and Pharmacology*, 40(3), 828-846.
- Abejón, A., Garea, A. and Irabien, A. (2015). Arsenic removal from drinking water by reverse osmosis: Minimization of costs and energy consumption. *Separation and Purification Technology*, 144, 46-53.
- Aber, S., Amani-Ghadim, A. and Mirzajani, V. (2009). Removal of Cr (VI) from polluted solutions by electrocoagulation: Modeling of experimental results using artificial neural network. *Journal of Hazardous Materials*, 171(1-3), 484-490.
- Adams, M. (2016). Summary of gold plants and processes *Gold Ore Processing (Second Edition)* (pp. 961-984): Elsevier.
- Afkar, E., Lisak, J., Saltikov, C., Basu, P., Oremland, R. S. and Stolz, J. F. (2003). The respiratory arsenate reductase from *Bacillus selenitireducens* strain MLS10. *FEMS Microbiology Letters*, 226(1), 107-112.
- Aksu. (2002). Determination of the equilibrium, kinetic and thermodynamic parameters of the batch biosorption of nickel (II) ions onto *Chlorella vulgaris*. *Process Biochemistry*, 38(1), 89-99.
- Aksu and Gonen, F. (2004). Biosorption of phenol by immobilized activated sludge in a continuous packed bed: prediction of breakthrough curves. *Process Biochemistry*, 39(5), 599-613.
- Al-Garni, S. M. (2005). Biosorption of lead by Gram-ve capsulated and noncapsulated bacteria. *Water Sa*, 31(3), 345-350.
- Al Lawati, W. M., Rizoulis, A., Eiche, E., Boothman, C., Polya, D. A., Lloyd, J. R., et al. (2012a). Characterisation of organic matter and microbial communities in contrasting arsenic-rich Holocene and arsenic-poor Pleistocene aquifers, Red River Delta, Vietnam. *Applied Geochemistry*, 27(1), 315-325.
- Al Lawati, W. M., Rizoulis, A., Eiche, E., Boothman, C., Polya, D. A., Lloyd, J. R., et al. (2012b). Characterisation of organic matter and microbial communities in contrasting arsenic-rich Holocene and arsenic-poor Pleistocene aquifers, Red River Delta, Vietnam. *Applied Geochemistry*, 27(1), 315-325.
- Alam, M., Tokunaga, S. and Maekawa, T. (2001). Extraction of arsenic in a synthetic arsenic-contaminated soil using phosphate. *Chemosphere*, 43(8), 1035-1041.
- Aleboyeh, A., Kasiri, M., Olya, M. and Aleboyeh, H. (2008). Prediction of azo dye decolorization by UV/H2O2 using artificial neural networks. *Dyes and Pigments*, 77(2), 288-294.
- Allievi, M. C., Florencia, S., Mariano, P. A., Mercedes, P. M. and Carmen, S. R. (2011). Metal biosorption by surface-layer proteins from *Bacillus* species. *Journal of Microbiology and Biotechnology*, 21(2), 147-153.
- Altowayti, W. A. H., Algaifi, H. A., Bakar, S. A. and Shahir, S. (2019). The adsorptive removal of As (III) using biomass of arsenic resistant *Bacillus thuringiensis* strain WS3: Characteristics and modelling studies. *Ecotoxicology and Environmental Safety*, 172, 176-185.
- Altowayti, W. A. H., Allozy, H. G. A., Shahir, S., Goh, P. S. and Yunus, M. A. M. (2019). A novel nanocomposite of aminated silica nanotube (MWCNT/Si/NH 2) and its potential on adsorption of nitrite. *Environmental Science and Pollution Research*, 1-12.
- Amato, K. R., Yeoman, C. J., Kent, A., Righini, N., Carbonero, F., Estrada, A., et al. (2013). Habitat degradation impacts black howler monkey (Alouatta pigra) gastrointestinal microbiomes. *The ISME Journal*, 7(7), 1344.
- Amini, M., Abbaspour, K. C., Berg, M., Winkel, L., Hug, S. J., Hoehn, E., et al. (2008). Statistical modeling of global geogenic arsenic contamination in groundwater. *Environmental Science & Technology*, 42(10), 3669-3675.
- Anah, L. and Astrini, N. (2018). Isotherm adsorption studies of Ni (II) ion removal from aqueous solutions by modified carboxymethyl cellulose hydrogel*.* Proceedings of the 2018 *IOP Conference Series: Earth and Environmental Science*, 012017.
- Anderson, C. R. and Cook, G. M. (2004). Isolation and characterization of arsenatereducing bacteria from arsenic-contaminated sites in New Zealand. *Current Microbiology*, 48(5), 341-347.
- Andres, J. and Bertin, P. N. (2016). The microbial genomics of arsenic. *FEMS Microbiology Reviews*, 40(2), 299-322.
- Annadurai, G., Ling, L. Y. and Lee, J. (2007). Biodegradation of phenol by Pseudomonas pictorum on immobilized with chitin. *African Journal of Biotechnology*, 6(3).
- Aryal, M., Ziagova, M. and Liakopoulou-Kyriakides, M. (2010). Study on arsenic biosorption using Fe (III)-treated biomass of Staphylococcus xylosus. *Chemical Engineering Journal*, 162(1), 178-185.
- Aryal, M., Ziagova, M. and Liakopoulou-Kyriakides, M. (2011). Comparison of Cr (VI) and As (V) removal in single and binary mixtures with Fe (III)-treated Staphylococcus xylosus biomass: Thermodynamic studies. *Chemical Engineering Journal*, 169(1-3), 100-106.
- Astuti, W. and Martiani, W. (2017). Competitive adsorption of Pb2+ and Zn2+ ions from aqueous solutions by modified coal fly ash*.* Proceedings of the 2017 *AIP Conference Proceedings*, 020007.

ATSDR. (2015). ATSDR's substance priority list.

