SYNERGISTIC CYTOTOXIC EFFECT OF THYMOQUINONE IN COMBINATION WITH CISPLATIN AGAINST MDA-MB-231 TRIPLE NEGATIVE CELL LINE

ZURFARAHANIM BINTI ZOLKEMRI

A thesis submitted in fulfilment of the requirements for the award of the degree of Master of Philosophy

> Faculty of Science Universiti Teknologi Malaysia

> > SEPTEMBER 2020

DEDICATION

This thesis is dedicated to my parents and siblings

ACKNOWLEDGEMENT

This thesis becomes a reality with many help of individuals. I would like to extend my sincere thanks to all of them.

Foremost, I wish to express my greatest gratitude to my respected advisor, Associate Professor Dr. Salehhuddin for helping me transformed a simple research question into an original project. Your evaluation and feedback made every step of this research valuable. My sincere thanks also goes to my co-supervisor, Dr. Khairudnadwa for her advice and knowledge she had imparted in this research.

Besides, I would like to thank my senior, Sayang Baba for being a great mentor. Your passion in sharing knowledge played a large role in shaping me into who I am today. I also want to thank Puan Syuhada for providing me with all the necessary facilities in Animal Tissue Culture Lab. Thank you to Department of Bioscience, Faculty of Science, and UTM for giving me opportunity to do this study.

Most important, thank you to my beloved father and mother, dearest Zolkemri and Fatimah for their love and supports in all of my endeavors over the years. I can always count on their encouragements and helps. I also want to extend my love, appreciation, and thanks to my siblings, who never questioned my abilities and always having confidence in my life decisions.

Not to forget, to my closest friend, Puteri Arien, Yonca, and Aqilah Zainal. I could never said enough words of thanks. I'm so grateful for your presence. Thank you so much for being there for me and for giving me advice and nice words whenever I need it the most.

Last but not least, thank you to anyone that were involved with me directly or indirectly in order for me to finish this thesis. I wish everyone to have a good life ahead! Sincerely.

ABSTRACT

The purpose of this study is to investigate the application of genetic algorithm (GA) in modelling linear and non-linear dynamic systems and develop an alternative model structure selection algorithm based on GA. Orthogonal least square (OLS), a gradient descent method was used as the benchmark for the proposed algorithm. A model structure selection based on modified genetic algorithm (MGA) has been proposed in this study to reduce problems of premature convergence in simple GA (SGA). The effect of different combinations of MGA operators on the performance of the developed model was studied and the effectiveness and shortcomings of MGA were highlighted. Results were compared between SGA, MGA and benchmark OLS method. It was discovered that with similar number of dynamic terms, in most cases, MGA performs better than SGA in terms of exploring potential solution and outperformed the OLS algorithm in terms of selected number of terms and predictive accuracy. In addition, the use of local search with MGA for fine-tuning the algorithm was also proposed and investigated, named as memetic algorithm (MA). Simulation results demonstrated that in most cases, MA is able to produce an adequate and parsimonious model that can satisfy the model validation tests with significant advantages over OLS, SGA and MGA methods, Furthermore, the case studies on identification of multivariable systems based on real experiment t al data from two systems namely a turbo alternator and a continuous stirred tank reactor showed that the proposed algorithm could be used as an alternative to adequately identify adequate and parsimonious models for those systems. Abstract must be bilingual. For a thesis written in Bahasa Melayu, the abstract must first be written in Bahasa Melayu and followed by the English translation. If the thesis is written in English, the abstract must be written in English and followed by the translation in Bahasa Melayu. The abstract should be brief, written in one paragraph and not exceed one (1) page. An abstract is different from synopsis or summary of a thesis. It should states the field of study, problem definition, methodology adopted, research process, results obtained and conclusion of the research. The abstract can be written using single or one and a half spacing. Example can be seen in Appendix 1 (Bahasa Melayu) and Appendix J (English).

ABSTRAK

Kajian ini dilakukan bertujuan mengkaji penggunaan algoritma genetik (GA) dalam pemodelan sistem dinamik linear dan tak linear dan membangunkan kaedah alternatif bagi pcmilihan struktur model menggunakan GA. Algorithma kuasa dua terkecil ortogon (OLS), satu kaedah penurunan kecerunan digunakan sebagai bandingan bagi kaedah yang dicadangkan. Pcmilihan struktur model mengunakan kaedah algoritma genetik yang diubahsuai (MGA) dicadangkan dalam kajian ini bagi mengurangkan masalah konvergens pramatang dalam algoritma genetik mudah (SGA). Kesan penggunaan gabungan operator MGA yang berbeza ke atas prestasi model yang terbentuk dikaji dan keberkesanan serta kekurangan MGA diu t arakan. Kajian simulasi dilakukan untuk membanding SGA, MGA dan OLS. Dengan meggunakan bilangan parameter dinamik yang setara kajian ini mendapati, dalam kebanyakan kes, prestasi MGA adalah lebih baik daripada SGA dalam mencari penyelesaian yang berpotensi dan lebih berkebolehan daripada OLS dalam menentukan bilangan sebutan yang dipilih dan ketepatan ramalan. Di samping itu, penggunaan carian tempatan dalam MGA untuk menambah baik algorithma tersebut dicadang dan dikaji, dinamai sebagai algoritma mcmetic (MA). Hasil simulasi menunjukkan, dalam kebanyakan kes, MA berkeupayaan menghasilkan model yang bersesuaian dan parsimoni dan mcmenuhi ujian pengsahihan model di samping mcmperolehi beberapa kelebihan dibandingkan dengan kaedah OLS, SGA dan MGA. Tambahan pula, kajian kes untuk sistcm berbilang pcmbolehubah menggunakan data eksperimental sebenar daripada dua sistem iaitu sistem pengulang-alik turbo dan reaktor teraduk berterusan menunjukkan algoritma ini boleh digunakan sebagai alternatif untuk mcmperolehi model termudah yang memadai bagi sistem tersebut.

TABLE OF CONTENTS

TITLE

DEC	iii	
DED	ICATION	iv
ACK	NOWLEDGEMENT	v
ABS	TRACT	vi
ABS	TRAK	vii
TAB	LE OF CONTENTS	viii
LIST	xii	
LIST	xiii	
LIST	TOF ABBREVIATIONS	xix
LIST	T OF SYMBOLS	XX
LIST	TOF APPENDICES	xxi
CHAPTER 1	INTRODUCTION	1
1.1	Research background	1
1.2	Problem statement	4
1.3	Research objectives	5
1.4	Research scopes	5
1.5	Research significance	6
CHAPTER 2	LITERATURE REVIEW	7
	2.1.1 Breast cancer statistic	8
2.2	Triple Negative Breast Cancer	9
	2.2.1 MDA-MB-231 TNBC cell line	10
2.3	Current treatment in TNBC	11
	2.3.1 Chemotherapy	11
2.4	Cisplatin	12
	2.4.1 Cytotoxic action of Cisplatin	14

2.4.2 Limitations of Cisplatin 15

2.5	Phytochemicals	16
	2.5.1 Thymoquinone	17
	2.5.1.1 Cytotoxic action of Thymoquinone	18
2.6	Drug combination	21
	2.6.1 Combination of Thymoquinone with Chemotherapeutic drug	23
	2.6.2 Combination of Thymoquinone with Cisplatin	25
2.7	Mechanism of cell-death	28
	2.7.1 Morphological and biochemical changes in apoptosis	28
	2.7.2 Apoptosis signaling pathways	32
	2.7.2.1 Extrinsic apoptosis signaling	32
	2.7.2.2 Intrinsic apoptosis signaling	34
	2.7.2.3 Roles of p53 and its family as transcription factor	35
CHAPTER 3	MATERIALS AND METHODS	39
3.1	General Workflow of Research Methodology	39
3.2	Materials	40
3.3	Cell lines	40
3.4	Cell culture	40
	3.4.1 Cell thawing	41
	3.4.2 Cell subculture	41
	3.4.3 Single cell suspension	42
3.5	Drugs preparation	42
3.6	Phase contrast microscopy	43
3.7	MTT assay	43
	3.7.1 Single drug treatment	44
	3.7.2 Drug combination treatment	45
3.8		
	Combination Index (CI) method	46
3.9	Combination Index (CI) method DAPI staining	46 47
3.9 3.10		

	3.12	Quant	itative Real-Time Polymerase Chain Reaction	50
		3.12.1	Primer design	50
		3.12.2	RNA extraction	52
		3.12.3	Synthesis of cDNA	53
		3.12.4	QPCR Assay	54
		3.12.5	Agarose gel electrophoresis	56
	3.13	Statist	ical analysis	57
СНАРТЕ	R 4	RESU	JLTS AND DISCUSSION	59
	4.1	Introd	uction	59
	4.2	Morpl	nological changes induced by TQ and CP	60
		4.2.1	Morphological changes in MDA-MB-231 TNBC cells	60
		4.2.2	Morphological changes in WRL-68 normal liver cells	62
	4.3	Cytote	oxicity of TQ, CP, and their combination	64
		4.3.1	Cytotoxicity of TQ and CP in MDA-MB-231 TNBC cell line	64
		4.3.2	Cytotoxicity of TQ and CP in WRL-68 normal liver cells	68
		4.3.3	Cytotoxicity of combination of TQ and CP in MDA-MB-231 cells and WRL-68 cells	72
		4.3.4	IC_{50} value comparison between TQ, CP, and their combination in both MDA-MB-231 cells and WRL-68 cells	74
	4.4	Syner	gistic effect of TQ and CP	75
	4.5	Apopt	osis induction	79
		4.5.1	Morphological changes in cell nuclei	79
		4.5.2	Cellular apoptosis induction	82
		4.5.3	Caspases activation	84
	4.6	Apopt	osis signaling pathways	86
		4.6.1	Activation of p53 and its family gene	87
		4.6.2	Cell cycle arrest	90
		4.6.3	Extrinsic apoptosis signaling pathway	91
		4.6.4	Intrinsic apoptosis signaling pathway	94

	4.6.5 Summary of apoptosis induction	98
4.7	Possible apoptosis mechanism	102
CHAPTER 5	CONCLUSION AND FUTURE WORK	105
5.1	Conclusion	105
5.2	Future Work	106
REFERENCES		107
APPENDICES		143

LIST OF PUBLICATION	103

LIST OF TABLES

TABLE NO.	TITLE	PAGE
Table 2.1	General nomenclature of Nigella sativa L. black cumin (Adapted from PLANTS database, 2019).	17
Table 2.2	Effect of thymoquinone (TQ) against cancer cells <i>in vitro</i> (\uparrow : upregulated, \downarrow : downregulated).	20
Table 2.3	The effect of combination of thymoquinone with cisplatin against cancer cell lines (\uparrow : upregulated, \downarrow : downregulated).	27
Table 3.1	List of primers sequences.	51
Table 3.2	The cDNA reaction components setup.	54
Table 3.3	The standard reverse transcription protocol.	54
Table 3.4	The qPCR reaction components setup.	55
Table 3.5	The qRT-PCR reaction standard protocol.	55
Table 3.6	Experimental design	56
Table 3.7	The asterisks scheme.	58
Table 4.1	The IC ₅₀ values of TQ, CP, and their combination (TQ+CP) in MDA-MB-231 TNBC cells and WRL-68 normal liver cells. The n/t and n/i in the table indicated 'not tested' and 'no 50% inhibition', respectively. The different letters represents the significant difference at p<0.05 between the treatment groups at the respective treatment duration based on One-way ANOVA followed by post-hoc Tukey's test.	74
Table 4.2	CompuSyn results of combination of TQ and CP against MDA-MB-231 cells.	77
Table 4.3	The fold-change in expression of target genes in TQ, CP, and TQ+CP treated MDA-MB-231 cells. The light pink shading indicated the positive regulation in specific treatment group. The asterisk represents the level of significant difference with the control cells at $p<0.05$ (*), $p<0.01$ (**), $p<0.001$ (***), and $p<0.0001$ (****) based on the paired t-test with a two-tailed P value.	99

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
Figure 2.1	The progression of normal cells to cancerous cells in breast ducts showing the ability to invade through the vessels and spread to distant sites (Mork, 2012).	7
Figure 2.2	The demographic changes of breast cancer deaths in females at all ages from 2018 to 2040 (Global Cancer Observatory, 2018, http://gco.larc.fr/).	8
Figure 2.3	MDA-MB 231 breast cancer cell line (ATCC, 2018, https://www.atcc.org/products/all/HTB-26.aspx).	10
Figure 2.4	Chemical structure of cisplatin (Dasari &Tchounwou, 2014)	13
Figure 2.5	Black Cumin seeds and chemical structure of thymoquinone (Mistry, 2016).	18
Figure 2.6	(A) The Unified Theory. (B) The CompuSyn program algorithm. (Chou, 2010).	22
Figure 3.1	General workflow of research methodology used in this study.	39
Figure 4.1	Phase contrast microscopy images of MDA-MB-231 TNBC cells at 100x magnification after being treated with TQ and CP at 10 μ M and 100 μ M for 24, 48, and 72 hours. Scale bar represents 200 μ m. The blue, red, and green arrows on the images shows the normal spindle-shaped cells, cell shrinkage, and cells rounding respectively.	61
Figure 4.2	Phase contrast microscopy images of MDA-MB-231 TNBC cells at 100x magnification after being treated with TQ and CP at 10 μ M and 100 μ M for 24, 48, and 72 hours. Scale bar represents 200 μ m. The blue, red, and yellow arrows on the images shows the normal polygonal-shaped cells, cell shrinkage, and membrane blebbing respectively.	63
Figure 4.3	The cytotoxicity effect of TQ (0-100 μ M) against MDA-MB-231 cells at (A) 24h, (B) 48h and (C) 72h of treatment. Each column represents mean \pm SD for three technical replicates from three independent experiments (n=3). The asterisk represents the level of significant difference with the control cells at p<0.05	

(*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value.

- Figure 4.4 The cytotoxicity effect of CP (0-100 μ M) against MDA-MB-231 cells at (A) 24h, (B) 48h and (C) 72h of treatment. Each column represents mean \pm SD for three technical replicates from three independent experiments (n=3). The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value.
- Figure 4.5 The cytotoxicity effect of TQ (0-100 μ M) against WRL-68 cells at (A) 24h, (B) 48h and (C) 72h of treatment. Each column represents mean \pm SD for three technical replicates from three independent experiments (n=3). The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value.
- Figure 4.6 The cytotoxicity effect of CP (0-100 μ M) against WRL-68 cells at (A) 24h, (B) 48h and (C) 72h of treatment. Each column represents mean \pm SD for three technical replicates from three independent experiments (n=3). The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value.
- Figure 4.7 The cytotoxicity effect of TQ (10.90 μ M) in combination with CP (10.39 μ M) against (A) MDA-MB-231 TNBC cells and (B) WRL-68 normal liver cells at 72h of treatment. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value.
- Figure 4.8 CompuSyn graphical results. (A) Dose-effect curves.(B) Median-effect plots. Fa : Fraction affected and Fu: Fraction unaffected.
- Figure 4.9 The Fa-CI plot generated from CompuSyn software.
- Figure 4.10 Classic isobologram at Fa = 0.5 (IC₅₀), Fa = 0.75 (IC₇₅), and Fa = 0.9 (IC₉₀) of TQ+CP generated by CompuSyn software.
- Figure 4.11 Morphological changes in cell nuclei. (A) Representative zoomed in monochrome images of DAPI staining of MDA-MB-231 cells showing normal

70

67

66

71

73

76

78

nuclei (a, b, and c), condensed nuclei (d, e, and f), and fragmented nuclei (g, h, and i) after being treated with TQ, CP, and their combination for 72 h. The images were taken at 200x magnification using fluorescence inverted microscope. Scale bar at 10 μ m. (B) The percentage of apoptotic cell nuclei analyzed by ImageJ. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.

