

**HEXADECYLTRIMETHYLMAMMONIUM-SILVER-KAOLINITE AS A MARINE
ANTI-BIOFOULING PAINT ADDITIVE**

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A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy

Faculty of Science
Universiti Teknologi Malaysia

NOVEMBER 2021

ACKNOWLEDGEMENT

Firstly, I would like to extend my deepest appreciation to my supervisor Ts. Dr. Wan Rosmiza Zana Wan Dagang for her constant drive and encouragement, unwavering positivity, helpful discussions, and valuable insights. Without her support and guidance, I would not be able to complete my research. Many thanks also to my co-supervisors, Assoc. Prof. Ts. ChM. Dr. Nik Ahmad Nizam Nik Malek and Dr. Haryati Binti Jamaluddin for the always enthusiastic support, useful suggestions, and wise words of encouragement.

I would like to take this opportunity to show my greatest appreciation and gratitude to my best colleagues, Mr. Herman Tuminoh, Mr. Amin Jumat and Dr. Jibrin Ndejiko for the stimulating discussions, sharing valuable comments and suggestions, cheered me on, celebrated each accomplishment and for all the fun we have had during these five years. Also, I thank these brilliant friends and colleagues who inspired me over the years with prompt support and fun memories: Dr. Chitra Selvaratnam, Dr. Suganthy Thevarajoo and Mrs. Kavita Supparmaniam.

To the staffs of the University Industry Research Laboratory (UIRL), especially Mrs. Amy Zuria Abdul Ajid, Mrs. Zahara Salleh and Mr. Izzam Idrus, I am forever indebted to all of you for your skilful technical support, helpful discussions and your kindness and enthusiasm in supporting my work. Thank you also to Mrs. Mas Linda Yunos from the School of Biomedical Engineering and Health Sciences, Faculty of Engineering for giving me all necessary assistance during CLSM analysis.

Not to be forgotten, to all the administration and laboratory staff at the Faculty of Science especially Ms. Zaleha Jaafar and Mr. Muhammad Hanif Muhammad Hatta, I am grateful for your wise tips, cheerfulness and unflagging enthusiasm when entertaining my self-doubts and uncertain questions.

I am ever grateful for the Zamalah scholarship from the Universiti Teknologi Malaysia (UTM) for sponsoring my studies for three years. I also thankful for the partial tuition fee waiver granted by Academic Management Division (AMD).

Last but not the least, I would like to thank my family: *my* parents (Hj. Noor Al Amin Yunos and Hjh. Rohani Ahmad @ Hassan) and siblings (Fatin, Fadhilah, Syafiq and Nasruddin) for their constant encouragement and unwavering support in all my pursuits. I am humbled by their love and loyalty. Special thanks with love to Mr. Hairol Anuar Haron, my husband for understanding me best and has been my best friend with great companion, loved, supported, encouraged, entertained, as well as helped me get through this agonizing period in the most positive way. This work is especially dedicated to my beloved sons, Abdul Qayyum and Abdul Wafiy, to show them that learning is a lifelong journey.

ABSTRACT

Copper-based anti-biofouling paint coating has been used to combat biofouling on man-made artificial marine structures for several decades. Despite its widespread use, the massive accumulation of copper and its compounds causes a serious threat to marine ecosystems. Thus, the search for efficient and environmentally friendly anti-biofouling coating has become more significant. Therefore, this current study focused on developing an anti-biofouling paint additive employing the active compounds, silver ion (Ag^+) and surfactant hexadecyltrimethylammonium (HDTMA) embedded in the kaolinite clay structure which is designated as Organo-silver-kaolinite (Or-Ag-Kao). The developed paint additive is intended for applications in marine artificial structures. The Or-Ag-Kao was prepared by adding Ag-kaolinite with HDTMA via a cation exchange process. The prepared samples; Ag-Kao, HDTMA-Kao and Or-Ag-Kao were characterized using ATR-FTIR and EDX. In this study, these analyses confirmed that the Ag^+ and HDTMA were successfully intercalated into the kaolinite without affecting their original structure. The samples were then incorporated into the commercial paint matrix to test for their anti-bacterial adhesion and anti-biofilm efficiency as an anti-biofouling coating additive against isolated marine biofouling bacteria, *Bacillus niabensis* and *Alteromonas litorea*. It was observed Or-Ag-Kao paint coating efficiently reduced the bacterial adhesion of *B. niabensis* (91.51%) and *A. litorea* (87.53%), as compared to commercial paint without additive. The Or-Ag-Kao paint coating also showed excellent anti-biofilm on *B. niabensis* and *A. litorea* with EPS mean volume of $0.219 \pm 0.129 \times 10^5 \mu\text{m}^3$ and $0.151 \pm 0.052 \times 10^5 \mu\text{m}^3$, respectively. A weak interaction was observed between the model bacteria and Or-Ag-Kao with commercial paint coated surface with the mean lateral detachment force 139.4004 nN (*B. niabensis*) and 146.2251 nN (*A. litorea*). Major contact surface area reduction was also observed in Or-Ag-Kao paint coating ($0.275 \mu\text{m}^2$ in *B. niabensis* and $0.391 \mu\text{m}^2$ in *A. litorea*) indicated that Or-Ag-Kao paint coating successfully minimized surface bacterial attachment. As a conclusion, the broad-spectrum effectiveness of Or-Ag-Kao paint coating minimized bacterial adhesion and biofilm formation may provide an environmental-friendly solution for marine biofouling. It is envisaged that the newly developed Or-Ag-Kao has high potential to be commercialized as an alternative marine anti-biofouling coating replacing the present traditional antifouling paints.

ABSTRAK

Lapisan cat anti-biopenempelan semula jadi berasaskan tembaga telah digunakan bagi menangani biopenempelan semula jadi pada struktur laut buatan selama beberapa dekad. Walaupun digunakan secara meluas, pengumpulan tembaga dan sebatianya secara besar-besaran mampu menyebabkan ancaman serius terhadap ekosistem marin. Oleh itu, pencarian lapisan anti- biopenempelan semula jadi yang cekap dan mesra alam menjadi lebih penting. Oleh itu, kajian semasa ini difokuskan kepada pembangunan bahan tambahan cat anti-biopenempelan semula jadi menggunakan sebatian aktif, ion perak (Ag^+) dan surfaktan heksadesiltrimetil ammonium (HDTMA) yang dibenam dalam struktur tanah liat kaolinit yang dinamakan sebagai Organo-perak-kaolinit (Or-Ag-Kao). Cat anti-biopenempelan semula jadi yang dibangunkan akan diaplikasikan kepada struktur buatan marin. Or-Ag-Kao disediakan dengan menambahkan Ag-kaolinit dengan HDTMA melalui proses pertukaran kation. Sampel yang disediakan; Ag-Kao, HDTMA-Kao dan Or-Ag-Kao dicirikan menggunakan ATR-FTIR dan EDX. Dalam kajian ini, analisis ini mengesahkan bahawa Ag^+ dan HDTMA telah berjaya dimuatkan ke dalam struktur kaolinit tanpa menjaskan struktur asalnya. Sampel dimasukkan ke dalam matriks cat komersial bagi menguji kecekapan anti-bakteria dan anti-biofilem sebagai bahan tambahan bagi lapisan cat anti-biopenempelan semula jadi terhadap bakteria biopenempelan semula jadi yang telah dipencarkan, *Bacillus niabensis* dan *Alteromonas litorea*. Diperhatikan bahawa lapisan cat Or-Ag-Kao dengan cekap mengurangkan lekatan bakteria *B. niabensis* (91.51%) dan *A. litorea* (87.53%) berbanding dengan cat komersial tanpa bahan tambahan. Lapisan cat Or-Ag-Kao juga menunjukkan anti-biofilem yang sangat baik terhadap *B. niabensis* dan *A. litorea* dengan purata isipadu EPS masing-masing bernilai $0.219 \pm 0.129 \times 10^5 \mu\text{m}^3$ dan $0.151 \pm 0.052 \times 10^5 \mu\text{m}^3$. Interaksi yang lemah juga diperhatikan diantara model bakteria dan permukaan bersalut cat komersial Or-Ag-Kao dengan daya pemisahan lateral 139.4004 nN (*B. niabensis*) dan 146.2251 nN (*A. litorea*). Pengurangan luas permukaan sentuhan yang besar juga diperhatikan di lapisan cat Or-Ag-Kao ($0.275 \mu\text{m}^2$ di *B. niabensis* dan $0.391 \mu\text{m}^2$ di *A. litorea*) menunjukkan bahawa lapisan cat Or-Ag-Kao berjaya meminimumkan lekatan bakteria permukaan. Sebagai kesimpulan, lapisan cat Or-Ag-Kao mempunyai spektrum keberkesanan yang luas dalam meminimumkan lekatan bakteria dan pembentukan biofilem mampu menyediakan penyelesaian yang mesra alam untuk biopenempelan semula jadi marin. Adalah dijangka bahawa, Or-Ag-Kao yang dibangunkan ini berpotensi tinggi untuk dikomersialkan sebagai lapisan anti-biopenempelan semula jadi marin alternatif menggantikan cat anti-biopenempelan semula jadi berasaskan tembaga yang ada sekarang.

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LIST OF ABBREVIATIONS

2-D	-	Two dimensional
3-D	-	Three Dimensional
AF	-	Anti-fouling
AFM	-	Atomic Force Microscope
Ag-Kao	-	Silver modified kaolinite
ANOVA	-	Analysis of variances
ATR-FTiR	-	Reflectance-Fourier Transform Infrared
BLAST	-	Basic Local Alignment Search Tool
Br	-	Bromide
Ca	-	Calcium
CA	-	Mean contact angle
CCD	-	Charge coupled device
CFD	-	Computational fluid dynamics
CFU	-	Colony forming units
Cl	-	Chloride
CLSM	-	Confocal Laser Scanning Microscope
CO	-	Carbon monoxide
CV	-	Crystal violet
CW	-	Calcofluor White M2R
CW	-	Calcofluor White M2R
DCA	-	Diiodomethane contact angle
DNA	-	Deoxyribonucleic acid
DVLO	-	Derjaguin, Verwey, Landau and Overbeek
E	-	East
e.g.	-	<i>exempli gratia</i> to mean “for example”
eDNA	-	Extracellular deoxyribonucleic acid
EDX	-	Energy-Dispersive X-ray
EPS	-	Extracellular polymeric substances
Eq.	-	Equation
FIB	-	Focused ion beam

GCA	-	Glycerol contact angle
H ₂ O	-	Chemical formula for water
HDTMA	-	Hexadecyltrimethylammonium
i.e.	-	<i>id est</i> to mean “that is” or "in other words."
IMO	-	International Maritime Organization
Kao	-	Kaolinite
KBr	-	Potassium Bromide
LAS X	-	Leica Application Suite X
MA	-	Marine Agar
Max	-	Maximum
MB	-	Marine broth
Mg	-	Magnesium
N	-	North
Na ⁺	-	Sodium ions
NCBI	-	National Centre for Biotechnology Information
NIS	-	Indigenous species
OD	-	Optical density
Or-Ag-Kao	-	Organo-silver-kaolinite
Or-Kao	-	Organo-kaolinite
PBS	-	Phosphate-buffered saline
PI	-	Propidium iodide
PS	-	Paint surface
QAC	-	Quaternary ammonium compounds
q-PCR	-	Quantitative polymerase chain reaction
QSI	-	Quorum sensing inhibitors
rRNA	-	16S ribosomal DNA
Sb	-	Antimony
SD	-	Standard deviation of negative control
SE	-	Surface energy
SEM	-	Scanning Electron Microscope
Si	-	Silica
sp.	-	Species

TBT-SPC	-	Tributyltin self-polishing
Ti	-	Titanium
TPT	-	Tributyltin
UV	-	Ultraviolet
VCA	-	Video contact angle
vdW	-	van de Waals
WCA	-	Water contact angle
<i>A. litorea</i>	-	<i>Alteromonas litorea</i>
<i>B. niabensis</i>	-	<i>Bacillus niabensis</i>
<i>et al.</i>	-	<i>et alii</i> to mean “and others”
<i>etc.</i>	-	<i>et cetera</i> to mean “and other similar things”, or “and so forth”
<i>I. baltica</i>	-	<i>Idiomarine baltica</i>
<i>M. communis</i>	-	<i>Marinomonas communis</i>
<i>P. aeruginosa</i>	-	<i>Pseudomonas aeruginosa</i>
<i>S. lutimaris</i>	-	<i>Salinimonas lutimaris</i>
Ag ⁺	-	Silver ion
CO ₂	-	Carbon dioxide
Cu ²⁺	-	Copper ion
Fe ²⁺	-	Iron divalent ion
Zn ⁺	-	Zinc ion
(CH ₃) ₃ N(CH ₂) ₁₅ CH ₃ Br	-	Chemical formula for Hexadecyltrimethylammonium bromide
AgNO ₃	-	Silver nitrate
Al (OH) ₃	-	Aluminium hydroxide
Al ₂ O ₃	-	Chemical formula for Aluminium oxide
Al ₂ Si ₂ O ₅ (OH) ₄	-	Chemical formula for kaolinite
MgCl ₂	-	Chemical formula for Magnesium chloride
NMs	-	Nanomaterials
NO _x	-	Nitrogen oxides
OD _{avcNeg}	-	Mean Optical density of negative control
OD _c	-	Cut-off Optical density
OD _c	-	Cut-off OD

SiNO_3	-	Silicon Nitride
SiO_2	-	Chemical formula for Silicon dioxide
SO_x	-	Sulphur oxide

LIST OF SYMBOLS

\$	-	Dollar
%	-	Percentage
~	-	Approximatively equal to
±	-	Range of values that a measured quantity may have
°	-	Degree
°C	-	Degree Celsius
µL	-	Microliter
µm	-	Micrometre
µm	-	Micrometre
µm ²	-	Micrometre square
µm ³	-	Micrometre cube
CFU/mL	-	Colony forming unit per millilitre
cm	-	Centimetre
cm ⁻¹	-	One per centimetre
eV	-	Electronvolt
g	-	Acceleration of gravity
G	-	Gram
h	-	Hours
k	-	Cantilever spring constant
kb	-	Kilobyte
kHz	-	Kilohertz
L	-	Cantilever length
L	-	Litre
m	-	Meter
mg	-	Milligram
min	-	Minutes
mJ/m ²	-	Millijoule per metre square
mL	-	Millilitre
mm	-	Millimetre
mM	-	Millimolar

N/m	-	Newton-metre
Nm	-	Nanometre
nm/V	-	Nano meter per voltage
nN	-	Nano newton
pH	-	Potential of hydrogen
<i>Ra</i>	-	Arithmetic mean roughness
rpm	-	Revolutions per minute
<i>Rq</i>	-	Root-Mean-Square roughness
s	-	Seconds
S	-	Sensitivity of applied cantilever
S	-	Sensitivity of applied cantilever
V	-	Voltage
V _{deflection}	-	Additional compression of the cantilever during bacterial detachment process
V _{setpoint}	-	User defined cantilever compression
V _{total}	-	Total vertical deflection
w/v	-	Weight per volume
α	-	Alpha
γ	-	Gamma
θ	-	Angle
θ and \varnothing	-	Angles of cantilever tip bending due to counterforce from bacterial cell to tip during detachment
λ_{emit}	-	Emission wavelength
λ_{excite}	-	Excitation wavelength

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CHAPTER 1

INTRODUCTION

1.1 Research Background

The ocean is the largest part on earth that contains an enormous amount of bacteria and always exposed to a variety of stress. In adapting these environmental challenges, marine bacteria usually form multicellular aggregates, known as biofilm. It is commonly visualized as a slimy layer that contains bacterial consortia embedded in an extracellular matrix. Biofilm development is a complex dynamic process forms gradually over time starting from the formation of a conditioning layer followed by bacterial adhesion, bacterial growth and biofilm expansion (Armbruster and Parsek, 2018). Biofilm formation have numerous advantages for bacteria development but detrimental to human and many industrial processes (Muhammad *et al.*, 2020).

Biofilm formation on marine man-made artificial structures such as ship body, vessels, cooling water systems, sensors, oil rigs and pipelines are a common ecological phenomenon. Surface colonization and subsequent biofilm formation and development provide numerous advantages to microorganism survival, but they can also cause serious deleterious problems to marine industries. Biofilm formation on marine structures contribute to biofouling, a process of deposition of living and non-living matter on man-made artificial structures. Biofilm formation mediates the attachment of other undesirable of microscopic (algae) or macroscopic (barnacles, mussels) organisms on surfaces. Biofouling formation interfere with process performance (Bucs *et al.*, 2018), cause material damage by the microbial attack (Fitridge *et al.*, 2012; Turan *et al.*, 2016), decrease product quality and quantity (Skovhus *et al.*, 2017), contamination of antifouling chemicals (Wang *et al.*, 2017) and shortening the life-time of plant components due to extended cleaning (Abioye *et al.*, 2019).

Another biofouling-related issue of concern is the effect of the introduction of species into a new environment, non-indigenous species (NIS) where they are not naturally present (invasive species) by the long-range transport of water in ballast tanks (Ojaveer *et al.*, 2018). Fouling communities developed in the submerged vessel communities have higher potential to be spread beyond their native distribution (Katsanevakis *et al.*, 2013). These invasive species may cause unwanted consequences for the environment, economics, and human health (Fletcher *et al.*, 2013; Lezzi *et al.*, 2018; Bonanno and Orlando-Bonaca, 2019).

Many conventional approaches such as mechanical cleaning, ultraviolet radiation and traditional chemical biocides have been utilized to counter biofilm effects in marine environment, but they are less effective. The use of antifouling paints with organometallic compounds or synthetic biocides might be the best way to combat biofilm and prevent biofouling (Salta *et al.*, 2013). For many years, tributyltin (TBT) was widely used as the active components in antifouling (Eklund and Watermann, 2018). TBT which is the most effective antifoulant has been used as a paint additive since the 1960s (Chambers *et al.*, 2006; Korkut and Atlar, 2012). However, due to its toxic effect on the wider marine environment, the application of TBT coatings was prohibited by Marine Environment Protection Committee (MEPC) of the International Maritime Organization (IMO) in September 2008 (Wang *et al.*, 2017).

After the ban on the use of TBT coating, copper has been increasingly used as the main biocide ingredient in antifouling paint coatings to tackle marine biofouling. Although copper is less toxic than TBT compounds, the extensive use of copper-based antifouling paints has led to the accumulation of copper and its compounds in the marine environment particularly beginning to pose a serious environmental problem (Srinivasan and Swain, 2007; Neira *et al.*, 2011; Siddiqui *et al.*, 2015). Previous studies shown that massive accumulation of copper ions concentrations toxic to aquatic organisms (Matthiessen *et al.*, 1999; Katranitsas *et al.*, 2003; Bao *et al.*, 2010). Studies have shown that dissolved copper may affect not only target species but also give negative effects on other marine species by disrupting enzymatic activity and increasing mortality (Katranitsas *et al.*, 2003; Bao *et al.*, 2010; Lindgren *et al.*, 2018).

The widespread environmental concerns of copper containing biocides had leading to the implementation of new, very strict regulations on limitation in application and residue levels of copper-containing biocides for boat, yacht and ship hulls. The new regulation on copper-based antifouling paints application was decided after considering serious consequences of biofouling and the benefits of effective copper-based antifouling paints that use safe and approved copper concentrations. Even though countries like Japan, Denmark and Sweden had prohibited the use of copper in anti-fouling systems (Dekinesh, 2018), many counties in the world including European Union, France, New Zealand, Canada, and United State still approved the use of copper-based antifouling coatings in their waters with requirement of a complete scientific risk assessments and additional safety measurements. The scientific risk assessments are including the eco-toxicology tests and environment impacts tests (Source:<http://echa.europa.eu/information-on-chemicals/biocidal-active-substances>) while the additional safety measurements are including safe operational procedures for the application, removal, enhanced labelling, and maximum residue levels in products entering the food chain (McNeil, 2018).

Due to the restrictions on toxic biocides active ingredients usage in antifouling paints and increasing consumer environmental concern, the study of marine biofilm and the strategies to combat biofouling is currently one of the most important fields of research. Current research has developed more approaches as potential environmentally-friendly alternatives to solve biofilm and biofouling problems on man-made artificial marine structures (Li and Ning, 2019). Novel approaches such as quorum sensing inhibitors (Song *et al.*, 2020; Muras *et al.*, 2021), antimicrobial peptides (Doiron *et al.*, 2018); Saha *et al.*, 2020) , enzymes (Aykin *et al.*, 2019; Mehrabi *et al.*, 2020), non-biocides compounds (Ventura *et al.*, 2017; Almeida *et al.*, 2018), bio-surfactants (Kim *et al.*, 2015; Vladkova, 2018), microorganism metabolites (Wang *et al.*, 2017; Li and Ning, 2019), nanomaterials (NMs) (Gutner-Hoch *et al.*, 2018; Figueiredo *et al.*, 2020) and biomimetic surfaces (Selim *et al.*, 2020; He *et al.*, 2020) had studied to solve biofilm formation on marine man-made artificial structures. However, many of these approaches still have limitations in application and efficiency (Tian *et al.*, 2021).

