

CHARACTERISATION OF MANNANASE FROM *Pontibacillus* sp. CL43

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DEDICATION

This thesis is dedicated to my beloved parents for their fully support and encouragement.

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ABSTRACT

Lignocellulosic biomass is a renewable feedstock for biofuel production. Pretreatment is performed to destroy the biomass recalcitrant structure subsequently allow enzymes to hydrolyse cellulose and hemicellulose for sugars production. These sugars could be used for biofuel production. In chemical pretreatment of lignocellulosic biomass, metal salts (KCl, CaCl₂ and MgCl₂) and surfactants (Tween 20, Tween 80 and Triton X-100) are used. These compounds remain in the biomass after the treatment and affect the performance of enzymes in following hydrolysis process. Hence, enzymes with metal- and surfactant-tolerance characteristics are preferable in biofuel production. This project focused on characterising *Pontibacillus* sp. CL43 and its mannanase which is responsible for degradation of mannan, a component present in lignocellulosic biomass. Strain CL43 is a Gram positive bacterium with rod-shaped, endospore-forming, catalase- and oxidase-positive. Strain CL43 could hydrolyse bile esculin, casein, starch, xylan, Tween 20, Tween 40, Tween 60 and Tween 80. Strain CL43 was susceptible to ampicillin, carbenicillin, doxycycline, gentamicin, minocycline, neomycin, oxacillin, penicillin G, piperacillin, polymyxin B, rifampicin, streptomycin and tetracycline. Mannanase of strain CL43 exhibited optimal activity at 60 °C, pH 7 and 2% (w/v) NaCl. Mannanase activity was found to be stable in the presence of metal ions (Ca²⁺, Cd²⁺, Mn²⁺, K⁺, Mg²⁺ and Co²⁺ with relative activity ranged from 87-128%) and surfactants (Tween 20, Tween 40, Tween 60, Tween 80 and Triton X-100 with relative activity ranged from 88-107%). Mannanase of strain CL43 with good tolerance to these metal ions and surfactants indicated it is a good candidate to be used for lignocellulosic biomass hydrolysis.

ABSTRAK

Biomass lignoselulosa adalah sumber bahan mentah yang boleh diperbaharui yang digunakan untuk penghasilan biofuel. Pra-rawatan dijalankan untuk memecahkan struktur biomass yang rekalsitran kemudiannya membolehkan enzim untuk menghidrolisis selulosa dan hemiselulosa bagi penghasilan gula. Gula jenis ini boleh digunakan untuk penghasilan biofuel. Di dalam pra-rawatan kimia biomass lignoselulosa, garam logam (KCl, CaCl₂ dan MgCl₂) dan surfaktan (Tween 20, Tween 80 dan Triton X-100) telah digunakan. Kompaun ini kekal dalam biomass lignoselulosa selepas pra-rawatan dan ianya dapat mempengaruhi prestasi enzim dalam proses hidrolisis yang berikutnya. Oleh itu, enzim yang mempunyai ciri-ciri logam- dan surfaktan-toleransi lebih disukai dalam penghasilan biofuel. Projek ini difokuskan pada pencirian *Pontibacillus* sp. CL43 dan mannanase yang bertanggungjawab untuk mendegradasi mannan, dimana ianya merupakan salah satu komponen yang terdapat dalam biomass lignoselulosa. Strain CL43 adalah Gram positif bakteria yang berbentuk *rod*, penghasilan endospora, katalase- dan oksidase-positif. Strain CL43 boleh menghidrolisis esculin, kasein, kanji, xilan, Tween 20, Tween 40, Tween 60 dan Tween 80. Strain CL43 sensitif kepada ampicillin, carbenicillin, doxycycline, gentamicin, minocycline, neomycin, oxacillin, penicillin G, piperacillin, polymyxin B, rifampicin, streptomycin and tetracycline. Mannanase daripada strain CL43 mempamerkan aktiviti optimum pada 60 °C, pH 7 dan 2% (w/v) NaCl. Aktiviti mannanase didapati stabil dengan kehadiran ion logam (Ca²⁺, Cd²⁺, Mn²⁺, K⁺, Co²⁺ dan Mg²⁺ dengan aktiviti relatif 87-128%) dan surfaktan (Tween 20, Tween 40, Tween 60, Tween 80 dan Triton X-100 dengan aktiviti relatif 88-107%). Mannanase daripada strain CL43 menunjukkan toleransi yang baik terhadap ion logam dan surfaktan. Oleh yang demikian, enzim ini boleh menjadi calon yang baik bagi penghidrolisisan biomass lignoselulosa.

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LIST OF ABBREVIATIONS

Al^{3+}	-	Aluminium (III) ion
$\text{Al}_2(\text{SO}_4)_3$	-	Aluminium (III) sulfate
ASW	-	Artificial seawater
Ca^{2+}	-	Calcium (II) ion
CaCl_2	-	Calcium (II) chloride
CAZy	-	Carbohydrate-active enzymes
Cd^{2+}	-	Cadmium (II) ion
$\text{Cd}(\text{NO}_3)_2$	-	Cadmium (II) nitrate
CE	-	Carbohydrate esterase
Co^{2+}	-	Cobalt (II) ion
CoCl_2	-	Cobalt (II) chloride
Cu^{2+}	-	Copper (II) ion
CuSO_4	-	Copper (II) sulfate
DMSO	-	Dimethylsulfoxide
DNS	-	Dinitrosalicylic acid
et al		et alia
Fe^{2+}	-	Ferum (II) ion
FeSO_4	-	Ferum (II) sulfate
GH	-	Glycoside hydrolase
H^+	-	Hydrogen ion
H_2O_2	-	Hydrogen peroxide
K^+	-	Potassium ion
KCl	-	Potassium chloride
LBG	-	Locust bean gum
MA	-	Marine agar
Mg^{2+}	-	Magnesium (II) ion
MgCl_2	-	Magnesium (II) chloride
Mn^{2+}	-	Manganese (II) ion
MnCl_2	-	Manganese (II) chloride
Na^+	-	Sodium ion

NaCl	-	Sodium chloride
Ni ²⁺	-	Nickel (II) ion
NiSO ₄	-	Nickel (II) sulfate
OD	-	Optical density
OF	-	Oxidative fermentative
SDS	-	Sodium dodecyl sulfate
-SH groups	-	Sulfhydryl groups
sp.	-	Species (singular)
spp.	-	Species (plural)
Zn ²⁺	-	Zinc (II) ion
ZnSO ₄	-	Zinc (II) sulfate