- Figure 4.12 The percentage of cell death induced by TQ, CP, and TQ+CP in MDA-MB-231 cells after 72h which include apoptotic and necrotic cells. Each column represents mean \pm S.D of three technical replicates from three independent experiments (n=3). The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.13 The fold-change of caspase-8, -9, and -3/7 activity induced by TQ, CP, and TQ+CP in MDA-MB-231 cells after 72h. Each column represents mean \pm S.D of three technical replicates from three independent experiments (n=3). The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.14 Fold-change of p53 mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.

81

- Figure 4.15 Fold-change of p63 mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.16 Fold-change of p73 mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.17 Fold-change of p21 mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.18 Fold-change of TNF mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.19 Fold-change of Fas mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column

91

89

90

represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (***), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.

- Figure 4.20 Fold-change of PUMA mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (***), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.21 Fold-change of NOXA mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.22 Fold-change of Bax mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01 (**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.23 Fold-change of Bak mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01

95

95

(**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p<0.05 between the treatment groups based on One-way ANOVA and Tukey's test.

- Figure 4.24 Fold-change of BCL2 mRNA expression level in MDA-MB-231 cells induced by TQ, CP, and TQ+CP. The untreated cells were used as control. Each column represents mean \pm S.D of three technical replicates. The asterisk represents the level of significant difference with the control cells at p<0.05 (*), p<0.01(**), p<0.001 (***), and p<0.0001 (****) based on the paired t-test with a two-tailed P value. The different letters represents the significant difference at p < 0.05between the treatment groups based on One-way ANOVA and Tukey's test.
- Figure 4.25 The PCR products of housekeeping gene, cell-cycle, and apoptosis-related genes visualized on agarose gel electrophoresis, showing single band and correct size of amplicon for specific gene. 101 Figure 4.26 The proposed apoptosis mechanism of the synergistic
- cytotoxic effect of TQ and CP in MDA-MB-231 cells. 103

97

LIST OF ABBREVIATIONS

ATCC	-	Americab Tissue Culture Collection
СР	-	Cisplatin
CI	-	Combination Index
DMEM	-	Dulbecco's Modified Eagle Medium
DMSO	-	Dimethylsulfoxide
DRI	-	Dose Reduction Index
ER	-	Estrogen Receptor
FBS	-	Fetal Bovine Serum
HER2	-	Human Epidermal Growth Factor Receptor 2
IC ₅₀	-	Half maximal Inhibitory Concentration
MTT	-	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium
		bromide
PBS	-	Phosphate Buffer Solution
PR	-	Progesterone Receptor
TQ	-	Thymoquinone
TQ+CP	-	Thymoquinone in combination with cisplatin
TNBC	-	Triple Negative Breast Cancer

LIST OF SYMBOLS

μm-MicrometreμL-MicrolitermL-Millilitrexg-Centrifugal force	μM	-	Micro Molar
mL - Millilitre	μm	-	Micrometre
	μL	-	Microliter
xg - Centrifugal force	mL	-	Millilitre
	xg	-	Centrifugal force

LIST OF APPENDICES

APPENDIX	TITLE	PAGE
Appendix A	Non-linear regression analysis of dose-effect curves of TQ and CP alone against MDA-MB-231 cells.	143
Appendix B	Non-linear regression analysis of dose-effect curves of CP alone against WRL-68 cells.	144
Appendix C	Non-linear regression analysis of dose-effect curves of TQ+CP against MDA-MB-231 cells	145
Appendix D	CompuSyn Report of TQ in combination with CP against MDA-MB-231 cells.	146

CHAPTER 1

INTRODUCTION

1.1 Research background

Breast cancer is the most common cancer and the first cause of death in women worldwide. It was estimated that 2.1 million new cases and 627 thousand of breast cancer related deaths occurred in 2018 (Bray *et al.*, 2018). In Malaysia, breast cancer incidence is on the increase and at present has become the first leading cause of cancer deaths among women in all ethnics. According to the Malaysia Cancer Registry, there were 17009 new cases and 7372 deaths due to breast cancer reported from 2007 to 2011, followed up to 2016. By ethnicity, Malays (57.9%) recorded the lowest 5-year survival rate, followed by Indians (70.5%), and Chinese (76.5%) (National Cancer Registry *et al.*, 2018). One of the common risk factor associated with the differences in survival rate between ethnics in Malaysia was the identification of distinct subtypes of breast cancer (Devi *et al.*, 2012). Approximately 15-20% of breast cancer cases accounts for triple negative breast cancer (Frank *et al.*, 2013).

Triple negative breast cancer (TNBC) is characterized by the absence of estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2) (Anders *et al.*, 2013). Due to the absence of these three biomarkers, the established treatments for breast cancer including endocrine and HER2-targeted therapy are unfortunately not effective for TNBC treatment (Isakoff, 2010). Besides, TNBC appears to be more aggressive, invasive, higher in grade, and highly metastatic as compared to other subtypes (Carey *et al.*, 2010). Given the lack of therapeutic options, cytotoxic chemotherapy remains the standard treatment for TNBC patients (Peddi *et al.*, 2012). However, only 50% of TNBC patients respond to primary cytotoxic agents and most cases are associated with early relapse within 3 years of diagnosis and higher risk of mortality less than 12 months (Dent *et al.*, 2007;

Sørlie *et al.*, 2001). Therefore, studies are now focusing on other cytotoxic agents that are not routinely used for breast cancer such as platinum-based drugs in order to find the most active agent for TNBC treatment. Indeed, numerous clinical studies have shown promising results of using platinum-based drugs for treatment of TNBC in adjuvant, neoadjuvant, and metastatic settings (Sparano *et al.*, 2008; Liedtke *et al.*, 2008; Byrski *et al.*, 2012).

One of the platinum-based drugs that has been actively studied for TNBC treatment is cisplatin (CP) (Hill *et al.*, 2019). Generally, CP exerts its cytotoxic properties by binding to N7-sites of purine residues on the nucleotides to form interand intra-strand DNA adducts and block the RNA transcription and DNA replication, signalling DNA damage in a cell (Basu & Krishnamurthy, 2010). If not repaired, DNA-damage can induce cell-cycle arrest and ultimately cause apoptosis (Tanida *et al.*, 2012). Apoptosis is a highly regulated natural process of cell death and in cancer, the loss of apoptosis has become a promising treatment strategy for decades (Pfeffer & Singh, 2018). Despite being a potent anticancer drug, the clinical use of CP is limited due to associated toxicity and side effects in patients including nausea and vomiting, hepatotoxicity, nephrotoxicity, and cardiotoxicity (Huang *et al.*, 2017; Pratibha *et al.*, 2006; Hayati *et al.*, 2015; Hu *et al.*, 2018). These CP-related toxicities are dose-dependent, thus an overdose of this drug might cause significant deaths in patients (Tsang *et al.*, 2009). Hence, another effective strategy to reduce CP-related toxicities for better cancer treatment is urgently needed.

There is a growing body of data that suggested drugs combination can minimize the toxicity effect towards normal cells while still providing therapeutic effect, such as cancer cell death (He *et al.*, 2019; Abe *et al.*, 2019; Monroe *et al.*, 2019). The drugs that are used in combination may interact in many unexpected ways and exhibit distinct effects for instance, synergism, additive, and antagonism (Lehar *et al.*, 2009). Among these interactions, achieving drug synergy is a highly pursued goal in anticancer drug combination studies (Yin *et al.*, 2014). Hypothetically, drug synergy could be achieved by using drugs that complement each other in terms of anticancer properties but with non-overlapping toxicity profiles. Recent experimental efforts are focusing on identifying synergistic drug pairs by using conventional cytotoxic drug with phytochemicals since natural compound has a safer toxicity profile compared to synthetic drugs (Ndreshkjana *et al.*, 2019; Tan *et al.*, 2019). In addition, phytochemicals have been proved to show wide range of biological activity including anti-inflammatory, anti-oxidant, and anti-cancer properties (Yimer *et al.*, 2019; Ozdemir *et al.*, 2018; Singh *et al.*, 2016). The combination of selected phytochemical with conventional chemotherapeutic may provide synergistic effect at a lower dose which can results to higher anticancer effect and reduced toxicity towards normal cells (Pezzani *et al.*, 2019).

Thymoquinone (TQ) is a phytochemical derived from Nigella sativa and has been reported to exert numerous pharmacological activities such as such as antiinflammatory, antimicrobial, antidiabetic, and also anticancer (Yazdi et al., 2018; Chaieb et al., 2011; Younus et al., 2018; Asaduzzaman et al., 2017). In the previous years, emerging evidence has shown anticancer activity of TQ in many types of cancer, for instance, bladder, cervical, colon, gastric, glioblastoma, and breast cancer (Iskender et al., 2016; Reindl et al., 2008; Kundu et al., 2014; Zhu et al., 2016; Gurung et al., 2010; Rajput et al., 2013). TQ exhibited anticancer response on cancer cell lines through many cellular mechanisms such as anti-proliferation, cell-cycle arrest, and apoptosis induction (ElKhoely et al., 2015; Bhattacharya et al., 2015; Woo et al., 2013). Particularly in TNBC, TQ inhibited growth of both MDA-MB-468 and MDA-MB-231 cell lines by arresting G1 phase of the cell cycle and induced apoptosis through caspase-dependent and independent pathways (Sutton et al., 2014). Moreover, TQ has been shown to protect normal cells from oxidative damage and prevents toxic side effects in many types of cells such as prostate epithelial cells, intestinal cells, lung fibroblasts, and breast epithelial cells (Kaseb et al., 2017; El-Najjar et al., 2010; Gurung et al., 2010; Kabil et al., 2018).

Previously, TQ has been used in combination with CP against cancer cell lines derived from colon, esophageal, oral squamous, gastric, ovarian, and also breast cancer (Zhang *et al.*, 2016; Hu *et al.*, 2018; Alaufi *et al.*, 2017; Ma *et al.*, 2017; Nessa *et al.*, 2011). Latest study by Liu *et al.* (2017) revealed that the combination of TQ and CP able to enhance the cytotoxicity against ovarian cancer cells (SK-OV-3) by upregulating BAX and downregulating BCL2 gene which contributed to apoptotic cell death (Liu *et al.*, 2017). Moreover, Sutton *et al.* (2014) has reported that TQ able to enhance the cytotoxic effect of CP against MDA-MB-468 TNBC cells, highlighting the potential of this drugs combination for TNBC treatment. However, no further evidence on the synergism of TQ and CP against any TNBC cells is discussed in the literature. Taken the evidence that TQ in combination with CP may enhance the cytotoxic effect against cancer cells, this study was aimed to investigate the synergistic cytotoxic effect of combining TQ with CP against MDA-MB-231 TNBC cells.

1.2 Problem statement

Cisplatin (CP) is cytotoxic, thus it targets all rapidly dividing cells including cancerous and normal cells. The use of cisplatin in the cancer treatment is limited with its severe toxicities in normal cells such as dose-dependent hepatotoxicity and nephrotoxicity (Pratibha et al., 2006; Sheth et al., 2017). One of the strategy to reduce the toxic effect of CP is by combining CP with non-toxic phytochemical such as thymoquinone (TQ) to achieve higher therapeutic index at lower doses, avoiding the dose-limiting effect of CP against normal cells (Jafri et al., 2010). The protective effect of TQ against cisplatin-induced toxicity have been well documented in many studies (Ulu et al., 2012; Al-Malki & Sayed, 2014). From these findings, combining TQ with CP may serves as an effective way to reduce the dose-limiting effect of CP. Previously, the combination of TQ with CP have been found to enhance the cytotoxic effects and induced apoptosis in several cancer including colon, esophageal, gastric, oral squamous, and ovarian cancer (Zhang et al., 2016; Hu et al., 2018; Ma et al., 2017; Alaufi et al., 2017; Nessa et al., 2011). Taken these findings into consideration, the present study was carried out to investigate the possible synergistic cytotoxic effect of combining TQ and CP against MDA-MB-231 TNBC cell line.

1.3 Research objectives

This project aims to investigate the possible synergistic cytotoxic effect of thymoquinone (TQ) in combination with cisplatin (CP) against MDA-MB-231 triple negative breast cancer (TNBC) cell line and the underlying mechanism of cell death induction. The specific objectives of this research were:

- (a) To evaluate the effect of TQ, CP, and their combination on the cell viability of MDA-MB-231 TNBC cells and WRL-68 liver cells using MTT colorimetric assay.
- (b) To determine the synergistic cytotoxic effect of TQ in combination with CP using Combination Index (CI) method by Chou-Talalay.
- (c) To determine the apoptotic effect of TQ, CP, and their synergistic combination on the cell nuclei morphology and cellular apoptosis/necrosis using fluorescent DAPI staining and Annexin-V FITC/PI staining assay respectively.
- (d) To evaluate the activity of caspase-8 and caspase-9 that are respectively responsible for initiation of extrinsic and intrinsic apoptosis and executioner caspase-3/7 using Caspase-Glo assay.
- (e) To evaluate the mRNA expression of target genes associated with transcription factor (p53, p63, and p73), cell cycle arrest (p21), extrinsic apoptosis (TNF and Fas), and intrinsic apoptosis signaling (PUMA, NOXA, Bak, Bax, and BCL2) using quantitative Real-Time Polymerase Chain Reaction (qRT-PCR).

1.4 Research scopes

In order to achieve the objectives of this study, the effect of TQ, CP, and their combination on the cell viability of MDA-MB-231 TNBC cells and WRL-68 liver cells were first evaluated using MTT assay. The synergistic combination dose of TQ and CP was determined using Chou-Talalay Combination Index (CI) method. The CI method utilized CompuSyn software for automated simulation of the CI values

and dose reduction index (DRI) together with generation of other graphs including dose-effect curves, median-effect plot, and isobologram. The cytotoxic effect of TQ, CP, and their synergistic combination on the cell nuclei morphology and cellular apoptosis/necrosis were determined using blue-fluorescent DAPI staining and Annexin-V FITC/PI staining assay. The photograph of DAPI stained cell nuclei was collected using Eclipse Ti Series inverted microscope equipped with Nikon's NIS elements imaging software (Nikon Instruments Inc., Tokyo, Japan). Whereas, the relative fluorescence unit (RFU) of Annexin V FITC/PI stained cells were measured using GloMax®-Multi Detection System (Promega Corp., Madison, Wisconsin, United States). Next, the apoptotic effect of the treatments on the activity of initiator caspases (caspase-8 and -9) and executioner caspases (caspase-3/7) were evaluated using luminescent Caspase-Glo assays. The results were recorded in the form of relative luminescent unit (RLU) using GloMax®-Multi Detection System (Promega Corp., Madison, Wisconsin, United States). The caspase-8 and -9 used in this study distinguished the involvement of the extrinsic and intrinsic apoptosis induced by the treatment. Further evaluation on the mRNA expression of the downstream proteins involved in the transcription factors, cell cycle, extrinsic, and intrinsic signalling pathways was performed using qRT-PCR. The markers used in this study were p53, p63, and p73 for transcription factor, p21 for cell cycle, TNF and Fas for extrinsic, and PUMA, NOXA, Bax, Bak, and BCL2 for intrinsic signalling. The qRT-PCR was performed using BioRad CFX96 Real Time PCR System (Bio-Rad Laboratories, Inc., Hercules, California, United States). The fold gene expression of the sample was calculated using the delta-delta CT ($2-\Delta\Delta$ Ct) method.

1.5 Research significance

The study of the cytotoxicity of TQ and CP can provide more therapeutic potential for cancer treatment. The synergistic combination dose of TQ with CP can possibly solve the toxicity issue of CP towards normal cells. Their synergistic interactions can provide more understanding about signalling pathways such as apoptosis induction in the treatment of cancer.

