IN SITU SYNTHESIS OF BIO-GREEN SILVER NANOPARTICLES-INCORPORATED ZEOLITE A USING *Orthosiphon aristatus* LEAVES EXTRACT FOR ANTIBACTERIAL WOUND HEALING

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DEDICATION

This thesis is fully dedicated to my late grandmother, supervisors, families, friends and colleagues for all the knowledge and sharing given to me along this challenging path. It is also dedicated to myself for the persistence and perseverance, which I was able to put upon myself from the starting line until the end of this meaningful journey.

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ABSTRACT

Silver in its various forms is well known to have a potent antibacterial property. Despite the high antibacterial activity and efficacy of silver nanoparticles (AgNP), its frequent use could lead to bacterial resistance. Without a proper release mechanism, the efficacy of AgNP is often questioned. Additionally, chemical and physical methods to synthesize AgNP pose threats to the environment and health. Thus, alternative approach using biological resources are desired. However, AgNP produced through this method still needs preclinical evaluation on toxicity and biocompatibility. Thus, a novel in situ biosynthesis of AgNP-incorporated synthesized zeolite A (AgNP-SZ) was developed. The AgNP-SZ was then assessed for their antibacterial activity, in vitro cytotoxicity and wound healing potency. Zeolite A (SZ) was synthesized from kaolinite through hydrothermal method whereas AgNP was produced from AgNO₃ using Orthosiphon aristatus leaves extract as the green reducing and capping agent. The AgNP-SZ was synthesized using 0.4 mL 5 % O. aristatus leaf extract solution and mixed physically with Ag-SZ. The synthesized materials SZ, Ag-SZ and AgNP-SZ were characterized for their morphological and physicochemical properties. In the present study, the characterization results validated that the synthesized product was zeolite A. Characterization by Transmission Electron Microscope (TEM) showed AgNP with particle size of 20.01 nm in diameter and area of 381.61 nm² was incorporated in the zeolite A. TEM analysis, surface and pore analysis (BET/BJH), thermogravimetric and differential temperature analysis (TGA-DTA), and inductively coupled plasma-optical emission spectrometry (ICP OES) were used to assess the synthesized products. These characterizations validated the O. aristatus leaves extract acted as natural reducing and capping agents with a timely release mechanism of AgNP from zeolite A. SZ, Ag-SZ and AgNP-SZ were assessed for antibacterial activity against E. coli and S. aureus using disc diffusion technique (DDT) and minimum inhibitory/bactericidal concentration (MIC/MBC), biofilm inhibition against P. aeruginosa, in vitro cytotoxicity against human skin fibroblast (HSF 1184) cells and wound healing potency through in vitro scratch assay. The powder form of the samples was pressed into pellets for DDT, whereas MIC/MBC and biofilm study utilized the powder form in both water and saline solution. Inhibition zones and bacterial growth inhibition were observed. The DDT showed clear zone of inhibitions for Ag-loaded materials on both bacteria, with E. coli was more susceptible than S. aureus in both water and saline solutions based on the MIC/MBC values. The AgNP-SZ also showed potential biofilm inhibition action against S. aureus compared to P. aeruginosa. SZ, Ag-SZ and AgNP-SZ at 0.5, 1.0, 1.5, and 2.0 mg/mL were tested for cytotoxicity. In vitro scratch assay determined the HSF 1184 cell migration rate after treatment with the synthesized products. The absence of cytotoxicity in all concentrations of AgNP-SZ proved that the material is biocompatible. Although cell migration rate by AgNP-SZ was slower compared to the SZ and control in *in vitro* scratch assay, the material did not hinder cell migration and proliferation. These findings show the potential of green synthesized AgNP-incorporated zeolite A using plant extract to substitute conventional methods, with good antibacterial application and sustainable production.

ABSTRAK

Argentum dalam pelbagai bentuk adalah sangat terkenal dengan ciri antibakteria yang kuat. Meskipun mempunyai aktiviti antibakteria yang tinggi, penggunaan nanozarah Argentum (AgNP) yang kerap menyebabkan kerintangan bakteria. Tanpa mekanisme pelepasan yang terkawal, keberkesanan AgNP sering dipertikaikan. Tambahan pula, kaedah sintesis secara kimia dan fizikal menjadi ancaman kepada alam sekitar dan kesihatan. Justeru, kaedah alternatif menggunakan sumber biologi adalah diperlukan. Namun, AgNP yang terhasil menggunakan sumber biologi masih memerlukan ujian pra-klinikal toksisiti dan bioserasi. Oleh itu, kaedah baharu biosintesis in situ AgNP sebatian sintesis zeolit A (AgNP-SZ) telah dibangunkan. AgNP-SZ telah dinilai berdasarkan aktiviti antibakteria, kadar ketoksikan sel, dan keupayaan menyembuh luka in vitro. Zeolit A (SZ) dihasilkan daripada kaolinit melalui proses hidroterma, manakala AgNP dihasilkan menggunakan ekstrak daun Orthosiphon aristatus sebagai agen penurun dan penukup. Penghasilan in situ ini telah dijalankan dengan menggunakan 0.4 mL larutan ekstrak daun O. aristatus (5%). Dalam kajian ini, kesemua sampel SZ, Ag-SZ dan AgNP-SZ telah dicirikan sifat morfologi dan fizikokimia mereka. Pencirian ini telah mengesahkan penghasilan zeolit A. Pencirian mikroskopi elektron transmisi (TEM) menunjukkan penyebatian AgNP dengan saiz zarah berdiameter 20.01 nm dan luas 381.61 nm² dalam struktur rangka zeolit A. Analisis TEM, kaedah BET/BJH, analisis termogravimetri (TGA-DTA), dan spektrometri pancaran optik-plasma terganding induktif (ICP-OES) telah digunakan untuk menilai produk yang disintesis. Pencirian tersebut mengesahkan bahawa ekstrak daun O. aristatus bertindak sebagai agen penukupan dan perlepasan terkawal AgNP daripada zeolit A. SZ, Ag-SZ dan AgNP-SZ telah dinilai menggunakan kaedah peresapan cakera (DDT) dan kepekatan merencat/bakteriasid minimum (MIC/MBC) terhadap Escherichia coli dan Staphylococcus aureus, perencatan biofilem terhadap Pseudomonas aeruginosa, serta analisis ketoksikan dan potensi menyembuh luka melalui ujian cakar in vitro ke atas sel fibroblaskulit manusia (HSF 1184). Sampel berbentuk serbuk diproses menjadi palet untuk digunakan dalam ujian DDT, manakala sampel berbentuk serbuk digunakan dalam ujian MIC/MBC dan biofilem di dalam larutan air dan garam. Zon perencatan dan pertumbuhan bakteria telah direkodkan. Hasil DDT telah menunjukkan zon perencatan bagi bahan terkandung Ag, dengan E. coli didapati lebih rentan berbanding S. aureus melalui ujian MIC/MBC. Ujian perencatan biofilem menggunakan AgNP-SZ juga menunjukkan potensi yang baik terhadap S. aureus berbanding P. aeruginosa. SZ, Ag-SZ dan AgNP-SZ pada kepekatan 0.5, 1.0, 1.5, dan 2.0 mg/mL telah dinilai bagi ujian ketoksikan sel. Ujian cakaran in vitro telah dilakukan untuk menentukan kadar penghijarahan sel HSF 1184. Ketiadaan aktiviti ketoksikan sel pada kesemua kepekatan AgNP-SZ telah membuktikan bahan tersebut adalah bersifat bioserasi. Walaupun kadar migrasi sel dilaporkan lebih perlahan bagi sampel AgNP-SZ berbanding zeolit A dan sampel kawalan dalam ujian cakaran in vitro, bahan tersebut tidak merencatkan migrasi sel. Penemuan ini menunjukkan kebolehupayaan sintesis hijau AgNP sebatian zeolit A daripada ekstrak tumbuhan berbanding teknologi lazim dengan sifat antibakteria dan bioserasi yang baik serta penghasilan produk yang lebih mampan.

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LIST OF ABBREVIATIONS

Kaol	-	Kaolinite
Meta-Kaol	-	Metakaolin
SZ	-	Synthesized zeolite A
Ag	-	Silver
AgNP	-	Silver nanoparticles
Ag-SZ	-	Silver-zeolite A
AgNP-SZ	-	Bio-synthesized AgNP incorporated zeolite A
XRD	-	X-ray diffraction
FTIR	-	Fourier transform infrared
FESEM	-	Field emission scanning electron microscope
TEM	-	Transmission electron microscope
EDX	-	Energy dispersive X-ray
FRIM	-	Forest Research Institute Malaysia
BET/ BJH	-	Brunauer-Emmett-Teller/ Barrett-Joyner-Halenda
LCMS	-	Liquid chromatography-mass spectrometry
TGA	-	Thermogravimetric analysis
DTA	-	Differential thermal analysis
NA	-	Nutrient agar
MHA	-	Mueller-Hinton agar
LB	-	Luria-Bertani
DDT	-	Disc diffusion technique
MIC	-	Minimum inhibition concentration
MBC	-	Minimum bactericidal concentration
ATCC	-	American-Type Cell Culture
HSF	-	Human skin fibroblast
MTT	-	(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium
		bromide)
PBS	-	Phosphate buffer saline
FBS	-	Fetal bovine serum

LIST OF SYMBOLS

°C	-	Degree Celsius
%	-	Percent
cm	-	Centimetre
g	-	Gram
mg	-	Milligram
μg	-	Microgram
h	-	Hour
М	-	Molar
L	-	Litre
mL	-	Millilitre
mm	-	Millimetre
μm	-	Micrometre
nm	-	Nanometre
μL	-	Microliter
v/v	-	Volume/volume
w/v	-	Weight/volume
kV	-	Kilovolt
kPa	-	Kilopascal
θ	-	Theta
Å	-	Angstrom
a.u.	-	Astronomical unit
mV	-	Millivolt
μV	-	Microvolt

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CHAPTER 1

INTRODUCTION

1.1 Research Background

Various types and compositions of clays and minerals have been frequently studied and utilized for their multipurpose applications. Studies have focused on zeolite type-A out of several types of zeolites that include A, X, and Y (common types). Zeolite A is widely used for various purposes, and it can be used as a carrier for silver ions and silver nanoparticles (AgNP) (Jiraroj, Tungasmita, and Tungasmita, 2014), biogas purification (Abdullah et al., 2018), and wastewater treatment (Mahmoodi and Saffar-Dastgerdi, 2019). It is a porous material with various molecular size channels, enabling specialized functions in catalysis, separation, and ion exchange (Kazemimoghadam, 2016). Zeolite A can be obtained through synthesis from kaolin for research purposes (Kwakye-Awuah et al., 2014; Loiola et al., 2012; Mgbemere et al., 2018). Kaolin, a type of clay, was used as the precursor for the synthesis of zeolite due to its abundant composition of silicon (Si), oxygen (O) and aluminium (Al). These elements are important in determining the outcome of the desired zeolite type. Therefore, the synthesized zeolite A was modified by the incorporation or ionexchanged with Ag and AgNP in advancing the development of composite material of zeolite A. This synergy has created potentials for advanced materials in biological and medical applications. Zeolite A composite is known for its antibacterial function after being modified with several appropriate and compatible antibacterial agents. Independently, zeolite A has no special feature to inhibit the growth of bacteria on its own. Antibacterial agents are compounds that can inhibit or kill bacteria. One of the antibacterial compounds such as silver, whether in the form of ion (Salim and Malek, 2016) or nanoparticle (Hu et al., 2016), has become a significant benchmark for incorporating onto these zeolites. Zeolite can promote wound healing with the addition of antibacterial agent (Neidrauer et al., 2014; Purnomo et al., 2018).

Malaysia is a tropical country which houses abundance of local herbs and plants. These plants contain a rich amount of phytochemical components beneficial for many uses. In this work, a local herb known scientifically as Orthosiphon aristatus or misai kucing in Malay is highlighted for its use as a green reducing agent. On the other hand, kaolinite, a type of clay, is also abundantly exist in Malaysia and is frequently used in many advanced construction technologies. These two resources are the main materials used in this work. Hence, this study is feasible and attainable to develop an impactful research finding. With reference to previous studies, the green reducing agent has become an alternative method to reduce silver (Ag) ions to silver nanoparticles (AgNP) (Helmy et al., 2020; Sytu & Camacho, 2018; Thi Lan Huong & Nguyen, 2019). Furthermore, zeolite A is also capable of being synthesized from the kaolinite (Cundy & Cox, 2005; Gougazeh & Buhl, 2014). AgNP is a worldwide nanomaterial known as an antibacterial agent and is conventionally synthesized using chemical and physical methods. This conventional method usually involves a risk and hazard to the user and requires a lot of energy input and a greener approach has been introduced over the decades using plant extract (Mittal et al., 2013). Plant extract contains a large number of phenolic compounds and alkaloids responsible for reducing Ag ions into AgNP (Mittal et al., 2013). Other than reducing capability, compounds found in plants are also able to cap the AgNP for the nanoparticles stability (Sytu & Camacho, 2018). Its nano-sized particle is also an interesting feature that is used for antibacterial application in which the very high surface area enables effective interaction with the targeted bacteria (Kim et al., 2007). AgNP is also reported as the most effective antibacterial agent against a wide range of pathogenic microorganisms (Sharma et al., 2009). The biocompatibility of the materials are often tested to assess their suitability for pre-clinical and clinical testing against human skin cells (HSF 1184) (Asraf et al., 2019), mice (Akkol et al., 2011), and zebrafish (Ramachandran et al., 2018). Following this trend, any wound healing research needs to adopt a biocompatibility test. Additionally, it can further enhance the value of the modified materials.