LIST OF SYMBOLS

α	-	Alpha
β	-	Beta
$^{\circ}\text{C}$	-	Degree Celsius
g	-	Gram
h	-	Hour
kPa	-	Kilo Pascal
mL	-	Millilitre
mm	-	Millimetre
mM	-	Millimolar
M	-	Molar
nm	-	Nanometre
n	-	Number
%	-	Percentage
®	-	Registered trademark
rpm	-	Revolutions per minute
U	-	Unit
v/v	-	Volume per volume
w/v	-	Weight per volume

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CHAPTER 1

INTRODUCTION

1.1 Background

Owing to the increasing demand of fossil fuels, global warming and emission of greenhouse gases, it is important to produce biofuels to maintain sustainable environment (Saini, Saini and Tewari, 2015). First generation biofuels are produced directly from food crops such as corn, sugarcane, soybean and others. While second generation biofuels are produced from non-edible biomass such as lignocellulosic biomass and do not compete with food crops production (Amoozegar et al., 2019). Bioethanol produced from lignocellulosic biomass is considered as a renewable and sustainable biofuel (Wang et al., 2012). Bioethanol production also reduces the environmental pollution arising from disposal of lignocellulosic waste (El-Naggar, Deraz and Khalil, 2014).

Lignocellulosic materials mainly composed of cellulose, hemicellulose and lignin components (Isikgor and Becer, 2015; Zhao et al., 2011a). Cellulose is the major component of lignocellulose biomass (Isikgor et al., 2015). Cellulose determines the structure and framework of cell wall. Hemicellulose acts as linkage that non-covalent bonded to cellulose and covalent bonded to lignin (Michelin et al., 2015). Hardwood hemicelluloses contain mostly xylans, whereas softwood hemicelluloses contain mostly glucomannans (Saha, 2003). Lignin protects plant from pathogen attack and environmental stress (Li, Pu and Ragauskas, 2016). Cross linking of lignin provides mechanical support for upward growth of plant (Chen, 2014).

Mannan is the important part of hemicellulose in plant cell walls and could be hydrolysed by mannanase (Mizutani et al., 2012). Mannanase could be produced from microorganisms, animals and plants (Luo et al., 2012). Fungal mannanase was found maximally active in acidic to neutral pH range (4-7) whereas bacterial mannanase is

found to be active at neutral to alkaline pH (6-12). For example, pH optimum of mannanase from *Bacillus nealsonii* PN11 and *Bacillus* sp. N16-5 are at pH 8 and 9.6 respectively. Besides, majority of mannanases show the thermo-tolerant property with optimum activity in the temperature range of 40-65 °C (Srivastava and Kapoor, 2017). Several halo-tolerant mannanase were also reported in the case of *Bacillus* sp. strain NN, *Bacillus* sp. HJ14, *Pantoea agglomerans* A021, *Enterobacter* sp. strain N18, *Sphingomonas* sp. JB13, *Scopulariopsis candida* strains LMK004 and LMK008 (Amoozegar et al., 2019; You et al., 2016; Zhang et al., 2016; Zhou et al., 2012). Searching for new manannase is an important aspect because manannase with additional features such as tolerant to metal ions and surfactants could make this enzyme more applicable in industries.

Microbial mannanase is widely used because of it is relatively low cost in production and easily controllable condition (El-refai et al., 2014). Mannanase is widely applied in various biotechnology industries such as bleaching of pulp and paper, laundry detergent, instant coffee processing and biofuel industries (Mou et al., 2011). This is the first report on mannanase characterisation for genus *Pontibacillus*.

1.2 Problem Statement

Fossil fuel is major source of energy and its demand gradually increase until may not meet global demand in the future. Burning of fossil fuel also causes global warming and environmental pollution (Amelio et al., 2016). Bioethanol produced from lignocellulosic biomass is considered as a renewable and sustainable biofuel to meet the increasing demand (Wang et al., 2012). Lignocellulosic biomass normally undergoes suitable pretreatment to remove the lignin and reduce crystallinity of cellulose (Baruah et al., 2018). Chemicals that are commonly used for the pretreatment are metal salts (KCl, CaCl₂ and MgCl₂) and surfactants (Tween 20, Tween 80 and Triton X-100) (Putro et al., 2016). However, these chemical residues remained in pre-treated lignocellulosic biomass, which affect the following enzymatic hydrolysis process. Therefore, investigation of enzyme with extra features including metal- and surfactant-tolerant is important to ensure activation of enzyme in the hydrolysis

process. Mangroves are salt-tolerant forest ecosystems that are situated between terrestrial and marine environments where seawater is mixed with freshwater (Wu et al., 2018). The swampy and saline environments make it possible for obtaining microorganisms and their enzymes with unique properties (Gao et al., 2010). Therefore, the aim of this study is to characterise *Pontibacillus* sp. CL43 isolated from mangrove sediment and its mannanase which is one of key enzymes in lignocellulosic biomass degradation.

1.3 Research Objectives

The objectives of the research are:

- (a) To characterise the *Pontibacillus* sp. CL43 phenotypically
- (b) To determine the effect of temperature, pH and salinity on activity and stability of mannanase
- (c) To assess the stability of mannanase in presence of metal ions, organic solvents and detergents

1.4 Scope of Study

The isolated bacterium *Pontibacillus* sp. CL43 was streaked on marine agar and the mannanase activity was screened using qualitative methods. Bacterial phenotypes were studied based on its morphology, biochemical tests, antibiotic-resistant tests, physiology and API kit. Next, activity and stability of mannanase were determined at different temperature, pH and salinity. Lastly, stability of mannanase was assessed in presence of metal ions, organic solvents and detergents.

1.5 Significance of Study

Lignocellulose wastes are generated in large amounts every year, including leaves, stalks, stems, wheat and others. Disposal of these wastes to the landfill causes environmental problems. However, they can be used to produce some value added products. For example, lignocellulose wastes could be converted to bioethanol. Mannan is the major component of hemicellulose. Mannanase is one of the hemicellulases that degrades mannan into smaller molecules (Cheng et al., 2016). This enzyme is widely used in various biotechnological applications, such as bioethanol production, coffee extraction and paper processing (Xia et al., 2016). So far, no mannanase from members of *Pontibacillus* has been reported. In this project, characteristics of mannanase from *Pontibacillus* sp. CL43 were investigated for its potential application such as saccharification process for bioethanol production in future.