Therefore, this research aims to offer a promising solution to the biofouling problem in its initial stage of bacterial adhesion and biofilm formation on surfaces. A considerable focus of this research lies in the development of anti-biofouling paint additive with an emphasis on active compounds, silver ion (Ag^+) and hexadecyltrimethylammonium (HDTMA) embedded in kaolinite clay structure. The aim is to transfer the developed anti-biofouling paint coating on man-made artificial structures in marine environment. Immobilization of cationic surfactants from the quaternary ammonium compounds (QAC) such as HDTMA in silver-kaolinite, termed as organo-silver-kaolinite clay (Or-Ag-Kao) may not only improve the antibacterial activity of the unmodified clay (Malachova *et al.*, 2011; Salim and Malek, 2017; Nur Aryantie *et al.*, 2019) but it also improve the absorption properties of the clay (Bagherifam *et al.*, 2014; Malik *et al.*, 2018). Kaolinite good controlled-release delivery system that could minimize cation residues in the environment and maintain toxic concentration of the loaded cation in close approximate to target microorganisms (Holešová *et al.*, 2013; (Abukhadra and Allah, 2019). The loading of Ag^+ and HDTMA onto the carrier system such as kaolinite may prevent the excessive release hence suppress the toxic side effects toward the marine ecology system as they are released into the environment at a controllable rate. Immobilization of HDTMA in silver modified kaolinite is also an alternative to prevent the formation of insoluble precipitates, AgCl when the Ag^+ is released into the seawater comes into contact with chloride anions (Karel *et al.*, 2015). Studies have shown that QACs such as HDTMA may act as the counter ions due to the great affinity towards chloride ions (Malek *et al.*, 2013).

1.2 Problem Statement

Biofouling in maritime causes environmental disasters and billions of dollars are being consumed annually in the shipping industry. Drag and surface smoothness decreases when fouling organisms attach to the hulls of the vessel resulting an increase in the hydrodynamic weight and subsequently lead to a reduction in top speed and loss of maneuverability (Lindholdt *et al.*, 2015). The effect of higher frictional resistance also increased fuel consumption (Cao *et al.*, 2011), increased bunker costs at a constant

speed of the vessel or reduced speed at a constant fuel consumption which later lead to additional expenses due to time delays or lost earnings (Dafforn *et al.*, 2011; Selim *et al.*, 2017). Moreover, biofouling can also cause an increase of the frequency of dry-docking operations either due to the need of additional hull cleaning or due to costly additional hull repair or coating replacement (Desher, 2018). Regarding to environmental impact, biofouling initiate transfer of invasive species into a new environment and has led to a new danger for the different oceans. Transported invasive species would potentially damage the conserved biodiversity to the particular new environment (Shevalkar *et al.*, 2020). Thus, efforts of managing biofouling are beneficial to both economically and to the environment.

The most effective and easy way to overcome biofouling problem is application of antifouling coatings containing varying concentrations and mixtures of biocides (Miller *et al.*, 2020). Presently, the most commonly used biocide in antifouling paints is cooper oxide (Gu *et al.*, 2020). Though copper oxide is less toxic than tin, its effect on marine ecology is gradually increasing as it continues to accumulate (Siddiqui *et al.*, 2015). This condition has caused much concern from regulators and researchers. The massive accumulation of copper ions concentrations in marine environment would significantly link to serious environmental hazard (Corcoll *et al.*, 2019). Thus, the search for efficient and environmentally friendly anti-biofouling coating has become an even more significant issue (Selim *et al.*, 2015).

Therefore, this research focused on investigating the anti-bacterial adhesion and anti-biofilm properties of the modified kaolinite, Or-Ag-Kao against isolated marine biofilm-forming bacteria. The finding would reveal the modified kaolinite potential as a new highly effective, broad-spectrum and environmental-friendly paint additive that can be applied in marine artificial structures. The working hypothesis was that immobilization of active compounds, silver ions and HDTMA in kaolinite structures would enhance the anti-bacterial adhesion and anti-biofilm properties of the paint and concurrently reduced the environmental hazard due to silver accumulation in marine environment. To date, no research has been done on producing new paint formulations containing Or-Ag-Kao to reduce or inhibit the biofilm formation on surfaces submerged in the seawater.

1.3 Objectives of the Study

In this study, the kaolinite sample's ability in reducing or minimizing biofilm formation were evaluated qualitatively and quantitatively. Hence, the following objectives were put forward:

- (a) To characterize the prepared modified kaolinite samples and compared with the commercial paint.
- (b) To select a paint coating technique that can minimize bacteria attachment on paint surfaces.
- (c) To isolate, characterize and identify the attached marine bacterial on painted surfaces exposed to natural seawater under hydrodynamic condition.
- (d) To evaluate the anti-bacterial adhesion and anti-biofilm efficiency of the modified kaolinite sample (with/without paint) against Gram-positive and Gram-negative marine biofilm-forming bacteria.
- (e) To investigate the lateral detachment force between the individual Gram-positive and Gram-negative marine biofilm-forming bacteria from modified kaolinite + commercial paint surfaces.

1.4 Scopes of the Study

The research was initiated by characterizing the prepared kaolinite samples: Kao (raw kaolinite), Ag-Kao (silver-kaolinite), HDTMA-Kao/Or-Kao (surfactant modified kaolinite), Or-Ag-Kao (surfactant-silver-kaolinite) and commercial paint using Attenuated Total Reflectance-Fourier Transform Infrared (ATR-FTiR) Spectroscopy and Energy-Dispersive X-ray (EDX) Spectroscopy. Characterization of the prepared kaolinite samples and commercial paint would provide information about the chemical elements present in the samples. Determinations of the sample's chemical elements constitution would be beneficial in generating correlation of the present elements and the anti-bacterial adhesion and anti-biofilm effects.

Subsequently, four different paint coating methods: brush, roller, dipping and air spray were evaluated to determine the most unfavourable surface condition that can reduce marine bacteria formation. The paint surface conditions were evaluated based on the paint surface physical properties and its microbial growth properties. The physical properties of the paint surfaces were analysed using Atomic Force Microscope (AFM), Eclipse Ni-U Upright Microscope and VCA Optima instrument. Meanwhile, the microbial growth properties of the surfaces were evaluated from the Colony Forming Unit (CFUs) counting. This was followed by the selection of coating technique that was applied to deposit the kaolinite samples incorporated into commercial paint. The selected coating technique would provide an inconducive environment for bacterial biofilm development which will influence the selection of modified kaolinite sample to minimize the bacterial adhesion.

Biofilm formation was induced when bacterial cells attach on a surface. The type of bacterial cells that attached on a surface depend on the surface structure. Therefore, the marine bacteria were isolated from exposed surfaces in natural filtered seawater at a standard temperature and flow rate for a setting period. The isolated bacteria were subjected to series of biochemical test and biofilm assay before identified using molecular identification (16S rRNA sequencing). Next, the isolated bacteria EPS components were determined by using Attenuated Total Reflectance-Fourier Transform Infrared (ATR-FTiR) Spectroscopy. Later, the EPS productivity of the isolated biofilm-forming marine bacteria were determined and compared using Jiao *et al.* (2010) method.

Anti-bacterial bioassay was conducted to determine the effectiveness of the anti-biofilm paint derived from modified kaolinite in inhibiting or reducing the biofilm development of the isolated biofilm-forming marine bacteria. From the anti-bacterial bioassay, two species of the bacteria with superior biofilm formation capability were selected to represent the Gram-negative and Gram-positive group of bacteria. Confocal Laser Scanning Microscope (CLSM) analysis was performed to enumerate the bacterial cells on the paint surface. CLSM analysis would provide clearer and specific information on bacterial attachment and biofilm formation as CLSM can observed living cells, dead cells and EPS.

The investigation on the strength of the bacterial attachment on paint surfaces were evaluated via interaction forces of cell-substrate surface interaction using AFM. For this purpose, lateral detachment force method in contact mode imaging was used to quantify the strength. Both qualitative and quantitative information collected from single bacterial cell interaction forces will determine which modified kaolinite sample was most effective in preventing biofilm formation.

1.5 Significance of the Study

The incorporation of silver ions and HDTMA in kaolinite structure would provide a promising solution to develop a new generation of efficient anti-biofouling additive for maritime coatings with minimal environmental hazard compared to currently available metal-based coatings. Kaolinite controlled-release delivery system would reduce the release rates of silver from antifouling paints in marine waters hence, the load of copper to the environment could be substantially reduced. The new coating is useful to extend the life of marine structures and reduce the maintenance cost. The outcome of this study will be a steppingstone in producing a broad-spectrum effective environmental-friendly anti-biofouling paint which can be used primarily in marine industry.

REFERENCES

- Abad-Álvaro, I., Trujillo, C., Bolea, E., Laborda, F., Fondevila, M., Latorre, M. A. and Castillo, J. R. (2019) ‘Silver nanoparticles-clays nanocomposites as feed additives: Characterization of silver species released during in vitro digestions. Effects on silver retention in pigs’, *Microchemical Journal*. Elsevier, 149(July), p. 104040.
- Abbott, A., Abel, P. D., Arnold, D. W. and Milne, A. (2000) ‘Cost–benefit analysis of the use of TBT: the case for a treatment approach’, *Science of The Total Environment*, 258(1–2), pp. 5–19.
- Abioye, O. P., Loto, C. A. and Fayomi, O. S. I. (2019) ‘Evaluation of Anti-biofouling Progresses in Marine Application’, *Journal of Bio- and Triboro-Corrosion*. Springer International Publishing, 5(1), p. 0.
- Abukhadra, M. R. and Allah, A. F. (2019) ‘Synthesis and characterization of kaolinite nanotubes (KNTs) as a novel carrier for 5-fluorouracil of high encapsulation properties and controlled release’, *Inorganic Chemistry Communications*. Elsevier, 103(February), pp. 30–36.
- Achinas, S., Charalampogiannis, N. and Euverink, G. J. W. (2019) ‘Brief Recap for Bacteria Adhesion’, *Applied Sciences*, 9, pp. 1–15.
- Agarwal, R. K., Singh, S., Bhilegaonkar, K. N. and Singh, V. P. (2011) ‘Optimization of microtitre plate assay for the testing of biofilm formation ability in different *Salmonella* serotypes’, *International Food Research Journal*, 18(4), pp. 1493–1498.
- Agogué, H., Joux, F., Obernosterer, I. and Lebaron, P. (2005) ‘Resistance of marine bacterioneuston to solar radiation’, *Applied and Environmental Microbiology*, 71(9), pp. 5282–5289.
- Agostini, V. O., Macedo, A. J., Muxagata, E., da Silva, M. V. and Pinho, G. L. L. (2019) ‘Natural and non-toxic products from Fabaceae Brazilian plants as a replacement for traditional antifouling biocides: an inhibition potential against initial biofouling’, *Environmental Science and Pollution Research*. Environmental Science and Pollution Research, 26(26), pp. 27112–27127.

- Ahmed, N. M. and El-sabbagh, S. H. (2015) ‘The Influence of Doped-Kaolin on the Properties of Styrene-Butadiene Rubber Composites’, *International Journal of Advanced Research*, 3(5), pp. 1–19.
- Akhigbe, L., Ouki, S. and Saroj, D. (2015) ‘Removal of Escherichia coli and heavy metals from aqueous solutions using silver-modified clinoptilolite’, *Desalination and Water Treatment*, 55(3), pp. 777–782.
- Al-Juboori, R. A. and Yusaf, T. (2012) ‘Biofouling in RO system: Mechanisms, monitoring and controlling’, *Desalination*. Elsevier B.V., 302, pp. 1–23.
- Almeida, J. R., Moreira, J., Pereira, D., Pereira, S., Antunes, J., Palmeira, A., Vasconcelos, V., Pinto, M., Correia-da-Silva, M. and Cidade, H. (2018) ‘Potential of synthetic chalcone derivatives to prevent marine biofouling’, *Science of the Total Environment*. Elsevier B.V., 643, pp. 98–106.
- Alvarez-Ordóñez, A., Mouwen, D. J. M., López, M. and Prieto, M. (2011) ‘Fourier transform infrared spectroscopy as a tool to characterize molecular composition and stress response in foodborne pathogenic bacteria’, *Journal of Microbiological Methods*. Elsevier B.V., 84(3), pp. 369–378.
- Alves, M. M., Mota Vieira, J. A., Alvares Pereira, M., Pereira, A. and Mota, M. (2001) ‘Effect of Lipids and Oleic Acid on Biomass Development in Anaerobic Fixed-Bed Reactors. Part I: Biofilm Growth and Activity’, *Water Research*, 35(1), pp. 255–263.
- Amadio, J., Casey, E. and Murphy, C. D. (2013) ‘Filamentous fungal biofilm for production of human drug metabolites’, *Applied Microbiology and Biotechnology*, 97(13), pp. 5955–5963.
- Ameh, T. and Sayes, C. M. (2019) ‘The potential exposure and hazards of copper nanoparticles: A review’, *Environmental Toxicology and Pharmacology*. Elsevier, 71, p. 103220.
- Anson, A., Fisher, P. J., Kennedy, A. F. D. and Sutherland, I. W. (1987) ‘A bacterium yielding a polysaccharide with unusual properties’, *Journal of Applied Bacteriology*, 62(2), pp. 147–150.
- Antunes, J., Leão, P. and Vasconcelos, V. (2019) ‘Marine biofilms: diversity of communities and of chemical cues’, *Environmental Microbiology Reports*, 11(3), pp. 287–305.

- Antunes, J. T., Sousa, A. G. G., Azevedo, J., Rego, A., Leão, P. N. and Vasconcelos, V. (2020) ‘Distinct Temporal Succession of Bacterial Communities in Early Marine Biofilms in a Portuguese Atlantic Port’, *Frontiers in Microbiology*, 11(August), pp. 1–17.
- Anwar, M. A. and Choi, S. (2014) ‘Gram-negative marine bacteria: Structural features of lipopolysaccharides and their relevance for economically important diseases’, *Marine Drugs*, 12(5), pp. 2485–2514.
- Arahal, D. R., Lucena, T., Macián, M. C., Ruvira, M. A., González, J. M., Lekumberri, I., Pinhassi, J. and Pujalte, M. J. (2016) ‘*Marinomonas blandensis* sp. Nov., a novel marine gammaproteobacterium’, *International Journal of Systematic and Evolutionary Microbiology*, 66(12), pp. 5544–5549.
- Armbruster, C. R. and Parsek, M. R. (2018) ‘New insight into the early stages of biofilm formation’, *Proceedings of the National Academy of Sciences of the United States of America*, 115(17), pp. 4317–4319.
- Armstrong, E., Boyd, K. G. and Burgess, J. G. (2000) ‘Prevention of marine biofouling using natural compounds from marine organisms. *Biotechnology Annual Review*, 6, pp. 221–241.
- Asséré, A., Oulahal, N. and Carpentier, B. (2008) ‘Comparative evaluation of methods for counting surviving biofilm cells adhering to a polyvinyl chloride surface exposed to chlorine or drying’, *Journal of Applied Microbiology*, 104(6), pp. 1692–1702.
- Awad, M. E., López-Galindo, A., Setti, M., El-Rahmany, M. M. and Iborra, C. V. (2017) ‘Kaolinite in pharmaceutics and biomedicine’, *International Journal of Pharmaceutics*. Elsevier, 533(1), pp. 34–48.
- Aykin, E., Omuzbuken, B. and Kacar, A. (2019) ‘Microfouling bacteria and the use of enzymes in eco-friendly antifouling technology’, *Journal of Coatings Technology and Research*. Springer US, 16(3), pp. 847–856.
- Azeredo, J., Azevedo, N. F., Briandet, R., Cerca, N., Costa, A. R., Desvaux, M., Bonaventura, G. Di, Jaglic, Z., Kačániová, M., Knøchel, S., Mergulhão, F., Meyer, R. L., Nychas, G., Tresse, O., Sternberg, C., Azeredo, J., Azevedo, N. F., Briandet, R., Cerca, N., Costa, A. R., Desvaux, M., Bonaventura, G. Di, Hébraud, M., Kačániová, M., Knøchel, S., Lourenço, A., Mergulhão, F., Meyer, L., Nychas, G., Simões, M., Tresse, O., Sternberg, C., Rita, A., Di, G., Michel, H., Meyer, R. L., Nychas, G., Sim, M. and Tresse, O. (2017)

- ‘Critical Reviews in Microbiology Critical review on biofilm methods a’, 7828(July).
- Badel, S., Laroche, C., Gardarin, C., Bernardi, T. and Michaud, P. (2008) ‘New method showing the influence of matrix components in *leuconostoc mesenteroides* biofilm formation’, *Applied Biochemistry and Biotechnology*, 151(2–3), pp. 364–370.
- Bagherifam, S., Komarneni, S., Lakzian, A., Fotovat, A., Khorasani, R., Huang, W., Ma, J., Hong, S., Cannon, F. S. and Wang, Y. (2014) ‘Highly selective removal of nitrate and perchlorate by organoclay’, *Applied Clay Science*. Elsevier B.V., 95, pp. 126–132.
- Bailey, S. A., Brown, L., Campbell, M. L., Canning-Clode, J., Carlton, J. T., Castro, N., Chainho, P., Chan, F. T., Creed, J. C., Curd, A., Darling, J., Fofonoff, P., Galil, B. S., Hewitt, C. L., Inglis, G. J., Keith, I., Mandrak, N. E., Marchini, A., McKenzie, C. H., Occhipinti-Ambrogi, A., Ojaveer, H., Pires-Teixeira, L. M., Robinson, T. B., Ruiz, G. M., Seaward, K., Schwindt, E., Son, M. O., Therriault, T. W. and Zhan, A. (2020) ‘Trends in the detection of aquatic non-indigenous species across global marine, estuarine and freshwater ecosystems: A 50-year perspective’, *Diversity and Distributions*, 26(12), pp. 1780–1797.
- Bao, W. Y., Lee, O. O., Chung, H. C., Li, M. and Qian, P. Y. (2010) ‘Copper affects biofilm inductiveness to larval settlement of the serpulid polychaete *hydroides elegans* (Haswell)’, *Biofouling*, 26(1), pp. 119–128.
- Barberán, A., Casamayor, E. O. and Fierer, N. (2014) ‘The microbial contribution to macroecology’, *Frontiers in Microbiology*, 5(May), pp. 1–8.
- Bartholomew, J. W. and Finkelstein, H. (1958) ‘Relationship of Cell Wall Staining To Gram Differentiation1’, *Journal of Bacteriology*, 75(1), pp. 77–84.
- Baumann, L., Baumann, P., Mandel, M. and Allen, R. D. (1972) ‘Taxonomy of aerobic marine eubacteria.’, *Journal of Bacteriology*, 110(1), pp. 402–429.
- Bazylinski, D. A., Li, M., Gong, J., Qing, Y., Zou, S., Fu, R., Su, L., Zhang, X. and Zhang, Q. (2016) ‘Protist-Bacteria Associations: Gammaproteobacteria and Alphaproteobacteria Are Prevalent as Digestion-Resistant Bacteria in Ciliated Protozoa’, *Front Microbiol*, 11(7), 498.
- Bellou, N., Papathanassiou, E., Dobretsov, S., Lykousis, V. and Colijn, F. (2012) ‘The effect of substratum type, orientation and depth on the development of

- bacterial deep-sea biofilm communities grown on artificial substrata deployed in the Eastern Mediterranean', *Biofouling*, 28(2), pp. 199–213.
- Benard, L. D., Tuah, P. M., Suadin, E. G. and Jamian, N. (2015) 'Isolation and characterization of surface and subsurface bacteria in seawater of Mantanani Island, Kota Belud, Sabah by direct and enrichment techniques', *IOP Conference Series: Materials Science and Engineering*, 78(1).
- Bendersky, M. and Davis, J. M. (2011) 'DLVO interaction of colloidal particles with topographically and chemically heterogeneous surfaces', *Journal of Colloid and Interface Science*. Elsevier Inc., 353(1), pp. 87–97.
- Bertani, B. and Ruiz, N. (2018) 'Function and Biogenesis of Lipopolysaccharides', *EcoSal Plus*, 8(1), pp. 1–33.
- Bharti, A., Velmourougane, K. and Prasanna, R. (2017) 'Phototrophic biofilms: diversity, ecology and applications', *Journal of Applied Phycology*. 29(6), pp. 2729–2744.
- Bhattacharyya, K. G. and Gupta, S. Sen (2006) 'Kaolinite, montmorillonite, and their modified derivatives as adsorbents for removal of Cu(II) from aqueous solution', *Separation and Purification Technology*, 50(3), pp. 388–397.
- Blackwood, D. J., Lim, C. S., Teo, S. L. M., Hu, X. and Pang, J. (2017) 'Macrofouling induced localized corrosion of stainless steel in Singapore seawater', *Corrosion Science*, 129(October), pp. 152–160.
- Bogachev, M. I., Volkov, V. Y., Markelov, O. A., Trizna, E. Y., Baydamshina, D. R., Melnikov, V., Murtazina, R. R., Zelenikhin, P. V., Sharafutdinov, I. S. and Kayumov, A. R. (2018) 'Fast and simple tool for the quantification of biofilm-embedded cells sub-populations from fluorescent microscopic images', *PLoS ONE*, 13(5), pp. 1–24.
- Boks, N. P., Norde, W., Mei, H. C. Van Der and Busscher, H. J. (2015) 'Forces involved in bacterial adhesion to hydrophilic and hydrophobic surfaces', (2008), pp. 3122–3133.
- Bonanno, G. and Orlando-Bonaca, M. (2019) 'Non-indigenous marine species in the Mediterranean Sea—Myth and reality', *Environmental Science and Policy*. Elsevier, 96(January), pp. 123–131.
- Bos, R., Van Der Mei, H. C. and Busscher, H. J. (1999) 'Physico-chemistry of initial microbial adhesive interactions - Its mechanisms and methods for study', *FEMS Microbiology Reviews*, 23(2), pp. 179–230.

