

ANTI-DENGUE VIRAL ACTIVITY OF *Carica papaya* LEAVES EXTRACT

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A thesis submitted in fulfilment of the
requirements for the award of the degree of
Doctor of Philosophy (Bioprocess Engineering)

School of Chemical and Energy Engineering
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SEPTEMBER 2021

ACKNOWLEDGEMENT

In the name of Allah, the Most Beneficent, the Most Merciful. All the praises and thanks be to Allah, the Lord of the 'Alamin for giving me the strength, knowledge, ability and opportunity to undertake this research study and to persevere and complete it satisfactorily. Without His blessings, this achievement would not have been possible.

I would like to first, express my sincere gratitude to my supervisor Dr. Harisun Yaakob for the continuous support of my PhD study, for her patience, motivation and immense knowledge. Her guidance helped me in all the time of research and writing of this thesis. I could not have imagined having a better advisor and mentor for my PhD study.

Many thanks also to Prof. Dr. Mohd Roji Sarmidi and Dr. Saleha Sahar for their insightful comments and encouragement. My sincere thanks go to Assoc. Prof. Dr. Keivan Zandi from Tropical Infectious Diseases Research & Education Centre (UM) and Dr. Rafidah Hanim Shueb from Department of Medical Microbiology & Parasitology (USM) who provided me an opportunity to join their team, and who gave access to the laboratory and research facilities. Without their precious support it would not be possible to conduct this research.

A very special thank you to Fitrien for her invaluable advice and for always being so supportive of my work. My deep appreciation goes out to the research team members and others who have provided assistance at various occasions. Their excellent work during data collection has made an invaluable contribution towards my PhD.

Finally, I would like to thank my family members whose love and guidance are with me in whatever I pursue.

ABSTRACT

Dengue fever is an infectious tropical disease, which is considered a worldwide public health problem. However, until today no licensed vaccine is approved for dengue virus. The effort to develop dengue vaccine is made complicated by the four closely related dengue virus serotypes. Nowadays, the search for anti-dengue viral activities in natural plant products has been increasing. Thus, the development of plant-based antiviral agent will have the potential to fight against dengue fever. This study aimed to investigate the anti-dengue activity of *Carica papaya* leaves extracts and its fraction as well as determination of possible active compound and mechanism involved. The analyze of polysaccharide, glycosaponin and total protein were carried out by standard methods. Viability of Vero cells treated with aqueous extract of *Carica papaya* leaves was estimated by neutral red uptake assay. The microscopic observation on cytopathic effect was observed to determine the highest tolerable dose of *Carica papaya* leaves extract in Vero cells. The cytotoxicity of *Carica papaya* extracts on Vero cells was determined by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide method. Antiviral assays of aqueous extract of *Carica papaya* leaves were performed in three different stages, pre-, post- and continuous treatment on Vero cells against DENV-2 by foci forming unit reduction assay. Further antiviral assays of *Carica papaya* leaves crude extract and its fraction was investigated. Foci reduction on DENV-2 infected cells treated with the extracts for post- and continuous treatment were determined. Number of RNA reduction of DENV-2 was investigated by SYBR® green quantitative reverse transcription polymerase chain reaction method. Finally, the possible active compounds in the extracts was carried out using high performance liquid chromatography (HPLC) analysis. In this study, the results showed that the methanol matured leaves contained the highest concentration of total polysaccharide than aqueous matured leaves with 0.23 mgmL^{-1} and 0.22 mgmL^{-1} respectively. In addition, glycosaponin content in methanol matured leaves was the highest followed by methanol young leaves with 82% and 75% respectively, as well as in total protein content of 35.9% and 29.6% respectively. The extract did not reveal cytotoxic effects on Vero and C6/36 cells at all tested concentrations as well as its fractions. However, the methanol and ethanol extracts were found to be more toxic than the water extract. Anti-adsorption effect was shown at $\text{IC}_{50} = 435.3 \text{ } \mu\text{gmL}^{-1}$ with $\text{SI} = 7.6$. The extract also inhibited DENV-2 replication in Vero cells with $\text{IC}_{50} = 1413 \text{ } \mu\text{gmL}^{-1}$ and $\text{SI} = 2.3$ when added after adsorption to the cells. The IC_{50} against DENV-2 was $137.6 \text{ } \mu\text{gmL}^{-1}$ and $\text{SI} = 23.9$ when cells were treated 5 hours before virus infection and continuously up to four-day post-infection. From the HPLC analyze, the possible active compound of catechin, quercetin and cinnamic acid were found in the water extract of *Carica papaya* leaves. In conclusion, the *Carica papaya* leaves extract showed slight inhibition of DENV-2 replication by decreasing foci number and size. The aqueous extract of *Carica papaya* leaves possesses the ability of inhibiting the activity of DENV-2. Thus, this extract is worth to be further investigated and might be advantageous as a primary source in the treatment of dengue and as a potential element for drugs formulation.

ABSTRAK

Demam denggi adalah penyakit tropika berjangkit, yang dianggap sebagai masalah kesihatan umum di seluruh dunia. Walaubagaimanapun, sehingga hari ini tiada vaksin berlesen diluluskan untuk virus denggi. Usaha untuk membangunkan vaksin denggi adalah rumit disebabkan oleh empat serotip virus denggi yang berkait rapat. Pada masa kini, pencarian aktiviti anti-denggi dalam produk tumbuhan semulajadi semakin meningkat. Oleh itu, pembangunan agen antiviral berasaskan tumbuhan akan mempunyai potensi untuk melawan demam denggi. Kajian ini bertujuan untuk mengkaji aktiviti anti-denggi dari ekstrak daun betik (*Carica papaya*) dan pecahannya serta penentuan kemungkinan sebatian dan mekanisme aktif yang terlibat. Analisis polisakarida, glikosaponin dan jumlah protein dilakukan dengan kaedah piawai. Daya tahan sel Vero yang dirawat dengan ekstrak air daun *Carica papaya* dianggarkan dengan esei pengambilan neutral merah. Pemerhatian mikroskopik pada kesan sitopatik diperhatikan untuk menentukan dos tertinggi daun *Carica papaya* yang boleh ditoleransi dalam sel Vero. Sitotoksiti ekstrak *Carica papaya* pada sel Vero ditentukan oleh kaedah 3-(4,5-dimetiltiazol-2-il)-2,5-difeniltetrazolium bromida. Pemeriksaan antiviral ekstrak air daun *Carica papaya* telah dilakukan dalam tiga peringkat rawatan berlainan, sebelum, selepas dan berterusan pada sel Vero terhadap virus DENV-2 melalui esei pengurangan unit pembentukan foci. Pemeriksaan lanjut antiviral ekstrak mentah *Carica papaya* dan pecahannya disiasat. Pengurangan foci terhadap sel-sel yang dijangkiti DENV-2 yang dirawat dengan ekstrak untuk rawatan selepas dan berterusan ditentukan. Bilangan pengurangan RNA DENV-2 disiasat oleh kaedah SYBR® hijau kuantitatif transkripsi terbalik tindak balas rantai polimer. Akhirnya, kemungkinan sebatian aktif dalam ekstrak dilakukan dengan menggunakan analisis kromatografi cecair berprestasi tinggi (HPLC). Dalam kajian ini, keputusan menunjukkan bahawa daun matang metanol mengandungi kepekatan tertinggi polisakarida daripada daun matang akues masing-masing dengan 0.23 mgmL^{-1} dan 0.22 mgmL^{-1} . Di samping itu, kandungan glikosaponin dalam daun matang metanol adalah yang tertinggi diikuti oleh daun muda metanol masing-masing dengan 82% dan 75%, serta jumlah protein masing-masing sebanyak 35.9% and 29.6%. Ekstrak itu tidak mendedahkan kesan sitotoksik pada sel Vero dan C6/36 pada semua kepekatan yang diuji serta pecahannya. Walaubagaimanapun, ekstrak metanol dan etanol didapati lebih toksik daripada ekstrak air. Kesan anti-penjerapan ditunjukkan pada $IC_{50} = 435.3 \text{ } \mu\text{gmL}^{-1}$ dengan $SI = 7.6$. Ekstrak ini juga menghalang replikasi virus DENV-2 dalam sel Vero dengan $IC_{50} = 1413 \text{ } \mu\text{gmL}^{-1}$ dan $SI = 2.3$ apabila ditambahkan selepas penjerapan ke sel. IC_{50} terhadap DENV-2 adalah $137.6 \text{ } \mu\text{gmL}^{-1}$ dan $SI = 23.9$ apabila sel dirawat selama 5 jam sebelum jangkitan virus dan berterusan sehingga empat-hari selepas jangkitan. Dari analisis HPLC, sebatian aktif katekin, kuersetin dan asid sinamik yang terdapat dalam ekstrak air daun *Carica papaya*. Sebagai kesimpulan, ekstrak daun *Carica papaya* menunjukkan sedikit perencatan replikasi DENV-2 dengan mengurangkan bilangan dan saiz foci. Ekstrak air daun *Carica papaya* mempunyai keupayaan merencat aktiviti DENV-2. Oleh itu, ekstrak ini adalah bernilai untuk disiasat lebih lanjut dan mungkin berfaedah sebagai sumber utama dalam rawatan denggi dan sebagai unsur berpotensi dalam formulasi ubat.