Therefore, this work is gearing up for green technology in the biosynthesis of AgNP and zeolite A from their sources and precursors, respectively. Primarily, this work introduced the functional term of *in situ* synthesis method of AgNP-incorporated zeolite A (AgNP-SZ) using the green reducing agent of *O. aristatus* which is the

novelty of this study. So far, Thus, the modification of synthesized zeolite A (SZ) with the biosynthesized AgNP promotes an opportunity and interesting feature in biomedical application. The research gap in this project includes the innovation of synthesizing a material using green technology and assessing the material in both applications on the antibacterial property and *in vitro* skin wound healing.

1.2 Problem Background

Synthetic zeolite has become a mainstream process due to its relevant features for the industrial application. Zeolite can be synthesized using chemical or aluminosilicate precursors which made them comparable materials to the one synthesized with biological sources. Each of the synthesis method or route defines its own strength and weakness. A chemical precursor affects a high reproducibility and purity of the end material (zeolite A) (Huang et al., 2012). However, chemicallysynthesized zeolite requires chemicals which are not readily available and may cause a health hazard. The chemical precursors in this study refer to chemical sources other than biological sources such as sodium borohydride and sodium citrate. This study focused on the abundance of kaolinite in Malaysia. Therefore, it is wise to utilize this country's abundant natural resource through the use of kaolinite as the aluminosilicate precursor to synthesize zeolite A. Besides, an important question is posed on the use of chemical and physical methods to synthesize AgNP. Although these chemicals such as sodium borohydride (NaBH₄) and trisodium citrate are good reducing agents, they are risks to the environment if not properly disposed of (Banfi et al., 2014). Other than that, a physical method such as laser ablation requires a vast amount of energy to project the beam (Wei et al., 2015). Thus, this has led to the alternative use of biological organisms, including plant, microbes, and yeast, to synthesize AgNP. This study has demonstrated the use of local herb named O. aristatus which the reducing potential rivals to that chemical process.

The frequent use of antibacterial agents such as silver, copper, and zinc has promoted bacterial resistance towards antibacterial agents. Although at a slow pace, bacteria can develop a resistive mechanism to counter the antibacterial materials (Ferreira et al., 2016). These resistances of gram-positive and gram-negative bacteria are dependent on the difference in their membrane structure such as the peptidoglycan and membrane thickness (Durán et al., 2016). In addition, available antibacterial agents such as antibiotic and antiseptic cream, gel or solution are not as effective as they were in certain cases. The reduced effectiveness is due to the allergic reaction of the skin to certain compounds and a low-dose ingredient. Thus, the development of a new material that can deliver antibacterial agents using suitable mechanisms such as controlled release, high surface area of contact and low to null toxicity is needed. The exploration of new antibacterial products also helps consumers to choose based on their preferences in the market.

There is rising concern of environmental factor when utilizing Ag in salts or nanoparticles although Ag is the most potent antibacterial agent to date. When the Ag is not properly disposed of, the environment will suffer a bioaccumulation phenomenon of heavy metals. A research was conducted to determine the level of bioaccumulation of AgNO₃ and AgNP in *daphnia magna* showing a higher amount of AgNP in the diet and water exposure (Ribeiro et al., 2017). A similar study also stated that Ag bio-accumulative index are higher in the form of AgNO₃ than AgNP in rainbow trout (Clark et al., 2019) The uncontrolled release of AgNP will accumulate in the host of the first consumer and later snowball to the end of the food chain. Humans as the end consumer will suffer the most contributing health hazard and illness. Current innovations are still lacking in utilizing natural materials such as plants and microbes in the production of AgNP. Therefore, green technology ranging from production to packaging has been the utmost concern for most industries and suppliers.

The toxicological property of the synthesized AgNP using a greener approach must also be assessed, although many claim that the green synthesis poses less threat to human health. Bio-synthesized AgNP has never been claimed to not cause any environmental issue at all. However, the use of biological synthesis would simply reduce the significant adverse effect on the environment by removing harmful byproducts of chemical synthesis. A review stated that there are many possible mechanisms that lead to an increase or decrease in the toxicity level of AgNP in general, depending on the factors such as interaction with organic matters, cations, pH, and oxygen (Clark et al., 2019). A high dose of Ag as in ions and nanoparticles will inhibit bacterial growth. However, a proper guideline is needed to assess the toxicity of these materials towards human health in general. Other than that, scientific data and results have to be produced to rectify the claim of toxicity level, thus creating a proper experimental setup on cytotoxicity testing of AgNP in the laboratory. On the other hand, wound infection is a frequent case that arises from the area of the wound being infected with bacteria. Wounded skin can be properly and quickly healed if personal hygiene is well-taken care of. However, in some cases, a wound can be severe due to this bacterial infection. According to a study done by swabbing infected wounds of patients in a hospital, common bacteria found on the swabs include *Staphylococcus* aureus, Pseudomonas aeruginosa, Proteus mirabilis, Escherichia coli, and Corynebacterium spp (Bessa et al., 2015). Undeniably, bacteria are the most common cause of wound persistence, according to the mentioned study. The wound can heal within a certain timeframe, but it can be impaired by bacterial infection. When skin is injured, bacteria can quickly obtain access to the underlying tissue making it difficult to heal (Bucknall, 1989). This will further give rise to inflammatory reaction on the skin since bacteria have the ability to form biofilm which act as their protective layer against antibacterial agent. The development of composite material with added value mainly the antibacterial property is sought. A good antibacterial activity shows decreases or kills a wide range of bacteria completely. Thus, these gaps in the research problem need to be explored and studied to produce a functional material with good antibacterial activity and biocompatible to the human skin cells.

1.3 Research Objectives

The objectives of this work are as follows:

- To synthesize zeolite A from kaolinite and bio-green silver nanoparticles from Orthosiphon aristatus extract and characterize them.
- 2. To incorporate bio-synthesized silver nanoparticles onto synthesized zeolite A using *in situ* method and characterize them.

- 3. To assess the antibacterial property of bio-synthesized silver nanoparticles immobilized on zeolite A towards gram positive and gram negative bacteria.
- 4. To evaluate the cytotoxicity and skin wound healing capability of the biosynthesized silver nanoparticles incorporated on zeolite A on human dermal fibroblast cells HSF 1184.

1.4 Research Scope

There are several important steps in producing the material for this research. Based on the objectives, the research scopes are divided into (1) the syntheses of zeolite A from kaolinite and the bio-synthesis of AgNP using *O. aristatus* leaf extract as the reducing and capping agent, (2) the *in situ* preparation method of biosynthesized AgNP-incorporated zeolite A, (3) antibacterial assay, and (4) *in vitro* biocompatibility and wound healing assessments on HSF 1184 cells.

The first scope included the synthesis of zeolite A from raw kaolinite and the green biosynthesis of AgNP using *O. aristatus* leaves extract as the reducing and capping agents. Zeolite A is well-known for its hydrothermal synthesis with various potential applications (Collins et al., 2020). This hydrothermal method transformed kaolinite into zeolite A. On the other hand, AgNP was biosynthesized using *O. aristatus* leaves extract as the reducing and capping agents. The synthesized materials (AgNP and SZ) were optimized and characterized before proceeding for *in situ* synthesis method.

In the next scope, both syntheses methods were combined into an *in situ* synthesis method through the synthesis, capping and incorporation of AgNP into the zeolite A. Similarly, the synthesized materials (Ag-SZ and AgNP-SZ) were characterized to confirm the incorporation of Ag and AgNP on or in the zeolite A framework. Figure 1.1 shows the schematic diagram of the synthesis pathway of *in situ* biosynthesized AgNP-incorporated zeolite A.

The characterization instruments used include Fourier transform infrared (FTIR) spectroscopy and X-ray powder diffraction (XRD). Furthermore, the surface morphology and properties were examined using field emission scanning electron microscopy (FESEM), energy dispersive X-ray (EDX) spectroscopy, transmission electron microscope (TEM), inductively coupled plasma-optical emission spectrometry (ICP-OES) and dispersion behaviour. Additional characterizations to complement the results were also carried out using thermogravimetric analysis (TGA), zeta potential, Brunauer-Emmett-Teller (BET) Surface Area Analysis and Barrett-Joyner-Halenda (BJH) Pore Size and Volume Analysis (BET-BJH), and inductively-coupled plasma optical emission spectroscopy (ICP-OES). Additionally, the *O. aristatus* was also identified and characterized for its phytochemical constituents. The plant extract was characterized using liquid chromatography-mass spectrometry (LCMS), and the synthesis of AgNP using the extract was optimized based on specific parameters.

The third scope emphasized the application and evaluation of the synthesized materials for antibacterial application. Disc diffusion technique (DDT) and minimum inhibitory/bactericidal concentration (MIC/MBC) were carried out against gram negative and gram positive of *Escherichia coli* ATCC 11229 and *Staphylococcus aureus* ATCC 6538. In addition, biofilm inhibition study was also conducted against *S. aureus* ATCC 6538 and *Pseudomonas aeruginosa* ATCC 15442. The synthesized materials were applied to the bacteria and assessed for the capability of Ag- and AgNP-associated materials to inhibit the bacterial growth or kill the bacteria completely.

The last scope involved the *in vitro* study of the synthesized materials towards human dermal fibroblast (HSF 1184) cells. The cells were grown and maintained until stable. Afterwards, the prepared samples were tested on the cells using cytotoxicity assay (direct method) and scratch assay (indirect method). The direct method involved a direct placement of the synthesized samples into the media containing cells. On the other hand, the indirect method required the sample extracts to be incorporated with the media for cell proliferation. Concentration of samples with no toxicity were selected and used for the scratch assay. The wound healing capability between the samples (SZ, Ag-SZ and AgNP-SZ) were evaluated and compared.

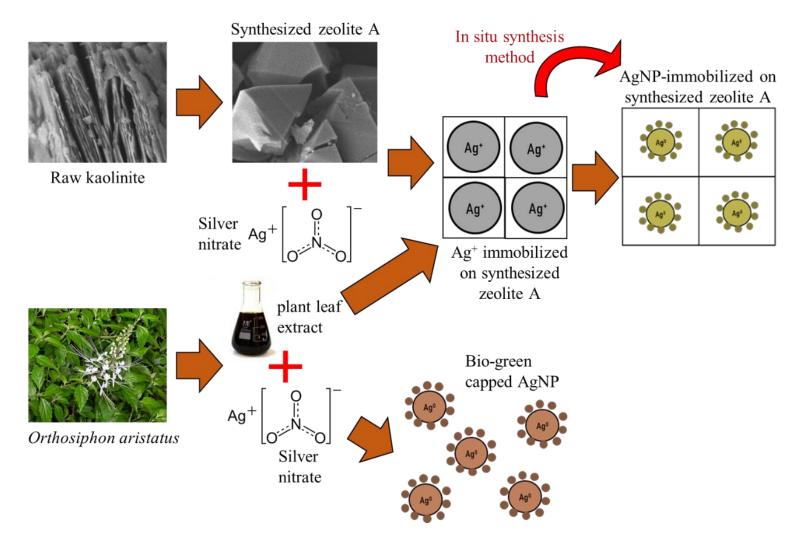


Figure 1.1 Schematic diagram of the *in situ* biosynthesized AgNP-incorporated zeolite A synthesis pathway

1.5 Research Outline

The outline of the research is shown schematically in Figure 1.2. The outline comprises of several stages divided into the synthesis of materials including zeolite A from kaolinite and AgNP using *O. aristatus*, the *in situ* synthesis antibacterial assessments and biofilm inhibition study, and biocompatibility and wound healing evaluation of the materials.

Stage 1 and 2 describes the preliminary synthesis of the respective zeolite A and AgNP. These syntheses were carried out firstly to determine the success of both synthesized materials (SZ and AgNP). SZ was synthesized from Kaol whereas AgNP was bio-synthesized from the *O. aristatus* leaves extract.

After the confirmation of the success of both synthesized materials, *in situ* method was carried out in Stage 3. In this stage, Ag-SZ and AgNP-SZ were synthesized. The difference between these two materials were the method of synthesis and the state of Ag. Ag was in the state of ions whereas AgNP was in the state of nanoparticles. In addition, Ag-SZ was used as a comparison to AgNP-SZ to assess the effect of reducing agent and capping agent derived from the phyto-compounds extracted from *O. aristatus* leaves.

Next stage was the application of the synthesized materials. Kaol, SZ, Ag-SZ and AgNP-SZ were the studied materials for antibacterial activity whereas SZ, Ag-SZ, and AgNP-SZ were tested for the *in vitro* cytotoxicity and wound healing assessments. Antibacterial assays were inclusive of DDT and MIC/MBC with the addition of biofilm inhibition study. Other than that, *in vitro* studies involved the use of cytotoxicity assay and scratch assay. In-depth methods are described in a later Chapter 3.