REFERENCES

- Adav, S. S. and Sze, S. K. (2014). *Trichoderma* secretome: an overview. In *Biotechnology and biology of Trichoderma* (pp. 103-114): Elsevier.
- Adiguzel, A., Nadaroglu, H. and Adiguzel, G. (2015). Purification and characterization of β -mannanase from *Bacillus pumilus* (M27) and its applications in some fruit juices. *Journal of Food Science and Technology*, 52(8), 5292-5298.
- Adiguzel, G., Sonmez, Z., Adiguzel, A. and Nadaroglu, H. (2016). Purification and characterization of a thermostable endo-beta-1,4 mannanase from *Weissella viridescens* LB37 and its application in fruit juice clarification. *European Food Research and Technology*, 242(5), 769-776.
- Ahmed, S., Luis, A. S., Bras, J. L., Ghosh, A., Gautam, S., Gupta, M. N. et al. (2013). A novel α -L-arabinofuranosidase of family 43 glycoside hydrolase (Ct43Araf) from *Clostridium thermocellum*. *PLoS One*, 8(9), e73575.
- Alalouf, O., Balazs, Y., Volkinshtein, M., Grimpel, Y., Shoham, G. and Shoham, Y. (2011). A new family of carbohydrate esterases is represented by a GDSL hydrolase/acetylxyloxyesterase from *Geobacillus stearothermophilus*. *Journal of Biological Chemistry*, 286(49), 41993-42001.
- Amelio, A., Van der Bruggen, B., Lopresto, C., Verardi, A., Calabro, V. and Luis, P. (2016). Pervaporation membrane reactors: biomass conversion into alcohols. In *Membrane technologies for biorefining* (pp. 331-381): Elsevier.
- Amoozegar, M. A., Safarpour, A., Noghabi, K. A., Bakhtiary, T. and Ventosa, A. (2019). Halophiles and their vast potential in biofuel production. *Frontiers in Microbiology*, 10, 1895.
- Andualem, B. and Gessesse, A. (2013). Production of microbial medium from defatted brebra (*Milletia ferruginea*) seed flour to substitute commercial peptone agar. *Asian Pacific Journal of Tropical Biomedicine*, 3(10), 790-797.
- Ariandi, Yopi and Meryandini, A. (2015). Enzymatic hydrolysis of copra meal by mannanase from *Streptomyces* sp. BF3.1 for the production of mannooligosaccharides. *Hayati Journal of Biosciences*, 22(2), 79-86.
- Bajpai, P. (2016). Structure of lignocellulosic biomass. In *Pretreatment of lignocellulosic biomass for biofuel production* (pp. 7-12): Springer.

- Baliah, N. T. and Begum, P. J. (2015). Isolation, identification and characterization of phosphate solubilizing bacteria (PSB) isolated from economically important crop plants. *Int. J. Curr. Microbiol. App. Sci*, 4(3), 915-924.
- Barnett, C. C., Demaso, C., Dodge, E., Harris, K. A. and Qi, R. B. (2015). Novel mannanase, compositions and methods of use thereof: Google Patents.
- Barton, L. L. (2005). *Structural and functional relationships in prokaryotes*: Springer Science & Business Media.
- Baruah, J., Nath, B. K., Sharma, R., Kumar, S., Deka, R. C., Baruah, D. C. et al. (2018). Recent trends in the pretreatment of lignocellulosic biomass for value-added products. *Frontiers in Energy Research*, 6(141).
- Behera, B. C., Yadav, H., Singh, S. K., Sethi, B. K., Mishra, R. R. and Kumari, S. (2017). Alkaline phosphatase activity of a phosphate solubilizing *Alcaligenes faecalis*, isolated from mangrove soil. *Biotechnology Research and Innovation*, 1(1), 101-111.
- Bhalla, A., Bischoff, K. M. and Sani, R. K. (2014). Highly thermostable GH39 β -xylosidase from a *Geobacillus* sp. strain WSUCF1. *BMC Biotechnology*, 14(1), 963.
- Bhunia, B. and Dey, A. (2012). Statistical approach for optimization of physiochemical requirements on alkaline protease production from *Bacillus licheniformis* NCIM 2042. *Enzyme Research*.
- Bibi, Z., Ansari, A., Zohra, R. R., Aman, A. and Ul Qader, S. A. (2014). Production of xylan degrading endo-1,4- β -xylanase from thermophilic *Geobacillus stearothermophilus* KIBGE-IB29. *Journal of Radiation Research and Applied Sciences*, 7(4), 478-485.
- Bibi, Z., Nawaz, M. A., Waqas, M., Aman, A. and Qader, S. A. U. (2019). Significance of metal ions, solvents and surfactants to improve the xylan degrading behavior of β -1,4-D-xylanohydrolase from *Geobacillus stearothermophilus* KIBGE-IB29. *Biocatalysis and Agricultural Biotechnology*, 17, 242-246.
- Biely, P., Westereng, B., Puchart, V., De Maayer, P. and Cowan, D. A. (2014). Recent progress in understanding the mode of action of acetylxylan esterases. *Journal of Applied Glycoscience*, 61(2), 35-44.
- Bora, N., Dodd, C. and Desmaures, N. (2015). *Diversity, dynamics and functional role of Actinomycetes on European smear ripened cheeses*: Springer.

