CHIRAL SEPARATION OF KETOCONAZOLE AND ITRACONAZOLE ANTI-FUNGAL DRUGS USING EXPERIMENTAL AND COMPUTATIONAL APPROACHES

SITI ROSILAH BINTI ARSAD

UNIVERSITI TEKNOLOGI MALAYSIA

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SITI ROSILAH BINTI ARSAD

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DEDICATION

In the loving memory of my late father, Arsad Dolah, my strength and my biggest influence in my life. My mother Rafeah Deraman, and my husband Ibrahim Bidin who is always supportive and patiently waiting for me to complete this thesis. I dedicate this thesis to all of you.

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"In the name of Allah, the Most Gracious and the Most Merciful"

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ABSTRACT

Drugs with multiple chiral centers were observed as very effective for treating various diseases. However, the enantiomeric resolution of multiple chiral center racemates is not much developed compared to racemates having a single asymmetric center. This work aimed to develop a chiral separation method for antifungal drugs using electrokinetic chromatography (EKC) and to elucidate mechanism of enantioseparation using a computer-aided molecular modelling study. Two azole antifungal drugs were selected namely ketoconazole and itraconazole, which consists of two and three chiral centers, respectively. The separation for ketoconazole was achieved using heptakis (2,3,6-tri-O-methyl)-β-cyclodextrin (TM- β -CD), a commonly used chiral selector, as it is relatively inexpensive and has a low UV absorbance in addition to an anionic surfactant, sodium dodecyl sulfate. The optimum conditions for chiral separation of ketoconazole was achieved using 10 mM phosphate buffer at pH 2.50 containing 20 mM TM-β-CD, 5 mM SDS, and 1.0% (v/v) methanol with an applied voltage of 25 kV at 25°C with a 5-s hydrodynamic injection time at 50 mbar. The four ketoconazole stereoisomers were successfully resolved within 17 min (total analysis time was 28 min including capillary conditioning). The migration time precision of this method was examined to give a repeatability and reproducibility with RSDs \leq 5.80% (n = 3) and RSDs \leq 8.88% (n = 9), respectively. A computational study, using quantum mechanics calculations with AutoDock and semi-empirical PM3 calculations, were used to predict the enantiodiscrimination of ketoconazole enantiomers. A Density Functional Theory (DFT) single-point calculation at the level of B3LYP/6-311G (d,p) was performed for the PM3-optimized complexes to obtain more accurate binding energy and also electronic structures of the complexes. Molecular docking and DFT were simulated to predict the enantioresolution of itraconazole with two types of cyclodextrins (CDs), TM-β-CD and (2-hydroxylpropyl)-γ-cyclodextrin (HP-\gamma-CD). The difference in energies of the inclusion complexes between the enantiomers and CD is a measure of chiral discrimination, which results in the separation of the enantiomers in the experimental studies. The dual-CD and triple-CD methods were developed for chiral separation of itraconazole using EKC. Highly sulfated β-cyclodextrin (S-β-CD), (2-hydroxylpropyl)-β-cyclodextrin (HP-β-CD), TM- β -CD and HP- γ -CD were screened as possible chiral selectors for enantioseparation of itraconazole. The enantioseparation of itraconazole was achieved using 10 mM phosphate buffer solution at pH 3.62 containing a mixture of 10 mM of each HP- β -CD, TM- β -CD and HP- γ -CD and an applied voltage of 25 kV at 25°C. Both computational and experimental investigations complement each other prior to chiral recognition mechanism. Combination of molecular modelling and capillary electrophoresis appears as a new emerging method for chiral analysis of pharmaceutical drugs.

ABSTRAK

Dadah dengan beberapa pusat kiral telah diamati sangat berkesan untuk merawat pelbagai penyakit. Walau bagaimanapun, resolusi enantiomer rasemat dengan pusat kiral berganda tidak banyak dibangunkan berbanding dengan rasemat yang mempunyai pusat asimetri tunggal. Kajian ini bertujuan untuk membangunkan kaedah pemisahan kiral bagi dadah antikulat menggunakan kromatografi elektrokinetik (EKC) dan menentukan mekanisme enantiopemisahan menggunakan kajian pemodelan molekul berbantukan komputer. Dua dadah antikulat azol telah dipilih iaitu ketokonazol dan itrakonazol, masing-masing mempunyai dua dan tiga pusat kiral. Pemisahan ketokonazol berjaya diperoleh menggunakan (2,3,6-tri-Ometil)-β-siklodekstrin (TM-β-CD), satu pemisah kiral yang biasa digunakan kerana ia relatif tidak mahal dan mempunyai keserapan UV yang rendah sebagai tambahan kepada surfaktan anion, natrium dodesil sulfat (SDS). Keadaan optimum bagi pemisahan kiral ketokonazol telah diperoleh menggunakan larutan penimbal fosfat 10 mM pada pH 2.50 yang mengandungi 20 mM TM-β-CD, 5 mM SDS, dan 1.0% (v/v) metanol pada voltan gunaan 25 kV pada 25°C dengan masa suntikan hidrodinamik 5-s pada 50 mbar. Empat stereoisomer ketokonazol berkenaan telah berjaya dipisahkan sepenuhnya dalam masa 17 min (jumlah masa analisis ialah 28 min termasuk pengkondisian kapilari). Kepresisan masa migrasi kaedah ini telah dikaji untuk memberi keterulangan dan kebolehulangan dengan masing-masing RSDs \leq 5.80% (n = 3) dan RSDs \leq 8.88% (n = 9). Satu kajian komputeran menggunakan pengiraan mekanik kuantum dengan AutoDock dan pengiraan semiempirik PM3 telah digunakan untuk meramalkan enantiodiskriminasi ketokonazol. Pengiraan titik tunggal Teori Fungsi Ketumpatan (DFT) pada tahap B3LYP/6-311G (d,p) telah dilakukan daripada kompleks PM3 yang dioptimumkan untuk mendapatkan tenaga ikatan dan struktur elektronik kompleks tersebut yang lebih tepat. Dok molekul dan DFT telah disimulasikan untuk meramalkan enantioresolusi itrakonazol dengan menggunakan dua jenis siklodekstrin (CD), TM-β-CD dan (2hidroksilpropil)-y-siklodekstrin (HP-y-CD). Perbezaan tenaga kompleks rangkuman antara enantiomer dan CD adalah ukuran diskriminasi kiral, yang menghasilkan pemisahan enantiomer dalam kajian eksperimen. Kaedah dwi-CD dan tri-CD telah dibangunkan untuk pemisahan kiral itrakonazol menggunakan EKC. β-siklodekstrin yang sangat tersulfat (S-β-CD), (2-hidroksilpropil)-β-siklodekstrin (HP-β-CD), TMβ-CD dan HP-γ-CD telah disaringkan sebagai pemilih kiral yang mungkin untuk pemisahan enantiomer itrakonazol. Pemisahan enantiomer itraconazol telah dicapai menggunakan larutan penimbal fosfat 10 mM pada pH 3.62 mengandungi campuran HP-β-CD, TM-β-CD dan HP-γ-CD, setiap satu 10 mM dan voltan gunaan 25 kV pada 25°C. Kedua-dua penyiasatan komputeran dan eksperimen saling melengkapkan antara satu sama lain sebelum mekanisme pengecaman kiral. Gabungan pemodelan molekul dan elektroforesis kapilari muncul sebagai kaedah baharu bagi analisis kiral dadah farmaseutikal.