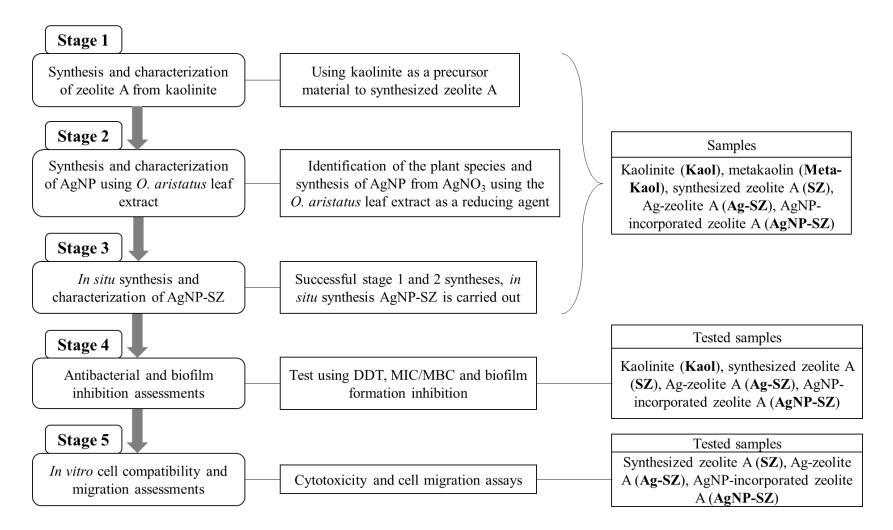


Figure 1.2 Outline of research stage-wise for the syntheses and applications of the materials.

1.6 Research Significance

The development of inorganic materials is important to enhance their attributes by giving them additional properties such as antibacterial, antifungal, promoting wound healing, and increasing livestock. It is important to show that synthetic zeolite can be synthesized abundantly because the source of raw kaolinite is available locally. Since Malaysia does not produce zeolite minerals, so our researchers can synthesize zeolites from abundantly available kaolinite sources with high aluminosilicate content (Abdullahi et al., 2019). Furthermore, the rising trend for green synthesis of nanoparticles using microbes and plants has provided vast opportunities for our researchers to utilize natural products and green approach. Our tropical region is rich with plants and herbs which are waiting to be discovered. There are many types of research in synthesizing nanoparticles using plants such as *Dodonaea visco*sa (Anandan et al., 2019), *Polygonum minus* (Ullah, Wilfred, and Shaharun, 2019) and *Sapindus mukorossi* (Thi Lan Huong & Nguyen, 2021) These green syntheses resulted in more biocompatibility, scalability, and applicability of the materials (Rajan et al., 2015).

Since there is no research conducted on the development of AgNP incorporated onto zeolite A using *O. aristatus*, this project is considered novel in its field. The intended use of this medicinal herb is due to its abundance and locality in the tropical climatic forest of the southeast of Asia (Febjislami et al. 2019). Additionally, Malaysia has many herbs for application in green technology. *O. aristatus* contains rich amounts of plant metabolites (Samidurai et al., 2020). These plant metabolites are responsible for reducing and capping the synthesized AgNP. The property of these natural metabolites is very much comparable to the conventional chemicals used to synthesize AgNP. Therefore, instead of using chemicals, plant metabolites are the potential substitutes. Potentially, all compatible materials or compounds can be adhered to or immobilized on the zeolite framework through ionic interaction (Nik, Chen, and Kaliaguine, 2011) or simply through the deposition of nanoparticles in the zeolitic pores (Jiraroj, Tungasmita, and Tungasmita, 2014). Ag ions from silver nitrate can be ion-exchanged with other cations such as sodium and magnesium ions (Ebadi Amooghin et al., 2016). On the other hand, AgNP can be deposited or adsorbed into

the zeolite A framework due to its nano-size (Shameli et al., 2011). The development of AgNP incorporated onto synthesized zeolite A using a greener approach is expected to set a benchmark for future researchers to continue discovering the potential of tropical plants for the biosynthesis of AgNP.

There is a limited finding related to the use of zeolite in wound healing. A review summarized the use of zeolite as a scaffold to enhance dermal tissue regeneration (Ninan et al., 2015). This report provided insight into the possible mode of actions of the synthesized materials and their antibacterial wound healing assessment. Thus, the outcome of this project has established a reproducible method and protocol in conducting antibacterial assay and *in vitro* study, which is acceptable and more scientific. Other than that, the scientific exploration of using local plants and herbs to synthesize AgNP will promote a greener technology application.

Wounded skin is prone to bacterial infection due to the exposed inner skin layer. Simply being said, wounded skin and bacterial infection are inseparable. Bacteria exist on the outermost layer of skin or dead skin layer. Minor wound opens up the skin which makes it easier for the bacteria to penetrate (Tomic-Canic et al., 2020). There are more than a type of bacterium resides on the skin each with different roles. Some are the contributing factors in developing chronic wound while the others are believed to promote wound healing on the skin. Therefore, antibacterial and wound healing assessments are suggested as a complementary for this type of work.

In a summary, this project is significant to promote an innovative and systematic approach in developing applied inorganic material with a greener method to solve wound infection on the skin. The method applied a biological synthesis method to replace the conventional chemical and physical methods. The systematic approach was reflected on the step-wise experimentation on antibacterial wound healing. The steps started with comprehensive antibacterial testing and followed by cytotoxicity and cell migration assessments.

REFERENCES

- Abdal Dayem, A., Hossain, M. K., Lee, S. Bin, Kim, K., Saha, S. K., Yang, G.-M., Choi, H. Y., & Cho, S.-G. (2017). The Role of Reactive Oxygen Species (ROS) in the Biological Activities of Metallic Nanoparticles. *International Journal of Molecular Sciences*, 18(1), 120. https://doi.org/10.3390/ijms18010120
- Abdullah, A. H., Mat, R., Somderam, S., Abd Aziz, A. S., & Mohamed, A. (2018). Hydrogen sulfide adsorption by zinc oxide-impregnated zeolite (synthesized from Malaysian kaolin) for biogas desulfurization. *Journal of Industrial and Engineering Chemistry*, 65, 334–342. https://doi.org/10.1016/j.jiec.2018.05.003
- Abdullahi, T., Harun, Z., Hafiz, M., Othman, D., & Nazri, K. (2019). Preliminary studies on hydrothermal synthesis of zeolite from Malaysian kaolinite clays. *Malaysian Journal of Fundamental and Applied Sciences*, 15(3), 421–425.
- Adabi, M., Naghibzadeh, M., Adabi, M., Zarrinfard, M. A., Esnaashari, S. S., Seifalian, A. M., Faridi-majidi, R., Aiyelabegan, H. T., & Ghanbari, H. (2017).
 Biocompatibility and nanostructured materials : applications in nanomedicine.
 Artificial Cells, Nanomedicine, and Biotechnology, 45(4), 833–842.
 https://doi.org/10.1080/21691401.2016.1178134
- Ahmad, N., Sharma, S., Alam, M. K., Singh, V. N., Shamsi, S. F., Mehta, B. R., & Fatma, A. (2010). Rapid synthesis of silver nanoparticles using dried medicinal plant of basil. *Colloids and Surfaces B: Biointerfaces*, 81, 81–86. https://doi.org/10.1016/j.colsurfb.2010.06.029
- Ahmed, S., Ahmad, M., Swami, B. L., & Ikram, S. (2016). A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: A green expertise. *Journal of Advanced Research*, 7(1), 17–28. https://doi.org/10.1016/j.jare.2015.02.007
- Akkol, E. K., Süntar, I., Koca, U., & Keleş, H. (2011). Wound healing activity of Rubus sanctus Schreber (Rosaceae): Preclinical study in animal models. *Evidence-Based Complementary and Alternative Medicine*, 2011, 1–6. https://doi.org/10.1093/ecam/nep137
- Akowuah, G. A., Zhari, I., Norhayati, I., Sadikun, A., & Khamsah, S. M. (2004). Sinensetin, eupatorin, 3'-hydroxy-5, 6, 7, 4 '-tetramethoxyflavone and rosmarinic

acid contents and antioxidative effect of Orthosiphon stamineus from Malaysia. *Food Chemistry*, 87(4), 559–566. https://doi.org/10.1016/j.foodchem.2004.01.008

- Amberg, M., Vandenbossche, M., & Hegemann, D. (2018). Controlled Ag release from electrically conductive coating systems. *Surface and Coatings Technology*, *336*(June 2017), 29–33. https://doi.org/10.1016/j.surfcoat.2017.10.021
- Ameer, O. Z., Salman, I. M., Asmawi, M. Z., Ibraheem, Z. O., & Yam, M. F. (2012). Orthosiphon stamineus: Traditional uses, phytochemistry, pharmacology, and toxicology. *Journal of Medicinal Food*, 15(8), 678–690. https://doi.org/10.1089/jmf.2011.1973
- Anandan, M., Poorani, G., Boomi, P., Varunkumar, K., Anand, K., Chuturgoon, A. A., Saravanan, M., & Gurumallesh Prabu, H. (2019). Green synthesis of anisotropic silver nanoparticles from the aqueous leaf extract of Dodonaea viscosa with their antibacterial and anticancer activities. *Process Biochemistry*, 80, 80–88. https://doi.org/10.1016/j.procbio.2019.02.014
- Anastas, P. T., & Warner, J. C. (1998). *Green Chemistry: Theory and Practice*. Oxford University Press. https://books.google.com.my/books?id=SrO8QgAACAAJ
- Andrews, J. M. (2005). BSAC standardized disc susceptibility testing method (version
 4). Journal of Antimicrobial Chemotherapy, 56(1), 60–76. https://doi.org/10.1093/jac/dki124
- Arnida, Malugin, A., & Ghandehari, H. (2010). Cellular uptake and toxicity of gold nanoparticles in prostate cancer cells: A comparative study of rods and spheres. *Journal of Applied Toxicology*, 30(3), 212–217. https://doi.org/10.1002/jat.1486
- Aroke, U. O., & El-Nafaty, U. A. (2014). XRF, XRD and FTIR Properties and Characterization of HDTMA-Br Surface Modified Organo-Kaolinite Clay. *International Journal of Emerging Technology and Advanced Engineering*, 4(4), 817–825.
- Arya, V., & Bala, M. (2013). Biological Synthesis of Silver Nanoparticles from Aqueous Extract of Endophytic Fungus Aspergillus fumigatus and Its Antibacterial Action. *International Journal of Nanomaterials and Biostructures*, 3(2), 37–41.
- Ashkarran, A. A., & Bayat, A. (2013). Surface plasmon resonance of metal nanostructures as a complementary technique for microscopic size measurement. *International Nano Letters*, 3(1), 1–10. https://doi.org/10.1186/2228-5326-3-50

- Asraf, M. H., Malek, N. A. N. N., Jemon, K., Sani, N. S. N. S., & Muhammad, M. S. (2019). Antibacterial, cytotoxicity and wound healing assessments of amine-functionalised zeolite Y. *Particuology*, 45, 116–123. https://doi.org/10.1016/j.partic.2018.09.006
- Autef, A., Joussein, E., Gasgnier, G., Pronier, S., Sobrados, I., Sanz, J., & Rossignol,
 S. (2013). Role of metakaolin dehydroxylation in geopolymer synthesis. In *Powder Technology* (Vol. 250, pp. 33–39).
 https://doi.org/10.1016/j.powtec.2013.09.022
- Ayele, L., Pérez-Pariente, J., Chebude, Y., & Díaz, I. (2015). Synthesis of zeolite A from Ethiopian kaolin. *Microporous and Mesoporous Materials*, 215, 29–36. https://doi.org/https://doi.org/10.1016/j.micromeso.2015.05.022
- Ayele, L., Pérez-Pariente, J., Chebude, Y., & Díaz, I. (2016). Conventional versus alkali fusion synthesis of zeolite A from low grade kaolin. *Applied Clay Science*, 132–133, 485–490. https://doi.org/10.1016/j.clay.2016.07.019
- Azizi, S., Mohamad, R., Abdul Rahim, R., Mohammadinejad, R., & Bin Ariff, A. (2017). Hydrogel beads bio-nanocomposite based on Kappa-Carrageenan and green synthesized silver nanoparticles for biomedical applications. *International Journal of Biological Macromolecules*, 104, 423–431. https://doi.org/10.1016/j.ijbiomac.2017.06.010
- B. Coyle, M. (2005). Manual of antimicrobial susceptibility testing. In M. B. Coyle (Ed.), *American Society for Microbiology*. American Society for Microbiology. https://doi.org/10.1007/s13398-014-0173-7.2
- Babu, P. J., Doble, M., & Raichur, A. M. (2018). Silver oxide nanoparticles embedded silk fibroin spuns: Microwave mediated preparation, characterization and their synergistic wound healing and anti-bacterial activity. *Journal of Colloid and Interface Science*, 513, 62–71. https://doi.org/10.1016/j.jcis.2017.11.001
- Bahuguna, A., Khan, I., Bajpai, V. K., & Kang, S. C. (2017). MTT assay to evaluate the cytotoxic potential of a drug. *Bangladesh Journal of Pharmacology*, 12(2), 115–118. https://doi.org/10.3329/bjp.v12i2.30892
- Bajpai, S. K., Mohan, Y. M., Bajpai, M., Tankhiwale, R., & Thomas, V. (2007).
 Synthesis of polymer stabilized silver and gold nanostructures. *Journal of Nanoscience and Nanotechnology*, 7(9), 2994–3010. https://doi.org/10.1166/jnn.2007.911

Balouiri, M., Sadiki, M., & Ibnsouda, S. K. (2016). Methods for in vitro evaluating

antimicrobial activity: A review. *Journal of Pharmaceutical Analysis*, 6(2), 71–79. https://doi.org/10.1016/j.jpha.2015.11.005