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LIST OF ABBREVIATIONS

Ab	-	Antibody
ADE	-	Antibody Dependent Enhancement
AML	-	Aqueous Mature Leaves
AYL	-	Aqueous Young Leaves
CMC	-	Carboxymethyl Cellulose
CPE	-	Cytopathic Effect
DAB	-	3'-diaminobenzidine
DENV	-	Dengue Virus
DHF	-	Dengue Hemorrhagic Fever
DMSO	-	Dimethyl Sulfoxide
DPPH	-	2,2-diphenyl-1-picrylhydrazyl
DSS	-	Dengue Shock Syndrome
FBS	-	Fetal Bovine Serum
FFURA	-	Foci Forming Unit Reduction Assay
HPLC	-	High Performance Liquid Chromatography
HTD	-	Highest Tolerated Dose
MML	-	Methanol Mature Leaves
MNTD	-	Maximum Non-Toxic Dose
MYL	-	Methanol Young Leaves
m.o.i	-	multiplicity of infection
MTT	-	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NRU	-	Neutral Red Uptake
O.D	-	Optical Density
PBS	-	Phosphate-Buffered Saline
PR	-	Protease
P/S	-	Penicillin/Streptomycin
RNA	-	Ribonucleic Acid
RT-PCR	-	Reverse Transcription Polymerase Chain Reaction
SPE	-	Solid Phase Extraction
TPB	-	Tryptose Phosphate Broth

CHAPTER 1

INTRODUCTION

1.1 Dengue Research Background

In recent decades, dengue fever has emerged as the most important viral disease of human and become major international public health concern. More than 100 countries worldwide have been affected by dengue fever (DF) and dengue hemorrhagic fever (DHF) (Sanyaolu *et al.*, 2017). Every year, up to 400 million people are infected mainly in tropical and sub-tropical regions around the world (Dos Santos *et al.*, 2019) including Southeast Asia. America and Asia have been expected with the highest risk zones of dengue transmission due to densely populated humans and vectors and surrounding climate suitability, with Asia bears up to 70% of this burden (Bhatt *et al.*, 2013). Until date, there is no specific anti-dengue therapeutic, but self-supportive care and specialized medical care for serious illness is very effective.

However, for dengue hemorrhagic fever, if not treated properly, it has a mortality rate as high as 10-20% (Zakaria *et al.*, 2019). A single dengue vaccine, Dengvaxia®, are first licensed vaccine for the prevention of dengue disease. However, due to its dengue-specific complexities, their long-term effectiveness against all four dengue virus serotypes still have an adverse effect on production (Godói *et al.*, 2017). The clinical development pathway brings a diverse of risk to all vaccine development efforts especially which were dengue-specific (Thomas and Yoon, 2019). Furthermore, this vaccine is no longer available due to the delay in the use of Dengvaxia® after some findings on the side effects of this vaccine in humans (Zakaria *et al.*, 2019).

Dengue fever, the tropical infectious disease has been rapidly expanding over the world. Dengue cases in Malaysia has becomes seriously increasing since the first reported case in 1902. The epidemic is not limited only in urban areas, but also involve rural areas in Malaysia. The awareness of dengue fever irrespective of age, led to the search for the best way to fight this virus. The current prevents of dengue infections are only in general based on public awareness such as vector control and surveillance alert. However, periodic vector control has not been able to stop its rapid emergence and global spread. To keep the infection rates low and to prevent outbreaks, some medical treatments have been practiced such as liquid injection for reducing body temperature and to avoid prolonged fever.

There is strong need of effort towards the development of safe and effective treatment to cure dengue. However, efforts to develop antiviral agents for dengue have encountered some difficulties. It is currently unclear mechanism since the four closely related, but antigenically distinct serotypes of dengue virus often undergo mutations. Therefore, antiviral would have to be effective against all the serotypes. Until now, there is no licensed, commercially available vaccines or specific therapeutics for dengue virus. Although numerous groups have already made vaccine formulation, they are still experiencing long-term problems and are still undergoing clinical trials to develop a safe, affordable and effective vaccine against all serotypes. In addition, any other possible treatments including traditional medicines should be investigated to test their effectiveness in controlling this problem.

Since ancient time, human has used plant for medicinal properties. Indian integrative medicine such as Rigveda, Atharvaveda and Ayurveda have categorized plants for drugs, essence, food, poisons and agricultural purpose (Jain *et al.*, 2008). Research efforts to explore the potential of new agents for antivirus increased in recent years because of low toxicity and high selective antiviral substances from products of plant origin (Newman and Cragg, 2016).

Medicinal plants are widely used to prevent infectious diseases and inflammation and has been supported by a lot of clinical evidence (Borchers *et al.*, 1997). Evidence linking plants to have antiviral properties are like *Parietaria diffusa* and *Urtica dioica* against Feline immunodeficiency virus (Uncini Manganelli *et al.*, 2005) and olive leaf extract inhibit spread of HIV-1 (Lee Huang *et al.*, 2003; Bao *et al.*, 2007).

In many part of the world, the leaves, fruits and latex are widely used traditionally to treat various types of ailments like asthma, rheumatism, fever, diarrhea, boils and hypertension (Zakaria *et al.*, 2006). Based on previous studies, this species have antibacterial, antifungal, anthelmintic and antimalarial properties (Melariri *et al.*, 2011) which these properties show that the activity of *Carica papaya* leaves extract against dengue infection.

Furthermore, a human study conducted by Ahmad *et al.* (2011) and Kala (2012) showed that juice of *Carica papaya* leaves can cure dengue fever. However, the interactions mechanism of *Carica papaya* leaves extract on dengue-infected cells are still remain unknown. Therefore, this study hypothesized that *Carica papaya* leaves extract can inhibit dengue by decrease the viability and change the morphological of dengue virus by certain active compound in the extracts. To prove that the cell culture study will be conducted and the interactions of *Carica papaya* leaves extract with dengue-infected cells will be investigated.

1.2 Problem Statement

Dengue fever remains a significant public health concern in Malaysia and was established in Malaysia ever since the first reported case of dengue in 1902 (Ahmad *et al.*, 2018). Rapid urbanization, climate changes and abandoned areas results in vector breeding causes rise in dengue outbreaks. Despite reactive efforts by the government in Malaysia, dengue cases continue to increase from years to years. Unfortunately, there are no specific therapeutics agents, and no vaccine commercially against dengue.

New anti-dengue drug leads are therefore urgently needed. Traditional healers have long used plants to prevent or cure infections. It is important that anti-dengue drug development has to be pursued further, with the highly active products, to preclinical and clinical testing.

In Malaysia, *Carica papaya* leaves juice have been used traditionally in folk medicine to cure dengue fever. This is possibly due to the presence of a wide variety compounds in juice of *Carica papaya* leaves known for their antioxidant and anti-inflammatory effect (Sudhakar and Theivanai, 2014; Gupta *et al.*, 2017). However, there is limited scientific evidence for its anti-dengue activity *in-vitro*.

The underlying mechanism and responsible compounds that act as potential anti-dengue agents remains unknown. Moreover, it is not known whether each fraction of *Carica papaya* leaves extract is able to give a similar positive effect to inhibit dengue virus and which compound of *Carica papaya* leaves fractions having a high anti-dengue inhibition. Therefore, this project was carried out in order to yield a novel insight and better understanding on how *Carica papaya* leaves extract and its fraction can inhibit dengue virus and likely lead to the development of anti-dengue supplement.

1.3 Objectives of the Study

The objectives of study were:

1. To evaluate the *in vitro* inhibitory potential of *Carica papaya* leaves extract and its fraction towards dengue virus infection type-2 (DENV-2).
2. To determine the mechanism of dengue inhibitory activities of *Carica papaya* leaves extract and its fraction.
3. To determine the possible active compounds which responsible for anti-dengue properties.

1.4 Scope of Study

The scope of this research are as listed below:

1. Extraction of *Carica papaya* leaves extract using Soxhlet extraction method. Water, methanol and ethanol were used as a solvent.
2. Phytochemical screening of primary metabolites (polysaccharides and glycosaponins) in *Carica papaya* leaves extract by standard methods of Malaysian Standard MS 2409:2011.
3. Fractionation of *Carica papaya* leaves extract using solid phase extraction (SPE).
4. Evaluation of cytotoxicity profile of *Carica papaya* leaves extract on different cell lines (Vero and C6/36 cells) by 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay (MTT).
5. Determination of antiviral activities of *Carica papaya* leaves extract against dengue-infected Vero and C6/36 cell line by foci forming unit reduction assay (FFURA).
6. Detection and quantification of viral RNA in the plasma by viral RNA quantification using quantitative transcription polymerase chain reaction analysis (qRT-PCR)
7. Determination of possible anti dengue active compound in *Carica papaya* leaves extract and its fraction using HPLC analysis.