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LIST OF ABBREVIATIONS

| BGE | - | Background electrolyte | | |
|---------|---|---|--|--|
| CD | - | Cyclodextrin | | |
| TM-β-CD | - | Heptakis-(2,3,6-tri-O-methyl)-β-cyclodextrin | | |
| HP-β-CD | - | Hydroxylpropyl-β-cyclodextrin | | |
| HP-γ-CD | - | Hydroxylpropyl-y-cyclodextrin | | |
| CD-EKC | - | Cyclodextrin-modified electrokinetic chromatography | | |
| CE | - | Capillary electrophoresis | | |
| CMC | - | Critical micelle concentration | | |
| CZE | - | Capillary zone electrophoresis | | |
| DAD | - | Diode-array detection | | |
| DFT | - | Density functional theory | | |
| EKC | - | Electrokinetic chromatography | | |
| EOF | - | Electroosmotic flow | | |
| GC | - | Gas chromatography | | |
| HPLC | - | High-performance liquid chromatography | | |
| MEKC | - | Micellar electrokinetic chromatography | | |
| PM3 | - | Parameterized model number 3 | | |
| SDS | - | Sodium dodecyl sulphate | | |
| SFC | - | Supercritical fluid chromatography | | |
| SPE | - | Solid-phase extraction | | |
| UV | - | Ultraviolet | | |

LIST OF SYMBOLS

| Å | - | Armstrong |
|----------------|---|------------------------------|
| α | - | Alpha |
| β | - | Beta |
| °C | - | Degree Celsius |
| Δ | - | Delta |
| γ | - | Gamma |
| μΑ | - | Micro ampere |
| μL | - | Micro litre |
| g | - | Gram |
| g/mol | - | Gram per mole |
| i.d. | - | Inner diameter |
| kV | - | Kilovolts |
| mg | - | Milligram |
| min | - | Minutes |
| µg/mL | - | Microgram per millilitre |
| mL/min | - | Millilitre per minute |
| mmol | - | Millimole |
| рКа | - | Acid dissociation constant |
| ppm | - | Part per million |
| R _s | - | Peak resolution |
| r^2 | - | Coefficient of determination |
| rpm | - | Rotation per minute |
| S | - | Second |
| t _m | - | Migration time |
| v/v | - | Volume per volume |
| | | |

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CHAPTER 1

INTRODUCTION

1.1 Research Background

Over the last 50 years drug stereochemistry has become a significant issue for both the pharmaceutical industry and the regulatory authorities since the problems associated with drug stereochemistry are complex. Many pharmaceutical drugs are known to be a racemic mixture. Such mixtures are regarded by some as 'compounds containing 50% impurity' which sometimes can cause toxic effects to the patients (Hutt and O'Grady, 1996). Since then, most of the top selling drug companies around the world are administered as single enantiomers that is worth of the desired physiological activity. Thus the development of methods for the production of enantiomerically pure compounds and the assessment of their enantiomeric purity have become more and more important specially in life science applications, such as biochemical, toxicological, forensic and pharmaceutical research. The development of single-enantiomer drugs is preferred because of the reduced risk of side effects.

In the last two decades, the incidence of serious fungal infections has grown dramatically due to the increase of risk groups: the advent of human immunodeficiency virus (HIV) or undergoing anticancer chemotherapy or the increased use of immunosuppressive therapies in organ transplantation (Thienpont *et al.*, 1999; Crego *et al.*, 2001; Castro-Puyana *et al.*, 2005). Amphotericin B (Fungizone) is a conventional topical antifungal drug was used to treat fungal infections. However, this drug is associated with significant toxicity, including infusion-related events, such as chills, fever, headache, nausea and vomiting (Saag and Dismukes, 1988). The availability of the azole antifungal agents represents a major advancement in the management of systemic fungal infections as they present low toxicity. Ketoconazole and itraconazole antifungal drugs have become frequently used as alternatives to Amphotericin B since they have versatility of

administration and a broad spectrum (Rotstein *et al.*, 1992; Ahmed *et al.*, 1998; Dilmaghanian *et al.*, 2004). However, since these two azoles are chiral compounds, they also are not free from adverse side effects due to different properties of stereoisomers. Therefore, it is important to promote the chiral separation for chiral ketoconazole and itraconazole antifungal drugs in order to eliminate the unwanted isomer. The structures of these two azoles used in the study use are shown in Figure 1.1. Ketoconazole and itraconazole consists of multiple chiral centers with two and three chiral centers, respectively.

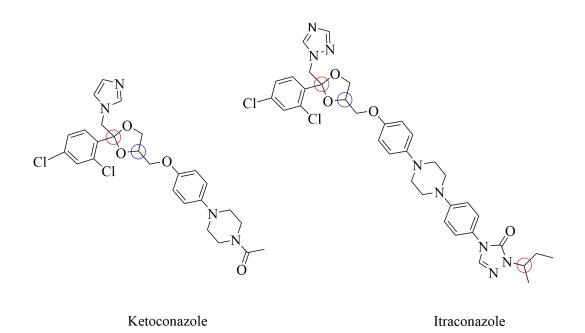


Figure 1.1 Structures of antifungal azole compounds, ketoconazole and itraconazole

Enantiomeric resolution becomes more challenging when dealing with multichiral center racemates since it is very difficult to find the identical properties of the enantiomers (Ali *et al.*, 2016). However, this challenge may be tackled by a versatile capillary electrophoresis (CE) technique which has rapidly attracted attention as a promising technique for enantioseparation due to its high separation efficiency and flexibility (Terabe and Otsuka 1994; Nishi and Terabe 1995). One of the most attractive advantages of CE for the separation of enantiomers is easy changes of separation media in the method development, that is, one can easily alter

the separation solution to find the optimum separation media and one can also use an expensive chiral selector because of the small amounts of media required (Nishi, 1996). Chiral separation using CE also do not require a chiral stationary phase (CSP) like in HPLC since the chiral selectors can be directly added into the background electrolyte (BGE) to provide a chiral environment and form enantiomer-chiral selector complexes with analytes (Li *et al.*, 2015a).

Mechanistic aspect of enantioseparation becomes interesting since it may provide valuable information such as prediction of elution order, appropriate chromatographic conditions and types of analytes separable with a given selector (Maier *et al.*, 2001). Additionally, it is desirable to obtain the structural models that explain the binding forces in chiral recognition to understand qualitatively the mechanism of enantioseparation. Computational investigations concerning chiral recognition of ketoconazole and itraconazole with cyclodextrins (CDs) have been performed to get further insight into the mechanism involved.

Molecular modelling might be used as a supportive tool to enhance our understanding of chiral recognition. Molecular docking approaches proved to be very useful for the evaluation of chiral recognition systems. Molecular docking is aimed at explaining possible chiral recognition mechanism between the selected antifungal drugs and CDs as chiral selector. Furthermore, the computational tools employed in molecular modelling such as quantum mechanics and molecular mechanics have reached high level of sophistication allowing prediction of intermolecular binding scenarios between molecules (Maier *et al.*, 2001).

1.2 Problem Statement

Currently, the separation of the enantiomers of a chiral compound with a single center of chirality is no more the issue since tremendous researches have shown an excellent separation in separating compounds with a single chiral center. However, the separation of enantiomers with multiple chiral centers becomes more challenging since it is very difficult to choose suitable chiral selectors that have the ability to differentiate several chiral centers simultaneously.

A variety of chiral selectors are available for chiral separation such as crown ethers, CDs, macrocyclic antibiotic and chiral surfactant. Among these chiral selectors, CDs offer great advantages in chiral separation as they can differ in selectivity of the enantiomers. Besides, CDs are commercially available at a cheaper price compared to macrocyclic antibiotic such as vancomycin. The application of CDs as chiral selector in CE has made CE a feasible technique for the separation of a large number of chiral compounds including azole antifungal drugs.

Previously, studies on ketoconazole (Castro-Puyana *et al.*, 2005) and itraconazole (Castro-Puyana *et al.*, 2006) antifungal drugs with multiple chiral centers were performed with single CDs using CE. However, only half of the stereoisomers were successfully resolved. Therefore, in order to improved and enhanced the resolution of stereoisomers of ketoconazole and itraconazole drugs, modifications in BGE solution was performed such as addition of surfactant and the use of dual and triple CDs.

Even though CE can offers a great advantage that is easy to choose a number of chiral selectors for chiral separation, however, the screening of suitable chiral selectors is a time consuming (Jimidar *et al.*, 2004). Furthermore, the lack of fundamental understanding of the chiral recognition mechanisms draw researchers' attention to employ computational techniques in chiral separation. Currently, tremendous articles were published on computational studies concerning chiral recognition mechanism. However, only one article was published on computational study of chiral recognition mechanism of ketoconazole with β -cyclodextrin (β -CD)

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as chiral selector (Redenti *et al.*, 1999), and the study of chiral recognition mechanism of itraconazole has not been reported so far. Hence, it is our interest to perform computational studies on these two antifungal drugs so that we can get further insight into the chiral interactions and mechanism in the enantioseparation process.

1.3 Aims and Objectives

The aim of the study is to develop computational and experimental studies for chiral separation of ketoconazole and itraconazole antifungal drugs using molecular modelling and electrokinetic chromatography (EKC), respectively. The interaction and possible chiral recognition mechanism of ketoconazole and itraconazole antifungal drugs with the CDs chiral selector will be elucidated using computational approach via molecular docking studies. The objectives of this study are to:

- 1. optimize separation of chiral ketoconazole and itraconazole antifungal drugs using CDs as chiral selector in EKC.
- 2. elucidate mechanism of ketoconazole antifungal drug enantioseparation using computer-aided molecular modelling study.
- 3. screen and predict potentials of dual and triple CD system for enantioseparation of itraconazole antifungal drug using molecular modelling technique, then testing the results using experimental approach.

