

PROTEIN EXPRESSION PROFILING OF *Orthosiphon aristatus* (Blume) Miq
AND MOLECULAR DYNAMICS SIMULATION OF TRANSKETOLASE WITH
ANTIDIABETIC POTENTIAL

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DEDICATION

This thesis is dedicated to my father, who always give me motivation, support and believed that I can do it whatever I want to. It is also dedicated to my beloved late mother, who always give me strength to accomplish all the tasks I have.

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ABSTRACT

Orthosiphon aristatus (Blume) Miq is a well-known medicinal plant widely commercialized especially as tea product to promote health. However, to date, the proteome of *O. aristatus* has never been studied. Hence, this study used four approaches; one-dimensional (1D) and two-dimensional (2D) proteomics, antidiabetic assay and bioinformatics to analyze the leaves proteome of *O. aristatus*. Protein from fresh and dried leaves (microwave-drying, halogen oven-drying and freeze-drying) was extracted using phenol/sodium dodecyl sulphate (SDS) buffer with three preliminary washes and separated by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) to check the quality. Subsequently, antioxidant assays; ferric reducing antioxidant power (FRAP) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) were performed on the protein to identify a suitable drying method with minimum effect on plant's beneficial content as it is highly consumed as tea product. Microwave-drying was the most suitable drying method for *O. aristatus* as it produced better protein quality ($859.49 \pm 5.98 \mu\text{g/mL}$, 14 protein bands detected) with high antioxidant activity (71.5% free radical scavenging activity, $1426.13 \text{ Fe}^{2+}\text{mM/g}$). Thus, only fresh and microwave-dried leaves protein were further used for subsequent study. Over 1000 proteins were identified from fresh and dried leaves using Liquid Chromatography Tandem Mass Spectrometry (LC-MS/MS) with 43 proteins found to be similar for both samples. The profiled proteins were obviously affected by heat stress, where proteins with known anticancer property such as thioredoxin were only present in fresh leaves whereas myrosinase and glutathione were only extracted from dried leaves. Protein expression profile of fresh and dried leaves using 2D approach was analyzed with PDQuest software. Fresh and dried leaves have 202 and 106 spots of different distribution pattern on 2D gel respectively. Via PDQuest analysis, 32 distinct spots were detected from fresh leaves but not dried leaves and 12 spots with 2-fold expression detected from both types of leaves. Fresh and dried leaves protein also showed potential antidiabetic activity from inhibitions of glucose diffusion, α -amylase and α -glucosidase. Subsequently, one of the identified proteins, Transketolase (TKT) which plays a crucial role in preventing diabetes complications, was studied further in terms of its thermostability using molecular dynamics (MD) simulation at different temperature (310 K, 368 K, 373 K, 423 K, 453 K and 473 K) and interaction with its ligand, thiamine diphosphate (TPP) via Autodock Vina. The MD results indicated that TKT is a thermostable protein but TKT and TKT-TPP complex may start to denature when exposed to temperature beyond boiling point. Gln237, Ser242, Cys246, Arg283 and Phe284 were identified as the possible TPP binding sites. Taken together, the findings provide a snapshot of fresh and dried leaves proteome of *O. aristatus* showing that the plant protein exhibited antioxidant and antidiabetic activities. The protein expression profile of *O. aristatus* could also be used for quality control purpose as a biomarker for *O. aristatus*. TKT bioinformatics information provided pharmacological insight that can accelerate drug design process.

ABSTRAK

Orthosiphon aristatus (Blume) Miq merupakan tumbuhan perubatan terkenal dikomersialkan secara meluas terutamanya sebagai produk teh untuk meningkatkan kesihatan. Walaubagaimanapun, sehingga kini, maklumat proteom *O. aristatus* masih belum pernah dikaji. Oleh itu, kajian ini menggunakan empat pendekatan; proteomik 1-dimensi (1D) dan 2-dimensi (2D), asai antidiabetik dan bioinformatik untuk menganalisa proteom daun *O. aristatus*. Protein daripada daun segar dan kering (pengeringan gelombang mikro, pengeringan ketuhar halogen dan pengeringan beku) telah diesktrak menggunakan penimbal fenol/sodium dodesil sulfat (SDS) dengan tiga pembersihan awal dan dipisahkan menggunakan electrophoresis gel sodium dodesil sulfat-poliakrilamida (SDS-PAGE) untuk pemeriksaan kualiti. Seterusnya, ujian antioksidan; *ferric reducing antioxidant power* (FRAP) dan 2,2-difenil-1-pikrilhidrazil (DPPH) dilakukan ke atas protein untuk mengenalpasti kaedah pengeringan yang sesuai dengan kesan minimum pada kandungan bermanfaat tumbuhan kerana ia digunakan sebagai produk teh. Pengeringan gelombang mikro adalah cara pengeringan yang paling sesuai untuk *O. aristatus* kerana menghasilkan kualiti protein yang lebih baik ($859.49 \pm 5.98 \mu\text{g/mL}$, 14 jalur protein dikesan) dengan aktiviti antioksidan yang tinggi (71.5% aktiviti skavengan radikal bebas, $1426.13 \text{ Fe}^{2+} \text{ mM/g}$). Oleh itu, hanya protein daun segar dan dikeringkan melalui gelombang mikro digunakan untuk kajian selanjutnya. Lebih 1000 protein telah dikenalpasti daripada daun segar dan kering menggunakan Liquid Chromatography Tandem Mass Spectrometry (LC-MS/MS) dengan 43 protein adalah sama bagi kedua-dua sampel. Protein terprofil didapati terjejas oleh haba, di mana protein yang diketahui bersifat antikanser seperti tioredoksin hanya terdapat dalam daun segar manakala mirosinase dan glutathion hanya diesktrak daripada daun kering. Profil pengekspressan protein daun segar dan kering menggunakan kaedah 2D telah dianalisa menggunakan perisian PDQuest. Daun segar dan kering masing-masing mempunyai 202 dan 106 titik protein dengan corak sebaran atas 2D yang berbeza. Melalui analisis PDQuest, 32 titik dikesan pada daun segar tetapi tidak pada daun kering dan 12 bintik dengan pengekspressan dua kali ganda dikesan pada kedua-dua jenis daun. Protein daun segar dan kering juga menunjukkan potensi aktiviti antidiabetik melalui perencanan sebaran glukosa, α -amilase dan α -glukosidase. Salah satu protein yang dikenalpasti, transketolase (TKT) yang diketahui memainkan peranan penting dalam pencegahan komplikasi diabetes, telah dikaji lebih lanjut dari segi termostabiliti menggunakan simulasi dinamik molekul (MD) pada suhu yang berbeza (310 K, 368 K, 373 K, 423 K, 453 K and 473 K) dan interaksi dengan ligannya, tiamin difosfat (TPP) melalui Autodock Vina. Keputusan MD menunjukkan TKT adalah protein stabil-suhu tetapi TKT dan juga kompleks TKT-TPP mungkin mula dinyahasli apabila terdedah kepada suhu di atas paras didih. Gln237, Ser242, Cys246, Arg283 and Phe284 telah dikenalpasti berkemungkinan menjadi tapak pengikatan TPP. Kesimpulannya, hasil kajian memberi gambaran proteom daun segar dan kering *O. aristatus* dan menunjukkan protein tumbuhan ini mempunyai bioaktiviti antioksidan dan antidiabetik. Profil ekspresi protein *O. aristatus* juga boleh digunakan sebagai biopenanda untuk pengawalan kualiti. Maklumat bioinformatik TKT memberi pandangan farmakologi yang dapat mempercepatkan proses reka bentuk ubat.