- Banerjee, S., Loza, K., Meyer-Zaika, W., Prymak, O., & Epple, M. (2014). Structural evolution of silver nanoparticles during wet-chemical synthesis. *Chemistry of Materials*, 26(2), 951–957. https://doi.org/10.1021/cm4025342
- Banfi, L., Narisano, E., Riva, R., Stiasni, N., Hiersemann, M., Yamada, T., & Tsubo,
 T. (2014). Sodium Borohydride. In *Encyclopedia of Reagents for Organic* Synthesis (pp. 1–13). American Cancer Society. https://doi.org/https://doi.org/10.1002/047084289X.rs052.pub3
- Barbieri, R., Coppo, E., Marchese, A., Daglia, M., Sobarzo-Sánchez, E., Nabavi, S. F., & Nabavi, S. M. (2017). Phytochemicals for human disease: An update on plantderived compounds antibacterial activity. *Microbiological Research*, 196, 44–68. https://doi.org/10.1016/j.micres.2016.12.003
- Bardestani, R., Patience, G. S., & Kaliaguine, S. (2019). Experimental methods in chemical engineering: specific surface area and pore size distribution measurements—BET, BJH, and DFT. *Canadian Journal of Chemical Engineering*, 97(11), 2781–2791. https://doi.org/10.1002/cjce.23632
- Behrends, V., Giskeødegård, G. F., Bravo-Santano, N., Letek, M., & Keun, H. C. (2019). Acetaminophen cytotoxicity in HepG2 cells is associated with a decoupling of glycolysis from the TCA cycle, loss of NADPH production, and suppression of anabolism. *Archives of Toxicology*, 93(2), 341–353. https://doi.org/10.1007/s00204-018-2371-0
- Bergaya, F., Dion, P., Alcover, J. F., Clinard, C., & Tchoubar, D. (1996). TEM study of kaolinite thermal decomposition by controlled-rate thermal analysis. *Journal* of Materials Science, 31(19), 5069–5075. https://doi.org/10.1007/BF00355907
- Berthomieu, C., & Hienerwadel, R. (2009). Fourier transform infrared (FTIR) spectroscopy. *Photosynthesis Research*, 101(2–3), 157–170. https://doi.org/10.1007/s11120-009-9439-x
- Bessa, L. J., Fazii, P., Di Giulio, M., & Cellini, L. (2015). Bacterial isolates from infected wounds and their antibiotic susceptibility pattern: Some remarks about wound infection. *International Wound Journal*, 12(1), 47–52. https://doi.org/10.1111/iwj.12049
- Bhuyar, P., Rahim, M. H. A., Sundararaju, S., Ramaraj, R., Maniam, G. P., & Govindan, N. (2020). Synthesis of silver nanoparticles using marine macroalgae

Padina sp. and its antibacterial activity towards pathogenic bacteria. *Beni-Suef University Journal of Basic and Applied Sciences*, 9(3), 1–15. https://doi.org/10.1186/s43088-019-0031-y

- Bliss, D. (2010). *Skin Anatomy*. National Cancer Institute. https://visualsonline.cancer.gov/details.cfm?imageid=4604
- Boncler, M., Rózalski, M., Krajewska, U., Podswdek, A., & Watala, C. (2014).
 Comparison of PrestoBlue and MTT assays of cellular viability in the assessment of anti-proliferative effects of plant extracts on human endothelial cells. *Journal of Pharmacological and Toxicological Methods*, 69(1), 9–16. https://doi.org/10.1016/j.vascn.2013.09.003
- Borah, D., Das, N., Das, N., Bhattacharjee, A., Sarmah, P., Ghosh, K., Chandel, M., Rout, J., Pandey, P., Ghosh, N. N., & Bhattacharjee, C. R. (2020). Alga-mediated facile green synthesis of silver nanoparticles: Photophysical, catalytic and antibacterial activity. *Applied Organometallic Chemistry*, 34(5), 1–10. https://doi.org/10.1002/aoc.5597
- Bucknall, T. E. (1989). Factors affecting wound healing. Problems in General Surgery, 6(2), 194–219. https://doi.org/10.1177/0022034509359125
- Bykkam, S., Ahmadipour, M., Narisngam, S., Kalagadda, V. R., & Chidurala, S. C. (2015). Extensive Studies on X-Ray Diffraction of Green Synthesized Silver Nanoparticles. *Advances in Nanoparticles*, 04(01), 1–10. https://doi.org/10.4236/anp.2015.41001
- Calonje, J. E., Brenn, T., Lazar, A. J., & McKee, P. H. (2011). *Pathology of the Skin E-Book*. Elsevier Health Sciences.
- Cameron, S. J., Hosseinian, F., & Willmore, W. G. (2018). A current overview of the biological and cellular effects of nanosilver. *International Journal of Molecular Sciences*, 19(7), 1–40. https://doi.org/10.3390/ijms19072030
- Carvalho, I., Curado, M., Palacio, C., Carvalho, S., & Cavaleiro, A. (2019). Ag release from sputtered Ag/a:C nanocomposite films after immersion in pure water and NaCl solution. *Thin Solid Films*, 671(August 2018), 85–94. https://doi.org/10.1016/j.tsf.2018.12.010
- Chae, S. Y., Lee, S. Y., & Joo, O. S. (2019). Directly synthesized silver nanoparticles on gas diffusion layers by electrospray pyrolysis for electrochemical CO 2 reduction. *Electrochimica Acta*, 303, 118–124. https://doi.org/10.1016/j.electacta.2019.02.046

- Chung, Y.-S., Choo, B. K. M., Ahmed, P. K., Othman, I., & Shaikh, M. F. (2020). A Systematic Review of the Protective Actions of Cat's Whiskers (Misai Kucing) on the Central Nervous System. *Frontiers in Pharmacology*, *11*, 692. https://doi.org/10.3389/fphar.2020.00692
- Clark, N. J., Boyle, D., Eynon, B. P., & Handy, R. D. (2019). Dietary exposure to silver nitrate compared to two forms of silver nanoparticles in rainbow trout: Bioaccumulation potential with minimal physiological effects. *Environmental Science: Nano*, 6(5), 1393–1405. https://doi.org/10.1039/c9en00261h
- Clogston, J. D., & Patri, A. K. (2011). Zeta potential measurement. In Methods in molecular biology (Clifton, N.J.) (Vol. 697, pp. 63–70). https://doi.org/10.1007/978-1-60327-198-1_6
- Collins, F., Rozhkovskaya, A., Outram, J. G., & Millar, G. J. (2020). A critical review of waste resources, synthesis, and applications for Zeolite LTA. *Microporous and Mesoporous Materials*, 291, 109667. https://doi.org/10.1016/j.micromeso.2019.109667
- Cooper, R., & Lawrence, J. C. (1996). The isolation and identification of bacteria from wounds. *Journal of Wound Care*, 5(7), 335–340. https://doi.org/10.12968/jowc.1996.5.7.335
- Cundy, C. S., & Cox, P. A. (2005). The hydrothermal synthesis of zeolites: Precursors, intermediates and reaction mechanism. *Microporous and Mesoporous Materials*, 82(1–2), 1–78. https://doi.org/10.1016/j.micromeso.2005.02.016
- Cyril, N., George, J. B., Joseph, L., Raghavamenon, A. C., & Sylas, V. P. (2019).
 Assessment of antioxidant, antibacterial and anti-proliferative (lung cancer cell line A549) activities of green synthesized silver nanoparticles from Derris trifoliata. *Toxicology Research*, 8(2), 297–308. https://doi.org/10.1039/C8TX00323H
- da Silva Filho, S. H., Vinaches, P., Silva, H. L. G., & Pergher, S. B. C. (2020). LTA zeolite synthesis using natural materials and its evaluation by Green Star methodology. *SN Applied Sciences*, 2(3), 1–6. https://doi.org/10.1007/s42452-020-2162-0
- Damsud, T., Grace, M. H., Adisakwattana, S., & Phuwapraisirisan, P. (2014).
 Orthosiphol A from the aerial parts of Orthosiphon aristatus is putatively responsible for hypoglycemic effect via α-glucosidase inhibition. *Natural Product Communications*, 9(5), 639–641.

https://doi.org/10.1177/1934578x1400900512

- Danaei, M., Dehghankhold, M., Ataei, S., Hasanzadeh Davarani, F., Javanmard, R., Dokhani, A., Khorasani, S., & Mozafari, M. R. (2018). Impact of particle size and polydispersity index on the clinical applications of lipidic nanocarrier systems. *Pharmaceutics*, 10(2), 1–17. https://doi.org/10.3390/pharmaceutics10020057
- Demirci, S., Ustaoğlu, Z., Yilmazer, G. A., Sahin, F., & Baç, N. (2014). Antimicrobial properties of zeolite-X and zeolite-A ion-exchanged with silver, copper, and zinc against a broad range of microorganisms. *Applied Biochemistry and Biotechnology*, 172(3), 1652–1662. https://doi.org/10.1007/s12010-013-0647-7
- Deng, Y., White, G. N., & Dixon, J. B. (2002). Effect of structural stress on the intercalation rate of kaolinite. *Journal of Colloid and Interface Science*, 250(2), 379–393. https://doi.org/10.1006/jcis.2001.8208
- Do, H. D., & Do, D. D. (2001). A new diffusion and flow theory for activated carbon from low pressure to capillary condensation range. *Chemical Engineering Journal*, 84(3), 295–308. https://doi.org/10.1016/S1385-8947(00)00383-1
- Dobias, J., & Bernier-Latmani, R. (2013). Silver release from silver nanoparticles in natural waters. *Environmental Science and Technology*, 47(9), 4140–4146. https://doi.org/10.1021/es304023p
- Domínguez, A. V., Algaba, R. A., Canturri, A. M., Villodres, Á. R., & Smani, Y. (2020). Antibacterial activity of colloidal silver against gram-negative and grampositive bacteria. *Antibiotics*, 9(1), 1–10. https://doi.org/10.3390/antibiotics9010036
- Durán, N., Durán, M., Jesus, M. B. De, Seabra, A. B., Fávaro, W. J., & Nakazato, G. (2016). Silver nanoparticles: A new view on mechanistic aspects on antimicrobial activity. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 12(3), 789–799. https://doi.org/10.1016/j.nano.2015.11.016
- Dutta, P., & Wang, B. (2019). Zeolite-supported silver as antimicrobial agents. Coordination Chemistry Reviews, 383, 1–29. https://doi.org/10.1016/j.ccr.2018.12.014
- Ebadi Amooghin, A., Omidkhah, M., Sanaeepur, H., & Kargari, A. (2016). Preparation and characterization of Ag+ ion-exchanged zeolite-Matrimid®5218 mixed matrix membrane for CO2/CH4 separation. *Journal of Energy Chemistry*, 25(3), 450–462. https://doi.org/10.1016/j.jechem.2016.02.004

- Febjislami, S., Kurniawati, A. N. I., Melati, M., & Wahyu, Y. (2019). Morphological characters, flowering and seed germination of the indonesian medicinal plant Orthosiphon aristatus. *Biodiversitas*, 20(2), 328–337. https://doi.org/10.13057/biodiv/d200204
- Ferdous, Z., Al-Salam, S., Greish, Y. E., Ali, B. H., & Nemmar, A. (2019). Pulmonary exposure to silver nanoparticles impairs cardiovascular homeostasis: Effects of coating, dose and time. *Toxicology and Applied Pharmacology*, 367(October 2018), 36–50. https://doi.org/10.1016/j.taap.2019.01.006
- Ferrag, C., Li, S., Jeon, K., Andoy, N. M., Sullan, R. M. A., Mikhaylichenko, S., & Kerman, K. (2021). Polyacrylamide hydrogels doped with different shapes of silver nanoparticles: Antibacterial and mechanical properties. *Colloids and Surfaces B: Biointerfaces, 197, 111397.* https://doi.org/10.1016/j.colsurfb.2020.111397
- Ferreira, L., Guedes, J. F., Almeida-Aguiar, C., Fonseca, A. M., & Neves, I. C. (2016). Microbial growth inhibition caused by Zn/Ag-Y zeolite materials with different amounts of silver. *Colloids and Surfaces B: Biointerfaces*, 142, 141–147. https://doi.org/10.1016/j.colsurfb.2016.02.042
- Floyd, M. M., Tang, J., Kane, M., & Emerson, D. (2005). Captured diversity in a culture collection: Case study of the geographic and habitat distributions of environmental isolates held at the American Type Culture Collection. *Applied and Environmental Microbiology*, 71(6), 2813–2823. https://doi.org/10.1128/AEM.71.6.2813-2823.2005
- Fratoddi, I. (2018). Hydrophobic and hydrophilic au and ag nanoparticles. Breakthroughs and perspectives. Nanomaterials, 8(1). https://doi.org/10.3390/nano8010011
- Fu, H., Li, Y., Yu, Z., Shen, J., Li, J., Zhang, M., Ding, T., Xu, L., & Lee, S. S. (2020). Ammonium removal using a calcined natural zeolite modified with sodium nitrate. *Journal of Hazardous Materials*, 393(October 2019). https://doi.org/10.1016/j.jhazmat.2020.122481
- Fultz, B., & Howe, J. M. (2012). Transmission Electron Microscopy and Diffractometry of Materials. Springer Berlin Heidelberg. https://books.google.com.my/books?id=tgOC3vlEY0sC
- García-Barrasa, J., López-De-luzuriaga, J. M., & Monge, M. (2011). Silver nanoparticles: Synthesis through chemical methods in solution and biomedical

applications. *Central European Journal of Chemistry*, 9(1), 7–19. https://doi.org/10.2478/s11532-010-0124-x

