# BIODEGRADATION OF AR-27 DYE AND ELECTRICITY GENERATION BY Enterobacter cloacae NF2015

NOOR FATEEN AFIKAH BINTI YAHYA

A thesis submitted in fulfilment of the requirements for the award of the degree of Master of Philosophy

> Faculty of Science Universiti Teknologi Malaysia

> > SEPTEMBER 2019

### DEDICATION

Dedicated to all people that I treasure most in life...

### To my Beloved Father and Mother

Yahya bin Abdullah and Norhayati binti Othman

# To my Beloved Father and Mother in Law

Mohamad bin Jusoh and Zainab binti Ali

## my Beloved Husband and daughter

Ahmad Zainal bin Mohamad and Nuur Hasya Ammara Hawani bt Ahmad Zainal

To my respectful supervisor

Dr. Norahim bin Ibrahim

#### ACKNOWLEDGEMENT

In the name of Allah s.w.t the Most Beneficent and the Most Merciful, who provides wisdom to face all the challenges throughout the completion of this master study.

In preparing this thesis, I was in contact with many people, researchers, academicians, and practitioners. They have contributed towards my understanding and thoughts. In particular, I wish to express my sincere appreciation to my main thesis supervisor, Dr. Norahim bin Ibrahim, for encouragement, guidance, critics and friendship. Without their continued support and interest, this thesis would not have been the same as presented here.

I am also indebted to Jabatan Perkhidmatan Awam (JPA) for funding my master study. My deepest appreciation is also extended to my husband who provide me physical and mental supports. Warmest gratitude to both of my parent who never failed to encourage me to pursue this study. I would also wish to convey my sincere gratitude to the Faculty of Science for providing me with all the necessary instruments and facilities. Thanks to all lab assistants for their kind assistance throughout this study. A special recognition goes to Dr. Bushra Ismail from Petroleum Research Centre, Iraq who did the isolation of all the bacteria used in this study.

My fellow postgraduate student should also be recognized for their support. My sincere appreciation also extends to all my colleagues Ms. Birintha Ganapathy, Mr. Muhamad Firdaus bin Sabaruddin, Mrs. Ameera Syaheerah bt Abdul Aziz, Dr. Mohamad Hanif Md Nor and Mr. Mohamad Fahmi Muhamad Mubarak who have provided assistance at various occasions. Their views and tips are useful indeed. Finally, I want to say thank you to all my friends in Fs for all their suggestions, helps, ideas, and sincere friendship.

#### ABSTRACT

One of the major concerns of developing countries worldwide is wastewater treatment and renewable energy generation. Wastewater by-product from production activity is the cause of pollution by industries. Microbial degradation is considered a good alternative to replace wastewater treatment approaches using physical and chemical method that are less desirable due to high costs and sludge production. This study was carried out to screen and identify the potential of unidentified bacterial strains POS, F2B, PCO OIL, and B1, previously isolated from Iraqi crude oil reservoir to decolorize azo dye and implementing the system in Microbial Fuel Cell (MFC) for bioelectricity generation. The selected bacterium was identified using 16S rDNA method. The effect of co-substrates was studied to determine the minimal conditions required for maximum decolorization of AR-27 dye. Analysis of degradation were performed using UV-Vis, FTIR, CV, COD, DNS, TPP content and HPLC under facultative anaerobic and sequential facultative anaerobic-aerobic condition. Bioelectricity study was conducted using two-chambered MFC using agar salt bridge and graphite felt electrodes. The effect of different diameter of salt bridge and electrode size to electricity generation was studied. Bacteria B1 identified as Enterobacter cloacae NF2015 was found to be the most dominant decolorizing bacteria that could decolorize 99 % of 100 mg/L of AR-27 within 2 h under facultative anaerobic condition, pH 7.0, and temperature of 29 °C  $\pm$  2. Yeast extract and glucose at 0.5 g/L each was found to be the optimum co-substrates for AR-27 decolorization. The disappearance of peak in the UV-Vis spectra at 521 nm indicated the biodegradation of azo dye. The presence of oxidation and reduction peak by cyclic voltammetry analysis showed an irreversible redox reaction during the degradation. FTIR-ATR analysis confirmed that the azo linkage was cleaved after decolorization had occurred. The reduction in concentration of metabolite catechol detected by HPLC analysis confirmed a successful degradation of AR-27 dye. There was 87.6 % and 100 % COD removal after 72 hours treatment in facultative anaerobic and facultative anaerobic-aerobic condition, respectively. Optimization of the MFC voltage using 4.5 cm diameter salt bridge and 25 cm<sup>2</sup> electrode surface area showed that the maximum OCV and CCV obtained were 809.7  $\pm$  12 mV and 108.3  $\pm$  19 mV, with current density and power density at 1.93  $\pm$  0.3  $mA/m^2$  and 0.21  $\pm$  0.04 mW/m<sup>2</sup>. SEM and FTIR analysis revealed biofilm and extracellular polymeric substances (EPSs) functional groups development on the electrode surface during MFC operation. In conclusion, bacteria E. cloacae NF2015 from crude oil reservoir has the potential to be used in simultaneous azo dye wastewater treatment and bioelectricity generation by MFC technologies.

#### ABSTRAK

Salah satu kebimbangan utama negara-negara membangun di seluruh dunia ialah rawatan air sisa dan penjanaan tenaga boleh diperbaharui. Air sisa yang terhasil aktiviti pengeluaran merupakan penyebab pencemaran daripada daripada perindustrian. Degradasi mikrob dianggap alternatif yang baik untuk menggantikan pendekatan semasa rawatan air sisa menggunakan kaedah fizikal dan kimia yang kurang diidami akibat kos tinggi dan pengeluaran enapcemar. Kajian ini dijalankan untuk menilai dan mengenal pasti potensi bakteria-bakteria yang belum dikenalpasti iaitu POS, F2B, PCO OIL, dan B1 yang dipencil daripada minyak mentah Iraq untuk menghuraikan pewarna azo dan seterusnya melaksanakan sistem tersebut dalam Sel Bahan Api Mikrob (MFC) untuk penjanaan bioelektrik. Bakteria yang dipilih telah dikenalpasti menggunakan kaedah rDNA 16S. Kesan substrat bersama telah dikaji untuk menentukan keadaan minimum yang diperlukan untuk penghuraian pewarna AR-27 oleh bakteria. Analisis degradasi telah dilakukan menggunakan UV-Vis, FTIR, CV, COD, DNS, kandungan TPP dan HPLC di bawah keadaan anaerobik fakultatif anaerobik fakultatif-aerobik berturutan. Kajian bioelektrik dijalankan dan menggunakan MFC dua ruang dengan jambatan garam dan elektrod karbon. Kesan diameter jambatan garam dan saiz elektrod yang berbeza kepada penjanaan elektrik telah dikaji. Bakteria B1 dikenalpasti sebagai Enterobacter cloacae NF2015 yang didapati sebagai penghurai paling dominan dapat menjernihkan 100 mg/L AR-27 sebanyak 99 % dalam masa 2 jam di bawah keadaan anaerobik fakultatif, pH 7.0, dan suhu 29 °C  $\pm$  2. Ekstrak yis dan glukosa masing-masing pada 0.5 g/L didapati substrat yang optimum untuk penghuraian AR-27. Kehilangan puncak dalam spektrum UV-Vis 521 nm menunjukkan biodegradasi pewarna azo. Kehadiran puncak pengoksidaan dan penurunan yang dikesan dalam analisis kitaran voltametri menunjukkan reaksi redoks tidak terbalikan berlaku semasa degradasi. Analisis FTIR-ATR mengesahkan bahawa rangkaian azo telah dipecahkan selepas penjernihan berlaku. Pengurangan kepekatan metabolit katekol yang dikesan oleh analisis HPLC mengesahkan degradasi pewarna AR-27. Terdapat 87.6 % dan 100 % penyingkiran COD selepas 72 jam rawatan dalam keadaan anaerobik fakultatif dan anaerobik-aerobik fakultatif. Pengoptimuman voltan MFC menggunakan jambatan garam berdiameter 4.5 cm dan luas permukaan elektrod 25 cm<sup>2</sup> menunjukkan maksimum OCV dan CCV iaitu 809.7  $\pm$  12 mV dan 108.3  $\pm$  19 mV, kepadatan arus dan ketumpatan kuasa sebanyak 1.93  $\pm$ 0.3 mA/m<sup>2</sup>, dan 0.21  $\pm$  0.04 mW/m<sup>2</sup>, masing-masing. Analisis SEM dan FTIR mendedahkan pembentukan kumpulan berfungsi biofilm dan bahan polimer ekstrasel (EPS) pada permukaan elektrod semasa operasi MFC. Sebagai kesimpulan, bakteria E. cloacae NF2015 dari takungan minyak mentah mempunyai potensi untuk digunakan dalam rawatan air sisa pewarna azo serentak dengan penjanaan bioelektrik oleh teknologi MFC.

# TABLE OF CONTENTS

# TITLE

DECLARATION	ii
DEDICATION	iii
ACKNOWLEDGEMENT	iv
ABSTRACT	V
ABSTRAK	vi
TABLE OF CONTENTS	vii
LIST OF TABLES	xiv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xxiii
LIST OF SYMBOLS	XXV
LIST OF APPENDICES	xxvii
<b>ER 1 INTRODUCTION</b>	1

# **CHAPTER 1 INTRODUCTION**

	1.1	Research Background	1
	1.2	Problem Statement	3
	1.3	Research Objectives	5
	1.4	Research Scope	6
	1.5	Significance of Research	7
CHAPTER 2	TTT	ERATURE REVIEW	9
		Г, КАТЦКГ, КГ, VIГ, VV	· · ·
			,
	2.1	Textile Wastewater	9
		Textile Wastewater	9
	2.1	Textile Wastewater	
	2.1 2.2	Textile Wastewater Azo Dyes in Textile Wastewater	9 10

	2.6 Decolorization of Azo Dye using Single Culture Bacteri	a 20
	2.7 Mechanism of Azo Dye Decolorization by Bacteria	26
	2.7.1 Azo dye Decolorization under Aerobic Condition	28
	2.7.2 Azo dye Decolorization under Anaerobic Condition	28
	2.8 Biodegradation by Sequential Facultative Aaerobic-Aer	obic
	Condition	31
	2.9 Factors Affecting Decolorization by Bacteria	32
	2.9.1 Effect of Aeration of Oxygen	33
	2.9.2 Effect of Nitrogen and Carbon Sources	33
	2.10 Microbial Fuel Cell: An Overview	34
	2.11 Mechanism of Bacteria Electron Transfer in MFC	38
	2.11.1 Anode Chamber	39
	2.11.2 Biofilm Formation at Anode	40
	2.12 Factor Affecting MFC Performance	41
	2.12.1 External Load	46
	2.12.2 Salt Bridge in MFC	47
	2.12.3 Electrode in MFC	51
	2.13 Bacteria from Crude Oil Reservoir	52
	2.13.1 Enterobacter cloacae Bacteria	53
	2.13.2 Bioelectricity Generation by <i>Enterobacter cloacae</i> bacteria	56
	2.14 Summary	58
CHAPTER 3	METHODOLOGY	59
	3.1 Experimental Design	59
	3.2 Media Preparation	61
	3.2.1 Nutrient Agar	61
	3.2.2 Nutrient Broth	61

3.2.3	Chemically Defined Medium with Modification	61
2.2		
3.3	Preparation of Stock Solutions	62
3.3.1	Dye Solution	62
3.3.2	Glucose and Yeast Extract	63
3.3.3	Chemical Oxygen Demand (COD) Reagent	64
3.3.4	Dinitrosalicylic Acid (DNS) Reagent	64
3.4	Bacterial Stock Culture	64
3.5	Screening of AR-27 Dye Decolorizing Bacteria	66
3.5.1	Screening using Mixed Culture	66
3.5.2	Screening using Pure Culture	68
3.6	Identification of Selected Dye-decolorizing Bacterial Stra	ain
		68
3.6.1	Biochemical Analysis	68
3.6.2	Molecular Identification	69
	3.6.2.1 Genomic DNA Extraction	69
	3.6.2.2 Polymerase Chain Reaction (PCR)	70
	3.6.2.3 16S rRNA Gene Sequence Analysis	71
3.7	Optimization of Co-substrates for AR-27 Decolorization	71
3.7.1	Screening of Nitrogen and Carbon Sources	72
3.7.2	Determination of Dye Removal from Standard Curve of AR-27 Dye	73
3.7.3	Dry Cell Weight	73
3.7.4	Indirect Determination of Bacteria Concentration	74
3.8	Biodegradation Analysis of AR-27 Decolorization under	
	Facultative Anaerobic and Sequential Facultative	
	Anaerobic-aerobic Conditions	74
3.8.1	Determination of AR-27 Dye Degradation	76
	3.8.1.1 UV-Vis Analysis	76

	3.8.1.2	HighPerformanceLiquidChromatography (HPLC)Analysis	76
	3.8.1.3	Determination of redox reaction by Cyclic Voltammetry (CV) Analysis	76
	3.8.1.4	Determination of Chemical Oxygen Demand (COD)	77
	3.8.1.5	Determination of Reducing Sugar by Dinitrosalicylic acid (DNS) Analysis	78
	3.8.1.6	Determination of Total Polyphenolic Content (TPC)	79
3.9	Bioelectric	city Generation Experiment using Selected	
	Bacteria S	train in modified CDM.	79
3.9.1	MFC Set	t Up	79
3.9.2	Preparati	ion of MFC Operation	80
	3.9.2.1	Bacterial Culture	80
	3.9.2.2	Preparation of CDM	81
	3.9.2.3	Preparation of Phosphate Buffer Solution	81
	3.9.2.4	Preparation of Salt Bridge	82
3.9.3	MFC Op	eration	82
3.9.4	Data Col	lection for MFC	83
3.9.5	Analysis	of Parameter	84
	3.9.5.1	External Load	84
	3.9.5.2	Diameter of Salt Bridge	85
	3.9.5.3	Size of Electrode	85
3.9.6	Bioelecti	ricity Analysis	86
3.9.7	Polarizat	ion Curve	86
3.9.8	Statistica	ıl Analysis	87
3.9.9	Characte Anode E	rization of Biofilm Formation on lectrode	87
	3.9.9.1	Pretreatment of MFC Anode	87
	3.9.9.2	Scanning Electron Microscope (SEM)	87