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LIST OF ABBREVIATIONS

1D	-	One-dimensional
1-DE	-	One-dimensional Gel Electrophoresis
2D		Two-dimensional
2-DE		Two-dimensional Gel Electrophoresis
3D	-	Three-dimensional
ABTS	-	2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonc acid)
ACE	-	Angiotensin-converting Enzyme
APS	-	Ammonium Persulfate
AtBRCA2	-	<i>Arabidopsis</i> Breast Cancer 2 Like 2A Protein
BRCA2	-	Breast Cancer 2 Like 2A Protein
BSA	-	Bovine Serum Albumin
CBB-R	-	Coomassie Brilliant Blue-Red
DAG-PKC	-	Diacylglycerol-Protein Kinase C
DIGE	-	Difference Gel Electrophoresis
DPP-4	-	Dipeptidyl Peptidase-4
DPPH	-	1,1-diphenyl-2-picrylhydrazyl
DTT	-	Dithiothreitol
FeSO ₄ .7H ₂ O	-	Iron (II) Sulphate Heptahydrate
FRAP	-	Ferric Reducing Antioxidant Power
GO	-	Gene Ontology
GROMACS	-	Groningen Machine for Chemical Simulations
GRX	-	Glutaredoxin
GST	-	Glutathione S-transferase
HMGB1	-	High Mobility Group Box-1
HSOS	-	Hepatic Sinusiodal Obstruction Syndrome
HSP	-	Heat Shock Protein
HTKT	-	Human TKT
IAA	-	Iodoacetamide
IEF	-	Isoelectric Focusing
IPG	-	Immobilized pH Gradient

I-TASSER	-	Iterative Threading Assembly Refinement
ITCs	-	Isothiocyanates
LC-MS/MS	-	Liquid Chromatography Tandem Mass Spectrometry
MD	-	Molecular Dynamics
MS	-	Mass Spectrometry
MS/MS	-	Tandem Mass Spectrometry
MTKT	-	Maize TKT
MW	-	Molecular Weight
NADPH	-	Nicotinamide Adenine Dinucleotide Phospahte
NCBI	-	National Center for Biotechnology Information
NF- κ B	-	Nuclear Factor kappa B
NMR	-	Nuclear Magnetic Resonance
OATKT	-	TKT from <i>O. aristatus</i>
PDB	-	Protein Data Bank
PPP	-	Pentose Phosphate Pathway
PRX	-	Peroxidase
R _g	-	Radius of Gyration
RMSD	-	Root Mean Square Deviation
RMSF	-	Root Mean Square Fluctuation
ROS	-	Reactive Oxygen Species
RSA	-	Radical Scavenging Activity
S.D	-	Standard Deviation
SDS	-	Sodium Dodecyl Sulphate
SDS-PAGE	-	Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis
SEM	-	Standard Error Mean
SM	-	Secondary Metabolism
SPC	-	Simple Point Charge
SPC/E	-	Extended Simple Point Charge
TEMED	-	Tetramethylethylenediamine
TIP3P	-	Transferable Intermolecular Potential 3P
TIP4P	-	Transferable Intermolecular Potential 4P
TKT	-	Transketolase

TPC	-	Total Phenolic Content
TPP	-	Thiamine Diphosphate
TPTZ	-	2,4,6-tri(2-pyridyl)-s-triazine
Tris-HCl	-	Tris Hydrochloride
UTM		Universiti Teknologi Malaysia
WHO	-	World Health Organization
YASARA	-	Yet Another Scientific Artificial Reality Application

LIST OF SYMBOLS

α		Alpha
Å	-	Angstrom
β		Beta
°C	-	Degree Celsius
<	-	Less than
%	-	Percent
Cl ⁻	-	Chloride ion
•OH	-	Hydroxyl Radical
Da	-	Dalton
IC ₅₀	-	Half of the Inhibition Concentration
K	-	Kelvin
K ⁺	-	Potassium ion
kDa	-	Kilodalton
M	-	Moles
µg	-	Microgram
µL	-	Microlitre
µm	-	Micrometre
mM	-	Millimoles
mmHg	-	Manometric Unit of Pressure
<i>m/z</i>	-	Mass/Charge
Na ⁺	-	Sodium ion
ng	-	Nanogram
nL	-	Nanolitre
nm	-	Nanometer
ns	-	Nanosecond
<i>pI</i>	-	Isoelectric Point
rpm	-	Revolution per Minute
U	-	Unit
V	-	Voltage
v/v	-	Volume per Volume

w/v - Weight per Volume

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CHAPTER 1

INTRODUCTION

1.1 Background of Study

In today's world, the medicinal plants have attracted scientists' interest to employ the bioactive compounds as primary sources for early drug discovery (Veeresham, 2012) due to the side effect of synthetic drugs. In recent years, there is a breakthrough in medical field where researchers found that protein or peptides from natural products could be used as therapeutic agent to treat diseases as they exhibit potent pharmacological activities such as plant protein and peptides possess anti-tumor activity (Tepkeeva *et al.*, 2008). At the same time, there is rapid growth in the consumption of herbal products as primary health care globally since public perceived herbal products as safe because they are naturally occurring and used for treatment from decades ago (Liu *et al.*, 2016). Thus, comprehensive authentication of herbs' components such as bioactive compounds and protein is necessary in order to ensure the quality of the medicines produced.