- Gaudreault, R., Van De Ven, T. G. M., & Whitehead, M. A. (2006). A theoretical study of the interactions of water with gallic acid and a PEO/TGG complex. *Molecular Simulation*, *32*(1), 17–27. https://doi.org/10.1080/08927020500492047
- Gemeay, A. H., Aboelfetoh, E. F., & El-Sharkawy, R. G. (2018). Immobilization of Green Synthesized Silver Nanoparticles onto Amino-Functionalized Silica and Their Application for Indigo Carmine Dye Removal. *Water, Air, and Soil Pollution*, 229(16), 1–17. https://doi.org/10.1007/s11270-017-3670-4
- Ghadiri, M., Chrzanowski, W., & Rohanizadeh, R. (2015). Biomedical applications of cationic clay minerals. *RSC Advances*, 5(37), 29467–29481. https://doi.org/10.1039/c4ra16945j
- Gholami-Shabani, M., Akbarzadeh, A., Norouzian, D., Amini, A., Gholami-Shabani,
 Z., Imani, A., Chiani, M., Riazi, G., Shams-Ghahfarokhi, M., & RazzaghiAbyaneh, M. (2014). Antimicrobial activity and physical characterization of
 silver nanoparticles green synthesized using nitrate reductase from Fusarium
 oxysporum. *Applied Biochemistry and Biotechnology*, 172(8), 4084–4098.
 https://doi.org/10.1007/s12010-014-0809-2
- Gopal, J., Chun, S., Anthonydhason, V., Jung, S., Mwang'ombe, B. N., Muthu, M., & Sivanesan, I. (2018). Assays Evaluating Antimicrobial Activity of Nanoparticles:
 A Myth Buster. *Journal of Cluster Science*, 29(2), 207–213. https://doi.org/10.1007/s10876-018-1334-1
- Gougazeh, M., & Buhl, J. C. (2014). Synthesis and characterization of zeolite A by hydrothermal transformation of natural Jordanian kaolin. *Journal of the Association of Arab Universities for Basic and Applied Sciences*, 15(1), 35–42. https://doi.org/10.1016/j.jaubas.2013.03.007
- Guo, D., Dou, D., Ge, L., Huang, Z., Wang, L., & Gu, N. (2015). A caffeic acid mediated facile synthesis of silver nanoparticles with powerful anti-cancer activity. *Colloids and Surfaces B: Biointerfaces*, 134, 229–234. https://doi.org/10.1016/j.colsurfb.2015.06.070
- Hall, B. G., Acar, H., Nandipati, A., & Barlow, M. (2013). Growth Rates Made Easy. *Molecular Biology and Evolution*, 31(1), 232–238.
 https://doi.org/10.1093/molbev/mst187
- Hanim, S. A. M., Malek, N. A. N. N., & Ibrahim, Z. (2017). Analyses of surface area,

porosity, silver release and antibacterial activity of amine-functionalized, silverexchanged zeolite NaY. *Vacuum*, *143*, 344–347. https://doi.org/10.1016/j.vacuum.2017.06.038

- Helmy, A., El-Shazly, M., Seleem, A., Abdelmohsen, U., Salem, M. A., Samir, A., Rabeh, M., Elshamy, A., & Singab, A. N. B. (2020). The synergistic effect of biosynthesized silver nanoparticles from a combined extract of parsley, corn silk, and gum arabic: In vivo antioxidant, anti-inflammatory and antimicrobial activities. *Materials Research Express*, 7(2), 025002. https://doi.org/10.1088/2053-1591/ab6e2d
- Herigstad, B., Hamilton, M., & Heersink, J. (2001). How to optimize the drop plate method for enumerating bacteria. In *Journal of Microbiological Methods* (Vol. 44, Issue 2, pp. 121–129). https://doi.org/10.1016/S0167-7012(00)00241-4
- Hissae Yassue-Cordeiro, P., Henrique Zandonai, C., Pereira Genesi, B., Santos Lopes,
 P., Sanchez-Lopez, E., Luisa Garcia, M., Regina Camargo Fernandes-Machado,
 N., Severino, P., B. Souto, E., & da Silva, C. (2019). Development of
 Chitosan/Silver Sulfadiazine/Zeolite Composite Films for Wound Dressing. *Pharmaceutics*, 11(10). https://doi.org/10.3390/pharmaceutics11100535
- Hollenbeck, B. L., & Rice, L. B. (2012). Intrinsic and acquired resistance mechanisms in enterococcus. *Virulence*, *3*(5), 421–569. https://doi.org/10.4161/viru.21282
- Hsu, C.-L., Hong, B.-H., Yu, Y.-S., & Yen, G.-C. (2010). Antioxidant and Anti-Inflammatory Effects of Orthosiphon aristatus and Its Bioactive Compounds. *Journal of Agricultural and Food Chemistry*, 58(4), 2150–2156. https://doi.org/10.1021/jf903557c
- Hu, X., Bai, J., Hong, H., & Li, C. (2016). Synthesis of Ag-loaded 4A-zeolite composite catalyst via supercritical CO2 fluid for styrene epoxidation. *Microporous and Mesoporous Materials*, 228, 224–230. https://doi.org/10.1016/j.micromeso.2016.03.042
- Huang, A., Wang, N., & Caro, J. (2012). Seeding-free synthesis of dense zeolite FAU membranes on 3-aminopropyltriethoxysilane-functionalized alumina supports. *Journal of Membrane Science*, 389, 272–279. https://doi.org/10.1016/j.memsci.2011.10.036
- Huang, X., Teng, X., Chen, D., Tang, F., & He, J. (2010). The effect of the shape of mesoporous silica nanoparticles on cellular uptake and cell function. *Biomaterials*, 31(3), 438–448.

https://doi.org/10.1016/j.biomaterials.2009.09.060

- Huang, Y., Lü, X., Chen, R., & Chen, Y. (2020). Comparative study of the effects of gold and silver nanoparticles on the metabolism of human dermal fibroblasts. *Regenerative Biomaterials*, 7(2), 221–232. https://doi.org/10.1093/rb/rbz051
- Humphrys, M. S., Creasy, T., Sun, Y., Shetty, A. C., Chibucos, M. C., Drabek, E. F., Fraser, C. M., Farooq, U., Sengamalay, N., Ott, S., Shou, H., Bavoil, P. M., Mahurkar, A., & Myers, G. S. A. (2013). Simultaneous transcriptional profiling of bacteria and their host cells. *PLoS ONE*, 8(12). https://doi.org/10.1371/journal.pone.0080597
- Javanmardi, S., & Divband, B. (2017). Beneficial Effects of Ag-Exchanged Zeolite Nanocomposite on Excisional Wound in Rats. *Iranian Journal of Veterinary* Surgery, 12(26), 25–31. https://doi.org/10.22034/ivsa.2017.50823
- Jha, B., & Singh, D. N. (2016). Basics of Zeolites. In Advanced Structured Materials (Vol. 78, pp. 5–30). Springer Science+Business Media. https://doi.org/10.1007/978-981-10-1404-8
- Jiang, Q., Luo, B., Wu, Z., Gu, B., Xu, C., Li, X., & Wang, X. (2021). Corn stalk/AgNPs modified chitin composite hemostatic sponge with high absorbency, rapid shape recovery and promoting wound healing ability. *Chemical Engineering Journal*, *421*(P1), 129815. https://doi.org/10.1016/j.cej.2021.129815
- Jiraroj, D., Tungasmita, S., & Tungasmita, D. N. (2014). Silver ions and silver nanoparticles in zeolite A composites for antibacterial activity. *Powder Technology*, 264, 418–422. https://doi.org/10.1016/j.powtec.2014.05.049
- Johnson, E. B. G., & Arshad, S. E. (2014). Applied Clay Science Hydrothermally synthesized zeolites based on kaolinite : A review. *Applied Clay Science*, 97–98, 215–221. https://doi.org/10.1016/j.clay.2014.06.005
- Jonkman, J. E. N., Cathcart, J. A., Xu, F., Bartolini, M. E., Amon, J. E., Stevens, K. M., & Colarusso, P. (2014). An introduction to the wound healing assay using live-cell microscopy. *Cell Adhesion and Migration*, 8(5), 440–451. https://doi.org/10.4161/cam.36224
- Jorge de Souza, T. A., Rosa Souza, L. R., & Franchi, L. P. (2019). Silver nanoparticles: An integrated view of green synthesis methods, transformation in the environment, and toxicity. *Ecotoxicology and Environmental Safety*, 171, 691– 700. https://doi.org/10.1016/j.ecoenv.2018.12.095

- Jou, S. K., & Malek, N. A. N. N. (2016). Characterization and antibacterial activity of chlorhexidine loaded silver-kaolinite. *Applied Clay Science*, 127–128, 1–9. https://doi.org/10.1016/j.clay.2016.04.001
- Kanta, J. (2015). Collagen matrix as a tool in studying fibroblastic cell behavior. *Cell Adhesion and Migration*, *9*(4), 308–316. https://doi.org/10.1080/19336918.2015.1005469
- Kar, S., Bagchi, B., Kundu, B., Bhandary, S., Basu, R., Nandy, P., & Das, S. (2014). Synthesis and characterization of Cu/Ag nanoparticle loaded mullite nanocomposite system: A potential candidate for antimicrobial and therapeutic applications. *Biochimica et Biophysica Acta - General Subjects*, 1840(11), 3264– 3276. https://doi.org/10.1016/j.bbagen.2014.05.012
- Kartini, K., Alviani, A., Anjarwati, D., Finna Fanany, A., Sukweenadhi, J., & Avanti,
 C. (2020). Process optimization for green synthesis of silver nanoparticles using indonesian medicinal plant extracts. *Processes*, 8(8). https://doi.org/10.3390/PR8080998
- Kazemimoghadam, M. (2016). Comparison of Kaolin and chemical source for preparation of Nano pore NaA Zeolite membranes. J. Water Environ. Nanotechnol, 1(1), 45–53. https://doi.org/10.7508/jwent.2016.01.006
- Kędziora, A., Speruda, M., Krzyżewska, E., Rybka, J., Łukowiak, A., & Bugla-Płoskońska, G. (2018). Similarities and differences between silver ions and silver in nanoforms as antibacterial agents. In *International Journal of Molecular Sciences* (Vol. 19, Issue 2). https://doi.org/10.3390/ijms19020444
- Khandaker, S., Toyohara, Y., Saha, G. C., Awual, M. R., & Kuba, T. (2020). Development of synthetic zeolites from bio-slag for cesium adsorption: Kinetic, isotherm and thermodynamic studies. *Journal of Water Process Engineering*, 33(October 2019), 101055. https://doi.org/10.1016/j.jwpe.2019.101055
- Kihara, T., Zhang, Y., Hu, Y., Mao, Q., Tang, Y., & Miyake, J. (2011). Effect of composition, morphology and size of nanozeolite on its in vitro cytotoxicity. *Journal of Bioscience and Bioengineering*, 111(6), 725–730. https://doi.org/10.1016/j.jbiosc.2011.01.017
- Kim, J. S., Kuk, E., Yu, K. N., Kim, J. H., Park, S. J., Lee, H. J., Kim, S. H., Park, Y. K., Park, Y. H., Hwang, C. Y., Kim, Y. K., Lee, Y. S., Jeong, D. H., & Cho, M. H. (2007). Antimicrobial effects of silver nanoparticles. *Nanomedicine: Nanotechnology, Biology, and Medicine, 3*(1), 95–101.

https://doi.org/10.1016/j.nano.2006.12.001

- Konan, K. L., Peyratout, C., Smith, A., Bonnet, J. P., Rossignol, S., & Oyetola, S. (2009). Comparison of surface properties between kaolin and metakaolin in concentrated lime solutions. *Journal of Colloid and Interface Science*, 339(1), 103–109. https://doi.org/10.1016/j.jcis.2009.07.019
- Król, M., Minkiewicz, J., & Mozgawa, W. (2016). IR spectroscopy studies of zeolites in geopolymeric materials derived from kaolinite. *Journal of Molecular Structure*, 1126, 200–206. https://doi.org/10.1016/j.molstruc.2016.02.027
- Kwakye-Awuah, B., Von-Kiti, E., Buamah, R., Nkrumah, I., & Williams, C. (2014).
 Effect of Crystallization Time on the Hydrothermal Synthesis of Zeolites from Kaolin and Bauxite. *International Journal of Scientific & Engineering Research*, 5(2), 734–741.
- Lai, M., Liao, Z., Zhou, Z., Kao, C., Li, Y., & Wang, H. (2019). Uses of near-infrared transmission spectra for the identification of hydrothermal kaolinite, dickite and kaolinite-dickite. *Spectrochimica Acta - Part A: Molecular and Biomolecular Spectroscopy*, 208, 179–184. https://doi.org/10.1016/j.saa.2018.09.063
- Lehman, S. E., & Larsen, S. C. (2014). Zeolite and mesoporous silica nanomaterials: greener syntheses, environmental applications and biological toxicity. *Environ. Sci.: Nano*, 1(3), 200–213. https://doi.org/10.1039/C4EN00031E
- Li, J., Corma, A., & Yu, J. (2015). Synthesis of new zeolite structures. *Chem. Soc. Rev.*, 44(20), 7112–7127. https://doi.org/10.1039/C5CS00023H
- Li, Q., Tanaka, Y., & Miwa, N. (2017). Influence of hydrogen-occluding-silica on migration and apoptosis in human esophageal cells *in vitro*. *Medical Gas Research*, 7(2), 76. https://doi.org/10.4103/2045-9912.208510
- Li, Y., Li, H., Xiao, L., Zhou, L., Shentu, J., Zhang, X., & Fan, J. (2012). Hemostatic efficiency and wound healing properties of natural zeolite granules in a lethal rabbit model of complex groin injury. *Materials*, 5(12), 2586–2596. https://doi.org/10.3390/ma5122586
- Liang, C.-C., Park, A. Y., & Guan, J.-L. (2007). In vitro scratch assay: a convenient and inexpensive method for analysis of cell migration in vitro. *Nature Protocols*, 2(2), 329–333. https://doi.org/10.1038/nprot.2007.30
- Lin, J., Miao, L., Zhong, G., Lin, C. H., Dargazangy, R., & Alexander-Katz, A. (2020). Understanding the synergistic effect of physicochemical properties of nanoparticles and their cellular entry pathways. *Communications Biology*, 3(1),