		3.9.9.3	Fourier Spectrosc	Transformed opy (FTIR)	Infrared	88
CHAPTER 4	Selec	tion and ic	lentificatio	n of Bacterial C	ulture	
	Colle	cted from	Iraqi Cruo	le Oil Reservoir	for	
	Deco	lorization	of Syntheti	c Azo Dye (AR-	27)	
	Wast	ewater.				89
	4.1	Introductio	on			89
	4.2	Methodolo	ogy			89
	4.3	Results an	d Discussio	ons		90
	4.3.1	Selectior	n of AR-27	dye decolorizing	bacteria	90
	4.3.2	Identifica bacteria	ation of s	elected dye de	colorizing	95
		4.3.2.1	Identificat morpholo characteri	gical and bi	eria by ochemical	95
		4.3.2.2		tion of bacteria	U	96
	4.4	Summary				101
CHAPTER 5	Asses	sment of A	AR-27 Dye	<b>Decolorization</b>	by the	
	Selec	ted Bacter	ia Strain T	hrough Optimiz	zation of	
	Co-su	ibstrates a	and Analys	is of Dye Degrad	lation in	
	CDM	[				103
	5.1	Introduction	on			103
	5.2	Methodolo	ogy			103
	5.3	Results an	d Discussio	ons		105
	5.3.1	Effect of 27 by <i>E</i> .		es to decolorizati	on of AR-	105
		5.3.1.1	Effect o decoloriza	of nitrogen so ation of AR-27	ource to	105
		5.3.1.2		concentration of decolorization of	-	107

		5.3.1.3	Effect of type of carbon source to decolorization of AR-27	111
		5.3.1.4	Effect of concentration of carbon source to decolorization of AR-27	112
		5.3.1.5	Percentage of dye removal and dry cell weight in control CDM without wo-substrate, unoptimized CDM, and optimized CDM.	117
	5.3.2	facultativ	dation analysis of AR-27 under ve anaerobic and sequential facultative c-aerobic conditions	119
		5.3.2.1	Determination of chemically oxygen demand (COD)	121
		5.3.2.2	Determination of AR-27 dye degradation by UV-Vis	123
		5.3.2.3	Determination of AR-27 dye degradation by FTIR Spectroscopy Analysis	124
	5.3.3		nation of redox reaction by Cyclic netry (CV)	128
		5.3.3.1	Determination of reducing sugar by DNS Analysis	131
		5.3.3.2	Determination of Total Polyphenolic Content (TPPC)	133
		5.3.3.3	Determination of catechol	135
	5.4	Summary		140
CHAPTER 6	Select	ted Bacter	f Bioelectricity Generation by ia using Different Salt Bridge Electrode Size in Double-chambered	
	MFC	•		141
	6.1	Introductio	on	141
	6.2	Methodolo	ogy	143
	6.3	Results an	d Discussions	144
	6.3.1	Evaluatio <i>cloacae</i> 1	on of electricity generation by <i>E</i> . NF2015	144

	6.3.2	Open cir	cuit voltage of non-optimized MFC	144
	6.3.3	Effect of	external loads in MFC	147
	6.3.4	Effect of	diameter of salt bridge in MFC	150
	6.3.5	Effect of in MFC	surface area of graphite felt electrode	155
	6.3.6	Polarizat	ion Curve of optimized MFC	159
	6.3.7	Cyclic V	oltammetry	162
	6.3.8	Characte electrode	rization of biofilm formation on anode	164
		6.3.8.1	Biofilm image characterization by SEM	164
		6.3.8.2	FTIR Analysis	167
	6.3.9	COD Re	moval	169
	6.4	Summary		170
CHAPTER 7	CON	CLUSION	NAND RECOMMENDATIONS	173
	7.1	Conclusio	n	173
	7.2	Contributi	on to knowledge	174
	7.3	Future Wo	orks	174
REFERENCES				177
List of publicatio	ns:			217

### LIST OF TABLES

TABLE NO.	TITLE	PAGE
Table 2.1	Structure-related toxicity of different type of azo dyes	13
Table 2.2	Detail of Amaranth dye (Sabnis R.W., 2010; Handbook of Biological Dyes and Stains	15
Table 2.3	Summary of the several treatment methods for the dye removal from wastewater effluent (Saratale <i>et al.</i> , 2011; Holkar <i>et al.</i> , 2016; Al-Mamun <i>et al.</i> , 2019)	18
Table 2.4	Summary of different type of azo dyes decolorization by pure bacterial strain.	22
Table 2.5	List of aromatic amines according to the EU Directive 2002/61/EC	31
Table 2.6	Factors affecting the performance of MFCs	43
Table 2.7	Summary review of previous salt bridge MFC studies	49
Table 2.8	Summary review of previous studies using bacteria E. cloacae for azo dye degradation	55
Table 2.9	Summary review of previous MFC studies using bacteria E. cloacae	57
Table 3.1	Original CDM composition (Nawahwi et al., 2013)	62
Table 3.2	Some details on the source of bacteria strains used in this study	65
Table 3.3	Bacteria combination in working solution	67
Table 3.4	The composition of bacteria culture in modified CDM medium for screening mixed culture.	67

Table 3.5	The composition of bacteria culture in modified CDM medium for screening pure culture.	68
Table 3.6	Universal primers used for amplification of 16S rRNA gene (Elmi, 2014; Bay, 2014)	71
Table 3.7	The composition of modified CDM for biodegradation analysis.	75
Table 3.8	The composition of modified CDM for MFC.	81
Table 4.1	AR-27 dye removal and dry cell weight of each bacteria consortium after 2 h in static flasks experiment. All bacterial combinations were grown in CDM supplemented with 0.1 g/L AR-27 dye and nutrient broth. Data are presented as the average of values obtained from three biological replicate, respectively.	92
Table 4.2	Dye removal and rate of dye removal of single culture bacterial strains	93
Table 4.3	Summary of biochemical characterization of bacteria B1.	96
Table 4.4	Accession number and strain name generated by GenBank	99
Table 5.1	Summary results of dye removal rate and bacterial growth after 2 h treatment in CDM contain AR-27 dye with different type of nitrogen sources.	106
Table 5.2	Summary results of dye removal rate and bacterial growth rate after 2 h treatment in CDM contain AR-27 dye with different concentration of yeast extract.	108
Table 5. 3	Summary results of dye removal rate and bacterial growth rate after 2 h treatment in CDM contain AR-27 dye with different type of carbon sources.	112
	anterent type of euroon sources.	114

Table 5.4	Summary results of dye removal rate and bacterial growth	
	rate after 2 h treatment in CDM contain AR-27 dye with	
	different concentration of glucose	113
Table 6.1	Summarized results of the MFC performance at different external loads	148
	external loads	140
Table 6.2	Various parameters showing the MFC performance at	
	different diameter of salt bridge	152
Table 6.3	(a)Diagram of close circuit voltage (CCV) (b) Current	
	density (c) Power density generation using different	
	diameter of salt bridge during 72 h operating time.	154
Table 6. 4	MFC performance at different external loads.	160
Table 6.5	Analysis of optimized MFC performance.	161
Table 6.6	Elemental analysis for control graphite felt and graphite felt	
	after 72 h MFC operation.	167

# LIST OF FIGURES

FIGURE NO	. TITLE	PAGE
Figure 2 1	Amaranth dye chemical structure (Coronila et al., 2014).	15
Figure 2.2	Schematic representation of different mechanisms of anaerobic azo dye reduction. RM=Redox mediator; ED=Electron donor; b=bacteria (enzymes) (Alabdraba &	
	Ali, 2014)	27
Figure 2.3	General overview of azo dyes and aromatic amines during anaerobic and aerobic treatment (Alabdraba & Albayati,	
	2014).	30
Figure 2.4	Quantitative analysis of the scientific literature on MFC (Source: ISI Web of Science, January 2017; Santoro <i>et al.</i> ,	
	2017)	36
Figure 2.5	MFC schematic (Holmberg et al., 2014)	37
Figure 2.6	Summarized view of two-chamber MFC with possible modes of electron transfer (1) Direct electron transfer via outer membrane cytochromes; (2) electron transfer through mediators; and (3) electron transfer through nanowires (Pant <i>et al.</i> , 2010).	40
Figure 2.7	Bacteria biofilm connected to electrode in MFC via pili (Franks & Nevin, 2010)	41
Figure 2.8	Schematic diagram of salt bridge in MFC (Khan <i>et al.</i> , 2012)	47
Figure 2.9	The proposed Grotthuss mechanism with the consecutive protons transfer indicated with the number arrows (Agmon,	
	1995).	48

Figure 3. 1	Flow chart of research methodology	60
Figure 3.2	Double-chambered MFC (a) setup in laboratory (b) Schematic view	80
Figure 3.3	Schematic view of MFC operations in (a) open circuit voltage and (b) close circuit.	84
Figure 4.1	Decolorization of CDM in facultative anaerobic condition (a) CDM media before decolorization (b) CDM media after 2h decolorization	91
Figure 4.2	AR-27 dye removal rate by pure bacteria NF2015 and its growth rate in static flask experiment. The bacteria grown in CDM supplemented with 0.2 g/L AR-27 dye and 10% NB. Data are presented as the average of values obtained from three biological replicate, respectively.	93
Figure 4.3	<ul><li>Gel electrophoresis image of bacterial NF2015 genomic</li><li>DNA extraction in 1% agarose gel was runned in triplicates.</li><li>(Lane 1: Ladder, Lane 2: extraction sample 1, Lane 3:</li><li>Extraction sample 2, and Lane 4: Extraction sample 3)</li></ul>	96
Figure 4.4	Gel electrophoresis image of PCR amplification product of bacteria NF2015 in 1% agarose gel was runned in triplicates (Lane 1: DNA Ladder, Lane 2: PCR sample 1, Lane 3: PCR sample 2, Lane 4: PCR sample 3)	97
Figure 4.5	NCBI Blast result that show almost 100% sequence of bacteria NF2015 matched to the database of <i>E.cloacae</i> .	98
Figure 4.6	Phylogenetic tree showing the relationships between isolated strain and closely related species based on 16S rRNA gene sequences. The sequence of Klebsiella pneumonia NJ02 KR476606 served as an outgroup sequence. The tree was constructed using the neighbour joining method. The numbers at nodes indicate the	

	percentages of occurrence of the branching order in 1000 bootstrapped trees. Scale bar = $0.2\%$ divergence.	99
Figure 5.1	(a)Percentage AR-27 dye removal vs time (b) Bacterial growth vs time in different type of nitrogen sources. Data are presented as the average of values obtained from three biological replicate, respectively.	109
Figure 5.2	(a)Percentage AR-27 dye decolorization vs time (b) Bacterial growth vs time) in different concentration of yeast extract as selected nitrogen source. Data are presented as the average of values obtained from three biological replicate, respectively	110
Figure 5.3	(a)Percentage of azo dye removal vs time (b) Bacterial growth vs time in different type of carbon sources based on the average. Data are presented as the average of values obtained from three biological replicate, respectively.	115
Figure 5.4	(a) Percentage AR-27 decolorization vs time (b) Bacterial growth vs time in different concentration of glucose as selected carbon sources. Data are presented as the average of values obtained from three biological replicate, respectively.	116
Figure 5.5	Comparison of (a) Bacterial growth, and (b) Percentage of AR-27 dye removal between CDM without co-substrate, unoptimized CDM and CDM with optimized co-substrates. Data are presented as the average of values obtained from three biological replicate, respectively	118
Figure 5.6	CDM (a) before and (b) after treatment with bacteria under facultative anaerobic (FA), and sequential facultative anaerobic-aerobic (FA-A) condition.	119
Figure 5.7(a)	Bacterial growth in CDM and (b) percentage of dye removal under aerobic (A), facultative anaerobic (FA), and	

	sequential facultative anaerobic-aerobic (FA-A) condition.	
	Data are presented as the average of values obtained from	
	three biological replicates, respectively.	120
Figure 5.8	COD reading vs time of CDM in facultative anaerobic and	
	sequential facultative anaerobic-aerobic condition. Data are	
	presented as the average of values obtained from three	
	biological replicate, respectively.	122
Figure 5.9	UV-Vis spectra of control CDM and CDM after	
	decolorization (2 h)	123
Figure 5.10	IR spectrum of (a) control CDM, (b) 2 h treatment in FA,	
	(c) sketched IR spectrum of both samples.	125
Figure 5.11	IR spectrum of (a) 72 h treatment in FA, and (b) 72 h	
	treatment in FA-A, and (c) sketched IR spectrum of both	
	conditions.	127
Figure 5.12	Cyclic voltammograms at 0.01 V/s in 2 h facultative	
	anaerobic condition.	128
Figure 5.13	Cyclic voltammograms at 0.01 V/s at different time	
	intervals during 72 h facultative anaerobic condition.	129
Figure 5.14	Cyclic voltammograms at 0.01 V/s at different time	
	intervals during 72 h sequential facultative anaerobic-	
	aerobic condition.	130
Figure 5.15	Reducing sugar concentration vs time in (a) facultative	
	anaerobic and (b) sequential facultative anaerobic-aerobic	
	condition.	132
Figure 5.16	Total polyphenol analysis and dry cell weight for AR-27	
	decolorization in (a) Facultative anaerobic condition and	
	(b) Sequential facultative anaerobic condition.	134
Figure 5.17	HPLC analysis of AR-27 treatment based on catechol	
	degradation (a) Standard catechol (400 mg/L), (b) 2 hours	

	of facultative anaerobic treatment, (c) 72 hours facultative anaerobic treatment, (d) 72 hours sequential facultative	
	anaerobic-aerobic treatment	137
Figure 518	Proposed AR-27 and catechol degradation pathway by <i>E. cloacae</i> NF2015 bacteria. (Chan <i>et al.</i> , 2012; Sabaruddin, 2018)	139
Figure 6.1	Open circuit voltage (OCV) reading of MFC during 48 hours operation time	145
Figure 6.2	Close circuit voltage (CCV) reading of MFC using 1 k $\Omega$ , 5 k $\Omega$ , and 10 k $\Omega$ during 72 h (b) MFC polarization curve	149
Figure 6.3	(a)Diagram of Close Circuit Voltage (CCV) (b) Current Density (c) Power Density generation using different electrode surface area during 72 h operating time.	158
Figure 6.4	Polarization curve of optimized MFC at 24 h for resistant 10 k $\Omega$ to 100 k $\Omega$ .	159
Figure 6.5	Cyclic voltammograms during 2 h treatment in MFC	163
Figure 6.6	Cyclic voltammograms at 24 h, 48 h, and 72 h treatment in MFC	164
Figure 6. 7	SEM images of (a) control graphite felt before MFC operation. (b) biofilm formation on the MFC graphite felt electrode after 72 h MFC operation (5000x magnification).	165
Figure 6.8	Energy Dispersive X-ray (EDX) analysis for (a) control graphite felt and (b) graphite felt electrode after MFC operation (100x magnification).	166
Figure 6.9	Infrared spectra for (a) control graphite felt and (b) graphite felt electrode after MFC operation	168

Figure 6.10 AR-27 treatment by *E. cloacae* NF2015 in facultative anaerobic flask treatment (FA), sequential facultative anaerobic-aerobic flask treatment (FA-A), and in facultative anaerobic MFC (MFC) in term of COD reading (mg/mL).