- Awala, H. A. and Jamal, M. M. (2011). Equilibrium and kinetics study of adsorption of some dyes onto feldspar. *Journal of the University of Chemical Technology and Metallurgy*, 46(1), 45-52.
- Ayawei, N., Ebelegi, A. N. and Wankasi, D. (2017). Modelling and interpretation of adsorption isotherms. *Journal of Chemistry*, 2017.
- Azcue, J. M. and Nriagu, J. O. (1995). Impact of abandoned mine tailings on the arsenic concentrations in Moira Lake, Ontario. *Journal of Geochemical Exploration*, 52(1-2), 81-89.
- Bahari, Z. M., Altowayti, W. A. H., Ibrahim, Z., Jaafar, J. and Shahir, S. (2013). Biosorption of As (III) by non-living biomass of an arsenic-hypertolerant *Bacillus cereus* strain SZ2 isolated from a gold mining environment: Equilibrium and kinetic study. *Applied Biochemistry and Biotechnology*, 171(8), 2247-2261.
- Bahari, Z. M., Ibrahim, Z., Jaafar, J. and Shahir, S. (2017). Draft Genome Sequence of Arsenic-Resistant *Microbacterium* sp. Strain SZ1 Isolated from Arsenic-Bearing Gold Ores. *Genome Announcements*, 5(43), e01183-01117.
- Bai, S. and Abraham, T. E. (2001). Biosorption of Cr (VI) from aqueous solution by Rhizopus nigricans. *Bioresource Technology*, 79(1), 73-81.
- Balaria, A. and Schiewer, S. (2008). Assessment of biosorption mechanism for Pb binding by citrus pectin. *Separation and Purification Technology*, 63(3), 577- 581.
- Banerjee, S., Dubey, S., Gautam, R. K., Chattopadhyaya, M. and Sharma, Y. C. (2017). Adsorption characteristics of alumina nanoparticles for the removal of hazardous dye, Orange G from aqueous solutions. *Arabian Journal of Chemistry*.
- Bang, S., Patel, M., Lippincott, L. and Meng, X. (2005). Removal of arsenic from groundwater by granular titanium dioxide adsorbent. *Chemosphere*, 60(3), 389-397.
- Barringer, J. L. and Reilly, P. A. (2013). Arsenic in groundwater: A summary of sources and the biogeochemical and hydrogeologic factors affecting arsenic occurrence and mobility *Current perspectives in contaminant hydrology and water resources sustainability*: InTech.
- Baskan, M. B. and Pala, A. (2010). A statistical experiment design approach for arsenic removal by coagulation process using aluminum sulfate. *Desalination*, 254(1-3), 42-48.
- Battesti, A., Majdalani, N. and Gottesman, S. (2011). The RpoS-mediated general stress response in Escherichia coli. *Annual Review of Microbiology*, 65, 189- 213.
- Bazrafshan, E., Kord, M. F. and Faridi, H. (2012). ARSENIC REMOVAL FROM AQUEOUS ENVIRONMENTS BY USING MORINGA PEREGRINA SEED EXTRACT AS A NATURAL COAGULANT. *Asian Journal of Chemistry*, 25(7).
- Berg, M., Tran, H. C., Nguyen, T. C., Pham, H. V., Schertenleib, R. and Giger, W. (2001). Arsenic contamination of groundwater and drinking water in Vietnam: a human health threat. *Environmental Science & Technology*, 35(13), 2621-2626.
- Bi, L., Yang, L., Bhunia, A. K. and Yao, Y. (2016). Emulsion stabilized with phytoglycogen octenyl succinate prolongs the antimicrobial efficacy of εpoly-l-lysine against Escherichia coli O157: H7. *LWT-Food Science and Technology*, 70, 245-251.
- Boddu, V. M., Abburi, K., Talbott, J. L., Smith, E. D. and Haasch, R. (2008). Removal of arsenic (III) and arsenic (V) from aqueous medium using chitosan-coated biosorbent. *Water Research*, 42(3), 633-642.
- Brahman, K. D., Kazi, T. G., Baig, J. A., Afridi, H. I., Arain, S. S., Saraj, S., et al. (2016). Biosorptive removal of inorganic arsenic species and fluoride from aqueous medium by the stem of Tecomella undulate. *Chemosphere*, 150, 320- 328.
- Bronner, I. F., Quail, M. A., Turner, D. J. and Swerdlow, H. (2014). Improved protocols for illumina sequencing. *Current Protocols in Human Genetics*, 18.12. 11-18.12. 42.
- Bundschuh, J., Litter, M. I., Parvez, F., Román-Ross, G., Nicolli, H. B., Jean, J.-S., et al. (2012). One century of arsenic exposure in Latin America: a review of history and occurrence from 14 countries. *Science of the Total Environment*, 429, 2-35.
- Button, Moriarty, M. M., Watts, M. J., Zhang, J., Koch, I. and Reimer, K. J. (2011). Arsenic speciation in field-collected and laboratory-exposed earthworms Lumbricus terrestris. *Chemosphere*, 85(8), 1277-1283.
- Button, Robertson, B. R., Lepp, P. W. and Schmidt, T. M. (1998). A small, dilutecytoplasm, high-affinity, novel bacterium isolated by extinction culture and having kinetic constants compatible with growth at ambient concentrations of dissolved nutrients in seawater. *Applied and Environmental Microbiology*, 64(11), 4467-4476.
- Castaldi, P., Silvetti, M., Enzo, S. and Melis, P. (2010). Study of sorption processes and FT-IR analysis of arsenate sorbed onto red muds (a bauxite ore processing waste). *Journal of Hazardous Materials*, 175(1-3), 172-178.
- Chakravarty, R. and Banerjee, P. C. (2012). Mechanism of cadmium binding on the cell wall of an acidophilic bacterium. *Bioresource Technology*, 108, 176-183.
- Chang, J.-S., Law, R. and Chang, C.-C. (1997). Biosorption of lead, copper and cadmium by biomass of Pseudomonas aeruginosa PU21. *Water Research*, 31(7), 1651-1658.
- Chattopadhyay, S., Bhaumik, S., Chaudhury, A. N. and Gupta, S. D. (2002). Arsenic induced changes in growth development and apoptosis in neonatal and adult brain cells in vivo and in tissue culture. *Toxicology Letters*, 128(1-3), 73-84.
- Chiancone, E. and Ceci, P. (2010). The multifaceted capacity of Dps proteins to combat bacterial stress conditions: detoxification of iron and hydrogen peroxide and DNA binding. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1800(8), 798-805.
- Chojnacka, K., Chojnacki, A. and Gorecka, H. (2005). Biosorption of Cr3+, Cd2+ and Cu2+ ions by blue–green algae Spirulina sp.: kinetics, equilibrium and the mechanism of the process. *Chemosphere*, 59(1), 75-84.
- Choong, T. S., Chuah, T., Robiah, Y., Koay, F. G. and Azni, I. (2007). Arsenic toxicity, health hazards and removal techniques from water: an overview. *Desalination*, 217(1-3), 139-166.
- Chowdhury and Mulligan. (2011). Biosorption of arsenic from contaminated water by anaerobic biomass. *Journal of Hazardous Materials*, 190(1-3), 486-492.
- Chu, K. (2003). Prediction of two-metal biosorption equilibria using a neural network. *European Journal of Mineral Processing and Environmental Protection*, 3(1), 119-127.
- Ciopec, M., Negrea, A., Lupa, L., Davidescu, C. M. and Negrea, P. (2014). Studies regarding As (V) adsorption from underground water by Fe-XAD8-DEHPA impregnated resin. Equilibrium sorption and fixed-bed column tests. *Molecules*, 19(10), 16082-16101.
- Clausen, C. A. (2000). Isolating metal‐tolerant bacteria capable of removing copper, chromium, and arsenic from treated wood. *Waste Management and Research*, 18(3), 264-268.
- Coates, J. (2000). Interpretation of infrared spectra, a practical approach. *Encyclopedia of Analytical Chemistry*, 12, 10815-10837.
- Cuebas, M., Sannino, D. and Bini, E. (2011). Isolation and characterization of arsenic resistant *Geobacillus kaustophilus* strain from geothermal soils. *Journal of Basic Microbiology*, 51(4), 364-371.
- Cullen, W. R. and Reimer, K. J. (1989). Arsenic speciation in the environment. *Chemical Reviews*, 89(4), 713-764.
- Cummings, Caccavo, F., Fendorf, S. and Rosenzweig, R. F. (1999). Arsenic mobilization by the dissimilatory Fe (III)-reducing bacterium *Shewanella* alga BrY. *Environmental Science & Technology*, 33(5), 723-729.
- Dada, A., Olalekan, A., Olatunya, A. and Dada, O. (2012). Langmuir, Freundlich, Temkin and Dubinin–Radushkevich isotherms studies of equilibrium sorption

of Zn2+ unto phosphoric acid modified rice husk. *IOSR Journal of Applied Chemistry*, 3(1), 38-45.

- Daneshvar, N., Khataee, A. and Djafarzadeh, N. (2006). The use of artificial neural networks (ANN) for modeling of decolorization of textile dye solution containing CI Basic Yellow 28 by electrocoagulation process. *Journal of Hazardous Materials*, 137(3), 1788-1795.
- Dao, T. D., Laborie, S. and Cabassud, C. (2016). Direct As (III) removal from brackish groundwater by vacuum membrane distillation: effect of organic matter and salts on membrane fouling. *Separation and Purification Technology*, 157, 35-44.
- Das, Bora, S. S., Yadav, R. and Barooah, M. (2017). A metagenomic approach to decipher the indigenous microbial communities of arsenic contaminated groundwater of Assam. *Genomics Data*, 12, 89-96.
- Das, Mondal, N., Bhaumik, R. and Roy, P. (2014). Insight into adsorption equilibrium, kinetics and thermodynamics of lead onto alluvial soil. *International Journal of Environmental Science and Technology*, 11(4), 1101-1114.
- Dash, C. and Payyappilli, R. J. (2016). KOH string and Vancomycin susceptibility test as an alternative method to Gram staining. *Journal of International Medicine and Dentistry*, 3(2), 88-90.
- Datta, S., Mailloux, B., Jung, H.-B., Hoque, M., Stute, M., Ahmed, K., et al. (2009). Redox trapping of arsenic during groundwater discharge in sediments from the Meghna riverbank in Bangladesh. *Proceedings of the National Academy of Sciences*, pnas. 0908168106.
- DeLong, E. F., Preston, C. M., Mincer, T., Rich, V., Hallam, S. J., Frigaard, N.-U., et al. (2006). Community genomics among stratified microbial assemblages in the ocean's interior. *Science*, 311(5760), 496-503.
- Despagne, F. and Massart, D. L. (1998). Neural networks in multivariate calibration. *Analyst*, 123(11), 157R-178R.
- Dey, U., Chatterjee, S. and Mondal, N. K. (2016). Isolation and characterization of arsenic-resistant bacteria and possible application in bioremediation. *Biotechnology Reports*, 10, 1-7.
- Dhar, R. K., Zheng, Y., Saltikov, C. W., Radloff, K. A., Mailloux, B. J., Ahmed, K. M., et al. (2011). Microbes enhance mobility of arsenic in Pleistocene aquifer

sand from Bangladesh. *Environmental Science & Technology*, 45(7), 2648- 2654.