1-10. https://doi.org/10.1038/s42003-020-0917-1

- Liu, Y. S., Chang, Y. C., & Chen, H. H. (2018). Silver nanoparticle biosynthesis by using phenolic acids in rice husk extract as reducing agents and dispersants. *Journal of Food and Drug Analysis*, 26(2), 649–656. https://doi.org/10.1016/j.jfda.2017.07.005
- Loiola, A. R., Andrade, J. C. R. A., Sasaki, J. M., & da Silva, L. R. D. (2012). Structural analysis of zeolite NaA synthesized by a cost-effective hydrothermal method using kaolin and its use as water softener. *Journal of Colloid and Interface Science*, 367(1), 502–508. https://doi.org/10.1016/j.jcis.2010.11.026
- Loo, Y. Y., Rukayadi, Y., Nor-Khaizura, M. A. R., Kuan, C. H., Chieng, B. W., Nishibuchi, M., & Radu, S. (2018). In Vitro antimicrobial activity of green synthesized silver nanoparticles against selected Gram-negative foodborne pathogens. *Frontiers in Microbiology*, 9(JUL), 1–7. https://doi.org/10.3389/fmicb.2018.01555
- Lopez-Chaves, C., Soto-Alvaredo, J., Montes-Bayon, M., Bettmer, J., Llopis, J., & Sanchez-Gonzalez, C. (2018). Gold nanoparticles: Distribution, bioaccumulation and toxicity. In vitro and in vivo studies. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 14(1), 1–12. https://doi.org/10.1016/j.nano.2017.08.011
- Ma, Y., Yan, C., Alshameri, A., Qiu, X., Zhou, C., & Li, D. (2014). Synthesis and characterization of 13X zeolite from low-grade natural kaolin. *Advanced Powder Technology*, 25(2), 495–499. https://doi.org/10.1016/j.apt.2013.08.002
- Machado, G. E., Pereyra, A. M., Rosato, V. G., Moreno, M. S., & Basaldella, E. I. (2019). Improving the biocidal activity of outdoor coating formulations by using zeolite-supported silver nanoparticles. *Materials Science and Engineering C*, 98(257), 789–799. https://doi.org/10.1016/j.msec.2019.01.040
- Madejová, J. (2003). FTIR techniques in clay mineral studies. *Vibrational Spectroscopy*, *31*(1), 1–10. https://doi.org/10.1016/S0924-2031(02)00065-6
- Maeno, S., Niki, Y., Matsumoto, H., Morioka, H., Yatabe, T., Funayama, A., Toyama, Y., Taguchi, T., & Tanaka, J. (2005). The effect of calcium ion concentration on osteoblast viability, proliferation and differentiation in monolayer and 3D culture. *Biomaterials*, 26(23), 4847–4855. https://doi.org/10.1016/j.biomaterials.2005.01.006
- Mahmoodi, N. M., & Saffar-Dastgerdi, M. H. (2019). Zeolite nanoparticle as a superior adsorbent with high capacity: Synthesis, surface modification and

pollutant adsorption ability from wastewater. *Microchemical Journal*, *145*(August 2018), 74–83. https://doi.org/10.1016/j.microc.2018.10.018

- Majeed, N. S., & Majeed, J. T. (2017). Synthesis of Nanozeolite NaA from Pure Source Material Using Sol Gel Method. *Journal of Engineering*, 23(6), 53–62.
- Mattila, P. K., & Lappalainen, P. (2008). Filopodia: Molecular architecture and cellular functions. *Nature Reviews Molecular Cell Biology*, 9(6), 446–454. https://doi.org/10.1038/nrm2406
- Meenakshi, S., Devi, S., Pandian, K., Devendiran, R., & Selvaraj, M. (2016). Sunlight assisted synthesis of silver nanoparticles in zeolite matrix and study of its application on electrochemical detection of dopamine and uric acid in urine samples. *Materials Science and Engineering C*, 69, 85–94. https://doi.org/10.1016/j.msec.2016.06.037
- Mgbemere, H. E., Lawal, G. I., Ekpe, I. C., & Chaudhary, A. L. (2018). Synthesis of Zeolite-a Using Kaolin Samples From Darazo, Bauchi State and Ajebo, Ogun State in Nigeria. *Nigerian Journal of Technology*, 37(1), 87–95.
- Miao, K., Gao, J., Zhang, J., Dong, L., Jia, Q., Wang, R., Yu, L., Wang, S., Hong, M., & Yang, S. (2021). Kinetics guided synthesis and performance of monodisperse zeolite LTA microspheres. *Microporous and Mesoporous Materials*, 323(May), 111194. https://doi.org/10.1016/j.micromeso.2021.111194
- Miller, W. R., Munita, J. M., & Arias, C. A. (2014). Mechanisms of antibiotic resistance in enterococci. *Expert Review of Anti-Infective Therapy*, 12(10), 1221– 1236. https://doi.org/10.1586/14787210.2014.956092
- Mittal, A. K., Chisti, Y., & Banerjee, U. C. (2013). Synthesis of metallic nanoparticles using plant extracts. *Biotechnology Advances*, 31(2), 346–356. https://doi.org/10.1016/j.biotechadv.2013.01.003
- Moral-Munoz, J. A., Lucena-Antón, D., Perez-Cabezas, V., Carmona-Barrientos, I., González-Medina, G., & Ruiz-Molinero, C. (2018). Highly cited papers in Microbiology: identification and conceptual analysis. *FEMS Microbiology Letters*, 365(20), 1–9. https://doi.org/10.1093/femsle/fny230
- Muryanto, S. (2014). Effect of Orthosiphon aristatus leaves extract on the crystallization behavior of struvite (MgNH4PO4.6H2O).
- Musyoka, N. M., Petrik, L. F., Hums, E., Kuhnt, A., & Schwieger, W. (2015). Thermal stability studies of zeolites A and X synthesized from South African coal fly ash.
 Research on Chemical Intermediates, 41(2), 575–582.

https://doi.org/10.1007/s11164-013-1211-3

- Muthukrishnan, L., Ramakrishnan, P., Muthukrishnan, L., & Ramakrishnan, P. (2021). Pretreatment evaluation of gallic acid tuned nanosilver for short-term preservation of goat skins in leather processing. *J Nanopart Res*, 23, 143–144. https://doi.org/10.1007/s11051-021-05247-9
- Narayanan, K. B., & Sakthivel, N. (2011). Green synthesis of biogenic metal nanoparticles by terrestrial and aquatic phototrophic and heterotrophic eukaryotes and biocompatible agents. *Advances in Colloid and Interface Science*, 169(2), 59–79. https://doi.org/10.1016/j.cis.2011.08.004
- Naz, S., Khaskheli, A. R., Aljabour, A., Kara, H., Talpur, F. N., Sherazi, S. T. H., Khaskheli, A. A., & Jawaid, S. (2014). Synthesis of Highly Stable Cobalt Nanomaterial Using Gallic Acid and Its Application in Catalysis. *Advances in Chemistry*, 2014, 1–6. https://doi.org/10.1155/2014/686925
- Neidrauer, M., Ercan, U. K., Bhattacharyya, A., Samuels, J., Sedlak, J., Trikha, R., Barbee, K. A., Weingarten, M. S., & Joshi, S. G. (2014). Antimicrobial efficacy and wound-healing property of a topical ointment containing nitric-oxide-loaded zeolites. *Journal of Medical Microbiology*, 63(PART 2), 203–209. https://doi.org/10.1099/jmm.0.067322-0
- Nik, O. G., Chen, X. Y., & Kaliaguine, S. (2011). Amine-functionalized zeolite FAU/EMT-polyimide mixed matrix membranes for CO2/CH4 separation. *Journal of Membrane Science*, 379(1–2), 468–478. https://doi.org/10.1016/j.memsci.2011.06.019
- Ninan, N., Muthiah, M., Aliza, N., Yahaya, B., Park, I., Elain, A., Wui, T., Thomas, S., & Grohens, Y. (2014). Biointerfaces Antibacterial and wound healing analysis of gelatin / zeolite scaffolds. *Colloids and Surfaces B: Biointerfaces*, 115, 244– 252. https://doi.org/10.1016/j.colsurfb.2013.11.048
- Ninan, N., Muthiah, M., Park, I.-K., Elain, A., Wong, T. W., Thomas, S., & Grohens, Y. (2013). Faujasites incorporated tissue engineering scaffolds for wound healing: in vitro and in vivo analysis. ACS Applied Materials & Interfaces, 5(21), 11194–11206. https://doi.org/10.1021/am403436y
- Ninan, N., Muthiah, M., Park, I.-K., Wong, T. W., Thomas, S., & Grohens, Y. (2015).
 Natural Polymer/Inorganic Material Based Hybrid Scaffolds for Skin Wound Healing. *Polymer Reviews*, 55(3), 453–490. https://doi.org/10.1080/15583724.2015.1019135

- Olsen, I. (2015). Biofilm-specific antibiotic tolerance and resistance. European Journal of Clinical Microbiology & Infectious Diseases, 34(5), 877–886. https://doi.org/10.1007/s10096-015-2323-z
- Onodera, A., Nishiumi, F., Kakiguchi, K., Tanaka, A., Tanabe, N., Honma, A., Yayama, K., Yoshioka, Y., Nakahira, K., Yonemura, S., Yanagihara, I., Tsutsumi, Y., & Kawai, Y. (2015). Short-term changes in intracellular ROS localisation after the silver nanoparticles exposure depending on particle size. *Toxicology Reports*, 2, 574–579. https://doi.org/10.1016/j.toxrep.2015.03.004
- Osonga, F. J., Akgul, A., Yazgan, I., Akgul, A., Eshun, G. B., Sakhaee, L., & Sadik, O. A. (2020). Size and shape-dependent antimicrobial activities of silver and gold nanoparticles: A model study as potential fungicides. *Molecules*, 25(11), 2682. https://doi.org/10.3390/molecules25112682
- Pal, S., Tak, Y. K., & Song, J. M. (2015). Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticle? A study of the gramnegative bacterium Escherichia coli. *Journal of Biological Chemistry*, 290(42), 1712–1720. https://doi.org/10.1128/AEM.02218-06
- Paul Das, M., Rebecca Livingstone, J., Veluswamy, P., & Das, J. (2018). Exploration of Wedelia chinensis leaf-assisted silver nanoparticles for antioxidant, antibacterial and in vitro cytotoxic applications. *Journal of Food and Drug Analysis*, 26(2), 917–925. https://doi.org/10.1016/j.jfda.2017.07.014
- Pauzi, N., & Amira, N. (2018). The potential of gallic acid and ascorbic acid as green reducing agent in ZnO nanoparticle synthesis. *Malaysian Journal of Catalysis*, 3(August 2017), 13–16.
- Pauzi, N., Zain, N., & Amira, N. (2017). The potential of gallic acid and ascorbic acid as green reducing agent in ZnO nanoparticle synthesis. *Malaysian Journal of Catalysis*, 3, 13–16.
- Pereira, P. M., Ferreira, B. F., Oliveira, N. P., Nassar, E. J., Ciuffi, K. J., Vicente, M. A., Trujillano, R., Rives, V., Gil, A., Korili, S., & de Faria, E. H. (2018).
 Synthesis of Zeolite A from Metakaolin and Its Application in the Adsorption of Cationic Dyes. *Applied Sciences*, 8(4), 608. https://doi.org/10.3390/app8040608
- Perruchas, F., Consoli, D., & Barbieri, N. (2020). Specialisation, diversification and the ladder of green technology development. *Research Policy*, 49(3), 103922. https://doi.org/10.1016/j.respol.2020.103922
- Prabhu, S., & Poulose, E. K. (2012). Silver nanoparticles: mechanism of antimicrobial

action, synthesis, medical applications, and toxicity effects. *International Nano Letters*, 2(1), 32. https://doi.org/10.1186/2228-5326-2-32