169

### LIST OF ABBREVIATIONS

Abs	-	Absorbances
Abs <sub>600nm</sub>	-	Absorbance at the wavelength of 600 nm
$A_{\mathrm{f}}$	-	Final absorbance
Ai	-	Initial absorbance
AR-27	-	Acid Red 27
Asa	-	Anode total surface area
BOD	-	Biological oxygen demand
CCV	-	Close circuit voltage
CDM	-	Chemically defined media
COD	-	Chemical oxygen demand
CV	-	Cyclic voltammetry
$C_i$	-	Initial COD
$C_{\mathrm{f}}$	-	Final COD
DNA	-	Deoxyribonucleic acid
DNS	-	Dinitrosalicyclic
EDX	-	Energy dispersive X-ray
EDTA	-	Ethylenediaminetetraacetic acid
FA	-	Facultative anaerobic
FA-A	-	Sequential facultative anaerobic-aerobic
FADH	-	Flavin adenine dinucleotide
FMNH <sub>2</sub>	-	1,5-dihydro flavin mononucleotide
FTIR	-	Fourier transform infrared spectroscopy
gDNA	-	Genomic DNA
$H_2SO_4$	-	Sulphuric acid
HgSO <sub>4</sub>	-	Mercury sulphate
HPLC	-	High performance liquid chromatography
Ι	-	Current
ID	-	Current density
$K_2Cr_2O_7$	-	Potassium dichromate
$K_2HPO_4$	-	Dipotassium hydrogen phosphate

KCl	-	Potassium chloride
$KH_2PO_4$	-	Potassium dihydrogen phosphate
MFC	-	Microbial fuel cell
Na <sub>2</sub> CO <sub>3</sub>	-	Sodium carbonate
NaCl	-	Sodium chloride
NADH	-	Nicotinamide adenine dinucleotide
NADPH	-	Nicotinamide adenine dinucleotide phosphate
NB	-	Nutrient broth
(NH4) <sub>2</sub> SO <sub>4</sub>	-	Ammonium sulphate
OCV	-	Open circuit voltage
OD	-	Optical density
Р	-	Power
PD	-	Power density
PEM	-	Proton exchange membrane
R	-	Resistance
RNA	-	Ribonucleic acid
rRNA	-	Ribosomal ribonucleic acid
SEM	-	Scanning electron microscope
sp	-	species
TPPC	-	Total polyphenolic content
TOC	-	Total organic carbon
UV-Vis	-	Ultra violet visible

### LIST OF SYMBOLS

%	-	Percent
μΑ	-	Microampere
μL	-	Microlitre
μm	-	Micrometre
μΜ	-	Micromolar
А	-	Ampere
cm	-	Centimetre
cm <sup>2</sup>	-	Centimetre square
e	-	Electrons
g	-	Gram
g-unit	-	G-Force
g/L	-	Gram per litre
h	-	Hours
$\mathrm{H}^+$	-	Hydrogen ion
kΩ	-	Kilo ohm
kg	-	kilogram
kPa	-	Kilopascal
L	-	Litre
М	-	Molar
mA	-	Milliampere
mA/m <sup>2</sup>	-	Milliampere per metre square
mg/mL	-	Milligram per millilitre
mL	-	Millilitre
mL/min	-	Millilitre per minute
mM	-	Millimolar
mV	-	Millivolt
mW	-	Miliwatt
$mW/m^2$	-	Miliwatts per metre square
min	-	minute
nm	-	Nanometre

°C	-	Degree Celsius
pmol	-	Picomoles
rpm	-	Revolution per minute
sec	-	second
V	-	Volt
v/v	-	Volume per volume
w/v	-	Weight per volume
λ	-	Wavelength
λmax	-	Maximum wavelength
Ω	-	Ohm

### LIST OF APPENDICES

APPENDIX	TITLE	PAGE
Appendix 1	Standard curve dry cell weight	211
Appendix 2	Growth profile of bacteria used in the screening study	211
Appendix 3	Biochemical Characterisation and Activities of Pure Culture Bacteria	212
Appendix 4	Standard curve of glucose concentration	214
Appendix 5	Standard curve total polyphenolic content	214
Appendix 6	Standard curve of catechol concentration	215
Appendix 7	Total Surface Area of Graphite Felt Calculation	215
Appendix 8	MFC Set up and Operation	216

#### **CHAPTER 1**

### **INTRODUCTION**

#### 1.1 Research Background

The management of industrial waste continues to be a major challenge in urban areas throughout the world, particularly in the rapidly growing cities and towns of the developing world. The lack of an effective and efficient industrial waste management system has had negative impact on the environment. According to Department of Environmental (DOE)'s Environmental Quality Report 2013, industrial effluent coming from manufacturing industries is one of the major sources of water pollution in Malaysia (DOE, 2013). The rise in the number of industries in Malaysia, including textile dyeing operations, has seriously increased the pollution that the country is experiencing. Textile industry is the major source of wastewater and accounts for 22 % of the total volume of industrial wastewater produced in Malaysia (Idris *et al.*, 2007).

Water pollution issue coming from industrial discharging effluents is life threatening. Among the various stages of textile production, the operations in the dyeing plant, which include pre-treatments, the dyeing and finishing, produce the most pollution. Previous report showed that the textile industry which contains various not eco-friendly dyes and organic compounds that are not eco-friendly is a major source of wastewater (Idris *et al.*, 2007). It also contains higher chemicals, high concentration of heavy metals, Biological Oxygen Demand (BOD), and also dissolved solid (Sharma *et al.*, 2007). The textile dyeing wastes are often of strong color and may also be of high temperature. When disposed into water bodies or onto land, these effluents result in the deterioration of ecology and aquatic life.

The biggest dye consumer is textile industry (Asad *et al.* 2007). The dye used in the printing and dyeing of textile product and discharging about approximately thirty

thousand tons of the dye material annually (Anjaneya *et al.*, 2011). 80% of annual commercial dyes production comes from azo dye in most countries due to their easiness of manufacturing methodology. There are over ten thousand dyes available commercially with a production of exceeding  $7 \times 10^5$  tons per year (Fu & Viraraghavan, 2001). Industries such as paper, textile, and food had major usage of azo dye (Bazin *et al.*, 2012; Anjaneya *et al.*, 2011). Their presence in wastewater affects transparency and photosynthesis by limiting the oxygen and light passing through (Sun *et al.*, 2009). Besides, the dye itself and the degradation products could be fatal and harmful to aquatic organisms which eventually affect human life via the food chain. Most of the azo dyes are either nontoxic or inert, but become toxic, mutagenic and carcinogenic upon their biotransformation (Kodam *et al.*, 2006).

Azo dyes are aromatic compounds that contain one or more azo bonds -N=Nor chromophore which make up the color (Anjaneya *et al.* 2011). Azo dyes are electron-deficient xenobiotic compounds as they contain the electron-withdrawing groups azo (N=N) and sulfonic (-SO<sub>3</sub><sup>-</sup>), causing the molecule to be an electron deficiency and subsequently less likely to undergo oxidative catabolism by bacteria (Sahasrabudhe & Pathade, 2012). Hence, azo dyes tend to persist under aerobic environmental conditions. Biological approach was chosen for wastewater treatment instead of other approaches such as physicochemical methods which involve the process of coagulation–flocculation, chemical precipitation, membrane filtration, and ion exchange (Kurniawan *et al.*, 2006) due to the costly operation and complicated process needed. In addition, biological approach is more environment-protective and economically feasible in wastewater treatment process.

In accordance with the Act and Environmental Rules (1997) it is mandatory for textile dyeing factories to install effluent treatment plants (ETPs) to treat wastewater before it leaves the factory premises. International pressure for effluent treatment is also increasing and many international buyers are now showing more concern over whether or not textiles are produced in an environmentally friendly way. Considering both factors of environmental pollution and serious health-risk caused by azo dyes, searching alternative ways to treat synthetic dyes are essential particularly for small scale textile industries. This is due to the fact that most of the textile dyeing and processing industries are located in developing countries whereby rivers are the main source of daily activities and drinking water. Most of the textile industries in Malaysia use conventional treatment methods to treat the textile effluents. the methods are effective in reduce the COD reading and removing the fiber of the wastewater however, they are not effective to decolorize the wastewater due to the presence of recalcitrant sulpho and azo groups in the zo dyes (Kulla *et al.*, 1983). This resulted in the remaining color in the treated textile wastewater.

Recently, fuel cells technology has received an exceptional attention as a new process for alternative wastewater treatment and renewable energy generation due to the high energy density, up scaling applicability and simple modular use (Evan *et al.*, 2013). In view of the overwhelming energy crisis, dependency upon fossil fuels has become a major concern. Among other sustainable and renewable resources, this bioenergy approach is considered as the most efficient way. A device that converts chemical energy from organic and inorganic matter into electrical energy through catalytic activities of microorganisms is called a microbial fuel cell (MFC). Electrons produced by the bacteria from substrates degradation are transferred from the anode to the cathode that is connected by a conductive material containing resistor. The protons in the anode migrate to the cathode and then combine with the electron and reduced by a catholyte such as oxygen to form water at the cathode surface.

The merging of the generation of power from wastewaters with the oxidation of organic or inorganic compounds is one of the vital areas of MFC research. Since the MFCs research in the pilot scale has reached a novelty success, it has boosted a research improvement globally in recent years. Even though MFC can directly generate electricity from the breakdown of organic matter, but optimization of MFC are required to increase and achieve sustainable power output.

### **1.2 Problem Statement**

One of the fastest growing industry that is significantly contributed to Malaysia's economic growth is the textile industry Previous report stated that Malaysia is the seventh largest producer and exporter of textile fibre in the Asian region in 2011 (Esho, 2015). Although the textile industry contributes positively toward the Malaysian economic growth, it was found that the industry poses a significant threat to the environmental quality, especially in terms of liquid effluent pollution and high energy consumption operational system. For the past ten years, it shows that total amount of scheduled waste generated by textile industry in Malaysia increased sharply from 744 tons in 2007 to 1559 tons in 2009 with the total water consumption in a textile industry reach as high as 3000 m3/day (Pang & Abdullah, 2012).

The Malaysia government has focused on improving its conservation policies in response to the increasing cost of energy. The government is also continually revising its energy policy to ensure sustainability of the energy resources (Mohamed & Lee, 2006) as it was estimated that the main energy consumption would increase by 2030 (Gan & Li, 2008). Besides, the depletion of fossil fuels and global warming issues together with unstable petroleum prices in the global market have encouraged the Malaysian government to start searching on renewable energy as an alternative source in line with the National Energy Policy (1979). The idea of treating the textile wastewater with simultaneous electricity generations in Microbial fuel cell ensure sustainability of the energy resources due to increasing cost of energy (Mohamed and Lee, 2006) and at the same time solving the water pollution problem in respond National Renewable Energy Policy and Action Plan (2009). Wastewaters show a convincing outcome as an energy source to be utilised as the MFC anolyte due to diverse types of organic substrate (Rahimnejad, 2015).

Hence, this pilot study focused on to enhance in wastewater quality analysis and to increase in electricity generation by using synthetic textile wastewater model. Several studies have recommended optimization which includes the electrogenic azo degrading bacteria and the MFC system itself. The optimization of textile wastewater treatment is crucial prior to MFC due to the fact that the system has to treat the wastewater efficiently while simultaneously produce electrical energy. The slow rate of substrate degradation is the main obstacle in biodegradation, so it is important for the newly isolated microbes to be acclimatized and the condition need to be optimized for the decolorization in flask. It was found that there is far greater diversity of electrogens which are electrochemically active biofilms in MFCs. Novel electrogenic bacteria remain undiscovered and unidentified. Meanwhile, many strains which have been identified by 16S rRNA studies remain to be cultured in the laboratory and their contribution to electricity generation have not yet to be confirmed (Zhang *et al.*, 2011). Besides, the maximum power production is limited by internal resistances, bacterial metabolic losses, electrochemical losses at the electrodes, and ohmic losses in the solution (Sleutels *et al.*, 2012).

Previously, and unpublished research by Bushra (Phd) has isolated and developed unidentified bacteria strains from sour crude oil. These bacteria strains have the ability to desulfurize the hydrocarbon. This finding had become a stepping stone for further implementation of the bacterial strains in wastewater treatment using synthetic textile wastewater and azo dye AR-27 (contain sulphur) used as a model.

The performance of decolorization of azo dye and bioelectricity generation via microbial fuel cell (MFC) by using *E. cloacae* bacteria from crude oil has yet been studied. Hence, this study focused on the azo dye treatment using a novel azo decolorizing bacterial strain *E. cloacae* NF2015 while simultaneously performing a series of optimization for the salt bridge MFC system in order to increase the bioelectricity generations in the form of stacked microbial fuel cell (MFC). In this study, MFC that use salt bridge for the proton exchange and the graphite felt electrode were optimized.

#### **1.3** Research Objectives

The main objective of this study is to screen the newly isolated bacteria from crude oil for AR-27 dye decolorization and optimize its condition to improve biodecolorization, biodegradation, and bioelectricity performance. The objectives of this study were as follows.

• To select and identify bacteria isolated from Iraqi crude oil for their ability to decolorize Azo dye (AR-27) wastewater.

- To revise AR-27 dye decolorization by the selected bacteria strain through optimization of co-substrates and treatment condition.
- To improve bioelectricity generation by the selected bacteria using different salt bridge diameter and electrode size in double-chambered microbial fuel cell.

### 1.4 Research Scope

The purpose of the current investigation is to identify that crude oil hydrocarbon degrading bacteria, and to elucidate their potential to degrade another form of hydrocarbon which is the azo dye AR-27. Four bacteria species POS, POS M2, B1, and F2B which were previously isolated from Iraqi crude oil reservoir were screened for the ability to decolorize Chemically Defined Media (CDM) containing azo dye Acid Red (AR) 27. The screening was performed using mixed bacteria culture under facultative anaerobic condition. The analysis of the azo dye degradation present in the synthetic wastewater sample was based on the absorbance reading at wavelength 521nm.