REFERENCES

- Abe, K., Yamamoto, N., Hayashi, K., Takeuchi, A., & Tsuchiya, H. (2019). Caffeine citrate enhanced cisplatin antitumor effects in osteosarcoma and fibrosarcoma in vitro and in vivo. *BMC cancer*, 19(1), 689.
- Aldossary S. A. (2019). Review on Pharmacology of Cisplatin: Clinical Use, Toxicity and Mechanism of Resistance of Cisplatin. Biomed Pharmacol J, 12(1).
- Alam, A., Farooq, U., Singh, R., Dubey, V.P., Kumar, S., Kumari, R. Naik, K.K., Tripathi, B.D., and Dhar, K.L. (2018). Chemotherapy Treatment and Strategy Schemes: A Review. *Open Acc J of Toxicol*, 2(5), OAJT.MS.ID.555600.
- Alaufi, O.M., Noorwali, A., Zahran, F., Al-Abd, A.M., and Al-Attas, S. (2017). Cytotoxicity of thymoquinone alone or in combination with cisplatin (CDDP) against oral squamous cell carcinoma in vitro. *Scientific Reports*, 7(1), 13131.
- Alhosin, M., Abusnina, A., Achour, M., Sharif, T., Muller, C., Peluso, J., Chataigneau, T., Lugnier, C., Schini-Kerth, V. B., Bronner, C., & Fuhrmann, G. (2010). Induction of apoptosis by thymoquinone in lymphoblastic leukemia Jurkat cells is mediated by a p73-dependent pathway which targets the epigenetic integrator UHRF1. *Biochemical pharmacology*, 79(9), 1251– 1260.
- Alhosin, M. (2020). Thymoquinone is a novel potential inhibitor of SIRT1 in cancers with p53 mutation: Role in the reactivation of tumor suppressor p73. *World Academy of Sciences Journal*, 2, 8.
- Abusnina, A., Alhosin, M., Keravis, T., Muller, C. D., Fuhrmann, G., Bronner, C., & Lugnier, C. (2011). Down-regulation of cyclic nucleotide phosphodiesterase PDE1A is the key event of p73 and UHRF1 deregulation in thymoquinone-induced acute lymphoblastic leukemia cell apoptosis. *Cellular signalling*, 23(1), 152–160.
- Al-Shabanah, O., Badary, O., Nagi, M., Al-Gharably, N., Al-Rikabi, A., and Al-Bekairi, A. (1998). Thymoquinone protects against doxorubicin-induced cardiotoxicity without compromising its antitumor activity. J. Exp. Clin. Cancer Res, 17, 193–198.

- Al-Malki, A. L., and Sayed, A. A. R. (2014). Thymoquinone attenuates cisplatininduced hepatotoxicity via nuclear factor kappa- β. BMC Complementary and Alternative Medicine, 14, 282.
- Alobaedi, O. H., Talib, W. H., & Basheti, I. A. (2017). Antitumor effect of thymoquinone combined with resveratrol on mice transplanted with breast cancer. *Asian Pacific journal of tropical medicine*, 10(4), 400–408.
- American Type Culture Collection (2018). MDA-MB-231 (ATCC® HTB-26[™]). Available at: <u>https://www.atcc.org/Products/All/HTB-26.aspx</u> (Accessed: 5 February 2020).
- Anders, C. K., Zagar, T. M., & Carey, L. A. (2013). The management of early-stage and metastatic triple-negative breast cancer: a review. *Hematology/oncology clinics of North America*, 27(4), 737–viii.
- Anders, C., & Carey, L. A. (2008). Understanding and treating triple-negative breast cancer. *Oncology (Williston Park, N.Y.)*, 22(11), 1233–1243.
- Anloo, K.H.G., Noordin, M.I., Kamalidehghan, B., Javar, H.A., Widodo, R.T., Majidzadeh, K., & Raisian, K. (2017). Involvement of NF-κB and HSP70 signaling pathways in the apoptosis of MDA-MB-231 cells induced by thymoquinone: An in vitro study. *Life Science Journal*, 14(5): 61-70.
- Arafa, S.A., Zhu, Q., Shah, Z.I., Wani, G., Barakat, B.M., Racoma, I., El-Mahdy, M.A., & Wani, A.A. (2011). Thymoquinone up-regulates PTEN expression and induces apoptosis in doxorubicin-resistant human breast cancer cells. *Mutat Res*, 706, 28–35.
- Ashkenazi, A., Pai, R. C., Fong, S., Leung, S., Lawrence, D. A., Marsters, S. A., & Schwall, R. H. (1999). Safety and antitumor activity of recombinant soluble Apo2 ligand. *The Journal of clinical investigation*, 104(2), 155–162.
- Ashkenazi, A. (2008). Targeting the extrinsic apoptosis pathway in cancer. *Cytokine Growth Factor Rev*, 19:325–331.
- Aslantürk, O. (2018). In Vitro Cytotoxicity and Cell Viability Assays: Principles, Advantages, and Disadvantages, in Larramendy, M.L. and Soloneski, S. Genotoxicity. IntechOpen, Chapter 1.
- Badr, G., Lefevre, E. A., & Mohany, M. (2011). Thymoquinone inhibits the CXCL12-induced chemotaxis of multiple myeloma cells and increases their susceptibility to Fas-mediated apoptosis. *PloS one*, 6(9), e23741.

- Bai, T., Yang, Y., Wu, Y.-L., Jiang, S., Lee, J. J., Lian, L.-H., et al. (2014). Thymoquinone alleviates thioacetamide-induced hepatic fibrosis and inflammation by activating LKB1–AMPK signaling pathway in mice. *Int. Immunopharmacol*, 19, 351–357.
- Balint, E., Phillips, A. C., Kozlov, S., Stewart, C. L., & Vousden, K. H. (2002). Induction of p57(KIP2) expression by p73beta. *Proceedings of the National Academy of Sciences of the United States of America*, 99(6), 3529–3534.
- Bansal, T., Jaggi, M., Khar, R. K., & Talegaonkar, S. (2009). Emerging significance of flavonoids as P-glycoprotein inhibitors in cancer chemotherapy. Journal of pharmacy & pharmaceutical sciences: a publication of the Canadian Society for Pharmaceutical Sciences, *Societe canadienne des sciences pharmaceutiques*, 12(1), 46–78.
- Bas, A., Forsberg, G., Hammarström, S., & Hammarström, M. L. (2004). Utility of the housekeeping genes 18S rRNA, beta-actin and glyceraldehyde-3phosphate-dehydrogenase for normalization in real-time quantitative reverse transcriptase-polymerase chain reaction analysis of gene expression in human T lymphocytes. *Scandinavian journal of immunology*, 59(6), 566–573.
- Basu, A., & Krishnamurthy, S. (2010). Cellular responses to Cisplatin-induced DNA damage. *Journal of nucleic acids*, 201367.
- Bayat Mokhtari, R., Homayouni, T. S., Baluch, N., Morgatskaya, E., Kumar, S., Das,
 B., & Yeger, H. (2017). Combination therapy in combating cancer. *Oncotarget*, 8(23), 38022-38043.
- Bhattacharya, S., Ahir, M., Patra, P., Mukherjee, S., Ghosh, S., Mazumdar, M., Chattopadhyay, S., Das, T., Chattopadhyay, D., & Adhikary, A. (2015).
 PEGylated-thymoquinone-nanoparticle mediated retardation of breast cancer cell migration by deregulation of cytoskeletal actin polymerization through miR-34a. *Biomaterials*, 51, 91–107.
- Bhattacharyya, A., Ear, U.S., & Koller, B.H. (2000). The breast cancer susceptibility gene BRCA1 is required for subnuclear assembly of Rad51 and survival following treatment with the DNA cross-linking agent cisplatin, *J Biol Chem* , 275(31): 23899-23903.
- Bhagya, N., Prabhu, A., Rekha, P.D., & Chandrashekar, K.R. Combination of tetrandrine and cisplatin synergises cytotoxicity and apoptosis in triple negative breast cancer. *Synergy*. 10, 100063.

- Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R.L., Torre, L.A., & Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: A Cancer Journal for Clinicians, 68: 394-424.
- Brenner, D., Blaser, H., & Mak, T. W. (2015). Regulation of tumour necrosis factor signalling: live or let die. *Nature reviews. Immunology*, 15(6), 362–374.
- Bucur, O., Nat, R., Cretoiu, D., & Popescu, L. M. (2001). Phagocytosis of apoptotic cells by microglia in vitro. *Journal of cellular and molecular medicine*, 5(4), 438–441.
- Budihardjo, I., Oliver, H., Lutter, M., Luo, X., & Wang, X. (1999). Biochemical pathways of caspase activation during apoptosis. *Annu Rev Cell Dev Biol*, 15:269–290.
- Bulut N, Kilickap S, Sari E, & Altundag K. (2008). Response to taxanes in triple negative breast cancer. *Cancer Chemother Pharmacol*, 63(1):189.
- Byrski T., Gronwald J , & Huzarski T. (2010). Pathologic complete response rates in young women with BRCA1-positive breast cancers after neoadjuvant chemotherapy. *J Clin Oncol.* 28:375–379.
- Byrski T., Dent R., Blecharz P., Foszczynska-Kloda M., Gronwald J., Huzarski T., Cybulski C., Marczyk E., Chrzan R., Eisen A., Lubinski J., & Narod S.A. (2012). Results of a phase II open-label, non-randomized trial of cisplatin chemotherapy in patients with BRCA1-positive metastatic breast cancer. *Breast Cancer Res*, 14(4):R110.
- Byrski, T., Huzarski, T., & Dent, R. (2014). Pathologic complete response to neoadjuvant cisplatin in BRCA1-positive breast cancer patients. *Breast Cancer Res Treat*, 147, 401–405.
- Cabal-Hierro, L., & Lazo, P. S. (2012). Signal transduction by tumor necrosis factor receptors. *Cellular signalling*, 24(6), 1297–1305.
- Caccuri, F., Sommariva, M., Marsico, S., Giordano, F., Zani, A., Giacomini, A., Fraefel, C., Balsari, A., & Caruso, A. (2019). Inhibition of DNA Repair Mechanisms and Induction of Apoptosis in Triple Negative Breast Cancer Cells Expressing the Human Herpesvirus 6 U94. *Cancers*, 11(7), 1006.
- Cailleau, R,. Olivé, M., & Cruciger, Q.V. (1978). Long-term human breast carcinoma cell lines of metastatic origin: preliminary characterization. *In Vitro*, 14(11):911-5.

- Carey, L., Winer, E., Viale, G., Cameron, D., & Gianni, L. (2010). Triple-negative breast cancer: disease entity or title of convenience? *Nat Rev Clin Oncol*, 7(12):683–92.
- Carneiro, B. A., & El-Deiry, W. S. (2020). Targeting apoptosis in cancer therapy. Nature reviews. *Clinical oncology*, 17(7), 395–417.
- Corda Y., Job C., Anin M.-F., Leng M. and Job D. (1991) Transcription by eucaryotic and procaryotic RNA polymerases of DNA modified at a d(GG) or a d(AG) site by the antitumor drug cis-diammindichloroplatinum(II). *Biochemistry*, 30: 222–230 29
- Costanzo, A., Pediconi, N., Narcisi, A., Guerrieri, F., Belloni, L., Fausti, F., Botti, E., & Levrero, M. (2014). TP63 and TP73 in cancer, an unresolved "family" puzzle of complexity, redundancy and hierarchy. *FEBS letters*, 588(16), 2590–2599.
- Chavez, K.J., Garimella, S.V., & Lipkowitz, S. (2010). Triple Negative Breast Cancer Cell Lines: One Tool in the Search for Better Treatment of Triple Negative Breast Cancer. *Breast Dis*, 32(1-2):35–48.
- Chen, D., Milacic, Vesna, Frezza, Michael, & Dou, Q. P. (2009). Metal Complexes, their Cellular Targets and Potential for Cancer Therapy. *Current Pharmaceutical Design*, 15(7), 777-791.
- Choi, K. H., Choi, H. Y., Ko, J. K., Park, S. S., Kim, Y. N., & Kim, C. W. (2004). Transcriptional regulation of TNF family receptors and Bcl-2 family by chemotherapeutic agents in murine CT26 cells. *Journal of cellular biochemistry*, 91(2), 410–422.
- Chou, T. C., Tan, Q. H., & Sirotnak, F. M. (1993). Quantitation of the synergistic interaction of edatrexate and cisplatin in vitro. *Cancer Chemotherapy*. *Pharmacol.* 31, 259–264.
- Chou, T.C. & Talalay, P. (1983). Analysis of combined drug effects: a new look at a very old problem. *Trends in Pharmacological Sciences*, 4, 450-454.
- Chou, T.C. & Talalay, P. (1984). Quantitative analysis of dose-effect relationships: the combined effects of multiple drugs or enzyme inhibitors. Advances in Enzyme Regulation, 22, 27-55.
- Chou, T.C. (2010). Drug Combination Studies and Their Synergy Quantification Using the Chou-Talalay Method. *Cancer Res*, 70(2), 440-446.

- Chaieb, K., Kouidhi, B., Jrah, H., Mahdouni, K., & Bakhrouf, A. (2011). Antibacterial activity of Thymoquinone, an active principle of Nigella sativa and its potency to prevent bacterial biofilm formation. *BMC Complement Altern Med*, 11, 29.
- Chavez, K.J., Garimella, S.V., & Lipkowitz, S. (2010). Triple Negative Breast Cancer Cell Lines: One Tool in the Search for Better Treatment of Triple Negative Breast Cancer. *Breast Dis*, 32(1-2):35–48.
- Chu, S. C., Hsieh, Y. S., Yu, C. C., Lai, Y. Y., & Chen, P. N. (2014). Thymoquinone induces cell death in human squamous carcinoma cells via caspase activationdependent apoptosis and LC3-II activation-dependent autophagy. *PloS one*, 9(7), e101579.
- Cui, L., Zhou, F., Chen, C., & Wang, C.C. (2019). Overexpression of CCDC69 activates p14ARF/MDM2/p53 pathway and confers cisplatin sensitivity. J Ovarian Res 12, 4.
- Czarnomysy, R., Surażyński, A., Popławska, B., Rysiak, E., Pawłowska, N., Czajkowska, A., Bielawski, K., & Bielawska, A. (2017). Synergistic action of cisplatin and echistatin in MDA-MB-231 breast cancer cells. *Molecular and cellular biochemistry*, 427(1-2), 13–22.
- Czarnomysy, R., Surażyński, A., Muszynska, A., Gornowicz, A., Bielawska, A., & Bielawski, K. (2018). A novel series of pyrazole-platinum(II) complexes as potential anti-cancer agents that induce cell cycle arrest and apoptosis in breast cancer cells. *Journal of enzyme inhibition and medicinal chemistry*, 33(1), 1006–1023.
- D'Arcy M. S. (2019). Cell death: a review of the major forms of apoptosis, necrosis and autophagy. *Cell biology international*, 43(6), 582–592.
- Da–lian, D., Ping, W., Haiyan, J., Coling, D., & Salvi, R. (2009). Gene expression in cisplatin ototoxicity and protection with p53 inhibitor. *Journal of Otology*, 4 (2), 61-70.
- Dai, X., Li, T., Bai, Z., Yang, Y., Liu, X., Zhan, J., & Shi, B. (2015). Breast cancer intrinsic subtype classification, clinical use and future trends. Am J Cancer Res, 5(10), 2929–2943.
- Daniel, P.S., Andrea, L.R., Aron, C.E., Zhigang, C.W., Zoltan, S., Qiyuan, L., Nicolai, J., Chee-Onn, L., Diana, C., Ayodele, B., Aquila, F., Rebecca, S.G., Paula, D.R., Nadine, M.T., Arcangela, D.N., Shridar G., Alexander M.,

Christian C., Dennis C.S., Leif W.E., Eric P.W., & Judy, E.G. (2010). Efficacy of Neoadjuvant Cisplatin in Triple-Negative Breast Cancer. *Journal of Clinical Oncology*, 28(7): 1145-1153.