- Botelho, P. S., Maciel, M. I. S., Bueno, L. A., Marques, M. D. F. F., Marques, D. N. and Sarmento Silva, T. M. (2014) ‘Characterisation of a new exopolysaccharide obtained from of fermented kefir grains in soymilk’, *Carbohydrate Polymers*. Elsevier Ltd., 107(1), pp. 1–6.
- Boyer, D., Mather, W., Mondragón-Palomino, O., Orozco-Fuentes, S., Danino, T., Hasty, J. and Tsimring, L. S. (2011) ‘Buckling instability in ordered bacterial colonies’, *Physical Biology*, 8(2).
- Bramhachari, P. V., Kavi Kishor, P. B., Ramadevi, R., Kumar, R., Rama Rao, B. and Dubey, S. T. (2007) ‘B . Rama Rao , and Santosh Kumar Dubey’, *Microbial biotechnology*, 17(1), pp. 44–51.
- Bressy, C. and Lejars, M. (2014) ‘Marine fouling : An overview marine fouling’, *Journal of Ocean Technology*, 9(4), pp. 19–28.
- Briand, J.-F., Barani, A., Garnier, C., Réhel, K., Urvois, F., LePoupon, C., Bouchez, A., Debroas, D. and Bressy, C. (2017) ‘Spatio-Temporal Variations of Marine Biofilm Communities Colonizing Artificial Substrata Including Antifouling Coatings in Contrasted French Coastal Environments’, *Microbial Ecology*. Springer US, 74(3), pp. 585–598.
- Brown, R., Russell, S., May, S., Regan, F. and Chapman, J. (2017) ‘Reproducible Superhydrophobic PVC Coatings; Investigating the Use of Plasticizers for Early Stage Biofouling Control’, *Advanced Engineering Materials*, 19(7), pp. 1–6.
- Brown, S., Santa Maria, J. P. and Walker, S. (2013) ‘Wall teichoic acids of gram-positive bacteria’, *Annual Review of Microbiology*, 67, pp. 313–336.
- Brück, H. L., Coutte, F., Dhulster, P., Gofflot, S., Jacques, P. and Delvigne, F. (2020) ‘Growth dynamics of bacterial populations in a two-compartment biofilm bioreactor designed for continuous surfactin biosynthesis’, *Microorganisms*, 8(5).
- Bryant, J. A., Aylward, F. O., Eppley, J. M., Karl, D. M., Church, M. J. and DeLong, E. F. (2016) ‘Wind and sunlight shape microbial diversity in surface waters of the North Pacific Subtropical Gyre’, *ISME Journal*. Nature Publishing Group, 10(6), pp. 1308–1322.
- Buchan, A., Shao, Z., Alexander Lever, M., Zürich, E., Engelen, B., Pohlner, M., Dlugosch, L., Wemheuer, B., Mills, H. and Kiel Reese, B. (2019) ‘The Majority of Active Rhodobacteraceae in Marine Sediments Belong to

- Uncultured Genera: A Molecular Approach to Link Their Distribution to Environmental Conditions'. *Front Microbiol*, 2(10), pp. 659.
- Bucs, S., Farhat, N., Kruithof, J. C., Picioreanu, C., van Loosdrecht, M. C. M. and Vrouwenvelder, J. S. (2018) 'Review on strategies for biofouling mitigation in spiral wound membrane systems', *Desalination*. Elsevier, 434(September 2017), pp. 189–197.
- Busseron, E., Ruff, Y., Moulin, E. and Giuseppone, N. (2013) 'Supramolecular self-assemblies as functional nanomaterials', *Nanoscale*, 5(16), pp. 7098–7140.
- Cao, S., Wang, J. D., Chen, H. S. and Chen, D. R. (2011) 'Progress of marine biofouling and antifouling technologies', *Chinese Science Bulletin*, 56(7), pp. 598–612.
- Capão, A., Moreira-Filho, P., Garcia, M., Bitati, S. and Procópio, L. (2020) 'Marine bacterial community analysis on 316L stainless steel coupons by Illumina MiSeq sequencing', *Biotechnology Letters*, 42(8), pp. 1431–1448.
- Carniello, V., Peterson, B. W., van der Mei, H. C. and Busscher, H. J. (2018) 'Physico-chemistry from initial bacterial adhesion to surface-programmed biofilm growth', *Advances in Colloid and Interface Science*. Elsevier B.V., pp. 1–14.
- Carpenter, T. S., Parkin, J. and Khalid, S. (2016) 'The Free Energy of Small Solute Permeation through the *Escherichia coli* Outer Membrane Has a Distinctly Asymmetric Profile', *The Journal of Physical Chemistry Letters*, 7(17), pp. 3446–3451.
- Caruso, G. (2020) 'Microbial colonization in marine environments: Overview of current knowledge and emerging research topics', *Journal of Marine Science and Engineering*, 8(2), pp. 1–22.
- de Carvalho, C. C. C. R. (2018) 'Marine biofilms: A successful microbial strategy with economic implications', *Frontiers in Marine Science*, 5(APR), pp. 1–11.
- De Carvalho, C. C. C. R. and Caramujo, M. J. (2012) 'Lipids of prokaryotic origin at the base of marine food webs', *Marine Drugs*, 10(12), pp. 2698–2714.
- Cassé, F. and Swain, G. W. (2006) 'The development of microfouling on four commercial antifouling coatings under static and dynamic immersion', *International Biodeterioration and Biodegradation*, 57(3), pp. 179–185.
- Celikkol-Aydin, S., Gaylarde, C. C., Lee, T., Melchers, R. E., Witt, D. L. and Beech, I. B. (2016) '16S rRNA gene profiling of planktonic and biofilm microbial populations in the Gulf of Guinea using Illumina NGS', *Marine*

- Environmental Research*. Elsevier Ltd, 122, pp. 105–112.
- Chalkiadakis, E., Dufourcq, R., Schmitt, S., Brandily, C., Kervarec, N., Coatanea, D., Amir, H., Loubersac, L., Chanteau, S., Guezennec, J., Dupont-Rouzeyrol, M. and Simon-Colin, C. (2013) ‘Partial characterization of an exopolysaccharide secreted by a marine bacterium, *Vibrio neocaldonicus* sp. nov., from new caledonia’, *Journal of Applied Microbiology*, 114(6), pp. 1702–1712.
- Chambers, L. D., Stokes, K. R., Walsh, F. C. and Wood, R. J. K. (2006) ‘Modern approaches to marine antifouling coatings’, *Surface and Coatings Technology*. Elsevier B.V., 201(6), pp. 3642–3652.
- Chang, W. S. and Halverson, L. J. (2003) ‘Reduced water availability influences the dynamics, development, and ultrastructural properties of *Pseudomonas putida* biofilms’, *Journal of Bacteriology*, 185(20), pp. 6199–6204.
- Chao, H. P. and Chen, S. H. (2012) ‘Adsorption characteristics of both cationic and oxyanionic metal ions on hexadecyltrimethylammonium bromide-modified NaY zeolite’, *Chemical Engineering Journal*. Elsevier B.V., 193–194, pp. 283–289.
- Chaumeil, F. and Crapper, M. (2013) ‘DEM simulations of initial deposition of colloidal particles around non-woven membrane spacers’, *Journal of Membrane Science*. Elsevier, 442, pp. 254–263.
- Chechetko, E. S., Tolpeshta, I. I. and Zavgorodnyaya, Y. A. (2017) ‘Application of dodecyltrimethylammonium-modified bentonite for water purification from oil and water-soluble oil components’, *Moscow University Soil Science Bulletin*, 72(3), pp. 119–124.
- Chen, D., Zhao, T. and Doyle, M. P. (2015) ‘Single- and mixed-species biofilm formation by *Escherichia coli* O157: H7 and *Salmonella*, and their sensitivity to levulinic acid plus sodium dodecyl sulfate’, *Food Control*. Elsevier Ltd, 57, pp. 48–53.
- Chen, X., Suwarno, S. R., Chong, T. H., McDougald, D., Kjelleberg, S., Cohen, Y., Fane, A. G. and Rice, S. A. (2013) ‘Dynamics of biofilm formation under different nutrient levels and the effect on biofouling of a reverse osmosis membrane system’, *Biofouling*, 29(3), pp. 319–330.
- Cheng, H., Zhou, Y. and Liu, Q. (2019) *Kaolinite nanomaterials: Preparation, properties and functional applications, Nanomaterials from Clay Minerals: A New Approach to Green Functional Materials*. Elsevier Inc.

- Cherifi, T., Jacques, M., Quessy, S. and Fravallo, P. (2017) ‘Impact of Nutrient Restriction on the Structure of *Listeria monocytogenes* Biofilm Grown in a Microfluidic System’, *Frontiers in Microbiology*, 8(MAY), pp. 1–13.
- Chernousova, S. and Epple, M. (2013) ‘Silver as antibacterial agent: Ion, nanoparticle, and metal’, *Angewandte Chemie - International Edition*, 52(6), pp. 1636–1653.
- Chiu, J. M. Y., Zhang, R., Wang, H., Thiagarajan, V. and Qian, P. Y. (2008) ‘Nutrient effects on intertidal community: From bacteria to invertebrates’, *Marine Ecology Progress Series*, 358(April), pp. 41–50.
- Cho, H. J., Jönsson, H., Campbell, K., Melke, P., Williams, J. W., Jedynak, B., Stevens, A. M., Groisman, A. and Levchenko, A. (2007) ‘Self-organization in high-density bacterial colonies: Efficient crowd control’, *PLoS Biology*, 5(11), pp. 2614–2623.
- Choi, S. H., Jang, Y. S., Jang, J. H., Bae, T. S., Lee, S. J. and Lee, M. H. (2019) ‘Enhanced antibacterial activity of titanium by surface modification with polydopamine and silver for dental implant application’, *Journal of Applied Biomaterials and Functional Materials*, 17(3).
- Chowdhury, S. R., Manna, S., Saha, P., Basak, R. K., Sen, R., Roy, D. and Adhikari, B. (2011) ‘Composition analysis and material characterization of an emulsifying extracellular polysaccharide (EPS) produced by *Bacillus megaterium* RB-05: A hydrodynamic sediment-attached isolate of freshwater origin’, *Journal of Applied Microbiology*, 111(6), pp. 1381–1393.
- Christensen, G. D., Simpson, W. A., Bisno, A. L. and Beachey, E. H. (1982) ‘Adherence of slime-producing strains of *Staphylococcus epidermidis* to smooth surfaces.’, *Infection and immunity*. American Society for Microbiology (ASM), 37(1), pp. 318–26.
- Christensen, G. D., Simpson, W. A., Younger, J. J., Baddour, L. M., Barrett, F. F., Melton, D. M. and Beachey, E. H. (1985) ‘Adherence of coagulase-negative staphylococci to plastic tissue culture plates: A quantitative model for the adherence of staphylococci to medical devices’, *Journal of Clinical Microbiology*, 22(6), pp. 996–1006.
- Chung, H. C., Lee, O. O., Huang, Y. L., Mok, S. Y., Kolter, R. and Qian, P. Y. (2010) ‘Bacterial community succession and chemical profiles of subtidal biofilms in relation to larval settlement of the polychaete *Hydroides elegans*’, *ISME*

- Journal*. Nature Publishing Group, 4(6), pp. 817–828.
- Conradi, M., Sever, T., Gregorčič, P. and Kocijan, A. (2019) ‘Short- and Long-Term Wettability Evolution and Corrosion Resistance of Uncoated and Polymer-Coated Laser-Textured Steel Surface’, *Coatings*, 9(9), p. 592.
- Cooksey, K. and Wigglesworth-Cooksey, B. (1995) ‘Adhesion of bacteria and diatoms to surfaces in the sea: a review’, *Aquatic Microbial Ecology*, 9, pp. 87–96.
- Copeland, M. F. and Weibel, D. B. (2009) ‘Bacterial swarming: A model system for studying dynamic self-assembly’, *Soft Matter*, 5(6), pp. 1174–1187.
- Corcoll, N., Yang, J., Backhaus, T., Zhang, X. and Eriksson, K. M. (2019) ‘Copper affects composition and functioning of microbial communities in marine biofilms at environmentally relevant concentrations’, *Frontiers in Microbiology*, 10(JAN), pp. 1–15.
- Cox, S. C., Jamshidi, P., Eisenstein, N. M., Webber, M. A., Burton, H., Moakes, R. J. A., Addison, O., Attallah, M., Shepherd, D. E. T. and Grover, L. M. (2017) ‘Surface Finish has a Critical Influence on Biofilm Formation and Mammalian Cell Attachment to Additively Manufactured Prosthetics’, *ACS Biomaterials Science and Engineering*, 3(8), pp. 1616–1626.
- Cravero, F., Gonzalez, I., Galan, E. and Dominguez, E. (1997) ‘Geology, mineralogy, origin and possible applications of some Argentinian kaolins in the Neuquen basin’, *Applied Clay Science*, 12(1–2), pp. 27–42.
- Cron, B., Macalady, J. L. and Cosmidis, J. (2021) ‘Organic stabilization of extracellular elemental sulfur in a Sulfurovum -rich biofilm : a new role for EPS ?’
- Czaczyk, K. and Myszka, K. (2007) ‘Biosynthesis of extracellular polymeric substances (EPS) and its role in microbial biofilm formation’, *Polish Journal of Environmental Studies*, 16(6), pp. 799–806.
- Dafforn, K. a., Lewis, J. a. and Johnston, E. L. (2011) ‘Antifouling strategies: History and regulation, ecological impacts and mitigation’, *Marine Pollution Bulletin*. Elsevier Ltd, 62(3), pp. 453–465.
- Dakal, T. C., Kumar, A., Majumdar, R. S. and Yadav, V. (2016) ‘Mechanistic basis of antimicrobial actions of silver nanoparticles’, *Frontiers in Microbiology*, 7(Nov), pp. 1–17.
- Dang, H., Li, T., Chen, M. and Huang, G. (2008) ‘Cross-Ocean Distribution of Rhodobacterales Bacteria as Primary Surface Colonizers in Temperate

- Coastal Marine Waters', *Applied and Environmental Microbiology*, 74(1), pp. 52–60.
- Dang, H. and Lovell, C. R. (2000) 'Bacterial primary colonization and early succession on surfaces in marine waters as determined by amplified rRNA gene restriction analysis and sequence analysis of 16S rRNA genes.', *Applied and environmental microbiology*. American Society for Microbiology (ASM), 66(2), pp. 467–75.
- Dang, H. and Lovell, C. R. (2016) 'Microbial Surface Colonization and Biofilm Development in Marine Environments', *Microbiology and Molecular Biology Reviews*, 80(1), pp. 91–138.
- Das, S. K., Ahmed, S., Ferdous, F., Farzana, F. D., Chisti, M. J., Latham, J. R., Talukder, K. A., Rahman, M., Begum, Y. A., Qadri, F., Golam Faruque, A. S. and Ahmed, T. (2013) 'Etiological diversity of diarrhoeal disease in Bangladesh', *Journal of Infection in Developing Countries*, 7(12), pp. 900–909.
- Das, S., Lyla, P. S. and Khan, S. A. (2006) 'Marine microbial diversity and ecology: Importance and future perspectives', *Current Science*, 90(10), pp. 1325–1335.
- Davarcioglu, B. (2010) 'Investigation of Central Anatolian region Nigde-Dikilitas (Turkey) clays by FTIR spectroscopy', *Epitoanyag - Journal of Silicate Based and Composite Materials*, 62(2), p. 55.
- Dave, S. R. (2016) 'Microbial Exopolysaccharide - An Inevitable Product for Living Beings and Environment', *Journal of Bacteriology & Mycology: Open Access*, 2(4), pp. 109–111.
- Davey, M. E. and O'toole, G. A. (2000) 'Microbial Biofilms: from Ecology to Molecular Genetics', *Microbiology and Molecular Biology Reviews*, 64(4), pp. 847–867.
- Deng, Y., White, G. N. and Dixon, J. B. (2002) 'Effect of structural stress on the intercalation rate of kaolinite', *Journal of Colloid and Interface Science*, 250(2), pp. 379–393.
- Desher, A. A. (2018) 'The Maritime Commons : Digital Repository of the World Maritime Biofouling impacts on the environment and ship energy efficiency'. Dissertations Thesis, World Maritime University.
- Desmond, P., Best, J. P., Morgenroth, E. and Derlon, N. (2018) 'Linking composition

- of extracellular polymeric substances (EPS) to the physical structure and hydraulic resistance of membrane biofilms', *Water Research*. Elsevier Ltd, 132, pp. 211–221.
- Detellier, C. (2018) 'Functional Kaolinite', *Chemical Record*, 18(7), pp. 868–877.
- Detty, M. R., Ciriminna, R., Bright, F. V. and Pagliaro, M. (2014) 'Environmentally benign sol-gel antifouling and foul-releasing coatings', *Accounts of Chemical Research*, 47(2), pp. 678–687.
- Ding, S., Zhang, L., Ren, X., Xu, B., Zhang, H. and Ma, F. (2012) 'The characteristics of mechanical grinding on kaolinite structure and thermal behavior', *Energy Procedia*, 16(PART B), pp. 1237–1240.
- Dobranksa, K., Nebesarova, J., Ruzicka, F., Dluhos, J. and Krzyzanek, V. (2013) 'Characterization of Yeast Biofilm by Cryo-SEM and FIB-SEM', *Microscopy and Microanalysis*, 19(S2), pp. 226–227.
- Dobretsov, S. and Rittschof, D. (2020) 'Love at first taste: Induction of larval settlement by marine microbes', *International Journal of Molecular Sciences*, 21(3).
- Dogan, M., Dogan, A. U., Aburub, A., Botha, A. and Wurster, D. E. (2012) 'Quantitative mineralogical properties (morphology-chemistry-structure) of pharmaceutical grade kaolinites and recommendations to regulatory agencies', *Microscopy and Microanalysis*, 18(1), pp. 143–151.
- Doghri, I., Rodrigues, S., Bazire, A., Dufour, A., Akbar, D., Sopena, V., Sablé, S. and Lanneluc, I. (2015) 'Marine bacteria from the French Atlantic coast displaying high forming-biofilm abilities and different biofilm 3D architectures', *BMC Microbiology*, 15(1), p. 231.
- Doiron, K., Beaulieu, L., St-Louis, R. and Lemarchand, K. (2018) 'Reduction of bacterial biofilm formation using marine natural antimicrobial peptides', *Colloids and Surfaces B: Biointerfaces*. Elsevier B.V., 167, pp. 524–530.
- Domeradzka-Gajda, K., Nocuń, M., Roszak, J., Janasik, B., Quarles, C. D., Wąsowicz, W., Grobelny, J., Tomaszewska, E., Celichowski, G., Ranoszek-Soliwoda, K., Cieślak, M., Puchowicz, D., Gonzalez, J. J., Russo, R. E. and Stępnik, M. (2017) 'A study on the in vitro percutaneous absorption of silver nanoparticles in combination with aluminum chloride, methyl paraben or di-n-butyl phthalate', *Toxicology Letters*, 272, pp. 38–48.
- Dong, B., Belkhair, S., Zaarour, M., Fisher, L., Verran, J., Tosheva, L., Retoux, R.,

- Gilson, J. P. and Mintova, S. (2014) ‘Silver confined within zeolite EMT nanoparticles: Preparation and antibacterial properties’, *Nanoscale*, Royal Society of Chemistry, 6(18), pp. 10859–10864.
- Donlan, R. M. (2002) ‘Biofilms: microbial life on surfaces’. *Emerging Infection*, 8(9), pp. 881–890.
- Donlan, R. M. and Costerton, J. W. (2002) ‘Biofilms: Survival mechanisms of clinically relevant microorganisms’, *Clinical Microbiology Reviews*, 15(2), pp. 167–193.
- Donnelly, B., Bedwell, I., Dimas, J., Scardino, A., Tang, Y. and Sammut, K. (2019) ‘Effects of various antifouling coatings and fouling on marine sonar performance’, *Polymers*, 11(4).
- Drago, L., Agrappi, S., Bortolin, M., Toscano, M., Romanò, C. L. and De Vecchi, E. (2016) ‘How to study biofilms after microbial colonization of materials used in orthopaedic implants’, *International Journal of Molecular Sciences*, 17(3).
- Du, H., Jiao, N., Hu, Y. and Zeng, Y. (2006) ‘Diversity and distribution of pigmented heterotrophic bacteria in marine environments’, *FEMS Microbiology Ecology*, 57(1), pp. 92–105.
- Durán, N., Durán, M., de Jesus, M. B., Seabra, A. B., Fávaro, W. J. and Nakazato, G. (2016) ‘Silver nanoparticles: A new view on mechanistic aspects on antimicrobial activity’, *Nanomedicine: Nanotechnology, Biology, and Medicine*. Elsevier Inc., 12(3), pp. 789–799.
- Eckhardt, S., Brunetto, P. S., Gagnon, J., Priebe, M., Giese, B. and Fromm, K. M. (2012) ‘Nanobio silver: its interactions with peptides and bacteria, and its uses in medicine’, *Chemical Review*, 113(7), 4708–54.
- Egan, S., Thomas, T. and Kjelleberg, S. (2008) ‘Unlocking the diversity and biotechnological potential of marine surface associated microbial communities’, *Current Opinion in Microbiology*, 11(3), pp. 219–225.
- Eilers, H., Pernthaler, J., Glöckner, F. O. and Amann, R. (2000) ‘Culturability and in situ abundance of pelagic Bacteria from the North Sea’, *Applied and Environmental Microbiology*, 66(7), pp. 3044–3051.
- Eklund, B. and Watermann, B. (2018) ‘Persistence of TBT and copper in excess on leisure boat hulls around the Baltic Sea’, *Environmental Science and Pollution Research*. Environmental Science and Pollution Research, 25(15), pp. 14595–14605.