REFERENCES

- Abdelkafi, S., Barouh, N., Fouquet, B., Fendri, I., Pina, M., Scheirlinckx, F., Villeneuve, P. and Carrière, F. (2011) 'Carica papaya lipase: A naturally immobilized enzyme with interesting biochemical properties', *Plant Foods for Human Nutrition*, 66, pp. 34–40.
- Abdul Ahmad, S. A., Palanisamy, U. D., Tejo, B. A., Chew, M. F., Tham, H. W. and Syed Hassan, S. (2017) 'Geraniin extracted from the rind of *Nephelium lappaceum* binds to dengue virus type-2 envelope protein and inhibits early stage of virus replication', *Virology Journal*, 14(1), p. 229.
- Abubakar, A. R. and Haque, M. (2020) 'Preparation of medicinal plants: Basic extraction and fractionation procedures for experimental purposes', *Journal of Pharmacy & Bioallied Sciences*, 12(1), pp. 1–10.
- Afzan, A., Abdullah, N. R., Halim, S. Z., Rashid, B. A., Semail, R. H. R., Abdullah, N., Jantan, I., Muhammad, H. and Ismail, Z. (2012) 'Repeated dose 28-days oral toxicity study of *Carica papaya* L. leaf extract in Sprague Dawley rats', *Molecules*, 17, pp. 4326–4342.
- Ahmad, N., Fazal, H., Ayaz, M., Abbasi, B. H., Mohammad, I. and Fazal, L. (2011) 'Dengue fever treatment with *Carica papaya* leaves extracts.', *Asian Pacific Journal of Tropical Biomedicine*, 1(4), pp. 330–333.
- Ahmad, R., Suzilah, I., Najdah, W. M. A. W., Topek, O., Mustafakamal, I. and Lee, H. L. (2018) 'Factors determining dengue outbreak in Malaysia', *PLoS ONE*, 13(2), pp. 1–13.
- Akhila, S. and Vijayalakshmi, N. G. (2015) 'Phytochemical studies on *Carica papaya* leaf juice', *International Journal of Pharmaceutical Sciences and Research*, 6(2), pp. 880–883.
- Alm, E., Lindegren, G., Falk, K. I. and Lagerqvist, N. (2015) 'One-step real-time RT-PCR assays for serotyping dengue virus in clinical samples', *BMC Infectious Diseases*, 15, p. 493.
- Amarasinghe, A. (2011) 'Dengue virus infection in Africa', *Emerging Infectious Diseases*, 17(8), pp. 1349–1354.

- Ammerman, N. C., Magda, B. S. and Azad, A. F. (2008) 'Growth and maintenance of Vero cell lines', *Current Protocols in Microbiology*, pp. 1–10.
- Amri, E. and Mamboya, F. (2012) 'Papain, a plant enzyme of biological importance: A review', *American Journal of Biochemistry and Biotechnology*, 8(2), pp. 99–104.
- Annegowda, H. V., Bhat, R., Min-Tze, L., Karim, A. A. and Mansor, S. M. (2012) 'Influence of sonication treatments and extraction solvents on the phenolics and antioxidants in star fruits', *Journal of Food Science and Technology*. Springer-Verlag, 49(4), pp. 510–514.
- Aravind, G., Bhowmik, D., Duraivel, S. and Harish, G. (2013) 'Traditional and medicinal uses of *Carica papaya*', *Journal of Medicinal Plants Studies*, 1(1), pp. 7–15.
- Arthanari, S., Vanitha, J., Krishnaswami, V., Renukadevi, P., Deivasigamani, K. and De Clercq, E. (2013) 'In vitro antiviral and cytotoxic screening of methanolic extract of *Cassia auriculata* flowers in HeLa, Vero, CRFK and HEL cell lines', *Drug Invention Today*, 5(1), pp. 28–31.
- Aruna, R. (2014) 'Review on Dengue viral replication, assembly and entry into the host cells', *International Journal of Microbial Applied Science*, 3(11), pp. 1025–1039.
- Ayoola, B. and Adeyeye, A. (2010) 'Phytochemical and nutrient evaluation of *Carica papaya* (pawpaw) leaves', *IJRRAS*, 5.
- Ayoola, P. B. and Adeyeye, A. (2010) 'Phytochemical and nutrient evaluation of *Carica papaya* (pawpaw) leaves', *International Journal of Recent Research and Applied Studies*, 5(3), pp. 325–328.
- Azarkan, M., El Moussaoui, A., Van Wuytswinkel, D., Dehon, G. and Looze, Y. (2003) 'Fractionation and purification of the enzymes stored in the latex of *Carica papaya*', *Journal of Chromatography B*, 790, pp. 229–238.
- De Azeredo, E. L., Monteiro, R. Q. and De-Oliveira Pinto, L. M. (2015) 'Thrombocytopenia in dengue: Interrelationship between virus and the imbalance between coagulation and fibrinolysis and inflammatory mediators', *Mediators of Inflammation*, 2015(314842), pp. 1–6.

- Bao, J., Zhang, D. W., Zhang, J. Z. H., Huang, P Lee, Huang, P Lin and Huang, S. L. (2007) ‘Computational study of bindings of olive leaf extract (OLE) to HIV-1 fusion protein gp41’, *Federation of European Biochemical Societies Letters*, 581(14), pp. 2737–2742.
- Barreto-Vieira, D. F., Jácome, F. C., da Silva, M. A. N., Caldas, G. C., de Filippis, A. M. B., de Sequeira, P. C., de Souza, E. M., Andrade, A. A., Manso, P. P. de A., Trindade, G. F., Lima, S. M. B. and Barth, O. M. (2017) ‘Structural investigation of C6/36 and Vero cell cultures infected with a Brazilian Zika virus’, *PloS ONE*, 12(9), pp. 1–18.
- Barrett, P. N., Mundt, W., Kistner, O. and Howard, M. K. (2009) ‘Vero cell platform in vaccine production: Moving towards cell culture-based viral vaccines’, *Expert Review of Vaccines*, 8(5), pp. 607–618.
- Baskaran, C., Bai, V. R., Velu, S. and Kumaran, K. (2012) ‘The efficacy of Carica papaya leaf extract on some bacterial and a fungal strain by well diffusion method’, *Asian Pacific Journal of Tropical Disease*, 2(2), pp. 658–622.
- Beauté, J. and Vong, S. (2010) ‘Cost and disease burden of dengue in Cambodia’, *BMC Public Health*, 10(521), pp. 2–7.
- Betancur-Galvis, L. A., Saez, J., Granados, H., Salazar, A. and Ossa, J. E. (1999) ‘Antitumor and antiviral activity of Colombian medicinal plant extracts’, *Memórias do Instituto Oswaldo Cruz*, 94(4), pp. 531–535.
- Bhatt, S., Gething, P. W., Brady, O. J., Messina, J. P., Farlow, A. W. and Moyes, C. L. (2013) ‘The global distribution and burden of dengue’, *Nature*, 496, pp. 504–507.
- Borchers, A. T., Hackman, R. M., Keen, C. L., Stern, J. S. and Gershwin, M. E. (1997) ‘Complementary medicine: A review of immunomodulatory effects of Chinese herbal medicines.’, *The American Journal of Clinical Nutrition*, 66, pp. 1303–1312.
- Canini, A., Alesiani, D., D’Arcangelo, G. and Tagliatesta, P. (2007) ‘Gas chromatography–mass spectrometry analysis of phenolic compounds from Carica papaya L. leaf’, *Journal of Food Composition and Analysis*, 20, pp. 584–590.
- Carrington, L. B. and Simmons, C. P. (2014) ‘Human to mosquito transmission of Dengue viruses’, *Frontiers in Immunology*, 5(290), pp. 1–8.