Out of these bacteria, the most potential azo dye decolorizing bacteria were characterized and identified using biochemical and molecular techniques. The effects of co-substrates (carbon and nitrogen source) were optimized by selecting the suitable carbon and nitrogen sources and adjusting the concentration thus improving decolorization efficiency simultaneously. The efficiency of selected bacteria to biodegrade azo dye medium was determined by analyzing the media before and after treatment using Ultraviolet-visible spectroscopy (UV-Vis), Fourier-transform infrared spectroscopy (FTIR), and High-performance liquid chromatography (HPLC) as well as the analysis of Chemically oxygen demand (COD) reduction, Total polyphenolic (TPP) content, total nitrogen, and reducing sugar. The electrochemical analysis was done using cyclic voltammetry (CV) analysis.

Finally, the biodegradation of azo dye by bacteria was applied in MFC to produce electricity. In order to test for the robustness of the MFC when treating with azo dye wastewater, the important limiting factors of MFC operation were suggested to improve MFC performance. The factors highlighted that increase power output in MFCs are the diameter of salt bridge (proton exchange system) and size of electrode at anode and cathode. The performance of MFCs was analyzed by calculating the power densities, current densities, voltage output with external resistance, and polarization curve. Besides, the surface morphology of electrodes had been examined using Scanning Electron Microscope (SEM).

#### **1.5** Significance of Research

This study focused on establish a bacterial culture capable in azo dye AR-27 removal and further applied in double chamber salt bridge-microbial fuel cell to generate electricity. The main idea for this study was to characterize and identify unknown strain on their ability to remove azo dye AR-27. The selected bacterial strain was further optimized on the effect of co-substrates to the efficiency of dye removal. The understanding on these potential isolates to degrade azo dye will increase the possibilities of developing strategies and models for azo dye pollutants removal from the environment focusing on the textile wastewater. Hence, this study could provide new idea for the current treatment of textile effluent and as an alternative green energy in the future

#### REFERENCES

- Adjemian, K., Lee, S., Srinivasan, S., Benziger, J. & Bocarsly, A. (2002). Silicon oxide nafion composite membranes for proton-exchange membrane fuel cell operation at 80-140 <sup>o</sup>C. *Journal of the Electrochemical Society*, 149, 256-261.
- Aelterman, P., Rabaey, K., Pham, T.H., Boon, N.W., & Verstraete, W. (2006). Continuous electricity generation at high voltages and currents using stacked microbial fuel cells, *Environmental Science and Technology*, 40 (10), 3388-3394.

Agmon, N. (1995). The Grotthuss mechanism. Chemical Physics Letters, 244, 456-462.

- Ahmad, R. & Kumar, R. (2011). Adsorption of amaranth dye onto alumina reinforced polystyrene. *Clean-Soil, Air, Water*, 39, 74-82.
- Akar, T., Safa-Ozcan, A., Tunali, S., & Ozcan, A. (2008). Biosorption of a textile dye (Acid Blue 40) by cone biomass of Thuja orientalis: Estimation of equilibrium, thermodynamic and kinetic parameters. *Bioresource Technology*, 99, 3057-3065.
- Alabdraba, W.M.S. & Albayati, M.B. (2014). Biodegradation of azo dyes-A review. International Journal of Environmental Engineering and Natural Resources, 1(4), 179-189.
- Al-Amrani, W.A., Lim, P.E., Seng, C.E., & Ngah, W.S.W. (2014). Factors affecting bio-decolorization of azo dyes and COD removal in anoxic–aerobic REACT operated sequencing batch reactor. *Journal of the Taiwan Institute* of Chemical Engineers, 45, 609-616.
- Albayati, M.B. (2010). Anaerobic/aerobic biological treatment of synthetic wastewater containing two types of Azo dye. Master Thesis: College of Engineering, Tikrit University, Iraq.

- Aldoury, M.M., Alabdraba, Waleed, M.S., & Albayati, M.B. (2014). Performance of sequential anaerobic/aerobic biological treatment of synthetic wastewater containing two types of Azo dye. 2nd International Conference of Environmental Science and Technology ICOEST2014-Side, Turkey.
- Allen, R.M. & Bennetto, H.P. (1993). Microbial fuel-cells: electricity production from carbohydrates. *Applied Biochemical Biotechnology*, 39(40), 27–40.
- Ali, A.H., Al-Mussawy, H.A., Hussein, M.J., & Hamadi, N.J. (2018). Experimental and theoretical study on the ability of microbial fuel cell for electricity generation. *Pollution*, 4(2), 359-368.
- Alibaba.com. Graphite felt for heat preservation. http://www.alibaba.com/ productgs/497968373/carbon\_graphite\_felt\_for\_heat\_preservation.html (2012).
- Anjaneya, O., Souche, S.Y., Santoshkumar, M., & Karegoudar, T. (2011). Decolorization of sulfonated azo dye Metanil Yellow by newly isolated bacterial strains: *bacillus sp.* strain AK1 and *Lysinibacillus sp.* Strain AK2. *Journal of Hazardous Materials*, 190(1), 351-358.
- Arora, R. (2012). Microbial Biotechnology: Energy and Environment. BioElectrochemistry, CABI, ISBN 1845939573, 9781845939571, 61.
- Artsanti P., Sudarlin, & Kirana E.F. (2017). The Effect of Increasing Surface Area of Graphite Electrode on the Performance of Dual Chamber Microbial Fuel Cells. *Proceeding International Conference on Science and Engineering*, 1, 137-140.
- Asad, S., Amoozegar, M., Pourbabae, A.A., Sarbolouki, M., & Dastgheib, S. (2007). Decolorization of textile azo dyes by newly isolated halophilic and halotolerant bacteria. *Bioresource Technology*, 98(1), 2082-2088.
- Aziz, A.S.B., Nasir, N.S.B., Ibrahim, N.B., & Yahya A.B. (2016). Decolorization of azo dy AR27 and bioelectricity generation in microbial fuel cell. *Science* and Engineering Technology, 5(1), 32-41.

- Bandary, B., Hussain, Z., & Kumar, R. (2016). Effect of carbon and nitrogen sources on Esterichia coli bacteria in removing dyes. *Materialstoday:Proceedings*, 3(10), 4023-4028.
- Barragan, B.E., Costa, C., & Marquez, M.C. (2007). Biodegradation of azo dyes by bacteria inoculated on solid media. *Dyes and Pigments*, 75, 75.
- Bazin, I. Ibn Hadi Hassine, A., Haj Hamouda, Y., Mnif, W., Bartegi, A., Lopez-Ferber, M., De Waard, M., & Gonzalez, C. (2012). Estrogenic and anti-estrogenic activity of 23 commercial textile dyes. *Ecotoxicology and Environmental Safety*, 85, 131-136.
- Belal, A.R., Cooper, S.M., & Khan, N.A. (2015). Corporate environmental responsibility and accountability: What chance in vulnerable Bangladesh? *Crit Perspect Account*, 33, 44–58.
- Bheemaraddi, M.C., Patil, S., Shivannavar, C.T., Gaddad, S.M. (2014). Isolation and characterization of Paracoccus sp. GSM2 capable of degrading textile azo dye reactive violet 5. *The Scientific World Journal*, 1-9.
- Biffinger, J.C., Pietron, J., Ray, R., Little, B., & Ringeisen, B.R. (2007). A bio film enhanced miniature microbial fuel cell using Shewanella oneidensis DSP10 and oxygen reduction cathodes. *Biosensor and Bioelectronics*, 22, 1672–1679.
- Bin, Y., Jiti, Z., Jing, W., Cuihong, D., Hongman, H., Zhiyong, S., & Yongming, B. (2004). Expression and characteristics of the gene encoding azo reductase from Rhodobacter sphaeroides AS1.1737. *FEMS Microbiology Letter*, 236, 129–136.
- Blumel, S., Contzen, M., Lutz, M., Stolz, M., & Knackmuss, H.J. (1998). Isolation of a bacterial strain with the ability to utilize the sulfonated Azo compound 4-carboxy-40-sulfoAzobenzene as sole source of carbon and energy. *Applied and Environmental Microbiology*, 64, 2315-2317.
- Bond, D. R., Holmes, D.R., Tender, L.M., & Lovley, D.R. (2002). Electrodereducing microorganisms that harvest energy from marine sediment. *Science*, 295, 483-485.

- Bond, D. R. & Lovley, D.R. (2003). Electricity production by Geobacter sulfurreducens attached to electrodes. Applied and Environmental Microbiology, 69, 1548-1555.
- Bond, D. R. & Lovley, D.R. (2005). Evidence for involvement of an electron shuttle in electricity generation by *Geothrix fermentans*. Applied and Enviromental Microbiology, 71, 2186-2189.
- Boye, K. & Hansen, D. (2003). Sequencing of 16S rDNA of *Klebsiella*: taxonomic relations within the genus and to other *Enterobacteriaceae*. *International Journal of Medical Microbiology*, 292, 495-503.
- Brown, D. & Hamburger, B. (1987). The degradation of dye stuffs. Part III. Investigation of their ultimate degradability. *Chemosphere*, 12, 397/404.
- Brown, D. & Laboureur, P. (1983). The degradation of dyestuffs: Part I. Primary biodegradation under anaerobic conditions. *Chemosphere*, 12, 397-404.
- Bullen, R. A., Arnot, T.C., Lakeman, J. B., & Walsh, F. C. (2006). Biofuel cells and their development. *Biosensor and Bioelectronics*, 21, 2015-2045.
- Busalmen, J.P., Esteve-Nunez, A., & Feliu, J.M. (2008). Whole cell electrochemistry of electricity-producing microorganisms evidences an adaptation for optimal exocellular electron transport. *Environmental Science and Technology*, 42, 2445-2450.
- Campo, A.G.D., Lobato, P.C.J. Rodrigo, M., & Morales, F.J.F. (2014). Effects of External Resistance on Microbial Fuel Cell's Performance. Environment, Energy and Climate Change II: *Energies from New Resources and the Climate Change*, 1-23
- Cappuccino, J.G. & Sherman, N. (2005). Microbiology: A Laboratory Manual. Benjamin Cummings New York, Science.
- Calignano, F., Tommasi, T., Manfredi, D., & Chiolerio, A. (2015). Additive Manufacturing of a Microbial Fuel Cell-A detailed study. *Scientific Reports*, 5, 17373.

- Carliell, C.M., Godefroy, S.J., Naidoo, N., Buckley, C.A., Senior, E., Mulholland, D.,
   & Martineigh, B.S. (1994). Anaerobic decolourisation of Azo dyes, in: Seventh International Symposium on Anaerobic Digestion, 303-306.
- Chacko, J.T. & Subramaniam, K. (2011). Enzymatic degradation of azo dyes: A review. International Journal of Environmental Sciences, 1(6), 1250-1260.
- Chae, K.J., Choi, M., Folusho, F., Ajayi, & Park, W., Chang, I.S., & Kim, I.S. (2008). Mass transport through a proton exchange membrane (nafion) in Microbial Fuel Cells. *Energy & Fuels*, 22, 169–176.
- Chae, K.J., Choi, M.J., Kim, K.Y., Ajayi, F.F., Park, W., Kim, C.W., & Kim, I.S. (2010). Methanogenesis control by employing various environmental stress conditions in two-chambered microbial fuel cells. *Bioresource Technology*, 101, 5350-5357.
- Chan, G.F., Rashid, N.A.A., Chua, L.S., Nasiri, R. & Ikubar, M.R.M. (2012). Communal microaerophilic-aerobic biodegradation of Amaranth by novel NAR-II bacterial consortium. *Bioresource Technology*, 105, 48-59.
- Chang, J.S. & Kuo, T.S. (2000). Kinetics of bacterial decolorization of azo dyes with *Escherichia coli* NO<sub>3</sub>. *Bioresource Technology*, 75, 107.
- Chang, J.S., Lin, Y.S. (2000). Fed-batch bioreactor strategies for microbial decolorization of azo dye using *Pseudomonas luteola* strain, *Biotechnology Progress*, 16, 979-985.
- Chang, J.S., Chou, C., Lin, Y., Ho, J., & Hu, T.L. (2001). Kinetic Characteristics of bacterial Azo-dye decolorization by *Pseudomonas luteola*. Water *Research*, 35, 2041-2850.
- Chang, O.S., Chen, B.Y., & Lin, Y.S. (2004). Stimulation of bacterial decolorization of an azo dye by extracellular metabolites from Escherichia coli strain NO3. *Bioresource Technology*, 91, 243-248.
- Chapman, K. L. (1992). Anaerobic/aerobic degradation of a textile dye wastewater. Virginia Polytechnic Institute and State University: Master Thesis

- Chaudhuri, S.K. & Lovley, D.R. (2003). Electricity generation by direct oxidation of glucose in mediator-less microbial fuel cells. *Nature Biotechnology*,21, 1229-1232.
- Chen, K.C., Huang, T., Wu, J.Y., & Houng, J.Y. (1999). Microbial decolorization of Azo dyes by Proteus mirabilis. Journal of Industrial Microbiology and Biotechnology, 23, 686-690.
- Chen, K.C., Jane-Yii, W., Liou, D.J., & Hwang, S.C.J. (2003). Decolorisation of the textile dyes by newly isolated bacterial strains. *Journal of Biotechnology*, 101,57-68.
- Chen, W., Li, J., Sun, X., Min, J., & Hu, X. (2017). High efficiency degradation of alkanes and crude oil by a salt-tolerant bacterium Dietziaspecies CN-3. *International Biodeterioration and Biodegradation*, 118, 110-118.
- Chengalroyen, M. D. & Dabbs, E. R. (2013). The microbial degradation of azo dyes: minireview. *World Journal of Microbiology and Biotechnology*, 29(3), 389-399.
- Chung, K. T., Stevens, S. E., & Cerniglia, C. E. (1992). The reduction of azo dyes by instential microflora. *Critical Review of Microbiology*, 18, 175-190.
- Clauwaert, P., Aelterman, P., Pham, T.H., De Schamphelaire, L., Carballa, M., Rabaey, K., & Verstraete, W. (2008). Minimizing losses in bioelectrochemical systems: the road to applications. *Applied Microbiology* and Biotechnology, 79, 901-913.
- Coronila, I.M., Barrera, L.M., Garrido, T.L.V, & Urbina, E.C. (2014). Bioabsorption of amaranth dye from aqeous solutions by roots, leaves, stems, and the whole plant of E. crassipes. *Environmental Engineering and Management Journal*, 13(8), 1917-1926.
- Coughlin, M.F., Kinkle, B.K., & Bishop, P.L. (1999). Degradation of Azo dyes containing amino naphthol by Sphingomonas sp. strain ICX. *Journal of Industrial Microbiology and Biotechnology*, 23, 341-346.