- Elifantz, H., Horn, G., Ayon, M., Cohen, Y. and Minz, D. (2013) 'Rhodobacteraceae are the key members of the microbial community of the initial biofilm formed in Eastern Mediterranean coastal seawater', *FEMS Microbiology Ecology*, 85(2), pp. 348–357.
- Fabres-Klein, M. H., Caizer Santos, M. J., Contelli Klein, R., Nunes de Souza, G. and de Oliveira Barros Ribon, A. (2015) 'An association between milk and slime increases biofilm production by bovine *Staphylococcus aureus*', *BMC Veterinary Research*, 11(1), pp. 1–8.
- Faille, C. and Carpentier, B. (2009) *Food contact surfaces, surface soiling and biofilm formation, Biofilms in the Food and Beverage Industries*. Woodhead Publishing Limited.
- De Falco, G., Ciardiello, R., Commodo, M., Del Gaudio, P., Minutolo, P., Porta, A. and D'Anna, A. (2018) 'TiO₂ nanoparticle coatings with advanced antibacterial and hydrophilic properties prepared by flame aerosol synthesis and thermophoretic deposition', *Surface and Coatings Technology*. Elsevier B.V, 349, pp. 830–837.
- Felicia, D. M., Rochiem, R. and Laia, S. M. (2018) 'The effect of silver (Ag) addition to mechanical and electrical properties of copper alloy (Cu) casting product', *AIP Conference Proceedings*, 1945.
- Fernández-Delgado, M., Duque, Z., Rojas, H., Suárez, P., Contreras, M., García-Amado, M. A. and Alciaturi, C. (2015) 'Environmental scanning electron microscopy analysis of *Proteus mirabilis* biofilms grown on chitin and stainless steel', *Annals of Microbiology*, 65(3), pp. 1401–1409.
- Festa, R. A. and Thiele, D. J. (2011) 'Copper: An essential metal in biology', *Current Biology*. Elsevier, 21(21), pp. R877–R883.
- Figueiredo, J., Loureiro, S. and Martins, R. (2020) 'Hazard of novel anti-fouling nanomaterials and biocides DCOIT and silver to marine organisms', *Environmental Science: Nano*, 7(6), pp. 1670–1680.
- Fitridge, I., Dempster, T., Guenther, J. and de Nys, R. (2012) 'The impact and control of biofouling in marine aquaculture: a review', *Biofouling*. Taylor & Francis, 28(7), pp. 649–669.
- Flemming, H.-C. and Ridgway, H. (2009) 'Biofilm Control: Conventional and Alternative Approaches', in *Marine and Industrial Biofouling*. Berlin, Heidelberg: Springer Berlin Heidelberg, pp. 103–117.

- Flemming, H. (2016) ‘EPS - Then and Now’, pp. 1–18.
- Flemming, H. C. (2020) ‘Biofouling and me: My Stockholm syndrome with biofilms’, *Water Research*. Elsevier Ltd, 173, p. 115576.
- Flemming, H. C., Neu, T. R. and Wozniak, D. J. (2007) ‘The EPS matrix: The “House of Biofilm Cells”’, *Journal of Bacteriology*, 189(22), pp. 7945–7947.
- Flemming, H. C. and Wingender, J. (2010) ‘The biofilm matrix’, *Nature Reviews Microbiology*. Nature Publishing Group, 8(9), pp. 623–633.
- Flemming, H. C., Wingender, J., Szewzyk, U., Steinberg, P., Rice, S. A. and Kjelleberg, S. (2016) ‘Biofilms: An emergent form of bacterial life’, *Nature Reviews Microbiology*. Nature Publishing Group, 14(9), pp. 563–575.
- Fletcher, M. (1991) ‘The Physiological Activity of Bacteria Attached to Solid Surfaces’, *Advances in Microbial Physiology*, 32(C), pp. 53–85.
- Fomina, M. and Skorochod, I. (2020) ‘Microbial interaction with clay minerals and its environmental and biotechnological implications’, *Minerals*, 10(10), pp. 1–54.
- Fox, C. J., Harrop, R. and Wimpenny, A. (1999) ‘Feeding ecology of herring (*Clupea harengus*) larvae in the turbid Blackwater Estuary’, *Marine Biology*, 134(2), pp. 353–365.
- Franklin, M. J., Chang, C., Akiyama, T. and Bothner, B. (2015) ‘New Technologies for Studying Biofilms’, *Microbiology Spectrum*, 3(4), pp. 1–23.
- Furtuna, D. K., Debora, K. and Warsito, E. B. (2018) ‘Comparison of Microbiological Examination by Test Tube and Congo Red Agar Methods to Detect Biofilm Production on Clinical Isolates’, *Folia Medica Indonesiana*, 54(1), p. 22.
- Gavhane, S. K., Sapkale, J. B., Susware, N. K. and Sapkale, S. J. (2021) ‘Impact of heavy metals in riverine and estuarine environment: A review’, *Research Journal of Chemistry and Environment*, 25(5), pp. 226–233.
- Gieroba, B., Sroka-Bartnicka, A., Kazimierczak, P., Kalisz, G., Lewalska-Graczyk, A., Vivcharenko, V., Nowakowski, R., Pieta, I. S. and Przekora, A. (2020) ‘Spectroscopic studies on the temperature-dependent molecular arrangements in hybrid chitosan/1,3-β-D-glucan polymeric matrices’, *International Journal of Biological Macromolecules*. Elsevier B.V, 159, pp. 911–921.
- Giovannoni and Rappé, M. (2000) ‘Evolution, diversity, and molecular ecology of marine prokaryotes’.
- Godwin, L. S. (2003) ‘Hull fouling of maritime vessels as a pathway for marine species

- invasions to the Hawaiian Islands', *Biofouling*, 19(SUPPL.), pp. 123–131.
- Gomes, L. C. and Mergulhão, F. J. (2017) 'SEM analysis of surface impact on biofilm antibiotic treatment', *Scanning*, 2017(c).
- Gomes, L. C., Moreira, J. M. R., Simões, M., Melo, L. F. and Mergulhão, F. J. (2014) 'Biofilm Localization in the Vertical Wall of Shaking 96-Well Plates', *Scientifica*, 2014, pp. 1–6.
- González-Machado, C., Capita, R., Riesco-Peláez, F. and Alonso-Calleja, C. (2018) 'Visualization and quantification of the cellular and extracellular components of *Salmonella agona* biofilms at different stages of development', *PLoS ONE*, 13(7).
- Grossart, H. P., Thorwest, M., Plitzko, I., Brinkhoff, T., Simon, M. and Zeeck, A. (2009) 'Production of a blue pigment (glaukothalin) by marine *Rheinheimera* spp.', *International Journal of Microbiology*, 2009.
- Gu, S., Kang, X., Wang, L., Lichtfouse, E. and Wang, C. (2019) 'Clay mineral adsorbents for heavy metal removal from wastewater: a review', *Environmental Chemistry Letters*. Springer International Publishing, 17(2), pp. 629–654.
- Gu, Y., Yu, L., Mou, J., Wu, D., Xu, M., Zhou, P. and Ren, Y. (2020) 'Research strategies to develop environmentally friendly marine antifouling coatings', *Marine Drugs*, 18(7).
- Guo, H., Li, K., Wang, W., Wang, C. and Shen, Y. (2017) 'Effects of Copper on Hemocyte Apoptosis, ROS Production, and Gene Expression in White Shrimp *Litopenaeus vannamei*', *Biological Trace Element Research*. Biological Trace Element Research, 179(2), pp. 318–326.
- Guo, X. pan, Niu, Z. shun, Lu, D. pei, Feng, J. nan, Chen, Y. ru, Tou, F. yun, Liu, M. and Yang, Y. (2017) 'Bacterial community structure in the intertidal biofilm along the Yangtze Estuary, China', *Marine Pollution Bulletin*. Elsevier, 124(1), pp. 314–320.
- Gutner-Hoch, E., Martins, R., Oliveira, T., Maia, F., Soares, A. M. V. M., Loureiro, S., Piller, C., Preiss, I., Weis, M., Larroze, S. B., Teixeira, T., Tedim, J. and Benayahu, Y. (2018) 'Antimicrofouling efficacy of innovative inorganic nanomaterials loaded with booster biocides', *Journal of Marine Science and Engineering*, 6(1), pp. 1–12.
- Guttenplan, S. B., Blair, K. M. and Kearns, D. B. (2010) 'The EpsE flagellar clutch is

- bifunctional and synergizes with EPS biosynthesis to promote bacillus subtilis biofilm formation', *PLoS Genetics*, 6(12), pp. 1–12.
- Guzman, M., Dille, J. and Godet, S. (2012) 'Synthesis and antibacterial activity of silver nanoparticles against gram-positive and gram-negative bacteria', *Nanomedicine: Nanotechnology, Biology and Medicine*, 8(1), pp. 37–45.
- Hadfield, M. G. (2011) 'Biofilms and marine invertebrate larvae: What bacteria produce that larvae use to choose settlement sites', *Annual Review of Marine Science*, 3, pp. 453–470.
- Hadi, A. A., Malek, N. A. N. N. and Williams, C. D. (2021) 'Structural characterization and antibacterial activity of antibiotic streptomycin immobilized on zeolite synthesized from natural kaolinite', *Biointerface Research in Applied Chemistry*, 11(5), pp. 13573–13586.
- Halla, N., Fernandes, I. P., Heleno, S. A., Costa, P., Boucherit-Otmani, Z., Boucherit, K., Rodrigues, A. E., Ferreira, I. C. F. R. and Barreiro, M. F. (2018) 'Cosmetics preservation: A review on present strategies', *Molecules*, 23(7), pp. 1–41.
- Haney, E. F., Trimble, M. J., Cheng, J. T., Vallé, Q. and Hancock, R. E. W. (2018) 'Critical assessment of methods to quantify biofilm growth and evaluate antibiofilm activity of host defence peptides', *Biomolecules*, 8(2), pp. 1–22.
- Harder, T., Campbell, A. H., Egan, S. and Steinberg, P. D. (2012) 'Chemical Mediation of Ternary Interactions Between Marine Holobionts and Their Environment as Exemplified by the Red Alga *Delisea pulchra*', *Journal of Chemical Ecology*, 38(5), pp. 442–450.
- Hassan, A., Usman, J., Kaleem, F., Omair, M., Khalid, A. and Iqbal, M. (2011) 'Evaluation of different detection methods of biofilm formation in the clinical isolates.', *The Brazilian journal of infectious diseases : an official publication of the Brazilian Society of Infectious Diseases*. Elsevier, 15(4), pp. 305–311.
- He, X., Tian, F., Bai, X., Yuan, C., Wang, C. and Neville, A. (2020) 'Biomimetic lubricant-infused titania nanoparticle surfaces via layer-by-layer deposition to control biofouling', *Applied Surface Science*. Elsevier B.V., 515, p. 146064.
- Heidarian, S., Mohammadipanah, F., Maghsoudlou, A., Dashti, Y. and Challis, G. L. (2019) 'Anti-microfouling activity of *Glycomyces sediminimaris* UTMC 2460 on dominant fouling bacteria of Iran marine habitats', *Frontiers in*

- Microbiology*, 10(JAN), pp. 1–14.
- Hellio, C., Tsoukatou, M., Maréchal, J. P., Aldred, N., Beaupoil, C., Clare, A. S., Vagias, C. and Roussis, V. (2005) ‘Inhibitory effects of Mediterranean sponge extracts and metabolites on larval settlement of the barnacle *Balanus amphitrite*’, *Marine Biotechnology*, 7(4), pp. 297–305.
- Herget, S., Toukach, P. V., Ranzinger, R., Hull, W. E., Knirel, Y. A. and Von Der Lieth, C. W. (2008) ‘Statistical analysis of the bacterial carbohydrate structure data base (BCSDB): Characteristics and diversity of bacterial carbohydrates in comparison with mammalian glycans’, *BMC Structural Biology*, 8.
- Herzberg, M., Kang, S. and Elimelech, M. (2009) ‘Role of extracellular polymeric substances (EPS) in biofouling of reverse osmosis membranes’, *Environmental Science and Technology*, 43(12), pp. 4393–4398.
- Hobley, L., Harkins, C., MacPhee, C. E. and Stanley-Wall, N. R. (2015) ‘Giving structure to the biofilm matrix: an overview of individual strategies and emerging common themes.’, *FEMS microbiology reviews*, 39(5), pp. 649–69.
- Holešová, S., Samlíková, M., Pazdziora, E. and Valášková, M. (2013) ‘Antibacterial activity of organomontmorillonites and organovermiculites prepared using chlorhexidine diacetate’, *Applied Clay Science*, 83–84, pp. 17–23.
- Hossain, M. S., Hossain, M. B., Rakib, M. R. J., Jolly, Y. N., Ullah, M. A. and Elliott, M. (2021) ‘Ecological and human health risk evaluation using pollution indices: A case study of the largest mangrove ecosystem of Bangladesh’, *Regional Studies in Marine Science*. Elsevier, p. 101913.
- Hou, X., Liu, S. and Zhang, Z. (2015) ‘Role of extracellular polymeric substance in determining the high aggregation ability of anammox sludge’, *Water Research*. Pergamon, 75, pp. 51–62.
- Huang, Y., Chakraborty, S. and Liang, H. (2020) ‘Methods to probe the formation of biofilms: Applications in foods and related surfaces’, *Analytical Methods*. Royal Society of Chemistry, 12(4), pp. 416–432.
- Huang, Y. L., Li, M., Yu, Z. and Qian, P. Y. (2011) ‘Correlation between pigmentation and larval settlement deterrence by *Pseudoalteromonas* sp. sf57.’, *Biofouling*, 27(3), pp. 287–293.
- Huggett, M. J., Williamson, J. E., De Nys, R., Kjelleberg, S. and Steinberg, P. D. (2006) ‘Larval settlement of the common Australian sea urchin *Heliocidaris*

- erythrogramma* in response to bacteria from the surface of coralline algae', *Oecologia*, 149(4), pp. 604–619.
- Hundáková, M., Valášková, M., Tomášek, V., Pazdziora, E. and Matějová, K. (2013) 'Silver and/or copper vermiculites and their antibacterial effect', *Acta Geodynamica et Geomaterialia*, 10(1), pp. 97–104.
- Hung, O. S., Thiyagarajan, V. and Qian, P. Y. (2008) 'Preferential attachment of barnacle larvae to natural multi-species biofilms: Does surface wettability matter?', *Journal of Experimental Marine Biology and Ecology*, 361(1), pp. 36–41.
- Hung, O. S., Thiyagarajan, V., Wu, R. S. S. and Qian, P. Y. (2005) 'Effect of ultraviolet radiation on biofilms and subsequent larval settlement of *Hydrodides elegans*', *Marine Ecology Progress Series*, 304(July 2014), pp. 155–166.
- Ibrišimović, M. A., I Mehmedinović, N. I., & Hukić, M. (2017) 'A Novel Spectrophotometric Assay for the Determination of Biofilm Forming Capacity of Causative Agents of Urinary Tract Infections.', *International Journal of Engineering Research & Technology (IJERT)*, 6(4), pp. 1225–1230.
- Inbakandan, D., Murthy, P. S., Venkatesan, R. and Khan, S. A. (2010) '16S rDNA sequence analysis of culturable marine biofilm forming bacteria from a ship's hull.', *Biofouling*, 26(8), pp. 893–899.
- Isah, M., Asraf, M. H., Malek, N. A. N. N., Jemon, K., Sani, N. S., Muhammad, M. S., Wahab, M. F. A. and Saidin, M. A. R. (2020) 'Preparation and characterization of chlorhexidine modified zinc-kaolinite and its antibacterial activity against bacteria isolated from water vending machine', *Journal of Environmental Chemical Engineering*. Elsevier B.V., 8(2), p. 103545.
- Ivče, R., Bakota, M., Kos, S. and Brčić, D. (2020) 'Advanced numerical method for determining the wetted area of container ships for increased estimation accuracy of copper biocide emissions', *Journal of Marine Science and Engineering*, 8(11), pp. 1–18.
- Jaime-Acuña, O. E., Meza-Villecas, A., Vasquez-Peña, M., Raymond-Herrera, O., Villavicencio-García, H., Petranovskii, V., Vazquez-Duhalt, R. and Huerta-Saquero, A. (2016) 'Synthesis and Complete Antimicrobial Characterization of CEOBACTER, an Ag-Based Nanocomposite', *PLOS ONE*. Edited by A.