- Broeker, J., Mechelke, M., Baudrexl, M., Mennerich, D., Hornburg, D., Mann, M. et al. (2018). The hemicellulose-degrading enzyme system of the thermophilic bacterium *Clostridium stercorarium*: comparative characterisation and addition of new hemicellulolytic glycoside hydrolases. *Biotechnology for Biofuels*, 11(1), 229.
- Calimlioglu, B. and Arga, K. Y. (2014). Proteins from halophilic bacteria: purification and their applications. In *Protein purification: principles and trends*: iConcept Press Ltd.
- Chauhan, P. S., Bharadwaj, A., Puri, N. and Gupta, N. (2014). Optimization of medium composition for alkali-thermostable mannanase production by *Bacillus nealsonii* PN-11 in submerged fermentation. *Int J Curr Microbiol Appl Sci*, 3(10), 1033-1045.
- Chauhan, P. S., Puri, N., Sharma, P. and Gupta, N. (2012). Mannanases: microbial sources, production, properties and potential biotechnological applications. *Applied Microbiology and Biotechnology*, 93(5), 1817-1830.
- Chekan, J. R., Kwon, I. H., Agarwal, V., Dodd, D., Revindran, V., Mackie, R. I. et al. (2014). Structural and biochemical basis for mannan utilization by *Caldanaerobius polysaccharolyticus* strain ATCC BAA-17. *Journal of Biological Chemistry*, 289(50), 34965-34977.
- Chen, H. (2014). Chemical composition and structure of natural lignocellulose. In *Biotechnology of lignocellulose* (pp. 25-71): Springer.
- Chen, Y. G., Zhang, Y. Q., Xiao, H. D., Liu, Z. X., Yi, L. B., Shi, J. X. et al. (2009). *Pontibacillus halophilus* sp. nov., a moderately halophilic bacterium isolated from a sea urchin. *International Journal of Systematic and Evolutionary Microbiology*, 59(7), 1635-1639.
- Chen, Y. G., Zhang, Y. Q., Yi, L. B., Li, Z. Y., Wang, Y. X., Xiao, H. D. et al. (2010). *Pontibacillus litoralis* sp. nov., a facultatively anaerobic bacterium isolated from a sea anemone and emended description of the genus *Pontibacillus*. *International Journal of Systematic and Evolutionary Microbiology*, 60(3), 560-565.
- Cheng, L. F., Duan, S. W., Feng, X. Y., Zheng, K., Yang, Q. and Liu, Z. C. (2016). Purification and characterization of a thermostable β -mannanase from *Bacillus subtilis* BE-91: potential application in inflammatory diseases. *BioMed Research International*, 2016.

- Cho, S. J. (2009). Isolation and characterization of mannanase producing *Bacillus amyloliquefaciens* CS47 from horse feces. *Journal of Life Science*, 19(12), 1724-1730.
- Cobucci-Ponzano, B., Strazzulli, A., Iacono, R., Masturzo, G., Giglio, R., Rossi, M. et al. (2015). Novel thermophilic hemicellulases for the conversion of lignocellulose for second generation biorefineries. *Enzyme and Microbial Technology*, 78, 63-73.
- Contesini, F. J., Liberato, M. V., Rubio, M. V., Calzado, F., Zubieta, M. P., Riaño-Pachón, D. M. et al. (2017). Structural and functional characterization of a highly secreted α -l-arabinofuranosidase (GH62) from *Aspergillus nidulans* grown on sugarcane bagasse. *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics*, 1865(12), 1758-1769.
- Da Cruz, A. F. (2013). Mannan-degrading enzyme system. In *Fungal enzymes*: Taylor & Francis Inc.
- David, A., Chauhan, P. S., Kumar, A., Angural, S., Kumar, D., Puri, N. et al. (2018). Coproduction of protease and mannanase from *Bacillus nealsonii* PN-11 in solid state fermentation and their combined application as detergent additives. *International Journal of Biological Macromolecules*, 108, 1176-1184.
- Debez, A., Belghith, I., Friesen, J., Montzka, C. and Elleuche, S. (2017). Facing the challenge of sustainable bioenergy production: Could halophytes be part of the solution? *Journal of Biological Engineering*, 11(1), 27.
- Dhawan, S. and Kaur, J. (2007). Microbial mannanases: an overview of production and applications. *Critical Reviews in Biotechnology*, 27(4), 197-216.
- Dutta, P., Deb, A. and Majumdar, S. (2016). Optimization of the medium for the production of extracellular amylase by the *Pseudomonas stutzeri* ISL B5 isolated from municipal solid waste. *International Journal of Microbiology*, 2016.
- El-Naggar, N., Deraz, S. and Khalil, A. (2014). Bioethanol production from lignocellulosic feedstocks based on enzymatic hydrolysis: Current status and recent developments. *Biotechnology*, 13(1), 1-21.
- El-refai, M. A., Khattab, O. K. H., Ismail, S. A., Hashem, A. M., Abo-Elnasr, A. A. and Nour, S. A. (2014). Improved mannanase production from *Penicillium humicola* and application for hydrolysis property. *Egyptian Pharmaceutical Journal*, 13(2), 160.

- Ergul, C. C. and Caliskan, E. (2018). Endospore formed bacteria and staining techniques. In *Science, ecology and engineering research in the globalizing world* (pp. 362).
- Fatima, S., Ajmal, R., Badr, G. and Khan, R. H. (2014). Harmful effect of detergents on lipase. *Cell Biochemistry and Biophysics*, 70(2), 759-763.
- Gao, Z. M., Ruan, L. W., Chen, X. L., Zhang, Y. Z. and Xu, X. (2010). A novel salt-tolerant endo- β -1,4-glucanase Cel5A in *Vibrio* sp. G21 isolated from mangrove soil. *Applied Microbiology and Biotechnology*, 87(4), 1373-1382.
- Ge, J. P., Du, R. P., Zhao, D., Song, G., Jin, M. and Ping, W. X. (2016). Kinetic study of a β -mannanase from the *Bacillus licheniformis* HDYM-04 and its decolorization ability of twenty-two structurally different dyes. *SpringerPlus*, 5(1), 1824.
- Giovanelli, E., Castellanos-Gomez, A. and Pérez, E. M. (2017). Surfactant-free polar-to-nonpolar phase transfer of exfoliated MoS₂ two-dimensional colloids. *ChemPlusChem*, 82(5), 732-741.
- Giuliano, C., Patel, C. R. and Kale-Pradhan, P. B. (2019). A guide to bacterial culture identification and results interpretation. *P & T*, 44(4), 192-200.
- Hadar, Y. (2013). Sources for lignocellulosic raw materials for the production of ethanol. In *Lignocellulose conversion* (pp. 21-38): Springer.
- Hakamada, Y., Ohkubo, Y. and Ohashi, S. (2014). Purification and characterization of β -mannanase from *Reinekea* sp. KIT-YO10 with transglycosylation activity. *Bioscience, Biotechnology and Biochemistry*, 78(4), 722-728.
- Hasan, F., Shah, A. A., Javed, S. and Hameed, A. (2010). Enzymes used in detergents: lipases. *African Journal of Biotechnology*, 9(31), 4836-4844.
- Hemmati, S., Seradj, H. and Mehrabi, N. (2017). Characterization of the lignin polymer in Brassicaceae family. *Research Journal of Pharmacognosy*, 4(2), 1-13.
- Higgins, D. and Dworkin, J. (2012). Recent progress in *Bacillus subtilis* sporulation. *FEMS Microbiology Reviews*, 36(1), 131-148.
- Huang, J., Qiao, Z. X., Tang, J. W. and Wang, G. (2015). High quality draft genome sequence of the moderately halophilic bacterium *Pontibacillus yanchengensis* Y32T and comparison among *Pontibacillus* genomes. *Standards in Genomic Sciences*, 10(1), 93.