- Das, S., Dey, K. K., Dey, G., Pal, I., Majumder, A., MaitiChoudhury, S., kundu, S. C., & Mandal, M. (2012). Antineoplastic and apoptotic potential of traditional medicines thymoquinone and diosgenin in squamous cell carcinoma. *PloS one*, 7(10), e46641.
- Dasari, S. & Tchounwou, P.B. (2014). Cisplatin in cancer therapy: Molecular mechanisms of action. *European Journal of Pharmacology*, 740, 364-378.
- Dastjerdi, M. N., Mehdiabady, E. M., Iranpour, F. G., & Bahramian, H. (2016). Effect of Thymoquinone on p53 Gene Expression and Consequence Apoptosis in Breast Cancer Cell Line. *International journal of preventive medicine*, 7, 66.
- Darakhshan, S., Bidmeshki, Pour, A., Hosseinzadeh, Colagar, A., & Sisakhtnezhad, S. (2015). Thymoquinone and its therapeutic potentials. *Pharmacol Res*, 95-96, 138-58.
- De Cola, A., Bongiorno-Borbone, L., Bianchi, E., Barcaroli, D., Carletti, E., Knight, R. A., Di Ilio, C., Melino, G., Sette, C., & De Laurenzi, V. (2012). FLASH is essential during early embryogenesis and cooperates with p73 to regulate histone gene transcription. *Oncogene*, 31(5), 573–582.
- Dehne, N., Lautermann, J., Petrat, F., Rauen, U., & de Groot, H. (2001). Cisplatin Ototoxicity: Involvement of Iron and Enhanced Formation of Superoxide Anion Radicals. *Toxicol Appl Pharmacol*, 174: 27-34.
- Delmastro, D.A., Li, J., Vaisman, A., Solle, M., & Chaney, S.G. (1997) DNA damage inducible-gene expression following platinum treatment in human ovarian carcinoma cell lines. *Cancer Chemother Pharmacol*, 39: 245–253.
- Demain, A. L., & Vaishnav, P. (2011). Natural products for cancer chemotherapy. *Microbial biotechnology*, 4(6), 687-99.
- Demchenko A. P. (2013). Beyond annexin V: fluorescence response of cellular membranes to apoptosis. *Cytotechnology*, 65(2), 157–172. https://doi.org/10.1007/s10616-012-9481-y
- Dent, R., Hanna, W.M., & Trudeau, M. (2009). Pattern of metastatic spread in triplenegative breast cancer. *Breast Cancer Res Treat*,115: 423-8.

- Devi, C. B., Tang, T. S. and Corbex, M. (2012), Incidence and risk factors for breast cancer subtypes in three distinct South-East Asian ethnic groups: Chinese, Malay and natives of Sarawak, Malaysia. *Int. J. Cancer*, 131: 2869-2877.
- Dirican, A., Atmaca, H., Bozkurt, E., Erten, C., Karaca, B., & Uslu, R. (2015). Novel combination of docetaxel and thymoquinone induces synergistic cytotoxicity and apoptosis in DU-145 human prostate cancer cells by modulating PI3K-AKT pathway. *Clinical & translational oncology : official publication of the Federation of Spanish Oncology Societies and of the National Cancer Institute of Mexico*, 17(2), 145–151.
- Dirican, A., Erten, C., Atmaca, H., Bozkurt, E., Kucukzeybek, Y., Varol, U., Oktay Tarhan, M., Karaca, B., & Uslu, R. (2014). Enhanced cytotoxicity and apoptosis by thymoquinone in combination with zoledronic acid in hormoneand drug-resistant prostate cancer cell lines. *Journal of B.U.ON. : official journal of the Balkan Union of Oncology*, 19(4), 1055–1061.
- di Pietro, A., Koster, R., Boersma-van Eck, W., Dam, W. A., Mulder, N. H., Gietema, J. A., de Vries, E. G., & de Jong, S. (2012). Pro- and anti-apoptotic effects of p53 in cisplatin-treated human testicular cancer are cell context-dependent. *Cell cycle (Georgetown, Tex.)*, 11(24), 4552–4562. https://doi.org/10.4161/cc.22803
- Effenberger-Neidnicht, K., & Schobert, R. (2011). Combinatorial effects of thymoquinone on the anti-cancer activity of doxorubicin. *Cancer chemotherapy and pharmacology*, 67(4), 867–874.
- El-Mahdy, M. A., Zhu, Q., Wang, Q., Wani, G. & Wani, A. A. (2005). Thymoquinone induces apoptosis through activation of caspase-8 and mitochondrial events in p53-null myeloblastic leukemia HL-60 cells. *Int. J. Cancer*, 117: 409-417.
- Eljack, N.D., Ma, H.Y.M., Drucker, J., Shen, C., Hambley, T.W., New, E.J., Clarke, R.J. (2014). Mechanism of cell uptake and toxicity of the anticancer drug cisplatin. *Metallomics*, 6(11), 2126-2133.
- Elmore, S. A., Dixon, D., Hailey, J. R., Harada, T., Herbert, R. A., Maronpot, R. R., Nolte, T., Rehg, J. E., Rittinghausen, S., Rosol, T. J., Satoh, H., Vidal, J. D., Willard-Mack, C. L., & Creasy, D. M. (2016). Recommendations from the INHAND Apoptosis/Necrosis Working Group. *Toxicologic pathology*, 44(2), 173–188.

- Elmore S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic pathology*, 35(4), 495–516. https://doi.org/10.1080/01926230701320337
- Estandarte, A. K., Botchway, S., Lynch, C., Yusuf, M., & Robinson, I. (2016). The use of DAPI fluorescence lifetime imaging for investigating chromatin condensation in human chromosomes. *Scientific reports*, 6, 31417.
- Fuertes, M.A., Alonso, C., & Pérez, J.M. (2003). Biochemical modulation of Cisplatin mechanisms of action: enhancement of antitumor activity and circumvention of drug resistance. *Chem Rev.* 103:645–662.
- Foucquier, J., & Guedj, M. (2015). Analysis of drug combinations: current methodological landscape. *Pharmacology research & perspectives*, 3(3), e00149.
- Fulda, S., Galluzzi, L., & Kroemer, G. (2010). Targeting mitochondria for cancer therapy. Nat. Rev. *Drug Discov*, 9, 447–464.
- Fulda, S., & Debatin, K.M. (2006). Extrinsic versus intrinsic apoptosis pathways in anticancer chemotherapy. *Oncogene*, 25, 4798–4811.
- Ferri, K.F., & Kroemer, G. (2001). Organelle-specific initiation of cell death pathways. *Nat. Cell Biol.*, 3, E255–E263.
- Fink, S. L., & Cookson, B. T. (2005). Apoptosis, pyroptosis, and necrosis: mechanistic description of dead and dying eukaryotic cells. *Infection and immunity*, 73(4), 1907–1916.
- Fischer, M. (2017). Census and evaluation of p53 target genes. *Oncogene*, 36, 3943–3956.
- Frank, G.A., Danilova, N.V., Andreeva, L.L., & Nefedova, N.A. (2013). WHO classification of tumors of the breast, 2012. *Arkh Patol*, 75(2), 53-63.
- Fan S, Chang JK, Smith ML, Duba D, Fornace Jr AJ and O'Connor PM . (1997). Cells lacking CIP1/WAF1 genes exhibit preferential sensitivity to cisplatin and nitrogen mustard. *Oncogene*, 14, 2127–2136.
- Fan S, Smith ML, Rivet DJ, Duba D, Zhan Q, Kohn KW, Fornace Jr AJ & O'Connor PM . (1995). Disruption of p53 function sensitizes breast cancer MCF-7 cells to cisplatin and pentoxifylline. *Cancer Res.*, 55, 1649–1654.
- Galanski, M., Jakupec, M.A., & Keppler, B.K. (2005). Update of the preclinical situation of anticancer platinum complexes: Novel design strategies and innovative analytical approaches. *Curr Med Chem*, 12:2075–2094.

- Gali-Muhtasib, H., Diab-Assaf, M., Boltze, C., Al-Hmaira, J., Hartig, R., Roessner, A., & Schneider-Stock, R. (2004). Thymoquinone extracted from black seed triggers apoptotic cell death in human colorectal cancer cells via a p53dependent mechanism. *International Journal of Oncology*, 25, 857-866.
- Gali-Muhtasib, H., Kuester, D., Mawrin, C., Bajbouj, K., Diestel, A., Ocker, M., Habold, C., Foltzer-Jourdainne, C., Schoenfeld, P., Peters, B., Diab-Assaf, M., Pommrich, U., Itani, W., Lippert, H., Roessner, A., & Schneider-Stock, R. (2008). Thymoquinone triggers inactivation of the stress response pathway sensor CHEK1 and contributes to apoptosis in colorectal cancer cells. *Cancer research*, 68(14), 5609–5618.
- Gamarra-Luques, C. D., Goyeneche, A. A., Hapon, M. B., & Telleria, C. M. (2012). Mifepristone prevents repopulation of ovarian cancer cells escaping cisplatinpaclitaxel therapy. *BMC cancer*, 12, 200. https://doi.org/10.1186/1471-2407-12-200
- Ganji-Harsini, S., Khazaei, M., Rashidi, Z., & Ghanbari, A. (2016). Thymoquinone Could Increase The Efficacy of Tamoxifen Induced Apoptosis in Human Breast Cancer Cells: An In Vitro Study. *Cell journal*, 18(2), 245–254.
- Geck, R. C., Foley, J. R., Murray Stewart, T., Asara, J. M., Casero, R. A., Jr, & Toker, A. (2020). Inhibition of the polyamine synthesis enzyme ornithine decarboxylase sensitizes triple-negative breast cancer cells to cytotoxic chemotherapy. *The Journal of biological chemistry*, 295(19), 6263–6277.
- Gonçalves, H., Guerra, M. R., Duarte Cintra, J. R., Fayer, V. A., Brum, I. V., & Bustamante Teixeira, M. T. (2018). Survival Study of Triple-Negative and Non-Triple-Negative Breast Cancer in a Brazilian Cohort. *Clinical Medicine Insights. Oncology*, 12, 1179554918790563.
- Gong, C., Qian, L., Yang, H., Ji, L. L., Wei, H., Zhou, W. B., Qi, C., & Wang, C. H. (2015). Hepatotoxicity and pharmacokinetics of cisplatin in combination therapy with a traditional Chinese medicine compound of Zengmian Yiliu granules in ICR mice and SKOV-3-bearing nude mice. *BMC complementary and alternative medicine*, 15, 283.
- Ghosh, S. (2019). Cisplatin: The first metal based anticancer drug. *Bioorganic Chemistry*, 88, 102925.
- Goyal, S.N., Prajapati, C.P., Gore, P.R., Patil, C.R., Mahajan, U.B., Sharma, C., Talla, S.P., & Ojha, S.K. (2017). Therapeutic Potential and Pharmaceutical

Development of Thymoquinone: A Multitargeted Molecule of Natural Origin. *Front. Pharmacol*, 8:656.

- Granada, A. E., Jiménez, A., Stewart-Ornstein, J., Blüthgen, N., Reber, S., Jambhekar, A., & Lahav, G. (2020). The effects of proliferation status and cell cycle phase on the responses of single cells to chemotherapy. *Molecular biology of the cell*, 31(8), 845–857.
- Greenwell, M., & Rahman, P. K. (2015). Medicinal Plants: Their Use in Anticancer Treatment. *International journal of pharmaceutical sciences and research*, 6(10), 4103-4112.
- Gressner, O., Schilling, T., Lorenz, K., Schulze Schleithoff, E., Koch, A., Schulze-Bergkamen, H., Lena, A. M., Candi, E., Terrinoni, A., Catani, M. V., Oren, M., Melino, G., Krammer, P. H., Stremmel, W., & Müller, M. (2005). TAp63alpha induces apoptosis by activating signaling via death receptors and mitochondria. *The EMBO journal*, 24(13), 2458–2471.
- Guo, X., Keyes, W. M., Papazoglu, C., Zuber, J., Li, W., Lowe, S. W., Vogel, H., & Mills, A. A. (2009). TAp63 induces senescence and suppresses tumorigenesis in vivo. *Nature cell biology*, 11(12), 1451–1457.
- Gurung, R.L., Lim, S.N., Khaw, A.K., Soon, J.F., Shenoy, K., & Mohamed, A.S. (2010). Thymoquinone induces telomere shortening, DNA damage and apoptosis in human glioblastoma cells. *PLoS One*, 5, 12124.
- Haffty, B.G., Yang, Q., Reiss, M., Kearney, T., Higgins, S.A., Weidhaas, J., Harris, L., Hait, W., & Toppmeyer, D. (2006). Locoregional Relapse and Distant Metastasis in Conservatively Managed Triple Negative Early-Stage Breast Cancer. *Journal of Clinical Oncology*, 24:36, 5652-5657.
- Ham, B., Fernandez, M. C., D'Costa, Z., & Brodt, P. (2016). The diverse roles of the TNF axis in cancer progression and metastasis. *Trends in cancer research*, 11(1), 1–27.
- Harpole, J. L., Tucci, M., & Benghuzzi, H. (2015). Pathophysiological Effects of Thymoquinone and Epigallocatechin-3-Gallate on SK-OV-3 Ovarian Cancer Like Cell Line. *Biomedical sciences instrumentation*, 51, 31–39.
- Hashemi, M., Fazaeli, A., Ghavami, S., Eskandari-Nasab, E., Arbabi, F., Mashhadi, M. A., Taheri, M., Chaabane, W., Jain, M. V., & Łos, M. J. (2013).
 Functional polymorphisms of FAS and FASL gene and risk of breast cancer pilot study of 134 cases. *PloS one*, 8(1), e53075.

- Hassan, M.S.U., Ansari, J., Spooner, D., & Hussain, S.A. (2010). Chemotherapy for breast cancer (Review). Oncology Reports, 24: 1121-1131.
- Hatakeyama, Y., Kobayashi, K., Nagano, T., Tamura, D., Yamamoto, M., Tachihara, M., Kotani, Y., & Nishimura, Y. (2014). Synergistic effects of pemetrexed and amrubicin in non-small cell lung cancer cell lines: Potential for combination therapy. *Cancer letters*, 343(1), 74–79.
- Hayati, F., Hossainzadeh, M., Shayanpour, S., Abedi-Gheshlaghi, Z., & Beladi Mousavi, S. S. (2015). Prevention of cisplatin nephrotoxicity. *Journal of nephropharmacology*, 5(1), 57-60.
- Hawkins DS, Demers GW & Galloway DA. (1996). Inactivation of p53 enhances sensitivity to multiple chemotherapeutic agents. *Cancer Res.*, 56, 892–898.
- He, W., Xia, Y., Cao, P., Hong, L., Zhang, T., Shen, X., & Zou, P. (2019). Curcuminoid WZ35 synergize with cisplatin by inducing ROS production and inhibiting TrxR1 activity in gastric cancer cells. *Journal of experimental* & clinical cancer research, CR, 38(1), 207.
- Hershberger, P.A., McGuire, T.F., Yu, W.D., Zuhowski, E.G., Schellens, J.H., Egorin, M.J., Trump, D.L., & Johnson, C.S. (2002). Cisplatin potentiates 1, 25-dihydroxyvitamin D3-induced apoptosis in association with increased mitogen-activated protein kinase kinase kinase 1 (MEKK-1) expression. *Mol. Cancer Ther*, 1, 821–829.
- Hill, D.P., Harper, A., Malcolm, J., McAndrews, M.S., Mockus, S.M., Patterson, S.E., Reynolds, T., baker, E.J., Bult, C.J., Chesler, E.J., & Blake, J.A. (2019) Cisplatin-resistant triple-negative breast cancer subtypes: multiple mechanisms of resistance. *BMC Cancer*, 19, 1039.
- Hoffman, R., Graham, L., & Newlands, E. S. (1989). Enhanced anti-proliferative action of busulphan by quercetin on the human leukaemia cell line K562. *British journal of cancer*, 59(3), 347–348.
- Holliday, D. L., & Speirs, V. (2011). Choosing the right cell line for breast cancer research. *Breast Cancer Research*, 13(4), 215.
- Hong, J. Y., Kim, G. H., Kim, J. W., Kwon, S. S., Sato, E. F., Cho, K. H., & Shim,E. B. (2012). Computational modeling of apoptotic signaling pathways induced by cisplatin. *BMC systems biology*, 6, 122.
- Hong, Y., Yang, J., Wu, W., Wang, W., Kong, X., Wang, Y., Yun, X., Zong, H., Wei, Y., Zhang, S., & Gu, J. (2008). Knockdown of BCL2L12 leads to

cisplatin resistance in MDA-MB-231 breast cancer cells. *Biochimica et biophysica acta*, 1782(11), 649–657.