- Crriddle, W.J.& Ellis, G.P. (1994). Spactraland chemical characterization of organic compounds. *Third Edition Great*, Britain.
- Crittenden, S.R., Sund, C.J., & Sumner, J.J. (2006). Mediating electron transfer from bacteria to a gold electrode via a self-assembled monolayer. *Langmuir*, 22, 9473-9476.
- Cuijie, F., Qin, D., Li, J., Chen, L., Zhao, F., Chen, S., Hu, H., & Yu, C.P. (2014). Characterization of Exoelectrogenic Bacteria Enterobacter Strains Isolated from a Microbial Fuel Cell Exposed to Copper Shock Load. *PLOS One*, 9(11), 1-10.
- Dandie, C. E.; Thomas, S. M. Bentham, R. H. & McClure, N. C. (2004). Physiological characterization of Mycobacterium sp. strain 1B isolated from a bacterial culture able to degrade high-molecular-weight polycyclic aromatic hydrocarbons. *Journal of Applied Microbiology*, 97, 246-255.
- Davin-Regli, A. & Pages, J.M. (2015). Enterobacter aerogenes and Enterobacter cloacae; versatile bacterial pathogens confronting antibiotic treatment. Frontiers in Microbiology, 6 (392), 1-10.
- Davis, V.M. & Bailey, J.E.J. (1993). Chemical reduction of FD and C yellow No. 5 to determine combined benzidine. *Journal of Chromatography*, 635, 160-164.
- Delaney, G.M., Bennetto, H.P., Mason, J.R., Roller, S.D., Stirling, J.L., & Thurston, C.F. (1984). Electron-transfer coupling in microbial fuel cells. II. Performance of fuel cells containing selected microorganism-mediator combinations. *Journal of Chemical Technology and Biotechnology*, 34, 13–27.
- Delee, W., O'Neill. C., & Hakes, F.R. (1998). Anaerobic treatment of textile effluents: a review. *Journal of Chemical Technology and Biotechnology*, 73, 323-335.
- Department of Environment (DOE), Malaysia. (2013). Malaysia Environmental Quality Report 2013. Misas Advertising Sdn. Bhd. 24-85.

Department of Statistic, Malaysia. (2011). Sustainability Report 2011. IWK. 101.

- DosSantos, A.B., Cervantes, F.J., & Van-Lier, J.B. (2007). Review paper on current technologies for decolourisation of textile wastewaters: perspectives for anaerobic biotechnology. *Bioresource Technology*, 98, 2369-2385.
- Dos Santos, A.B., Madrid, M.P., De Bok, F.A.M., Stams, A.J.M., Van Lier, J.B., & Cervantes, F.J. (2006). The contribution of fermentative bacteria and methanogenic archaea to azo dye reduction by thermophilic anaerobic consortium. *Enzyme and Microbial Technology*, 39, 38-46.
- Dudonné, S., Vitrac, X., Coutière, P., Woillez, M., & Mérillon, J. M. (2009). Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *Journal of Agricultural and Food Chemistry*, 57(5), 1768-1774.
- Eggleston, C.M., Voros, J., Shi, L., Lower, B.H., Droubay, T.C., & Colberg P.J.S. (2008). Binding and direct electrochemistry of OmcA, an outer-membrane cytochrome from an iron reducing bacterium, with oxide electrodes: a candidate biofuel cell system. *Inorganica Chimica Acta*, 361, 769-777.
- Emde, R. & Schink, B. (1990). Oxidation of glycerol, lactate, and propionate by Propionibacterium freudenreichii in a poisedpotential amperometric culture system. *Archieves of Microbiology*, 153, 506–512.
- Emde, R., Swain, A., & Schink, B. (1989). Anaerobic oxidation of glycerol by Escherichia coli in an amperometric poisedpotential culture system. *Applied Microbiology and Biotechnology*, 32, 170–175.
- Esho, H. (2015). Dynamics of the Textiles and Apparel Industries in Southeast Asia: A Preliminary Analysis. Journal of International Economic Studies, No.29, 85–106.
- Evans, D.H., James, B., Claiborne, & Currie, S. (2013). *The physiology of fishes*. (4<sup>th</sup> Ed). CRC Press. (pp. 14).

- Field, S.J., Dobbin, P.S., Cheesman, M.R., Watmough, N.J., Thomson, A.J., & Richardson, D.J. (2000). Purification and magneto-optical spectroscopic characterization of cytoplasmic membrane and outer membrane multiheme c-type cytochromes from Shewanella frigidimarina NCIMB400. *The Journal of Biological Chemistry*, 275, 8515-8522.
- Field, J.A., Stams, A.J.M., Kato, M., Schraa, G. (1995). Enhanced biodegradation of aromatic pollutants in cocultures of anaerobic and aerobic bacterial consortia. *Antoine Van Leeuwenhoek*, 67, 47-77.
- Filipkowska, Z. (2003). Sanitary and bacteriological aspects of sewage treatment. *Acta Microbiologica Polonica*, 52, 57-66.
- Forgacs, E., Cserhati, T., & Oros, G. (2004). Removal of synthetic dyes from wastewaters: a review. *Environment International*, 30, 953-997.
- Franciscon, E., Zille, A., Fantiatti-Garboggini, F., Silva, I.S., Cavaco-Paulo, A., & Durrant, L.R. (2009). Microaerophilic-aerobic sequential decolourization/biodegradation of textile azo dyes by a facultative *Klebsiella sp.* Strain VN-31. *Process Biochemistry*, 44.446-452.
- Franks, A.E & Nevin, K.P., (2010). Microbial Fuel Cells, A Current Review. *Energies*, 3, 899-919.
- Fricke, K., Harnisch, F., & Schroder, U. (2008). On the use of cyclic voltammetry for the study of anodic electron transfer in microbial fuel cells. *Energy and Environmental Science*, 1, 144-147.
- Fu, Y. & Viraraghavan, T. (2001). Fungal decolorization of dye wastewaters: A Review. *Bioresource Technology*, 79, 251-262.
- Gan, P. Y., and Li, Z. (2008). An econometric study on long-term energy outlook and the implications of renewable energy utilization in Malaysia. Energy Policy, 36(2), 890-899.
- Ganapathy, B., Ying, C. Y., & Ibrahim, N. (2016). Bioelectricity generation based on Acid Red 27 decolourisation in microbial fuel cell. *Journal of Advanced Research in Applied Sciences and Engineering Technology*, 5(1), 42-52.

- Ghangrekar, M.M. & Shinde, V.B. (2006). Wastewater Treatment in Microbial Fuel Cell and Electricity Generation: A Sustainable Approach. 12th international sustainable development research conference, 1-9.
- Ghangrekar, M.M. & Shinde, V.B. (2007). Performance of membrane-less microbial fuel cell treating wastewater and effect of electrode distance and area on electricity production. *Bioresource Technology*, 98(15), 2879-2885.
- German, B., Maribel, Q., & Gloria, M. (2004). Aerobic degradation of the azo dye acid red 151 in a sequencing bactch biofilter. *Bioresource Technology*, 92, 143-149.
- Gingell, R. & Walker, R. (1971). Mechanisms of Azo reduction by *Streptococcus faecalis* II. The role of soluble flavins. *Xenobiotica*, 1, 231-239.
- Godhake, G., Jadhav, S., Dawkar, S., & Govindwar, V. (2009). Biodegradation of diazo dye direct brown MR by Acinetobacter calcoaceticus NCIM 2890. *International Biodeterioration and Biodegradation*, 63, 433-439.
- Gorby, Y. A., Yanina, S., McLean, J. S., Rosso, K. M., Moyles, D., & Dohnalkova, A. (2006). Electrically conductive bacterial nanowires produced by Shewanella oneidensis strain MR-1 and other microorganisms. *Proceeding of National Academy of Science* USA, 103, 11358-11363,81-2285.
- Gude, V.G. (2016). Microbial Fuel Cells for Wastewater Treatment and Energy Generation: an Overview. *Journal of Cleaner Production*, 122, 287-307.
- Guo, J., Zhou, J., Wang, D., Tian, C., Wang, P. & Salah Uddin, M. (2008). A novel moderately halophilic bacterium for decolorizing azo dye under high salt condition. *Biodegradation*, 19, 15-19.
- Grzebyk, M. & Pozniak, G. (2005). Microbial fuel cells (MFCs) with interpolymer cation exchange membranes. *Separation and Purification Technology*, 41, 321–328.

- Handayani, W., Meitiniarti, V.I., & Timotius, K.H. (2007). Decolorization of Acid Red 27 and Reactive Red 2 by *Enterococcus faecalis* under a batch system. *World Journal of Microbiology and Biotechnology*, 23, 1239-1244.
- Hao, J.J., Song, F.Q., Huang, F., Yang, C.L., Zhang, Z.J., Zheng, Y., & Tian, X.J. (2007). Production of laccase by a newly isolated deuteromycete fungus Pestalotiopsis sp. And its decolorization of azo dyes. *Journal of Industrial Microbiology and Biotechnology*, 34, 233-240.
- Harnisch, F. & Schroder, U. (2009). Selectivity versus mobility: separation of anode and cathode in microbial bioelectrochemical systems. *ChemSusChem*, 2, 921-926.
- Hassanshahian, M. (2014). Isolation and characterization of biosurfactant producing bacteria from Persian Gulf (Bushehr provenance). *Marine Pollution Bulletin*, 86, 361-366.
- Haug, W., Schmidt, A., Nortemann, B., Hempel, D.C., Stolz, A., Knackmuss, H.J. (1991). Mineralization of the sulfonated Azo dye Mordant Yellow 3 by a 6-aminonaphthalene-2- sulfonate-degrading bacterial consortium. *Applied and Environmental Microbiology*, 57, 3144-3149.
- He, Z., Minteer, S.D., & Angenent, L.T. (2005). Electricity generation from artificial wastewater using an upflow microbial fuel cell. *Environmental Science* and Technology, 39, 5262-5267.
- Hinteregger, C., Leitner, R., Loidl, M., Ferschl, A., & Streichsbier, F. (1992).
   Degradation of phenol and phenolic compounds by *Pseudomonas putida* EKII. *Applied Microbiology and Biotechnology*, 37(2), 252-259
- Holmes, D. E., Bond, D.R., & Lovley, D.R. (2004). Electron transfer by *Desulfobulbus* propionicus to Fe (III) and graphite electrodes. *Applied and Environmental Microbiology*, 70, 1234-1237.
- Holmes, D. E., Nicoll, J.S., Bond, D.R., & Lovley, D.R. (2004). Potential role of a novel psychrotolerant member of the family *Geobacteraceae*, *Geopsychrobacter electrodiphilus* gen. nov., sp. nov., in electricity

production by a marine sediment fuel cell. *Applied and Environmental Microbiology*, 70, 6023-6030.

- Hong, Y., Guo, J., Zhicheng, X., Cuiyun, M., Meiying, X., & Guoping, S. (2007). Reduction and partial degradation mechanisms of naphthylaminesulfonic azo dye Amaranth by Shewanella decolorationis S12. *Applied Microbial Biotechnology*, 75, 647–654.
- Hu, T.L. (1998). Degradation of azo dye RP2B by *Pseudomonas luteola*. Water Science and Technology, 38, 299-306.
- Huggins, T., Wang, H., Kearns, J., Jenkins, P., & Ren, Z.J. (2014). Biochar as a sustainable electrode material for electricity production in microbial fuel cells. *Bioresource Technology*, 157, 114-119.
- Ibrahim, S.Z. (2015). Environmental legislation for safe guarding our water. Forum on Environmental Legislation 2015 in conjunction with ENSEARCH 31<sup>th</sup> AGM.
- Idris, A., Hashim, R., Abdul Rahman, R., Ahmad, W.A., Ibrahim, Z., Abdul Razak, P.R., Mohd Zin, H., & Bakar, I. (2007). International Journal of Engineering and Technology, 4(2), 228-234.
- Jain, K., Shah, V., Chapla, D., & Madamwar, D. (2012). Decolorization and degradation of azo dye- Reactive violet 5R by an acclimatized indigenous bacterial mixed cultures-SB4 isolated from anthropogenic dye contaminated soil. *Journal of Hazardous Materials*, (213-214), 378-386.
- Ilamathi, R. & Jayaprint, J. (2018). Microbial Fuel Cell for dye decolorization. Environmental *Chemistry Letters*, 16, 239-250.
- Ismail, A.F. & Matsuura, T. (2012). Sustainable membrane technology for energy, water, and environmental. *Technology and Engineering, John Wiley & Sons*, 127-130.
- Isik, M. & Sponza, D.T. (2007). Fate and toxicity of azo dye metabolites under batch longterm anaerobic incubations. *Enzyme Microbiology Technology*, 40(4), 934–939.

- Jabeen, H.S., Rahman, S.U., Mahmood, S., & Anwer, S. (2013). Genotoxicity assessment of amaranth and allura red using *Saccharomyces cerevisiae*. *Bulletin of Environmental Contamination and Toxicology*, 90, 22-26.
- Jadhav, S.B., Patil, N.S., Watharkar, A.D., Apine, O.A., & Jadhav, J.P., (2013). Batch and continuous biodegradation of amaranth in plain distilled water by *P*. *aeruginosa* BCH and toxicological scrutiny using oxidative stress studies. *Environmental Science and Pollution Research*, 20, 2854-2866.
- Jadhav, J.P., Parshetti, G.K., & Kalme, S.D. (2007). Decolourization of azo dye methyl red by Saccharomyces cerevisiae MTCC 463. Chemosphere, 68(2), 394– 400.
- Jang, J.K., Pham, T.H., Chang, I.S., Kang, K.H., Moon, H., Cho, K.S., & Kim, B.H. (2004). Construction and operation of a novel mediator-and membraneless microbial fuel cell. *Process Biochemistry*, 39, 1007-1012.
- Jatoi, A.S., Aziz, S., Mahar H., & Furqan M. (2016). To Investigate the Optimized Conditions of Salt Bridge for Bio- Electricity Generation from Distillery Waste Water Using Microbial Fuel Cell. NUST Journal of Engineering Sciences, 9(2), 29-34
- Kabir, E.S., Kabir, M., & Islam, S.M. (2002). Assessment of effluent quality of Dhaka export processing zone with special emphasis to the textile and dying industries. *Jahangirnagar Uni J Science*, 25, 137–8.
- Kapdan, I.K., Kargi, F., McMullan, G., & Marchant, R. (2000). Decolorization of textile dye stuffs by a mixed bacterial consortium. *Biotechnology Letters*, 22, 1179-1181.
- Kapdan, I.K., Tekol, M., & Sengul, F. (2003). Decolorization of simulated textile wastewater in an anaerobic/ aerobic sequential treatment system. *Process Biochemistry*, 38, 1031-1037.
- Kardi, S.N., Ibrahim, N., Rashid, N.A.A., Darzi, G.N. (2015). Simultaneous acid red 27 decolourisation and bioelectricity generation in a (H-type) microbial fuel cell using NAR-2. Environmental Science and Pollution Research, 23(4), 3358-3364.