- Isikgor, F. H. and Becer, C. R. (2015). Lignocellulosic biomass: a sustainable platform for the production of bio-based chemicals and polymers. *Polymer Chemistry*, 6(25), 4497-4559.
- Jamaludin, Z., Muhammad Salleh, M. and Yahya, A. (2017). Potential use of lignobiomass for sugar production. *J Appl Biotechnol Bioeng*, 3(6), 476-477.
- Jia, X., Mi, S., Wang, J., Qiao, W., Peng, X. and Han, Y. (2014). Insight into glycoside hydrolases for debranched xylan degradation from extremely thermophilic bacterium *Caldicellulosiruptor lactoaceticus*. *PloS One*, 9(9), e106482.
- Joe, M. M., Deivaraj, S., Benson, A., Henry, A. J. and Narendrakumar, G. (2018). Soil extract calcium phosphate media for screening of phosphate-solubilizing bacteria. *Agriculture and Natural Resources*, 52(3), 305-308.
- Kamal, M. Z., Yedavalli, P., Deshmukh, M. V. and Rao, N. M. (2013). Lipase in aqueous-polar organic solvents: activity, structure and stability. *Protein Science*, 22(7), 904-915.
- Kameshwar, A. K. S. and Qin, W. (2017). Qualitative and quantitative methods for isolation and characterization of lignin-modifying enzymes secreted by microorganisms. *BioEnergy Research*, 10(1), 248-266.
- Kansoh, A. L. and Nagieb, Z. A. (2004). Xylanase and mannanase enzymes from *Streptomyces galbus* NR and their use in biobleaching of softwood kraft pulp. *Antonie Van Leeuwenhoek*, 85(2), 103-114.
- Katrolia, P., Jia, H., Yan, Q., Song, S., Jiang, Z. and Xu, H. (2012). Characterization of a protease-resistant α -galactosidase from the thermophilic fungus *Rhizomucor miehei* and its application in removal of raffinose family oligosaccharides. *Bioresource Technology*, 110, 578-586.
- Kumar, S. and Sani, R. K. (2018). *Biorefining of biomass to biofuels*: Springer International Publishing
- Lal, M., Dutt, D., Kumar, A. and Gautam, A. (2015). Optimization of submerged fermentation conditions for two xylanase producers *Coprinellus disseminatus* MLK-01NTCC-1180 and MLK-07NTCC-1181 and their biochemical characterization. *Cellulose Chemistry and Technology*, 49(5-6), 471-483.
- Lam, M. Q., Nik Mut, N. N., Thevarajoo, S., Chen, S. J., Selvaratnam, C., Hussin, H. et al. (2018). Characterization of detergent compatible protease from halophilic *Virgibacillus* sp. CD6. *3 Biotech*, 8(2), 104.

- Lama, L., Calandrelli, V., Gambacorta, A. and Nicolaus, B. (2004). Purification and characterization of thermostable xylanase and β -xylosidase by the thermophilic bacterium *Bacillus thermantarcticus*. *Research in Microbiology*, 155(4), 283-289.
- Lee, J. C., Kim, Y. S., Yun, B. S. and Whang, K. S. (2015). *Pontibacillus salicampi* sp. nov., a moderately halophilic bacterium isolated from saltern soil. *International Journal of Systematic and Evolutionary Microbiology*, 65(2), 375-380.
- Lee, Y. S., Zhou, Y., Park, I. H., Chandra, M. R. G. S., Ahn, S. C. and Choi, Y. L. (2010). Isolation and purification of thermostable β -mannanase from *Paenibacillus illinoisensis* ZY-08. *Journal of the Korean Society for Applied Biological Chemistry*, 53(1), 1-7.
- Li, M., Pu, Y. and Ragauskas, A. J. (2016). Current understanding of the correlation of lignin structure with biomass recalcitrance. *Frontiers in Chemistry*, 4, 45.
- Li, T., Li, C. T., Butler, K., Hays, S. G., Guarneri, M. T., Oyler, G. A. et al. (2017). Mimicking lichens: incorporation of yeast strains together with sucrose-secreting cyanobacteria improves survival, growth, ROS removal and lipid production in a stable mutualistic co-culture production platform. *Biotechnol Biofuels* 10(1), 55.
- Liao, H., Li, S., Zheng, H., Wei, Z., Liu, D., Raza, W. et al. (2014). A new acidophilic thermostable endo-1,4- β -mannanase from *Penicillium oxalicum* GZ-2: cloning, characterization and functional expression in *Pichia pastoris*. *BMC Biotechnology*, 14(1), 90.
- Lim, J. M., Jeon, C. O., Park, D. J., Kim, H. R., Yoon, B. J. and Kim, C. J. (2005a). *Pontibacillus marinus* sp. nov., a moderately halophilic bacterium from a solar saltern and emended description of the genus *Pontibacillus*. *International Journal of Systematic and Evolutionary Microbiology*, 55(3), 1027-1031.
- Lim, J. M., Jeon, C. O., Song, S. M. and Kim, C. J. (2005b). *Pontibacillus chungwhensis* gen. nov., sp. nov., a moderately halophilic Gram-positive bacterium from a solar saltern in Korea. *International Journal of Systematic and Evolutionary Microbiology*, 55(1), 165-170.
- Liu, Q., Yang, P., Luo, H., Shi, P., Huang, H., Meng, K. et al. (2012). A novel endo-1,4- β -mannanase from *Bispora antennata* with good adaptation and stability