Orthosiphon aristatus (Blume) Miq is one of the famous traditional medicinal plants in Malaysia which is widely used as folk medicines for the treatment of many diseases such as diabetes and kidney problems (Chin *et al.*, 2008). Most of the studies related to the plant was mainly focused on its phytochemical content. The first paper of *O. aristatus* was published in 1973, focusing flavonoid (Matsuura *et al.*, 1973). Alshawsh *et al.* (2011) reported that the most important constituents from *O. aristatus* are nine lipophilic flavones, two flavonol glycosides and nine caffeic acid derivatives. The presence of phytochemical contents such as phenolic compounds were attributed to the pharmacological activities of the plant including diuretic, antidiabetic, antioxidant, anti-inflammatory, antihypertensive, antiobesity and antitumor activities (Adnyana *et al.*, 2013). Thus, the leaves of the plant had been commercialized as herbal product especially tea product aiming to benefit human health as people

awareness about the health benefits of herbal tea is increasing. Drying process is the common practice applied to the commercialized products in order to extend the products' shelf life. Despite of it, the drying process may cause side effect on food quality if improper drying technique was applied. Although many compounds from this plant have been isolated and studied, detailed proteomics study on both *O. aristatus* fresh and dried leaf remains unknown.

Proteomics can be defined as a large scale protein analysis from any organism, tissue or cell and now it becomes a powerful tool in the field of biology research as it is able to reveal details about how the protein function, regulation and the interaction of proteins (Mirzaei *et al.*, 2016). Until now, many proteome of medicinal plants' have been studied using two dimensional gel electrophoresis (2-DE) and tandem mass spectrometry including *Gynura procumbens* (Hew and Gam, 2011), *Piper sarmentosum* (Shim and Gam, 2012), *Clematis chinensis* (Ishtiaq *et al.*, 2014), *Gingko biloba* (Uvackova *et al.*, 2014), *Phyllanthus niruri* (Nail and Zin, 2015) and *Withania somnifera* (Singh *et al.*, 2017). The two-dimensional (2-DE) result or protein spot distribution pattern can be used as a reference map for quality control of different herbal drugs (Ishtiaq, 2013). Meanwhile, mass spectrometry analysis is able to explore the complete proteome of the plant. Usually, the main focus will be the defense and stress-related proteins as they may also contribute to the plant's medicinal value such as antibacterial and antioxidant activities (Shim and Gam, 2012).

Antioxidant sources are gaining high demands in market as many reports suggested that antioxidant compounds is able to eliminate free radicals or reactive oxygen species (ROS) in the body and thus can prevent oxidative damage-related diseases such as coronary heart disease, cancer, aging and Alzheimer's disease (Szymanska *et al.*, 2016). Antioxidant can be defined as any compound which can prevent, eliminate or delays oxidative damage in cells (Halliwell, 2007) through donation of proton or electron (Aruoma *et al.*, 1993). Diabetes is one of the diseases without cure even in today's world and the worst thing is the current treatments for diabetes such as insulin injection and antidiabetic drugs will cause side effects on human health. For example, metformin may cause gastrointestinal problems (Chaudhury *et al.*, 2017) and deficiency of vitamin B12 and folic acid that leading to

anemia and neuropathy (Fogelman *et al.*, 2017). Meanwhile, insulin injection may cause lipoatrophy and lipohypertrophy at sites of injection and allergy reaction (Chaudhury *et al.*, 2017). Hence, the researchers are trying to find alternative therapeutic approach from natural sources. To date, there are many antidiabetic proteins or peptides have been reported (Rajasekhar *et al.*, 2010; Mendes *et al.*, 2014; Admassu *et al.*, 2018). For instance, catalase and trehalose phosphorylase are the identified proteins from grey oyster mushroom that may associate in combating diabetes (Wahab *et al.*, 2014) as catalase is able to enhance insulin secretion and sensitization whereas trehalose phosphate plays a crucial role in starch, sucrose and glucose metabolism (Christian and Nidetzky, 1999). Besides, bean peptide from Pinto bean (Ngoh and Gan, 2016) and LAPSLPGKPKPD peptide from egg yolk protein (Zambrowicz *et al.*, 2015) are considered as antidiabetic peptides due to its possible role in inhibiting α -amylase and α -glucosidase respectively. Nevertheless, the potential of *O. aristatus* protein in combating diabetes is still not yet known.

Bioinformatics is gaining popularity in plant proteome studies in order to obtain detailed understanding of protein three-dimensional (3D) structure, protein-protein interaction and protein-ligand interaction. Molecular dynamics (MD) simulation analysis is always being used to study the behaviour of the protein at varied conditions. For example, MD simulation is able to reveal the structural flexibility of malate dehydrogenase from marine molluscs and how the protein can adapt to the different temperature conditions (Dong *et al.*, 2017). Transketolase (TKT) protein from *O. aristatus* is of particular interest because we believed that TKT may be a useful protein that can be used to prevent the side effects of diabetes based on its function which can inhibit three biochemical pathways implicated in the pathogenesis of hyperglycemia induced-vascular damage (hexosamine pathway, advanced glycation end product formation pathway and diacylglycerol-protein kinase c pathway) by converting fructose-6-phosphate and glyceraldehyde-3-phosphate into pentose-5-phosphate and other sugars (Hammes *et al.*, 2003). Nevertheless, the properties of TKT little known.

1.2 Problem Statement

O. aristatus is a plant known for its medicinal properties from both the traditional background and scientific research and is commonly known as Java tea. In Malaysia, *O. aristatus* has been used as traditional medicine especially the leaves in diabetes, urinary lithiasis, edema, hypertension, diuretic, inflammation, jaundice, eruptive fever, hepatitis, rheumatism, and as a remedy for kidney stones (Shafaei *et al.*, 2015). Due to its pharmacological properties, the plant has been widely produced as herbal products to maintain human health. To date, lots of studies have been done on the plant, majorly on its phytochemical contents and many chemical compounds especially the secondary metabolites of the plant were isolated and identified. Many studies reported that the pharmacological activities of the plant are associated with the presence of the phytochemical compounds (Arafat *et al.* 2008; Mohamed *et al.*, 2011; Al-Suede *et al.*, 2012; Ripin *et al.*, 2018). Nevertheless, protein is arising as a therapeutic source to treat human diseases nowadays because protein is able to be engineered to enhance its functionality, half-life and drug efficacy (Lutz, 2010; Tobin *et al.*, 2014). According to Lagasse *et al.* (2017), certain therapeutic protein drugs have been approved as therapies for the patients. Thus, it is important to explore the potent proteins' bioactivity especially protein from medicinal plants as the protein may be associated with its medicinal value. Yet, the information of *O. aristatus* protein is lacking although many studies have been done on the plant. It is worth to explore the *O. aristatus* protein due to the fact that protein can become an alternative source in producing medicines for diseases. Moreover, it is necessary to know the drying effect on *O. aristatus* protein since it has been highly produced as products in dry form.