- Diamadopoulos, E., Ioannidis, S. and Sakellaropoulos, G. P. (1993). As (V) removal from aqueous solutions by fly ash. *Water Research*, 27(12), 1773-1777.
- Dickson, D., Liu, G. and Cai, Y. (2017). Adsorption kinetics and isotherms of arsenite and arsenate on hematite nanoparticles and aggregates. *Journal of Environmental Management*, 186, 261-267.
- Ding, Y., Jing, D., Gong, H., Zhou, L. and Yang, X. (2012). Biosorption of aquatic cadmium (II) by unmodified rice straw. *Bioresource Technology*, 114, 20-25.
- Dombrowski, P. M., Long, W., Farley, K. J., Mahony, J. D., Capitani, J. F. and Di Toro, D. M. (2005). Thermodynamic analysis of arsenic methylation. *Environmental Science & Technology*, 39(7), 2169-2176.
- Dong, G., Huang, Y., Yu, Q., Wang, Y., Wang, H., He, N., et al. (2014). Role of nanoparticles in controlling arsenic mobilization from sediments near a realgar tailing. *Environmental Science & Technology*, 48(13), 7469-7476.
- Drewniak, L., Stasiuk, R., Uhrynowski, W. and Sklodowska, A. (2015). *Shewanella* sp. O23S as a driving agent of a system utilizing dissimilatory arsenatereducing bacteria responsible for self-cleaning of water contaminated with arsenic. *International Journal of Molecular Sciences*, 16(7), 14409-14427.
- Escalante, G., Campos, V., Valenzuela, C., Yañez, J., Zaror, C. and Mondaca, M. (2009). Arsenic resistant bacteria isolated from arsenic contaminated river in the Atacama Desert (Chile). *Bulletin of Environmental Contamination and Toxicology*, 83(5), 657-661.
- Fakruddin, M. and Mannan, S. (2013). Methods for Analyzing Diversity of Microbial Communities in Natural Environments. *Ceylon Journal of Science (Biological Sciences)*, 42, 19-33. doi: 10.4038/cjsbs.v42i1.5896
- Fan, H., Su, C., Wang, Y., Yao, J., Zhao, K., Wang, Y., et al. (2008). Sedimentary arsenite‐oxidizing and arsenate‐reducing bacteria associated with high arsenic groundwater from Shanyin, Northwestern China. *Journal of Applied Microbiology*, 105(2), 529-539.
- Fang, J. and Deng, B. (2014). Rejection and modeling of arsenate by nanofiltration: Contributions of convection, diffusion and electromigration to arsenic transport. *Journal of Membrane Science*, 453, 42-51.
- Fernandes, F. and Lona, L. (2005). Neural network applications in polymerization processes. *Brazilian Journal of Chemical Engineering*, 22(3), 401-418.
- Ferris, M., Ruff-Roberts, A., Kopczynski, E., Bateson, M. and Ward, D. (1996). Enrichment culture and microscopy conceal diverse thermophilic Synechococcus populations in a single hot spring microbial mat habitat. *Applied and Environmental Microbiology*, 62(3), 1045-1050.
- Flanagan, S. V., Johnston, R. B. and Zheng, Y. (2012). Arsenic in tube well water in Bangladesh: health and economic impacts and implications for arsenic mitigation. *Bulletin of the World Health Organization*, 90, 839-846.
- Flemming, H.-C. and Wingender, J. (2010). The biofilm matrix. *Nature Reviews Microbiology*, 8(9), 623.
- Fontana, K. B., Chaves, E. S., Sanchez, J. D., Watanabe, E. R., Pietrobelli, J. M. and Lenzi, G. G. (2016). Textile dye removal from aqueous solutions by malt bagasse: isotherm, kinetic and thermodynamic studies. *Ecotoxicology and Environmental Safety*, 124, 329-336.
- Gault, A., Islam, F., Polya, D., Charnock, J., Boothman, C., Chatterjee, D., et al. (2005). Microcosm depth profiles of arsenic release in a shallow aquifer, West Bengal. *Mineralogical Magazine*, 69(5), 855-863.
- Ge, Y., Wang, D.-Z., Chiu, J.-F., Cristobal, S., Sheehan, D., Silvestre, F., et al. (2013). Environmental OMICS: current status and future directions. *Journal of Integrated Omics*, 3(2), 75-87.
- Giri, A., Patel, R. and Mahapatra, S. (2011). Artificial neural network (ANN) approach for modelling of arsenic (III) biosorption from aqueous solution by living cells of *Bacillus cereus* biomass. *Chemical Engineering Journal*, 178, 15-25.
- Giri, A., Patel, R., Mahapatra, S. and Mishra, P. (2012). Biosorption of arsenic (III) from aqueous solution by living cells of Bacillus cereus. *Environmental Science and Pollution Research*, 1-11.
- Giri, A., Patel, R., Mahapatra, S. and Mishra, P. (2013). Biosorption of arsenic (III) from aqueous solution by living cells of *Bacillus cereus*. *Environmental Science and Pollution Research*, 20(3), 1281-1291.
- Gnanasangeetha, D. and Sarala, T., D. (2015). Modelling of As3+ adsorption from aqueous solution using Azadirachta indica by artificial neural network. *Desalination and Water Treatment*, 56(7), 1839-1854.
- Gob, S., Oliveros, E., Bossmann, S. H., Braun, A. M., Guardani, R. and Nascimento, C. A. (1999). Modeling the kinetics of a photochemical water treatment process by means of artificial neural networks. *Chemical Engineering and Processing: Process Intensification*, 38(4-6), 373-382.
- Goswami, R., Mukherjee, S., Rana, V. S., Saha, D. R., Raman, R., Padhy, P. K., et al. (2015). Isolation and characterization of arsenic-resistant bacteria from contaminated water-bodies in West Bengal, India. *Geomicrobiology Journal*, 32(1), 17-26.
- Guo, J. and Chen, C. (2017). Removal of arsenite by a microbial bioflocculant produced from swine wastewater. *Chemosphere*, 181, 759-766.
- Gupta, Ahuja, P., Khan, S., Saxena, R. and Mohapatra, H. (2000). Microbial biosorbents: meeting challenges of heavy metal pollution in aqueous solutions. *Current Science* 78(8), 967-973.
- Gupta, Nayak, A. and Agarwal, S. (2015). Bioadsorbents for remediation of heavy metals: current status and their future prospects. *Environmental Engineering Research*, 20(1), 1-18.
- Halem, D. v., Bakker, S., Amy, G. and Van Dijk, J. (2009). Arsenic in drinking water: a worldwide water quality concern for water supply companies. *Drinking Water Engineering and Science*, 2(1), 29-34.
- Hamed, M. M., Khalafallah, M. G. and Hassanien, E. A. (2004). Prediction of wastewater treatment plant performance using artificial neural networks. *Environmental Modelling & Software*, 19(10), 919-928.
- Handley, K. M., Héry, M. and Lloyd, J. R. (2009). Redox cycling of arsenic by the hydrothermal marine bacterium Marinobacter santoriniensis. *Environmental Microbiology*, 11(6), 1601-1611.
- Hansda, A. and Kumar, V. (2015). Biosorption of copper by bacterial adsorbents: a review. *Research Journal of Environmental Toxicology*, 9(2), 45.
- Hansen, H. K., Ribeiro, A. and Mateus, E. (2006). Biosorption of arsenic (V) with Lessonia nigrescens. *Minerals Engineering*, 19(5), 486-490.
- Hao, L., Wang, N., Wang, C. and Li, G. (2018). Arsenic removal from water and river water by the combined adsorption-UF membrane process. *Chemosphere*, 202, 768-776.
- Haris, S. A., Altowayti, W. A. H., Ibrahim, Z. and Shahir, S. (2018). Arsenic biosorption using pretreated biomass of *psychrotolerant Yersinia* sp. strain

SOM-12D3 isolated from Svalbard, Arctic. *Environmental Science and Pollution Research*, 25(28), 27959-27970.