1.4 Scope of the Research

The focus of the research is to separate two azole chiral drugs, ketoconazole and itraconazole using EKC due to its high efficiency, short analysis time and wide application range. EKC is known as a suitable and simple method for separation of either acidic or basic drugs. The mechanism of enantioseparation was elucidated using a computer-aided molecular modelling study. The interaction of enantiomers with CD as chiral selector and their binding energies were studied for the chiral recognition mechanism. Separation of four and eight stereoisomers of ketoconazole and itraconazole, respectively still has not been achieved by far using EKC technique. Therefore, studies on these two antifungal drugs should be interesting for the purpose of enantioseparation mechanism.

However, it is very important to note that the results derived from molecular modelling generally do not account for the presence of modifiers, ions, effects of differential solvation of the diastereomeric complexes and the underlying support (Maier *et al.*, 2001). Therefore, there is still limited knowledge on chiral discrimination processes and phenomena when comparing experimental enantioselectivity data with the results of molecular modelling.

1.5 Significance of Research

Chirality is a major concern in the modern pharmaceutical industry and still a scientific challenge especially to predict successful baseline separations of chiral compounds. Hence there is a great need to develop suitable method for analysis and separation of chiral compounds especially for compounds with multiple chiral centers. In this study, multiple chiral centers ketoconazole and itraconazole antifungal drugs were successfully separated and developed using EKC method with multiple CDs. For the first time we have succeeded in the enantioseparation of four stereoisomers of ketoconazole using EKC with addition of small amount of anionic surfactant, sodium dodecyl sulphate (SDS) and addition of three different type of

CDs simultaneously in the CE BGE. In addition, the use of dual and triple CDs shows significant results in obtaining a complete separation of itraconazole stereoisomers.

This study also contributed to the combination of experimental and computational studies using molecular modelling approaches to obtain further insight into formation of complexes of chiral compounds with CDs as chiral selectors. Molecular docking was successfully performed using Autodock 4.2 software which minimized the cost as the usage of certain software are quite expensive. To the best of our knowledge, the use of dual CDs in modelling the host-guest system was also the first to be performed using Autodock software.

1.6 Outline of the Thesis

This thesis consists of seven chapters. Chapter 1 describes the research background, problem statement, objectives, scope as well as significance of the study.

Chapter 2 reports the literature search related to stereoisomers of chiral compounds, the importance of chiral separation, introduction to azole antifungal drug, review on previous enantioseparation of ketoconazole and itraconazole drugs, capillary electrophoresis as a chiral separation technique, cyclodextrin as a chiral selector and last but not least the trends in molecular modelling studies related to chiral analysis.

Chapter 3 explores the enantioseparation of ketoconazole using CD-EKC. This chapter reports on the optimization of several parameters of CD-EKC system including concentration of heptakis (2,3,6-tri-O-methyl)- β -cyclodextrin (TM- β -CD) used, concentration of sodium dodecyl sulphate (SDS), pH effect, buffer phosphate concentration, effect of separation temperature, effect applied voltage, and effect of addition organic modifiers. Method validation is also reported in this chapter which

discussed on linearity, precision and limit of detection (LOD) of the method developed. Furthermore, the method developed was applied to real sample analysis in urine and cream formulation using solid phase extraction (SPE) as a sample pretreatment.

Chapter 4 describes the computational investigations on chiral recognition of ketoconazole and TM- β -CD. Molecular docking was performed to investigate the mechanism of chiral recognition of ketoconazole and TM- β -CD using MS Modelling software and Autodock software. In this chapter, four stereoisomers of ketoconazole was placed into CD cavity manually to study the inclusion mechanism and obtain the binding energy of the complexes using MS Modelling software. The binding energy of the complexes using PM3 with Gaussian 09 software. Next, molecular docking of ketoconazole with TM- β -CD was performed using Autodock software. Further calculations of binding energies from molecular docking with Autodock were performed with Gaussian 09 software using PM3 and DFT method. Results from molecular docking showed that the magnitude of binding energy difference which indicates the enantiodiscrimination of the separation. The interactions involved in the formation of complexes are also discussed in this chapter.

Chapter 5 explores the molecular docking of itraconazole drugs with two types of CDs. TM- β -CD and HP- γ -CD were selected as hosts to differentiate the ability of different size cavity in the formation of complexes. Difference of functional groups were also taken into accounts as the factors in the complexes formation. In this chapter, the formation of the complexes was explored using single macromolecule and dual macromolecules as host/s. The binding energy was calculated with Gaussian 09 using DFT method. The binding energy differences of complexes obtain from TM- β -CD and HP- γ -CD was also compared. Furthermore, the chapter discussed on chiral separation of itraconazole using CD-EKC. The discussion involved the screening process to find the suitable chiral selector using dual and triple CDs system. Native CDs namely TM- β -CD, hydroxylpropyl- γ -cyclodextrin (HP- γ -CD), and hydroxylpropyl- β -cyclodextrin (HP- β -CD) were used for screening. Charged CDs namely highly sulphated β -CD was also used for

screening purposes. This chapter also reports the optimization of several parameters of CD-EKC system including concentration of CDs used, pH effect, buffer phosphate concentration, effect of separation temperature, effect applied voltage, and effect of addition organic modifiers. The optimum separation achieved in experimental studies was compared with the results obtained in computational study.

Lastly, chapter 6 presents the overall conclusions and suggestion for further studies. This chapter summarizes the result obtained throughout the study such as the optimum conditions of the enantioseparation of ketoconazole and itraconazole. In addition, the chiral discrimination for both ketoconazole and itraconazole obtained from molecular docking are also summarized in this chapter. Suggestions for further studies are presented.

REFERENCES

- Abushoffa, A. M., Fillet, M., Marini, R. D., Hubert, P., and Crommen, J. (2003). Enantiomeric Separation of Aminoglutethimide by Capillary Electrophoresis Using Native Cyclodextrins in Single and Dual Systems. J. Sep. Sci., 26, 536-542.
- Abushoffa, A. M., Burjanadze, N., Blaschke, G., Crommen, J., and Chankvetadze, B. (2002). Comparative Study on the Enantioseparation of Glutethimide Using Dual Cyclodextrin Systems and Cyclodextrin Modified MEKC in Capillary Electrophoresis. J. Sep. Sci., 25, 10-16.
- Ahmed, M., El-Gibaly, I., and Ahmed, S. (1998). Effect of Cyclodextrins on the Physicochemical Properties and Antimycotic Activity of Clotrimazole. *Int. J. Pharm.*, 171, 111–121.
- Al Azzam, K.M., Saad, B., Adnan, R., and Aboul-Enein, H.Y. (2010).
 Enantioselective Analysis of Ofloxacin and Ornidazole in Pharmaceutical Formulations by Capillary Electrophoresis Using Single Chiral Selector and Computational Calculation of Their Inclusion Complexes. *Anal. Chim. Acta*, 674, 249–255.
- Al Azzam, K.M. (2015). Host-Guest Inclusion Complexes Between Amlodipine Enantiomers in Biphasic Recognition Chiral Extraction System Using Tartaric Acid and β-Cyclodextrin Derivatives as Positive Confirmation Using of Their Enantioselective Extraction. *Sci. Pharm.*, 83, 683–698.
- Al-Burtomani, S. K. S., and Suliman F. E. O. (2018). Experimental and Theoretical Study of the Inclusion Complexes of Epinephrine with β-Cyclodextrin, 18-Crown-6 and Cucurbit[7]uril. *New J. Chem.*, 42, 5785-5797.
- Ali, I., Suhail, M., Al Othman, Z. A., Alwarthan A. (2017). Chiral Separation and Modeling of Baclofen, Bupropion, and Etodolac Profens on Amylose Reversed Phase Chiral Column. *Chirality*, 29, 386-397.
- Ali, I., Suhail, M., AL-Othman, Z. A., Alwarthan, A. and Aboul-Enein, H. Y. (2016). Enantiomeric Resolution of Multiple Chiral Centres Racemates by Capillary Electrophoresis. *Biomed. Chromatogr.*, 30, 683-694.