- over a broad pH range. *Applied Biochemistry and Biotechnology*, 166(6), 1442-1453.
- Luo, H., Wang, K., Huang, H., Shi, P., Yang, P. and Yao, B. (2012). Gene cloning, expression and biochemical characterization of an alkali-tolerant β -mannanase from *Humicola insolens* Y1. *Journal of Industrial Microbiology & Biotechnology*, 39(4), 547-555.
- Luo, Y., Li, Z., Li, X., Liu, X., Fan, J., Clark, J. H. et al. (2019). The production of furfural directly from hemicellulose in lignocellulosic biomass: a review. *Catalysis Today*, 319, 14-24.
- Madigan, M. T., Bender, K. S., Buckley, D. H., Parker, J. and Stahl, D. A. (2003). *Brock biology of microorganisms*: Upper Saddle River (NJ): Prentice-Hall, 2003.
- Mah, M. H. (2019). *Characterisation of xylanase from Microbulbifer sp. CL37*. Unpublished Master Thesis, Universiti Teknologi Malaysia, Skudai.
- Makela, M. R., Dilokpimol, A., Koskela, S. M., Kuuskeri, J., de Vries, R. P. and Hilden, K. (2018). Characterization of a feruloyl esterase from *Aspergillus terreus* facilitates the division of fungal enzymes from Carbohydrate Esterase family 1 of the carbohydrate-active enzymes (CAZy) database. *Microbial Biotechnology*, 11(5), 869-880.
- Michelin, M., Ruiz, H. A., Silva, D. P., Ruzene, D. S., Teixeira, J. A. and Polizeli, M. L. T. M. (2015). Cellulose from lignocellulosic waste. In *Polysaccharides: bioactivity and biotechnology* (pp. 475-511): Springer International Publishing.
- Mizutani, K., Tsuchiya, S., Toyoda, M., Nanbu, Y., Tominaga, K., Yuasa, K. et al. (2012). Structure of β -1,4-mannanase from the common sea hare *Aplysia kurodai* at 1.05 Å resolution. *Acta Crystallographica Section F: Structural Biology and Crystallization Communications*, 68(10), 1164-1168.
- Mohamad Roslan, M. A., Zainal Abidin, Z. Z. and Mohd Omar, S. (2016). Screening of ligninase-producing bacteria from south east Pahang peat swamp forest soil. *Malaysian Journal of Microbiology*, 12(6), 433-437.
- Morales-Delarosa, S. and Campos-Martin, J. M. (2014). Catalytic processes and catalyst development in biorefining. In *Advances in biorefineries* (pp. 152-198): Elsevier.

- Moreira, L. R. S. (2008). An overview of mannan structure and mannan-degrading enzyme systems. *Applied Microbiology and Biotechnology*, 79(2), 165.
- Mou, H., Zhou, F., Jiang, X. and Liu, Z. (2011). Production, purification and properties of β -mannanase from soil bacterium *Bacillus circulans* m-21. *Journal of Food Biochemistry*, 35(5), 1451-1460.
- Muhammad, A., Bokhari, S. A. I., Vernoux, J. P., Ali, M. I., Faryal, R., Desmaures, N. et al. (2019). Purification, characterization and thermodynamic assessment of an alkaline protease by *Geotrichum Candidum* of dairy origin. *Iranian Journal of Biotechnology*, 17(2).
- Mussatto, S. I. and Teixeira, J. A. (2010). Lignocellulose as raw material in fermentation processes. In *Current research, technology and education topics in applied microbiology and microbial biotechnology* (pp. 897-907): Formatex Research Center.
- Nadaroglu, H. and Dikbas, N. (2018). Purification and characterization of a β -mannanase from *Lactobacillus plantarum* (ATCC® 14917TM). *International Journal of Innovative Research and Reviews*, 2(1), 1-6.
- Nagar, S., Mittal, A. and Gupta, V. K. (2012). Enzymatic clarification of fruit juices (apple, pineapple and tomato) using purified *Bacillus pumilus* SV-85S xylanase. *Biotechnology and Bioprocess Engineering*, 17(6), 1165-1175.
- Nakamura, A. M., Nascimento, A. S. and Polikarpov, I. (2017). Structural diversity of carbohydrate esterases. *Biotechnology Research and Innovation*, 1(1), 35-51.
- Panda, T. and Gowrishankar, B. S. (2005). Production and applications of esterases. *Applied Microbiology and Biotechnology*, 67(2), 160-169.
- Pangsri, P., Piwpankaew, Y., Ingkakul, A., Nitisinprasert, S. and Keawsompong, S. (2015). Characterization of mannanase from *Bacillus circulans* NT 6.7 and its application in mannooligosaccharides preparation as prebiotic. *SpringerPlus*, 4(1), 771.
- Pletnev, P., Osterman, I., Sergiev, P., Bogdanov, A. and Dontsova, O. (2015). Survival guide: *Escherichia coli* in the stationary phase. *Acta Naturae*, 7(4), 22-33.
- Poli, A., Finore, I., Tramice, A., Di Donato, P., Nicolaus, B. and Lama, L. (2016). Technical developments for vegetable waste biomass degradation by thermophiles. In *Biotechnology of extremophiles* (pp. 539-579): Springer.
- Pradeep, G. C., Cho, S. S., Choi, Y. H., Choi, Y. S., Jee, J. P., Seong, C. N. et al. (2016). An extremely alkaline mannanase from *Streptomyces* sp. CS428

- hydrolyzes galactomannan producing series of mannoooligosaccharides. *World Journal of Microbiology and Biotechnology*, 32(5), 84.
- Puchart, V., Vrsanska, M., Svoboda, P., Pohl, J., Ogel, Z. B. and Biely, P. (2004). Purification and characterization of two forms of endo- β -1,4-mannanase from a thermotolerant fungus, *Aspergillus fumigatus* IMI 385708 (formerly *Thermomyces lanuginosus* IMI 158749). *Biochimica et Biophysica Acta* 1674(3), 239-250.
- Putro, J. N., Soetaredjo, F. E., Lin, S. Y., Ju, Y. H. and Ismadji, S. (2016). Pretreatment and conversion of lignocellulose biomass into valuable chemicals. *RSC Advances*, 6(52), 46834-46852.
- Rahmani, N., Amaniyah, M. and Djohan, A. C. (2017). Cloning of partial β -mannanase gene from Indonesia marine bacteria *Bacillus Subtilis* LBF-005. *Squalen Bulletin of Marine and Fisheries Postharvest and Biotechnology*, 12(1), 19-27.
- Rattanasuk, S. and Ketudat-Cairns, M. (2009). *Chryseobacterium indologenes*, novel mannanase-producing bacteria. *Songklanakarin Journal of Science & Technology*, 31(4).
- Reiner, K. (2010). Catalase test protocol. *American Society For Microbiology*.
- Reynolds, J., Moyes, R. and Breakwell, D. P. (2009). Differential staining of bacteria: endospore stain. *Current Protocols in Microbiology*, Appendix 3, Appendix 3J.
- Robak, K. and Balcerek, M. (2018). Review of second generation bioethanol production from residual biomass. *Food Technology and Biotechnology*, 56(2), 174.
- Rodloff, A., Bauer, T., Ewig, S., Kujath, P. and Müller, E. (2008). Susceptible, intermediate and resistant—the intensity of antibiotic action. *Deutsches Ärzteblatt International*, 105(39), 657.
- Saha, B. C. (2003). Hemicellulose bioconversion. *Journal of Industrial Microbiology and Biotechnology*, 30(5), 279-291.
- Saini, J. K., Saini, R. and Tewari, L. (2015). Lignocellulosic agriculture wastes as biomass feedstocks for second-generation bioethanol production: concepts and recent developments. *3 Biotech*, 5(4), 337-353.
- Sakthivel, M., Karthikeyan, N., Jayaveny, R. and Palani, P. (2010). Optimization of culture conditions for the production of extracellular cellulase from *Corynebacterium lipophiloflavum*. *Journal of Ecobiotechnology*.