- Karkmaz, M., Puzenat, E., Guillard, C., & Herrmann, J.M., (2004). Photocatalytic degradation of the alimentary azo dye amaranth: Mineralization of the azo group to nitrogen. *Applied Catalysis B: Environmental*, 51, 183-194.
- Karnnet, S., Potiyaraj, P., & Pimpan, V. (2005). Preparation and properties of biodegradable stearic acid-modified gelatin films. *Polymer Degradation* and Stability, 90(1), 106-110.
- Keane, J., & Velde, D.W. (2017). The role of textile and clothing industries in growth and development strategies. *Investment and Growth Programme Overseas Development Institute (ODI).*
- Khalid, A., Arshad, M., & Crowley, D.E. (2008). Accelerated decolorization of structurally different azo dyes by newly isolated bacterial strains. *Applied Microbiology and Biotechnology*, 78, 361-369.
- Khan, M.R, Bhattacharjee, R., & Amin, M.S.A. (2012). Performance of the Salt Bridge Based Microbial Fuel Cell, *Intenational Journal of Engineering and Technology*, 1(2),115-123
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S., & Chimni, S.S. (2005). Decolorization of various Azo dyes by bacterial consortia. *Dyes and Pigments*, 67, 55-61.
- Klimiuk, E. & Łebkowska, M. (2005). Biotechnology in protection of environmental (in Polish). Wydawnictwo Naukowe PWN S.A, *Warszawa*, 5, 114-136.
- Kim, J.R., Cheng, S., Oh, S.E., & Logan, B.E. (2007). Power generation using different cation, anion, and ultrafiltration membranes in microbial fuel cells. *Environmental Science and Technology*, 41, 1004-1009.
- Kim, J. R, Min, B., & Logan, B. E. (2005). Evaluation of procedures to acclimate a microbial fuel cell for electricity production. *Applied Microbiology and Biotechnology*, 68, 23–30.
- Kim, H.J., Hong, S.W., Kim, H.D., Yang, S.Y., & Chung, S.H. (2002). A study on quenching meshes as a possible controlling tool of hydrogen explosion in

nuclear power plants. 10th International Conference on Nuclear Engineering, American Society of Mechanical Engineers, 2, 145-151.

- Kim, B. H., Kim, H.J., Hyun, M.S., & Park, D.S. (1999a). Direct electrode reaction of Fe (III) reducing bacterium, *Shewanella putrefaciens*. *Journal of Microbiology and Biotechnology*, 9, 127-131
- Kim, H.J., Hyun, M.S., Chang, I.S., & Kim, B.H. (1999b). A microbial fuel cell type lactate biosensor using a metal reducing bacterium, *Shewanella putrefaciens*. *Journal of Microbiology and Biotechnology*, 9, 365–367.
- Kim, H. J., Park, H.S., Hyun, M.S., Chang, I.S., Kim, M., & Kim, B.H. (2002). A mediator-less microbial fuel cell using a metal reducing bacterium, *Shewanella putrefaciens. Enzyme and Microbial Technology*, 30, 145-152.
- Kimura, M. (1980). A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16(2), 111-120.
- Knapp, J.S. & Newby, P.S. (1995). The microbiological decolorization of an industrial effluent containing a diazo linked chromophore. *Water Resources*, 7, 1807-1809.
- Kodam, K., Soojhawon, I., Lokhande, P., & Gawai, K. (2006). Microbial decolorization of reactive azo dyes under aerobic conditions. World Journal of Microbiology and Biotechnology, 21, 367-370.
- Kulla, H.G., Klausener, F., Meyer, O., Ludeke, B., & Leisinger, T. (1983). Interference of aromatic sulfo groups in the microbial degradation of the azo dyes orange II and I. Archives of Microbiology, 135, 1-7.
- Kumar, N. & Das D. (2000). Enhancement of hydrogenproduction by Enterobacter cloacae IIT-BT 08. Process Biochemistry, 35(6), 589-593
- Kurniawan, T.A., Chan, G., Lo, W.H., & Babel, S. (2006). Physico-chemical treatment techniques for wastewater laden with heavy metals. *Chemical Engineering Journal*, 118, 83-98.

- Laczi, K., Kis, A., Horvath, B., Maroti, G., Hegedus, B., Perei, K., & Rakhely, G. (2015). Metabolic responses of *Rhodococcus erythropolis* PR4 grown on diesel oil and various hydrocarbons. *Applied Microbiology and Biotechnology*, 99(22), 9745-9759.
- Li, W., Raoult, D., & Fournier, P. (2009). Bacterial strain typing in the genomic era. *FEMS Microbiology Review*, 33, 892-916.
- Lithgow, A.M., Romero, L., Sanchez, I.C., Souto, F.A., & Vega, C.A. (1986). Interception of electron-transport chain in bacteria with hydrophilic redox mediators. *Journal of Chemical Research*, Synopses, 5, 178–179
- Liu, H., Cheng, S., Logan, B. E. (2005). Production of electricity from acetate or butyrate using a single-chamber microbial fuel cell. *Environmental Science Technology*, 39, 658–662.
- Liu, H. & Logan, B.E. (2004). Electricity generation using an air cathode single chamber microbial fuel cell in the presence and absence of a proton exchange membrane. *Environmental Science Technology*, 38, 4040–4046.
- Liu, H., Ramnarayanan, R., & Logan, B.E. (2004). Production of electricity during wastewater treatment using a single chamber microbial fuel cell. *Environmental Science Technology*, 38, 2281–2285.
- Liu, W.Y., Wong, C.F., Chung, K.M.K., Jiang, J.W., & Leung F.C.C. (2013). Comparative genome analysis of Enterobacter cloacae. *Plos One*, 8(9), e74487.
- Logan, B. E (2007). Microbial Fuel Cell. John Wiley and Sons Inc. New Jersey, US.
- Logan, B.E., Aelterman, P., Hamelers, B., Rozendal, R., Schroder, U., Keller, J., Freguiac, S., Verstraete, W., & Rabaey, K. (2006). Microbial fuel cells: methodology and technology. *Environmental Science and Technology*, 40(17), 5181-5192.
- Logan, B.E., Hamelers, B., Rozendal, R., Schrorder, U., Keller, J., Freguia, S., Aelterman, P., Verstraeta, W., & Rabaey, K. (2006). Microbial fuel cells:

Methodology and technology. *Environmental Sciences and Technology*, 40, 5181-5192.

- Logan, B. E. & Regan, J.M. (2006). Microbial fuel cells: Challenges and applications. *Environmental. Science and Technology*, 41, 5172-5180.
- Lovley, D. R. (2006). Bug juice: harvesting electricity with microorganisms. *Nature Reviews Microbiology*, 4, 497-508.
- Lv, Z., Xie, D., Yue, X., Feng, C, & Wei, C. (2012). Ruthenium oxide-coated carbon felt electrode: A highly active anode for microbial fuel cell applications. *Journal of Power Sources*, 210, 26-31.
- Lyon, D.Y., Buret, F., Vogel, T.M., & Monier, J.M. (2010). Is resistance futile? Changing external resistance does not improve microbial fuel cell performance. *Bioelectrochemistry*, 78, 2-7.
- Magnuson, T. S., Isoyama, N., Hodges-Myerson, A.L., Davidson, G., Maroney, M.J., Geesey, G.G., & Lovley, D.R. (2001). Isolation, characterization and gene sequence analysis of a membraneassociated 89 kDa Fe (III) reducing cytochrome c from *Geobacter sulfurreducens*. *Biochemical Journal*, 359, 147-152.
- Mahadevan, R., Bond, D.R., Butler, J.E., Esteve-Nunez, A., Coppi, M.V., Palsson, B.O., Schilling, C.H., & Lovley, D.R. (2006). Characterization of metabolism in the Fe (III)-reducing organism *Geobacter sulfurreducens* by constraint-based modeling. *Applied and Environmental Microbiology*, 72, 1558-1568.
- Majeed, H.A.S.A., Al-Ahmad, A.Y., & Hussain, K.A. (2011). The preparation, characterization, and the study of the linear properties of a new azo compound. *Journal of Basrah research (Sciences)*, 37(2).
- Maiti, S., Sinha, S.S., & Singh, M. (2017). Microbial decolorization and detoxification of emerging environmental pollutant: cosmetic hair dyes. *Journal of Hazard Matereology*, 338, 356–363.

- Malaysia's Environmental Law, ENVIRONMENTAL QUALITY ACT, 1974, the Malaysia Environmental Quality (Industrial Effluents) Regulations 2009 (PU(A) 434).
- Marcus (2008, January 7). Fuel Cell that Uses Bacteria to Generate Electricity, ScienceDaily. Retrieved December 12, 2018 from www.sciencedaily.com/releases/2008/01/080103101137.htm
- Marshall, R.T. (1985). Standard methods for the microbiological examination of dairy products, 15<sup>th</sup> Edition. American Public Health Association, Washington, D.C.
- Marsili, E., Rollefson, J.B., Baron, D.B., Hozalski, R.M., Bond, D.R. (2008). Microbial biofilm voltammetry: direct electrochemical characterization of catalytic electrode-attached biofilms. *Applied and Environmental Microbiology*, 74(23), 7329-7337.
- Mazari-Hiriart, M., Ponce-de-Leon, S., Lopez-Vidal, Y., Islas-Macias, P., Amieva-Fernandez, R.I., & Quinones-Falconi, F. (2008). Microbiological implications of periurban agriculture and water reuse in Mexico City. *PLoS One*, 3, 2305.
- Meckenstock, R. U., Safinowski, M. & Griebler, C. (2004). Anaerobic degradation of polycyclic aromatic hydrocarbons. *FEMS Microbiology Ecology*, 49, 27-36.
- Meerbergen, K., Willems, K.A., Dewil, R., Impe, J.V., Appels, L., & Lievens, B. (2018). Isolation and screening of bacterial isolates from wastewater treatment plants to decolorize azo dyes. *Journal of Bioscience and Bioengineering*, 125 (4), 448-456.
- Meitl, L.A., Eggleston, C.M., Colberg, P.J., Khare, N., Reardon, C.L., & Shi, L. (2009). Electrochemical interaction of Shewanella oneidensis MR-1 and its outer membrane cytochromes OmcA and MtrC with hematite electrodes. *Geochimica Et Cosmochimica Acta*, 73, 5292-5307.
- Menicucci, J., Beyenal, H., Marsili, E., Veluchamy, R.A., Demir, G., Lewandowski, Z. (2006). Procedure for determining maximum sustainable power

generated by microbial fuel cell. *Environmental Science Technology*, 40, 1062-1068.

- Mezzatesta M. L., Gona F., & Stefani S. (2012). Enterobacter cloacae complex: clinical impact and emerging antibiotic resistance. Future Microbiology, 7, 887-902.
- Min, B., Cheng, S., & Logan, B.E. (2005). Electricity generation using membrane and salt bridge microbial fuel cells. *Water Research*, 9, 1675-1686.
- Min, B., Kim, J.R., Oh, S.E., Regan, J.M, & Logan, B.E. (2005). Electricity generation from swine wastewater using microbial fuel cells. *Water Resources*, 39(20), 4961-4968.
- Mohamed, A. R., and Lee, K. T. (2006). Energy for sustainable development in Malaysia: Energy policy and alternative energy. Energy Policy, 34(15), 2388-2397.
- Mohan, Y., Kumar, S.M.M., & Das, D. (2008). Electricity generation using microbial fuel cells. *International Journal of Hydrogen Energy*, 33, 423-426.
- Mohan, S.V., Saravanan, R., Veer, S.R., Mohanakrishna, G., & Sarma, P.N. (2006).
   Bioelectricity production from wastewater treatment in dual chambered microbial fuel cell (MFC) using selectively enriched mixed microflora: Effect of catholyte. *Bioresource Technology*, 99(3), 596-603.
- Moosvi, S., Keharia, H., & Madamwar, D. (2005). Decolourization of textile dye Reactive Violet 5 by a newly isolated bacterial consortium RVM 11.1. *World Journal of Microbiology and Biotechnology*, 21, 667-672.
- Morita, M., Malvankar, N.S., Franks, A.E., Summers, Z.M., Giloteaux, L., Rotaru, A.E., Rotaru, C., & Lovley, D.R. (2011). Potential for direct interspecies electron transfer in methanogenic wastewater digester aggregates. *MBio*, 2, e00159-00111.
- Muhamad Firdaus b. Sabaruddin (2018). Assessment of Acid Red 27 Biodecolourisation and Biogenic Electricity Generation in Stacked

Microbial Fuel Cell by Citrobacter freudii A1 and Enterococcus casseliflavus C1. University of Technology Malaysia: Master Thesis

- Nachiyar, C.V. & Rajkumar, G.S. (2003). Degradation of tannery and textile dye, Navitan Fast Blue S5R by *Pseudomonas aeruginosa*. World Journal of Microbiology and Biotechnology, 19, 609-614.
- Nakanish, N. & Solomon, P. (1977). Infrared Absorption Spectroscopy. *Holden-Day*, Inc London, Sydney, 8(11).
- Nadzri, Y. (2013). The Way Forward: Solid Waste Managment in Malaysia. 10th Annual Waste Management Conference & Exhibition, Malaysia, 10-19 July, 2013.
- Nawahwi, M., Ibrahim, Z., and Yahya, A. (2013). Degradation of the Azo Dye Reactive Red 195 by *Paenibacillus* spp. R2. *Journal of Bioremedation and Biodegradation*, 4: 174.
- Nazina, T. N., Grigor'yan, A. A. Shestakova, N. M. Babich, T. L. Ivoilov, V. S., Feng, Q. Ni, F. Wang, J. She, Y. Xiang, T. Luo, Z. Belyaev, S. S. & Ivanov M. V. (2007). Microbiological investigations of high-temperature horizons of the Kongdian petroleum reservoir in connection with field trial of a biotechnology for enhancement of oil recovery. *Microbiology*, 76, 287-296.
- Nevin, K.P., Kim, B.C., Glaven, R.H., Johnson, J.P., Woodard, T.L., Methe B.A., Jr, R.J.D., Covalla, S.F., Franks, A.E., Liu, A., Lovley, D.R. (2009). Anode biofilm transcriptomics reveals outer surface components essential for high density current production in Geobacter sulfurreducens fuel cells. *PloS one*, 4, 5628.
- Nevin, K. P., Richter, F., Covalla, S.F., Johnson, J.P., Woodard, T.L., Orloff, A.L., Jia, H.,Zhang, M., & Lovley, D.R. (2008). Power output and columbic efficiencies from biofilms of Geobacter sulfurreducens comparable to mixed community microbial fuel cells. *Environmental Microbiology*, 10, 2505-2514.