- Horácek P & Drobník J. (1971). Interaction of cis-dichlorodiammineplatinum (II) with DNA. *Biochim Biophys Acta*, 254:341–347.
- Huang, J., Yu, S., Ji, C., & Li, J. (2015). Structural basis of cell apoptosis and necrosis in TNFR signaling. *Apoptosis : an international journal on* programmed cell death, 20(2), 210–215.
- Hu, X., Ma, J., Vikash, V., Li, J., Wu, D., Liu, Y., Zhang, J., & Dong, W. (2018a).
 Thymoquinone Augments Cisplatin-Induced Apoptosis on Esophageal Carcinoma Through Mitigating the Activation of JAK2/STAT3 Pathway. *Digestive diseases and sciences*, 63(1), 126–134.
- Hu, Y., Sun, B., Zhao, B., Mei, D., Gu, Q., & Tian, Z. (2018b). Cisplatin-induced cardiotoxicity with midrange ejection fraction: A case report and review of the literature. *Medicine*, 97(52), e13807.
- Hu, S., Li, X., Xu, R., Ye, L., Kong, H., Zeng, X., Wang, H., & Xie, W. (2016). The synergistic effect of resveratrol in combination with cisplatin on apoptosis via modulating autophagy in A549 cells. *Acta biochimica et biophysica Sinica*, 48(6), 528-35.
- Huang, L., Liu, Q., Chen, S., & Shao, Z. (2017). Cisplatin versus carboplatin in combination with paclitaxel as neoadjuvant regimen for triple negative breast cancer. *OncoTargets and therapy*, 10, 5739–5744.
- Huang, R. Y., Pei, L., Liu, Q., Chen, S., Dou, H., Shu, G., Yuan, Z. X., Lin, J., Peng,
 G., Zhang, W., & Fu, H. (2019). Isobologram Analysis: A Comprehensive
 Review of Methodology and Current Research. *Frontiers in pharmacology*, 10, 1222.
- Huszno, J., & Grzybowska, E. (2018). TP53 mutations and SNPs as prognostic and predictive factors in patients with breast cancer. *Oncology letters*, 16(1), 34– 40.
- Ibiyeye, K. M., Nordin, N., Ajat, M., & Zuki, A. (2019). Ultrastructural Changes and Antitumor Effects of Doxorubicin/Thymoquinone-Loaded CaCO3 Nanoparticles on Breast Cancer Cell Line. *Frontiers in oncology*, 9, 599.
- Imran, M., Raub, A., Khan, I.A., Shahbaz, M., Qaisrani, T.B., Fatmawati, S., Abu-Izneid, T., Imran, A., Rahman, K.U., & Gonda, T.A. (2018). Thymoquinone:

A novel strategy to combat cancer: A review. *Biomedicine & Pharmacotherapy*, 106 (2018): 390–402.

- Isakoff S. J. (2010). Triple-negative breast cancer: role of specific chemotherapy agents. *Cancer journal (Sudbury, Mass.)*, 16(1), 53–61.
- Israel, B.B., Tilghman, S.L., Parker-Lemieux, K., & Payton-Stewart, F. (2018). Phytochemicals: Current strategies for treating breast cancer (Review). Oncology Letters, 15: 7471-7478.
- Ishida S, Lee J, Thiele DJ, & Herskowitz I. (2002). Uptake of the anticancer drug cisplatin mediated by the copper transporter Ctr1 in yeast and mammals. *Proc Natl Acad Sci U S A*. 99:14298–14302.
- Iskender B, Izgi K, & Canatan H. (2016). Novel anti-cancer agent myrtucommulone-A and thymoquinone abrogate epithelial-mesenchymal transition in cancer cells mainly through the inhibition of PI3K/AKT signalling axis. *Mol Cell Biochem*, 416, 71–84.
- Islam, SS, Al-Sharif, I, Sultan, A, Al-Mazrou, A, Remmal, A, & Aboussekhra, A. (2018). Eugenol potentiates cisplatin anti-cancer activity through inhibition of ALDH-positive breast cancer stem cells and the NF-κB signaling pathway. *Molecular Carcinogenesis*, 57: 333–346.
- Jabbour, A. M., Heraud, J. E., Daunt, C. P., Kaufmann, T., Sandow, J., O'Reilly, L. A., Callus, B. A., Lopez, A., Strasser, A., Vaux, D. L., & Ekert, P. G. (2009).
 Puma indirectly activates Bax to cause apoptosis in the absence of Bid or Bim. *Cell death and differentiation*, 16(4), 555–563.
- Jafri, S. H., Glass, J., Shi, R., Zhang, S., Prince, M., & Kleiner-Hancock, H. (2010). Thymoquinone and cisplatin as a therapeutic combination in lung cancer: *In vitro* and *in vivo*. *Journal of Experimental & Clinical Cancer Research*, CR, 29(1), 87.
- Jan, R., & Chaudhry, G. E. (2019). Understanding Apoptosis and Apoptotic Pathways Targeted Cancer Therapeutics. Advanced pharmaceutical bulletin, 9(2), 205–218. https://doi.org/10.15171/apb.2019.024
- Ji, S., Orlikova, B., & Diederich, M. (2014). Non-edible plants as an attractive source of compounds with chemopreventive potential. *Journal of cancer prevention*, 19(1), 1-6.
- Jiang, Y., Guo, C., Vasko, M.R., & Kelley, M.R. (2008). Implications of Apurinic/Apyrimidinic Endonuclease in Reactive Oxygen Signaling

Response after Cisplatin Treatment of Dorsal Root Ganglion Neurons. *Cancer Res*, 68: 6425-6434.

- Jiang, Y., Ji, F., Liu, Y., He, M., Zhang, Z., Yang, J., Wang, N., Zhong, C., Jin, Q., Ye, X., & Chen, T. (2017). Cisplatin-induced autophagy protects breast cancer cells from apoptosis by regulating yes-associated protein. *Oncology reports*, 38(6), 3668–3676.
- Josephs, S. F., Ichim, T. E., Prince, S. M., Kesari, S., Marincola, F. M., Escobedo, A. R., & Jafri, A. (2018). Unleashing endogenous TNF-alpha as a cancer immunotherapeutic. *Journal of translational medicine*, 16(1), 242.
- Jordan, P. & Carmo-Fonseca, M. (2000). Molecular mechanisms involved in cisplatin cytotoxicity. *Cell. Mol. Life Sci*, 57: 1229.
- Juric, V., Chen, C. C., & Lau, L. F. (2009). Fas-mediated apoptosis is regulated by the extracellular matrix protein CCN1 (CYR61) in vitro and in vivo. *Molecular and cellular biology*, 29(12), 3266–3279.
- Kabil, N., Bayraktar, R., & Kahraman, N. (2018). Thymoquinone inhibits cell proliferation, migration, and invasion by regulating the elongation factor 2 kinase (eEF-2K) signaling axis in triple-negative breast cancer. *Breast Cancer Res Treat*, 171(3): 593–605.
- Kannan, K., Amariglio, N., Rechavi, G., Jakob-Hirsch, J., Kela, I., Kaminski, N., Getz, G., Domany, E., & Givol, D. (2001). DNA microarrays identification of primary and secondary target genes regulated by p53. *Oncogene*, 20(18), 2225–2234.
- Kaseb, A.O., Chinnakannu, K., Chen, D. (2007). Androgen Receptor– and E2F-1– Targeted Thymoquinone Therapy for Hormone-Refractory Prostate Cancer. *Cell, Tumor, and Stem Biology*, 67(16), 7782-7788.
- Katoh I, Aisaki KI, Kurata SI, Ikawa S and Ikawa Y. (2000). p51A (TAp63gamma), a p53 homolog, accumulates in response to DNA damage for cell regulation. *Oncogene*, 19, 3126–3130.
- Kay, J. G., & Grinstein, S. (2011). Sensing phosphatidylserine in cellular membranes. Sensors (Basel, Switzerland), 11(2), 1744–1755. https://doi.org/10.3390/s110201744
- Kensara, O. A., El-Shemi, A. G., Mohamed, A. M., Refaat, B., Idris, S., & Ahmad, J. (2016). Thymoquinone subdues tumor growth and potentiates the chemopreventive effect of 5-fluorouracil on the early stages of colorectal

carcinogenesis in rats. *Drug design, development and therapy*, 10, 2239–2253.

- Kerr, J. F., Wyllie, A. H., & Currie, A. R. (1972). Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British journal of cancer*, 26(4), 239–257.
- Koraneekit, A., Limpaiboon, T., Sangka, A., Boonsiri, P., Daduang, S., & Daduang, J. (2018). Synergistic effects of cisplatin-caffeic acid induces apoptosis in human cervical cancer cells via the mitochondrial pathways. *Oncology letters*, 15(5), 7397-7402.
- Kundu J, Choi BY, Jeong CH, Kundu JK, & Chun KS. (2014). Thymoquinone induces apoptosis in human colon cancer HCT116 cells through inactivation of STAT3 by blocking JAK2- and Src mediated phosphorylation of EGF receptor tyrosine kinase. *Oncol Rep*, 32, 821–828.
- Kharbanda, S., Pandey, P., Yamauchi, T., Kumar, S., Kaneki, M., Kumar, V., & Kufe, D. (2000). Activation of MEK kinase 1 by the c-Abl protein tyrosine kinase in response to DNA damage. *Molecular and cellular biology*, 20(14), 4979–4989.
- Kapinova, A., Kubatka, P., Golubnitschaja, O., Kello, M., Zubor, P., Solar, P., & Pec, M. (2018). Dietary phytochemicals in breast cancer research: anticancer effects and potential utility for effective chemoprevention. *Environmental health and preventive medicine*, 23(1), 36.
- Khader, M., & Eckl, P. M. (2014). Thymoquinone: an emerging natural drug with a wide range of medical applications. *Iranian journal of basic medical sciences*, 17(12), 950-7.
- Khan, M. A., Chen, H. C., Tania, M., & Zhang, D. Z. (2011). Anticancer activities of Nigella sativa (black cumin). *African journal of traditional, complementary,* and alternative medicines : AJTCAM, 8(5 Suppl), 226-32.
- Khan, M. A., Tania, M., Wei, C., Mei, Z., Fu, S., Cheng, J., & Fu, J. (2015). Thymoquinone inhibits cancer metastasis by downregulating TWIST1 expression to reduce epithelial to mesenchymal transition. *Oncotarget*, 6(23), 19580–19591.
- Khan, A., Aldebasi, Y. H., Alsuhaibani, S. A., & Khan, M. A. (2019a). Thymoquinone Augments Cyclophosphamide-Mediated Inhibition of Cell

Proliferation in Breast Cancer Cells. *Asian Pacific journal of cancer prevention : APJCP*, 20(4), 1153–1160.

- Khan, M. A., Tania, M., & Fu, J. (2019b). Epigenetic role of thymoquinone: impact on cellular mechanism and cancer therapeutics. *Drug discovery today*, 24(12), 2315–2322.
- Kleih, M., Böpple, K., Dong, M., Gaißler, A., Heine, S., Olayioye, M. A., Aulitzky, W. E., & Essmann, F. (2019). Direct impact of cisplatin on mitochondria induces ROS production that dictates cell fate of ovarian cancer cells. *Cell death & disease*, 10(11), 851.
- Koka, P.S., Mondal, D., Schultz, M., Abdel-Mageed, A.B. & Agrawal, K.C. Studies on molecular mechanisms of growth inhibitory effects of thymoquinone against prostate cancer cells: role of reactive oxygen species. *Exp Biol Med*, 235, 751–760.
- Korbakis, D., & Scorilas, A. (2012). Quantitative expression analysis of the apoptosis-related genes BCL2, BAX and BCL2L12 in gastric adenocarcinoma cells following treatment with the anticancer drugs cisplatin, etoposide and taxol. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*, 33(3), 865–875.
- Kotowski, U., Heiduschka, G., Kadletz, L., Fahim, T., Seemann, R., Schmid, R., & Thurnher, D. (2017). Effect of thymoquinone on head and neck squamous cell carcinoma cells in vitro: Synergism with radiation. *Oncology Letters*, 14(1), 1147–1151.
- Krajcí, D., Mares, V., Lisá, V., Spanová, A., & Vorlícek, J. (2000). Ultrastructure of nuclei of cisplatin-treated C6 glioma cells undergoing apoptosis. *European journal of cell biology*, 79(5), 365–376.
- Kroemer, G., Galluzzi, L., & Brenner, C. (2007). Mitochondrial membrane permeabilization in cell death. *Physiol. Rev*, 87, 99–163.
- Kroemer, G., El-Deiry, W. S., Golstein, P., Peter, M. E., Vaux, D., Vandenabeele, P., Zhivotovsky, B., Blagosklonny, M. V., Malorni, W., Knight, R. A., Piacentini, M., Nagata, S., Melino, G., & Nomenclature Committee on Cell Death (2005). Classification of cell death: recommendations of the Nomenclature Committee on Cell Death. Cell death and differentiation, 12 Suppl 2, 1463–1467.

- Kutuk, O., Arisan, E. D., Tezil, T., Shoshan, M. C., & Basaga, H. (2009). Cisplatin overcomes Bcl-2-mediated resistance to apoptosis via preferential engagement of Bak: critical role of Noxa-mediated lipid peroxidation. *Carcinogenesis*, 30(9), 1517–1527.
- Laurenzi, V.D. & Melino, G. (2000). Evolution of functions within the p53/p63/p73 family. *Annals of the New York Academy of Sciences*, 926, 90–100.
- Lamson, D. W., & Brignall, M. S. (2000). Antioxidants and cancer, part 3: quercetin. *Alternative medicine review : a journal of clinical therapeutic*, 5(3), 196–208.
- LeBlanc, H., Lawrence, D., Varfolomeev, E., Totpal, K., Morlan, J., Schow, P., Fong, S., Schwall, R., Sinicropi, D., & Ashkenazi, A. (2002). Tumor-cell resistance to death receptor--induced apoptosis through mutational inactivation of the proapoptotic Bcl-2 homolog Bax. *Nat Med*, 8 (3), 274-281.
- Lee, S., Han, S., Jeong, A.L., Park, J.S., Jung, S.H., Choi, K.D., & Yang, Y. (2015). Combined Treatment of Herbal Mixture Extract H9 with Trastuzumab Enhances Anti-tumor Growth Effect. *J.Microbiol. Biotechnol*, 25(7), 1036-1046.
- Lebwohl, D., & Canetta, R. (1998). Clinical development of platinum complexes in cancer therapy: an historical perspective and an update. *European Journal of Cancer*, 34(10), 1522 1534.
- Leong, C.O., Vidnovic, N. DeYoung, M.P., Sgroi, D., & Ellisen, L.W. (2007). The p63/p73 network mediates chemosensitivity to cisplatin in a biologically defined subset of primary breast cancers. *J Clin Invest*, 117(5), 1370–1380.
- Lebert, J. M., Lester, R., Powell, E., Seal, M., & McCarthy, J. (2018). Advances in the systemic treatment of triple-negative breast cancer. *Current oncology* (*Toronto, Ont.*), 25(1): S142-S150.
- Lehar J, Krueger AS, Avery W, Heilbut AM, Johansen LM, et al. (2009) Synergistic drug combinations tend to improve therapeutically relevant selectivity. *Nat Biotechnol* 27: 659–666.
- Lehmann, B.D., Bauer, J.A., Chen, X., Sanders, M.E., Chakravarthy, A.B., & Shyr, Y. (2011). Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J Clin Invest*, 121(7): 2750-2767.
- Leist, M., & Jaattela, M. (2001). Four deaths and a funeral: from caspases to alternative mechanisms. Nat. Rev. *Mol. Cell Biol.*, 2, 589–598.