1.3 Significance of Work

Through this study, protein profiling and protein expression profile of fresh and dried leaf were provided. It is valued to study the protein of *O. aristatus* because protein is the direct product from central dogma and involved in all the biological process as all the enzymes are protein. From the protein profiling of fresh and dried leaf, it can provide knowledge on the plant growth and development process and how

the plant cope with heat stress. In addition, the protein profile could also be useful for finding proteins with potent beneficial property and the expression profile could be used as a reference map for drug quality control. Besides, the antioxidant and antidiabetic activity of *O. aristatus* fresh and dried leaf protein were explored in this study using *in vitro* study. The results obtained were very crucial as it is revealed that proteins from *O. aristatus* also attributed to therapeutic value besides its phytochemical compounds. At the same time, it also provides insight that the antioxidant and antidiabetic activity of the protein was affected by drying process. Furthermore, potential protein with antidiabetic activity from the plant was studied further using bioinformatics analysis to explore its thermostability and docking mechanism with the ligand because the plant had been claimed to be effective in alleviate diabetes. This kind of insight may give a great impact in pharmaceutical industries as it could reveal the properties of the protein and thus can save time for the whole process of drug design.

1.4 Objectives of Research

The objectives of this study were:

- (a) To profile the protein of *O. aristatus* from fresh and manually dried leaf for further comparison of 2-DE expression.
- (b) To determine the antidiabetic activity of fresh and manually dried leaf protein from *O. aristatus*.
- (c) To predict thermostability and docking mechanism of selected protein with antidiabetic potential from profiled protein.

1.5 Scope of Research

The scope of this study is to profile and analyse protein of *O. aristatus* fresh and dried leaf. In this study, the protein from fresh and dried leaf (microwave-dried, freeze-dried and halogen oven-dried) were used for extraction by using modified phenol/SDS buffer with three preliminary washes. The protein obtained was quantified using Bradford assay and the quality was checked via sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) where its electrophoretic pattern was determined. Furthermore, the protein from fresh and dried leaf was further used to quantify antioxidant activity via 1,1-diphenyl-2-picrylhydrazyl (DPPH) and ferric reducing antioxidant power (FRAP) assays in order to determine the antioxidant activity of the protein. Then, the most suitable drying method for the leaf was chosen based on the protein quantity, quality and its antioxidant activity. After that, the protein from fresh and the best dried leaf was tryptic-digested and sent for liquid chromatography tandem mass spectrometry (LC-MS/MS). Spectral results obtained from LC-MS/MS was analyzed and searched against National Center for Biotechnology Information (NCBI) database. After that, the identified proteins were classified based on functional categorization of gene ontology (GO). To compare 2-DE expression pattern of fresh and dried leaf, the protein from fresh and dried leaf was run using 17 cm IPG strip, pH 4-7, 8000 focusing voltage. The number of protein spots and the expression level of the protein in fresh and dried leaf were compared using PDQuest software (Biorad, CA, USA). Apart from that, the antidiabetic activity of the fresh and dried leaf protein was determined in the study through *in vitro* studies via glucose diffusion, α -amylase and α -glucosidase inhibition. Lastly, TKT as one of the identified proteins with potential antidiabetic property was selected to perform molecular simulation at different temperatures (310 K, 368 K, 373 K, 423 K, 453K and 473 K) using GRONingen MACHine for Chemical Simulations (GROMACS) software version 5.0.4 to determine its thermostability as well as molecular docking with its ligand, TPP using Autodock in order to know the protein-ligand interaction.

REFERENCES

- Abd El-Kader, A. S. S., El-Dosouky, M. A., Abouwarda, A., Mohammad Abdel All, S. and Osman, M. I. (2012). Isolation, screening and identification and optimization of cultural conditions for selected local bacterial β -galactosidase producers. *Journal of Applied Science and Research*, 8(4), 2010-2017.
- Abdelwahab, S. I., Mohan, S., Mohamed E. M., Al-Mekhlafi, N., Mariod, A. A., Abdul, A. B., Abdulla, M. A. and Alkharfy, K. M. (2010). Antiapoptotic and antioxidant properties of *Orthosiphon stamineus* benth (Cat's Whiskers): intervention in the Bcl-2-mediated apoptotic pathway. *Evidence-Based Complementary and Alternative Medicine*, 2011(1), 1-11.
- Abdu, A., Aderis, N., Abdul Hamid, H., Majid, N. A., Muhamad, N., Jusop, S., Singh, K., Singh, D. and Ahmad, K. (2011). Using *Orthosiphon stamineus* B. for phytoremediation of heavy metals in soils amended with sewage sludge. *American Journal of Applied Sciences*, 8(4), 323-331.
- Abdullah, S., Ahmad, M. S., Shaari, A. R., Johar, H. M., & Noor, N. F. M. (2011). Drying characteristics and herbal metabolites composition of misai kucing (*Orthosiphon stamineus* Benth.) leaves. In *Proceedings of International Conference on Food Engineering and Biotechnology, 7th-9th May, Bangkok, Thailand*.
- Abdullah, S., Shaari, A. R. and Azimi, A. (2012). Effect of drying methods on metabolites composition of misai kucing (*Orthosiphon stamineus*) leaves. *APCBEE Procedia*, 2, 178-182.
- Abirami, A., Nagarani, G. and Siddhuraju, P. (2014). *In vitro* antioxidant, anti-diabetic, cholinesterase and tyrosinase inhibitory potential of fresh juice from *Citrus hystrix* and *C. maxima* fruits. *Food Science and Human Wellness*, 3(1), 16-25.
- Abraham, M. J., Murtola, T., Schulz, R., Páll, S., Smith, J. C., Hess, B. and Lindahl, E. (2015). GROMACS: High performance molecular simulations through multi-level parallelism from laptops to supercomputers. *SoftwareX*, 1(1), 19-25.