- Ali, I., Aboul-Enein, H.Y., Gaitonde, V.D., Singh, P., Rawat, M.S.M., and Sharma, B. (2009). Chiral Separations of Imidazole Antifungal Drugs on AmyCoat RP Column in HPLC. *Chromatographia*, 70, 223–227.
- Ali, I., Kumerer, K., and Aboul-Enein, H.Y. (2006). Mechanistic Principles in Chiral Separations Using Liquid Chromatography and Capillary Electrophoresis. *Chromatographia*, 63, 295–307.
- Al Omari, A., Al Omari, M.M., Badwan, A., and Al-Sou'od, K. (2011). Effect of Cyclodextrins on the Solubility and Stability of Candesartan Cilexetil in Solution and Solid State. J. Pharm. Biomed. Anal., 54, 503–509.
- Al-Othman, Z., Al-Warthan, A., Alam, S.D., and Ali, I. (2014). Enantioseparation of Drugs with Multiple Chiral Centers by Chromatography and Capillary Electrophoresis. *Biomed. Chromatogr.*, 28, 1514–1524.
- Altria, K., Marsh, A., and Sänger-van de Griend, C. (2006). Capillary Electrophoresis for the Analysis of Small-Molecule Pharmaceuticals. *Electrophoresis*, 27, 2263–2282.
- Antczak, A., Ramstad, T., and Johnson, R. (2006). A CD-MEKC Method Utilizing a Neutral Surfactant for Enantiomeric Purity Determination of an Oxazolidinone Drug Candidate. *Chromatographia*, 64, 57–64.
- Bahri, M. A., Hoebeke, M., Grammenos, A., Delanaye, L., Vandewalle N. and Seret,
 A. (2006). Investigation of SDS, DTAB and CTAB Micelle Microviscosities
 by Electron Spin Resonance. *Colloid Surf. A.*, 290, 206-212.
- Barzegar, A., Moosavi-Movahedi, A., Mahnam, K., and Ashtiani, S.H. (2010). Chaperone-Like Activity of Alpha-Cyclodextrin via Hydrophobic Nanocavity to Protect Native Structure of ADH. *Carbohydr. Res.*, 345, 243–249.
- Becuwe, M., Landy, D., Delattre, F., Cazier, F., and Fourmentin, S. (2008). Fluorescent Indolizine-β-Cyclodextrin Derivatives for the Detection of Volatile Organic Compounds. *Sensors*, 8, 3689–3705.
- Bernal, J.L., del Nozal, M.J., Toribio, L., Montequi, M.I., and Nieto, E.M. (2000). Separation of Ketoconazole Enantiomers by Chiral Subcritical-Fluid Chromatography. J. Biochem. Bioph. Meth., 43, 241–250.
- Bernal, J.L., Toribio, L., del Nozal, M.J., Nieto, E.M., and Montequi, M.I. (2002). Separation of Antifungal Chiral Drugs by SFC and HPLC: A Comparative Study. J. Biochem. Bioph. Meth., 54, 245–254.

- Blanco, M. and Valverde, I. (2003). Choice of Chiral Selector for Enantioseparation by Capillary Electrophoresis. *TrAC-Trends Anal. Chem.*, 22, 428–439.
- Breadmore, M.C. and Thormann, W. (2003). Capillary Electrophoresis Evidence for the Stereoselective Metabolism of Itraconazole in Man. *Electrophoresis*, 24, 2588–2597.
- Burke, D. and Henderson, D. J. (2002). Chirality: A Blueprint for the Future. *Br. J. Anaesth.*, 88:4, 563-576.
- Cârcu-Dobrin, M., Budâu, M., Hancu, G., Gagyi, L., Rusu, A., Kelemen, H. (2016). Enantioselective Analysis of Fluoxetine in Pharmaceutical Formulations by Capillary Zone Electrophoresis. *Saudi Pharm. J.*, 25:3, 397-403.
- Cârje, A. G., Balint, A., Muntean, D. L., Hancu, G., Ion, V., and Imre, S. (2018). Comparative Enantioseparation of Amlodipine by HPLC and Capillary Electrophoresis. *Acta Med. Marisiensis*, 64:1, 28-33.
- Carvalho, P.O., Cass, Q.B., Calafatti, S. a., Contesini, F.J., and Bizaco, R. (2006). Review- Alternatives for the Separation of Drug Enantiomers: Ibuprofen as a Model Compound. *Braz. J. Chem. Eng.*, 23, 291–300.
- Castro-Puyana, M., Crego, A.L., Marina, M.L., and García-Ruiz, C. (2007). Enantioselective Separation of Azole Compounds by EKC. Reversal of Migration Order of Enantiomers with CD Concentration. *Electrophoresis*, 28, 2667–2674.
- Castro-Puyana, M., Crego, A.L., and Marina, M.L. (2006). Separation and Quantitation of the Four Stereoisomers of Itraconazole in Pharmaceutical Formulations by Electrokinetic Chromatography. *Electrophoresis*, 27, 887– 895.
- Castro-Puyana, M., Crego, A.L., and Marina, M.L. (2005). Enantiomeric Separation of Ketoconazole and Terconazole Antifungals by Electrokinetic Chromatography: Rapid Quantitative Analysis of Ketoconazole in Pharmaceutical Formulations. *Electrophoresis*, 26, 3960–3968.
- Chang, M. W., Ayeni, C., Breuer, S., Torbett, B. E. (2010). Virtual Screening for HIV Protease Inhibitors: A Comparison of AutoDock 4 and Vina. *Plos One*, 5:8, 11955.
- Chhabra, N., Aseri, M. L. and Padmanabhan, D. (2013). A Review of Drug Isomerism and Its Significance. *Int. J. Appl. Basic. Med. Res.*, 3:1, 16-18.

- Chen, J., Du, Y., Zhu, F., Chen, B., Zhang, Q., Du, S., and Li, P. (2015). Study of the Enantioseparation Capability of Chiral Dual System Based on Chondroitin Sulfate C in CE. *Electrophoresis*, 36, 607–614.
- Chen, Y., Zhang, J., Zhang, L., and Chen, G. (2010). Separation of Dipeptides with Two Chiral Centers Using 2-Hydroxypropyl-β-CD-Modified MEKC. *Electrophoresis*, 31, 1493–1497.
- Cosconati, S., Forli, S., Perryman A. L., Harris, R., Goodsell D. S., and Olson A. J. (2010). Molecular Dynamics Simulations with Quantum Mechanics/Molecular Mechanics and Adaptive Neural Networks. *Expert Opin. Drug Discov.*, 5:6, 597-607.
- Crego, L., Gómez, J., Marina, M.L., and Lavandera, J.L. (2001). Application of Capillary Zone Electrophoresis with Off-Line Solid-Phase Extraction to in Vitro Metabolism Studies of Antifungals. *Electrophoresis*, 22, 2503–25 11.
- Danel, C., Foulon, C., Goossens, J.F., Bonte, J.P., and Vaccher, C. (2006). Validation of Chiral Electrokinetic Chromatography Methods Using Highly Sulfated Cyclodextrins: Determination of Enantiomeric Purity of Aromatase Inhibitors. *Chromatographia*, 63, 353–358.
- Del Valle, E. M. M. (2003). Cyclodextrins and Their Uses: A Review. *Process Biochem.*, 39:9, 1033-1046.
- Demissie, H. and Duraisamy, R. (2016). Effects of Electrolytes on the Surface and Micellar Characteristics of Sodium Dodecyl Sulphate Surfactant Solution. J. Sci. Innovative Res., 5:6, 208-214.
- Domínguez, A., Fernández, A., González, N., Iglesias, E., and Montenegro, L. (1997). Determination of Critical Micelle Concentration of Some Surfactants by Three Techniques. J. Chem. Edu., 74, 1227–1231.
- Duchene, D. (2011). Cyclodextrins in Pharmaceutics, Cosmetics, and Biomedicine: Current and Future Industrial Applications. 1st edition. John Wiley & Sons, Inc. 2-18
- Eder, A.R., Chen, J.S., and Arriaga, E. (2006). Separation of Doxorubicin and Doxorubicinol by Cyclodextrin-Modified Micellar Electrokinetic Capillary Chromatography. *Electrophoresis*, 27, 3263–3270.
- Ekroos, M., and Sjogren, T. (2006). Structural Basis for Ligand Promiscuity in Cytochrome P450 3A4. *PNAS*, 103:37, 13682-13687.