- Lei, X., Lv, X., Liu, M., Yang, Z., Ji, M., & Guo, X. (2012). Thymoquinone inhibits growth and augments 5-fluorouracil-induced apoptosis in gastric cancer cells both in vitro and in vivo. *Biochem. Biophys. Res. Commun.* 417, 864–868.
- Lemaire M. A., Schwartz A., Rahmouni A. R. and Leng M. (1991) Interstrand crosslinks are preferentially formed at the d(GC) sites in the reaction between cisdiamminedichloroplatinum (II) and DNA. *Proc. Natl. Acad. Sci. USA* 88: 1982–1985 30
- Lieberthal, W., Triaca, V., & Levine, J. (1996). Mechanisms of death induced by cisplatin in proximal tubular epithelial cells: apoptosis vs. necrosis. Am J Physiol, 270, F700–F708.
- Lim, L. Y., Vidnovic, N., Ellisen, L. W., & Leong, C. O. (2009). Mutant p53 mediates survival of breast cancer cells. *British journal of cancer*, 101(9), 1606–1612.
- Li, X., Miao, X., Wang, H., Xu, Z., & Li, B. (2015a). The tissue dependent interactions between p53 and Bcl-2 in vivo. *Oncotarget*, 6(34), 35699-709.
- Li, S., Tan, H. Y., Wang, N., Zhang, Z. J., Lao, L., Wong, C. W., & Feng, Y. (2015b). The Role of Oxidative Stress and Antioxidants in Liver Diseases. *International journal of molecular sciences*, 16(11), 26087-124.
- Li, F., Rajendran, P., & Sethi, G. (2010). Thymoquinone inhibits proliferation, induces apoptosis and chemosensitizes human multiple myeloma cells through suppression of signal transducer and activator of transcription 3 activation pathway. *British Journal of Pharmacology*, 161(3), 541–554.
- Li, J., & Yuan, J. (2008). Caspases in apoptosis and beyond. *Oncogene*, 27(48), 6194–6206.
- Li, S., Li, Q., Lü, J., Zhao, Q., Li, D., Shen, L., Wang, Z., Liu, J., Xie, D., Cho, W. C., Xu, S., & Yu, Z. (2020). Targeted Inhibition of miR-221/222 Promotes Cell Sensitivity to Cisplatin in Triple-Negative Breast Cancer MDA-MB-231 Cells. *Frontiers in genetics*, 10, 1278.
- Liedtke, C., Mazouni, C., Hess, K.R., André, F., Tordai, A., Mejia, J.A., Symmans, W.F., Gonzalez-Angulo, A.M., Hennessy, B., Green, M., Cristofanilli, M., Hortobagyi, G.N., & Pusztai, L. (2008). Response to neoadjuvant therapy and long-term survival in patients with triple-negative breast cancer. J Clin Oncol, 26(8), 1275-1281.

- Liu, X., Dong, J., Cai, W., Pan, Y., Li R., & Li, B. (2017). The Effect of Thymoquinone on Apoptosis of SK-OV-3 Ovarian Cancer Cell by Regulation of Bcl-2 and Bax. *Int J Gynecol Cancer*, 27(8), 596-1601.
- Liu, X.D., Li, M., Li, W.X., Wang, Q.Y., & Zhang, H.X. (2019). Combined Effect of Lentinan and Cisplatin on Cytokines IL-6, TNF-α, and TGF-β in Tumor Therapy. *International Journal of Polymer Science, Hindawi*, 11 pages.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) *Method. Methods (San Diego, Calif.)*, 25(4), 402–408.
- Ma, Y., Zhu, B., Yong, L., Song, C., Liu, X., Yu, H., & Liu, X. (2016). Regulation of Intrinsic and Extrinsic Apoptotic Pathways in Osteosarcoma Cells Following Oleandrin Treatment. *International journal of molecular sciences*, 17(11), 1950.
- Ma, J., Hu, X., Li, J., Wu, D., Lan, Q., Wang, Q., & Dong, W. (2017). Enhancing conventional chemotherapy drug cisplatin-induced anti-tumor effects on human gastric cancer cells both in vitro and in vivo by Thymoquinone targeting PTEN gene. *Oncotarget*, 8(49), 85926–85939.
- Maciejczyk A, & Surowiak P (2013). Quercetin inhibits proliferation and increases sensitivity of ovarian cancer cells to cisplatin and paclitaxel. *Ginekol Pol.*, 84(7):590-595.
- Makki, J. (2015). Diversity of Breast Carcinoma: Histological Subtypes and Clinical Relevance. *Clin Med Insights Pathol*, 8, 23–31.
- Malhotra, G. K., Zhao, X., Band, H., & Band, V. (2010). Histological, molecular and functional subtypes of breast cancers. *Cancer biology & therapy*, 10(10), 955–960.
- Mansour, M., Ginawi, O., El-Hadiyah, T., El-Khatib, A., Al-Shabanah, O., and Al-Sawaf, H. (2001). Effects of volatile oil constituents of Nigella sativa on carbon tetrachloride-induced hepatotoxicity in mice: evidence for antioxidant effects of thymoquinone. Res. Commun. Mol. Pathol. *Pharmacol.* 110, 239–252.
- Mantri, Y., Lippard, S. J., & Baik, M. H. (2007). Bifunctional binding of cisplatin to DNA: why does cisplatin form 1,2-intrastrand cross-links with ag but not with GA?. *Journal of the American Chemical Society*, 129(16), 5023–5030.

- Martinho, N., Santos, T., Florindo, H. F., & Silva, L. C. (2019). Cisplatin-Membrane Interactions and Their Influence on Platinum Complexes Activity and Toxicity. *Frontiers in physiology*, 9, 1898.
- Matsuzaki, I., Suzuki, H., Kitamura, M., Minamiya, Y., Kawai, H., & Ogawa, J. (2000). Cisplatin induces fas expression in esophageal cancer cell lines and enhanced cytotoxicity in combination with LAK cells. *Oncology*, 59(4), 336– 343.
- Mello J. A., Lippard S. J. and Essigmann J. M. (1995) DNA adducts of cisdiamminedichloroplatinum (II) and its trans-isomer inhibit RNA polymerase II differentially in vivo. *Biochemistry* 34: 14783–14791.
- Misra, S., Zhang, X., Wani, N. A., Sizemore, S., & Ray, A. (2020). Both BRCA1wild type and -mutant triple-negative breast cancers show sensitivity to the NAE inhibitor MLN4924 which is enhanced upon MLN4924 and cisplatin combination treatment. *Oncotarget*, 11(8), 784–800.
- Mistry, V.D. (2016). Understanding the mechanistic aspect of Thymoquinone in breast cancer by employing different nano composites (Doctoral dissertation). Retrieved from RMIT Research Repository.
- Michalak, E. M., Jansen, E. S., Happo, L., Cragg, M. S., Tai, L., Smyth, G. K., Strasser, A., Adams, J. M., & Scott, C. L. (2009). Puma and to a lesser extent Noxa are suppressors of Myc-induced lymphomagenesis. *Cell death and differentiation*, 16(5), 684–696.
- Micheau, O., Solary, E., Hammann ,A., Martin, F., & Dimanche-Boitrel, M.T. (1997). Sensitization of cancer cells treated with cytotoxic drugs to fasmediated cytotoxicity. J. Natl. Cancer Inst., 89, 783–789.
- Mondal, J., Panigrahi, A.K., and Khuda-Bukhsh, A.R. (2015). Physico-chemical and ultra-structural characterizations of PLGA-loaded nanoparticles of Boldine and their efficacy in ameliorating cisplatin induced hepatotoxicity in normal liver cells in vitro. *JIPBS*, 2 (4), 506-521.
- Monneret, C. (2011). Platinum anticancer drugs. From serendipity to rational design. Ann Pharm Fr, 69(6), 286-95.
- Monroe, J. D., Hodzic, D., Millay, M. H., Patty, B. G., & Smith, M. E. (2019). Anti-Cancer and Ototoxicity Characteristics of the Curcuminoids, CLEFMA and EF24, in Combination with Cisplatin. *Molecules (Basel, Switzerland)*, 24(21), 3889.

- Moll UM, Erster S & Zaika A . (2001). p53, p63 and p73--solos, alliances and feuds among family members. *Biochim. Biophys. Acta*, 1552, 47–59.
- Möltgen, S., Piumatti, E., Massafra, G., Metzger, S., Jaehde, U., & Kalayda, G. (2020). Cisplatin Protein Binding Partners and Their Relevance for Platinum Drug Sensitivity. *Cells*, 9(6), 1322. MDPI AG.
- Mostofa, A.G.M., Hossain, M.K., Basak, D., & Sayeed, M.S. (2017). Thymoquinone as a Potential Adjuvant Therapy for Cancer Treatment: Evidence from Preclinical Studies. *Front Pharmacol*, 8, 295.
- Motulsky, H.J. (2016). GraphPad Curve Fitting Guide. Available at: <u>http://www.graphpad.com/guides/prism/7/curve-fitting/index.htm</u> (Accessed 21 July 2020).
- Mork, H.H. (2012). *Molecular profiling of Ductal Carcinoma In Situ*. Master Thesis, Norwegian University of Science and Technology.
- Mu, G. G., Zhang, L. L., Li, H. Y., Liao, Y., & Yu, H. G. (2015). Thymoquinone Pretreatment Overcomes the Insensitivity and Potentiates the Antitumor Effect of Gemcitabine Through Abrogation of Notch1, PI3K/Akt/mTOR Regulated Signaling Pathways in Pancreatic Cancer. *Digestive diseases and sciences*, 60(4), 1067–1080.
- Mustacchi, G., & De Laurentiis, M. (2015). The role of taxanes in triple-negative breast cancer: literature review. *Drug design, development and therapy*, 9, 4303-18.
- Müller, M., Wilder, S., Bannasch, D., Israeli, D., Lehlbach, K., Li-Weber, M., & Krammer, P. H. (1998). p53 activates the CD95 (APO-1/Fas) gene in response to DNA damage by anticancer drugs. *The Journal of experimental medicine*, 188(11), 2033–2045.
- National Cancer Institute. (2018). Understanding Cancer. Available at: http://ncinagpur.in/detail/understanding-cancer
- National Cancer Registry, National Cancer Institute, and Ministry of Health Malaysia (2018). Malaysian Study on Cancer Survival (MySCan). Available at: http://nci.moh.gov.my/index.php/ms/list-penerbitan/35-laporan/522myscan
- Nessa, M.U., Beale, P., Chan, C., Yu, J.Q., & Huq, F. (2011). Synergism from Combinations of Cisplatin and Oxaliplatin with Quercetin and

Thymoquinone in Human Ovarian Tumour Models. *Anticancer Res*, 31 (11), 3789-3797.

- Ndreshkjana, B., Çapci, A., Klein, V., Chanvorachote, P., Muenzner, J. K., Huebner, K., & Schneider-Stock, R. (2019). Combination of 5-fluorouracil and thymoquinone targets stem cell gene signature in colorectal cancer cells. *Cell death* & *disease*, 10(6), 379.
- Ng, W.K., Yazan, L.S., & Ismail, M. (2011). Thymoquinone from Nigella sativa was more potent than cisplatin in eliminating of SiHa cells via apoptosis with down-regulation of Bcl-2 protein. *Toxicology in Vitro*, 25(7), 392-1398.
- Niedner, H., Christen, R., Lin, X., Kondo, A., & Howell, S. B. (2001). Identification of genes that mediate sensitivity to cisplatin. *Molecular pharmacology*, 60(6), 1153–1160.
- Ning, Y., Hui, N., Qing, B., Zhuo, Y., Sun, W., Du, Y., Liu, S., Liu, K., & Zhou, J. (2019). ZCCHC10 suppresses lung cancer progression and cisplatin resistance by attenuating MDM2-mediated p53 ubiquitination and degradation. *Cell Death Dis* 10, 414.
- Norwood, A. A., Tucci, M., & Benghuzzi, H. (2007). A comparison of 5-fluorouracil and natural chemotherapeutic agents, EGCG and thymoquinone, delivered by sustained drug delivery on colon cancer cells. *Biomedical sciences instrumentation*, 43, 272–277.
- Norbury, C. J., & Hickson, I. D. (2001). Cellular Response to DNA Damage. *Annual Review of Pharmacology and Toxicology*, 41(1), 367–401.
- Odeh, L.H., Talib, W.H., & Basheti, I.A. (2018). Synergistic effect of thymoquinone and melatonin against breast cancer implanted in mice. *Journal of Cancer Research and Therapeutics*, 1-7.
- Olszewski, U., Ach, F., Ulsperger, E., Baumgartner, G., Zeillinger, R., Bednarski, P., & Hamilton, G. (2009). In Vitro Evaluation of Oxoplatin: An Oral Platinum (IV) Anticancer Agent. *Metal-Based Drugs*, 348916.
- Olivero, O.A., Chang, P.K., Lopez-Larraza, D.M., Semino-Mora, C.M., & Poirier, M.C. (1997). Preferential formation and decreased removal of cisplatin–DNA adducts in Chinese hamster ovary cell mitochondrial DNA as compared to nuclear DNA. *Mutation; Research /Genetic Toxicology and Environmental. Mutagenesis*, 391: 79-86.

- Oun, R., Moussa, Y.E., & Wheate, N.J. (2018). The side effects of platinum-based chemotherapy drugs: a review for chemists. *Dalton Trans*, 47, 6645.
- Ozdemir, N., Kantekin-Erdogan, M. N., Tat, T., & Tekin, A. (2018). Effect of black cumin oil on the oxidative stability and sensory characteristics of mayonnaise. *Journal of food science and technology*, 55(4), 1562–1568.
- Pan, T., Mao, T., Yang, H., Wang, H., & Wang, Y. (2018). Silencing of TGIF sensitizes MDA-MB-231 human breast cancer cells to cisplatin-induced apoptosis. *Experimental and therapeutic medicine*, 15(3), 2978–2984. https://doi.org/10.3892/etm.2018.5780
- Paramasivam, A., Sambantham, S., Shabnam, J., Raghunandhakumar, S., Anandan, B., & Rajiv, R. (2012). Anti-cancer effects of thymoquinone in mouse neuroblastoma (Neuro-2a) cells through caspase-3 activation with downregulation of XIAP. *Toxicol. Lett.* 213, 151–159.
- Park, E. J., Chauhan, A. K., Min, K. J., Park, D. C., & Kwon, T. K. (2016).
 Thymoquinone induces apoptosis through downregulation of c-FLIP and Bcl-2 in renal carcinoma Caki cells. *Oncology reports*, 36(4), 2261–2267.
- Park, J.H., Ahn, J., & Kim, S. (2018). How shall we treat early triple-negative breast cancer (TNBC): from the current standard to upcoming immuno-molecular strategies *ESMO Open*, 3:e000357.
- Payet D, Gaucheron F, Sip M, & Leng M. (1993). Instability of the monofunctional adducts in cis-[Pt(NH3)2(N7-N-methyl-2- diazapyrenium)Cl](2+)-modified DNA: rates of -linking reactions in cis-platinum-modified DNA. *Nucleic* Acids Res. 21:5846–5851.
- Pauzi, A. Z., Yeap, S. K., Abu, N., Lim, K. L., Omar, A. R., Aziz, S. A., Chow, A. L., Subramani, T., Tan, S. G., & Alitheen, N. B. (2016). Combination of cisplatin and bromelain exerts synergistic cytotoxic effects against breast cancer cell line MDA-MB-231 in vitro. *Chinese medicine*, 11, 46.
- Peddi, P. F., Ellis, M. J., & Ma, C. (2012). Molecular basis of triple negative breast cancer and implications for therapy. *International journal of breast cancer*, 2012, 217185.
- Perri, F., Pisconti, S., & Della Vittoria Scarpati, G. (2016). P53 mutations and cancer: a tight linkage. *Annals of translational medicine*, 4(24), 522.