- Salwoom, L., Raja Abd Rahman, R. N. Z., Salleh, A. B., Mohd Shariff, F., Convey, P. and Mohamad Ali, M. S. (2019). New recombinant cold-adapted and organic solvent tolerant lipase from psychrophilic *Pseudomonas* sp. LSK25, isolated from Signy Island Antarctica. *International Journal of Molecular Sciences*, 20(6), 1264.
- Shaheen, S., Aman, A. and Siddiqui, N. N. (2017). Influence of metal ions, surfactants and organic solvents on the catalytic performance of levansucrase from *Zymomonas mobilis* KIBGE-IB14. *Journal of Basic and Applied Sciences*, 13, 41-46.
- Shi, H., Huang, Y., Zhang, Y., Li, W., Li, X. and Wang, F. (2013). High-level expression of a novel thermostable and mannose-tolerant β -mannosidase from *Thermotoga thermarum* DSM 5069 in *Escherichia coli*. *BMC Biotechnology*, 13(1), 83.
- Shields, P. and Cathcart, L. (2010). Oxidase test protocol. *American Society For Microbiology*
- Shukla, P. and Pletschke, B. I. (2013). *Advances in enzyme biotechnology*: Springer.
- Simões, L. C., Silva, R. R., Nascimento, C. E., Boscolo, M., Gomes, E. and Silva, R. (2019). Purification and physicochemical characterization of a novel thermostable xylanase secreted by the fungus *Myceliophthora heterothallica* F.2.1.4. *Applied Biochemistry and Biotechnology*, 188(4), 991-1008.
- Singh, G. and Hoondal, G. S. (2011). Biobleaching of wheat straw richsoda pulp by the application of thermophilic mannanase from *Bacillus* sp. MG33. *International Journal of Environmental Sciences*
- Sinha, R. and Khare, S. K. (2014). Effect of organic solvents on the structure and activity of moderately halophilic *Bacillus* sp. EMB9 protease. *Extremophiles*, 18(6), 1057-1066.
- Soumya, R. S. and Abraham, E. T. (2010). Isolation of β -mannanase from *Cocos nucifera* Linn haustorium and its application in the depolymerization of β -(1,4)-linked D-mannans. *International Journal of Food Sciences and Nutrition*, 61(3), 272-281.
- Srivastava, P. K. and Kapoor, M. (2017). Production, properties and applications of endo- β -mannanases. *Biotechnology Advances*, 35(1), 1-19.
- Sriyapai, P., Kawai, F., Siripoke, S., Chansiri, K. and Sriyapai, T. (2015). Cloning, expression and characterization of a thermostable esterase HydS14 from

- Actinomadura* sp. strain S14 in *Pichia pastoris*. *International Journal of Molecular Sciences*, 16(6), 13579-13594.
- Sun, F., Wang, B., Du, Y., Liu, X., Lai, Q., Li, G. et al. (2010). *Arenibacter nanhaiticus* sp. nov., isolated from marine sediment of the South China Sea. *International Journal of Systematic and Evolutionary Microbiology*, 60(1), 78-83.
- Suresh, C., Kitaoka, M. and Hayashi, K. (2003). A thermostable non-xylanolytic α -glucuronidase of *Thermotoga maritima* MSB8. *Bioscience, Biotechnology and Biochemistry*, 67(11), 2359-2364.
- Suwanto, A., Thenawidjaja, M. and Purwadaria, T. (2005). Isolation and characterization of mannanolytic thermophilic bacteria from palm oil shell and their mannanase enzyme production properties. *Biotropia: The Southeast Asian Journal of Tropical Biology*(25).
- Suzuki, K., Michikawa, M., Sato, H., Yuki, M., Kamino, K., Ogasawara, W. et al. (2017). Purification, cloning, functional expression, structure and characterization of a thermostable β -Mannanase from *Talaromyces trachyspermus* B168 and its efficiency in production of mannoooligosaccharides from coffee wastes. *Journal of Applied Glycoscience*, jag. JAG-2017_2018.
- Tamaru, Y., Araki, T., Amagoi, H., Mori, H. and Morishita, T. (1995). Purification and characterization of an extracellular beta-1,4-mannanase from a marine bacterium, *Vibrio* sp. strain MA-138. *Appl. Environ. Microbiol.*, 61(12), 4454-4458.
- Trinh, D. K., Quyen, D. T., Do, T. T. and Nghiem, N. M. (2013). Purification and characterization of a novel detergent-and organic solvent-resistant endo-beta-1,4-glucanase from a newly isolated basidiomycete *Peniophora* sp. NDVN01. *Turkish Journal of Biology*, 37(4), 377-384.
- Virk, A. P., Sharma, P. and Capalash, N. (2012). Use of laccase in pulp and paper industry. *Biotechnology Progress*, 28(1), 21-32.
- Wang, J., Shao, Z., Hong, Y., Li, C., Fu, X. and Liu, Z. (2010). A novel β -mannanase from *Pantoea agglomerans* A021: gene cloning, expression, purification and characterization. *World Journal of Microbiology and Biotechnology*, 26, 1777-1784.
- Wang, J., Zeng, D., Liu, G., Wang, S. and Yu, S. (2014). Truncation of a mannanase from *Trichoderma harzianum* improves its enzymatic properties and