- Purnomo, Setyarini, P. H., & Sulistyaningsih, D. (2018). Zeolite-based biomaterials for biomedical application: A review. AIP Conference Proceedings, 1977(030013), 1–7. https://doi.org/10.1063/1.5042933
- Qayyum, H., Ali, R., Rehman, Z. U., Ullah, S., Shafique, B., Dogar, A. H., Shah, A., & Qayyum, A. (2019). Synthesis of silver and gold nanoparticles by pulsed laser ablation for nanoparticle enhanced laser-induced breakdown spectroscopy. *Journal of Laser Applications*, 31(2), 022014. https://doi.org/10.2351/1.5086838
- Radwan, I. M., Gitipour, A., Potter, P. M., Dionysiou, D. D., & Al-Abed, S. R. (2019). Dissolution of silver nanoparticles in colloidal consumer products: effects of particle size and capping agent. *Journal of Nanoparticle Research*, 21(7). https://doi.org/10.1007/s11051-019-4597-z
- Radzig, M. A., Nadtochenko, V. A., Koksharova, O. A., Kiwi, J., Lipasova, V. A., & Khmel, I. A. (2013). Antibacterial effects of silver nanoparticles on gramnegative bacteria: Influence on the growth and biofilms formation, mechanisms of action. *Colloids and Surfaces B: Biointerfaces*, 102, 300–306. https://doi.org/10.1016/j.colsurfb.2012.07.039
- Rafique, M., Rafique, M. S., Kalsoom, U., Afzal, A., Butt, S. H., & Usman, A. (2019).
 Laser ablation synthesis of silver nanoparticles in water and dependence on laser nature. *Optical and Quantum Electronics*, 51(6), 1–11. https://doi.org/10.1007/s11082-019-1902-0
- Rajan, R., Chandran, K., Harper, S. L., Yun, S. II, & Kalaichelvan, P. T. (2015). Plant extract synthesized silver nanoparticles: An ongoing source of novel biocompatible materials. *Industrial Crops and Products*, 70, 356–373. https://doi.org/10.1016/j.indcrop.2015.03.015
- Ramachandran, R., Krishnaraj, C., Kumar, V. K. A., Harper, S. L., Kalaichelvan, T. P., & Yun, S. Il. (2018). In vivo toxicity evaluation of biologically synthesized silver nanoparticles and gold nanoparticles on adult zebrafish: a comparative study. *3 Biotech*, 8(10), 1–12. https://doi.org/10.1007/s13205-018-1457-y
- Ramos-Vivas, J., Chapartegui-González, I., Fernández-Martínez, M., González-Rico,
 C., Fortún, J., Escudero, R., Marco, F., Linares, L., Montejo, M., Aranzamendi,
 M., Muñoz, P., Valerio, M., Aguado, J. M., Resino, E., Ahufinger, I. G., Vega,
 A. P., Martínez-Martínez, L., Fariñas, M. C., Ruiz San Millán, J. C., ... The

ENTHERE Study Group, the G. for S. of I. in T. of the S. S. of I. D. and C. M. (GESITRA-S. and the S. N. for R. in I. D. (REIPI). (2019). Biofilm formation by multidrug resistant Enterobacteriaceae strains isolated from solid organ transplant recipients. *Scientific Reports*, *9*(1), 8928. https://doi.org/10.1038/s41598-019-45060-y

- Ribeiro, F., Van Gestel, C. A. M., Pavlaki, M. D., Azevedo, S., Soares, A. M. V. M., & Loureiro, S. (2017). Bioaccumulation of silver in Daphnia magna: Waterborne and dietary exposure to nanoparticles and dissolved silver. *Science of the Total Environment*, 574, 1633–1639. https://doi.org/10.1016/j.scitotenv.2016.08.204
- Robson, H. (2001). Verified Synthesis of Zeolitic Materials: Second Edition. Elsevier Science. https://books.google.com.my/books?id=nSNVDThj2DsC
- Rueden, C. T., Schindelin, J., Hiner, M. C., DeZonia, B. E., Walter, A. E., Arena, E. T., & Eliceiri, K. W. (2017). ImageJ2: ImageJ for the next generation of scientific image data. *BMC Bioinformatics*, 18(1), 1–26. https://doi.org/10.1186/s12859-017-1934-z
- Saheban, M., Bakhsheshi-Rad, H. R., Kasiri-Asgarani, M., Hamzah, E., Ismail, A. F., Aziz, M., & Dayaghi, E. (2019). Effect of zeolite on the corrosion behavior, biocompatibility and antibacterial activity of porous magnesium/zeolite composite scaffolds. *Materials Technology*, 34(5), 258–269. https://doi.org/10.1080/10667857.2018.1549803
- Saidan, N. H., Aisha, A. F. A., Hamil, M. S. R., Majid, A. M. S. A., & Ismail, Z. (2015). A novel reverse phase high-performance liquid chromatography method for standardization of Orthosiphon stamineus leaf extracts. *Pharmacognosy Research*, 7(1), 23–31. https://doi.org/10.4103/0974-8490.147195
- Saien, J., & Bahrami, M. (2016). Understanding the effect of different size silica nanoparticles and SDS surfactant mixtures on interfacial tension of n-hexane– water. *Journal of Molecular Liquids*, 224, 158–164. https://doi.org/10.1016/j.molliq.2016.09.112
- Salehi-Abari, M., Koupaei, N., & Hassanzadeh-Tabrizi, S. A. (2020). Synthesis and Characterisation of semi-interpenetrating network of Polycaprolactone/polyethylene glycol diacrylate/zeolite-CuO as wound dressing. *Materials Technology*, 35(5), 290–299. https://doi.org/10.1080/10667857.2019.1678088
- Samidurai, D., Pandurangan, A. K., Krishnamoorthi, S. K., Perumal, M. K., & Nanjian,

R. (2020). Sinensetin isolated from Orthosiphon aristatus inhibits cell proliferation and induces apoptosis in hepatocellular carcinoma cells. *Process Biochemistry*, 88, 213–221. https://doi.org/10.1016/j.procbio.2019.09.031

- Sang, S., Liu, Z., Tian, P., Liu, Z., Qu, L., & Zhang, Y. (2006). Synthesis of small crystals zeolite NaY. *Materials Letters*, 60, 1131–1133. https://doi.org/10.1016/j.matlet.2005.10.110
- Sani, N. S., Malek, N. A. N. N., Jemon, K., Kadir, M. R. A., & Hamdan, H. (2017). Effect of mass concentration on bioactivity and cell viability of calcined silica aerogel synthesized from rice husk ash as silica source. *Journal of Sol-Gel Science and Technology*, 82(1), 120–132. https://doi.org/10.1007/s10971-016-4266-y
- Schoch, C. L., Ciufo, S., Domrachev, M., Hotton, C. L., Kannan, S., Khovanskaya, R., Leipe, D., Mcveigh, R., O'Neill, K., Robbertse, B., Sharma, S., Soussov, V., Sullivan, J. P., Sun, L., Turner, S., & Karsch-Mizrachi, I. (2020). NCBI Taxonomy: a comprehensive update on curation, resources and tools. *Database : The Journal of Biological Databases and Curation*, 2020. https://doi.org/10.1093/database/baaa062
- Schüring, A., Auerbach, S., Fritzsche, S., & Haberlandt, R. (2002). On entropic barriers for diffusion in zeolites: A molecular dynamics study. *The Journal of Chemical Physics*, 116, 10890–10894. https://doi.org/10.1063/1.1480011
- Seliem, M. K., Mohamed, E. A., Selim, A. Q., Shahien, M. G., & Abukhadra, M. R. (2015). Synthesis of Na-a Zeolites From Natural and Thermally Activated Egyptian Kaolinite: Characterization and Competitive Adsorption of Copper Ions From Aqueous Solutions. *International Jornal of Bioassays*, 4(10), 4423–4430.
- Seo, Y. S., Ch, S. H., Cho, S., Yoon, H. R., Kang, Y. H., & Park, Y. (2015). Caffeic acid: Potential applications in nanotechnology as a green reducing agent for sustainable synthesis of gold nanoparticles. *Natural Product Communications*, 10(4), 627–630. https://doi.org/10.1177/1934578x1501000424
- Serati-Nouri, H., Jafari, A., Roshangar, L., Dadashpour, M., Pilehvar-Soltanahmadi,
 Y., & Zarghami, N. (2020). Biomedical applications of zeolite-based materials:
 A review. *Materials Science and Engineering C*, *116*(February), 111225.
 https://doi.org/10.1016/j.msec.2020.111225
- Shameli, K., Bin Ahmad, M., Zargar, M., Wan Yunus, W. M. Z., & Ibrahim, N. A. (2011). Fabrication of silver nanoparticles doped in the zeolite framework and

antibacterial activity. *International Journal of Nanomedicine*, 6, 331. https://doi.org/10.2147/ijn.s16964

- Sharma, V. K., Yngard, R. A., & Lin, Y. (2009). Silver nanoparticles: Green synthesis and their antimicrobial activities. *Advances in Colloid and Interface Science*, 145(1–2), 83–96. https://doi.org/10.1016/j.cis.2008.09.002
- Shenoy, N., Stenson, M., Lawson, J., Abeykoon, J., Patnaik, M., Wu, X., & Witzig, T. (2017). Drugs with anti-oxidant properties can interfere with cell viability measurements by assays that rely on the reducing property of viable cells. *Laboratory Investigation*, 97(5), 494–497. https://doi.org/10.1038/labinvest.2017.18
- Shi, X., Zou, J., Chen, X., Zheng, H., Jin, Z., Li, F., & Piao, J. G. (2020). The effect of size on the surface enhanced Raman scattering property of SiO2@PDA@AGNP core-shell-satellite nanocomposite. *Chemistry Letters*, 49(5), 534–537. https://doi.org/10.1246/cl.200040
- Siddique, M. H., Aslam, B., Imran, M., Ashraf, A., Nadeem, H., Hayat, S., Khurshid, M., Afzal, M., Malik, I. R., Shahzad, M., Qureshi, U., Khan, Z. U. H., & Muzammil, S. (2020). Effect of Silver Nanoparticles on Biofilm Formation and EPS Production of Multidrug-Resistant Klebsiella pneumoniae. *BioMed Research International*, 2020, 9. https://doi.org/10.1155/2020/6398165
- Son, J. S., Hwang, E. J., Kwon, L. S., Ahn, Y. G., Moon, B. K., Kim, J., Kim, D. H., Kim, S. G., & Lee, S. Y. (2021). Antibacterial activity of propolis-embedded zeolite nanocomposites for implant application. *Materials*, 14(5), 1–13. https://doi.org/10.3390/ma14051193
- Sondi, I., & Salopek-Sondi, B. (2004). Silver nanoparticles as antimicrobial agent: A case study on E. coli as a model for Gram-negative bacteria. *Journal of Colloid and Interface Science*, 275(1), 177–182. https://doi.org/10.1016/j.jcis.2004.02.012
- Subramanian, P., Ravichandran, A., Manoharan, V., Muthukaruppan, R., Somasundaram, S., Pandi, B., Krishnan, A., Marimuthu, P. N., Somasundaram, S. S. N., & You, S. G. (2019). Synthesis of Oldenlandia umbellata stabilized silver nanoparticles and their antioxidant effect, antibacterial activity, and biocompatibility using human lung fibroblast cell line WI-38. *Process Biochemistry*, 86(June), 196–204. https://doi.org/10.1016/j.procbio.2019.08.002

Sudarmono, Kim, S. Y., & Paik, J. H. (2020). Contradictory between morphology and

phylogenetic trees of Orthosiphon spp. (Lamiaceae) from Indonesia. *IOP Conference Series: Earth and Environmental Science*, 457(1). https://doi.org/10.1088/1755-1315/457/1/012030

Sun, Q., Cai, X., Li, J., Zheng, M., Chen, Z., & Yu, C.-P. P. (2014). Green synthesis of silver nanoparticles using tea leaf extract and evaluation of their stability and antibacterial activity. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 444, 226–231.

https://doi.org/https://doi.org/10.1016/j.colsurfa.2013.12.065

- Suryanarayana, C., & Norton, M. G. (2013). *X-Ray Diffraction: A Practical Approach*. Springer US. https://books.google.com.my/books?id=RRfrBwAAQBAJ
- Sytu, M. R. C., & Camacho, D. H. (2018). Green Synthesis of Silver Nanoparticles (AgNPs) from Lenzites betulina and the Potential Synergistic Effect of AgNP and Capping Biomolecules in Enhancing Antioxidant Activity. *BioNanoScience*, 8(3), 835–844. https://doi.org/10.1007/s12668-018-0548-x
- Tauanov, Z., & Inglezakis, V. J. (2019). Removal of iodide from water using silver nanoparticles-impregnated synthetic zeolites. *Science of the Total Environment*, 682, 259–270. https://doi.org/10.1016/j.scitotenv.2019.05.106
- Tazaki, H., Taguchi, D., Hayashida, T., & Nabeta, K. (2001). Stable isotope-labeling studies on the oxidative coupling of caffeic acid via o-quinone. In *Bioscience*, *Biotechnology and Biochemistry* (Vol. 65, Issue 12, pp. 2613–2621). https://doi.org/10.1271/bbb.65.2613
- Thi Lan Huong, V., & Nguyen, N. T. (2019). Green synthesis, characterization and antibacterial activity of silver nanoparticles using Sapindus mukorossi fruit pericarp extract. *Materials Today: Proceedings*, 42, 88–93. https://doi.org/10.1016/j.matpr.2020.10.015
- Thi Lan Huong, V., & Nguyen, N. T. (2021). Green synthesis, characterization and antibacterial activity of silver nanoparticles using Sapindus mukorossi fruit pericarp extract. *Materials Today: Proceedings*, 42, 88–93. https://doi.org/https://doi.org/10.1016/j.matpr.2020.10.015
- Tomic-Canic, M., Burgess, J. L., O'Neill, K. E., Strbo, N., & Pastar, I. (2020). Skin Microbiota and its Interplay with Wound Healing. *American Journal of Clinical Dermatology*, 21(s1), 36–43. https://doi.org/10.1007/s40257-020-00536-w
- Trusilewicz, L., Fernández-Martínez, F., Rahhal, V., & Talero, R. (2012). TEM and SAED characterization of metakaolin. Pozzolanic activity. *Journal of the*