- Pegram, M. D., Lopez, A., Konecny, G., & Slamon, D. J. (2000). Trastuzumab and chemotherapeutics: drug interactions and synergies. *Semin. Oncol.* 27(6 *Suppl.* 11), 21–25. Discussion: 92–100.
- Pfeffer, C. M., & Singh, A. (2018). Apoptosis: A Target for Anticancer Therapy. International journal of molecular sciences, 19(2), 448.
- Pflaum, J., Schlosser, S., & Müller, M. (2014). p53 Family and Cellular Stress Responses in Cancer. *Frontiers in oncology*, 4, 285.
- Pistritto, G., Trisciuoglio, D., Ceci, C., Garufi, A., & D'Orazi, G. (2016). Apoptosis as anticancer mechanism: function and dysfunction of its modulators and targeted therapeutic strategies. *Aging*, 8(4), 603–619. https://doi.org/10.18632/aging.100934
- PLANTS Database. (2019). *Nigella sativa L. black cumin*. Available at: <u>https://plants.usda.gov/core/profile?symbol=NISA2</u> (Accessed: 6 February 2020).
- Pratibha, R., Sameer, R., Padmanabh, Rataboli, V., Dayanand, Bhiwgade, A., & Dhume, C.Y. (2006). Enzymatic studies of cisplatin induced oxidative stress in hepatic tissue of rats, *European Journal of Pharmacology*, 532(3), 290-293.
- Pezzani, R., Salehi, B., Vitalini, S., Iriti, M., Zuñiga, F. A., Sharifi-Rad, J., & Martins, N. (2019). Synergistic Effects of Plant Derivatives and Conventional Chemotherapeutic Agents: An Update on the Cancer Perspective. *Medicina* (*Kaunas, Lithuania*), 55(4), 110.
- Prokhorova, E. A., Kopeina, G. S., Lavrik, I. N., & Zhivotovsky, B. (2018). Apoptosis regulation by subcellular relocation of caspases. *Scientific reports*, 8(1), 12199.
- Ramesh, G., & Reeves, W. B. (2003). TNFR2-mediated apoptosis and necrosis in cisplatin-induced acute renal failure. *American journal of physiology. Renal physiology*, 285(4), F610–F618.
- Rajput S, Kumar BNP, Sarkar S, Das S, & Azab B. (2013). Targeted Apoptotic Effects of Thymoquinone and Tamoxifen on XIAP Mediated Akt Regulation in Breast Cancer. *PLOS ONE*, 8(4), e61342.
- Raina, R., Hussain, A., & Sharma, R. (2020). Molecular insight into apoptosis mediated by flavones in cancer (Review). World Academy of Sciences Journal, 2, 1-1.

- Relles, D., Chipitsyna, G. I., Gong, Q., Yeo, C. J., & Arafat, H. A. (2016). Thymoquinone Promotes Pancreatic Cancer Cell Death and Reduction of Tumor Size through Combined Inhibition of Histone Deacetylation and Induction of Histone Acetylation. *Advances in preventive medicine*, 2016, 1407840.
- Riley, T., Sontag, E., Chen, P., & Levine, A. (2008). Transcriptional control of human p53-regulated genes. Nature reviews. *Molecular cell biology*, 9(5), 402–412.
- Riss, T.L., Moravec, R.A., Niles, A.L. (2013). Cell Viability Assays. In: Sittampalam, G.S., Grossman, A., & Brimacombe, K., editors. Assay Guidance Manual [Internet]. Bethesda (MD): Eli Lilly & Company and the National Center for Advancing Translational Sciences; 2004-. Available from: https://www.ncbi.nlm.nih.gov/books/NBK144065/
- Rocha, C., Silva, M. M., Quinet, A., Cabral-Neto, J. B., & Menck, C. (2018). DNA repair pathways and cisplatin resistance: an intimate relationship. Clinics (Sao Paulo, Brazil), 73(suppl 1), e478s.
- Rouzier R, Perou CM, & Symmans WF. (2005). Breast cancer molecular subtypes respond differently to preoperative chemotherapy. *Clin Cancer Res*, 11:5678-85.
- Roepke, M., Diestel, A., Bajbouj, K., Walluscheck, D., Schonfeld, P., Roessner, A., Schneider-Stock, R., & Gali-Muhtasib, H. (2007). Lack of p53 augments thymoquinone-induced apoptosis and caspase activation in human osteosarcoma cells. *Cancer Biol Ther*. 6, 160–169.
- Rosenberg, B., Van Camp, L., & Krigas, T. (1965). Inhibition of cell division in Escherichia coli by electrolysis products from a platinum electrode. *Nature*, 205(4972), 698–699, 1965.
- Rosenberg, B., & VanCamp, L.(1970). The successful regression of large solid sarcoma 180 tumors by platinum compounds. *Cancer Research*, 30(6): 1799– 1802.
- Reindl W, Yuan J, Krämer A, Strebhardt K, & Berg T. (2008). Inhibition of polo-like kinase 1 by blocking polo-box domain-dependent protein-protein interactions. *Chem Biol*, 15, 459–466.
- Sadhukhan, P., Saha, S., Dutta, S., & Sil, P. C. (2018). Mangiferin Ameliorates Cisplatin Induced Acute Kidney Injury by Upregulating Nrf-2 via the

Activation of PI3K and Exhibits Synergistic Anticancer Activity With Cisplatin. *Frontiers in pharmacology*, 9, 638.

- Sakalar, C., Yuruk, M., Kaya, T., Aytekin, M., Kuk, S., & Canatan, H. (2013). Pronounced transcriptional regulation of apoptotic and TNF-NF-kappa-B signaling genes during the course of thymoquinone mediated apoptosis in HeLa cells. *Molecular and cellular biochemistry*, 383(1-2), 243–251.
- Şakalar, C., İzgi, K., İskender, B., Sezen, S., Aksu, H., Çakır, M., Kurt, B., Turan, A., & Canatan, H. (2016). The combination of thymoquinone and paclitaxel shows anti-tumor activity through the interplay with apoptosis network in triple negative breast.
- Seki, K., Yoshikawa, H., Shiiki, K., Hamada, Y., Akamatsu, N., & Tasaka, K. (2000). Cisplatin (CDDP) specifically induces apoptosis via sequential activation of caspase-8, -3 and -6 in osteosarcoma. *Cancer chemotherapy and pharmacology*, 45(3), 199–206.
- Sandra, M., Sancho-Martínez, F., Javier Piedrafita, Jorge B. Cannata-Andía, José, M., López-Novoa, Francisco, J., & López-Hernández (2011). Necrotic Concentrations of Cisplatin Activate the Apoptotic Machinery but Inhibit Effector Caspases and Interfere with the Execution of Apoptosis, *Toxicological Sciences*, 122 (1), 73–85.
- Santos, N.A., Catão, C.S., Martins, N.M., Curti, C., & Bianchi, M.L. (2007). Cisplatin-induced nephrotoxicity is associated with oxidative stress, redox state unbalance, impairment of energetic metabolism and apoptosis in rat kidney mitochondria. *Arch Toxicol*, 81: 495-504.
- Sarosiek, K. A., Chi, X., Bachman, J. A., Sims, J. J., Montero, J., Patel, L., Flanagan, A., Andrews, D. W., Sorger, P., & Letai, A. (2013). BID preferentially activates BAK while BIM preferentially activates BAX, affecting chemotherapy response. *Molecular cell*, 51(6), 751–765.
- Schmittgen, T. D., Zakrajsek, B. A., Mills, A. G., Gorn, V., Singer, M. J., & Reed, M. W. (2000). Quantitative reverse transcription-polymerase chain reaction to study mRNA decay: comparison of endpoint and real-time methods. *Analytical biochemistry*, 285(2), 194–204.
- Scian, M. J., Carchman, E. H., Mohanraj, L., Stagliano, K. E., Anderson, M. A., Deb, D., Crane, B. M., Kiyono, T., Windle, B., Deb, S. P., & Deb, S. (2008). Wild-

type p53 and p73 negatively regulate expression of proliferation related genes. *Oncogene*, 27(18), 2583–2593.

- Sen, S., Sharma, H., & Singh, N. (2005). Curcumin enhances Vinorelbine mediated apoptosis in NSCLC cells by the mitochondrial pathway. *Biochemical and biophysical research communications*, 331(4), 1245–1252.
- Seca, A., & Pinto, D. (2018). Plant Secondary Metabolites as Anticancer Agents: Successes in Clinical Trials and Therapeutic Application. *International journal of molecular sciences*, 19(1), 263.
- Shahraki, S., Mohebbati, R., Shafei, M. N., Mahmoudi, M., Hosseinian, S., Parhizgar, S., Yazd, Z., Heravi, N. E., Abadi, R., & Khajavirad, A. (2019). Induction of Apoptosis and Growth-Inhibition by Thymoquinone in ACHN and GP-293 Cell Lines in Comparable with Cis-Platinum. *Journal of pharmacopuncture*, 22(3), 176–183.
- Shamloo, B., & Usluer, S. (2019). p21 in Cancer Research. Cancers, 11(8), 1178.
- Sharma, G.N., Dave, R., Sanadya, J., Sharma, P., & Sharma, K.K. (2010). VARIOUS TYPES AND MANAGEMENT OF BREAST CANCER: AN OVERVIEW. J Adv Pharm Technol Res, 1(2), 109–126.
- Sharma, K., Vu, T. T., Cook, W., Naseri, M., Zhan, K., Nakajima, W., & Harada, H. (2018). p53-independent Noxa induction by cisplatin is regulated by ATF3/ATF4 in head and neck squamous cell carcinoma cells. *Molecular oncology*, 12(6), 788–798.
- Shen, M., Duan, W. M., Wu, M. Y., Wang, W. J., Liu, L., Xu, M. D., Zhu, J., Li, D. M., Gui, Q., Lian, L., Gong, F. R., Chen, K., Li, W., & Tao, M. (2015). Participation of autophagy in the cytotoxicity against breast cancer cells by cisplatin. *Oncology reports*, 34(1), 359–367.
- Sheth, S., Mukherjea, D., Rybak, L. P., & Ramkumar, V. (2017). Mechanisms of Cisplatin-Induced Ototoxicity and Otoprotection. Frontiers in cellular neuroscience, 11, 338. cancer. *Tumour Biol*, 37, 4467–4477.
- Shibue, T., Suzuki, S., Okamoto, H., Yoshida, H., Ohba, Y., Takaoka, A., & Taniguchi, T. (2006). Differential contribution of Puma and Noxa in dual regulation of p53-mediated apoptotic pathways. *The EMBO journal*, 25(20), 4952-62.

- Shoieb, A. M., Elgayyar, M., Dudrick, P. S., Bell, J. L., & Tithof, P. K. (2003). In vitro inhibition of growth and induction of apoptosis in cancer cell lines by thymoquinone. *International journal of oncology*, 22(1), 107–113.
- Simpson, P.V., Desai, N.M., Casari, I., Massi, M., & Falasca, M. (2019). Metalbased antitumor compounds: beyond cisplatin. Future Medicinal Chemistry, Epub ahead of print.
- Singh, S., Sharma, B., Kanwar, S. S., & Kumar, A. (2016). Lead Phytochemicals for Anticancer Drug Development. *Frontiers in plant science*, 7, 1667.
- Sutton, K.M., Greenshields, A.L., & Hoskin, D.W. (2014). Thymoquinone: A bioactive component of black caraway seeds, causes G1 phase cell cycle arrest and apoptosis in triple-negative breast cancer cells with mutant p53. *Nutr Cancer*, 66, 408-18.
- Sørensen, B. H., Nielsen, D., Thorsteinsdottir, U. A., Hoffmann, E. K., & Lambert, I. H. (2016). Downregulation of LRRC8A protects human ovarian and alveolar carcinoma cells against Cisplatin-induced expression of p53, MDM2, p21Waf1/Cip1, and Caspase-9/-3 activation. American journal of physiology. *Cell physiology*, 310(11), C857–C873.
- Salem, M.L. (2005). Immunomodulatory and therapeutic properties of the Nigella sativa L. seed. *Int. Immunopharmacol.*, 5, 1749-1770.
- Salim, L. Z., Mohan, S., Othman, R., Abdelwahab, S. I., Kamalidehghan, B., Sheikh, B. Y., & Ibrahim, M. Y. (2013). Thymoquinone induces mitochondriamediated apoptosis in acute lymphoblastic leukaemia in vitro. *Molecules* (*Basel, Switzerland*), 18(9), 11219–11240.
- Salmani, J.M., Asghar, S., Lv, H., & Zhou, J. (2014). Aqueous solubility and degradation kinetics of the phytochemical anticancer thymoquinone; probing the effects of solvents, pH and light. *Molecules*, 19, 5925-5939.
- Scambia, G., Ranelletti, F. O., Panici, P. B., De Vincenzo, R., Bonanno, G., Ferrandina, G., Piantelli, M., Bussa, S., Rumi, C., & Cianfriglia, M. (1994). Quercetin potentiates the effect of adriamycin in a multidrug-resistant MCF-7 human breast-cancer cell line: P-glycoprotein as a possible target. *Cancer chemotherapy and pharmacology*, 34(6), 459–464.
- Sledge, G.W., Loehrer, P.J., Roth, B.J., & Einhorn, L.H. (1988). Cisplatin as firstline therapy for metastatic breast cancer. *J Clin Oncol*, 14:1811–1818.

- Siddik, Z.H. (2003). Cisplatin: mode of cytotoxic action and molecular basis of resistance. *Oncogene*, 22(47), 7265-79.
- Siveen, K. S., Mustafa, N., Li, F., Kannaiyan, R., Ahn, K. S., Kumar, A. P., Chng, W. J., & Sethi, G. (2014). Thymoquinone overcomes chemoresistance and enhances the anticancer effects of bortezomib through abrogation of NF-κB regulated gene products in multiple myeloma xenograft mouse model. *Oncotarget*, 5(3), 634–648.
- Sørlie, T., Perou, C. M., Tibshirani, R., Aas, T., Geisler, S., Johnsen, H., & Børresen-Dale, A. L. (2001). Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proceedings of the National Academy of Sciences of the United States of America, 98(19), 10869–10874.
- Sparano, J. A., Wang, M., Martino, S., Jones, V., Perez, E. A., Saphner, T., & Davidson, N. E. (2008). Weekly paclitaxel in the adjuvant treatment of breast cancer. *The New England journal of medicine*, 358(16), 1663–1671.
- Sullivan, K. D., Galbraith, M. D., Andrysik, Z., & Espinosa, J. M. (2018). Mechanisms of transcriptional regulation by p53. *Cell death and differentiation*, 25(1), 133–143.
- Štarha, P., Trávníček, Z., Popa, I., & Dvořák, Z. (2014). Synthesis, characterization and in vitro antitumor activity of platinum(II) oxalato complexes involving 7azaindole derivatives as coligands. *Molecules (Basel, Switzerland)*, 19(8), 10832–10844.
- Sun, C.Y., Zhang, Q.Y., Zheng, G.J., & Feng, B. (2019). Phytochemicals: Current strategy to sensitize cancer cells to cisplatin. *Biomedicine & Pharmacotherapy*, 110: 518-527.
- Syntichaki, P., & Tavernarakis, N. (2002). Death by necrosis. Uncontrollable catastrophe, or is there order behind the chaos?. EMBO reports, 3(7), 604–609.
- Tallarida, R.J. (2001). Drug synergism: its detection and applications. *J Pharmacol Exp Ther*, 298(3), 865-72.
- Tanida, S., Mizoshita, T., Ozeki, K., Tsukamoto, H., Kamiya, T., Kataoka, H., Sakamuro, D., & Joh, T. (2012). Mechanisms of Cisplatin-Induced Apoptosis and of Cisplatin Sensitivity: Potential of BIN1 to Act as a Potent Predictor of Cisplatin Sensitivity in Gastric Cancer Treatment. *International journal of* surgical oncology, 862879.