- Adam, Y., Somchit, M., Sulaiman, M., Nasaruddin, A., Zuraini, A., Bustamam, A. and Zakaria, Z. (2009). Diuretic properties of *Orthosiphon stamineus* Benth. *Journal of Ethnopharmacology*, 124(1), 154-158.
- Adisakwattana, S., Jiphimai, P., Prutanopajai, P., Chanathong, B., Sapwarabol, S. and Ariyapitipan, T. (2010). Evaluation of α -glucosidase, α -amylase and protein glycation inhibitory activities of edible plants. *International Journal of Food Science and Nutrition*, 61(3), 295-305.
- Adler, V., Yin, Z., Fuchs, S. Y., Benezra, M., Rosario, L., Tew, K. D., Pincus, M. R., Sardana, M., Henderson, C. J., Wolf, C. R., Davis, R. J. and Ronai, Z. (1999). Regulation of JNK signaling by GSTp. *The EMBO Journal*, 18(5), 1321-1334.
- Admassu, H., Gasmalla, M. A. A., Yang, R. and Zhao, W. (2018). Bioactive peptides derived from seaweed protein and their health benefits: Antihypertensive, antioxidant, and antidiabetic properties. *Journal of Food Science*, 83(1), 6-16.
- Adnyana, I. K., Setiawan, F. and Insanu, M. (2013). From ethnopharmacology to clinical study of *Orthosiphon stamineus* Benth. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 66-73.
- Agrahar-Murugkar, D. and Jha, K. (2010). Effect of drying on nutritional and functional quality and electrophoretic pattern of soyflour from sprouted soybean (*Glycine max*). *Journal of Food Science and Technology*, 47(5), 482-487.
- Agoreyo, B. O., Akpiroroh, O., Orukpe, O. A., Osaweren, O. R. and Owabor, C. N. (2011). The effects of various drying methods on the nutritional composition of *Musa paradisiaca*, *Dioscorea rotundata* and *Colocasia esculenta*. *Asian Journal of Biochemistry*, 6(6), 458-464.
- Ahrné, L., Pereira, N., Staack, N. and Floberg, P. (2007). Microwave convective drying of plant foods at constant and variable microwave power. *Drying Technology*, 25(7-8), 1149-1153.
- Akhtar, M. N. and Gayathri, M. (2016). Anti-oxidant, anti-microbial and glucose diffusion inhibition activities of the aqueous and chloroform extract of *Phyllanthus urinaria*. *International Journal of Pharmacy and Pharmaceutical Sciences*, 8(4), 278-280.
- Akowuah, G., Zhari, I., Norhayati, I. and Sadikun, A. (2005). Radical scavenging activity of methanol leaf extract of *Orthosiphon stamineus*. *Pharmaceutical Biology*, 42(8), 629-635.

- Akowuah, G., Zhari, I., Norhayati, I., Sadikun, A. and Khamsah, S. (2004). Sinensetin, eupatorin, 3'-hydroxy-5, 6, 7, 4'-tetramethoxyflavone and rosmarinic acid contents and antioxidative effect of *Orthosiphon stamineus* from Malaysia. *Food Chemistry*, 87(4), 559-566.
- Akpan, E. J. and Umoh, I. B. (2004). Effect of heat and tetracycline treatments on the food quality and acidity factors in cocoyam [*Xanthosoma sagittifolium* (L) Schott]. *Pakistan Journal of Nutrition*, 3(4), 240-243.
- Alam, I., Sharmin, S., Kim, K. H., Kim, Y. G., Lee, J. and Lee, B. H. (2013). An improved plant leaf protein extraction for high resolution two-dimensional polyacrylamide gel electrophoresis and comparative proteomics. *Biotechnic and Histochemistry*, 88(2), 61-75.
- Alban, A., David, S. O., Bjorkesten, L., Andersson, C., Sloge, E., Lewis, S. and Currie, I. (2003). A novel experimental design for comparative two-dimensional gel analysis: Two-dimensional difference gel electrophoresis incorporating a pooled internal standard. *Proteomics*, 3(1), 36-44.
- Ali, Z.M, Armugam, S. and Lazan, H. (1995) β -Galactosidase and its significance in ripening mango fruit. *Phytochemistry*, 38(5), 1109-1114.
- Alshawsh, M. A., Abdulla, M. A., Ismail, S. and Amin, Z. A. (2011). Hepatoprotective effects of *Orthosiphon stamineus* extract on thioacetamide-induced liver cirrhosis in rats. *Evidence-Based Complementary and Alternative Medicine*, 2011(1), 1-6.
- Al-Suede, F. S. R., Farsi, E., Ahamed, M. K. B., Ismail, Z., Majid, A. S. A. and Majid, A. M. S. A. (2014). Marked antitumor activity of cat's whiskers tea (*Orthosiphon stamineus*) extract in orthotopic model of human colon tumor in nude mice. *Journal of Biochemical Technology*, 3(5), 170-176.
- Ameer, O. Z., Salman, I. M., Asmawi, M. Z., Ibraheem, Z. O. and Yam, M. F. (2012). *Orthosiphon stamineus*: traditional uses, phytochemistry, pharmacology, and toxicology. *Journal of Medicinal Food*, 15(8), 678-690.
- Amira, A. B., Bauwens, J., De Pauw, E., Besbes, S., Attia, H., Francis, F. and Blecker, C. (2017). Identification of proteins from wild cardoon flowers (*Cynara cardunculus* L.) by a proteomic approach. *Journal of Chemical Biology*, 10(1), 25-33.

- Angel, G. R., Vimala, B. and Nambisan, B. (2013). Antioxidant and anti-inflammatory activities of proteins isolated from eight *Curcuma* species. *Phytopharmacology*, 4(1), 96-105.
- Angelino, D., Dosz, E. B., Sun, J., Hoeflinger, J. L., Van Tassell, M. L., Chen, P., Harnly, J. M., Miller, M. J. and Jeffery, E. H. (2015). Myrosinase-dependent and -independent formation and control of isothiocyanate products of glucosinolate hydrolysis. *Frontiers in Plant Science*, 6(1), 831.
- Anker, L. (1974). Auxin synthesis inhibition by sugars, notably by galactose. *Acta Botanica Neerlandica*, 23(1), 705-714.
- Antunes, D. A., Devaurs, D. and Kavraki, L. E. (2015). Understanding the challenges of protein flexibility in drug design. *Expert Opinion on Drug Discovery*, 10(12), 1301-1313.
- Arafat, O., Tham, S., Sadikun, A., Zhari, I., Haughton, P. and Asmawi, M. (2008). Studies on diuretic and hypouricemic effects of *Orthosiphon stamineus* methanol extracts in rats. *Journal of Ethnopharmacology*, 118(3), 354-360.
- Arcan, I. and Yemenicioglu, A. (2007). Antioxidant activity of protein extracts from heat-treated or thermally processed chickpeas and white beans. *Food Chemistry*, 103(2), 301-312.
- Aruoma, O. I., Murcia, A., Butler, J. and Halliwell, B. (1993). Evaluation of the antioxidant and prooxidant actions of gallic acid and its derivatives. *Journal of Agricultural and Food Chemistry*, 41(11), 1880-1885.
- Asad, B. S., Iqbal, M. M. and Kiranmai, M. (2012). Hepatoprotective activity of *Phyllanthus Amarus* seeds extracts in CCl₄ treated rats: *In vitro* and *in vivo*. *Global Journal of Medical Research*, 12(6), 38-49.
- Asraf, S. S. and Gunasekaran, P. (2010). Current trends of β -galactosidase research and application. *Current research, technology and education topics in applied microbiology and microbial biotechnology*. Microbiology book series Formatex Research Center, Spain, 880-890.
- Avigan, M., I., Mozersky, R., P. and Seeff, L., B. (2016). Scientific and regulatory perspectives in herbal and dietary supplement associated hepatotoxicity in the United States. *International Journal of Molecular Sciences*, 17(3), 331-361.
- Awale, S., Tezuka, Y., Banskota, A. H., Shimoji, S., Taira, K. and Kadota, S. (2002). Norstaminane- and isopimarane-type diterpenes of *Orthosiphon stamineus* from Okinawa. *Tetrahedron*, 58(27), 5503-5512.