- Hassan, Z., Sultana, M., van Breukelen, B. M., Khan, S. I. and Röling, W. F. (2015). Diverse arsenic-and iron-cycling microbial communities in arseniccontaminated aquifers used for drinking water in Bangladesh. *FEMS Microbiology Ecology*, 91(4).
- Hayes, R. B. (1997). The carcinogenicity of metals in humans. *Cancer Causes & Control*, 8(3), 371-385.
- Herath, I., Vithanage, M., Bundschuh, J., Maity, J. P. and Bhattacharya, P. (2016). Natural Arsenic in Global Groundwaters: Distribution and Geochemical Triggers for Mobilization. [journal article]. *Current Pollution Reports*, 2(1), 68-89. doi: 10.1007/s40726-016-0028-2
- Héry, M., Gault, A. G., Rowland, H. A., Lear, G., Polya, D. A. and Lloyd, J. R. (2008). Molecular and cultivation-dependent analysis of metal-reducing bacteria implicated in arsenic mobilisation in south-east asian aquifers. *Applied Geochemistry*, 23(11), 3215-3223.
- Héry, M., Rizoulis, A., Sanguin, H., Cooke, D. A., Pancost, R. D., Polya, D. A., et al. (2015). Microbial ecology of arsenic‐mobilizing C ambodian sediments: lithological controls uncovered by stable‐isotope probing. *Environmental Microbiology*, 17(6), 1857-1869.
- Higgins, D. and Dworkin, J. (2012). Recent progress in *Bacillus subtilis* sporulation. *FEMS Microbiology Reviews*, 36(1), 131-148.
- Hohmann, C., Morin, G., Ona-Nguema, G., Guigner, J.-M., Brown Jr, G. E. and Kappler, A. (2011). Molecular-level modes of As binding to Fe (III)(oxyhydr) oxides precipitated by the anaerobic nitrate-reducing Fe (II)-oxidizing *Acidovorax* sp. strain BoFeN1. *Geochimica et Cosmochimica Acta*, 75(17), 4699-4712.
- Horneman, A., van Geen, A., Kent, D. V., Mathe, P., Zheng, Y., Dhar, R., et al. (2004). Decoupling of as and fe release to bangladesh groundwater under reducing conditions. Part I: Evidence from sediment profiles1. *Geochimica et Cosmochimica Acta*, 68(17), 3459-3473.
- Huang, Y. F., Ang, S. Y., Lee, K. M. and Lee, T. S. (2015). Quality of water resources in Malaysia. *Research and Practices in Water Quality*.
- Hughes, M. F. (2002). Arsenic toxicity and potential mechanisms of action. *Toxicology Letters*, 133(1), 1-16.
- Igwe, J., Nwokennaya, E. and Abia, A. (2005). The role of pH in heavy metal detoxification by biosorption from aqueous solutions containing chelating agents. *African Journal of Biotechnology*, 4(10).
- Ike, M., Miyazaki, T., Yamamoto, N., Sei, K. and Soda, S. (2008). Removal of arsenic from groundwater by arsenite-oxidizing bacteria. *Water Science and Technology*, 58(5), 1095-1100.
- Islam, F. S., Gault, A. G., Boothman, C., Polya, D. A., Charnock, J. M., Chatterjee, D., et al. (2004). Role of metal-reducing bacteria in arsenic release from Bengal delta sediments. *Nature*, 430(6995), 68.
- Jadhav, S. V., Bringas, E., Yadav, G. D., Rathod, V. K., Ortiz, I. and Marathe, K. V. (2015). Arsenic and fluoride contaminated groundwaters: a review of current technologies for contaminants removal. *Journal of Environmental Management*, 162, 306-325.
- Jain and Ali. (2000). Arsenic: occurrence, toxicity and speciation techniques. *Water Research*, 34(17), 4304-4312.
- Jain, A. and Agarwal, M. (2017). Kinetic equilibrium and thermodynamic study of arsenic removal from water using alumina supported iron nano particles. *Journal of Water Process Engineering*, 19, 51-59.
- Jebelli, M. A., Maleki, A., Amoozegar, M. A., Kalantar, E., Shahmoradi, B. and Gharibi, F. (2017). Isolation and identification of indigenous prokaryotic bacteria from arsenic-contaminated water resources and their impact on arsenic transformation. *Ecotoxicology and Environmental Safety*, 140, 170- 176.
- Jiang, J.-Q., Ashekuzzaman, S., Jiang, A., Sharifuzzaman, S. and Chowdhury, S. R. (2012). Arsenic contaminated groundwater and its treatment options in Bangladesh. *International Journal of Environmental Research and Public Health*, 10(1), 18-46.
- Joshi, Flora, S. S. and Kalia, K. (2009). Bacillus sp. strain DJ-1, potent arsenic hypertolerant bacterium isolated from the industrial effluent of India. *Journal of Hazardous Materials*, 166(2), 1500-1505.
- Kanamarlapudi, S. L. R. K., Chintalpudi, V. K. and Muddada, S. (2018). Application of Biosorption for Removal of Heavy Metals from Wastewater. *Biosorption*, 69.
- Kao, A.-C., Chu, Y.-J., Hsu, F.-L. and Liao, V. H.-C. (2013). Removal of arsenic from groundwater by using a native isolated arsenite-oxidizing bacterium. *Journal of Contaminant Hydrology*, 155, 1-8.
- Kapoor, A., Viraraghavan, T. and Cullimore, D. R. (1999). Removal of heavy metals using the fungus Aspergillus niger. *Bioresource Technology*, 70(1), 95-104.
- Karas, V. O., Westerlaken, I. and Meyer, A. S. (2015). The DNA-binding protein from starved cells (Dps) utilizes dual functions to defend cells against multiple stresses. *Journal of Bacteriology*, JB. 00475-00415.
- Kartinen, E. O. and Martin, C. J. (1995). An overview of arsenic removal processes. *Desalination*, 103(1-2), 79-88.
- Khaliq, W. and Ehsan, M. B. (2016). Crack healing in concrete using various bio influenced self-healing techniques. *Construction and Building Materials*, 102, 349-357.
- Kim, Kim, K. W. and Cho, J. (2006). Removal and transport mechanisms of arsenics in UF and NF membrane processes. *Journal of Water and Health*, 4(2), 215- 223.
- Kim, Lee, C., Lee, S. M. and Jung, J. (2018). Chemical and toxicological assessment of arsenic sorption onto Fe-sericite composite powder and beads. *Ecotoxicology and Environmental Safety*, 147, 80-85.
- Kinniburgh, D. and Smedley, P. (2001). Arsenic contamination of groundwater in Bangladesh. *Risk Management and Healthcare Policy* 251–261.
- Kiran, M., Shrikant, A., Parag, S., Lele, S. and Rekha, S. (2008). Comparison of artificial neural network (ANN) and response surface methodology (RSM) in fermentation media optimization: Case study of fermentative production of scleroglucan. *Biochemical Engineering Journal*, 41(3), 266-273.
- Kitchin, K. T. (2001). Recent advances in arsenic carcinogenesis: modes of action, animal model systems, and methylated arsenic metabolites. *Toxicology and Applied Pharmacology*, 172(3), 249-261.
- Kord, M. F., Bazrafshan, E., Farzadkia, M. and Amini, S. (2012). Arsenic removal from aqueous solutions by Salvadora persica stem ash. *Journal of Chemistry*, 2013.
- Kosutic, K., Furac, L., Sipos, L. and Kunst, B. (2005). Removal of arsenic and pesticides from drinking water by nanofiltration membranes. *Separation and Purification Technology*, 42(2), 137-144.
- Kotmakci, M., Çetintaş, V. B. and Kantarcı, A. G. (2017). Preparation and characterization of lipid nanoparticle/pDNA complexes for STAT3 downregulation and overcoming chemotherapy resistance in lung cancer cells. *International Journal of Pharmaceutics*, 525(1), 101-111.
- Krafft, T. and Macy, J. M. (1998). Purification and characterization of the respiratory arsenate reductase of Chrysiogenes arsenatis. *European Journal of Biochemistry*, 255(3), 647-653.
- Krautler, B. (1990). Chemistry of methylcorrinoids related to their roles in bacterial C1 metabolism. *FEMS Microbiology Letters*, 87(3‐4), 349-354.
- Krika, F., Azzouz, N. and Ncibi, M. C. (2016). Adsorptive removal of cadmium from aqueous solution by cork biomass: Equilibrium, dynamic and thermodynamic studies. *Arabian Journal of Chemistry*, 9, S1077-S1083.
- Kulp, T., Hoeft, S., Miller, L., Saltikov, C., Murphy, J., Han, S., et al. (2006). Dissimilatory arsenate and sulfate reduction in sediments of two hypersaline, arsenic-rich soda lakes: Mono and Searles Lakes, California. *Applied and Environmental Microbiology*, 72(10), 6514-6526.
- Kumar, S., Nair, R. R., Pillai, P. B., Gupta, S. N., Iyengar, M. and Sood, A. (2014). Graphene oxide–MnFe2O4 magnetic nanohybrids for efficient removal of lead and arsenic from water. *ACS Applied Materials & Interfaces*, 6(20), 17426-17436.
- Kumari, P., Sharma, P., Srivastava, S. and Srivastava, M. (2006). Biosorption studies on shelled Moringa oleifera Lamarck seed powder: removal and recovery of arsenic from aqueous system. *International Journal of Mineral Processing*, 78(3), 131-139.
- Kwok, K. C., Koong, L. F., Al Ansari, T. and McKay, G. (2018). Adsorption/desorption of arsenite and arsenate on chitosan and nanochitosan. *Environmental Science and Pollution Research*, 25(15), 14734-14742.
- Kyle, J., Breuer, P., Bunney, K. and Pleysier, R. (2012). Review of trace toxic elements (Pb, Cd, Hg, As, Sb, Bi, Se, Te) and their deportment in gold processing: Part II: Deportment in gold ore processing by cyanidation. *Hydrometallurgy*, 111, 10-21.
- Layton, A. C., Chauhan, A., Williams, D. E., Mailloux, B., Knappett, P. S., Ferguson, A. S., et al. (2014). Metagenomes of microbial communities in arsenic-and pathogen-contaminated well and surface water from Bangladesh. *Genome Announc.*, 2(6), e01170-01114.
- Lek, S. and Guégan, J.-F. (1999). Artificial neural networks as a tool in ecological modelling, an introduction. *Ecological Modelling*, 120(2-3), 65-73.
- Li and Logan, B. E. (2004). Bacterial adhesion to glass and metal-oxide surfaces. *Colloids and Surfaces B: Biointerfaces*, 36(2), 81-90.
- Li, Tian, H., Wang, L. and Duan, J. (2016). Bacterial Diversity in Linglong Gold Mine, China. *Geomicrobiology Journal*, 1-7.
- Lievremont, D., N'negue, M.-A., Behra, P. and Lett, M.-C. (2003). Biological oxidation of arsenite: batch reactor experiments in presence of kutnahorite and chabazite. *Chemosphere*, 51(5), 419-428.
- Lin, L., Qiu, W., Wang, D., Huang, Q., Song, Z. and Chau, H. W. (2017). Arsenic removal in aqueous solution by a novel Fe-Mn modified biochar composite: Characterization and mechanism. *Ecotoxicology and Environmental Safety*, 144, 514-521.
- Liu, P. Y., Liu, H., Li, Y. J. and Dong, C. X. (2014). Remediation of arsenic contaminated soils and treatment of washing effluent using calcined Mn-Fe Layered double hydroxide*.* Proceedings of the 2014 *Advanced Materials Research*,
- Loukidou, M. X., Matis, K. A., Zouboulis, A. I. and Liakopoulou-Kyriakidou, M. (2003). Removal of As (V) from wastewaters by chemically modified fungal biomass. *Water Research*, 37(18), 4544-4552.
- Lu, X., Zhang, Y., Liu, C., Wu, M. and Wang, H. (2018). Characterization of the antimonite-and arsenite-oxidizing bacterium Bosea sp. AS-1 and its potential application in arsenic removal. *Journal of Hazardous Materials*, 359, 527- 534.
- Lugli, G. A., Milani, C., Duranti, S., Alessandri, G., Turroni, F., Mancabelli, L., et al. (2019). Isolation of novel gut bifidobacteria using a combination of metagenomic and cultivation approaches. *Genome Biology*, 20(1), 96. doi: 10.1186/s13059-019-1711-6
- Luo, J., Bai, Y., Liang, J. and Qu, J. (2014). Metagenomic approach reveals variation of microbes with arsenic and antimony metabolism genes from highly contaminated soil. *PLoS One*, 9(10), e108185.
- Maeda, S., Ohki, A., Miyahara, K., Takeshita, T. and Higashi, S. (1990). Growth characteristics and arsenic metabolism of two species of arsenic‐tolerant bacteria. *Applied Organometallic Chemistry*, 4(3), 245-250.
- Mahajan, R., Attri, S., Sharma, K., Singh, N., Sharma, D. and Goel, G. (2018). Statistical assessment of DNA extraction methodology for cultureindependent analysis of microbial community associated with diverse environmental samples. [journal article]. *Molecular Biology Reports*, 45(3), 297-308. doi: 10.1007/s11033-018-4162-3
- Maji, S. K., Pal, A. and Pal, T. (2008). Arsenic removal from real-life groundwater by adsorption on laterite soil. *Journal of Hazardous Materials*, 151(2-3), 811- 820.
- Malasarn, D., Saltikov, C., Campbell, K., Santini, J., Hering, J. and Newman, D. (2004). arrA is a reliable marker for As (V) respiration. *Science*, 306(5695), 455-455.
- Mandal, Sahu, M. K. and Patel, R. K. (2013). Adsorption studies of arsenic (III) removal from water by zirconium polyacrylamide hybrid material (ZrPACM-43). *Water Resources and Industry*, 4, 51-67.
- Mandal and Suzuki, K. T. (2002). Arsenic round the world: a review. *Talanta*, 58(1), 201-235.
- Mangaiyarkarasi, M. M., Vincent, S., Janarthanan, S., Rao, T. S. and Tata, B. (2011). Bioreduction of Cr (VI) by alkaliphilic *Bacillus subtilis* and interaction of the membrane groups. *Saudi Journal of Biological Sciences*, 18(2), 157-167.
- Marsden, J. and House, I. (2006). *The chemistry of gold extraction*: SME.
- Martinez, A. and Kolter, R. (1997). Protection of DNA during oxidative stress by the nonspecific DNA-binding protein Dps. *Journal of Bacteriology*, 179(16), 5188-5194.
- McCreadie, H., Jambor, J., Blowes, D. W., Ptacek, C. and Hiller, D. (1998). *Geochemical behavior of autoclave-produced ferric arsenates and jarosite in a gold-mine tailings impoundment*: National Water Research Institute.
- McHardy, A. C. and Rigoutsos, I. (2007). What's in the mix: phylogenetic classification of metagenome sequence samples. *Current Opinion in Microbiology*, 10(5), 499-503.
- Meena, A. K., Mishra, G., Rai, P., Rajagopal, C. and Nagar, P. (2005). Removal of heavy metal ions from aqueous solutions using carbon aerogel as an adsorbent. *Journal of Hazardous Materials*, 122(1-2), 161-170.
- Meroufel, B., Benali, O., Benyahia, M., Benmoussa, Y. and Zenasni, M. (2013). Adsorptive removal of anionic dye from aqueous solutions by Algerian kaolin: Characteristics, isotherm, kinetic and thermodynamic studies. *Journal of Materials and Environmental Science*, 4(3), 482-491.
- Meyer and Baker, T. A. (2011). Proteolysis in the Escherichia coli heat shock response: a player at many levels. *Current Opinion in Microbiology*, 14(2), 194-199.
- Meyer and Grainger, D. C. (2013). The Escherichia coli nucleoid in stationary phase *Advances in applied microbiology* (Vol. 83, pp. 69-86): Elsevier.
- Meyer, Michalke, K., Kouril, T. and Hensel, R. (2008). Volatilisation of metals and metalloids: an inherent feature of methanoarchaea? *Systematic and Applied Microbiology*, 31(2), 81-87.
- Michalke, K., Wickenheiser, E., Mehring, M., Hirner, A. and Hensel, R. (2000). Production of volatile derivatives of metal (loid) s by microflora involved in anaerobic digestion of sewage sludge. *Applied and Environmental Microbiology*, 66(7), 2791-2796.
- Miller, W. H., Schipper, H. M., Lee, J. S., Singer, J. and Waxman, S. (2002). Mechanisms of Action of Arsenic Trioxide. *Cancer research*, 62(14), 3893- 3903.
- Miyatake and Hayashi. (2009). Characteristics of arsenic removal from aqueous solution by *Bacillus megaterium* strain UM-123. *Journal of Environmental Biotechnology*, 9(2), 123-129.
- Miyatake and Hayashi. (2011). Characteristics of arsenic removal by *Bacillus cereus* strain W2. *Resources Processing*, 58(3), 101-107.
- Mohamad, O. A., Hao, X., Xie, P., Hatab, S., Lin, Y. and Wei, G. (2009). Biosorption of copper (II) from aqueous solution using non-living *Mesorhizobium amorphae* strain CCNWGS0123. *Microbes and Environments*, 1202170359-1202170359.
- Mohamed, E. A. and Farag, A. G. (2015). Arsenic Removal from Aqueous Solutions by Different *Bacillus* and *Lysinibacillus* Species. *Bioremediation Journal*, 19(4), 269-276.
- Mohan, Dinesh, Pittman Jr and Charles U. (2007). Arsenic removal from water/wastewater using adsorbents—a critical review. *Journal of Hazardous Materials*, 142(1-2), 1-53.
- Mohan and Pittman. (2007). Arsenic removal from water/wastewater using adsorbents—a critical review. *Journal of Hazardous Materials*, 142(1), 1-53.
- Mondal, P., Bhowmick, S., Chatterjee, D., Figoli, A. and Van der Bruggen, B. (2013). Remediation of inorganic arsenic in groundwater for safe water supply: a critical assessment of technological solutions. *Chemosphere*, 92(2), 157-170.
- Mondal, P., Hermans, N., Tran, A. T. K., Zhang, Y., Fang, Y., Wang, X., et al. (2014). Effect of physico-chemical parameters on inorganic arsenic removal from aqueous solution using a forward osmosis membrane. *Journal of Environmental Chemical Engineering*, 2(3), 1309-1316.
- Mukherjee, A., Sengupta, M. K., Hossain, M. A., Ahamed, S., Das, B., Nayak, B., et al. (2006). Arsenic contamination in groundwater: a global perspective with emphasis on the Asian scenario. *Journal of Health, Population and Nutrition*, 142-163.
- Muñiz, G., Fierro, V., Celzard, A., Furdin, G., Gonzalez-Sánchez, G. and Ballinas, M. (2009). Synthesis, characterization and performance in arsenic removal of iron-doped activated carbons prepared by impregnation with Fe (III) and Fe (II). *Journal of Hazardous Materials*, 165(1-3), 893-902.
- Murugesan, G., Sathishkumar, M. and Swaminathan, K. (2006). Arsenic removal from groundwater by pretreated waste tea fungal biomass. *Bioresource Technology*, 97(3), 483-487.
- Nair, S. and Finkel, S. E. (2004). Dps protects cells against multiple stresses during stationary phase. *Journal of Bacteriology*, 186(13), 4192-4198.
- Nanthakumar, K., Karthikeyan, K. and Lakshmanaperumalsamy, P. (2009). Investigation on Biosorption of Reactive Blue 140 by Dead Biomass of Aspergillus niger HM11: Kinetics and Isotherm Studies. *Global Journal of Biotechnology & Biochemistry*, 4(2), 169-178.
- Naujokas, M. F., Anderson, B., Ahsan, H., Aposhian, H. V., Graziano, J. H., Thompson, C., et al. (2013). The broad scope of health effects from chronic arsenic exposure: update on a worldwide public health problem. *Environmental Health Perspectives*, 121(3), 295-302.
- Nazari, A. M., Radzinski, R. and Ghahreman, A. (2017). Review of arsenic metallurgy: Treatment of arsenical minerals and the immobilization of arsenic. *Hydrometallurgy*, 174, 258-281.
- Neelakanta, G. and Sultana, H. (2013). The use of metagenomic approaches to analyze changes in microbial communities. *Microbiology Insights*, 6, MBI. S10819.
- Nickson, R., McArthur, J., Ravenscroft, P., Burgess, W. and Ahmed, K. (2000). Mechanism of arsenic release to groundwater, Bangladesh and West Bengal. *Applied Geochemistry*, 15(4), 403-413.
- Nicomel, N., Leus, K., Folens, K., Van Der Voort, P. and Du Laing, G. (2016). Technologies for arsenic removal from water: current status and future perspectives. *International Journal of Environmental Research and Public Health*, 13(1), 62.
- Nidheesh, P. and Singh, T. A. (2017). Arsenic removal by electrocoagulation process: Recent trends and removal mechanism. *Chemosphere*, 181, 418-432.
- Nold, S. C., Kopczynski, E. D. and Ward, D. M. (1996). Cultivation of aerobic chemoorganotrophic proteobacteria and gram-positive bacteria from a hot spring microbial mat. *Applied and Environmental Microbiology*, 62(11), 3917-3921.
- Ogundipe, K. D. and Babarinde, A. (2017). Comparative study on batch equilibrium biosorption of Cd (II), Pb (II) and Zn (II) using plantain (Musa paradisiaca) flower: kinetics, isotherm, and thermodynamics. *Chemistry International*, 3(2), 135-149.
- Omoregie, E. O., Couture, R.-M., Van Cappellen, P., Corkhill, C. L., Charnock, J. M., Polya, D. A., et al. (2013). Arsenic bioremediation by biogenic iron oxides and sulfides. *Applied and Environmental Microbiology*, AEM. 00683- 00613.
- Onnby, L., Kumar, P. S., Sigfridsson, K. G., Wendt, O. F., Carlson, S. and Kirsebom, H. (2014). Improved arsenic (III) adsorption by Al2O3 nanoparticles and