- Castillo, J. A., Castrillón, J. C., Diosa-Toro, M., Betancur, J. G., St Laurent, G., Smit, J. M. and Urcuqui-Inchima, S. (2016) 'Complex interaction between dengue virus replication and expression of miRNA-133a', *BMC Infectious Diseases*, 16(1), p. 29.
- CDC (2010) 'Supplemental issue: Dengue outbreaks worldwide'. The Dengue Update.
- Chansang, U., Zahiri, N. S., Bansiddhi, J., Boonruad, T., Thongsrirak, P., Mingmuang, J., Benjapong, N. and Mulla, M. S. (2005) 'Mosquito larvicidal activity of aqueous extracts of long pepper (*Piper retrofractum* vahl) from Thailand', *Journal of Vector Ecology*, 30(2), pp. 195–200.
- Chen, C. D., Seleena, B., Nazni, W. A., Lee, H. L., Masri, S. M., Chiang, Y. F. and Sofian-Azirun, M. (2006) 'Dengue vectors surveillance in endemic areas in Kuala Lumpur City Centre and Selangor State, Malaysia', *Dengue Bulletin*, 30, pp. 197–203.
- Chen, C. L., Lin, C. F., Wan, S. W., Wei, L. S., Chen, M. C., Yeh, T. M., Liu, H. S., Anderson, R. and Lin, Y. S. (2013) 'Anti-dengue virus nonstructural protein 1 antibodies cause NO-mediated endothelial cell apoptosis via ceramide-regulated glycogen synthase kinase-3 β and NF- κ B activation.', *Journal of Immunology*, 191(4), pp. 1744–1752.
- Chen, H. R., Lai, Y. C. and Yeh, T. M. (2018) 'Dengue virus non-structural protein 1: A pathogenic factor, therapeutic target, and vaccine candidate', *Journal of Biomedical Science*, 25(58), pp. 1–11.
- Choochote, W., Tuetun, B., Kanjanapothi, D., Rattanachanpichai, E., Chaithong, U., Chaiwong, P., Jitpakdi, A., Tippawangkosol, P., Riyong, D. and Pitasawat, B. (2004) 'Potential of crude seed extract of celery, *Apium graveolens* L., against the mosquito *Aedes aegypti* (L.) (Diptera: Culicidae).', *Journal of Vector Ecology*, 29(2), pp. 340–346.
- Chukwuka, K. S., Iwuagwu, M. and Uka, U. N. (2013) 'Evaluation of nutritional components of *Carica papaya* L. at different stages of ripening', *Journal of Pharmacy and Biological Sciences*, 6(4), pp. 13–16.
- Clyde, K., Kyle, J. L. and Harris, E. (2006) 'Recent advances in deciphering viral and host determinants of Dengue virus replication and pathogenesis', *Journal of Virology*, 80(23), pp. 11418–11431.
- Dai, J. and Mumper, R. J. (2010) 'Plant phenolics: Extraction, analysis and their antioxidant and anticancer properties', *Molecules*, 15(10), pp. 7313–7352.

- Desftischer, P., Talarico, L., Nosedá, M., Pitabguimaraes, S., Damonte, E. and Duarte, M. (2006) 'Chemical structure and antiviral activity of carrageenans from *Meristiella gelidium* against herpes simplex and dengue virus', *Carbohydrate Polymers*, 63(4), pp. 459–465.
- Dharmarathna, S. L. C. A., Wickramasinghe, S., Waduge, R. N., Rajapakse, R. P. V. J. and Kularatne, S. A. M. (2013) 'Does *Carica papaya* leaf-extract increase the platelet count? An experimental study in a murine model', *Asian Pacific Journal of Tropical Biomedicine*, 3(9), pp. 720–724.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S. and Ju, Y.-H. (2014) 'Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*', *Journal of Food and Drug Analysis*, 22(3), pp. 296–302.
- Domínguez de María, P., Sinisterra, J. V, Tsai, S.-W. and Alcántara, A. R. (2006) '*Carica papaya* lipase (CPL): An emerging and versatile biocatalyst', *Biotechnology Advances*, 24(5), pp. 493–499.
- Drew, R. (2003) 'Micropropagation of *Carica papaya* and related species', in Jain, S. M. and Ishii, K. (eds) *Micropropagation of Woody Trees and Fruits*, pp. 543–564.
- Falleh, H., Ksouri, R., Lucchessi, M. E., Abdelly, C. and Magné, C. (2012) 'Ultrasound-assisted extraction: Effect of extraction time and solvent power on the levels of polyphenols and antioxidant activity of *Mesembryanthemum edule* L. Aizoaceae shoots', *Tropical Journal of Pharmaceutical Research*, 11(2), pp. 243–249.
- Friebe, P. and Harris, E. (2010) 'Interplay of RNA elements in the Dengue virus 5' and 3' ends required for viral RNA replication', *Journal of Virology*, 84(12), pp. 6103–6118.
- Galang, M. G. M., Macabeo, A. P. G., Chang, W.-C., Isobe, M. and Aguinaldo, M. A. M. (2016) 'Glucosides from the unripe fruit juice of *Carica papaya* Linn. (Caricaceae) cultivar "Red Lady" with antioxidant activity', *Journal of Functional Foods*, 22, pp. 358–362.
- García, C. C., Talarico, L., Almeida, N., Colombres, S., Duschatzky, C. and Damonte, E. B. (2003) 'Virucidal activity of essential oils from aromatic plants of San Luis, Argentina', *Phytotherapy Research*, 17, pp. 1073–1075.

- García, L. L., Padilla, L. and Castaño, J. C. (2017) 'Inhibitors compounds of the flavivirus replication process', *Virology Journal*, 14(1), p. 95.
- Giacometti, J., Žauhar, G. and Žuvić, M. (2018) 'Optimization of ultrasonic-assisted extraction of major phenolic compounds from olive leaves (*Olea europaea* L.) using response surface methodology', *Foods*, 7(9), pp. 1–14.
- Godói, I. P., Lemos, L. L. P., de Araújo, V. E., Bonoto, B. C., Godman, B. and Guerra Júnior, A. A. (2017) 'CYD-TDV dengue vaccine: Systematic review and meta-analysis of efficacy, immunogenicity and safety', *Journal of Comparative Effectiveness Research*, 6(2), pp. 165–180.
- Goel, A., Patel, D. N., Lakhani, K. K., Agarwal, S. B. and Agarwal, A. (2004) 'Dengue fever: A dangerous foe', *Journal Indian Academy of Clinical Medicine*, 5(3), pp. 247–258.
- Grzybowski, A., Tiboni, M. and Silva, M. A. N. (2011) 'The combined action of phytolarvicides for the control of dengue fever vector, *Aedes aegypti*', *Brazilian Journal of Pharmacognosy*, 22(3), pp. 1–9.
- Gubler, D. J. (2006) 'Dengue/dengue haemorrhagic fever: History and current status', *Novartis Foundation Symposium*, 277, pp. 3–16.
- Gupta, A., Patil, S. S. and Pendharkar, N. (2017) 'Antimicrobial and anti-inflammatory activity of aqueous extract of *Carica papaya*', *Journal of Herbmed Pharmacology*, 6(4), pp. 148–152.
- Guzman, A. and Istúriz, R. E. (2010) 'Update on the global spread of dengue', *International Journal of Antimicrobial Agents*, 36, pp. 40–42.
- Guzman, M. G., Halstead, S. B., Artsob, H., Buchy, P., Farrar, J., Gubler, D. J., Hunsperger, E., Kroeger, A., Margolis, H. S., Martínez, E., Nathan, M. B., Pelegrino, J. L., Simmons, C., Yoksan, S. and Peeling, R. W. (2010) 'Dengue: A continuing global threat', *Nature Reviews Microbiology*, 8, pp. 7–16.
- Halstead, S. B. (2012) 'Controversies in dengue pathogenesis', *Paediatrics and International Child Health*, 32(1), pp. 5–9.
- He, J. F., Luo, H. M., Liang, W. J., Zheng, K., Kang, M. and Liu, L. P. (2007) 'Epidemic situation of dengue fever in Guangdong province, China, 1990-2005', *Dengue Bulletin*, 31, pp. 1–9.
- He, X., Ma, Y., Yi, G., Wu, J., Zhou, L. and Guo, H. (2017) 'Chemical composition and antifungal activity of *Carica papaya* Linn. seed essential oil against *Candida* spp.', *Letters in Applied Microbiology*, 64(5), pp. 350–354.