- Awale, S., Tezuka, Y., Banskota, A. H., Adnyana, I. K. and Kadota, S. (2003a). Highly-oxygenated isopimarane-type diterpenes from *Orthosiphon stamineus* of Indonesia and their nitric oxide inhibitory activity. *Chemical and Pharmaceutical Bulletin*, 51(3), 268-275.
- Awale, S., Tezuka, Y., Banskota, A. H. and Kadota, S. (2003b). Siphonols A–E: novel nitric oxide inhibitors from *Orthosiphon stamineus* of Indonesia. *Bioorganic & Medicinal Chemistry Letters*, 13(1), 31-35.
- Awale, S., Tezuka, Y., Kobayashi, M., Ueda, J. Y. and Kadota, S. (2004). Neoorthosiphonone A; a nitric oxide (NO) inhibitory diterpene with new carbon skeleton from *Orthosiphon stamineus*. *Tetrahedron Letters*, 45(7), 1359-1362.
- Baginsky, S. (2009). Plant proteomics: concepts, applications, and novel strategies for data interpretation. *Mass Spectrometry Reviews*, 28(1), 93-120.
- Baheri, M. and Dayer, M. R. (2016). Temperature and pH Effects on Insulin Structure: A Molecular Dynamic Approach. *Jentashapir Journal of Health Research*, 7(4), e36931.
- Balasuriya, B. N. and Rupasinghe, H. V. (2011). Plant flavonoids as angiotensin converting enzyme inhibitors in regulation of hypertension. *Functional Foods in Health and Disease*, 1(5), 172-188.
- Bandaranayake, W. M. (2006). Quality control, screening, toxicity, and regulation of herbal drugs. *Modern phytomedicine: turning medicinal plants into drugs*, 25-57.
- Baptista, J. A. B., Tavares, J. F. D. and Carvalho, R. C. B. (1998). Comparison of catechins and aromas among different green teas using HPLC/SPME-GC. *Food Research International*, 31(10), 729–736.
- Basha, S. K. and Kumari, V. S. (2012). *In vitro* antidiabetic activity of *Psidium guajava* leaves extracts. *Asian Pacific Journal of Tropical Disease*, 2(1), 98-100.
- Basheer, M. K. A. and Majid, A. M. S. A. (2010). Medicinal potentials of *Orthosiphon stamineus* Benth. *WebmedCentral CANCER*, 1(12), 1-7.
- Basu, S. and Sen, S. (2009). Turning a mesophilic protein into a thermophilic one: a computational approach based on 3D structural features. *Journal of Chemical Information and Modeling*, 49(7), 1741-1750.

- Basu, S. and Sen, S. (2013). Do homologous thermophilic–mesophilic proteins exhibit similar structures and dynamics at optimal growth temperatures? A molecular dynamics simulation study. *Journal of Chemical Information and Modeling*, 53(2), 423-434.
- Baudin, B. (2012). Two-Dimensional Gel Electrophoresis (2-DE). In *Gel Electrophoresis-Principles and Basics*. London, United Kingdom: InTechOpen.
- Bello, M. O., Ibrahim, A. O., Ogunwande, I. A. and Olawore, N. O. (2004). Heavy trace metals and macronutrients status in herbal plants of Nigeria. *Food Chemistry*, 85(1), 67-71.
- Berendsen, H., J., C., Grigera, J., R. and Straatsma, T., P. (1987). The missing term in effective pair potentials. *Journal of Physical Chemistry*, 91(24), 6269-6271.
- Berggren, K., Chernokalskaya, E., Steinberg, T. H., Kemper, C., Lopez, M. F., Diwu, Z., Haugland, R. P. and Patton, W. F. (2000). Background-free, high sensitivity staining of proteins in one- and two-dimensional sodium dodecyl sulfate-polyacrylamide gels using a luminescent ruthenium complex. *Electrophoresis*, 21(12), 2509-2521.
- Berrone, E., Beltramo, E., Solimine, C., Ape, A. U. and Porta, M. (2006). Regulation of intracellular glucose and polyol pathway by thiamine and benfotiamine in vascular cells cultured in high glucose. *Journal of Biological Chemistry*, 281(14), 9307-9313.
- Bevevino, L. H. and Aires, M. M. (1994). Effect of crude extract of roots of *Bredemeyera floribunda* Willd. II. Effect on glomerular filtration rate and renal tubular function of rats. *Journal of Ethnopharmacology*, 43(3), 203-207.
- Beyer, K., Grishina, G., Bardina, L., Grishin, A. and Sampson, H., A. (2002). Identification of an 11S globulin as a major hazelnut food allergen in hazelnut-induced systemic reactions. *Journal of Allergy and Clinical Immunology*, 110(3), 517-523.
- Boguth, G., Harder, A., Scheibe, B., Wildgruber, R. and Weiss, W. (2000). The current state of two-dimensional electrophoresis with immobilized pH gradients. *Electrophoresis*, 21(6), 1037-1053.
- Bohm, H. J. (1992). The computer program LUDI: a new method for the de novo design of enzyme inhibitors. *Journal of Computer-Aided Molecular Design*, 6(1), 61-78.