H2O2: Evidence of oxidation to arsenic (V) from X-ray absorption spectroscopy. *Chemosphere*, 113, 151-157.

- Opio, F. (2013). *Investigation of Fe (III)-As (III) bearing phases and their potential for arsenic disposal.*
- Oremland, R. S., Hoeft, S. E., Santini, J. M., Bano, N., Hollibaugh, R. A. and Hollibaugh, J. T. (2002). Anaerobic oxidation of arsenite in Mono Lake water and by a facultative, arsenite-oxidizing chemoautotroph, strain MLHE-1. *Applied and Environmental Microbiology*, 68(10), 4795-4802.
- Oremland, R. S., Kulp, T. R., Blum, J. S., Hoeft, S. E., Baesman, S., Miller, L. G., et al. (2005). A microbial arsenic cycle in a salt-saturated, extreme environment. *Science*, 308(5726), 1305-1308.
- Padmavathy, V. (2008). Biosorption of nickel (II) ions by baker's yeast: Kinetic, thermodynamic and desorption studies. *Bioresource Technology*, 99(8), 3100- 3109.
- Pal, A., Dutta, S., Mukherjee, P. and Paul, A. (2005). Occurrence of heavy metalresistance in microflora from serpentine soil of Andaman. *Journal of Basic Microbiology*, 45(3), 207-218.
- Pandey and Bhatt, R. (2015). Arsenic resistance and accumulation by two bacteria isolated from a natural arsenic contaminated site. *Journal of Basic Microbiology*, 55(11), 1275-1286.
- Pandey, Choubey, S., Verma, Y., Pandey, M. and Chandrashekhar, K. (2009). Biosorptive removal of arsenic from drinking water. *Bioresource Technology*, 100(2), 634-637.
- Park, Y.-S., Chon, T.-S., Kwak, I.-S. and Lek, S. (2004). Hierarchical community classification and assessment of aquatic ecosystems using artificial neural networks. *Science of the Total Environment*, 327(1-3), 105-122.
- Peng, T., Hubele, N. and Karady, G. (1992). Advancement in the application of neural networks for short-term load forecasting. *IEEE Transactions on Power Systems*, 7(1), 250-257.
- Pepi, M., Volterrani, M., Renzi, M., Marvasi, M., Gasperini, S., Franchi, E., et al. (2007). Arsenic‐resistant bacteria isolated from contaminated sediments of the Orbetello Lagoon, Italy, and their characterization. *Journal of Applied Microbiology*, 103(6), 2299-2308.
- Perez-Jimenez, J. R., DeFraia, C. and Young, L. (2005). Arsenate respiratory reductase gene (arrA) for *Desulfosporosinus* sp. strain Y5. *Biochemical and Biophysical Research Communications*, 338(2), 825-829.
- Podder, M. and Majumder, C. (2015). Phycoremediation of arsenic from wastewaters by *Chlorella pyrenoidosa*. *Groundwater for Sustainable Development*, 1(1-2), 78-91.
- Pokhrel, D. and Viraraghavan, T. (2008). Arsenic removal from an aqueous solution by modified A. niger biomass: batch kinetic and isotherm studies. *Journal of Hazardous Materials*, 150(3), 818-825.
- Polya, D. A. and Middleton, D. R. (2017). Arsenic in drinking water: sources & human exposure. *IWA Publishing*, 1.
- Prakash, N., Manikandan, S., Govindarajan, L. and Vijayagopal, V. (2008). Prediction of biosorption efficiency for the removal of copper (II) using artificial neural networks. *Journal of Hazardous Materials*, 152(3), 1268- 1275.
- Prasad, Ramanathan, A., Paul, J., Subramanian, V. and Prasad, R. (2013). Biosorption of arsenite $(As+3)$ and arsenate $(As+5)$ from aqueous solution by Arthrobacter sp. biomass. *Environmental Technology*, 34(19), 2701-2708.
- Prasad, Srivastava, P., Subramanian, V. and Paul, J. (2011). Biosorption of As (III) ion on Rhodococcus sp. WB-12: biomass characterization and kinetic studies. *Separation Science and Technology*, 46(16), 2517-2525.
- Prasenjit, Mohanty, B., Balomajumder, C. and Saraswati, S. (2012). Modeling of the removal of arsenic species from simulated groundwater containing As, Fe, and Mn: a neural network based approach. *CLEAN - Soil Air Water*, 40(3), 285-289.
- Prieto, D., Devesa-Rey, R., Rubinos, D., Díaz-Fierros, F. and Barral, M. (2013). Arsenate retention by epipsammic biofilms developed on streambed sediments: influence of phosphate. *BioMed Research International*, 2013.
- Puranik, P. and Paknikar, K. (1997). Biosorption of lead and zinc from solutions using Streptoverticillium cinnamoneum waste biomass. *Journal of Biotechnology*, 55(2), 113-124.
- Qin, Rosen, B. P., Zhang, Y., Wang, G., Franke, S. and Rensing, C. (2006). Arsenic detoxification and evolution of trimethylarsine gas by a microbial arsenite S-