- Almeida. Wiley, 11(11), p. e0166205.
- Jain, K., Parida, S., Mangwani, N., Dash, H. R. and Das, S. (2013) ‘Isolation and characterization of biofilm-forming bacteria and associated extracellular polymeric substances from oral cavity’, *Annals of Microbiology*, 63(4), pp. 1553–1562.
- Jarrell, K. F. and McBride, M. J. (2008) ‘The surprisingly diverse ways that prokaryotes move’, *Nature Reviews Microbiology*, 6(6), pp. 466–476.
- Jeong, S., Kim, J., Kim, H., Chung, K. and Yoon, H. (2018) ‘Identification of preponderant marine bacteria and their biofouling characteristics on adsorbents of different sizes and shapes in seawater’, 26(3), pp. 458–464.
- Jiang, T., Yang, Y., Huang, Z. and Qiu, G. (2003) ‘Simultaneous leaching of manganese and silver from manganese–silver ores at room temperature’, *Hydrometallurgy*. Elsevier, 69(1–3), pp. 177–186.
- Jiao, Y., Cody, George D., Harding, A. K., Wilmes, P., Schrenk, M., Wheeler, K. E., Banfield, J. F. and Thelen, M. P. (2010) ‘Characterization of extracellular polymeric substances from acidophilic microbial biofilms.’, *Applied and environmental microbiology*. American Society for Microbiology, 76(9), pp. 2916–22.
- Jiao, Y., Cody, George D., Harding, A. K., Wilmes, P., Schrenk, M., Wheeler, K. E., Banfield, J. F. and Thelen, M. P. (2010) ‘Characterization of extracellular polymeric substances from acidophilic microbial biofilms’, *Applied and Environmental Microbiology*, 76(9), pp. 2916–2922.
- Jin, X. and Marshall, J. S. (2020) ‘Influence of cell interaction forces on growth of bacterial biofilms’, *Physics of Fluids*. AIP Publishing, LLC, 32(9).
- Jones, P. R., Cottrell, M. T., Kirchman, D. L. and Dexter, S. C. (2007a) ‘Bacterial community structure of biofilms on artificial surfaces in an estuary’, *Microbial Ecology*, 53(1), pp. 153–162.
- Jones, P. R., Cottrell, M. T., Kirchman, D. L. and Dexter, S. C. (2007b) ‘Bacterial Community Structure of Biofilms on Artificial Surfaces in an Estuary’, *Microbial Ecology*, 53(1), pp. 153–162.
- Jonsson, P. R., Berntsson, K. M. and Larsson, A. I. (2004) ‘Linking larval supply to recruitment: Flow-mediated control of initial adhesion of barnacle larvae’, *Ecology*, 85(10), pp. 2850–2859.
- Jou, S. K. and Malek, N. A. N. N. (2016) ‘Characterization and antibacterial activity

- of chlorhexidine loaded silver-kaolinite', *Applied Clay Science*. Elsevier B.V., 127–128, pp. 1–9.
- Jubeh, B., Breijyeh, Z. and Karaman, R. (2020) 'Resistance of gram-positive bacteria to current antibacterial agents and overcoming approaches', *Molecules*, 25(12).
- Juergensmeyer, M. A., Nelson, E. S. and Juergensmeyer, E. A. (2007) 'Shaking alone, without concurrent aeration, affects the growth characteristics of *Escherichia coli*', *Letters in Applied Microbiology*, 45(2), pp. 179–183.
- Karel, F. B., Koparal, A. S. and Kaynak, E. (2015a) 'Development of silver ion doped antibacterial clays and investigation of their antibacterial activity', *Advances in Materials Science and Engineering*, 2015.
- Karel, F. B., Koparal, A. S. and Kaynak, E. (2015b) 'Development of Silver Ion Doped Antibacterial Clays and Investigation of Their Antibacterial Activity', *Advances in Materials Science and Engineering*. Hindawi, 2015, pp. 1–6.
- Karimi, K., Amini, J., Harighi, B. and Bahramnejad, B. (2012) 'Evaluation of biocontrol potential of *Pseudomonas* and *Bacillus* spp. against Fusarium wilt of chickpea', *Australian Journal of Crop Science*, 6(4), pp. 695–703.
- Karunakaran, E., Mukherjee, J., Ramalingam, B. and Biggs, C. A. (2011) "Biofilmology": A multidisciplinary review of the study of microbial biofilms', *Applied Microbiology and Biotechnology*, 90(6), pp. 1869–1881.
- Karygianni, L., Ren, Z., Koo, H. and Thurnheer, T. (2020) 'Biofilm Matrixome: Extracellular Components in Structured Microbial Communities', *Trends in Microbiology*. Elsevier Current Trends, 28(8), pp. 668–681.
- Kataoka, C., Kato, Y., Ariyoshi, T., Takasu, M., Narazaki, T., Nagasaka, S., Tatsuta, H. and Kashiwada, S. (2018) 'Comparative toxicities of silver nitrate, silver nanocolloids, and silver chloro-complexes to Japanese medaka embryos, and later effects on population growth rate', *Environmental Pollution*. Elsevier Ltd, 233, pp. 1155–1163.
- Katranitsas, A., Castritsi-Catharios, J. and Persoone, G. (2003) 'The effects of a copper-based antifouling paint on mortality and enzymatic activity of a non-target marine organism', *Marine Pollution Bulletin*. Elsevier Ltd, 46(11), pp. 1491–1494.
- Katsanevakis, S., Zenetos, A., Belchior, C. and Cardoso, A. C. (2013) 'Invading European Seas: Assessing pathways of introduction of marine aliens', *Ocean*

- and Coastal Management*. Elsevier Ltd, 76, pp. 64–74.
- Katsikogianni, M. and Missirlis, Y. F. (2004) ‘Concise review of mechanisms of bacterial adhesion to biomaterials and of techniques used in estimating bacteria-material interactions - True Open Access’, *MEu. rKopatesaink oCgeialnsn ain adn dM Ya.tFe.r iMaliss*, 8(August), pp. 37–57.
- Kavitha, S. and Raghavan, V. (2018) ‘Isolation and characterization of marine biofilm forming bacteria from a ship’s hull’, *Frontiers in Biology*, 13(3), pp. 208–214.
- Kędziora, A., Speruda, M., Krzyżewska, E., Rybka, J., Lukowiak, A. and Bugała-Płoskońska, G. (2018) ‘Similarities and differences between silver ions and silver in nanoforms as antibacterial agents’, *International Journal of Molecular Sciences*, 19(2).
- Khajotia, S. S., Smart, K. H., Pilula, M. and Thompson, D. M. (2013) ‘Concurrent quantification of cellular and extracellular components of biofilms.’, *Journal of visualized experiments : JoVE*, (82), pp. 1–7.
- Kilian, M. (2018) ‘The oral microbiome – friend or foe?’, *European Journal of Oral Sciences*, 126, pp. 5–12.
- Kim, L. H., Jung, Y., Kim, S. J., Kim, C. M., Yu, H. W., Park, H. D. and Kim, I. S. (2015) ‘Use of rhamnolipid biosurfactant for membrane biofouling prevention and cleaning’, *Biofouling*, 31(2), pp. 211–220.
- Kim, M., Oh, H. S., Park, S. C. and Chun, J. (2014) ‘Towards a taxonomic coherence between average nucleotide identity and 16S rRNA gene sequence similarity for species demarcation of prokaryotes’, *International Journal of Systematic and Evolutionary Microbiology*, 64(PART 2), pp. 346–351.
- Klausen, M. M., Thomsen, T. R., Nielsen, J. L., Mikkelsen, L. H. and Nielsen, P. H. (2004) ‘Variations in microcolony strength of probe-defined bacteria in activated sludge flocs’, *FEMS Microbiology Ecology*, 50(2), pp. 123–132.
- Klein, M. I., Scott-Anne, K. M., Gregoire, S., Rosalen, P. L. and Koo, H. (2012) ‘Molecular approaches for viable bacterial population and transcriptional analyses in a rodent model of dental caries’, *Molecular Oral Microbiology*, 27(5), pp. 350–361.
- Klöckner, W. and Büchs, J. (2012) ‘Advances in shaking technologies’, *Trends in Biotechnology*, 30(6), pp. 307–314.
- Koedoeder, C., Stock, W., Willems, A., Mangelinckx, S., De Troch, M., Vyverman,

- W. and Sabbe, K. (2019) ‘Diatom-bacteria interactions modulate the composition and productivity of benthic diatom biofilms’, *Frontiers in Microbiology*, 10(JUN), pp. 1–11.
- Kokare, C. R., Chakraborty, S., Khopade, A. N. and Mahadik, K. R. (2009) ‘Biofilm: Importance and applications’, *Indian Journal of Biotechnology*, 8(2), pp. 159–168.
- Kolter, R. (2010) ‘Biofilms in lab and nature: A molecular geneticist’s voyage to microbial ecology’, *International Microbiology*, 13(1), pp. 1–7.
- Koo, H., Allan, R. N., Howlin, R. P., Stoodley, P. and Hall-Stoodley, L. (2017) ‘Targeting microbial biofilms: Current and prospective therapeutic strategies’, *Nature Reviews Microbiology*. Nature Publishing Group, 15(12), pp. 740–755.
- Korkut, E. and Atlar, M. (2012) ‘An experimental investigation of the effect of foul release coating application on performance, noise and cavitation characteristics of marine propellers’, *Ocean Engineering*, 41(June 2014), pp. 1–12.
- Kostakioti, M., Hadjifrangiskou, M. and Hultgren, S. J. (2013) ‘Bacterial biofilms: Development, dispersal, and therapeutic strategies in the dawn of the postantibiotic era’, *Cold Spring Harbor Perspectives in Medicine*, 3(4), pp. 1–23.
- Kotal, M. and Bhowmick, A. K. (2015) ‘Polymer nanocomposites from modified clays: Recent advances and challenges’, *Progress in Polymer Science*. Elsevier Ltd, 51, pp. 127–187.
- Kraeling, M. E. K., Topping, V. D., Keltner, Z. M., Belgrave, K. R., Bailey, K. D., Gao, X. and Yourick, J. J. (2018) ‘In vitro percutaneous penetration of silver nanoparticles in pig and human skin’, *Regulatory Toxicology and Pharmacology*, 95(January), pp. 314–322.
- Kragh, K. N., Hutchison, J. B., Melaugh, G., Rodesney, C., Roberts, A. E. L., Irie, Y., Jensen, P., Diggle, S. P., Allen, R. J., Gordon, V. and Bjarnsholt, T. (2016) ‘Role of multicellular aggregates in biofilm formation’, *mBio*, 7(2), pp. 1–11.
- Krajišnik, D., Daković, A., Milojević, M., Malenović, A., Kragović, M., Bogdanović, D. B., Dondur, V. and Milić, J. (2011) ‘Properties of diclofenac sodium sorption onto natural zeolite modified with cetylpyridinium chloride’, *Colloids and Surfaces B: Biointerfaces*, 83(1), pp. 165–172.

- Krishnan, S., Weinman, C. J. and Ober, C. K. (2008) ‘Advances in polymers for anti-biofouling surfaces’, *Journal of Materials Chemistry*, 18(29), pp. 3405–3413.
- Kumar, A. S., Mody, K. and Jha, B. (2007) ‘Bacterial exopolysaccharides - A perception’, *Journal of Basic Microbiology*, 47(2), pp. 103–117.
- Kumar, S., Hiremath, S. S., Ramachandran, B. and Muthuvijayan, V. (2019). ‘Effect of Surface Finish on Wettability and Bacterial Adhesion of Micromachined Biomaterials’, *Biotribology*, 100095.
- Kumar, M. A., Anandapandian, K. T. K. and Parthiban, K. (2011) ‘Production and characterization of exopolysaccharides (EPS) from biofilm forming marine bacterium’, *Brazilian Archives of Biology and Technology*, 54(4), pp. 259–265.
- Kumar, S., Shukla, A., Baul, P. P., Mitra, A. and Halder, D. (2018) ‘Biodegradable hybrid nanocomposites of chitosan/gelatin and silver nanoparticles for active food packaging applications’, *Food Packaging and Shelf Life*. Elsevier, 16(March), pp. 178–184.
- Kumari, G. V., Asha, S., Ananth, A. N., Rajan, M. A. J. and Mathavan, T. (2018) ‘Polyethyleneglycol/silver functionalized reduced graphene oxide aerogel for environmental application’, *AIP Conference Proceedings*, 1942.
- Kwon, K. K., Lee, H. S., Jung, S. Y., Yim, J. H., Lee, J. H. and Lee, H. K. (2002) ‘Isolation and identification of biofilm-forming marine bacteria on glass surfaces in Dae-Ho Dike, Korea’, *Journal of Microbiology*, 40(4), pp. 260–266.
- De la Fuente-Núñez, C., Reffuveille, F., Fernández, L. and Hancock, R. E. W. (2013) ‘Bacterial biofilm development as a multicellular adaptation: Antibiotic resistance and new therapeutic strategies’, *Current Opinion in Microbiology*, 16(5), pp. 580–589.
- Lahir, Y. K. (2020) ‘A Interactions at Interface between Nanomaterial’s and Biofilm : A General Survey’, *Advances in Clinical Toxicology*, 5(3).
- Landoulsi, J., Cooksey, K. E. and Dupres, V. (2011) ‘Review - Interactions between diatoms and stainless steel: Focus on biofouling and biocorrosion’, *Biofouling*, 27(10), pp. 1105–1124.
- Lara-Martín, P. A., Li, X., Bopp, R. F. and Brownawell, B. J. (2010) ‘Occurrence of alkyltrimethylammonium compounds in urban estuarine sediments: Behentrimonium as a new emerging contaminant’, *Environmental Science*

- and Technology*, 44(19), pp. 7569–7575.
- Lau, P. C. Y., Dutcher, J. R., Beveridge, T. J. and Lam, J. S. (2009) ‘Absolute Quantitation of Bacterial Biofilm Adhesion and Viscoelasticity by Microbead Force Spectroscopy’, *Biophysical Journal*. Cell Press, 96(7), pp. 2935–2948.
- Lau, S. C. K., Harder, T. and Qian, P. (2003) ‘Induction of larval settlement in the serpulid polychaete *Hydroides elegans* (Haswell): Role of bacterial extracellular polymers’, *Biofouling*, 19(3), pp. 197–204.
- Lau, S., Thiagarajan, V., Cheung, S. and Qian, P. (2005) ‘Roles of bacterial community composition in biofilms as a mediator for larval settlement of three marine invertebrates’, *Aquatic Microbial Ecology*, 38, pp. 41–51.
- Leary, D. H., Li, R. W., Hamdan, L. J., Hervey, W. J., Lebedev, N., Wang, Z., Deschamps, J. R., Kusterbeck, A. W. and Vora, G. J. (2014) ‘Integrated metagenomic and metaproteomic analyses of marine biofilm communities’, *Biofouling*, 30(10), pp. 1211–1223.
- Lee, J.-W., Nam, J.-H., Kim, Y.-H., Lee, K.-H. and Lee, D.-H. (2008) ‘Bacterial communities in the initial stage of marine biofilm formation on artificial surfaces’, *The Journal of Microbiology*, 46(2), pp. 174–182.
- Lee, K.-J., Lee, M.-A., Hwang, W., Park, H. and Lee, K.-H. (2016) ‘Deacylated lipopolysaccharides inhibit biofilm formation by Gram-negative bacteria.’, *Biofouling*. Taylor & Francis, 32(7), pp. 711–723.
- Lee, O. O., Chung, H. C., Yang, J., Wang, Y., Dash, S., Wang, H. and Qian, P. Y. (2014) ‘Molecular Techniques Revealed Highly Diverse Microbial Communities in Natural Marine Biofilms on Polystyrene Dishes for Invertebrate Larval Settlement’, *Microbial Ecology*, 68(1), pp. 81–93.
- Lefèvre, C. T., Bernadac, A., Yu-Zhang, K., Pradel, N. and Wu, L. F. (2009) ‘Isolation and characterization of a magnetotactic bacterial culture from the Mediterranean Sea’, *Environmental Microbiology*, 11(7), pp. 1646–1657.
- Lekamge, S., Miranda, A. F., Abraham, A., Li, V., Shukla, R., Bansal, V. and Nugegoda, D. (2018) ‘The toxicity of silver nanoparticles (AgNPs) to three freshwater invertebrates with different life strategies: *Hydra vulgaris*, *Daphnia carinata* and *Paratya australiensis*’, *Frontiers in Environmental Science*, 6 (Dec), pp. 1–13.
- Lem, K. W., Choudhury, A., Lakhani, A. A., Kuyate, P., Haw, J. R., Lee, D. S., Iqbal, Z. and Brumlik, C. J. (2012) ‘Use of Nanosilver in Consumer Products

- Products Delivery Systems Hierarchy Formulation / Compounding Clusters / Structures Molecules Features Identify the Unmet Needs from Materials to / from Applications', *Bentham Science Publishers*, 6(1), pp. 60–72.
- Lewis, J.A. (1998) 'Marine Biofouling and its prevention on underwater surfaces', *Materials Forum*, 22, pp. 41-61.
- Lewis, C., Ellis, R. P., Vernon, E., Elliot, K., Newbatt, S. and Wilson, R. W. (2016) 'Ocean acidification increases copper toxicity differentially in two key marine invertebrates with distinct acid-base responses', *Scientific Reports*. Nature Publishing Group, 6(Sept), pp. 1–10.
- Lezzi, M., Del Pasqua, M., Pierri, C. and Giangrande, A. (2018) 'Seasonal non-indigenous species succession in a marine macrofouling invertebrate community', *Biological Invasions*. Springer International Publishing, 20(4), pp. 937–961.
- Li, L., Feng, Y., Liu, Y., Wei, B., Guo, J., Jiao, W., Zhang, Z. and Zhang, Q. (2016) 'Titanium dioxide nanoparticles modified by salicylic acid and arginine: Structure, surface properties and photocatalytic decomposition of p-nitrophenol', *Applied Surface Science*, 363, pp. 627–635.
- Li, S. W., Sheng, G. P., Cheng, Y. Y. and Yu, H. Q. (2016) 'Redox properties of extracellular polymeric substances (EPS) from electroactive bacteria', *Scientific Reports* 6, 39098, pp. 1–7.
- Li, T., Huang, X., Wang, Q. and Yang, G. (2020) 'Adsorption of metal ions at kaolinite surfaces: Ion-specific effects, and impacts of charge source and hydroxide formation', *Applied Clay Science*, 194(May).
- Li, X., Yan, Z. and Xu, J. (2003) 'Quantitative variation of biofilms among strains in natural populations of *Candida albicans*', *Microbiology*, 149(2), pp. 353–362.
- Li, Y. and Ning, C. (2019) 'Latest research progress of marine microbiological corrosion and bio-fouling, and new approaches of marine anti-corrosion and anti-fouling', *Bioactive Materials*. Elsevier, 4(Jan), pp. 189–195.
- Li, Z. and Gallus, L. (2005) 'Surface configuration of sorbed hexadecyltrimethylammonium on kaolinite as indicated by surfactant and counterion sorption, cation desorption, and FTIR', *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 264(1–3), pp. 61–67.
- Lima, L. F. O., Habu, S., Gern, J. C., Nascimento, B. M., Parada, J. L., Noseda, M. D.,

- Gonçalves, A. G., Nisha, V. R., Pandey, A., Soccol, V. T. and Soccol, C. R. (2008) 'Production and characterization of the exopolysaccharides produced by *Agaricus brasiliensis* in submerged fermentation', *Applied Biochemistry and Biotechnology*, 151(2–3), pp. 283–294.
- Limoli, D. H., Jones, C. J. and Wozniak, D. J. (2015) 'Bacterial Extracellular Polysaccharides in Biofilm Formation and Function', *Microbiology Spectrum*, 3(3), pp. 1–30.
- Lin, Q., Lim, J. Y. C., Xue, K., Yew, P. Y. M., Owh, C., Chee, P. L. and Loh, X. J. (2020) 'Sanitizing agents for virus inactivation and disinfection', *View*, 1(2).
- Lindgren, J. F., Ytreberg, E., Holmqvist, A., Dahlström, Magnus, Dahl, P., Berglin, M., Wrangle, A.-L. and Dahlström, Mia (2018) 'Biofouling The Journal of Bioadhesion and Biofilm Research Copper release rate needed to inhibit fouling on the west coast of Sweden and control of copper release using zinc oxide Copper release rate needed to inhibit fouling on the west coast of Sweden and control of copper release using zinc oxide', *Biofouling*, 34(4), pp. 453–463.
- Lindholdt, A., Olsen, S. M., Yebra, D. M. and Kiil, S. (2015) 'Effects of biofouling development on drag forces of hull coatings for ocean-going ships: A review', *Journal of Coatings Technology and Research*. Springer US, 12(3), pp. 415–444.
- Liu, Y., Yin, Y., Wang, L., Zhang, W., Chen, X., Yang, X., Xu, J. and Ma, G. (2013) 'Surface hydrophobicity of microparticles modulates adjuvanticity', *Journal of Materials Chemistry B*, 1(32), pp. 3888–3896.
- Londono, S. C., Hartnett, H. E. and Williams, L. B. (2017) 'Antibacterial Activity of Aluminum in Clay from the Colombian Amazon', *Environmental Science and Technology*, 51(4), pp. 2401–2408.
- López, D., Vlamakis, H. and Kolter, R. (2010) 'Biofilms.', *Cold Spring Harbor perspectives in biology*, 2(7).
- Lorite, G. S., Rodrigues, C. M., de Souza, A. A., Kranz, C., Mizaikoff, B. and Cotta, M. A. (2011) 'The role of conditioning film formation and surface chemical changes on *Xylella fastidiosa* adhesion and biofilm evolution', *Journal of Colloid and Interface Science*. Elsevier Inc., 359(1), pp. 289–295.
- Lüdecke, C., Roth, M., Yu, W., Horn, U., Bossert, J. and Jandt, K. D. (2016) 'Nanorough titanium surfaces reduce adhesion of *Escherichia coli* and