- Tan, G.H., Taib, N.A., Choo, W.Y., Teo, S.H., & Yip, C.H. (2009). Clinical characteristics of triple-negative breast cancer: experience in an Asian developing country. *Asian Pac J Cancer Prev*, 10(3): 395-8
- Tan, B. L., & Norhaizan, M. E. (2019). Curcumin Combination Chemotherapy: The Implication and Efficacy in Cancer. *Molecules (Basel, Switzerland)*, 24(14), 2527.
- Tang, D., Kang, R., Berghe, T. V., Vandenabeele, P., & Kroemer, G. (2019). The molecular machinery of regulated cell death. *Cell research*, 29(5), 347–364.
- Thomadaki, H., & Scorilas, A. (2007). Breast cancer cells response to the antineoplastic agents cisplatin, carboplatin, and doxorubicin at the mRNA expression levels of distinct apoptosis-related genes, including the new member, BCL2L12. *Annals of the New York Academy of Sciences*, 1095, 35–44.
- Toné, S., Sugimoto, K., Tanda, K., Suda, T., Uehira, K., Kanouchi, H., Samejima, K., Minatogawa, Y., & Earnshaw, W. C. (2007). Three distinct stages of apoptotic nuclear condensation revealed by time-lapse imaging, biochemical and electron microscopy analysis of cell-free apoptosis. *Experimental cell research*, 313(16), 3635–3644. https://doi.org/10.1016/j.yexcr.2007.06.018
- Torres, M.P., Ponnusamy, M.P., Chakraborty, S., Smith, L.M., Das, S., Arafat, H.A., & Batra, S.K. (2010). Effects of thymoquinone in the expression of mucin 4 in pancreatic cancer cells: implications for the development of novel cancer therapies. *Mol Cancer Ther.*, 9:1419–1431.
- Torrey, H., Butterworth, J., Mera, T., Okubo, Y., Wang, L., Baum, D., Defusco, A., Plager, S., Warden, S., Huang, D., Vanamee, E., Foster, R., & Faustman, D. L. (2017). Targeting TNFR2 with antagonistic antibodies inhibits proliferation of ovarian cancer cells and tumor-associated Tregs. *Science signaling*, 10(462), eaaf8608.
- Toyozumi, Y., Arima, N., Izumaru, S., Kato, S., Morimatsu, M., & Nakashima, T. (2004). Loss of caspase-8 activation pathway is a possible mechanism for CDDP resistance in human laryngeal squamous cell carcinoma, HEp-2 cells. *International journal of oncology*, 25(3), 721–728.
- Tsang, R.Y., Al-Fayea, T. & Au, HJ. (2009). Cisplatin Overdose, Toxicities and Management *Drug-Safety*, 32(12), 1109–1122.

- Tsuruya, K., Ninomiya, T., Tokumoto, M., Hirakawa, M., Masutani, K., Taniguchi, M., Fukuda, K., Kanai, H., Kishihara, K., Hirakata, H., & Iida, M. (2003).
 Direct involvement of the receptor-mediated apoptotic pathways in cisplatin-induced renal tubular cell death. *Kidney international*, 63(1), 72–82.
- Turner, N., Tutt, A., & Ashworth, A. (2004). Hallmarks of 'BRCAness' in sporadic cancers, *Nat Rev Cancer*, 4(10): 814-819.
- Turashvili, G., & Brogi, E. (2017). Tumor Heterogeneity in Breast Cancer. *Frontiers in medicine*, 4, 227.
- Ulu R., Dogukan A., Tuzcu M., Gencoglu H., Ulas M., Ilhan N., Muqbil I., Mohammad R.M., Kucuk O., & Sahin K. (2012). Regulation of renal organic anion and cation transporters by thymoquinone in cisplatin induced kidney injury. Food Chem. *Toxicol*, 50:1675–1679.
- US NIH. (n.d). ClinicalTrials.gov. https://clinicaltrials.gov/. Accessed 1 January 2020.
- Viale, G. (2012). The current state of breast cancer classification. *Ann. Oncol*, 23, x207–x210.
- van Tonder, A., Joubert, A. M., & Cromarty, A. D. (2015). Limitations of the 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay when compared to three commonly used cell enumeration assays. *BMC research notes*, 8, 47.
- Vicar, T., Raudenska, M., Gumulec, J., & Balvan, J. (2020). The Quantitative-Phase Dynamics of Apoptosis and Lytic Cell Death. Scientific reports, 10(1), 1566.
- Walker, N.I., Harmon, B.V., Gobe, G.C. & Kerr, J.F. (1988) Patterns of cell death. Methods Achiev. Exp. Pathol., 13, 18–54.
- Wang, Q., Yu, X., Li, F., Lv, X., Fu, X., Gu, H., & Zhang, B. (2019). Efficacy of celastrol combined with cisplatin in enhancing the apoptosis of U-2OS osteosarcoma cells via the mitochondrial and endoplasmic reticulum pathways of apoptosis. *Oncology Letters*, 17, 3305-3313.
- Wang, S., Xie, J., Li, J., Liu, F., Wu, X., & Wang, Z. (2016). Cisplatin suppresses the growth and proliferation of breast and cervical cancer cell lines by inhibiting integrin β5-mediated glycolysis. *American journal of cancer research*, 6(5), 1108–1117.
- Wahba, H. A., & El-Hadaad, H. A. (2015). Current approaches in treatment of triplenegative breast cancer. *Cancer biology & medicine*, 12(2), 106–116.

- Wawruszak, A., Luszczki, J. J., Grabarska, A., Gumbarewicz, E., Dmoszynska-Graniczka, M., Polberg, K., & Stepulak, A. (2015). Assessment of Interactions between Cisplatin and Two Histone Deacetylase Inhibitors in MCF7, T47D and MDA-MB-231 Human Breast Cancer Cell Lines - An Isobolographic Analysis. *PloS one*, 10(11), e0143013.
- Wen, C. J., Wu, L. X., Fu, L. J., Yu, J., Zhang, Y. W., Zhang, X., & Zhou, H. H. (2013). Genomic screening for targets regulated by berberine in breast cancer cells. Asian Pacific journal of cancer prevention : *APJCP*, 14(10), 6089– 6094.
- Williams, S., Tucci, M. A., & Benghuzzi, H. A. (2014). The effect of combination treatments of epigallocatechin-3-gallate, thymoquinone, and 5-Fluorouracil on fadu nasopharyngeal carcinoma cells. *Biomedical sciences instrumentation*, 50, 361–366.
- Wilson, A. J., Saskowski, J., Barham, W., Yull, F., & Khabele, D. (2015).
 Thymoquinone enhances cisplatin-response through direct tumor effects in a syngeneic mouse model of ovarian cancer. *Journal of ovarian research*, 8, 46.
- Wong R. S. (2011). Apoptosis in cancer: from pathogenesis to treatment. *Journal of experimental & clinical cancer research* : *CR*, 30(1), 87.
- Woo, C.C., Kumar, A.P., Sethi, G., & Tan, K.H. (2012). Thymoquinone: potential cure for inflammatory disorders and cancer. *Biochem Pharmacol*, 83(4):443-51.
- Woo, C.C., Loo, S.Y., Gee, V., Yap, C.W., Sethi, G., Kumar, A.P., & Tan, K.H.B. (2011). Anticancer activity of thymoquinone in breast cancer cells: Possible involvement of PPAR-γ pathway, *Biochemical Pharmacology*, 82(5), 464-475.
- Wu, W. S., Heinrichs, S., Xu, D., Garrison, S. P., Zambetti, G. P., Adams, J. M., & Look, A. T. (2005). Slug antagonizes p53-mediated apoptosis of hematopoietic progenitors by repressing puma. *Cell*, 123(4), 641–653.
- Wu, Y., Tibrewal, N., & Birge, R. B. (2006). Phosphatidylserine recognition by phagocytes: a view to a kill. *Trends in cell biology*, 16(4), 189–197.
- Wu, Y. M., Zhang, K. J., Yue, X. T., Wang, Y. Q., Yang, Y., Li, G. C., Li, N., & Wang, Y. G. (2009). Enhancement of tumor cell death by combining cisplatin

with an oncolytic adenovirus carrying MDA-7/IL-24. *Acta pharmacologica Sinica*, 30(4), 467–477.

- Wu, W., Wang, H.D., Guo, W., Yang, K., Zao, Y.P., Jiang, Y.G. & He, P. (2010). Up-regulation of Fas reverses cisplatin resistance of human small cell lung cancer cells. *Journal of Experimental & Clinical Cancer Research*; 29, 49.
- Xie, Y., Gou, Q., Wang, Q., Zhong, X., & Zheng, H. (2017). The role of BRCA status on prognosis in patients with triple-negative breast cancer. *Oncotarget*, 8(50), 87151-87162.
- Xie, X., He, G., & Siddik, Z.H. (2017). Functional Activation of Mutant p53 by Platinum Analogues in Cisplatin-Resistant Cells Is Dependent on Phosphorylation. *Mol Cancer Res*, 15 (3), 328-339.
- Yao, H., He, G., Yan, S., Chen, C., Song, L., Rosol, T. J., & Deng, X. (2017). Triplenegative breast cancer: is there a treatment on the horizon?. *Oncotarget*, 8(1), 1913–1924.
- Yang, Z., Schumaker, L.M., Egorin, M.J., Zuhowski, E.G., & Guo, Z. (2006). Cisplatin Preferentially Binds Mitochondrial DNA and Voltage-Dependent Anion Channel Protein in the Mitochondrial Membrane of Head and Neck Squamous Cell Carcinoma: Possible Role in Apoptosis. *Clin Cancer Res* 12: 5817-5825.
- Yazan LS. (2009). Cytotoxicity of thymoquinone (TQ) from Nigella sativa towards human cervical carcinoma cells (HeLa) *Journal of Pharmacy Research*, 2(4).
- Yeh, J., Chun, J., & Schwartz, S. (2017). Clinical Characteristics in Patients with Triple Negative Breast Cancer, *International Journal of Breast Cancer*, vol. 2017, Article ID 1796145, 5 pages.
- Yen, H.C., Tang, Y.C., Chen, F.Y., Chen, S.W., & Majima, H.J. (2005). Enhancement of cisplatin-induced apoptosis and caspase 3 activation by depletion of mitochondrial DNA in a human osteosarcoma cell line. *Ann N Y Acad Sci.* 1042: 516–522.
- Yildirim, I.H., Azzawri, A.A., & Duran, T. (2019). Thymoquinone induces apoptosis via targeting the Bax/BAD and Bcl-2 pathway in breast cancer cells. *Dicle Med J*, 46 (3), 411 – 417.
- Yimit, A., Adebali, O., Sancar, A., & Jiang, Y. (2019). Differential damage and repair of DNA-adducts induced by anti-cancer drug cisplatin across mouse organs. *Nat Commun* 10, 309.

- Yin, Z., Deng, Z., Zhao, W., & Cao, Z. (2018a). Searching Synergistic Dose Combinations for Anticancer Drugs. *Frontiers in pharmacology*, 9, 535.
- Yin, L. L., Wen, X. M., Lai, Q. H., Li, J., & Wang, X. W. (2018b). Lenalidomide improvement of cisplatin antitumor efficacy on triple-negative breast cancer cells in vitro. *Oncology letters*, 15(5), 6469–6474.
- Yip, C.H., Pathy, N.B., & SH Teo, S.H. (2014). A Review of Breast Cancer Research in Malaysia. *Med J Malaysia*, 69, 8-22.
- Yimer, E. M., Tuem, K. B., Karim, A., Ur-Rehman, N., & Anwar, F. (2019). Nigella sativa L. (Black Cumin): A Promising Natural Remedy for Wide Range of Illnesses. Evidence-based complementary and alternative medicine: eCAM.
- Yazdi, H.S, Noorbakhsh, M.F., Hayati, F., Samarghandian, S., & Farkhondeh, T. (2018). Immunomodulatory and Anti-inflammatory Effects of Thymoquinone. *Cardiovascular & Hematological Disorders-Drug Targets*, 18(1), 52-60.
- Younus H. (2018) Antidiabetic Action of Thymoquinone. In: Younus H. (eds) Molecular and Therapeutic actions of Thymoquinone, p 7-17.
- Zhang, L. N., Li, J. Y., & Xu, W. (2013). A review of the role of Puma, Noxa and Bim in the tumorigenesis, therapy and drug resistance of chronic lymphocytic leukemia. *Cancer gene therapy*, 20(1), 1–7.
- Zhang, X. Z., Wang, L., Liu, D. W., Tang, G. Y., & Zhang, H. Y. (2014). Synergistic inhibitory effect of berberine and d-limonene on human gastric carcinoma cell line MGC803. *Journal of medicinal food*, 17(9), 955–962.
- Zhang, L., Bai, Y., & Yang, Y. (2016). Thymoquinone chemosensitizes colon cancer cells through inhibition of NF-κB. Oncology Letters, 12(4), 2840–2845.
- Zhang, M., Zheng, J., Nussinov, R., & Ma, B. (2017). Release of Cytochrome C from Bax Pores at the Mitochondrial Membrane. *Scientific reports*, 7(1), 2635.
- Zhang, Y., Wu, J., Ye, M., Wang, B., Sheng, J., Shi, B., & Chen, H. (2018). ETS1 is associated with cisplatin resistance through IKKα/NF-κB pathway in cell line MDA-MB-231. *Cancer cell international*, 18, 86.
- Zhang, H., Shao, F., Guo, W., Gao, Y. and He, J. (2019), Knockdown of KLF5 promotes cisplatin-induced cell apoptosis via regulating DNA damage checkpoint proteins in non-small cell lung cancer. *Thorac Cancer*, 10: 1069-1077.

- Zhang, J., & Xie, T. (2020). Ghrelin inhibits cisplatin-induced MDA-MB-231 breast cancer cell apoptosis via PI3K/Akt/mTOR signaling. *Experimental and therapeutic medicine*, 19(3), 1633–1640.
- Zhu WQ, Wang J, Guo XF, Liu Z, & Dong WG. (2016). Thymoquinone inhibits proliferation in gastric cancer via the STAT3 pathway in vivo and in vitro. *World J Gastroenterol*, 22, 4149–4159.

LIST OF PUBLICATION

Non-Indexed Conference Proceeding

 Zolkemri, Z., Kabi, K., Baba, S., & Hamdan, S. (2018). Optimization of spheroid formation of HeLa cells using combination of hanging drop method and 96-well plate coated agarose. In 2018 *The 7th International Graduate Conference on Engineering, Science & Humanities.* (pp 491-493). ISBN: 978-967-2171-27-0