- Hidari, K. I. P. J., Takahashi, N., Arihara, M., Nagaoka, M., Morita, K. and Suzuki, T. (2008) 'Structure and anti-dengue virus activity of sulfated polysaccharide from a marine alga', *Biochemical and Biophysical Research Communications*, 376(1), pp. 91–95.
- Huet, J., Looze, Y., Bartik, K., Raussens, V., Wintjens, R. and Boussard, P. (2006) 'Structural characterization of the papaya cysteine proteinases at low pH', *Biochemical and Biophysical Research Communications*, 341(2), pp. 620–626.
- Husin, F., Chan, Y. Y., Gan, S. H., Sulaiman, S. A. and Shueb, R. H. (2015) 'The effect of *Hydrocotyle sibthorpioides* Lam. extracts on in vitro dengue replication', *Evidence-Based Complementary and Alternative Medicine*, 2015.
- Husin, F., Yaakob, H., Rashid, S. N. A., Shahar, S. and Soib, H. H. (2019) 'Cytotoxicity study and antioxidant activity of crude extracts and SPE fractions from *Carica papaya* leaves', *Biocatalysis and Agricultural Biotechnology*, 19, pp. 1–6.
- Hussain, K., Ismail, Z., Sadikun, A., Ibrahim, P. and Roxb, P. (2008) 'Analysis of proteins, polysaccharides, glycosaponins contents of *Piper sarmentosum* Roxb. and anti-TB evaluation for bio-enhancing/interaction effects of leaf extracts with Isoniazid (INH)', *Natural Product Radiance*, 7(5), pp. 402–408.
- Ikram, E. H. K., Stanley, R., Netzel, M. and Fanning, K. (2015) 'Phytochemicals of papaya and its traditional health and culinary uses: A review', *Journal of Food Composition and Analysis*, 41, pp. 201–211.
- Imaga, N. A. and Adepoju, O. A. (2010) 'Analyses of antisickling potency of *Carica papaya* dried leaf extract and fractions', *Journal of Pharmacognosy and Phytotherapy*, 2(7), pp. 97–102.
- Jain, M., Ganju, L., Katiyal, A., Padwad, Y., Mishra, K. P., Chanda, S., Karan, D., Yogendra, K. M. S. and Sawhney, R. C. (2008) 'Effect of *Hippophae rhamnoides* leaf extract against dengue virus infection in human blood-derived macrophages', *Phytomedicine*, 15, pp. 793–799.
- Jamal, J. A. (2006) 'Malay traditional medicine: An overview of scientific and technological progress', *Asia Pacific Tech Monitor*, 23, pp. 37–49.
- Jassim, S. A. A. and Naji, M. A. (2003) 'Novel antiviral agents: A medicinal plant perspective', *Journal of Applied Microbiology*, 95(3), pp. 412–427.

- Jiang, W. L., Luo, X. L. and Kuang, S. J. (2005) 'Effects of *Alternanthera philoxeroides* Griseb against dengue virus in vitro', *Journal of First Military Medical University*, 25(4), pp. 454–456.
- Joseph, B., Sankarganesh, P., Ichiyama, K. and Yamamoto, N. (2015) 'In vitro study on cytotoxic effect and anti-DENV2 activity of *Carica papaya* L. leaf', *Frontiers in Life Science*, 8(1), pp. 18–22.
- Joshi, B., Sah, G. P., Basnet, B. B., Bhatt, M. R., Sharma, D., Subedi, K., Pandey, J. and Malla, R. (2011) 'Phytochemical extraction and antimicrobial properties of different medicinal plants: *Ocimum sanctum* (Tulsi), *Eugenia caryophyllata* (Clove), *Achyranthes bidentata* (Datiwan) and *Azadirachta indica* (Neem)', *Journal of Microbiology and Antimicrobials*, 3(1), pp. 1–7.
- Kala, C. P. (2012) 'Medicinal & aromatic plants leaf juice of *Carica papaya* L.: A remedy of dengue fever', *Medicinal and Aromatic Plants*, 1(6), pp. 1–2.
- Kaliwantoro, N., Soesatyo, M. H., Indarto, Juffrie, M. and Dharmastiti, R. (2016) 'Effect of dengue virus infection on the permeability of vero cells line', in *AIP Conference Proceedings*, pp. 1–6.
- Kaliyaperumal, K., Kim, H. M., Jegajeevanram, K., Xavier, J. and Vijayalakshmi, J. (2014) 'Papaya: A gifted nutraceutical plant - A critical review of recent human health research', *International Journal of Genuine Traditional Medicine*, 4, pp. 1–17.
- Kannan, A., Narayanan, K. S., Sasikumar, S., Philipose, J. and Surendran, S. A. (2014) 'Coagulopathy in dengue fever patients', *International Journal of Research in Medical Sciences*, 2(3), pp. 1070–1072.
- Kiat, T. S., Phippen, R., Yusof, R., Ibrahim, H., Khalid, N. and Rahman, N. A. (2006) 'Inhibitory activity of cyclohexenyl chalcone derivatives and flavonoids of fingerroot, *Boesenbergia rotunda* (L.), towards dengue-2 virus NS3 protease', *Bioorganic and Medicinal Chemistry Letters*, 16(12), pp. 3337–3340.
- Kim, M. S., Moore, P. H., Zee, F., Fitch, M. M. M., Steiger, D. L., Manshardt, R. M., Paull, R. E., Drew, R. A., Sekioka, T. and Ming, R. (2002) 'Genetic diversity of *Carica papaya* as revealed by AFLP markers', *Genome*, 45(3), pp. 503–512.
- Klawikkan, N., Nukoolkarn, V., Jirakanjanakit, N., Yoksan, S., Wiwat, C. and Thirapanmethee, K. (2011) 'Effect of Thai medicinal plant extracts against dengue virus in vitro', *Mahidol University Journal of Pharmaceutical Science*, 38(1–2), pp. 13–18.

- Krishna, K. L., Paridhavi, M. and Patel, J. A. (2008) 'Review on nutritional, medicinal and pharmacological properties of Papaya (*Carica papaya* Linn.)', *Natural Product Radiance*, 7(4), pp. 364–373.
- Kudi, A. C. and Myint, S. H. (1999) 'Antiviral activity of some Nigerian medicinal plant extracts', *Journal of Ethnopharmacology*, 68, pp. 289–294.
- Kuhn, R. J., Zhang, W., Rossmann, M. G., Pletnev, S. V, Corver, J., Lenches, E., Jones, C. T., Mukhopadhyay, S., Chipman, P. R., Strauss, E. G., Baker, T. S. and Strauss, J. H. (2002) 'Structure of Dengue virus: Implications for Flavivirus organization, maturation, and fusion', *Cell*. Elsevier, 108(5), pp. 717–725.
- Kulasinghe, S., Ediriweera, R. and Kumara, P. (2016) 'Association of abnormal coagulation tests with dengue virus infection and their significance as early predictors of fluid leakage and bleeding', *Sri Lanka Journal of Child Health*, 45(3), pp. 184–188.
- Kyle, J. L. and Harris, E. (2008) 'Global spread and persistence of dengue.', *Annual Review of Microbiology*, 62, pp. 71–92.
- Lam, S. K. (1993) 'Two decades of dengue in Malaysia', *Tropical Medicine*, 35(4), pp. 195–200.
- Leardkamolkarn, V., Sirigulpanit, W., Phurimsak, C., Kumkate, S., Himakoun, L. and Sripanidkulchai, B. (2012) 'The inhibitory actions of *Houttuynia cordata* aqueous extract on dengue virus and dengue-infected cells', *Journal of Food Biochemistry*, 36(1), pp. 86–92.
- Lee Huang, S., Zhang, L., Lin Huang, P., Chang, Y. T. and Huang, P. L. (2003) 'Anti-HIV activity of olive leaf extract (OLE) and modulation of host cell gene expression by HIV-1 infection and OLE treatment', *Biochemical and Biophysical Research Communications*, 307(4), pp. 1029–1037.
- Lescar, J., Soh, S., Lee, L. T., Vasudevan, S. G., Kang, C. and Lim, S. P. (2018) 'The Dengue virus replication complex: From RNA replication to protein-protein interactions to evasion of innate immunity', *Advances in Experimental Medicine and Biology*, 1062, pp. 115–129.
- Leyssen, P., de Clercq, E. and Neyts, J. (2000) 'Perspectives for the treatment of infections with Flaviviridae', *Clinical Microbiology Reviews*, 13(1), pp. 67–82.

- Lim, S. P., Wang, Q. Y., Noble, C. G., Chen, Y. L., Dong, H., Zou, B., Yokokawa, F., Nilar, S., Smith, P., Beer, D., Lescar, J. and Shi, P. Y. (2013) 'Ten years of dengue drug discovery: Progress and prospects', *Antiviral Research*, 100, pp. 500–519.
- Lin, C. F., Chiu, S. C., Hsiao, Y. L., Wan, S. W., Lei, H. Y., Shiau, A. L., Liu, H. S., Yeh, T. M., Chen, S. H., Liu, C. C. and Lin, Y. S. (2005) 'Expression of cytokine, chemokine, and adhesion molecules during endothelial cell activation induced by antibodies against dengue virus nonstructural protein 1', *The Journal of Immunology*, 174, pp. 395–403.
- Liza, M. S., Abdul Rahman, R., Mandana, B., Jinap, S., Rahmat, A., Zaidul, I. S. M. and Hamid, A. (2010) 'Supercritical carbon dioxide extraction of bioactive flavonoid from *Strobilanthes crispus* (Pecah Kaca)', *Food and Bioprocess Processing*, 88, pp. 319–326.
- Longdet, I. Y. and Adoga, E. A. (2017) 'Effect of methanolic Leaf extract of *Carica papaya* on *Plasmodium berghei* infection in albino mice', *European Journal of Medicinal Plants*, 20(1), pp. 1–7.
- Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. (1951) 'Protein measurement with the Folin phenol reagent.', *The Journal of Biological Chemistry*, 193(1), pp. 265–275.
- Makanjuola, S. A. (2017) 'Influence of particle size and extraction solvent on antioxidant properties of extracts of tea, ginger, and tea-ginger blend', *Food Science & Nutrition*, 5(6), pp. 1179–1185.
- Malavige, G. N. and Ogg, G. S. (2017) 'Pathogenesis of vascular leak in dengue virus infection', *Immunology*, 151(3), pp. 261–169.
- Mariana, C. P. and Juan, N. F. (2017) 'Domestication and genetics of papaya: A review', *Frontiers in Ecology and Evolution*, 5(12), pp. 1–9.
- Martins, S. D. T., Silveira, G. F., Alves, L. R., Duarte dos Santos, C. N. and Bordignon, J. (2012) 'Dendritic cell apoptosis and the pathogenesis of dengue.', *Viruses*, 4(11), pp. 2736–2753.
- Md Salleh, L., Hartati, H., Jamaludin, R., Che Yunus, M. A., Yakub, H. and Abd. Aziz, A. (2014) 'Antioxidant activity and total phenolic contents in methanol extracts from *Swietenia mahagoni* and *Andrographis paniculata*', *Jurnal Teknologi*, 69(4), pp. 51–53.