- Bokesch, H. R., Pannell, L. K., Cochran, P. K., Sowder, R. C., McKee, T. C. and Boyd, M. R. (2001). A novel anti-hiv macrocyclic peptide from *Palicourea condensata*. *Journal of Natural Products*, 64(2), 249-250.
- Boumendjel, M. E. and Boutebba, A. (2003). Heat treatment effects on the biochemical and nutritional content of double concentrate tomato paste. *Acta Horti*, 613(1), 429-432.
- Bonneau, R. and Baker, D. (2001). *Ab initio* protein structure prediction: progress and prospects. *Annual Review of Biophysics and Biomolecular Structure*, 30(1), 173-189.
- Bouley, J., Meunier, B., Chambon, C., De Smet, S., Hocquette, J., F. and Picard, B. (2005). Proteomic analysis of bovine skeletal muscle hypertrophy. *Proteomics*, 5(2), 490-500.
- Bowie, J. U., Luthy, R. and Eisenberg, D. (1991). A method to identify protein sequences that fold into a known three-dimensional structure. *Science*, 253(5016), 164-170.
- Boyle, L., Wamelink, M. M., Salomons, G. S., Roos, B., Pop, A., Dauber, A., Hwa, V., Andrew, M., Douglas, J. and Feingold, M. (2016). Mutations in TKT are the cause of a syndrome including short stature, developmental delay, and congenital heart defects. *The American Journal of Human Genetics*, 98(6), 1235-1242.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1-2), 248-254.
- Bray, T. M. (1999). Antioxidants and oxidative stress in health and disease: Introduction. *Proceedings of the Society for Experimental Biology and Medicine*, 222(3), 195-195.
- Brechenmacher, L., Lee, J., Sachdev, S., Song, Z., Nguyen, T. H. N., Joshi, T., Oehrle, N., Libault, M., Mooney, B., Xu, D., Cooper, B. and Stacey, G. (2009). Establishment of a protein reference map for soybean root hair cells. *Plant Physiology*, 149(2), 670-682.
- Brenelli, S., L., Campos, S. D. and Saad, M. J. (1997). Viscosity of gums *in vitro* and their ability to reduce postprandial hyperglycemia in normal subjects. *Brazilian Journal of Medical and Biological Research*, 30(12), 1437-1440.

- Brownlee, M., Vlassara, H., and Cerami, A. (1984). Nonenzymatic glycosylation and the pathogenesis of diabetic complications. *Annals of Internal Medicine*, 101(1), 527-537.
- Bruno, G., Runzo, C., Cavallo-Perin, P., Merletti, F., Rivetti, M., Pinach, S., Novelli, G., Trovati, M., Cerutti, Pagano, G. and Piedmont Study Group for Diabetes Epidemiology. (2005). Incidence of type 1 and type 2 diabetes in adults aged 30–49 years: The population-based registry in the province of Turin, Italy. *Diabetes Care*, 28(11), 2613-2619.
- Bull, H. B. and Breese, K. (1973). Thermal transitions of proteins. *Archives of Biochemistry and Biophysics*, 156(2), 604-612.
- Burkhart, J. M., Schumbrutzki, C., Wortelkamp, S., Sickmann, A. and Zahedi, R. P. (2012). Systemic and quantitative comparison of digest efficiency and specificity reveals the impact of trypsin quality on MS-based proteomics. *Journal of Proteomics*, 75(4), 1454-1462.
- Bushra, R. and Aslam, N. (2010). An overview of clinical pharmacology of ibuprofen. *Oman Medical Journal*, 25(3), 1550-1661.
- Butterfield, D. A., Perluigi, M. and Sultana, R. (2006). Oxidative stress in Alzheimer's disease brain: new insights from redox proteomics. *European journal of pharmacology*, 545(1), 39-50.
- Cai, L., Wu, X., Zhang, Y., Li, X., Ma, S. and Li, J. (2015). Purification and characterization of three antioxidant peptides from protein hydrolysate of grass carp (*Ctenopharyngodon idella*) skin. *Journal of Functional Foods*, 16(1), 234-242.
- Cai, X., Xiao, C., Xue, H., Xiong, H., Hang, Y., Xu, J. and Lu, Y. (2018). A comparative study of the antioxidant and intestinal protective effects of extracts from different parts of Java tea (*Orthosiphon stamineus*). *Food Science & Nutrition*, 6(3), 579-584.
- Calahan, J., Howard, D., Almalki, A. J., Gupta, M. P. and Calderon, A. I. (2016). Chemical adulterants in herbal medicinal products: A review. *Planta Medica*, 82(6), 505-515.
- Calligaris, D., Villard, C. and Lafitte, D. (2011). Advances in top-down proteomics for disease biomarker discovery. *Journal of Proteomics*, 74 (7), 9209-34.

- Camilloni, C., Bonetti, D., Morrone, A., Giri, R., Dobson, C. M., Brunori, M., Gianni, S. and Vendruscolo, M. (2016). Towards a structural biology of the hydrophobic effect in protein folding. *Scientific reports*, 6(1), 28285-28294.
- Canovas, F. M., Dumas-Gaudot, E., Recorbet, G., Jorin, J., Mock, H. P. and Rossignol, M. (2004). Plant proteome analysis. *Proteomics*, 4(2), 285-298.
- Carpentier, S. C., Witters, E., Laukens, K., Deckers, P., Swennen, R. and Panis, B. (2005). Preparation of protein extracts from recalcitrant plant tissues: an evaluation of different methods for two-dimensional gel electrophoresis analysis. *Proteomics*, 5(10), 2497-2507.
- Carrasco-Castilla, J., Hernandez-Alvarez, A. J., Jimenez-Martinez, C., Gutierrez-Lopez, G. F. and Davila-Ortiz, G. (2012). Use of proteomics and peptidomics methods in food bioactive peptide science and engineering. *Food Engineering Reviews*, 4(4), 224-243.
- Carstensen, B., Jorgensen, M. E. and Friis, S. (2014). The epidemiology of diabetes and cancer. *Current Diabetes Report*, 14(10), 535-535.
- Carter, C., Pan, S., Zouhar, J., Avila, E. L., Girke, T. and Raikhel, N. V. (2004). The vegetative vacuole proteome of *Arabidopsis thaliana* reveals predicted and unexpected proteins. *The Plant Cell*, 16(12), 3285-3303.
- Cereto-Massagué, A., Ojeda, M. J., Joosten, R. P., Valls, C., Mulero, M., Salvado, M. J., Arola-Arnal, A., Arola, L., Garcia-Vallvé, S. and Pujadas, G. (2013). The good, the bad and the dubious: VHELIBS, a validation helper for ligands and binding sites. *Journal of Cheminformatics*, 5(1), 36-44.
- Chaitanya, M., Babajan, B., Anuradha, C., Naveen, M., Rajasekhar, C., Madhusudana, P. and Kumar, C. S. (2010). Exploring the molecular basis for selective binding of *Mycobacterium tuberculosis* Asp kinase toward its natural substrates and feedback inhibitors: a docking and molecular dynamics study. *Journal of Molecular Modeling*, 16(8), 1357-1367.
- Chakrabarti, S., Jahandideh, F. and Wu, J. (2014). Food-derived bioactive peptides on inflammation and oxidative stress. *BioMed Research International*, 2014(1), 608979-608990.
- Chan, E. W. and Lim, Y. Y. (2006). Antioxidant activity of *Thunbergia laurifolia* tea. *Journal of Tropical Forest Science*, 18(2), 130-136.