- Niessen, J., Schroder, U., & Scholz F. (2004). Exploiting complex carbohydrates for microbial electricity generation-a bacterial fuel cell operating on starch. *Electrochemistry Communications*, 6, 955-958.
- Nigam, P., Banat, I.M., Singh, D., & Marchant, R. (1996). Microbial process for the decolonization of textile effluent containing Azo, diAzo and reactive dyes. *Process Biochemistry*, 31, 435-442.
- Nimje, V.R., Chen, C.Y., Chen, C.C., Tsai, J.Y., Chen, H.R., Huang, Y.M., Jean, J.S., Chang, Y.F., & Shih, R.C. (2011). Microbial fuel cell of *Enterobacter cloacae*: Effect of anodic pH microenvironment on current, power density, internal resistances and electrochemical losses. *International Journal of Hydrogen Energy*, 36, 11093-11101.
- Nishijima, K. A., Couey, H. M., & Alvarez, A. M. (1987). Internal yellowing, a bacterial disease of papaya fruits caused by *Enterobacter cloacae*. *Plant Disease*, 71, 1029-1034.
- Nishijima, K.A. (1999). Enterobacter cloacae. Crop Knowledge Master, http://www.extento.hawaii.edu/Kbase/crop/Type/e\_cloac.htm
- Offei, F., Thygesen, A., Mensah, M., Tabbicca, K., Fernando, D., Petrushina, I., & Daniel, G. (2016). A viable electrode material for use in microbial fuel cells for tropical regions. *Energies*, 9(35), en9010035.
- Oh, S.E., Min, B., & Logan, B.E. (2004). Cathode performance as a factor in electricity generation in microbial fuel cells. *Environmental Science Technology*, 38, 4900–4904.
- Oh, S.E. & Logan, B.E. (2005). Hydrogen and electricity production from a food processing wastewater using fermentation and microbial fuel cell technologies. *Water Research*, 39, 4673-4682.
- Olivier, S., Fredric, B., & Keith, B. (2008). Bacteria and yeasts as catalysts in microbial fuel cells: electron transfer from microorganisms to electrodes for green electricity. *Energy and Environmental Science*,1, 607-620.

- Ong, S.A., Tooriska, E., Hirata, M., Hano, T. (2005). Treatment of azo dye OrangeII in a sequential anaerobic and aerobic-sequencing batch reactor system. *Environmental Chemistry Letters*, 2, 203-207.
- Padmavathy, S., Sandhya, S., Swaminathan, K., Subrahmanyam, Y.V., & Kaul, S.N. (2003). Comparison of decolorization of reactive Azo dyes by microorganisms isolated from various sources. *Journal of Environmental Science*, 15, 628-633.
- Padmavathy, S., Sandhya, S., Swaminathan, K., Subramanium, Y.V., Chakrabarti, T., & Kaul, S.N. (2003). Aerobic decolorization of reactive azo dyes in presence of various co-substrates. *Chemical and Biochemical Engineering Quarterly*, 17(2), 147-151.
- Pagga, U. & Brown, D. (1986). The degradation of dyestuffs: Part II. Behavior of dyestuffs in aerobic biodegradation tests. *Chemosphere*, 15, 479-491.
- Palanivelan, R., Rajakumar, S. & Ayyasamy, P.M. (2011). Effect of various carbon and nitrogen sources on decolorzation of textile dye remazol golden yellow using bacterial species. *Journal of Environmental Biology*, 35, 781-787.
- Pandey, A., Singh, P., & Lyenger, L. (2007). Bacterial decolourisation and degradation of azo dyes. *International Biodeterioration and Biodegradation*, 59, 73-84.
- Pang, Y. L., and Abdullah, A. Z. (2013). Current status of textile industry w astewater management and research progress in Malaysia: a review. CLEAN–Soil, Air, Water, 41(8), 751-764.
- Pant, D., Bogaert, G.V., Diels, L., & Vanbroekhoven, K. (2010). A review of the substrates used in microbial fuel cells (MFCs) for sustainable energy production. *Bioresource Technology*, 101(6), 1533-1543.
- Park, H. S., Kim, B.H., Kim, H.S., Kim, H.J., Kim, G.T., Kim, M., Chang, I.S., Park, Y.K., & Chang, H.I. (2001). A novel electrochemically active and Fe (III)reducing bacterium phylogenetically related to *Clostridium butyricum* isolated from a microbial fuel cell. *Anaerobe*, 7, 297-306.

- Park, D.H., & Zeikus, J.G. (2000). Electricity generation in microbial fuel cells using neutral redas an electronophore. *Applied and Environmental Microbiology*, 66 (4), 1292–1297.
- Park, D.H. & Zeikus, J.G. (2003). Improved fuel cell and electrode designs for producing electricity from microbial degradation. *Biotechnology and Bioengineering*, 81, 348-355.
- Parkash, A. (2016). Utilization of distillery wastewater for electricity generation using microbial fuel cell. *Journal of Applied and Emerging Science*, 6(2), 79-86
- Parkash, A., Aziz, S., & Sa, S. (2015). Impact of Salt Concentrations on Electricity Generation using Hostel Sludge Based Dual Chambered Microbial Fuel Cell. *Journal of Bioprocessing & Biotechniques*, 5(8), 1-6.
- Parthipan, P., Elumalai, P., Karthikeyan, O.P., Ting, Y.P., & Rajasekar, A. (2017). Review on biodegradation of hydrocarbon and their influence on corrosion of carbon steel with special reference to petroleum industry. *Journal of Environmental Biotechnology Resources*, 6 (1), 12-33.
- Parthipan, P., Preetham, E., Machuca, L.L., Rahman, P.K.S.M., Murugan, K., & Rajasekar, A. (2017). Biosurfactant and degradative enzymes mediated crude oil degradation by bacterium Bacillus subtilis A1. *Frontiers in Microbiology*, 8, 193.
- Parvin, F., Rahman, M.M., & Islam, M.M. (2015). Isolation of Mixed Bacterial Culture from Rajshahi Silk Industrial Zone and Their Efficiency in Azo Dye Decolorization. *Indian Journal of Science and Technology*, 8(10), 950–957.
- Peju, G. & Rajat, J. (1906). Note sur le polymorphisme des bactéries sans l'urée. Comptes *Rendus Social Biologies*, 61, 477.
- Pham, C. A., Jung, S.J., Phung, N.T., Lee, J., Chang, I.S., Kim, B.H., Yi, H., & Chun, J. (2003). A novel electrochemically active and Fe (III)-reducing bacterium phylogenetically related to *Aeromonas hydrophila*, isolated from a microbial fuel cell. *FEMS Microbiology Letters*, 223, 129-134.

- Pinto, R.P., Srinivasan, B., Guiot, S.R., & Tartakovsky, B. (2011). The effect of realtime external resistance optimization on microbial fuel cell performance. *Water Resources*, 45, 1571-1578.
- Potter, M.C. (1912). Electrical effects accompanying the decomposition of organic compounds. *Proceedings of the Royal Society B: Biological Sciences*, 84, 260-276.
- Prasad, S.S. & Aikat, K. (2014). Study of bio-degradation and bio-decolourisation of azo dye by *Enterobacter sp.* SXCR. *Environmental Technology*, 35(8), 956-965.
- Priya, B., Uma, L., Ahamed, A.K., Subramaniam, G., & Prabaharan, D. (2011). Ability to use the diazo dye, CI Acid Black 1 as a nitrogen source by the marine cyanobacterium; Oscillatoria curviceps BDU92191. Bioresource Technology, 102, 7218-7223.
- Qiao, Y., Li, C.M., Bao, S.J., & Bao, Q.L. (2007). Carbon nanotube/ polyaniline composite as anode material for microbial fuel cells. *Journal of Power Sources*, 170, 79-84.
- Quintanilha, A. G., Zilberstein, B., Santos, M.A., Pajecki, D., Moura, E.G., Alves, P.R., Maluf-Filho, F., & Cecconello, I. (2007). A novel sampling method for the investigation of gut mirobiota. *World Journal of Gastroenterol*, 13, 3990-3995.
- Rabaey, K., Boon, N., Dendf, V., Verhaege, M., Ho<sup>--</sup>fte, M., & Verstraete, W., (2004a).
   Bacteria produce and use redox mediators for electron transfer in microbial fuel cells. Extended abstract, Proceedings of the 228th ACS Meeting, *American Chemical Society*, Philadelphia, PA.
- Rabaey, K., Boon, N., Siciliano, S.D., Verhaege, M., & Verstraete, W. (2004b).
  Biofuel cells select for microbial consortia that self-mediate electron transfer. *Applied and Environmental Microbiology*, 70 (9), 1–10.
- Rabaey, K., Boon, N., Siciliano, S.D., Verhaege, M., & Verstraete, W. (2004c). Biofuel cells select for microbial consortia that self-mediate electron transfer, *Applied Environmental Microbiology*, 70, 5373-5382.

- Rabaey, K., Boon, N., Hofte, M., & Verstraete, W. (2005). Microbial phenazine production enhances electron transfer in biofuel cells, *Environmental Science and Technology*, 39, 3401-3408.
- Rabaey, K., Lissens, G., Sicilliano, S.D., & Verstraete, W. (2003). A microbial fuel cell capable of converting glucose to electricity at high rate and efficiency. *Biotechnology Letters*, 25, 1531–1535.
- Rabaey, K. & Verstraete, W. (2005). Microbial fuel cells: novel biotechnology for energy generation. *Trends Biotechnology*, 23, 291-298.
- Rafii, F. & Cerniglia, C.E. (1995). Reduction of Azo dyes and nitroaromatic compounds by bacterial enzymes from the human intestinal tract. *Environmental Health Perspectives*, 103, 17-19.
- Rahimnejad, M., Adhami, A., Darvari, S., Zirepour, A., and Oh, S. E. (2015). Microbial fuel cell as new technology for bioelectricity generation: a review. Alexandria Engineering Journal, 54(3), 745-756.
- Rahimnejad, M., Jafari, T., Haghparast, F., Najafpour, G.D., & Goreyshi, A.A. (2010).
   Nafion as a nanoproton conductor in microbial fuel cells. *Turkish Journal Engineering and Environmental Science*, 34, 289-292.
- Ramya, M., Iyappan, S., Manju, A., & Jiffe, J.S. (2010). Biodegradation and decolorization of Acid Red by Acinetobacter radioresistens. *Journal of Bioremediation and Biodegradation*, 1, 105.
- Reguera, G., McCarthy, K.D., Mehta, T., Nicoll, J.S., Tuominen, M.T., & Lovley, D.R. (2005). Extracellular electron transfer via microbial nanowires. *Nature*, 435, 1098-1101.
- Rezaei, F., Xing, D., Wagner, R., Regan, J.M., Richard, T.L., & Logan, B.E. (2009) Simultaneous cellulose degradation and electricity production by *Enterobacter cloacae* in a microbial fuel cell. *Applied and Environmental Microbiology*, 75 (11), 3673-3678.
- Ringeisen, B.R., Henderson, E., Wu, P.K., Pietron, J., Ray, R., Little, B., Biffinger, J.C., & Jones-Meehan, J.M. (2006). High power density from a miniature

microbial fuel cell using Shewanella oneidensis DSP10. *Environmental Science and Technology*, 40, 2629–2634.

- Rieger, P.G., Meir, H.M., Gerle, M., Vogt, U., Groth, T., & Knackmuss, H.J. (2002). Xenobiotics in the environment: present and future strategies to obviate the problem of biological persistence. *Journal of Biotechnology*, 94,101-123.
- Richter, H., Nevin, K.P., Jia, H.F., Lowy, D.A., Lovley, D.R., & Tender, L.M. (2009). Cyclic voltammetry of biofilms of wild type and mutant *Geobacter sulfurreducens* on fuel cell anodes indicates possible roles of OmcB, OmcZ, type IV pili, and protons in extracellular electron transfer. *Energy and Environmental Science*, 2, 506–516.
- Rismani, Y.H., Christy, A., Carver, S.M., Yu, Z., Dehority, B.A., & Tuoviven, O.H. (2011). Effect of external resistance on bacterial diversity and metabolism in cellulose-fed microbial fuel cells. *Bioresources Technology*, 102, 278-283.
- Rosado, A.S., Duarte, G.F., Seldin, L., & Van Elsas, J.D. (1998). Genetic diversity of nifH gene sequences in *Paenibacillus azotofixans* strains and soil samples analyzed by denaturing gradient gel electrophoresis of PCR-amplified gene fragments. *Applied and Environmental Microbiology*, 64(8), 2770-2779.
- Roxon, J.J., Ryan, A.J., & Wright, S.E. (1967). Enzymatic reduction of tartrazine by Proteus vulgaris from rats. *Food and Cosmetics Toxicology*, 5, 645-656.
- Roy, J.N., Babanova, S., Garcia, K.E., Cornejo, J., Ista, L.K., & Atanassov, P. (2014). Catalytic biofilm formation by *Shewanella oneidensis* MR-1 and anode characterization by expanded uncertainty. *Electrochimica Acta*, 126, 3-10.
- Rozendal, A.R., Hamelers, M. V. H., & Buisman, N.C. (2006). Effects of membrane cation transport on pH and microbial fuel cell performance. *Environmental Science and Technology*, 40, 5206-5211.