- Staphylococcus aureus* via nano adhesion points', *Colloids and Surfaces B: Biointerfaces*. Elsevier B.V., 145, pp. 617–625.
- Luppens, S. B. I., Reij, M. W., Van der Heijden, R. W. L., Rombouts, F. M. and Abée, T. (2002) 'Development of a standard test to assess the resistance of *Staphylococcus aureus* biofilm cells to disinfectants', *Applied and Environmental Microbiology*, 68(9), pp. 4194–4200.
- Ma, C. and Eggleton, R. A. (1999) 'Cation exchange capacity of kaolinite', *Clays and Clay Minerals*, 47(2), pp. 174–180.
- Ma, W., Peng, D., Walker, S. L., Cao, B., Gao, C. H., Huang, Q. and Cai, P. (2017) '*Bacillus subtilis* biofilm development in the presence of soil clay minerals and iron oxides', *npj Biofilms and Microbiomes*. Springer US, 3(1), pp. 0–1.
- Madsen, J. S., Røder, H. L., Russel, J., Sørensen, H., Burmølle, M. and Sørensen, S. J. (2016) 'Coexistence facilitates interspecific biofilm formation in complex microbial communities', *Environmental microbiology*, 18(8), pp. 2565–2574.
- Maeda, H. and Ishida, N. (1967) 'Specificity of binding of hexopyranosyl polysaccharides with fluorescent brightener', *Journal of Biochemistry*, 62(2), pp. 276–278.
- Mahajan, P. G., Desai, N. K., Dalavi, D. K., Bhopate, D. P., Kolekar, G. B. and Patil, S. R. (2015) 'Cetyltrimethylammonium bromide capped 9-anthraldehyde nanoparticles for selective recognition of phosphate anion in aqueous solution based on fluorescence quenching and application for analysis of chloroquine', *Journal of Fluorescence*, 25(1), pp. 31–38.
- Maia, F., Silva, a. P., Fernandes, S., Cunha, a., Almeida, a., Tedim, J., Zheludkevich, M. L. and Ferreira, M. G. S. (2015) 'Incorporation of biocides in nanocapsules for protective coatings used in maritime applications', *Chemical Engineering Journal*. Elsevier B.V., 270, pp. 150–157.
- Majdan, M., Bujacka, M., Sabah, E., Gladysz-Płaska, A., Pikus, S., Sternik, D., Komosa, Z. and Padewski, A. (2009) 'Unexpected difference in phenol sorption on PTMA- and BTMA-bentonite', *Journal of Environmental Management*, 91(1), pp. 195–205.
- Maki, J. S., Rittschof, D., Samuelsson, M., Szewzyk, U., Yule, a B., I, S. and Costlow, J. D. (1990) 'Effect of marine bacteria and their exopolymers on the attachment of barnacle cypris larvae settlement and metamorphosis into the adult', *Bulletin of Marine Science*, 46(2), pp. 499–511.

- Malachová, K., Praus, P., Rybková, Z., and Kozák, O. (2011) ‘Antibacterial and antifungal activities of silver, copper and zinc montmorillonites’ *Applied Clay Science*, 53(4), 642–645.
- Malek, N. A. N. N., Ishak, S. A. and Kadir, M. R. A. (2013) ‘Antibacterial Activity of Copper and CTAB Modified Clays against *Pseudomonas aeruginosa*’, *Advanced Materials Research*, 626, pp. 178–182.
- Malek, N. A. N. N. and Ramli, N. I. (2015) ‘Characterization and antibacterial activity of cetylpyridinium bromide (CPB) immobilized on kaolinite with different CPB loadings’, *Applied Clay Science*. Elsevier B.V., 109–110, pp. 8–14.
- Malik, N., Shrivastava, S. and Ghosh, S. B. (2018) ‘Moisture Absorption Behaviour of Biopolymer Polycapralactone (PCL) / Organo Modified Montmorillonite Clay (OMMT) biocomposite films’, *IOP Conference Series: Materials Science and Engineering*, 346(1).
- Mandlik, A., Swierczynski, A., Das, A. and Ton-That, H. (2008) ‘Pili in Gram-positive bacteria: assembly, involvement in colonization and biofilm development’, *Trends in Microbiology*, 16(1), pp. 33–40.
- Mangala Nagasundari, S., Muthu, K., Kaviyarasu, K., Farraj, D. A. A. and Alkufeyd, R. M. (2021) ‘Current trends of Silver doped Zinc oxide nanowires photocatalytic degradation for energy and environmental application’, *Surfaces and Interfaces*. Elsevier, 23, p. 100931.
- Marassi, V., Di Cristo, L., Smith, S. G. J., Ortelli, S., Blosi, M., Costa, A. L., Reschiglian, P., Volkov, Y. and Prina-Mello, A. (2018) ‘Silver nanoparticles as a medical device in healthcare settings: A five-step approach for candidate screening of coating agents’, *Royal Society Open Science*, 5(1).
- María, A., Solarte, F., Massani, M. B., Molina, V., Guerrero, M. B. and Sánchez, R. M. T. (2018) ‘Correlating Antimicrobial activity and Structure in Montmorillonite modified with Hexadecyltrimethylammonium and Silver’, *International Journal of ChemTech Research*, 11(06), pp. 209–223.
- Di Martino, P. (2018) ‘Extracellular polymeric substances, a key element in understanding biofilm phenotype’, *AIMS Microbiology*, 4(2), pp. 274–288.
- Mason, D. J., Lopéz-Amorós, R., Allman, R., Stark, J. M. and Lloyd, D. (1995) ‘The ability of membrane potential dyes and calcafluor white to distinguish between viable and non-viable bacteria’, *Journal of Applied Bacteriology*, 78(3), pp. 309–315.

- Mathias, J.D. and Stoodley, P. (2011) *Advances in Biofilm Mechanics*. in Biofilm Highlights, 111–139.
- Matile, S., Jentzsch, A. V., Montenegro, J. and Fin, A. (2011) ‘Recent synthetic transport systems’, *Chemical Society Reviews*, 40(5), pp. 2453–2474.
- Matthiessen, P., Reed, J. and Johnson, M. (1999) ‘Sources and potential effects of copper and zinc concentrations in the estuarine waters of Essex and Suffolk, United Kingdom’, *Marine Pollution Bulletin*. Pergamon, 38(10), pp. 908–920.
- McCutcheon, J. and Southam, G. (2018) ‘Advanced biofilm staining techniques for TEM and SEM in geomicrobiology: Implications for visualizing EPS architecture, mineral nucleation, and microfossil generation’, *Chemical Geology*, 498, pp. 115–127.
- McDonald, M. J., Gehrig, S. M., Meintjes, P. L., Zhang, X. X. and Rainey, P. B. (2009) ‘Adaptive divergence in experimental populations of *Pseudomonas fluorescens*. IV. Genetic constraints guide evolutionary trajectories in a parallel adaptive radiation’, *Genetics*, 183(3), pp. 1041–1053.
- McNeil, E. M. (2018) ‘Antifouling: Regulation of biocides in the UK before and after Brexit’, *Marine Policy*. Elsevier Ltd, 92, pp. 58–60.
- Md Saad, N. S. S., Nik Malek, N. A. N. and Chong, C. S. (2016) ‘Antimicrobial Activity Of Copper Kaolinite And Surfactant Modified Copper Kaolinite Against Gram Positive And Gram Negative Bacteria’, *Jurnal Teknologi*, 78(3-2).
- Mehrabi, Z., Taheri-Kafrani, A., Asadnia, M. and Razmjou, A. (2020) ‘Bienzymatic modification of polymeric membranes to mitigate biofouling’, *Separation and Purification Technology*. Elsevier B.V., 237, p. 116464.
- Melaiye, A. and Youngs, W. J. (2005) ‘Silver and its application as an antimicrobial agent’, *Expert Opinion on Therapeutic Patents*, 15(2), pp. 125–130.
- Merino, D., Tomadoni, B., Salcedo, M. F., Mansilla, A. Y., Casalongué, C. A. and Alvarez, V. A. (2020) ‘Nanoclay as Carriers of Bioactive Molecules Applied to Agriculture’, *Handbook of Nanomaterials and Nanocomposites for Energy and Environmental Applications*, pp. 1–22.
- de Messano, L. V. R., Sathler, L., Reznik, L. Y. and Coutinho, R. (2009) ‘The effect of biofouling on localized corrosion of the stainless steels N08904 and UNS S32760’, *International Biodeterioration and Biodegradation*. Elsevier Ltd,

- 63(5), pp. 607–614.
- Mikhailova, E. O. (2020) ‘Silver Nanoparticles: Mechanism of Action and Probable Bio-Application’, *Journal of Functional Biomaterials*, 11(4), p. 84.
- Miller, R. J., Adeleye, A. S., Page, H. M., Kui, L., Lenihan, H. S. and Keller, A. A. (2020) ‘Nano and traditional copper and zinc antifouling coatings: metal release and impact on marine sessile invertebrate communities’, *Journal of Nanoparticle Research. Journal of Nanoparticle Research*, 22(5).
- Mintova, S., Jaber, M. and Valtchev, V. (2015) ‘Nanosized microporous crystals: emerging applications’, *Chemical Society Reviews*. Royal Society of Chemistry, 44(20), pp. 7207–7233.
- Miranda-Trevino, J. C. and Coles, C. a. (2003) ‘Kaolinite properties, structure and influence of metal retention on pH’, *Applied Clay Science*, 23(1–4), pp. 133–139.
- Mitchell, D. J., Mecholsky, J. J. and Adair, J. H. (2000) ‘All-steel and Si₃N₄-steel hybrid rolling contact fatigue under contaminated conditions’, *Wear*, 239(2), pp. 176–188.
- Mitik-Dineva, N., Wang, J., Mocanasu, R. C., Stoddart, P. R., Crawford, R. J. and Ivanova, E. P. (2008) ‘Impact of nano-topography on bacterial attachment’, *Biotechnology Journal*, 3(4), pp. 536–544.
- Molino, P. J., Campbell, E. and Wetherbee, R. (2009) ‘Development of the initial diatom microfouling layer on antifouling and fouling-release surfaces in temperate and tropical Australia.’, *Biofouling*, 25(8), pp. 685–694.
- Moosa, S., Mohd Faisol Mahadeven, A. N. and Shameli, K. (2021) ‘Physicochemical synthesis of Silver/Kaolinite nanocomposites and study their antibacterial properties’, *Journal of Research in Nanoscience and Nanotechnology*, 1(1), pp. 1–11.
- Moreira, J. M. R., Teodósio, J. S., Silva, F. C., Simões, M., Melo, L. F. and Mergulhão, F. J. (2013) ‘Influence of flow rate variation on the development of Escherichia coli biofilms’, *Bioprocess and Biosystems Engineering*, 36(11), pp. 1787–1796.
- Morikawa, M., Kagihiro, S., Haruki, M., Takano, K., Branda, S., Kolter, R. and Kanaya, S. (2006) ‘Biofilm formation by a *Bacillus subtilis* strain that produces γ-polyglutamate’, *Microbiology*, 152(9), pp. 2801–2807.
- Morrison, K. D., Misra, R. and Williams, L. B. (2016) ‘Unearthing the Antibacterial

- Mechanism of Medicinal Clay: A Geochemical Approach to Combating Antibiotic Resistance', *Scientific Reports* 6, 19043, pp. 1–13.
- Morsy, F. A., El-Sherbiny, S., Samir, M. and Fouad, O. A. (2016) 'Application of nanostructured titanium dioxide pigments in paper coating: a comparison between prepared and commercially available ones', *Journal of Coatings Technology and Research*. Springer US, 13(2), pp. 307–316.
- Motshekga, S. C., Ray, S. S., Onyango, M. S. and Momba, M. N. B. (2015) 'Preparation and antibacterial activity of chitosan-based nanocomposites containing bentonite-supported silver and zinc oxide nanoparticles for water disinfection', *Applied Clay Science*. Elsevier B.V., 114, pp. 330–339.
- Mounier, J., Gelsomino, R., Goerges, S., Vancanneyt, M., Vandemeulebroecke, K., Hoste, B., Scherer, S., Swings, J., Fitzgerald, G. F. and Cogan, T. M. (2005) 'Surface microflora of four smear-ripened cheeses', *Applied and Environmental Microbiology*, 71(11), pp. 6489–6500.
- Muhammad, M. H., Idris, A. L., Fan, X., Guo, Y., Yu, Y., Jin, X., Qiu, J., Guan, X. and Huang, T. (2020) 'Beyond Risk: Bacterial Biofilms and Their Regulating Approaches', *Frontiers in Microbiology*, 11(May), pp. 1–20.
- Müller, W. E. G., Wang, X., Proksch, P., Perry, C. C., Osinga, R., Gardères, J. and Schröder, H. C. (2013) 'Principles of Biofouling Protection in Marine Sponges: A Model for the Design of Novel Biomimetic and Bio-inspired Coatings in the Marine Environment?', *Marine Biotechnology*, 15(4), pp. 375–398.
- Muras, A., Parga, A., Mayer, C. and Otero, A. (2021) 'Use of Quorum Sensing Inhibition Strategies to Control Microfouling', *Marine drugs*, 19(2).
- Murray, H. (2002) 'Industrial clays case study', *Mining, Minerals and Sustainable Development*, 1(64), pp. 1–9.
- Murray, H. H. (2006). Applied clay mineralogy: occurrences, processing and applications of kaolins, bentonites, palygorskitesepiolite, and common clays (Vol. 2): Elsevier , pp. 85–109.
- Mustapha, S., Tijani, J. O., Ndamitso, M. M., Abdulkareem, S. A., Shuaib, D. T., Mohammed, A. K. and Sumaila, A. (2020) 'The role of kaolin and kaolin/ZnO nanoadsorbents in adsorption studies for tannery wastewater treatment', *Scientific Reports*, 10(1), pp. 1–22.
- Narsing Rao, M. P., Xiao, M. and Li, W. J. (2017) 'Fungal and bacterial pigments:

- Secondary metabolites with wide applications', *Frontiers in Microbiology*, 8(Jun), pp. 1–13.
- Naumann, D. (2000) 'Infrared Spectroscopy in Microbiology', *Encyclopedia of Analytical Chemistry*, pp. 102–131.
- Neira, C., Mendoza, G., Levin, L. A., Zirino, A., Delgadillo-Hinojosa, F., Porrachia, M. and Deheyn, D. D. (2011) 'Macrofaunal community response to copper in Shelter Island Yacht Basin, San Diego Bay, California', *Marine Pollution Bulletin*. Pergamon, 62(4), pp. 701–717.
- Nešporová, K., Pavlík, V., Šafránková, B., Vágnerová, H., Odráška, P., Žídek, O., Císařová, N., Skoroplyas, S., Kubala, L. and Velebný, V. (2020) 'Effects of wound dressings containing silver on skin and immune cells', *Scientific Reports*. Nature Publishing Group UK, 10(1), pp. 1–14.
- Newberry, C. J., Webster, G., Cragg, B. A., Parkes, R. J., Weightman, A. J. and Fry, J. C. (2004) 'Diversity of prokaryotes and methanogenesis in deep subsurface sediments from the Nankai Trough, Ocean Drilling Program Leg 190', *Environmental Microbiology*, 6(3), pp. 274–287.
- Ni, B., Colin, R., Link, H., Endres, R. G. and Sourjik, V. (2020) 'Growth-rate dependent resource investment in bacterial motile behavior quantitatively follows potential benefit of chemotaxis', *Proceedings of the National Academy of Sciences of the United States of America*, 117(1), pp. 595–601.
- Nicholas A. Lyons and Roberto Kolter (2015) 'On The Evolution of Bacterial Multicellularity Nicholas', *Proceedings of the Royal Society of London*, 33(4), pp. 395–401.
- Nocker, A., Lepo, J. E. and Snyder, R. A. (2004) 'Influence of an oyster reef on development of the microbial heterotrophic community of an estuarine biofilm', *Applied and Environmental Microbiology*, 70(11), pp. 6834–6845.
- Le Norcy, T., Niemann, H., Proksch, P., Linossier, I., Vallée-Réhel, K., Hellio, C. and Faÿ, F. (2017) 'Anti-biofilm effect of biodegradable coatings based on hemibastadin derivative in marine environment', *International Journal of Molecular Sciences*, 18(7).
- Nur Aryantie, W., Zulhilmi Amir Awaluddin, M. and Ahmad Nizam Nik Malek, N. (2019) 'Characterization and Antibacterial Activity of Streptomycin Antibiotic Loaded Organo-Kaolinite', *IOP Conference Series: Earth and Environmental Science*, 276(1).

- Nurioglu, A. G., Esteves, A. C. C. and de With, G. (2015) ‘Non-toxic, non-biocide-release antifouling coatings based on molecular structure design for marine applications’, *Journal of Materials Chemistry B*. Royal Society of Chemistry, 3(32), pp. 6547–6570.
- Nwosu, I. G., Abu, G. O. and Agwa, K. O. (2019) ‘Isolation, Screening and Characterization of Exopolysaccharide Producing Bacteria’, *Microbiology Research Journal International*, 29(5), pp. 1–9.
- O’Leary, N. A., Wright, M. W., Brister, J. R., Ciufo, S., Haddad, D., McVeigh, R., Rajput, B., Robbertse, B., Smith-White, B., Ako-Adjei, D., Astashyn, A., Badretdin, A., Bao, Y., Blinkova, O., Brover, V., Chetvernin, V., Choi, J., Cox, E., Ermolaeva, O., Farrell, C. M., Goldfarb, T., Gupta, T., Haft, D., Hatcher, E., Hlavina, W., Joardar, V. S., Kodali, V. K., Li, W., Maglott, D., Masterson, P., McGarvey, K. M., Murphy, M. R., O’Neill, K., Pujar, S., Rangwala, S. H., Rausch, D., Riddick, L. D., Schoch, C., Shkeda, A., Storz, S. S., Sun, H., Thibaud-Nissen, F., Tolstoy, I., Tully, R. E., Vatsan, A. R., Wallin, C., Webb, D., Wu, W., Landrum, M. J., Kimchi, A., Tatusova, T., DiCuccio, M., Kitts, P., Murphy, T. D. and Pruitt, K. D. (2016) ‘Reference sequence (RefSeq) database at NCBI: Current status, taxonomic expansion, and functional annotation’, *Nucleic Acids Research*, 44(D1), pp. D733–D745.
- O’Toole, G. A. (2010) ‘Microtiter dish Biofilm formation assay’, *Journal of Visualized Experiments*, (47), pp. 10–11.
- O’Toole, G. A. and Kolter, R. (1998) ‘Flagellar and twitching motility are necessary for *Pseudomonas aeruginosa* biofilm development’, *Molecular Microbiology*, 30(2), pp. 295–304.
- O, A. U. and A, E.-N. U. (2008) ‘International Journal of Emerging Technology and Advanced Engineering XRF, XRD and FTIR Properties and Characterization of HDTMA-Br Surface Modified Organo-Kaolinite Clay’, *Certified Journal*, 9001(4), pp. 817–825.
- Oates, A., Bowling, F. L., Boulton, A. J. M., Bowler, P. G., Metcalf, D. G. and McBain, A. J. (2014) ‘The visualization of biofilms in chronic diabetic foot wounds using routine diagnostic microscopy methods’, *Journal of Diabetes Research*, 2014.
- Obaje, S. O., Omada, J. I. and Dambatta, U. A. (2013) ‘Clays and their Industrial Applications: Synoptic Review’, *International Journal of Science and*

Technology, 3(5).