- Elbashir, A.A., Aboul-enein, H.Y., Elbashir, A.A., and Aboul-enein, H.Y. (2015). Capillary Electrophoresis and Molecular Modeling as a Complementary Technique for Chiral Recognition Mechanism. *Crit. Rev. Anal. Chem.* 43:3, 131-137.
- Escuder-Gilabert, L., Martín-Biosca, Y., Medina-Hernández, M.J., and Sagrado, S. (2014). Cyclodextrins in Capillary Electrophoresis: Recent Developments and New Trends. J. Chromatogr. A, 1357, 2–23.
- Fang, L., Du, Y., Hu, X., Luo, L., Guo, X., and Yu, J. (2017). Carboxymethyl β-Cyclodextrin as Chiral Selector in Capillary Electrophoresis: Enantioseparation of 16 Basic Chiral Drugs and Its Chiral Recognition Mechanism Associated with Drugs' Structural Features. *Biomed. Chromatogr.*, 31, 1-7.
- Fassihi, A. R. (1993). Racemates and Enantiomers in Drug Development. *Int. J. Pharm.*, 92:1-3, 1-14.
- Fatiha, M., Leila, L., Leila, N., and Eddine, K.D. (2012). Theoretical Study of the Inclusion Processes of Ethyl p-hydroxybenzoate with β-cyclodextrin: PM3MM and ONIOM2 Calculations. J. Taiwan Ins. Chem. Eng., 43, 868–872.
- FDA Drug Safety Communication. (2016). FDA Limits Usage of Nizoral (Ketoconazole) Oral Tablets Due To Potentially Fatal Liver Injury and Risk of Drug Interactions and Adrenal Gland Problems.
- Fejős, I., Urbancsok, Z., Hu, W., and Szente, L. (2014). Separation of Alogliptin Enantiomers in Cyclodextrin-Modified Capillary Electrophoresis : A Validated Method. Electrophoresis, 35, 2885–2891.
- Fifere, A., Marangoci, N., Maier, S., Coroaba, A., Maftei, D., and Pinteala, M. (2012). Theoretical Study on β-cyclodextrin Inclusion Complexes with Propiconazole and Protonated Propiconazole. *Beilstein J. Org. Chem.*, 8, 2191–2201.
- Frisch, MJ, Trucks, GW, Schlegel HB, Scuseria GE, Robb MA, Cheeseman JR, Scalmani G, Barone V, Mennucci B, Petersson GA, Nakatsuji H, Caricato M, Li X, Hratchian HP, Izmaylov AF, Bloino J, Zheng G, Sonnenberg JL, Hada M, Ehara M, Toyota K, Fukuda R, Hasegawa J, Ishida M, Nakajima T, Honda Y, Kitao O, Nakai H, Vreven T, Montgomery JA Jr, Peralta JE, Ogliaro F, Bearpark M, Heyd JJ, Brothers E, Kudin KN, Staroverov VN, Keith T, Kobayashi R, Normand J, Raghavachari K, Rendell A, Burant JC, Iyengar SS, Tomasi J, Cossi M, Rega N, Millam JM, Klene M, Knox JE, Cross JB, Bakken V, Adamo C, Jaramillo J, Gomperts R, Stratmann RE, Yazyev O, Austin AJ,

Cammi R, Pomelli C, Ochterski JW, Martin RL, Morokuma K, Zakrzewski VG, Voth GA, Salvador P, Dannenberg JJ, Dapprich S, Daniels AD, Farkas O, Foresman JB, Ortiz JV, Cioslowski J, Fox DJ. Gaussian 09, Revision D.01. Gaussian, Inc: Wallingford CT; 2013.

- Grosdidier, A., Zoete, V., and Michielin, O. (2009). Blind Docking of 260 Protein-Ligand Complexes with EADock 2.0. J. Comp. Chem., 30, 2021–2030.
- Gübitz, G. and Schmid, M.G. (2008). Chiral Separation by Capillary Electromigration Techniques. J. Chromatogr. A, 1204, 140–156.
- Hancu, G., Hilochie, A., Vlad, A. R., Cârje, A., and Vescan, A. T., (2016). Enantiomeric Separation of Sibutramine by Capillary Zone Electrophoresis. J. Braz. Chem. Soc., 27:6, 1116-1120.
- Hancu, G., Attila, L., and Aura, P. (2015). Chiral Separation of the Enantiomers of Omeprazole and Pantoprazole by Capillary Electrophoresis. *Chromatographia*, 78, 279–284.
- Hancu, G., Kelemen, H., Rusu, A., and Gyéresi, Á. (2013). Development of A Capillary Electrophoresis Method for the Simultaneous Determination of Cephalosporins. J. Serb. Chem. Soc., 78, 1413–1423.
- Hazai, E., Hazai, I., Demko, L., Kovacs, S., Malik, D., Akli, P., Hari, P., Szeman, J., Fenyvesi, E., Benes, E., Szente, L., and Bikadi, Z. (2010). Cyclodextrin KnowledgeBase A Web-Based Service Managing CD-Ligand Complexation Data. J. Comp. Aided Mol. Des., 24, 713–717.
- Heeres, J., Meerpoel, L., and Lewi, P. (2010). Conazoles. Molecules, 15, 4129-4188.
- Hermawan, D., Wan Ibrahim, W.A., Sanagi, M.M., and Aboul-Enein, H.Y. (2010). Chiral Separation of Econazole Using Micellar Electrokinetic Chromatography with Hydroxypropyl-γ-Cyclodextrin. J. Pharm. Biomed. Anal, 53, 1244–1249.
- Hou, X., Du, J., Zhang, J., Du, L., Fang, H., and Li, M. (2012). How to Improve Docking Accuracy of AutoDock4.2: A Case Study Using Different Electrostatic Potentials. J. Chem. Inf. Model, 53, 188–200.
- Huang, L., Lin, J., Xu, L., and Chen, G. (2007). Nonaqueous and Aqueous-Organic Media for the Enantiomeric Separations of Neutral Organophosphorus Pesticides by CE. *Electrophoresis*, 28, 2758–2764.
- Hutt, A. J. and O'Grady, J. (1996). Drug Chirality: A Consideration of the Significance of the Stereochemistry of Antimicrobial Agents. J. Antimicrob. Chemother. 37, 7-32.

- Indra, KR. Chirality In Drug Design And Development, Marcel Dekker, New York, 2004.
- Iorga, B., Herlem, D., Barré, E., and Guillou, C. (2006). Acetylcholine Nicotinic Receptors: Finding the Putative Binding Site of Allosteric Modulators Using the "Blind Docking" Approach. J. Mol. Model., 12, 366–372.
- Issaraseriruk, N., Shitangkoon, A., and Aree, T. (2010). Molecular Docking Study for the Prediction of Enantiodifferentiation of Chiral Styrene Oxides by Octakis(2,3-di-O-Acetyl-6-O-Tert-Butyldimethylsilyl)-γ-Cyclodextrin. J. Mol. Graph. Model., 28, 506–512.
- Izake E.L. Chiral Discrimination and Enantioselective Analysis of Drug: An Overview. (2007). J. Pharm. Sci. 96:7, 1659–1676.
- Jamali, B., Bjørnsdottir, I., Cornett, C., and Honoré Hansen, S. (2009). Investigation of a Dual CD Chiral CE System for Separation of Glitazone Compounds. *Electrophoresis*, 30, 2853–2861.
- de Jesus, M.B., de Matos Alves Pinto, L., Fraceto, L.F., Takahata, Y., Lino, A.C.S., Jaime, C., and de Paula, E. (2006). Theoretical and Experimental Study of A Praziquantel and β-Cyclodextrin Inclusion Complex Using Molecular Mechanic Calculations and H1-Nuclear Magnetic Resonance. J. Pharm. Biomed. Anal., 41, 1428–1432.
- Jimidar, M. I., Ael, W. V., Nyen, P. V., Peeters, M., Redlich, D. and Smet, M. D. (2004). A Screening Strategy for the Development of Enantiomeric Separation Methods in Capillary Electrophoresis. *Electrophoresis*, 25, 2772–2785.
- Kavran-Belin, G., Rudaz, S., and Veuthey, J.-L. (2005). Enantioseparation of Baclofen with Highly Sulfated β-Cyclodextrin by Capillary Electrophoresis with Laser-Induced Fluorescence Detection. J. Sep. Sci., 28, 2187–2192.
- Károly, M., Vámos, J., Nemes, A., Rácz, A., and Noszál, B. (2003). Lipophilicity of Vinpocetine and Related Compounds Characterized by Reversed-Phase Thin-Layer Chromatography. J. Chromatogr. A, 996, 195–203.
- Khan Beigi, F. A., Imani, M., Payehghadr, M. and Hosseini, H. (2011). SPE-HPLC Method for Determination of Ketoconazole and Clotrimazole Residues in Cow's Milk. J. Braz. Chem. Soc. 22:9, 1679-1685.
- Kicuntod, J., Sangpheak, K., Mueller, M., Wolschann, P., Viernstein, H., Kato, K. and Chavasiri, W. (2018). Theoretical and Experimental Studies on Inclusion Complexes of Pinostrobin and β-Cyclodextrins. *Sci. Pharm.*, 86:5, 1-15.