- Melariri, P., Campbell, W., Etusim, P. and Smith, P. (2011) 'Antiplasmodial properties and bioassay-guided fractionation of ethyl acetate extracts from *Carica papaya* leaves', *Journal of Parasitology Research*, 2011, pp. 1–7.
- Meneses, R., Ocazonez, R. E., Martínez, J. R. and Stashenko, E. E. (2009) 'Inhibitory effect of essential oils obtained from plants grown in Colombia on yellow fever virus replication in vitro', *Annals of Clinical Microbiology and Antimicrobials*, 8, pp. 1–8.
- Ministry of Health Malaysia (2006) 'Health facts 2006'.
- Mir, A., Ismatullah, H., Rauf, S. and Niazi, U. H. K. (2016) 'Identification of bioflavonoid as fusion inhibitor of dengue virus using molecular docking approach', *Informatics in Medicine Unlocked*, 3, pp. 1–6.
- Mohan, L., Amberkar, M. and Meena, K. (2011) '*Ocimum sanctum* Linn (Tulsi): An overview', *International Journal of Pharmaceutical Science Review and Research*, 7(1), pp. 51–53.
- Mohd Zaki, A. H., Brett, J., Ismail, E. and L'Azou, M. (2014) 'Epidemiology of dengue disease in Malaysia (2000-2012): A systematic literature review', *PLoS Neglected Tropical Diseases*, 8(11), pp. 1–9.
- Mojisola, C. O. C., Anthony, E. A. and Alani, D. M. (2009) 'Antisickling properties of the fermented mixture of *Carica papaya* Linn and *Sorghum bicolor* (L.) Moench', *African Journal of Pharmacy and Pharmacology*, 3(4), pp. 140–143.
- Muhammad Azami, N. A., Salleh, S. A., Neoh, H. M., Syed Zakaria, S. Z. and Jamal, R. (2011) 'Dengue epidemic in Malaysia: Not a predominantly urban disease anymore', *BMC Research Notes*, 4(1), pp. 216–219.
- Muliawan, S. Y., Kit, L. S., Devi, S., Hashim, O. and Yusof, R. (2006) 'Inhibitory potential of *Quercus lusitanica* extract on dengue virus type-2 replication', *Southeast Asian Journal Tropical Medicine Public Health*, 37(3), pp. 132–135.
- Newman, D. J. and Cragg, G. M. (2016) 'Natural products as sources of new drugs from 1981 to 2014', *Journal of Natural Products*, 79(3), pp. 629–661.
- Newman, D. J., Cragg, G. M. and Snader, K. M. (2003) 'Natural products as sources of new drugs over the period 1981-2002', *Journal of Natural Products*, 66, pp. 1022–1037.

- Ng, L. C., Chem, Y. K., Koo, C., Mudin, R. N. B., Amin, F. M., Lee, K. S. and Kheong, C. C. (2015) '2013 dengue outbreaks in Singapore and Malaysia caused by different viral strains', *American Journal of Tropical Medicine and Hygiene*, 92(6), pp. 1150–1155.
- Nguyen, T. T., Parat, M. O., Hodson, M. P., Pan, J., Shaw, P. N. and Hewavitharana, A. K. (2015) 'Chemical characterization and in vitro cytotoxicity on Squamous cell Carcinoma cells of Carica papaya leaf extracts', *Toxins*, 8(1), p. 7.
- Ocazonez, R. E., Meneses, R., Torres, F. Á. and Stashenko, E. (2010) 'Virucidal activity of Colombian Lippia essential oils on dengue virus replication in vitro', *Memorias do Instituto Oswaldo Cruz*, 105(3), pp. 304–309.
- Ojha, A., Nandi, D., Batra, H., Singhal, R., Annarapu, G. K., Bhattacharyya, S., Seth, T., Dar, L., Medigeshi, G. R., Vrati, S., Vikram, N. K. and Guchhait, P. (2017) 'Platelet activation determines the severity of thrombocytopenia in dengue infection', *Scientific Reports*, 7(41697), pp. 1–10.
- Okeniyi, J. A. O., Ogunlesi, T. A., Oyelami, O. A. and Adeyemi, L. A. (2007) 'Effectiveness of dried Carica papaya seeds against human intestinal parasitosis: A pilot study', *Journal of Medicinal Food*, 10(1), pp. 194–196.
- Okoduwa, S. I. R., Umar, I. A., James, D. B., Inuwa, H. M. and Habila, J. D. (2016) 'Evaluation of extraction protocols for anti-diabetic phytochemical substances from medicinal plants', *World Journal of Diabetes*, 7(20), p. 605—614.
- Okpe, O., Habila, N., Ikwebe, J., Upev, V. A., Okoduwa, S. I. R. and Isaac, O. T. (2016) 'Antimalarial potential of Carica papaya and Vernonia amygdalina in mice infected with Plasmodium berghei', *Journal of Tropical Medicine*, 2016, pp. 1–6.
- Ono, L., Wollinger, W., Rocco, I. M., Coimbra, T. L. M., Gorin, P. a J. and Sierakowski, M. R. (2003) 'In vitro and in vivo antiviral properties of sulfated galactomannans against yellow fever virus (BeH111 strain) and dengue 1 virus (Hawaii strain)', *Antiviral Research*, 60(3), pp. 201–208.
- Otu, A., Ebenso, B., Etokidem, A. and Chukwuekezie, O. (2019) 'Dengue fever: An update review and implications for Nigeria and similar countries', *African Health Sciences*, 19(2), pp. 2000–2007.
- Owoyele, B. V., Adebukola, O. M., Funmilayo, A. A. and Soladoye, A. O. (2008) 'Anti-inflammatory activities of ethanolic extract of Carica papaya leaves', *Inflammopharmacology*, 16, pp. 168–173.

- Pang, E. L. and Loh, H. S. (2016) 'Current perspectives on dengue episode in Malaysia', *Asian Pacific Journal of Tropical Medicine*, 9(4), pp. 395–401.
- Pang, T., Hassan, H. and Ramalingam, S. (1988) *Demam denggi dan demam denggi berdarah*. Kuala Lumpur: Dewan Bahasa dan Pustaka.
- Parekh, J., Jadeja, D. and Chanda, S. (2005) 'Efficacy of aqueous and methanol extracts of some medicinal plants for potential antibacterial activity', *Turkish Journal of Biology*, 29, pp. 203–210.
- Parida, M. M., Upadhyay, C., Pandya, G. and Jana, a. M. (2002) 'Inhibitory potential of neem (*Azadirachta indica* Juss) leaves on dengue virus type-2 replication', *Journal of Ethnopharmacology*, 79, pp. 273–278.
- Patel, K., Panchal, N. and Ingle, P. (2019) 'Review of extraction techniques. Extraction methods: Microwave, ultrasonic, pressurized fluid, Soxhlet extraction, Etc', *International Journal of Advanced Research in Chemical Science*, 6(3), pp. 6–21.
- Pedro, C. Q., Tania, G. F., Ingrid, R. B. and Santiago, G. T. (2011) 'Antifungal activity in ethanolic extracts of *Carica papaya* L. cv. Maradol leaves and seeds.', *Indian Journal of Microbiology*, 51(1), pp. 54–60.
- Pujol, C. A., Estevez, J. M., Carlucci, M. J., Ciancia, M., Cerezo, A. S. and Damonte, E. B. (2002) 'Novel DL-galactan hybrids from the red seaweed *Gymnogongrus torulosus* are potent inhibitors of Herpes Simplex Virus and dengue virus', *Antiviral Chemistry and Chemotherapy*, 13(2), pp. 83–89.
- Qi, R., Zhang, L. and Chi, C. (2008) 'Biological characteristics of dengue virus and potential targets for drug design', *Acta Biochimica et Biophysica Sinica*, 40(2), pp. 91–101.
- Raj Kapoor, B., Jayakar, B., Kavimani, S. and Muruges, N. (2002) 'Effect of dried fruits of *Carica papaya* Linn on hepatotoxicity', *Biological & Pharmaceutical Bulletin*, 25(12), pp. 1645–1646.
- Rates, S. M. . (2001) 'Plants as source of drugs', *Toxicon*, 39(5), pp. 603–613.
- Rees, C. R., Costin, J. M., Fink, R. C., McMichael, M., Fontaine, K. A., Isern, S. and Michael, S. F. (2008) 'In vitro inhibition of dengue virus entry by p-sulfoxy-cinnamic acid and structurally related combinatorial chemistries', *Antiviral research*, 80(2), pp. 135–142.