- expression efficiency in *Trichoderma reesei*. *Journal of Industrial Microbiology & Biotechnology*, 41(1), 125-133.
- Wang, M., Li, Z., Fang, X., Wang, L. and Qu, Y. (2012). Cellulolytic enzyme production and enzymatic hydrolysis for second-generation bioethanol production. In *Biotechnology in China III: biofuels and bioenergy* (pp. 1-24): Springer.
- Wang, M., You, S., Zhang, S., Qi, W., Liu, Z., Wu, W. et al. (2013). Purification, characterization and production of β -mannanase from *Bacillus subtilis* TJ-102 and its application in gluco-mannooligosaccharides preparation. *European Food Research and Technology*, 237(3), 399-408.
- Widdel, F. (2007). Theory and measurement of bacterial growth. *Di dalam Grundpraktikum Mikrobiologie*, 4(11), 1-11.
- Wu, J., Qiu, C., Ren, Y., Yan, R., Ye, X. and Wang, G. (2018). Novel salt-tolerant xylanase from a mangrove-isolated fungus *Phoma* sp. MF13 and its application in chinese steamed bread. *ACS Omega*, 3(4), 3708-3716.
- Xia, W., Lu, H., Xia, M., Cui, Y., Bai, Y., Qian, L. et al. (2016). A novel glycoside hydrolase family 113 endo- β -1,4-mannanase from *Alicyclobacillus* sp. strain A4 and insight into the substrate recognition and catalytic mechanism of this family. *Appl. Environ. Microbiol.*, 82(9), 2718-2727.
- Yamabhai, M., Sak-Ubol, S., Srila, W. and Haltrich, D. (2016). Mannan biotechnology: from biofuels to health. *Critical Reviews in Biotechnology*, 36(1), 32-42.
- Yang, H., Shi, P., Lu, H., Wang, H., Luo, H., Huang, H. et al. (2015). A thermophilic β -mannanase from *Neosartorya fischeri* P1 with broad pH stability and significant hydrolysis ability of various mannan polymers. *Food Chemistry*, 173, 283-289.
- Yang, X., Ma, R., Shi, P., Huang, H., Bai, Y., Wang, Y. et al. (2014). Molecular characterization of a highly-active thermophilic β -glucosidase from *Neosartorya fischeri* P1 and its application in the hydrolysis of soybean isoflavone glycosides. *PLoS One*, 9(9), e106785.
- Yang, Y., Zou, Z., He, M. and Wang, G. (2011). *Pontibacillus yanchengensis* sp. nov., a moderately halophilic bacterium isolated from salt field soil. *International Journal of Systematic and Evolutionary Microbiology*, 61(8), 1906-1911.

- Yildiz, S. Y. and Oner, E. T. (2014). Mannan as a promising bioactive material for drug nanocarrier systems. *Application of Nanotechnology in Drug Delivery*, 9, 311-342.
- Yin, L. J., Tai, H. M. and Jiang, S. T. (2012). Characterization of mannanase from a novel mannanase-producing bacterium. *Journal of Agricultural and Food Chemistry*, 60(25), 6425-6431.
- Yoo, H. Y., Pradeep, G. C., Kim, S. W., Park, D. H., Choi, Y. H., Suh, J. W. et al. (2015). A novel low-molecular weight alkaline mannanase from *Streptomyces tendae*. *Biotechnology and Bioprocess Engineering*, 20(3), 453-461.
- Yoshida, M., Igarashi, K., Kawai, R., Aida, K. and Samejima, M. (2004). Differential transcription of β -glucosidase and cellobiose dehydrogenase genes in cellulose degradation by the basidiomycete *Phanerochaete chrysosporium*. *FEMS Microbiology Letters*, 235(1), 177-182.
- You, J., Liu, J. F., Yang, S. Z. and Mu, B. Z. (2016). Low-temperature-active and salt-tolerant β -mannanase from a newly isolated *Enterobacter* sp. strain N18. *Journal of Bioscience and Bioengineering*, 121(2), 140-146.
- You, X., Qin, Z., Yan, Q., Yang, S., Li, Y. and Jiang, Z. (2018). Structural insights into the catalytic mechanism of a novel glycoside hydrolase family 113 β -1,4-mannanase from *Amphibacillus xylanus*. *Journal of Biological Chemistry*, 293(30), 11746-11757.
- Zeng, Y., Himmel, M. E. and Ding, S. Y. (2017). Visualizing chemical functionality in plant cell walls. *Biotechnology for Biofuels*, 10(1), 263.
- Zhang, R., Song, Z., Wu, Q., Zhou, J., Li, J., Mu, Y. et al. (2016). A novel surfactant-, NaCl- and protease-tolerant β -mannanase from *Bacillus* sp. HJ14. *Folia Microbiologica*, 61(3), 233-242.
- Zhao, R., Zhao, R., Tu, Y., Zhang, X., Deng, L. and Chen, X. (2018). A novel α -galactosidase from the thermophilic probiotic *Bacillus coagulans* with remarkable protease-resistance and high hydrolytic activity. *PloS One*, 13(5), e0197067.
- Zhao, X. Q., Zi, L. H., Bai, F. W., Lin, H. L., Hao, X. M., Yue, G. J. et al. (2011a). Bioethanol from lignocellulosic biomass. In *Biotechnology in China III: biofuels and bioenergy* (pp. 25-51): Springer.

- Zhao, Y., Zhang, Y., Cao, Y., Qi, J., Mao, L., Xue, Y. et al. (2011b). Structural analysis of alkaline β -mannanase from alkaliphilic *Bacillus* sp. N16-5: implications for adaptation to alkaline conditions. *PLoS One*, 6(1), e14608.
- Zhou, C., Xue, Y. and Ma, Y. (2018). Characterization and high-efficiency secreted expression in *Bacillus subtilis* of a thermo-alkaline β -mannanase from an alkaliphilic *Bacillus clausii* strain S10. *Microbial Cell Factories*, 17(1), 124.
- Zhou, J., Zhang, R., Gao, Y., Li, J., Tang, X., Mu, Y. et al. (2012). Novel low-temperature-active, salt-tolerant and proteases-resistant endo-1,4- β -mannanase from a new *Sphingomonas* strain. *Journal of Bioscience and Bioengineering*, 113(5), 568-574.
- Zyl, W. H. V., Rose, S. H., Trollope, K. and Görgens, J. F. (2010). Fungal β -mannanases: mannan hydrolysis, heterologous production and biotechnological applications. *Process Biochemistry*, 45(8), 1203-1213.