adenosylmethionine methyltransferase. *Proceedings of the National Academy of Sciences*, 103(7), 2075-2080.

- Qin, Shi, B. and Liu, J. (2006). Application of chitosan and alginate in treating waste water containing heavy metal ions. *Indian Journal of Chemical Technology* 13(5).
- Rahman, M., Tondel, M., Ahmad, S. A., Chowdhury, I. A., Faruquee, M. H. and Axelson, O. (1999). Hypertension and arsenic exposure in Bangladesh. *Hypertension*, 33(1), 74-78.
- Ramachandran, S. K., Ramakrishnan, V. and Bang, S. S. (2001). Remediation of concrete using micro-organisms. *ACI Materials Journal-American Concrete Institute*, 98(1), 3-9.
- Ramrakhiani, L., Majumder, R. and Khowala, S. (2011). Removal of hexavalent chromium by heat inactivated fungal biomass of Termitomyces clypeatus: Surface characterization and mechanism of biosorption. *Chemical Engineering Journal*, 171(3), 1060-1068.
- Ranjan, Mishra, D. and Hasan, S. (2011). Bioadsorption of arsenic: an artificial neural networks and response surface methodological approach. *Industrial & Engineering Chemistry Research*, 50(17), 9852-9863.
- Ranjan, Talat, M. and Hasan, S. (2009). Biosorption of arsenic from aqueous solution using agricultural residue 'rice polish'. *Journal of Hazardous Materials*, 166(2-3), 1050-1059.
- Rastogi, G. and Sani, R. K. (2011). Molecular techniques to assess microbial community structure, function, and dynamics in the environment *Microbes and Microbial Technology* (pp. 29-57): Springer.
- Ravenscroft, P., Brammer, H. and Richards, K. (2009). *Arsenic pollution: a global synthesis* (Vol. 28): John Wiley & Sons.
- Riesenfeld, C. S., Schloss, P. D. and Handelsman, J. (2004). Metagenomics: genomic analysis of microbial communities. *Annual Review of Genetics*, 38, 525-552.
- Rizoulis, A., Al Lawati, W. M., Pancost, R. D., Polya, D. A., van Dongen, B. E. and Lloyd, J. R. (2014). Microbially mediated reduction of FeIII and AsV in Cambodian sediments amended with 13C-labelled hexadecane and kerogen. *Environmental Chemistry*, 11(5), 538-546.
- Robey, N. M., Solo-Gabriele, H. M., Jones, A. S., Marini, J. and Townsend, T. G. (2018). Metals content of recycled construction and demolition wood before

and after implementation of best management practices. *Environmental Pollution*, 242, 1198-1205.

- Robins, R. and Jayaweera, L. D. (1992). Arsenic in gold processing. *Mineral Procesing and Extractive Metallurgy Review*, 9(1-4), 255-271.
- Roddick-Lanzilotta, A. J., McQuillan, A. J. and Craw, D. (2002). Infrared spectroscopic characterisation of arsenate (V) ion adsorption from mine waters, Macraes Mine, New Zealand. *Applied Geochemistry*, 17(4), 445-454.
- Rosewarne, C. P., Pettigrove, V., Stokes, H. W. and Parsons, Y. M. (2010). Class 1 integrons in benthic bacterial communities: abundance, association with Tn 402-like transposition modules and evidence for coselection with heavy-metal resistance. *FEMS Microbiology Ecology*, 72(1), 35-46.
- Rowland, H., Boothman, C., Pancost, R., Gault, A., Polya, D. and Lloyd, J. (2009). The role of indigenous microorganisms in the biodegradation of naturally occurring petroleum, the reduction of iron, and the mobilization of arsenite from West Bengal aquifer sediments. *Journal of Environmental Quality*, 38(4), 1598-1607.
- Rowland, H., Pederick, R., Polya, D., Pancost, R., Van Dongen, B., Gault, A., et al. (2007). The control of organic matter on microbially mediated iron reduction and arsenic release in shallow alluvial aquifers, Cambodia. *Geobiology*, 5(3), 281-292.
- Roy, Mondal, N. and Das, K. (2014). Modeling of the adsorptive removal of arsenic: a statistical approach. *Journal of Environmental Chemical Engineering*, 2(1), 585-597.
- Roy, Mondal, N. K., Bhattacharya, S., Das, B. and Das, K. (2013). Removal of arsenic (III) and arsenic (V) on chemically modified low-cost adsorbent: batch and column operations. *Applied Water Science*, 3(1), 293-309.
- Saha, D., Bhowal, A. and Datta, S. (2010). Artificial neural network modeling of fixed bed biosorption using radial basis approach. *Heat and Mass Transfer*, 46(4), 431-436.
- Salari, D., Daneshvar, N., Aghazadeh, F. and Khataee, A. (2005). Application of artificial neural networks for modeling of the treatment of wastewater contaminated with methyl tert-butyl ether (MTBE) by UV/H2O2 process. *Journal of Hazardous Materials*, 125(1-3), 205-210.
- Salmassi, T. M., Venkateswaren, K., Satomi, M., Newman, D. K. and Hering, J. G. (2002). Oxidation of arsenite by Agrobacterium albertimagni, AOL15, sp. nov., isolated from Hot Creek, California. *Geomicrobiology Journal*, 19(1), 53-66.
- Samal, A. C., Kar, S., Maity, J. P. and Santra, S. C. (2013). Arsenicosis and its relationship with nutritional status in two arsenic affected areas of West Bengal, India. *Journal of Asian Earth Sciences*, 77, 303-310.
- San, N. O. and Dönmez, G. (2012). Biosorption of chromium (VI), nickel (II) and Remazol Blue by Rhodotorula muciloginosa biomass. *Water Science and Technology*, 65(3), 471-477.
- Santoyo, G., Pacheco, C. H., Salmerón, J. H. and León, R. H. (2017). The role of abiotic factors modulating the plant-microbe-soil interactions: toward sustainable agriculture. A review. *Spanish Journal of Agricultural Research*, 15(1), 13.
- Saqib, A. N. S., Waseem, A., Khan, A. F., Mahmood, Q., Khan, A., Habib, A., et al. (2013). Arsenic bioremediation by low cost materials derived from Blue Pine (Pinus wallichiana) and Walnut (Juglans regia). *Ecological Engineering*, 51, 88-94.
- Sarı, A., Uluozlü, Ö. D. and Tüzen, M. (2011). Equilibrium, thermodynamic and kinetic investigations on biosorption of arsenic from aqueous solution by algae (Maugeotia genuflexa) biomass. *Chemical Engineering Journal*, 167(1), 155-161.
- Sarkar, D., Makris, K. C., Vandanapu, V. and Datta, R. (2007). Arsenic immobilization in soils amended with drinking-water treatment residuals. *Environmental Pollution*, 146(2), 414-419.
- Satyapal, G. K., Mishra, S. K., Srivastava, A., Ranjan, R. K., Prakash, K., Haque, R., et al. (2018). Possible bioremediation of arsenic toxicity by isolating indigenous bacteria from the middle Gangetic plain of Bihar, India. *Biotechnology Reports*, 17, 117-125.
- Schloss, P. D. and Handelsman, J. (2005). Metagenomics for studying unculturable microorganisms: cutting the Gordian knot. *Genome Biology*, 6(8), 229.
- Seki, H., Suzuki, A. and Maruyama, H. (2005). Biosorption of chromium (VI) and arsenic (V) onto methylated yeast biomass. *Journal of Colloid and Interface Science*, 281(2), 261-266.
- Sengupta, D., Mitra, A. K., Choudhury, S. S. and Chandra, A. (2014). Isotherm Study in Arsenic Tolerant Bacteria Isolated from Arsenic Affected Area in West-Bengal, India. *IOSR Journal Of Environmental Science*, 8(1), 08-19.
- Sezonov, G., Joseleau-Petit, D. and d'Ari, R. (2007). Escherichia coli physiology in Luria-Bertani broth. *Journal of Bacteriology*, 189(23), 8746-8749.
- Shahid, M., Niazi, N. K., Dumat, C., Naidu, R., Khalid, S., Rahman, M. M., et al. (2018). A meta-analysis of the distribution, sources and health risks of arsenic-contaminated groundwater in Pakistan. *Environmental Pollution*, 242, 307-319.
- Shakya, S., Pradhan, B., Smith, L., Shrestha, J. and Tuladhar, S. (2012). Isolation and characterization of aerobic culturable arsenic-resistant bacteria from surfacewater and groundwater of Rautahat District, Nepal. *Journal of Environmental Management*, 95, S250-S255.
- Shariatpahani, M., Anderson, A., Abdelghani, A. and Englande, A. (1983). Microbial metabolism of an organic arsenical herbicide*.* Proceedings of the 1983 *Biodeterioration 5: papers presented at the 5th International Biodeterioration Symposium, Aberdeen, September, 1981/edited by TA Oxley and S. Barry*,
- Shetty, G. R. and Chellam, S. (2003). Predicting membrane fouling during municipal drinking water nanofiltration using artificial neural networks. *Journal of Membrane Science*, 217(1-2), 69-86.
- Shibata, J. and Murayama, N. (2006). Engineering aspect on the removal of As (V), As (III), Cr (VI), B (III) and Se (IV) with functional inorganic ion exchanger*.* Proceedings of the 2006 *Proceedings of the 2006 TMS Fall Extraction and Processing Division, Sohn International Symposium, San Diego, CA, USA*, 339-346.
- Shojaeimehr, T., Rahimpour, F., Khadivi, M. A. and Sadeghi, M. (2014). A modeling study by response surface methodology (RSM) and artificial neural network (ANN) on Cu2+ adsorption optimization using light expended clay aggregate (LECA). *Journal of Industrial and Engineering Chemistry*, 20(3), 870-880.
- Sibi, G. (2014). Biosorption of arsenic by living and dried biomass of fresh water microalgae-potentials and equilibrium studies. *Journal of Bioremediation & Biodegredation*, 5(6), 1.
- Silver, S. and Phung, L. T. (2005). Genes and enzymes involved in bacterial oxidation and reduction of inorganic arsenic. *Applied and Environmental Microbiology*, 71(2), 599-608.
- Simeonova, D. D., Lievremont, D., Lagarde, F., Muller, D. A., Groudeva, V. I. and Lett, M.-C. (2004). Microplate screening assay for the detection of arseniteoxidizing and arsenate-reducing bacteria. *FEMS Microbiology Letters*, 237(2), 249-253.
- Singh, Bajpai, J., Bajpai, A. and Shrivastava, R. (2011). Removal of arsenic ions and bacteriological contamination from aqueous solutions using chitosan nanospheres. *Indian Journal of Chemical Technology* 18(5).
- Singh, Kiran, Sinha, TJM, Srivastava and Shalini. (2015). Functionalized nanocrystalline cellulose: smart biosorbent for decontamination of arsenic. *International Journal of Mineral Processing*, 139, 51-63.
- Singh, Kumar, D. and Sahu, A. P. (2007). Arsenic in the environment: effects on human health and possible prevention. *Journal of Environmental Biology*, 28(2), 359.
- Singh, Kumar, S., Bansal, A., Jha, M. and Dey, A. (2012). An integrated approach to remove Cr (VI) using immobilized *Chlorella minutissima* grown in nutrient rich sewage wastewater. *Bioresource Technology*, 104, 257-265.
- Singh, Singh, S., Parihar, P., Singh, V. P. and Prasad, S. M. (2015). Arsenic contamination, consequences and remediation techniques: a review. *Ecotoxicology and Environmental Safety*, 112, 247-270.
- Smedley and Kinniburgh, D. (2002). A review of the source, behaviour and distribution of arsenic in natural waters. *Applied Geochemistry*, 17(5), 517- 568.
- Smedley and Kinniburgh, D. G. (2001). Source and behavior of arsenic in natural waters. *United Nations synthesis report on arsenic in drinking water. World Health Organization, Geneva, Switzerland.* , 1-61.
- Smith, Cotrufo, M., Rumpel, C., Paustian, K., Kuikman, P., Elliott, J., et al. (2015). Biogeochemical cycles and biodiversity as key drivers of ecosystem services provided by soils. *Soil Discussions*, 2(1), 537-586.
- Smith, Koch, I. and Reimer, K. J. (2007). Arsenic Speciation Analysis of Cultivated White Button Mushrooms (Agaricus bisporus) Using High-Performance Liquid Chromatography− Inductively Coupled Plasma Mass Spectrometry,