- Reis, S. R. I. N., Valente, L. M. M., Sampaio, A. L., Siani, A. C., Gandini, M., Azeredo, E. L., D'Avila, L. A., Mazzei, J. L., Henriques, M. das G. M. and Kubelka, C. F. (2008) 'Immunomodulating and antiviral activities of *Uncaria tomentosa* on human monocytes infected with dengue virus-2', *International Immunopharmacology*, 8(3), pp. 468–476.
- Robards, K. (2003) 'Strategies for the determination of bioactive phenols in plants, fruit and vegetables', *Journal of Chromatography A*, 1000(1), pp. 657–691.
- Robinson, M. M. and Zhang, X. (2011) *The world medicines situation 2011 traditional medicine: global situation, issues and challenges*. Geneva: World Health Organization.
- Saleeza, S. N. R., Norma, R. Y. and Azirun, M. S. (2011) 'Mosquitoes larval breeding habitat in urban and suburban areas, Peninsular Malaysia', *World Academy of Science, Engineering and Technology*, 58, pp. 569–573.
- Sánchez, I., Gómez-Garibay, F., Taboada, J. and Ruiz, B. H. (2000) 'Antiviral effect of flavonoids on the dengue virus', *Phytotherapy Research*, 14(2), pp. 89–92.
- Dos Santos, M., Teixeira, T. R., Santos, F. R. da S., Lima, W. G., Ferraz, A. C., Silva, N. L., Leite, F. J., Siqueira, J. M., Luyten, W., de Castro, A. H. F., de Magalhães, J. C. and Ferreira, J. M. S. (2019) 'Bauhinia holophylla (Bong.) Steud. leaves-derived extracts as potent anti-dengue serotype-2', *Natural Product Research*, pp. 1–6.
- Sanyaolu, A., Okorie, C., Badaru, O., Adetona, K., Ahmed, M., Akanbi, O., Foncham, J., Kadavil, S., Likaj, L., Miraj-Raza, S., Pearce, E., Sylvester, R. and Wallis, E. (2017) 'Global epidemiology of dengue hemorrhagic fever: An update', *Journal of Human Virology & Retrovirology*, 5(6), pp. 1–7.
- Sathasivam, K., Ramanathan, S., Mansor, S. M., Haris, M. R. M. H. and Wernsdorfer, W. H. (2009) 'Thrombocyte counts in mice after the administration of papaya leaf suspension', *Wiener Klinische Wochenschrift The Middle European Journal of Medicine*, 121(3), pp. 19–22.
- Sekeli, R., Hamid, M. H., Razak, R. A., Wee, C. and Ong-abdullah, J. (2018) 'Malaysian *Carica papaya* L. var. eksotika: Current research strategies fronting challenges', *Frontiers in Plant Science*, 9(9), pp. 1–9.
- Sellahewa, K. H. (2013) 'Pathogenesis of dengue haemorrhagic fever and its impact on case management', *ISRN Infectious Diseases*, pp. 1–6.

- Senthilvel, P., Lavanya, P., Kumar, K. M., Swetha, R., Anitha, P., Bag, S., Sarveswari, S., Vijayakumar, V., Ramaiah, S. and Anbarasu, A. (2013) 'Flavonoid from *Carica papaya* inhibits NS2B-NS3 protease and prevents Dengue 2 viral assembly.', *Bioinformation*, 9(18), pp. 889–895.
- Sherwani, S. K., Bokhari, T. Z., Nazim, K., Gilani, S. A. and Kazmi, S. U. (2013) 'Qualitative phytochemical screening and antifungal activity of *Carica papaya* leaf extract against human and plant pathogenic fungi', *International Research Journal of Pharmacy*, 4(7), pp. 83–86.
- Shi, J., Nawaz, H., Pohorly, J., Mittal, G., Kakuda, Y. and Jiang, Y. (2005) 'Extraction of polyphenolics from plant material for functional food: Engineering and technology', *Food Reviews International*. Taylor & Francis, 21(1), pp. 139–166.
- Siler, J. F., Hall, M. W. and Hitchens, A. P. (1926) 'Dengue: Its history, epidemiology, mechanism of transmission, etiology, clinical manifestations, immunity, and prevention', *Philippine Journal of Science*. Manila, 29(1–2), pp. 1–304.
- Sinha, N., Gupta, N., Jhamb, R., Gulati, S. and Kulkarni Ajit, V. (2008) 'The 2006 dengue outbreak in Delhi, India', *The Journal of Communicable Diseases*, 40(4), pp. 243–248.
- Soares Melecchi, M. I., Péres, V. F., Dariva, C., Zini, C. A., Abad, F. C., Martinez, M. M. and Caramão, E. B. (2006) 'Optimization of the sonication extraction method of *Hibiscus tiliaceus* L. flowers', *Ultrasonics Sonochemistry*, 13(3), pp. 242–250.
- Srivastava, M. and Kapoor, V. P. (2005) 'Seed galactomannans: An overview', *Chemistry and Biodiversity*, 2(3), pp. 295–317.
- Suaya, J. A., Shepard, D. S., Siqueira, J. B., Martelli, C. T., Lum, L. C. S., Tan, L. H., Kongsin, S., Jiamton, S., Garrido, F., Montoya, R., Armien, B., Huy, R., Castillo, L., Caram, M., Sah, B. K., Sughayyar, R., Tyo, K. R. and Halstead, S. B. (2009) 'Cost of dengue cases in eight countries in the Americas and Asia: A prospective study', *American Journal of Tropical Medicine and Hygiene*, 80(5), pp. 846–855.
- Sudhakar, N. and Theivanai, V. R. M. (2014) 'Potential medicinal properties of *Carica papaya* Linn.: A mini review', *International of Pharmacy and Pharmaceutical Sciences*, 6(2), pp. 1–4.

- Sugawara, K., Nishiyama, K., Ishikawa, Y., Abe, M., Sonoda, K., Komatsu, K., Horikawa, Y., Takeda, K., Honda, T., Kuzuhara, S., Kino, Y., Mizokami, H., Mizuno, K., Oka, T. and Honda, K. (2002) 'Development of Vero cell-Derived inactivated Japanese encephalitis vaccine', *Biologicals*, 30(4), pp. 303–314.
- Takahashi, H. and Suzuki, Y. (2017) 'Cellular control of Dengue virus replication: Role of interferon-inducible genes', in Sperança, M. A. (ed.) *Dengue - Immunopathology and Control Strategies*. Rijeka: IntechOpen.
- Talarico, L. B., Pujol, C. a, Zibetti, R. G. M., Faría, P. C. S., Nosedá, M. D., Duarte, M. E. R. and Damonte, E. B. (2005) 'The antiviral activity of sulfated polysaccharides against dengue virus is dependent on virus serotype and host cell', *Antiviral Research*, 66(2–3), pp. 103–110.
- Tan, P. W., Tan, C. P. and Ho, C. W. (2011) 'Antioxidant properties: Effects of solid-to-solvent ratio on antioxidant compounds and capacities of pegaga (*Centella asiatica*)', *International Food Research Journal*, 18(2), pp. 557–562.
- Tang, L. I. C., Ling, A. P. K., Koh, R. Y., Chye, S. M. and Voon, K. G. L. (2012) 'Screening of anti-dengue activity in methanolic extracts of medicinal plants', *BMC Complementary and Alternative Medicine*, 12(3), pp. 1–10.
- Tecelão, C., Rivera, I., Sandoval, G. and Ferreira Dias, S. (2012) 'Carica papaya latex: A low-cost biocatalyst for human milk fat substitutes production', *European Journal of Lipid Science and Technology*, 114(3), pp. 266–276.
- Teixeira da Silva, J. A., Rashid, Z., Nhut, D., Sivakumar, D., Gera, A., Souza Junior, M. and Tennant, P. (2007) 'Papaya (*Carica papaya* L.) biology and biotechnology', *Tree and Forestry Science and Biotechnology*, 1, pp. 47–73.
- Teng, W. C., Chan, W., Suwanarusk, R., Ong, A., Ho, H. K., Russell, B., Rénia, L. and Koh, H. L. (2019) 'In vitro antimalarial evaluations and cytotoxicity investigations of *Carica papaya* leaves and carpaine', *Natural Product Communications*, 14(1), pp. 33–36.
- Thomas, S. J. and Yoon, I. K. (2019) 'A review of Dengvaxia®: Development to deployment', *Human vaccines & immunotherapeutics*, 15(10), pp. 2295–2314.
- Trujillo Correa, A. I., Quintero Gil, D. C., Diaz Castillo, F., Quiñones, W., Robledo, S. M. and Martinez Gutierrez, M. (2019) 'In vitro and in silico anti-dengue activity of compounds obtained from *Psidium guajava* through bioprospecting', *BMC Complementary and Alternative Medicine*, 19(1), p. 298.