American Ceramic Society, 95(9), 2989–2996. https://doi.org/10.1111/j.1551-2916.2012.05325.x

- Twentyman, P. R., & Luscombe, M. (1987). A study of some variables in a tetrazolium dye (MTT) based assay for cell growth and chemosensitivity. *British Journal of Cancer*, 56, 279–285.
- Tyler, G. (2001). ICP-OES, ICP-MS and AAS Techniques Compared. In *Technical note 05: ICP Optical Spectroscopy* (Issue 3).
- Ullah, H., Wilfred, C. D., & Shaharun, M. S. (2019). Green synthesis of copper nanoparticle using ionic liquid-based extraction from Polygonum minus and their applications. *Environmental Technology*, 40(28), 3705–3712. https://doi.org/10.1080/09593330.2018.1485751
- Van Dong, P., Ha, C. H., Binh, L. T., & Kasbohm, J. (2012). Chemical synthesis and antibacterial activity of novel-shaped silver nanoparticles. *International Nano Letters*, 2(1), 1–9. https://doi.org/10.1186/2228-5326-2-9
- Vargas, A., Angeli, M., Pastrello, C., Mcquaid, R., Li, H., Jurisicova, A., & Jurisica, I. (2016). Robust quantitative scratch assay. *Bioinformatics*, 32(9), 1439–1440. https://doi.org/10.1093/bioinformatics/btv746
- Vijayan, R., Joseph, S., & Mathew, B. (2018). Augmented antimicrobial, antioxidant and catalytic activities of green synthesised silver nanoparticles. *Materials Research Express*, 5(8), 85022. https://doi.org/10.1088/2053-1591/aaaf33
- Walton, K. S., & Snurr, R. Q. (2007). Applicability of the BET method for determining surface areas of microporous metal-organic frameworks. *Journal of the American Chemical Society*, 129(27), 8552–8556. https://doi.org/10.1021/ja071174k
- Wang, C., Leng, S., Guo, H., Yu, J., Li, W., Cao, L., & Huang, J. (2019). Quantitative arrangement of Si/Al ratio of natural zeolite using acid treatment. *Applied Surface Science*, 498(August), 143874. https://doi.org/10.1016/j.apsusc.2019.143874
- Wang, J. Q., Huang, Y. X., Pan, Y., & Mi, J. X. (2014). Hydrothermal synthesis of high purity zeolite A from natural kaolin without calcination. *Microporous and Mesoporous Materials*, 199, 50–56. https://doi.org/10.1016/j.micromeso.2014.08.002
- Wang, S., Li, R., Qing, Y., Wei, Y., Wang, Q., Zhang, T., Sun, C., Qin, Y., Li, D., & Yu, J. (2019). Antibacterial activity of Ag-incorporated zincosilicate zeolite scaffolds fabricated by additive manufacturing. *Inorganic Chemistry Communications*, 105(March), 31–35.

https://doi.org/10.1016/j.inoche.2019.04.026

- Wang, W., Feng, Q., Liu, K., Zhang, G., Liu, J., & Huang, Y. (2015). A novel magnetic 4A zeolite adsorbent synthesised from kaolinite type pyrite cinder (KTPC). *Solid State Sciences*, *39*, 52–58. https://doi.org/10.1016/j.solidstatesciences.2014.11.012
- Wang, X., Chang, J., & Wu, C. (2018). Bioactive inorganic / organic nanocomposites for wound healing. *Applied Materials Today*, 11, 308–319. https://doi.org/10.1016/j.apmt.2018.03.001
- Wang, Y. (2020). Measurements and Modeling of Water Adsorption Isotherms of Zeolite Linde-Type A Crystals. *Industrial and Engineering Chemistry Research*, 59(17), 8304–8314. https://doi.org/10.1021/acs.iecr.9b06891
- Wei, L., Lu, J., Xu, H., Patel, A., Chen, Z. S., & Chen, G. (2015). Silver nanoparticles: Synthesis, properties, and therapeutic applications. *Drug Discovery Today*, 20(5), 595–601. https://doi.org/10.1016/j.drudis.2014.11.014
- Williams, D. F. (2014). *There is no such thing as a biocompatible material*. *35*, 10009–10014.
- Wu, H., Tian, H., Li, J., Liu, L., Wang, Y., Qiu, J., Wang, S., & Liu, S. (2020). Self-detoxifying hollow zinc silica nanospheres with tunable Ag ion release-recapture capability: A nanoantibiotic for efficient MRSA inhibition. *Composites Part B: Engineering*, 202(September), 108415. https://doi.org/10.1016/j.compositesb.2020.108415
- Wu, J., Zheng, Y., Song, W., Luan, J., Wen, X., Wu, Z., Chen, X., Wang, Q., & Guo, S. (2014). In situ synthesis of silver-nanoparticles/bacterial cellulose composites for slow-released antimicrobial wound dressing. *Carbohydrate Polymers*, 102(1), 762–771. https://doi.org/10.1016/j.carbpol.2013.10.093
- Xiong, R., Lu, C., Wang, Y., Zhou, Z., & Zhang, X. (2013). Nanofibrillated cellulose as the support and reductant for the facile synthesis of Fe3O4/Ag nanocomposites with catalytic and antibacterial activity. *Journal of Materials Chemistry A*, 1(47), 14910–14918. https://doi.org/10.1039/c3ta13314a
- Xu, H., Qu, F., Xu, H., Lai, W., Wang, Y. A., Aguilar, Z. P., & Wei, H. (2012). Role of reactive oxygen species in the antibacterial mechanism of silver nanoparticles on Escherichia coli O157:H7. *BioMetals*, 25(1), 45–53. https://doi.org/10.1007/s10534-011-9482-x
- Xue, Z., Ma, J., Hao, W., Bai, X., Kang, Y., Liu, J., & Li, R. (2012). Synthesis and

characterization of ordered mesoporous zeolite LTA with high ion exchange ability. *Journal of Materials Chemistry*, 22(6), 2532–2538. https://doi.org/10.1039/c1jm14740d

- Yassue-Cordeiro, P. H., Zandonai, C. H., Genesi, B. P., Lopes, P. S., Sanchez-Lopez, E., Garcia, M. L., Fernandes-Machado, N. R. C., Severino, P., Souto, E. B., & da Silva, C. F. (2019). Development of chitosan/silver sulfadiazine/zeolite composite films for wound dressing. *Pharmaceutics*, 11(10), 535. https://doi.org/10.3390/pharmaceutics11100535
- Yee, M. S. L., Khiew, P. S., Tan, Y. F., Chiu, W. S., Kok, Y. Y., & Leong, C. O. (2015). Low temperature, rapid solution growth of antifouling silver-zeolite nanocomposite clusters. *Microporous and Mesoporous Materials*, 218, 69–78. https://doi.org/10.1016/j.micromeso.2015.07.004
- Yee, M. S. L., Khiew, P. S., Tan, Y. F., Kok, Y. Y., Cheong, K. W., Chiu, W. S., & Leong, C. O. (2014). Potent antifouling silver-polymer nanocomposite microspheres using ion-exchange resin as templating matrix. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 457(1), 382–391. https://doi.org/10.1016/j.colsurfa.2014.06.010
- Yin, L. H., Ran, B., Hu, T. J., Yang, C., Fei, J. J., & Li, Y. H. (2017). Preparation of highly efficient antibacterial polymeric films: Via the modulation of charge density and hydrophobicity. *RSC Advances*, 7(10), 6006–6012. https://doi.org/10.1039/c6ra26071c
- You, C., Li, Q., Wang, X., Wu, P., Ho, J. K., Jin, R., Zhang, L., Shao, H., & Han, C. (2017). Silver nanoparticle loaded collagen/chitosan scaffolds promote wound healing via regulating fibroblast migration and macrophage activation. *Scientific Reports*, 7(1), 1–11. https://doi.org/10.1038/s41598-017-10481-0
- Yu, O., Yu, E., & Shamova, O. V. (2018). Applied Clay Science Biological activity and sorption ability of synthetic montmorillonite modi fi ed by silver / lysozyme nanoparticles. *Applied Clay Science*, 163(July), 56–62. https://doi.org/10.1016/j.clay.2018.07.015
- Yunusov, K. E., Sarymsakov, A. A., Jalilov, J. Z. o`g`li, & Atakhanov, A. A. o`g`li. (2021). Physicochemical properties and antimicrobial activity of nanocomposite films based on carboxymethylcellulose and silver nanoparticles. *Polymers for Advanced Technologies*, 32(4), 1822–1830. https://doi.org/https://doi.org/10.1002/pat.5223

- Yusof, A. M., Nizam, N. A., & Rashid, N. A. A. (2010). Hydrothermal conversion of rice husk ash to faujasite-types and NaA-type of zeolites. *Journal of Porous Materials*, 17(1), 39–47. https://doi.org/10.1007/s10934-009-9262-y
- Zarrintaj, P., Mahmodi, G., Manouchehri, S., Mashhadzadeh, A. H., Khodadadi, M., Servatan, M., Ganjali, M. R., Azambre, B., Kim, S., Ramsey, J. D., Habibzadeh, S., Saeb, M. R., & Mozafari, M. (2020). Zeolite in tissue engineering: Opportunities and challenges. *MedComm*, 1(1), 5–34. https://doi.org/10.1002/mco2.5
- Zhang, S., Ye, J., Liu, X., Wang, Y., Li, C., Fang, J., Chang, B., Qi, Y., Li, Y., & Ning,
 G. (2021). Titanium carbide/zeolite imidazole framework-8/polylactic acid electrospun membrane for near-infrared regulated photothermal/photodynamic therapy of drug-resistant bacterial infections. *Journal of Colloid and Interface Science*, 599, 390–403. https://doi.org/10.1016/j.jcis.2021.04.109
- Zhang, Yu, Shao, D., Yan, J., Jia, X., Li, Y., Yu, P., & Zhang, T. (2016). The pore size distribution and its relationship with shale gas capacity in organic-rich mudstone of Wufeng-Longmaxi Formations, Sichuan Basin, China. *Journal of Natural Gas Geoscience*, 1(3), 213–220. https://doi.org/10.1016/j.jnggs.2016.08.002
- Zhang, Yuan, Leng, Z., Zou, F., Wang, L., Chen, S. S., & Tsang, D. C. W. (2018). Synthesis of zeolite A using sewage sludge ash for application in warm mix asphalt. *Journal of Cleaner Production*, 172, 686–695. https://doi.org/10.1016/J.JCLEPRO.2017.10.005
- Zheng, K., Setyawati, M. I., Leong, D. T., & Xie, J. (2018). Antimicrobial silver nanomaterials. *Coordination Chemistry Reviews*, 357, 1–17. https://doi.org/10.1016/j.ccr.2017.11.019
- Zhou, B., Xiao, J. F., Tuli, L., & Ressom, H. W. (2012). LC-MS-based metabolomics. *Molecular BioSystems*, 8(2), 470–481. https://doi.org/10.1039/c1mb05350g
- Zou, X., Li, P., Lou, J., Fu, X., & Zhang, H. (2017). Stability of single dispersed silver nanoparticles in natural and synthetic freshwaters: Effects of dissolved oxygen. *Environmental Pollution*, 230, 674–682. https://doi.org/https://doi.org/10.1016/j.envpol.2017.07.007

LIST OF PUBLICATIONS

Journal with Impact Factor

- Asraf, M. H., Malek, N. A. N. N., Jemon, K., Sani, N. S., Muhammad, M. S. (2019). Cytotoxicity and Wound Healing Assessments of Amine-Functionalized Zeolite Y, *Particuology*, 45, 116 123. https://doi.org/10.1016/j.partic.2018.09.006. (Q2, IF: 3.067)
- Asraf, M. H., Sani, N. S., Williams, C. D., Jemon, K., Malek, N. A. N. N. (2021). In Situ Biosynthesized Silver Nanoparticle-Incorporated Synthesized Zeolite A Using Orthosiphon Aristatus Extract for In Vitro Antibacterial Wound Healing, *Particuology*, In Press, https://doi.org/10.1016/j.partic.2021.09.007. (Q2, IF: 3.067)
- Isah, M., Asraf, M. H., Malek, N. A. N. N., Jemon, K., Sani, N. S., Muhammad, M. S., Wahab, M. F. A., Saidin, M. A. R. (2020). Preparation and Characterization of Chlorhexidine Modified Zinc-Kaolinite and Its Antibacterial Activity Against Bacteria Isolated from Water Vending Machine, *Journal of Environmental Chemical Engineering*, 8, 103545. https://doi.org/10.1016/j.jece.2019.103545 (Q1, IF: 4.3)

Indexed Conference Proceedings

 Asraf, M. H., Malek, N. A. N. N. (2020). Effect of Different HDTMA Loading on Silver Modified Kaolinite on Its Antibacterial Activity, *AIP Conference Proceeding*, 2231, 020003. https://doi.org/10.1063/5.0002423. (Indexed in WOS & Scopus)