- Korting, H.C. and Schöllmann, C. (2009). The significance of Itraconazole for Treatment of Fungal Infections of Skin, Nails and Mucous Membranes. J. Ger. Soc. Dermatol., 7, 11–20.
- Krait, S., Heuermann, M., Scriba, G. K. E. (2017). Development of a Capillary Electrophoresis Method for the Determination of the Chiral Purity of Dextromethorphan by a Dual Selector System using Quality by Design Methodology. J. Sep. Sci., 41:6, 1405-1413.
- Kroemer, R.T. (2007). Structure-Based Drug Design: Docking and Scoring. *Curr. Protein Pept. Sci.*, 8, 312–328.
- Li, L., Wu, C., Ma, Y., Zhou, S., Li, Z., Sun, T. (2017). Effectively Enhancing the Enantioseparation Ability of β-cyclodextrin Derivatives by *de novo* Design and Molecular Modeling. Analyst, 142:19, 3699-3706.
- Li, L., Li, X., Luo, Q., and You, T. (2015a). A Comprehensive Study of the Enantioseparation of Chiral Drugs by Cyclodextrin Using Capillary Electrophoresis Combined with Theoretical Approaches. *Talanta*, 142, 28–34.
- Li W., Ding G. S. and Tang A. N. (2015b). Enantiomer Separation of Propranolol and Tryptophan Using Bovine Serum Albumin. *RSC Adv.*, 5, 93850-93857. functionalized silica nanoparticles as adsorbents
- Li, W., Liu, C., Tan, G., Zhang, X., Zhu, Z., and Chai, Y. (2012). Molecular Modeling Study of Chiral Separation and Recognition Mechanism of β-Adrenergic Antagonists by Capillary Electrophoresis. *Int. J. Mol. Sci.*, 13, 710–725.
- Lin, X., Zhu, C., and Hao, A. (2005). Enantioseparation in Capillary Electrophoresis Using 2-O-(2-hydroxybutyl)-β-CD as a Chiral Selector. *Electrophoresis*, 26, 3890–3896.
- Lin, X., Hou, W., and Zhu, C. (2003). Enantiomer Separation of Miconazole by Capillary Electrophoresis with Dual Cyclodextrin Systems. *Anal. Sci.*, 19, 1509–1512.
- Lin X., Li G., Jiang W., Chua Y., Wu P., Yafeng G. (2001). Chiral Separation of Lobeline and Benzhexol by Capillary Electrophoresis Using the Reaction Mixture of β-cyclodextrin, Phosphorous-pentoxide and L-glutamic Acid as Chiral Selector. Anal. Chim. Acta, 431, 41-48.
- Lipka, E., Danel, C., Bonte, J.P., and Vaccher, C. (2010). Dual CD System in Capillary Electrophoresis for Direct Separation of the Four Stereoisomers of Agonist and Antagonist Melatoninergic Ligands. *Electrophoresis*, 31, 1529–1532.

- Liu, Y and You, C. C. (2001). Molecular Recognition Studies on Modified Cyclodextrins. *Chinese J. Chem.* 19:6, 533-544.
- Liu, Y., Fu, X., Ma, C., Zhong, J., Liao, Y., and Liu, H. (2009). Chiral Separation of Raltitrexed by Cyclodextrin-Modified Micellar Electrokinetic Chromatography. Anal. Bioanal. Chem., 393, 321–326.
- Liu, C., Zhang, J., Zhang, X., Zhao, L., and Li, S. (2018). Enantiomeric Separation of Adrenaline, Noradrenaline, and Isoprenaline by Capillary Electrophoresis Using Streptomycin-Modified Gold Nanoparticles. *Microchim. Acta*, 185:4, 227.
- Lourenco, T. C., Batista Jr, J. M., Furlan, M., He, Y., Nafie, L. A., Santana C. C., Cass
 Q. B. (2012). Albendazole Sulfoxide Enantiomers: Preparative Chiral Separation and Absolute Stereochemistry. *J. Chromatogr. A*, 1230, 61-65.
- Maier, N.M., Franco, P., and Lindner, W. (2001). Separation of Enantiomers: Needs, Challenges, Perspectives. J. Chromatogr. A, 906, 3–33.
- Mangelings, D. and Vander Heyden, Y. (2008). Chiral Separations in Sub- and Supercritical Fluid Chromatography. J. Sep. Sci., 31, 1252–1273.
- Mbuna, J., Kaneta, T., and Imasaka, T. (2010). Measurement of Intracellular Accumulation of Anthracyclines in Cancerous Cells by Direct Injection of Cell Lysate in MEKC/LIF Detection. *Electrophoresis*, 31, 1396–1404.
- McConathy, J. and Owens, M.J. (2003). Stereochemistry in Drug Action. *Prim. Care Companion J. Clin. Psychiatry*, 5, 70–73.
- Meiler, G.L. and J. (2013). RosettaLigand Docking with Flexible XML Protocols. *Methods Mol Biol*, 819, 143–155.
- Messner, M., Kurkov, S. V, Jansook, P., and Loftsson, T. (2010). Self-Assembled Cyclodextrin Aggregates and Nanoparticles. *Int. J. Pharm.*, 387, 199–208.
- Mohan, S.J., Mohan, E.C., and Yamsani, M.R. (2009). Chirality and its Importance in Pharmaceutical Field-An Overview. Int. J. Pharm. Sci. Nanotechnology, 1, 309–316.
- Morris, G.M., Goodsell, D.S., Halliday, R.S., Huey, R., Hart, W.E., Belew, R.K., Olson, A.J., and Al, M.E.T. (1998). Automated Docking Using A Lamarckian Genetic Algorithm and An Empirical Binding Free Energy Function. *J. Comp. Chem.*, 19, 1639–1662.

- Mulisa, E., Ndorbor, T., Xiao, D., and He, H. (2014). Molecular Docking Method for the Prediction of Enantiorecognition of Trihexyphenidyly and Its Derivatives on Carboxy Methyl-Cyclodextrins. *Ind. J. Sci. Res. and Tech.* 2, 31–37.
- Nadendla, R.R. (2004). Molecular modeling: A Powerful Tool for Drug Design and Molecular Docking. *Resonance*, 9, 51–60.
- Nakamura, H., Sano, A., and Matsura K. (1998). Determination of Critical Micelle Concentration of Anionic Surfactants by Capillary Electrophoresis Using 2-Naphtalenemethanol as a Marker for Micelle Formation. *Anal. Sci.*, 14, 379-382.
- Nascimento, C.S., Lopes, F., and Guimar, L. (2014). Molecular Modeling Study of the Recognition Mechanism and Enantioseparation of 4-Hydroxypropranolol by Capillary Electrophoresis Using Carboxymethyl-β-Cyclodextrin As The Chiral Selector. *Analyst*, 139, 3901–3910.
- Neumajera, G., Sohajda, T., Darcsi, A., Tóth G., Szente, L., Noszál, B., Béni, S. (2012). Chiral Recognition of Dapoxetine Enantiomers with Methylated-Gamma-Cyclodextrin: A Validated Capillary Electrophoresis Method. J. Pharm. Biomed. Anal., 62, 42–47.
- Nishi, H. (1996). Enantiomer Separation of Drugs by Electrokinetic Chromatography. J. Chromatogr. A, 735, 57–76.
- Nishi, H. and Terabe, S. (1996). Micellar Electrokinetic Chromatography Perspectives in Drug Analysis. J. Chromatogr. A, 735, 3–27.
- Nishi, H. and Terabe, S. (1995). Optical Resolution of Drugs by Capillary Electrophoretic Techniques. J. Chromatogr. A, 694, 245–276.
- Pasquinia, B., Melanic, F., Caprinia, C., Bubba M. D., Pinzauti, S., Orlandini, S., and Furlanetto S. (2017). Combined Approach Using Capillary Electrophoresis, NMR Andmolecular Modeling for Ambrisentan Related Substances Analysis: Investigation of Intermolecular Affinities, Complexation Andseparation Mechanism. J. Pharmceut. Biomed. Anal., 144, 220-229.
- Passos, J. J., de Souza, F. B., Lula, I. S., Barreto, E. A., Lopes, J. F., de Almeida W. B., and Sinisterra, R. D. (2011). Multi-Equilibrium System Based on Sertraline and β-Cyclodextrin Supramolecular Complex in Aqueous Solution. *Int. J. Pharm.*, 421:1, 24–33.