and X-ray Absorption Spectroscopy. *Environmental Science & Technology*, 41(20), 6947-6954.

- Sokker, H., El-Sawy, N. M., Hassan, M. and El-Anadouli, B. E. (2011). Adsorption of crude oil from aqueous solution by hydrogel of chitosan based polyacrylamide prepared by radiation induced graft polymerization. *Journal of Hazardous Materials*, 190(1-3), 359-365.
- Song, B., Chyun, E., Jaffé, P. R. and Ward, B. B. (2009). Molecular methods to detect and monitor dissimilatory arsenate-respiring bacteria (DARB) in sediments. *FEMS Microbiology Ecology*, 68(1), 108-117.
- Srinath, T., Verma, T., Ramteke, P. and Garg, S. (2002). Chromium (VI) biosorption and bioaccumulation by chromate resistant bacteria. *Chemosphere*, 48(4), 427-435.
- Staley, J. T. and Konopka, A. (1985). Measurement of in situ activities of nonphotosynthetic microorganisms in aquatic and terrestrial habitats. *Annual Reviews in Microbiology*, 39(1), 321-346.
- Staunton, W. P. (2016). Carbon-in-pulp *Gold Ore Processing (Second Edition)* (pp. 535-552): Elsevier.
- Stefani, F. O., Bell, T. H., Marchand, C., Ivan, E., El Yassimi, A., St-Arnaud, M., et al. (2015). Culture-dependent and-independent methods capture different microbial community fractions in hydrocarbon-contaminated soils. *PLoS One*, 10(6), e0128272.
- Strik, D. P., Domnanovich, A. M., Zani, L., Braun, R. and Holubar, P. (2005). Prediction of trace compounds in biogas from anaerobic digestion using the MATLAB Neural Network Toolbox. *Environmental Modelling & Software*, 20(6), 803-810.
- Stute, M., Zheng, Y., Schlosser, P., Horneman, A., Dhar, R., Datta, S., et al. (2007). Hydrological control of As concentrations in Bangladesh groundwater. *Water Resources Research*, 43(9).
- Sun. (2004). Arsenic contamination and arsenicosis in China. *Toxicology and Applied Pharmacology*, 198(3), 268-271.
- Sun, Bostick, Mailloux, B. J., Ross, J. M. and Chillrud, S. N. (2016). Effect of oxalic acid treatment on sediment arsenic concentrations and lability under reducing conditions. *Journal of Hazardous Materials*, 311, 125-133.
- Sun, Chillrud, Mailloux, B. J., Stute, M., Singh, R., Dong, H., et al. (2016). Enhanced and stabilized arsenic retention in microcosms through the microbial oxidation of ferrous iron by nitrate. *Chemosphere*, 144, 1106-1115.
- Sun, Li, X., Pi, J., Sun, Y., Li, B., Jin, Y., et al. (2006). Current research problems of chronic arsenicosis in China. *Journal of Health, Population and Nutrition*, 176-181.
- Sun, Liu, S., Li, B., Sun, X., Guo, X., Qian, C., et al. (2001). Current situation of endemic arsenicosis in China. *Environmental Sciences: an International Journal of Environmental Physiology and Toxicology*, 8(5), 425-434.
- Sun, Xiao, Dong, Y., Tang, S., Krumins, V., Ning, Z., et al. (2016). Profiling microbial community in a watershed heavily contaminated by an active antimony (Sb) mine in Southwest China. *Science of the Total Environment*, 550, 297-308.
- Suresh, K., Prabagaran, S., Sengupta, S. and Shivaji, S. (2004). *Bacillus* indicus sp. nov., an arsenic-resistant bacterium isolated from an aquifer in West Bengal, India. *International Journal of Systematic and Evolutionary Microbiology*, 54(4), 1369-1375.
- Tabaraki, R. and Heidarizadi, E. (2018). Simultaneous biosorption of Arsenic (III) and Arsenic (V): Application of multiple response optimizations. *Ecotoxicology and Environmental Safety*, 166, 35-41.
- Tabassum, R. A., Shahid, M., Dumat, C., Niazi, N. K., Khalid, S., Shah, N. S., et al. (2019). Health risk assessment of drinking arsenic-containing groundwater in Hasilpur, Pakistan: effect of sampling area, depth, and source. *Environmental Science and Pollution Research*, 26(20), 20018-20029.
- Tamaki, S. and Frankenberger, W. T. (1992). Environmental biochemistry of arsenic *Reviews of environmental contamination and toxicology* (pp. 79-110): Springer.
- Teclu, D., Tivchev, G., Laing, M. and Wallis, M. (2008). Bioremoval of arsenic species from contaminated waters by sulphate-reducing bacteria. *Water Research*, 42(19), 4885-4893.
- Texier, A.-C., Andres, Y., Faur-Brasquet, C. and Le Cloirec, P. (2002). Fixed-bed study for lanthanide (La, Eu, Yb) ions removal from aqueous solutions by immobilized Pseudomonas aeruginosa: experimental data and modelization. *Chemosphere*, 47(3), 333-342.
- Theivarasu, C. and Mylsamy, S. (2010). Equilibrium and kinetic adsorption studies of Rhodamine-B from aqueous solutions using cocoa (Theobroma cacao) shell as a new adsorbent. *International Journal of Engineering Science and Technology*, 2, 6284-6292.
- Titah, H. S., Halmi, M. I. E. B., Abdullah, S. R. S., Hasan, H. A., Idris, M. and Anuar, N. (2018). Statistical optimization of the phytoremediation of arsenic by Ludwigia octovalvis-in a pilot reed bed using response surface methodology (RSM) versus an artificial neural network (ANN). *International Journal of Phytoremediation*, 20(7), 721-729.
- Tiwari, S., Sarangi, B. K. and Thul, S. T. (2016). Identification of arsenic resistant endophytic bacteria from Pteris vittata roots and characterization for arsenic remediation application. *Journal of Environmental Management*, 180, 359- 365.
- Tokunaga, S. and Hakuta, T. (2002). Acid washing and stabilization of an artificial arsenic-contaminated soil. *Chemosphere*, 46(1), 31-38.
- Torsvik, V., Goksøyr, J. and Daae, F. L. (1990). High diversity in DNA of soil bacteria. *Applied and Environmental Microbiology*, 56(3), 782-787.
- Tsai, Y.-P., You, S.-J., Pai, T.-Y. and Chen, K.-W. (2005). Effect of cadmium on composition and diversity of bacterial communities in activated sludges. *International Biodeterioration & Biodegradation*, 55(4), 285-291.
- Turan, N. G., Mesci, B. and Ozgonenel, O. (2011a). Artificial neural network (ANN) approach for modeling Zn (II) adsorption from leachate using a new biosorbent. *Chemical Engineering Journal*, 173(1), 98-105.
- Turan, N. G., Mesci, B. and Ozgonenel, O. (2011b). The use of artificial neural networks (ANN) for modeling of adsorption of Cu (II) from industrial leachate by pumice. *Chemical Engineering Journal*, 171(3), 1091-1097.
- Turpeinen, R., Pantsar-Kallio, M. and Kairesalo, T. (2002). Role of microbes in controlling the speciation of arsenic and production of arsines in contaminated soils. *Science of the Total Environment*, 285(1-3), 133-145.
- Tyson, G. W., Chapman, J., Hugenholtz, P., Allen, E. E., Ram, R. J., Richardson, P. M., et al. (2004). Community structure and metabolism through reconstruction of microbial genomes from the environment. *Nature*, 428(6978), 37.
- Uddin, M. T., Mozumder, M. S. I., Figoli, A., Islam, M. A. and Drioli, E. (2007). Arsenic removal by conventional and membrane technology: An overview. *Indian Journal of Chemical Technology*, 14(5), 441-450.
- Ungureanu, G., Santos, S., Boaventura, R. and Botelho, C. (2015). Arsenic and antimony in water and wastewater: overview of removal techniques with special reference to latest advances in adsorption. *Journal of Environmental Management*, 151, 326-342.
- Valls, M. and De Lorenzo, V. (2002). Exploiting the genetic and biochemical capacities of bacteria for the remediation of heavy metal pollution. *FEMS Microbiology Reviews*, 26(4), 327-338.
- Vancanneyt, M., Schut, F., Snauwaert, C., Goris, J., Swings, J. and Gottschal, J. (2001). Sphingomonas alaskensis sp. nov., a dominant bacterium from a marine oligotrophic environment. *International Journal of Systematic and Evolutionary Microbiology*, 51(1), 73-79.
- Veglio, F. and Beolchini, F. (1997). Removal of metals by biosorption: a review. *Hydrometallurgy*, 44(3), 301-316.
- Velasquez, L. and Dussan, J. (2009). Biosorption and bioaccumulation of heavy metals on dead and living biomass of *Bacillus sphaericus*. *Journal of Hazardous Materials*, 167(1-3), 713-716.
- Vijayaraghavan, K. and Yun, Y. S. (2008). Bacterial biosorbents and biosorption. *Biotechnology Advances*, 26(3), 266-291.
- Volesky, B. (1987). Biosorbents for metal recovery. *Trends in Biotechnology*, 5(4), 96-101.
- Wan, W. S., Kamari, A. and Koay, Y. J. (2004). Equilibrium and kinetics studies of adsorption of copper (II) on chitosan and chitosan/PVA beads. *International Journal of Biological Macromolecules*, 34, 155-161.
- Wang, Jeng, J.-S., Yip, P.-K., Chen, C.-L., Hsu, L.-I., Hsueh, Y.-M., et al. (2002). Biological gradient between long-term arsenic exposure and carotid atherosclerosis. *Circulation*, 105(15), 1804-1809.
- Wang and Mulligan, C. N. (2006). Occurrence of arsenic contamination in Canada: sources, behavior and distribution. *Science of the total Environment*, 366(2- 3), 701-721.
- Wang, Sheng, H.-F., He, Y., Wu, J.-Y., Jiang, Y.-X., Tam, N. F.-Y., et al. (2012). Comparison of bacterial diversity in freshwater, intertidal wetland, and