- Ogunsona, E. O., Muthuraj, R., Ojogbo, E., Valerio, O. and Mekonnen, T. H. (2020) ‘Engineered nanomaterials for antimicrobial applications: A review’, *Applied Materials Today*. Elsevier, 18, p. 100473.
- Ojaveer, H., Galil, B. S., Carlton, J. T., Alleway, H., Gouletquer, P., Lehtiniemi, M., Marchini, A., Miller, W., Occhipinti-Ambrogi, A., Peharda, M., Ruiz, G. M., Williams, S. L. and Zaiko, A. (2018) *Historical baselines in marine bioinvasions: Implications for policy and management*, *PLoS ONE*.
- Olivier, F., Tremblay, R., Bourget, E. and Rittschof, D. (2000) ‘Barnacle settlement: Field experiments on the influence of larval supply, tidal level, biofilm quality and age on *Balanus amphitrite cyprids*’, *Marine Ecology Progress Series*, 199, pp. 185–204.
- Olson, M. E., Ceri, H., Morck, D. W., Buret, A. G. and Read, R. R. (2002) ‘Biofilm bacteria: Formation and comparative susceptibility to antibiotics’, *Canadian Journal of Veterinary Research*, 66(2), pp. 86–92.
- Ordax, M., Marco-Noales, E., López, M. M. and Biosca, E. G. (2010) ‘Exopolysaccharides favor the survival of *Erwinia amylovora* under copper stress through different strategies’, *Research in Microbiology*, 161(7), pp. 549–555.
- Orsod, M., Joseph, M. and Huyop, F. (2012) ‘Characterization of exopolysaccharides produced by *Bacillus cereus* and *Brachybacterium* sp. isolated from Asian sea bass (*Lates calcarifer*)’, *Malaysian Journal of Microbiology*, 8(3), pp. 170–174.
- Ortega-Morales, B. O., Santiago-García, J. L., Chan-Bacab, M. J., Moppert, X., Miranda-Tello, E., Fardeau, M. L., Carrero, J. C., Bartolo-Pérez, P., Valadéz-González, A. and Guezennec, J. (2007) ‘Characterization of extracellular polymers synthesized by tropical intertidal biofilm bacteria’, *Journal of Applied Microbiology*, 102(1), pp. 254–264.
- Osman, M. E., El-shouny, W., Talat, R. and El-zahaby, H. (2012) ‘Polysaccharides Production From Some *Pseudomonas syringae*’, *Journal of microbioloy, biotechnology and food science*, 1(5), pp. 1305–1318.
- Özdemir, G., Limoncu, M. H. and Yapar, S. (2010) ‘The antibacterial effect of heavy metal and cetylpyridinium-exchanged montmorillonites’, *Applied Clay Science*, 48(3), pp. 319–323.

- Pandey, A., Naik, M. and Dubey, S. K. (2010) 'Hemolysin, Protease, and EPS Producing Pathogenic *Aeromonas hydrophila* Strain An4 Shows Antibacterial Activity against Marine Bacterial Fish Pathogens', *Journal of Marine Biology*, 2010, pp. 1–9.
- Pang, C. M., Hong, P., Guo, H. and Liu, W. T. (2005) 'Biofilm formation characteristics of bacterial isolates retrieved from a reverse osmosis membrane', *Environmental Science and Technology*, 39(19), pp. 7541–7550.
- Park, M. B., Ahn, S. H., Nicholas, C. P., Lewis, G. J. and Hong, S. B. (2017) 'Charge density mismatch synthesis of zeolite beta in the presence of tetraethylammonium, tetramethylammonium, and sodium ions: Influence of tetraethylammonium decomposition', *Microporous and Mesoporous Materials*. Elsevier Ltd, 240, pp. 159–168.
- Parkar, D., Jadhav, R. and Pimpliskar, M. (2017) 'Marine bacterial extracellular polysaccharides: A review', *Journal of Coastal Life Medicine*, 5(1), pp. 29–35.
- Parretti, P., Canning-Clode, J., Ferrario, J., Marchini, A., Botelho, A. Z., Ramalhosa, P. and Costa, A. C. (2020) 'Free rides to diving sites: the risk of marine non-indigenous species dispersal', *Ocean and Coastal Management*, 190(March).
- Paula, A. J., Hwang, G. and Koo, H. (2020) 'Dynamics of bacterial population growth in biofilms resemble spatial and structural aspects of urbanization', *Nature Communications*. Springer US, 11(1), pp. 1–14.
- Percival, S. L., Hill, K. E., Malic, S., Thomas, D. W. and Williams, D. W. (2011) 'Antimicrobial tolerance and the significance of persister cells in recalcitrant chronic wound biofilms', *Wound Repair and Regeneration*, 19(1), pp. 1–9.
- Pereni, C. I., Zhao, Q., Liu, Y. and Abel, E. (2006) 'Surface free energy effect on bacterial retention', *Colloids and Surfaces B: Biointerfaces*, 48(2), pp. 143–147.
- Pinto, R. M., Soares, F. A., Reis, S., Nunes, C. and Van Dijck, P. (2020) 'Innovative Strategies Toward the Disassembly of the EPS Matrix in Bacterial Biofilms', *Frontiers in Microbiology*, 11(May), pp. 1–20.
- Plascencia-Jatomea, R., Almazán-Ruiz, F. J., Gómez, J., Rivero, E. P., Monroy, O. and González, I. (2015) 'Hydrodynamic study of a novel membrane aerated biofilm reactor (MABR): Tracer experiments and CFD simulation', *Chemical Engineering Science*. Elsevier, 138, pp. 324–332.
- Pochon, X., Zaiko, A., Hopkins, G. A., Banks, J. C. and Wood, S. A. (2015) 'Early

- detection of eukaryotic communities from marine biofilm using high-throughput sequencing: an assessment of different sampling devices', *Biofouling*, 31(3), pp. 241–251.
- Pollet, T., Berdjeeb, L., Garnier, C., Durrieu, G., Le Poupon, C., Misson, B. and Jean-François, B. (2018) 'Prokaryotic community successions and interactions in marine biofilms: the key role of Flavobacteriia', *FEMS microbiology ecology*, 94(6), pp. 1–13.
- Popescu, A. and Doyle, R. J. (1996) 'The gram stain after more than a century', *Biotechnic and Histochemistry*, 71(3), pp. 145–151.
- Potter, P. M., Navratilova, J., Rogers, K. R. and Al-Abed, S. R. (2019) 'Transformation of silver nanoparticle consumer products during simulated usage and disposal', *Environmental Science: Nano*. Royal Society of Chemistry, 6(2), pp. 592–598.
- Pour, N. K., Dusane, D. H., Dhakephalkar, P. K., Zamin, F. R., Zinjarde, S. S. and Chopade, B. A. (2011) 'Biofilm formation by *Acinetobacter baumannii* strains isolated from urinary tract infection and urinary catheters', *FEMS Immunology and Medical Microbiology*, 62(3), pp. 328–338.
- Prabhu, S. and Poulose, E. K. (2012) 'Silver nanoparticles: mechanism of antimicrobial', *Int. Nano Lett.*, 2, pp. 32–41.
- Prasad, M. S., Reid, K. J. and Murray, H. H. (1991) 'Kaolin: processing, properties and applications', *Applied Clay Science*, 6(2), pp. 87–119.
- Preedy, E., Perni, S., Nipić, D., Bohinc, K. and Prokopovich, P. (2014) 'Surface roughness mediated adhesion forces between borosilicate glass and gram-positive bacteria', *Langmuir*, 30(31), pp. 9466–9476.
- Prokopovich, P. and Starov, V. (2011) 'Adhesion models: From single to multiple asperity contacts', *Advances in Colloid and Interface Science*. Elsevier B.V., 168(1–2), pp. 210–222.
- Ptáček, P., Frajkorová, F., Šoukal, F. and Opravil, T. (2014) 'Kinetics and mechanism of three stages of thermal transformation of kaolinite to metakaolinite', *Powder Technology*. Elsevier B.V., 264, pp. 439–445.
- Radix, P., Léonard, M., Papantoniou, C., Roman, G., Saouter, E., Gallotti-Schmitt, S., Thiébaud, H. and Vasseur, P. (2000) 'Comparison of Four Chronic Toxicity Tests Using Algae, Bacteria, and Invertebrates Assessed with Sixteen Chemicals', *Ecotoxicology and Environmental Safety*, 47(2), pp. 186–194.

- Rajasekar, A., Maruthamuthu, S., Palaniswamy, N. and Rajendran, A. (2007) ‘Biodegradation of corrosion inhibitors and their influence on petroleum product pipeline’, *Microbiological Research*, 162(4), pp. 355–368.
- Rampadarath, S., Bandhoa, K., Puchooa, D., Jeewon, R. and Bal, S. (2017) ‘Early bacterial biofilm colonizers in the coastal waters of Mauritius’, *Electronic Journal of Biotechnology*. Elsevier España, S.L.U., 29, pp. 13–21.
- Rees, C. M., Brady, B. A. and Fabris, G. J. (2001) ‘Incidence of impossex in *Thais orbita* from Port Phillip Bay (Victoria, Australia), following 10 years of regulation on use of TBT’, *Marine Pollution Bulletin*, 42(10), pp. 873–878.
- Reichhardt, C. and Parsek, M. R. (2019) ‘Confocal laser scanning microscopy for analysis of *Pseudomonas aeruginosa* biofilm architecture and matrix localization’, *Frontiers in Microbiology*, 10(APR).
- Renner, L. D. and Weibel, D. B. (2011) ‘Physicochemical regulation of biofilm formation’, *MRS Bulletin*, 36(5), pp. 347–355.
- Renner, L. D. and Weibel, D. B. (no date) ‘Physicochemical regulation of biofilm formation’.
- Rocke, E., Cheung, S., Gebe, Z., Dames, N. R., Liu, H. and Moloney, C. L. (2020) ‘Marine Microbial Community Composition During the Upwelling Season in the Southern Benguela’, *Frontiers in Marine Science*, 7(April), pp. 1–17.
- Rodríguez-Calvo, A., Silva-Castro, G. A., Uad, I., Robledo-Mahón, T., Menéndez, M., González-López, J. and Calvo, C. (2017) ‘A comparative study of adhesion by bacterial isolates of marine origin’, *International Biodegradation and Biodegradation*. Elsevier Ltd, 123, pp. 87–95.
- Rogers, K. L., Carreres-Calabuig, J. A., Gorokhova, E. and Posth, N. R. (2020) ‘Special Issue-Current Evidence Micro-by-micro interactions: How microorganisms influence the fate of marine microplastics’, *Limnology and Oceanography Letters*, 5, pp. 18–36.
- Romano, G., Costantini, M., Sansone, C., Lauritano, C., Ruocco, N. and Ianora, A. (2017) ‘Marine microorganisms as a promising and sustainable source of bioactive molecules’, *Marine Environmental Research*. Elsevier Ltd, 128, pp. 58–69.
- Roy, R., Tiwari, M., Donelli, G. and Tiwari, V. (2018) ‘Strategies for combating bacterial biofilms: A focus on anti-biofilm agents and their mechanisms of action’, *Virulence*. Taylor & Francis, 9(1), pp. 522–554.

- Ruan, X., Li, L. and Liu, J. (2013) ‘Flocculating characteristic of activated sludge flocs: Interaction between Al³⁺ and extracellular polymeric substances’, *Journal of Environmental Sciences (China)*. The Research Centre for Eco-Environmental Sciences, Chinese Academy of Sciences, 25(5), pp. 916–924.
- Ruiz-Linares, M., Baca, P., Arias-Moliz, M. T., Ternero, F. J., Rodríguez, J. and Ferrer-Luque, C. M. (2019) ‘Antibacterial and antibiofilm activity over time of guttaflow bioseal and AH plus’, *Dental Materials Journal*, 38(5), pp. 701–706.
- Rumampuk, N. D., Rumengan, I. F., Rompas, R. M., Undap, S. L., Boneka, F. B., Jensen, K. R. and Lasut, M. T. (2018) ‘Tributyltin (TBT) contamination and impacts on imposex in *Thalessa aculeata* (mollusca: Neogastropoda: muricidae) in Minahasa Peninsula coastal waters, North Sulawesi, Indonesia’, *AACL Bioflux*, 11(1), pp. 184–193.
- Sabater, S., Guasch, H., Ricart, M., Romaní, A., Vidal, G., Klünder, C. and Schmitt-Jansen, M. (2007) ‘Monitoring the effect of chemicals on biological communities. the biofilm as an interface’, *Analytical and Bioanalytical Chemistry*, 387(4), pp. 1425–1434.
- Sadekuzzaman, M., Yang, S., Mizan, M. F. R. and Ha, S. D. (2015) ‘Current and Recent Advanced Strategies for Combating Biofilms’, *Comprehensive Reviews in Food Science and Food Safety*, 14(4), pp. 491–509.
- Saha, A., Nir, S. and Reches, M. (2020) ‘Amphiphilic Peptide with Dual Functionality Resists Biofouling’, *Langmuir*, 36(15), pp. 4201–4206.
- Salazar, G. and Sunagawa, S. (2017) ‘Marine microbial diversity’, *Current Biology*. Elsevier, 27(11), pp. R489–R494.
- Salim, M. M. and Malek, N. A. N. N. (2017) ‘Review of modified Zeolites by surfactant and Silver as antibacterial agents’, *Journal of Advanced Research in Materials Science*, 36(1), pp. 1–20.
- Salta, M., Wharton, J. A., Blache, Y., Stokes, K. R. and Briand, J.F. (2013) ‘Marine biofilms on artificial surfaces: structure and dynamics’, *Environmental Microbiology*, 15(11), p. n/a-n/a.
- Salta, M., Wharton, J. A., Blache, Y., Stokes, K. R. and Briand, J. F. (2013) ‘Marine biofilms on artificial surfaces: Structure and dynamics’, *Environmental Microbiology*, 15(11), pp. 2879–2893.
- Salta, M., Wharton, J. A., Stoodley, P., Dennington, S. P., Goodes, L. R., Werwinski,

- S., Mart, U., Wood, R. J. K. and Stokes, K. R. (2010) ‘Designing biomimetic antifouling surfaces’, *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 368(1929), pp. 4729–4754.
- Sanli, K., Bengtsson-Palme, J., Henrik Nilsson, R., Kristiansson, E., Rosenblad, M. A., Blanck, H. and Eriksson, K. M. (2015) ‘Metagenomic sequencing of marine periphyton: Taxonomic and functional insights into biofilm communities’, *Frontiers in Microbiology*, 6(OCT), pp. 1–14.
- Saur, T., Morin, E., Habouzit, F., Bernet, N. and Escudié, R. (2017) ‘Impact of wall shear stress on initial bacterial adhesion in rotating annular reactor’, *PLoS ONE*, 12(2), pp. 1–19.
- Selim, M. S., El-Safty, S. A., Shenashen, M. A., Higazy, S. A. and Elmarakbi, A. (2020) ‘Progress in biomimetic leverages for marine antifouling using nanocomposite coatings’, *Journal of Materials Chemistry B*, 8(17), pp. 3701–3732.
- Selim, M. S., Shenashen, M. A., El-Safty, S. A., Higazy, S. A., Isago, H. and Elmarakbi, A. (2017) ‘Recent Progress in Marine Foul-Release Polymeric Nanocomposite Coatings’, *Progress in Materials Science*. Elsevier Ltd, 87, pp. 1–32.
- Semeniuk, D. M., Bundy, R. M., Payne, C. D., Barbeau, K. A. and Maldonado, M. T. (2015) ‘Acquisition of organically complexed copper by marine phytoplankton and bacteria in the northeast subarctic Pacific Ocean’, *Marine Chemistry*. Elsevier B.V., 173, pp. 222–233.
- Serri, C., De Gennaro, B., Catalanotti, L., Cappelletti, P., Langella, A., Mercurio, M., Mayol, L. and Biondi, M. (2016) ‘Surfactant-modified phillipsite and chabazite as novel excipients for pharmaceutical applications?’, *Microporous and Mesoporous Materials*. Elsevier Ltd, 224, pp. 143–148.
- Shackleton, R. T., Shackleton, C. M. and Kull, C. A. (2019) ‘The role of invasive alien species in shaping local livelihoods and human well-being: A review’, *Journal of Environmental Management*. Elsevier, 229(October 2017), pp. 145–157.
- Shahariar, H., Kim, I., Soewardiman, H. and Jur, J. S. (2019) ‘Inkjet Printing of Reactive Silver Ink on Textiles’, *ACS Applied Materials and Interfaces*. American Chemical Society, 11, pp. 6208–6216.

- Sheng, X. X., Ting, Y. P. and Pehkonen, S. O. (2008) ‘The influence of ionic strength, nutrients and pH on bacterial adhesion to metals’, *Journal of Colloid and Interface Science*, 321(2), pp. 256–264.
- Shevalkar, M., Mishra, A. and Meenambiga, S. S. (2020) ‘A review on invasive species in marine biofouling’, *Research Journal of Pharmacy and Technology*, 13(9), pp. 4517–4521.
- Shobana, C., Rangasamy, B., Hemalatha, D. and Ramesh, M. (2021) ‘Bioaccumulation of silver and its effects on biochemical parameters and histological alterations in an Indian major carp *Labeo rohita*’, *Environmental Chemistry and Ecotoxicology*. Elsevier B.V., 3, pp. 51–58.
- Shukla, P. J. and Dave, B. P. (2018) ‘Screening And Molecular Identification Of Potential Exopolysaccharides (EPSSs) Producing Marine Bacteria From The Bhavnagar Coast, Gujarat’, *International Journal of Pharmaceutical Sciences and Research*, 9(7), pp. 2973–2981.
- Siddiqui, S., Goddard, R. H. and Bielmyer-Fraser, G. K. (2015) ‘Comparative effects of dissolved copper and copper oxide nanoparticle exposure to the sea anemone, *Exaiptasia pallida*’, *Aquatic Toxicology*. Elsevier B.V., 160, pp. 205–213.
- Sim, S. T. V., Suwarno, S. R., Chong, T. H., Krantz, W. B. and Fane, A. G. (2013) ‘Monitoring membrane biofouling via ultrasonic time-domain reflectometry enhanced by silica dosing’, *Journal of Membrane Science*. Elsevier, 428, pp. 24–37.
- Sim, W., Barnard, R. T., Blaskovich, M. A. T. and Ziora, Z. M. (2018) ‘Antimicrobial silver in medicinal and consumer applications: A patent review of the past decade (2007–2017)’, *Antibiotics*, 7(4), pp. 1–15.
- Simbine, E. O., Rodrigues, L. da C., Lapa-Guimarães, J., Kamimura, E. S., Corassin, C. H. and de Oliveira, C. A. F. (2019) ‘Application of silver nanoparticles in food packages: A review’, *Food Science and Technology*, 39(4), pp. 793–802.
- Simões, L. C., Simões, M. and Vieira, M. J. (2010) ‘Adhesion and biofilm formation on polystyrene by drinking water-isolated bacteria’, *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, 98(3), pp. 317–329.
- Simões, M., Simões, L. C. and Vieira, M. J. (2010) ‘A review of current and emergent

- biofilm control strategies', *Food Science and Technology*, 43(4), pp. 573–583.
- Sinde, E. and Carballo, J. (2000) 'Attachment of *Salmonella* spp. and *Listeria monocytogenes* to stainless steel, rubber and polytetrafluorehtylene: The influence of free energy and the effect of commercial sanitizers', *Food Microbiology*, 17(4), pp. 439–447.
- Singh, A., Dhiman, N., Kar, A. K., Singh, D., Purohit, M. P., Ghosh, D. and Patnaik, S. (2020) 'Advances in controlled release pesticide formulations: Prospects to safer integrated pest management and sustainable agriculture', *Journal of Hazardous Materials*. Elsevier B.V., 385, p. 121525.
- Song, K., Lee, J., Choi, S. O. and Kim, J. (2019) 'Interaction of surface energy components between solid and liquid on wettability', *Polymers*, 11(3).
- Song, Y., Zhang, S., Zeng, Y., Zhu, J., Du, X., Cai, Z. and Zhou, J. (2020) 'The rhodamine isothiocyanate analogue as a quorum sensing inhibitor has the potential to control microbially-induced biofouling', *Marine Drugs*, 18(9).
- Sousa, A. M., Machado, I., Nicolau, A. and Pereira, M. O. (2013) 'Improvements on colony morphology identification towards bacterial profiling', *Journal of Microbiological Methods*, 95(3), pp. 327–335.
- Spooner, N., Gibbs, P. E., Bryan, G. W. and Goad, L. J. (1991) 'The effect of tributyltin upon steroid titres in the female dogwhelk, *Nucella lapillus*, and the development of imposex', *Marine Environmental Research*, 32(1–4), pp. 37–49.
- Srey, S., Jahid, I. K. and Ha, S. Do (2013) 'Biofilm formation in food industries: A food safety concern', *Food Control*. Elsevier Ltd, 31(2), pp. 572–585.
- Srinivasan, M. and Swain, G. W. (2007) 'Managing the use of copper-based antifouling paints', *Environmental Management*, 39(3), pp. 423–441.
- Stackebrandt, E. and Jonas, E. (2006) 'Taxonomic parameters revisited: tarnished gold standards', *Microbiology Today*, 33, pp. 152–155.
- Stallard, C. P., McDonnell, K. A., Onayemi, O. D., O'Gara, J. P. and Dowling, D. P. (2012) 'Evaluation of protein adsorption on atmospheric plasma deposited coatings exhibiting superhydrophilic to superhydrophobic properties', *Biointerphases*, 7(1–4), pp. 1–12.
- Stepanović, S., Vuković, D., Dakić, I., Savić, B. and Švabić-Vlahović, M. (2000) 'A modified microtiter-plate test for quantification of staphylococcal biofilm