- Pérez-Fernández, V., García, M.A., and Marina, M.L. (2010). Enantiomeric Separation Of Cis-Bifenthrin by CD-MEKC: Quantitative Analysis in A Commercial Insecticide Formulation. *Electrophoresis*, 31, 1533–1539.
- Phyo, Y. Z., Cravo, S., Palmeira, A., Tiritan, M. E., Anake, K., Pinto, M. M. M., and Fernandes, C. (2018). Enantiomeric Resolution and Docking Studies of Chiral Xanthonic Derivatives on Chirobiotic Columns. *Molecules*, 23, 142.
- Płotka, J. M., Simeonov, V., Morrison, C., Biziuk, M., Namieśnik, J. (2014). Capillary Gas Chromatography Using A γ-Cyclodextrin for Enantiomeric Separation of Methylamphetamine, Its Precursors and Chloro Intermediates After Optimization of the Derivatization Reaction. J. Chromatogr. A, 1347, 146–156.
- Pyrgaki, C., Bannister, S.J., Gera, L., Gerber, J.G., and Gal, J. (2011). Stereoselective Determination of the Epimer Mixtures of Itraconazole in Human Blood Plasma using HPLC and Fluorescence Detection. *Chirality*, 503, 495–503.
- Qiang Lin G., Ming Li Y., and Chan A. S. C. *Principles and Applications of Assymetric Synthesis.* (2001). John Wiley & Sons, Inc. 4-8.
- Quaglia, M.G., Donati, E., Desideri, N., Fanali, S., D'auria, F.D., and Tecca, M. (2002). Chiral Discrimination by HPLC and CE and Antifungal Activity of Racemic Fenticonazole and its Enantiomers. *Chirality*, 14, 449–454.
- Quirino, J.P., Dulay, M.T., Fu, L., Mody, T.D., Zare, R.N., and Avenue, A. (2002). Capillary Electrophoresis Separation and Native Laser-Induced Fluorescence Detection of Metallotexaphyrins. *J. Sep. Sci.*, 2002, 25, 819–824.
- Rasheed A., Ashok Kumar C. K., Sravanthi V. V. N. S. S. (2008). Cyclodextrins as Open Access Drug Carrier Molecule: A Review. *Sci. Pharm.*, 76, 567–598.
- Rawjee, Y.Y., Staerk, D.U., and Vigh, G. (1993). Capillary Electrophoretic Chiral Separations with Cyclodextrin Additives. I. Acids: Chiral Selectivity As A Function of pH and the Concentration of β-Cyclodextrin for Fenoprofen and Ibuprofen. J. Chromatogr. A, 635, 291–306.
- Redenti, E., Ventura, P., Fronza, G., Selva, a, Rivara, S., Plazzi, P. V, and Mor, M. (1999). Experimental and Theoretical Analysis of the Interaction of (+/-)-Cis-Ketoconazole with β-cyclodextrin in the Presence of (+)-L-Tartaric Acid. *J. Pharm. Sci.*, 88, 599–607.
- Redhu S. and Jindal A. (2013). Molecular Modelling: A New Scaffold For Drug Design. Int. J. Pharm. Pharm. Sci., 5:1, 5-8.

- Riley C. M., Lough W. J. and Wainer I. W. (1994). *Pharmaceutical and Biomedical Application of Liquid Chromatography*. Oxford: Pergamon.
- Rodriguez-Delgado, M. A., Francisco J. Garcia-Montelongo, and Cifuentes, A. (2002). Ultrafast Sodium Dodecyl Sulfate Micellar Electrokinetic Chromatography with Very Acidic Running Buffers. *Anal. Chem.*, 74, 257-260.
- Rotstein, D.M. and Denis J. Kertesz, Keith A. M. Walker, and D.C.S. (1992). Stereoisomers of Ketoconazole: Preparation and Biological Activity. J. Med. Chem., 35:15, 2818–2825.
- Saag, M.S. and Dismukes, W.E. (1988). Azole Antifungal Agents : Emphasis a New Triazole. *Antimicrob. Agents Chemother*. 32:1, 1-8.
- Sakina, H., Abdelaziz, B., Leila, N., Imene, D., Fatiha, M., and Eddine, K.D. (2012). Molecular Docking Study on β-Cyclodextrin Interactions of Metobromuron and [3-(p-bromophenyl)-1-methoxy-1-methylurea]. *J. Inc. Phenom. Macrocyc. Chem.*, 74, 191–200.
- Samakashvili, S., Salgado, A., Scriba, G. K. E., and Chankvetadze, B. (2013). Comparative Enantioseparation of Ketoprofen with Trimethylated α-, β-, and γ-Cyclodextrins in Capillary Electrophoresis and Study of Related Selector – Selectand Interactions Using Nuclear Magnetic Resonance Spectroscopy. *Chirality*, 25, 79–88.
- Sawarkar H. S. (2011). Cyclodextrines A Multidimensional Pharmaceutical Tool. *Int. J. Herbal Drug*, 1:11, 11-21.
- Schiaffella F., Macchiarulo A., Milanese L., Vecchiarelli A., and Fringuellia R. (2006). Novel Ketoconazole Analogues Based on the Replacement of 2,4-Dichlorophenyl Group With 1,4-Benzothiazine Moiety: Design, Synthesis, And Microbiological Evaluation. *Bioorg. Med. Chem.*, 14, 5196–5203.
- Schmid, M.G. and Gubitz, G. (1997). Chiral Separation Principles in Capillary Electrophoresis. J. Chromatogr. A., 792, 179–225.
- Scriba, G.K.E. V. (2008). Cyclodextrins in Capillary Electrophoresis Enantioseparations-Recent Developments and Applications. J. Sep. Sci., 31, 1991–2011.
- Sekhon B. S. (2010). Enantioseparation of Chiral Drugs An Overview. Int. J. ChemTech Res., 2:2, 1584-1594.
- Shakil, S. (2013). A Simple Click By Click Protocol To Perform Docking. Experimental and Clinical (EXCLI) J., 12, 831–857.

- Shen, L., and Yang, W. (2018). Molecular Dynamics Simulations with Quantum Mechanics/ Molecular Mechanics and Adaptive Neural Networks. J. Chem. Theory Comput., 14:3, 1442-1455.
- Shi, J., Su, Y., and Jiang, W. (2013). Enantioseparation and Chiral Recognition of α-cyclohexylmandelic Acid and Methyl α-cyclohexylmandelate on Hydroxypropyl-β-cyclodextrin as Chiral Selector: HPLC and Molecular Modeling. J. Chromatogr. Sci., 51, 8–16.
- Shi, W., Nacev, B., Bhat, S., and Liu, J.O. (2010). Impact of Absolute Stereochemistry on the Antiangiogenic and Antifungal Activities of Itraconazole. ACS Med. Chem. Lett., 1, 155–159.
- Singh A. K., Kedor-Hackmann E. R. M., Santoro M. I. R. M. (2004). Enantiomeric Separation and Quantitative Determination of Propranolol Enantiomers in Pharmaceutical Preparations by Chiral Liquid Chromatography. *Braz. J. Pharm. Sci.*, 40: 3, 301-308.
- Smith, S.W. (2009). Chiral Toxicology: It's The Same Thing...Only Different. *Toxicol. Sci.*, 110:1, 4–30.
- de Sousa, F.B., Denadai, Â.M.L., Lula, I.S., Lopes, J.F., Dos Santos, H.F., De Almeida, W.B., and Sinisterra, R.D. (2008). Supramolecular Complex of Fluoxetine with β-cyclodextrin: An Experimental and Theoretical Study. *Int. J. Pharm.*, 353, 160–169.
- Suliman, F.O. and Elbashir, A.A. (2012). Enantiodifferentiation of Chiral Baclofen by β-cyclodextrin Using Capillary Electrophoresis: A Molecular Modeling Approach. J. Mol. Struct., 1019, 43–49.
- Surpateanu, G. and Iorga, and B.I. (2012). Evaluation of Docking Performance in A Blinded Virtual Screening of Fragment-Like Trypsin Inhibitors. J. Comp. Aided Mol. Des., 26, 595–601.
- Szabo, Z., Völgyi, G., Komjáti, B., Hancu, G., and Nosza, B. (2016). Chiral Separation of Asenapine Enantiomers by Capillary Electrophoresis and Characterization of Cyclodextrin Complexes by NMR Spectroscopy, Mass Spectrometry and Molecular Modeling. J. Pharm. Biomed. Anal., 117, 398–404.
- Terabe, S. and Otsuka, K. (1994). Separation Techniques of Enantiomers by Capillary Electrophoretic. *J. Chromatogr. A.*, 666, 295–319.