marine sediments using millions of Illumina tags. *Applied and Environmental Microbiology*, AEM. 01821-01812.

- Weeger, W., Lievremont, D., Perret, M., Lagarde, F., Hubert, J.-C., Leroy, M., et al. (1999). Oxidation of arsenite to arsenate by a bacterium isolated from an aquatic environment. *Biometals*, 12(2), 141-149.
- Wirsen, C. O., Sievert, S. M., Cavanaugh, C. M., Molyneaux, S. J., Ahmad, A., Taylor, L., et al. (2002). Characterization of an autotrophic sulfide-oxidizing marine Arcobacter sp. that produces filamentous sulfur. *Applied and Environmental Microbiology*, 68(1), 316-325.
- Wu, Feng, S. X., Li, B. and Mi, X. M. (2010). The characteristics of Escherichia coli adsorption of arsenic (III) from aqueous solution. *World Journal of Microbiology and Biotechnology*, 26(2), 249-256.
- Wu, Liu, Q., Li, Z., Cheng, W., Sun, J., Guo, Z., et al. (2018). Environmental factors shaping the diversity of bacterial communities that promote rice production. *BMC Microbiology*, 18(1), 51.
- Xie, Z., Luo, Y., Wang, Y., Xie, X. and Su, C. (2013). Arsenic resistance and bioaccumulation of an indigenous bacterium isolated from aquifer sediments of Datong Basin, Northern China. *Geomicrobiology Journal*, 30(6), 549-556.
- Yamamura, S. and Amachi, S. (2014). Microbiology of inorganic arsenic: from metabolism to bioremediation. *Journal of Bioscience and Bioengineering*, 118(1), 1-9.
- Yan, L., Yin, H., Zhang, S., Leng, F., Nan, W. and Li, H. (2010). Biosorption of inorganic and organic arsenic from aqueous solution by *Acidithiobacillus ferrooxidans* BY-3. *Journal of Hazardous Materials*, 178(1-3), 209-217.
- Yannarell, A. C. and Triplett, E. W. (2005). Geographic and environmental sources of variation in lake bacterial community composition. *Applied and Environmental Microbiology*, 71(1), 227-239.
- Yetilmezsoy, K. (2006). Determination of optimum body diameter of air cyclones using a new empirical model and a neural network approach. *Environmental Engineering Science*, 23(4), 680-690.
- Yetilmezsoy, K. and Demirel, S. (2008). Artificial neural network (ANN) approach for modeling of Pb (II) adsorption from aqueous solution by Antep pistachio (Pistacia Vera L.) shells. *Journal of Hazardous Materials*, 153(3), 1288-1300.
- Yooseph, S., Nealson, K. H., Rusch, D. B., McCrow, J. P., Dupont, C. L., Kim, M., et al. (2010). Genomic and functional adaptation in surface ocean planktonic prokaryotes. *Nature*, 468(7320), 60.
- Yoshida, T., Yamauchi, H. and Sun, G. F. (2004). Chronic health effects in people exposed to arsenic via the drinking water: dose–response relationships in review. *Toxicology and Applied Pharmacology*, 198(3), 243-252.
- Yu, H. S., Lee, C. H. and Chen, G. S. (2002). Peripheral vascular diseases resulting from chronic arsenical poisoning. *The Journal of Dermatology*, 29(3), 123- 130.
- Zaini, H., Mohd, N. A. R., Siti, M. S., Yamin, Y. and Ahmad, S. (2011). Removal of copper from aqueous solution by adsorption using magnesium aluminium hydrogen phosphate layered double hydroxides. *Journal of Nuclear and Related Technologies*, 8(2), 60-67.
- Zhang, P., Zhang, N., Li, Z., Yean, S., Li, H., Shipley, H. J., et al. (2018). Identification of a new high-molecular-weight Fe− citrate species at low citrate-to-Fe molar ratios: Impact on arsenic removal with ferric hydroxide. *Chemosphere*.
- Zheng, Y., Stute, M., Van Geen, A., Gavrieli, I., Dhar, R., Simpson, H., et al. (2004). Redox control of arsenic mobilization in Bangladesh groundwater. *Applied Geochemistry*, 19(2), 201-214.
- Zhu, Y.-G., Xue, X.-M., Kappler, A., Rosen, B. P. and Meharg, A. A. (2017). Linking genes to microbial biogeochemical cycling: lessons from arsenic. *Environmental Science & Technology*, 51(13), 7326-7339.

LIST OF PUBLICATIONS

- 1) Altowayti, W.A.H., Algaifi, H.A., Bakar, S.A. and Shahir, S., 2019. **The adsorptive removal of As (III) using biomass of arsenic resistant** *Bacillus thuringiensis* **strain WS3: Characteristics and modelling studies.** *Ecotoxicology and environmental safety*, *172*, pp.176-185.
- 2) Altowayti, W.A.H and Shahir, S., 2018. **The Effect of Acid-Washing Pretreatment on Arsenic Removal by Dried Biomass of Indigenous Arsenic Resistant Bacteria**. THE INTERNATIONAL CONFERENCE ON GLOBAL & EMERGING TRENDS 2018 (ICGET) held on 2nd – 4th May 2018 at the Baze University, Abuja, Nigeria.
- 3) Altowayti, W.A.H, Dahawi, A.A., and Shahir, S., (**Significance of Bio-treatment by Acid Washing for Enlargement of Arsenic Desorption in Indigenous Arsenic-Resistant Bacteria from Gold Mine**) *Malaysian Journal of Fundamental and Applied Sciences* (Accepted)

JOURNAL UNDER REVIEW/COMMUNICATED:

- 1) Altowayti, W.A.H, Almoalemi, H., and Shahir, S., (Isolation And Identification Of Indigenous Arsenic Resistant Microbes From Arsenic-Rich Mine Tailings Using Culture-Independent And Dependent Approaches)
- 2) Altowayti, W.A.H, Haris, S. A., Shahir, S., Zakaria, Z., Ibrahim, S. (Removal of As (III) and As (V) By Using Mixed Dried Biomass of Three Indigenous Arsenic Resistant Microbes Isolated From Gold Mining Environment)