- Russ, R., Rau, J., & Stolz, A. (2000). The function of cytoplasmic flavin reductases in the reduction of Azo dyes by bacteria. *Applied and Environmental Microbiology*, 66, 1429-1434.
- Sabnis, R.W. (2010). Handbook of Biological Dyes and Stains: Synthesis and Industrial Applications. *John Wiley & Sons*, 19.
- Sahasrabudhe, M. & Pathade, G. (2012). Decolourisation and degradation of C.I. Reactive Red 195 by Georgenia sp.CC-NMPT-T3. Indian Journal of Experimental Biology, 50, 290-299.
- Samrot, A.V., Senthilkumar, P., Pawankumar, K., Akilandeswar, G.C., Rajalakshmi, N., & Dhathathreyan, K.S. (2010). Electricity generation by *Enterobacter cloacae* SU-1 in mediator less microbial fuel cell. *International Journal of Hydrogen Energy*, 35(15), 7723-7729.
- Sarioglu, M., Bali, U., & Bisgin, T. (2007). The removal of C.I basic red 46 in a mixed methanogenic anaerobic culture. *Dyes and Pigments*, 74, 223-229.
- Sarvajith, M., Reddy, G.K.K., & Nancharaiah, Y. (2018). Textile dye biodecolourization and ammonium removal over nitrite in aerobic granular sludge sequencing batch reactors. *Journal of Hazardous Material*, 342, 536-543.
- Schroder, U. (2007). Anodic electron transfer mechanisms in microbial fuel cells and their energy efficiency, *Physical Chemistry Physics*, 2619-2629.
- Semdé, R., Pierre, D., Geuskens, G., Devleeschouwer, M., & Moes, A.J. (1998).Study of some important factors involved in Azo derivative reduction by *Clostridium perfringens. International Journal of Pharmaceuticals*, 161, 45-54.
- Sevda, S. & Sreekrishnan, T.R. (2012). Effect of salt concentration and mediators in salt bridge microbial fuel cell for electricity generation from synthetic wastewater. *Journal of Environmental Science and Health*, 47, 878-886.
- Seviour, T., Doyle, L.E., Lauw, S.J.L., Hinks, J., Rice, S.A., Nesatyy, V.J., Webster, R.D., Kjelleberg, S., & Marsili, E. (2015). Voltammetric profiling of

redox-active metabolites expressed by Pseudomonas aeruginosa for diagnostic purposes. *Chemical Community*, 51, 3789-3792.

- Sleutels, T. H., Ter Heijne, A., Buisman, C. J., and Hamelers, H. V. (2012). Bioelectrochemical systems: an outlook for practical applications. ChemSusChe, 5(6), 1012-1019.
- Shabnam, S.M. & Moghaddam, M.R.A. (2016). Aerobic granular sludge for dye biodegradation in a sequencing batch reactor with anaerobic/aerobic cycles. *Clean Soil Air Water*, 44, 438-443.
- Shahmoradi, B., Maleki, A., & Byrappa, K. (2011). Photocatalytic degradation of amaranth and brilliant blue FCF dyes using in situ modified tungsten doped TiO2 hybrid nanoparticles. *Catalysis Science and Technology*, 1, 1216-1223.
- Shankar, R., Pathak, N., Mondal, P., Chaurasia, A.K., & Chand, S. (2017). Energy Production through Microbial Fuel Cell. Sustainable Utilization of Natural Resources, 353-380
- Sharifuddin, S.S. & Ying, A.S. (2017). Current situation and issues of industrial wastewater management in Malaysia. 13<sup>th</sup> WEPA Annual Meeting and WEPA International Workshop on Industrial Wastewater Management, 26-28 September, Jakarta Indonesia.
- Sharma, K., Sharma, S., Sharma, S., Singh, P., Kumar, S., Grover, R. and Sharma, P. (2007). A comparative study on characterization of textile wastewaters (untreated and treated) toxicity by chemical and biological tests.Chemosphere, 69(1), 48-54.
- Shi, L., Squier, T.C., Zachara, J.M., & Fredrickson, J.K. (2007). Respiration of metal (hydr) oxides by *Shewanella* and *Geobacter*: a key role for multihaem ctype cytochromes. *Molecular Microbiology*, 65, 12-20.
- Shijie, Y., Zhao, Q., Zhang, J., Jiang, J., & Zhao, S. (2006). A microbial fuel cell using permanganate as the cathodic electron acceptor. *Journal of power sources*, 162(2), 1409-1415.

- Siebel, D., Bennetto, H.P., Delaney, G.M., Mason, J.R., Stirling, J.L., & Thurston, C.F. (1984). Electron-transfer coupling in microbial fuel cells: 1. Comparison of redoxmediator reduction rates and respiratory rates of bacteria. *Journal* of Chemical Technology and Biotechnology, 34, 3–12.
- Sigma. Sigma Aldrich catalogue. http://www.sigmaaldrich.com/catalog/product/ aldrich/204048 (2012)
- Song, T.S., Yan, Z.S., Zhao, Z.W., & Jiang, H.L. (2010). Removal of organic matter in freshwater sediment by microbial fuel cells at various external resistances. *Journal of Chemical Technology and Biotechnology*, 85, 1489-1493.
- Srikanth, S., Marsili, E., Flickinger, M.C., & Bond, D.R. (2008). Electrochemical characterization of Geobacter sulfurreducens cells immobilized on graphite paper electrodes. *Biotechnology and Bioengineering*, 99, 1065-1073.
- Steter, J. R., Barros, W. R., Lanza, M. R., & Motheo, A. J. (2014). Electrochemical and sonoelectrochemical processes applied to amaranth dye degradation. *Chemosphere*, 117, 200-207.
- Stroud, J. L. Paton, G. I. & Semple K. T. (2007). Microbe-aliphatic hydrocarbon interactions in soil implication for biodegradation and bioremediation. *Journal of Applied Microbiology*, 102, 1239-1253.
- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of Azo dyes. Applied Microbiology and Biotechnology, 56, 69-80.
- Sugiura, W., Miyashita, T., Yokoyama, T., Arai, M. (1999). Isolation of Azo-dye degrading microorganisms and their application to white discharge printing of fabric. *Journal of Bioscience*, 88(5), 577-581.
- Surygała, J. (2001). Crude oil and environment (in Polish). Oficyna Wydawnicza Politechniki Wrocławskiej, Wrocław, 5, 114-136.

- Summers, Z.M., Fogarty, H.E., Leang, C., Franks, A.E., Malvankar, N.S., & Lovley, D.R. (2010). Direct exchange of electrons within aggregates of an evolved syntrophic coculture of anaerobic bacteria. *Science*, 330, 1413-1415.
- Sun, J., Hu, Y.Y., Bi, Z., & Cao, Y.Q. (2009). Simultaneous decolorization of azo dye and bioelectricity generation using a microfiltration membrane air-cathode single-chamber microbial fuel cell. *Bioresource Technology*, 100, 3185-3192.
- Sun, M., Mu, Z., Chen, G., Sheng, G., Liu, X., & Chen, Y. (2009). Microbe-assisted sulfide oxidation in the anode of a microbial fuel cell. *Environmental Science Technology*, 43, 3372-3377.
- Suzuki, S., Karube, I., & Matsunaga, T. (1978). Application of a biochemical fuel cell to wastewaters. *Biotechnology and Bioengineering*. *Symposium*, 8, 501– 511
- Suzuki, Y., Yoda, T., Ruhul, A., & Sugiura, W. (2001). Molecular cloning and characterization of the gene coding for azoreductase from *Bacillus sp* OY1-2 isolated from soil. *Journal of Biological Chemistry*, 276, 9059-9065
- Toledo, F.L., Calvo, C., Rodelas, B., & Gonzalez-Lopez, J. (2006). Selection and identification of bacteria isolated from waste crude oil with polycyclic aromatic hydrocarbons removal capacities. *Systematic and Applied Microbiology*, 29(3), 244-252.
- Vilar, E.O., de Freitas, N.L., de Lirio, F.R., & de Sousa, F.B. Study of the electrical conductivity of graphite felt employed as a porous electrode. *Brazilian Journal of Chemical Engineering*, 15(3), 295-302.
- Virdis, B., Harnisch, F., Batstone, D.J., Rabaey, K., & Donose, B.C. (2012). Noninvasive characterization of electrochemically active microbial biofilms using confocal Raman microscopy. *Energy and Environmental Science*, 5, 7017-7024.
- Vos, P.D., Garrity, G.M., Jones, D., Krieg, N.R., Ludwig, W., Rainey, F.A., Schleifer,K.H., & Whitman, W.B. (2009). Bergey's manual of systematic

bacteriology: The firmicutes. *Springer*, Dordrecht Heidelberg London New York, 2(3), Ed: Whitman, W.B., Parte, A.C., Goodfellow, M., Kampfer, P., Vos, P.D., Rainey, F.A., Schleifer, K.H. ISBN: 978-0-387-95041-9, 865.

- Walker, R. (1970). The metabolism of Azo compounds: A review of the literature. Food and Cosmetics Toxicology, 8, 659-676.
- Wang, H., Park, J.D., & Ren, Z.J. (2015). Practical Energy Harvesting for Microbial Fuel Cells: A Review. *Environmental Science and Technology*, 49(6), 3267–3277.
- Wang, H., Zheng, Z.W., Su, J.Q., Tian, Y., Xiong, X.J., Zheng, T.L. (2009). Biological decolorization of the reactive dyes Reactive Black 5 by a novel isolated bacterial strain *Enterobacter sp.* EC3. *Journal of Hazardous Materials*, 171, 654-659.
- Weber, E.J. & Wolfe, L.N. (1987). Kinetic studies of the reduction of aromatic Azo compounds in anaerobic sediment/water systems. *Environmental Toxicology and Chemistry*, 6, 911-919.
- Widdel, F. & Rabus, R. (2001). Anaerobic biodegradation of saturated and aromatic hydrocarbons. *Biotechnology*, 12, 259-276.
- WingardJr., L.B., Shaw, C.H., & Castner, J.F. (1982). Bioelectrochemical fuel cells. *Enzyme Microbial Technology*, 4, 137–142.
- Wolicka, D. & Borkowski, A. (2012). Microorganisms and Crude Oil, Introduction to Enhanced Oil Recovery (EOR) Processes and Bioremediation of Oil-Contaminated Sites, Dr. Laura Romero-Zerón (Ed.), ISBN: 978-953-51-0629-6, InTech, 114-142.
- Wuhrmann, K., Mechsner, K., & Kappeler, T. (1980). Investigation on ratedetermining factors in the microbial reduction of Azo dyes. *European Journal of Applied Microbiolology and Biotechnology*, 9, 325-338.

- Xing, D., Zuo, Y., Cheng, S., Regan, J.M., & Logan, B.E. (2008). Electricity generation by *Rhodopseudomonas palustris* DX-1. *Environmental Science* and Technology, 42, 4146-4151.
- Xu, M., Guo, J., Sun, G. (2007). Biodegradation of textile azo dye by Shewanella decolorationis S12 under microaerophilic conditions. *Applied Microbiology and Biotechnology*, 76, 719-726.
- Yang, Y., Wyatt II, D.T., & Bahorshky, M. (1998). Decolorization of Dyes Using UV/H2O2 photochemical oxidation. *Textile Chemist and Colorist*, 30, 27-35.
- Yi, H., Nevin, K.P., Kim, B.C., Franks, A.E., Klimes, A., Tender, L.M., & Lovley, D.R. (2009). Selection of a variant of *Geobacter sulfurreducens* with enhanced capacity for current production in microbial fuel cells. *Biosensor* and Bioelectronics, 24, 3498-3503.
- Yu, J., Wang, X., & Yue, P.L. (2001). Optimal decolorization and kinetic modeling of synthetic dyes by *Pseudomonas* strains. *Water Research*, 35, 3579-3586.
- Zaharia, C., Suteu, D., & Muresan, A. (2012). Options and solutions for textile effluent decolorization using some specific physico-chemical treatment steps. *Environmental Engineering and Management Journal*, 11, 493-509.
- Zhang, T., Cui, C., Chen, S., Ai, X., Yang, H., Shen, P., & Peng, Z. (2006). A novel mediatorless microbial fuel cell based on direct biocatalysis of *Escherichia coli. Chemical Communications*, 2257-2259.
- Zhang L, Zhu X, Li J, Qiang L, and Ye D (2011) Biofilm formation and electricity generation of a microbial fuel cell started up under different external resistances. Journal of Power Sources, 196, 6029–6035
- Zhu, N., Chen, X., Zhang, T., Wu, P., Li, P., & Wu, J. (2011). Improved performance of membrane free single-chamber air-cathode microbial fuel cells with nitric acid and ethylenediamine surface modified activated carbon fiber felt anodes. *Bioresource Technology*, 102, 422-426.

- Zhuang, L., Zhou, S., Yuan, Y., Liu, T., Wu, Z., & Cheng, J. (2011). Development of *Enterobacter aerogenes* fuel cells: From in situ biohydrogen oxidization to direct electroactive biofilm. *Bioresource Technology*, 102, 284-289.
- Zhuang, W.Q., Tay, J.H., Maszenan, A.M., Krumholz, L.R/, & Tay, S.T. (2003). Importance of gram-positive naphthalene-degrading bacteria in oilcontaminated tropical marine sediments. *Letter of Applied Microbiology*, 36(4), 251-257.
- Zhuwei, D., Li, H., & Gu, T. (2007). A state-of-the-art review on microbial fuel cells: a promising technology for wastewater treatment and bioenergy. *Biotechnology advances*, 25(5), 464-482.
- Zimmermann, T., Kulla, H., & Leisinger, T. (1982). Purification and properties of orange II Azoreductase from *Pseudomonas* KF46. *Experientia*, 38, 1380-1384.
- Zimmermann, T., Kulla, H.G., & Leisinger, T. (1982). Properties of purified orange II azoreductase the enzyme initiating azo dye degradation by *Pseudomonas* KF46. *European Journal of Biochemistry*,129, 197-203.
- Zuo, Y., Xing, D., Regan, J.M., & Logan, B.E. (2008). Isolation of the exoelectrogenic bacterium Ochrobactrum anthropi YZ-1 by using a U-tube microbial fuel cell. Applied and Environmental Microbiology, 74, 3130-3137.

## LIST OF PUBLICATIONS:

Yahya, N.F.A.B., Khiavi, N.D., & Ibrahim, N. (2016). Green Electricity Production by *Epipremnum aureum* and Bacteria in Plant Microbial Fuel Cell. Journal of Advanced Research in Applied Sciences and Engineering Technology, 5(1), 22-31