- Uncini Manganelli, R. E., Zaccaro, L. and Tomei, P. E. (2005) 'Antiviral activity in vitro of *Urtica dioica* L., *Parietaria diffusa* M. et K. and *Sambucus nigra* L.', *Journal of Ethnopharmacology*, 98(3), pp. 323–327.
- Veeraseatakul, P., Wongchompoo, B., Thichak, S., Yananto, Y., Waneesorn, J. and Chutipongvivate, S. (2007) 'Circulation of dengue serotypes in five provinces of northern Thailand during 2002-2006', *Dengue Bulletin*, 31, pp. 19–25.
- Vij, T. and Prashar, Y. (2015) 'A review on medicinal properties of *Carica papaya* Linn.', *Asian Pacific Journal of Tropical Disease*, 5(1), pp. 1–6.
- Vijayan, P., Raghu, C., Ashok, G., Dhanaraj, S. A. and Suresh, B. (2004) 'Antiviral activity of medicinal plants of Nilgiris', *Indian Journal of Medicinal Research*, 120, pp. 24–29.
- Weyermann, J., Lochmann, D. and Zimmer, A. (2005) 'A practical note on the use of cytotoxicity assays', *International Journal of Pharmaceutics*, 288(2), pp. 369–376.
- White, L. A. (1987) 'Susceptibility of *Aedes albopictus* C6/36 cells to viral infection', *Journal of clinical microbiology*, 25(7), pp. 1221–1224.
- Wittenauer, J., Mäckle, S., Sußmann, D., Ute, S. W. and Carle, R. (2015) 'Inhibitory effects of polyphenols from grape pomace extract on collagenase and elastase activity', *Fitoterapia*, 101, pp. 179–187.
- Wong, S. S., Abd Jamil, J. and Abu Bakar, S. (2007) 'Antibody neutralization and viral virulence in recurring dengue virus type-2 outbreaks', *Viral Immunology*, 20(3), pp. 359–368.
- World Health Organization (1997) *Dengue haemorrhagic fever: Diagnosis, treatment, prevention and control*. 2nd edn. Geneva: World Health Organization.
- World Health Organization (2009) *Dengue: Guidelines for diagnosis, treatment, prevention and control*. New edn. Geneva: World Health Organization.
- Yogiraj, V., Goyal, P. K., Chauhan, C. S., Goyal, A. and Vyas, B. (2014) '*Carica papaya* Linn: An overview', *International Journal of Herbal Medicine*, 2(5), pp. 1–8.
- Yohan, B., Kendarsari, R. I., Mutia, K., Bowolaksono, A., Harahap, A. R. and Sasmono, R. T. (2014) 'Growth characteristics and cytokine/chemokine induction profiles of dengue viruses in various cell lines', *Acta virologica*, 58(1), p. 20–27.

- Yunita, F., Hanani, E. and Kristianto, J. (2012) 'The effect of *Carica papaya* L. leaves extract capsules on platelets count and hematocrit level in dengue fever patient', *International Journal of Medicinal and Aromatic Plants*, 2(4), pp. 573–578.
- Zakaria, I. I., Salin, N. H., Amanah, A., Othman, S., Khairuddin, F., Khawory, M. H., Wahab, R. A., Rahaman, M. R. A., Chern, P. P., Johari, N. A. and Wahab, H. (2019) 'Potential anti-viral compounds from Malaysian plant natural product repository and database (MyNature50000) for DENV2', *Biotechnology & Biotechnological Equipment*, 33(1), pp. 379–389.
- Zakaria, Z. A., Mat Jais, A. M., Sulaiman, M. R., Mohamed Isa, S. S. P. and Riffin, S. (2006) 'The in vitro antibacterial activity of methanol and ethanol extracts of *Carica papaya* flowers and *Magnifera indica* leaves', *Journal of Pharmacology and Toxicology*, 1(3), pp. 278–283.
- Zandi, K., Lani, R., Wong, P. F., Teoh, B. T., Sam, S. S., Johari, J., Mustafa, M. R. and Abu Bakar, S. (2012) 'Flavone enhances dengue virus type-2 (NGC strain) infectivity and replication in Vero cells', *Molecules*, 17, pp. 2437–2445.
- Zandi, K., Teoh, B. T., Sam, S. S., Wong, P. F. and Mustafa, M. R. (2011) 'In vitro antiviral activity of fisetin, rutin and naringenin against dengue virus type-2', *Journal of Medicinal Plants Research*, 5(23), pp. 5534–5539.
- Zandi, K., Teoh, B. T., Sam, S. S., Wong, P. F., Mustafa, M. R. and Abu Bakar, S. (2011) 'Antiviral activity of four types of bioflavonoid against dengue virus type-2.', *Virology Journal*, 8(1), p. 560.
- Zandi, K., Teoh, B. T., Sam, S. S., Wong, P. F., Mustafa, M. R. and Abu Bakar, S. (2012) 'Novel antiviral activity of baicalein against dengue virus.', *BMC Complementary and Alternative Medicine*, 12(1), p. 214.
- Zhang, Q. W., Lin, L. G. and Ye, W. C. (2018) 'Techniques for extraction and isolation of natural products: A comprehensive review', *Chinese Medicine*, 13, p. 20.
- Zubairi, S. I., Sarmidi, M. R. and Aziz, R. A. (2014) 'The effects of raw material particles size, types of solvents and solvent-to-solid ratio on the yield of rotenone extracted from *Derris elliptica* roots', *Sains Malaysiana*, 43(5), pp. 707–713.

Zunjar, V., Dash, R. P., Jivrajani, M., Trivedi, B. and Nivsarkar, M. (2016)
‘Antithrombocytopenic activity of carpaine and alkaloidal extract of *Carica papaya* Linn. leaves in busulfan induced thrombocytopenic Wistar rats’,
Journal of Ethnopharmacology, 181, pp. 20–25.

LIST OF PUBLICATIONS

Journal with Impact Factor

1. **Abd Kadir, S. L.**, Ya'akob, H. & Mohamed Zulkefli, R. (2013). Potential anti-dengue medicinal plants: A review. *Journal of Natural Medicines*, 67 (4), pp. 677-689. doi: [10.1007/s11418-013-0767-y](https://doi.org/10.1007/s11418-013-0767-y) (IF: 1.966)

Non-Indexed Conference Proceedings

1. **Abd Kadir, S. L.**, Ya'akob, H., Mohamed Zulkefli, R. and Moghaddam, E. (2014) Screening of antiviral activity in *Carica papaya* aqueous extract on dengue virus type-2. *5th International Conference on Biotechnology for the Wellness Industry*. Kuala Lumpur.
2. **Abd Kadir, S. L.**, Ya'akob, H., Mohamed Zulkefli, R. and Moghaddam, E. (2014) The effects of *Carica papaya* leaves extract against dengue virus infected Vero cells. *2014 Asian Conference on the Life Sciences and Sustainability*. Hiroshima, Japan.
3. Ya'akob, H., Mohamed Zulkefli, R., Abd Rashid, S. N. A., Zainol, N. A., Sadek, N. A., Aman Nor, N. F., **Abd Kadir, S. L.**, Shaiful Amri, A. S., and Nik Mat Daud, N. N. N. (2015) Anti-Dengue: A novel Polyherbal anti-dengue capsule. *17th Industrial Art and Technology Exhibition (INATEX) 2015*. Universiti Teknologi Malaysia, Johor.
4. Shaiful Amri, A. S., Ya'akob H., **Abd Kadir, S. L.**, and Husin, F. (2016) Acute oral toxicity study and anti-viral properties of *Carica papaya* leaves extract against DENV. *6th International Conference on Biotechnology for the Wellness Industry*. Melaka.

5. **Abd Kadir, S. L.**, Ya'akob, H., Shueb, R. H., Shaiful Amri, A. S. and Husin, F. (2016) Cytotoxic effects of *Carica papaya* leaves aqueous extract on C6/36 and Vero 76 cell line. *Asia Dengue Conference*. Kuala Lumpur.
6. Shaiful Amri, A. S., Ya'akob H., **Abd Kadir, S. L.**, and Husin, F. (2016) Prototype development of anti-dengue supplement from *Carica papaya* leaves, polyherbal extract and virgin coconut oil. *Asia Dengue Conference*. Kuala Lumpur.