- formation', *Journal of Microbiological Methods*, 40(2), pp. 175–179.
- Stepanović, S., Vuković, D., Hola, V., Bonaventura, G. Di, Djukić, S., Ćirković, I. and Ruzicka, F. (2007) 'Quantification of biofilm in microtiter plates: overview of testing conditions and practical recommendations for assessment of biofilm production by Staphylococci', *APMIS*, 115(8), pp. 891–899.
- Stoodley, P., Sauer, K., Davies, D. G. and Costerton, J. W. (2002) 'Biofilms as complex differentiated communities.', *Annual review of microbiology*, 56, pp. 187–209.
- Sui, C., de Vos, P., Stapersma, D., Visser, K. and Ding, Y. (2020) 'Fuel consumption and emissions of ocean-going cargo ship with hybrid propulsion and different fuels over voyage', *Journal of Marine Science and Engineering*, 8(8).
- Sul, W. J., Oliver, T. A., Ducklow, H. W., Amaral-Zettler, L. A. and Sogin, M. L. (2013) 'Marine bacteria exhibit a bipolar distribution', *Proceedings of the National Academy of Sciences of the United States of America*, 110(6), pp. 2342–2347.
- Sun, K., Shi, Y., Wang, X. and Li, Z. (2017) 'Sorption and retention of diclofenac on zeolite in the presence of cationic surfactant', *Journal of Hazardous Materials*. Elsevier B.V., 323, pp. 584–592.
- Sun, T. and Qing, G. (2011) 'Biomimetic smart interface materials for biological applications', *Advanced Materials*, 23(12), pp. 57–77.
- Sunagawa, S., Coelho, L. P., Chaffron, S., Kultima, J. R., Labadie, K., Salazar, G., Djahanschiri, B., Zeller, G., Mende, D. R., Alberti, A., Cornejo-Castillo, F. M., Costea, P. I., Cruaud, C., D'Ovidio, F., Engelen, S., Ferrera, I., Gasol, J. M., Guidi, L., Hildebrand, F., Kokoszka, F., Lepoivre, C., Lima-Mendez, G., Poulain, J., Poulos, B. T., Royo-Llonch, M., Sarmento, H., Vieira-Silva, S., Dimier, C., Picheral, M., Searson, S., Kandels-Lewis, S., Boss, E., Follows, M., Karp-Boss, L., Krzic, U., Reynaud, E. G., Sardet, C., Sieracki, M., Velayoudon, D., Bowler, C., De Vargas, C., Gorsky, G., Grimsley, N., Hingamp, P., Iudicone, D., Jaillon, O., Not, F., Ogata, H., Pesant, S., Speich, S., Stemmann, L., Sullivan, M. B., Weissenbach, J., Wincker, P., Karsenti, E., Raes, J., Acinas, S. G. and Bork, P. (2015) 'Structure and function of the global ocean microbiome', *Science*, 348(6237), pp. 1–10.
- Sutherland, I. W. (2001) 'Biofilm exopolysaccharides: A strong and sticky framework', *Microbiology*, 147(1), pp. 3–9.

- Syafiuddin, A., Salmiati, S., Hadibarata, T., Kueh, A. B. H., Salim, M. R. and Zaini, M. A. A. (2018) ‘Silver Nanoparticles in the Water Environment in Malaysia: Inspection, characterization, removal, modeling, and future perspective’, *Scientific Reports*. Springer US, 8(1), pp. 1–15.
- Syakti, A. D., Lestari, P., Simanora, S., Sari, L. K., Lestari, F., Idris, F., Agustiadi, T., Akhlus, S., Hidayati, N. V. and Riyanti (2019) ‘Culturable hydrocarbonoclastic marine bacterial isolates from Indonesian seawater in the Lombok Strait and Indian Ocean’, *Helion*. Elsevier Ltd, 5(5), p. e01594.
- Sylvester, F. and MacIsaac, H. J. (2010) ‘Is vessel hull fouling an invasion threat to the Great Lakes?’, *Diversity and Distributions*, 16(1), pp. 132–143.
- Talapko, J., Matijević, T., Juzbašić, M., Antolović-Požgain, A. and Škrlec, I. (2020) ‘Antibacterial activity of silver and its application in dentistry, cardiology and dermatology’, *Microorganisms*, 8(9), pp. 1–13.
- Tan, D., Yuan, P., Annabi-Bergaya, F., Liu, D. and He, H. (2015) ‘Methoxy-modified kaolinite as a novel carrier for high-capacity loading and controlled-release of the herbicide amitrole’, *Scientific Reports*, 5, pp. 1–6.
- Teixeira, A. P., Dias, J. M. L., Carinhas, N., Sousa, M., Clemente, J. J., Cunha, A. E., von Stosch, M., Alves, P. M., Carrondo, M. J. T. and Oliveira, R. (2011) ‘Cell functional enviromics: Unravelling the function of environmental factors’, *BMC Systems Biology*, 5.
- De Tender, C., Devriese, L. I., Haegeman, A., Maes, S., Vangeyte, J., Catrysse, A., Dawyndt, P. and Ruttink, T. (2017) ‘Temporal Dynamics of Bacterial and Fungal Colonization on Plastic Debris in the North Sea’, *Environmental Science and Technology*, 51(13), pp. 7350–7360.
- Thallinger, B., Prasetyo, E. N., Nyanhongo, G. S. and Guebitz, G. M. (2013) ‘Antimicrobial enzymes: An emerging strategy to fight microbes and microbial biofilms’, *Biotechnology Journal*, 8(1), pp. 97–109.
- Thamilselvi, V. and Radha, K. V. (2017) ‘A Review On The Diverse Application Of Silver Nanoparticle’, *IOSR Journal of Pharmacy (IOSRPHR)*, 07(01), pp. 21–27.
- Tian, L., Yin, Y., Bing, W. and Jin, E. (2021) ‘Antifouling Technology Trends in Marine Environmental Protection’, *Journal of Bionic Engineering*, 18(2), pp. 239–263.
- Tsai, P. C. and Ding, W. H. (2004) ‘Determination of alkyltrimethylammonium

- surfactants in hair conditioners and fabric softeners by gas chromatography-mass spectrometry with electron-impact and chemical ionization', *Journal of Chromatography A*, 1027(1–2), pp. 103–108.
- Tsang, C. S. P., Ng, H. and McMillan, A. S. (2007) 'Antifungal susceptibility of *Candida albicans* biofilms on titanium discs with different surface roughness', *Clinical Oral Investigations*, 11(4), pp. 361–368.
- Tunega, D., Gerzabek, M. H. and Lischka, H. (2004) 'Ab initio molecular dynamics study of a monomolecular water layer on octahedral and tetrahedral kaolinite surfaces', *Journal of Physical Chemistry B*, 108(19), pp. 5930–5936.
- Turan, O., Demirel, Y. K., Day, S. and Tezdogan, T. (2016) 'Experimental Determination of Added Hydrodynamic Resistance Caused by Marine Biofouling on Ships', *Transportation Research Procedia*, 14, pp. 1649–1658.
- Valášková, M., Hundáková, M., Kutláková, K. M., Seidlerová, J., Čapková, P., Pazdziora, E., Matějová, K., Heřmánek, M., Klemm, V. and Rafaja, D. (2010) 'Preparation and characterization of antibacterial silver/vermiculites and silver/montmorillonites', *Geochimica et Cosmochimica Acta*, 74(22), pp. 6287–6300.
- Vandamme, P., Pot, B., Gillis, M., De Vos, P., Kersters, K. and Swings, J. (1996) 'Polyphasic taxonomy, a consensus approach to bacterial systematics', *Microbiological Reviews*, 60(2), pp. 407–438.
- Vassilev, N., Vassileva, M., Martos, V., Garcia del Moral, L. F., Kowalska, J., Tylkowski, B. and Malusá, E. (2020) 'Formulation of Microbial Inoculants by Encapsulation in Natural Polysaccharides: Focus on Beneficial Properties of Carrier Additives and Derivatives', *Frontiers in Plant Science*, 11(March), pp. 1–9.
- Velankar, B. Y. N. K. (1951) 'Bacteria in the Inshore Environment At Mandapam', pp. 96–112.
- Ventura, C., Guerin, A. J., El-Zubir, O., Ruiz-Sanchez, A. J., Dixon, L. I., Reynolds, K. J., Dale, M. L., Ferguson, J., Houlton, A., Horrocks, B. R., Clare, A. S. and Fulton, D. A. (2017) 'Marine antifouling performance of polymer coatings incorporating zwitterions', *Biofouling*. Taylor & Francis, 33(10), pp. 892–903.
- Verhoef, R., De Waard, P., Schols, H. A., Rättö, M., Siika-Aho, M. and Voragen, A. G. J. (2002) 'Structural elucidation of the EPS of slime producing

- Brevundimonas vesicularis* sp. isolated from a paper machine', *Carbohydrate Research*, 337(20), pp. 1821–1831.
- Videla, H. A. (2010) 'Biocorrosion and biofouling of metals and alloys of industrial usage. Present state of the art at the beginning of the new millennium', *Revista de Metalurgia*, 39(Extra), pp. 256–264.
- Villaseñor, M. J. and Ríos, Á. (2018) 'Nanomaterials for water cleaning and desalination, energy production, disinfection, agriculture and green chemistry', *Environmental Chemistry Letters*, 16(1), pp. 11–34.
- Villeneuve, A., Bouchez, A. and Montuelle, B. (2011) 'In situ interactions between the effects of season, current velocity and pollution on a river biofilm', *Freshwater Biology*, 56(11), pp. 2245–2259.
- Vinagre, P. A., Simas, T., Cruz, E., Pinori, E. and Svenson, J. (2020) 'Marine biofouling: A European database for the marine renewable energy sector', *Journal of Marine Science and Engineering*, 8(8).
- Viseras, C., Aguzzi, C., Cerezo, P. and Lopez-Galindo, A. (2007) 'Uses of clay minerals in semisolid health care and therapeutic products', *Applied Clay Science*, 36(1–3), pp. 37–50.
- Vizcayno, C., de Gutiérrez, R. M., Castello, R., Rodriguez, E. and Guerrero, C. E. (2010) 'Pozzolan obtained by mechanochemical and thermal treatments of kaolin', *Applied Clay Science*. Elsevier B.V., 49(4), pp. 405–413.
- Vladkova, T. G. (2018) 'Low adhesive surfaces for biofouling control', *Journal of Chemical Technology and Metallurgy*, 53(3), pp. 408–413.
- Volfson, D., Cookson, S., Hasty, J. and Tsimring, L. S. (2008) 'Biomechanical ordering of dense cell populations', *Proceedings of the National Academy of Sciences of the United States of America*, 105(40), pp. 15346–15351.
- Vu, B., Chen, M., Crawford, R. J. and Ivanova, E. P. (2009) 'Bacterial extracellular polysaccharides involved in biofilm formation', *Molecules*, 14(7), pp. 2535–2554.
- Wang, D., Liu, Y., Yang, Y. and Xiao, D. (2016) 'Theoretical and experimental study on surface roughness of 316L stainless steel metal parts obtained through selective laser melting', *Rapid Prototyping Journal*, 22(4), pp. 706–716.
- Wang, K.-L., Wu, Z.-H., Wang, Y., Wang, C.-Y. and Xu, Y. (2017) 'Mini-Review: Antifouling Natural Products from Marine Microorganisms and Their Synthetic Analogs', *Marine Drugs*. Multidisciplinary Digital Publishing

- Institute, 15(9), p. 266.
- Wang, K., Li, W., Rui, X., Chen, X., Jiang, M. and Dong, M. (2014) ‘Characterization of a novel exopolysaccharide with antitumor activity from *Lactobacillus plantarum* 70810’, *International Journal of Biological Macromolecules*. Elsevier B.V., 63, pp. 133–139.
- Wang, L., Chen, W., Song, X., Li, Y., Zhang, W., Zhang, H. and Niu, L. (2020) ‘Cultivation substrata differentiate the properties of river biofilm EPS and their binding of heavy metals: A spectroscopic insight’, *Environmental Research*. Academic Press, 182, p. 109052.
- Wang, L., Hu, C. and Shao, L. (2017) ‘The antimicrobial activity of nanoparticles present situation’, *International journal of nanomedicine*, 12, pp. 1227–1249.
- Wang, M., Wang, J., Zhou, Y., Zhang, M., Xia, Q., Bi, W. and Chen, D. D. Y. (2017) ‘Ecofriendly Mechanochemical Extraction of Bioactive Compounds from Plants with Deep Eutectic Solvents’, *ACS Sustainable Chemistry and Engineering*, 5(7), pp. 6297–6303.
- Wassmann, T., Kreis, S., Behr, M. and Buergers, R. (2017) ‘The influence of surface texture and wettability on initial bacterial adhesion on titanium and zirconium oxide dental implants’, *International Journal of Implant Dentistry*. International Journal of Implant Dentistry, 3(1).
- Webster, N. S. and Negri, A. P. (2006) ‘Site-specific variation in Antarctic marine biofilms established on artificial surfaces’, *Environmental Microbiology*, 8(7), pp. 1177–1190.
- Whitehead, K. A. and Verran, J. (2006) ‘The effect of surface topography on the retention of microorganisms’, *Food and Bioproducts Processing*, 84(4 C), pp. 253–259.
- Wilson, C., Lukowicz, R., Merchant, S., Valquier-Flynn, H., Caballero, J., Sandoval, J., Okuom, M., Huber, C., Brooks, T. D., Wilson, E., Clement, B., Wentworth, C. D. and Holmes, A. E. (2017) ‘Quantitative and Qualitative Assessment Methods for Biofilm Growth: A Mini-review.’, *Research & reviews. Journal of engineering and technology*, 6(4).
- Woinarski, A. Z., Stevens, G. W. and Snape, I. (2006) ‘A Natural Zeolite Permeable Reactive Barrier to Treat Heavy-Metal Contaminated Waters in Antarctica’, *Process Safety and Environmental Protection*, 84(2), pp. 109–116.
- Woo, K. J., Hye, C. K., Ki, W. K., Shin, S., So, H. K. and Yong, H. P. (2008)

- ‘Antibacterial activity and mechanism of action of the silver ion in *Staphylococcus aureus* and *Escherichia coli*’, *Applied and Environmental Microbiology*, 74(7), pp. 2171–2178.
- Wu, T., Xie, A. G., Tan, S. Z. and Cai, X. (2011) ‘Antimicrobial effects of quaternary phosphonium salt intercalated clay minerals on *Escherichia coli* and *Staphylococcus aureus*’, *Colloids and Surfaces B: Biointerfaces*. Elsevier B.V., 86(1), pp. 232–236.
- Xiang, Q. Q., Gao, Y., Li, Q. Q., Ling, J. and Chen, L. Q. (2020) ‘Proteomic profiling reveals the differential toxic responses of gills of common carp exposed to nanosilver and silver nitrate’, *Journal of Hazardous Materials*. Elsevier, 394(November 2019), p. 122562.
- Xochitl, D. B., Sevda, S., Vanbroekhoven, K. and Pant, D. (2012) ‘The accurate use of impedance analysis for the study of microbial electrochemical systems’, *Chemical Society Reviews*, 41(21), pp. 7228–7246.
- Yang, W. J., Neoh, K. G., Kang, E. T., Teo, S. L. M. and Rittschof, D. (2014) ‘Polymer brush coatings for combating marine biofouling’, *Progress in Polymer Science*. Elsevier Ltd, 39(5), pp. 1017–1042.
- Yebra, D. M., Kiil, S. and Dam-Johansen, K. (2004) ‘Antifouling technology - Past, present and future steps towards efficient and environmentally friendly antifouling coatings’, *Progress in Organic Coatings*, 50(2), pp. 75–104.
- Yin, W., Wang, Y., Liu, L. and He, J. (2019) ‘Biofilms: The microbial “protective clothing” in extreme environments’, *International Journal of Molecular Sciences*, 20(14).
- Young, K. D. (2006) ‘The Selective Value of Bacterial Shape’, *Microbiology and Molecular Biology Reviews*, 70(3), pp. 660–703.
- Young, K. D. (2007) ‘Bacterial morphology: why have different shapes?’, *Current Opinion in Microbiology*, 10(6), pp. 596–600.
- Yu, Q., Wu, Z. and Chen, H. (2015) ‘Acta Biomaterialia Dual-function antibacterial surfaces for biomedical applications’, *Acta Biomaterialia*. Acta Materialia Inc., (January).
- Yu, W., Li, X., He, J., Chen, Y., Qi, L., Yuan, P., Ou, K., Liu, F., Zhou, Y. and Qin, X. (2021) ‘Graphene oxide-silver nanocomposites embedded nanofiber core-spun yarns for durable antibacterial textiles’, *Journal of Colloid and Interface Science*. Elsevier Inc., 584, pp. 164–173.

- Yue, Z. B., Li, Q., Li, C. chuan, Chen, T. hu and Wang, J. (2015) ‘Component analysis and heavy metal adsorption ability of extracellular polymeric substances (EPS) from sulfate reducing bacteria’, *Bioresource Technology*. Elsevier Ltd, 194, pp. 399–402.
- Zhan, G. and Zeng, H. C. (2017) ‘Alternative synthetic approaches for metal-organic frameworks: transformation from solid matters’, *Chemical Communications*. Royal Society of Chemistry, 53(1), pp. 72–81.
- Zhang, C., Li, B., Tang, J. Y., Wang, X. L., Qin, Z. and Feng, X. Q. (2017) ‘Experimental and theoretical studies on the morphogenesis of bacterial biofilms’, *Soft Matter*, 13(40), pp. 7389–7397.
- Zhang, S., Liu, Q., Cheng, H., Gao, F., Liu, C. and Teppen, B. J. (2017) ‘Thermodynamic Mechanism and Interfacial Structure of Kaolinite Intercalation and Surface Modification by Alkane Surfactants with Neutral and Ionic Head Groups’, *Journal of Physical Chemistry C*, 121(16), pp. 8824–8831.
- Zhang, T., Chao, Y., Shih, K., Li, X. and Fang, H. H. P. (2011) ‘Ultramicroscopy Quantification of the lateral detachment force for bacterial cells using atomic force microscope and centrifugation’, *Ultramicroscopy*. Elsevier, 111(2), pp. 131–139.
- Zhang, X., Chen, X., Yang, J., Jia, H. R., Li, Y. H., Chen, Z. and Wu, F. G. (2016) ‘Quaternized Silicon Nanoparticles with Polarity-Sensitive Fluorescence for Selectively Imaging and Killing Gram-Positive Bacteria’, *Advanced Functional Materials*, 26(33), pp. 5958–5970.
- Zhang, X., Wang, L. and Levänen, E. (2013) ‘Superhydrophobic surfaces for the reduction of bacterial adhesion’, *RSC Advances*, 3(30), pp. 12003–12020.
- Zhang, Z. J., Chen, S. H., Wang, S. M. and Luo, H. Y. (2011) ‘Characterization of extracellular polymeric substances from biofilm in the process of starting-up a partial nitrification process under salt stress’, *Applied Microbiology and Biotechnology*, 89(5), pp. 1563–1571.
- Zhao, W., Deng, J., Ren, Y., Xie, L., Li, W., Wang, Q., Li, S. and Liu, S. (2021) ‘Antibacterial application and toxicity of metal-organic frameworks’, *Nanotoxicology*. Taylor and Francis, 15(3), pp. 311–330.
- Zheng, S., Bawazir, M., Dhall, A., Kim, H., He, L., Heo, J. and Hwang, G. (2021) ‘Implication of Surface Properties , Bacterial Motility , and Hydrodynamic

Conditions on Bacterial Surface Sensing and Their Initial Adhesion', 9(Feb), pp. 1–22.

Zhu, L., Chen, B. and Shen, X. (2000) 'Sorption of phenol, p-nitrophenol, and aniline to dual-cation organobentonites from water', *Environmental Science and Technology*, 34(3), pp. 468–475.

LIST OF PUBLICATION

Indexed Journal

1. **Amin, M. N. A.**, Wan Dagang, W. R. Z., Malek, N. A. N. N. and Jamaluddin, H. (2021) ‘Isolation and identification of early marine biofilm-forming bacteria on commercial paint surface’, *Malaysian Journal of Microbiology*, Vol. 7(2), pp. 130 - 142 (**Indexed by SCOPUS**).