- Terabe, S., Otsuka, K., Ichikawa, K., Tsuchiya, and Ando, T. (1984). Electrokinetic Separations with Micellar Solutions and Open-Tubular Capillaries. *Anal. Chem.*, 56, 111–113.
- Testa, B., Vistoli, G. and Pedretti, A. (2013). Symmetry Elements and Operations, Classification of Stereoisomers. *Helv. Chim. Acta*, 96, 1-3.
- Thiel, W. Modern Methods and Algorithms of Quantum Chemistry (2000). J. Grotendorst (Ed). 261-283.
- Thienpont, Gal, J., Aeschlimann, C., and Félix, G. (1999). Studies on Stereoselective Separations of the "Azole" Antifungal Drugs Ketoconazole and Itraconazole Using HPLC and SFC on Silica-Based Polysaccharides. *Analusis*, 27, 713–718.
- Toribio, L., del Nozal, M.J., Bernal, J.L., Alonso, C., and Jiménez, J.J. (2007). Enantiomeric Separation of Several Antimycotic Azole Drugs Using Supercritical Fluid Chromatography. J. Chromatogr. A, 1144, 255–261.
- Tsai, C. S. An Introduction to Computational Biochemistry. (2002). Wiley-Liss, Inc. 258–314.
- Tseng, W. B., Hsieh, M. M., Chiu, T. C., Yu, P. L. and Chen S. H. (2018). Enantioseparation of Phenothiazines Through Capillary Electrophoresis with Solid Phase Extraction and Polymer Based Stacking. *J. Food Drug Anal.*, 26:3, 1171-1179.
- Tucker, G.T. (2000). Chiral Switches. Lancet, 355, 1085-1087.
- Upadhyay, S. K., and Ali S. M. (2018). Molecular Recognition of Flunarizine Dihydrochloride and β-Cyclodextrin Inclusion Complex by NMR and Computational Approaches. *Chem. Cent. J.*, 12, 33.
- Velikinac, I., Cudina, O., Janković, I., Agbaba, D., and Vladimirov, S. (2004). Comparison of Capillary Zone Electrophoresis and High Performance Liquid Chromatography Methods for Quantitative Determination of Ketoconazole in Drug Formulations. *Farmaco*, 59, 419–424.
- Wan Ibrahim, W.A., Munirah, S., Wahib, A.B.D., Hermawan, D., and Sanagi, M.M. (2013). Separation of Selected Imidazole Enantiomers Using Dual Cyclodextrin System in Micellar Electrokinetic Chromatography. *Chirality*, 335, 328–335.
- Wan Ibrahim, W.A., Munirah, S., Wahib, A.B.D., Hermawan, D., and Sanagi, M.M. (2012). Chiral Separation of Vinpocetine Using Cyclodextrin-Modified Micellar Electrokinetic Chromatography. *Chirality*, 254, 252–254.

- Wan Ibrahim, W.A., Hermawan, D., Sanagi, M.M., and Aboul-Enein, H.Y. (2009). Cyclodextrin-modified MEKC for Enantioseparation of Hexaconazole, Penconazole, and Myclobutanil. J. Sep. Sci., 32, 466–471.
- Wang, Z., Tang, Z., Gu, Z., Hu, Z., Ma, S., and Kang, J. (2005). Enantioseparation of Chiral Allenic Acids by Micellar Electrokinetic Chromatography with Cyclodextrins As Chiral Selector. *Electrophoresis*, 26, 1001–1006.
- Wang, Z., Cai, C., Lin, Y., Bian, Y., Guo, H., and Chen, X. (2011). Enantioselective Separation of Ketoconazole Enantiomers by Membrane Extraction. *Sep. Purif. Technol.*, 79, 63–71.
- Wienen, F., Laug, S., Baumann, K., Schwab, A., Just, S., and Holzgrabe, U. (2003). Determination of Clotrimazole in Mice Plasma by Capillary Electrophoresis. J. Pharm. Biomed. Anal., 30, 1879–1887.
- Willems, L., van der Geest, R.., and de Beule, K. (2001). Itraconazole Oral Solution and Intravenous Formulations: Review of Pharmacokinetics and Pharmacodynamics. J. Clin. Pharm. Ther., 26, 159–169.
- Yang G.S., Chen D. M., Yang M. Y., Tang B., Gao J. J., Aboul-Enein H. Y., and Koppenhoefer B. (2005). Enantioseparation of Some Clinically Used Drugs by Capillary Electrophoresis Using Sulfated β-Cyclodextrin as a Chiral Selector. *Chromatographia*, 62, 441–445.
- Yang, L. J., Chang, Q., Zhou, S. Y., Yang, Y. H., Xia, F. T., Chen, W., Li, M., and Yang, X. D. (2018). Host–Guest Interaction Between Brazilin and Hydroxypropyl-β-Cyclodextrin: Preparation, Inclusion Mode, Molecular Modelling and Characterization. *Dyes Pigm.*, 150, 193-201.
- Yao, Y., Song, P., Wen, X., Deng, M., Wang, J., and Guo, X. (2017). Chiral Separation of 12 Pairs of Enantiomers by Capillary Electrophoresis Using Heptakis-(2,3-Diacetyl-6-Sulfato)-β-Cyclodextrin As the Chiral Selector and the Elucidation of the Chiral Recognition Mechanism by Computational Methods. *J. Sep. Sci.*, 40:14, 2849-3014.
- Ye J., Yu W., Chen G., Shen Z. and Zeng S. (2009). Enantiomeric Separation of 2-Arylpropionic Acid Nonsteroidal Anti-inflammatory Drugs by HPLC with Hydroxypropyl-β-cyclodextrin as Chiral Mobile Phase Additive. *Biomed. Chromatogr.* 24, 799–807.

- Yu, J., Liang, X., Wang, Z., Guo, X., Sun, T., and Guo, X. (2015). Separation of Folinic Acid Diastereomers in Capillary Electrophoresis Using a New Cationic β-Cyclodextrin Derivative. *PLOS One*, 1–10.
- Zhang, Y.I.J.U.N., Huang, M.X., Zhang, Y.U.P., Armstrong, D.W., and Breitbach, Z.S. (2013). Use of Sulfated Cyclofructan 6 and Sulfated Cyclodextrins for the Chiral Separation of Four Basic Pharmaceuticals by Capillary Electrophoresis. *Chirality*, 742, 735–742.
- Zhao, M., Cui, Y., Yu, J., Xu, S., and Guo, X. (2014). Combined Use of Hydroxypropyl-β-cyclodextrin and Ionic Liquids for the Simultaneous Enantioseparation of Four Azole Antifungals by CE and A Study of the Synergistic Effect. J. Sep. Sci., 37, 151–157.
- Zhang, X., Li, Z., Shen, B., Chen, J., Xu, X. (2012). Enantioseparation of Three Non-Steroidal Anti-Inflammatory Agents on Chiral Stationary Phase by HPLC. *JASMI*, 2, 18-23.
- Zhu, C., Lin, X., and Wei, Y. (2002). Chiral Separation of Pemoline Enantiomers by Cyclodextrin-Modified Micellar Capillary Chromatography. J. Pharm. Biomed. Anal., 30, 293–298.
- Znaleziona, J., Fejős, I., Ševčík, J., Douša, M., Béni, S. and Maier, V. (2015). Enantiomeric Separation of Tapentadol by Capillary Electrophoresis - Study of Chiral Selectivity Manipulation by Various Types of Cyclodextrins. *J. Pharm. Biomed. Anal.*, 105, 10-16.