- Chan, E. W., Lim, Y. Y., Wong, S. K., Lim, K. K., Tan, S. P., Lianto, F. S. and Yong, M. Y. (2009). Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. *Food Chemistry*, 113(1), 166-172.
- Chan, E. W., Lye, P. Y., Eng, S. Y. and Tan, Y. P. (2013). Antioxidant properties of herbs with enhancement effects of drying treatments: A synopsis. *Free Radicals and Antioxidants*, 3(1), 2-6.
- Chan, K. (2003). Some aspects of toxic contaminants in herbal medicines. *Chemosphere*, 52(9), 1361-1371.
- Chandramouli, K. and Qian, P. Y. (2009). Proteomics: challenges, techniques and possibilities to overcome biological sample complexity. *Human Genomics and Proteomics*, 2009(1), 1-22.
- Chang, B. S., and Patro, S. Y. (2004). Freeze-drying process development for protein pharmaceuticals. *Lyophilization of Biopharmaceuticals*, 2(1), 113-138.
- Chaplin, M. (2010). Water's hydrogen bond strength. In *Water and life: The unique properties of H₂O* (pp. 69-86). CRC Press, Boca Raton, FL.
- Chaudhury, A., Duvoor, C., Reddy Dendi, V. S., Kraleti S., Chada, A., Ravilla, R., Marco, A., Shekhawat, N. S., Montales, M. T., Kuriakose, K., Sasapu, A., Beebe, A., Patil, N., Musham, C. K., Lohani, G. P. and Mirza, W. (2017). Clinical review of antidiabetic drugs: Implications for type 2 diabetes mellitus management. *Frontiers in Endocrinology*, 8(6), 1-12.
- Chen, L., Tuo, B. and Dong, H. (2016). Regulation of intestinal glucose absorption by ion channels and transporter. *Nutrients*, 8(1), 43-43.
- Chen, S. and Harmon, A. C. (2006). Advances in plant proteomics. *Proteomics*, 6(20), 5504-5516.
- Chen, Y., Liu, W., Xue, J., Yang, J., Chen, X., Shao, Y., Kwok, L., Y., Bilige, M., Mang, L. and Zhang, H. (2014). Angiotensin-converting enzyme inhibitory activity of *Lactobacillus helveticus* strains from traditional fermented dairy foods and antihypertensive effect of fermented milk of strain H9. *Journal of Dairy Science*, 97(11), 6680–6692.
- Chen, Z., Fu, Y., Xu, W. and Li, M. (2013). Molecular dynamics simulation of barnase: contribution of noncovalent intramolecular interaction to thermostability. *Mathematical Problems in Engineering*, 2013(2013), 1-12.

- Cheng, H. T., Hsieh, S. Y., Sung, C. M., Pai, B. C. J., Liu, N. J. and Chen, C. P. (2016). Optimizing human bile preparation for two-dimensional gel electrophoresis. *BioMed Research International*, 2016(2016), 1-6.
- Chi, C., F., Hu, F., Y., Wang, B., Li, T. and Ding, G., F. (2015). Antioxidant and anticancer peptides from the protein hydrolysate of blood clam (*Tegillarca granosa*) muscle. *Journal of Functional Foods*, 15(1), 301-313.
- Chin, J., Abas, H. and Sabariah, I. (2008). Toxicity study of *Orthosiphon stamineus* Benth (misai kucing) on sprague dawley rats. *Tropical Biomedicine*, 25(1), 9-16.
- Chinta, S. J. and Andersen, J. K. (2008). Redox imbalance in Parkinson's disease. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1780(11), 1362-1367.
- Cho, N. H., Shaw, J. E., Karuranga, S., Huang, Y., da Rocha Fernandes, J. D., Ohlrogge, A. W., and Malanda, B. (2018). IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and projections for 2045. *Diabetes Research and Clinical Practice*, 138(2018), 271-281.
- Chong, K. L. and Lim, Y. Y. (2011). Effects of drying on the antioxidant properties of herbal tea from selected *Vitex* species. *Journal of Food Quality*, 35(1), 51-59.
- Choong, Y. S., Tye, G. J. and Lim, T. S. (2013). Minireview: Applied structural bioinformatics in proteomics. *The Protein Journal*, 32(7), 505-511.
- Christian, E. I. S. and Nidetzky, B. (1999). Characterization of trehalose phosphorylase from *Schizopyllum commune*. *Biochemical Journal*, 341(2), 385-393.
- Chua, K. and Chou, S. (2003). Low-cost drying methods for developing countries. *Trends in Food Science & Technology*, 14(12), 519-528.
- Chung, F. L., Kelloff, G., Steele, V., Pittman, B., Zang, E., Jiao, D., Rigotty, J., Choi, C. I. and Rivenson, A. (1996). Chemopreventive efficacy of arylalkyl isothiocyanates and N-acetylcysteine for lung tumorigenesis in Fischer rats. *Cancer Research*, 56(4), 772-778.
- Clifton, J., G., Huang, F., Kovac, S., Yang, X., Hixson, D., C. and Josic, D., J. (2009). Proteomic characterization of plasma-derived clotting factor VIII-von Willebrand facotr concentrates. *Electrophoresis*, 30(20), 3636-3646.
- Colovos, C. and Yeates, T. O. (1993). Verification of protein structures: patterns of nonbonded atomic interactions. *Protein Science*, 2(9), 1511-1519.

- Conaway, C. C., Getahun, S. M., Liebes, L. L., Pusateri, D. J., Topham, D. K., Botero-Omary, M. and Chung, F. L. (2000). Disposition of glucosinolates and sulforaphane in humans after ingestion of steamed and fresh broccoli. *Nutrition and Cancer*, 38(2), 168–178.
- Costantini, S., Colonna, G. and Facchiano, A. M. (2008). ESBRI: A web server for evaluating salt bridges in proteins. *Bioinformatics*, 3(3), 137-138.
- Cristoni, S., & Mazzuca, S. (2011). *Bioinformatics Applied to Proteomics*. London, United Kingdom: INTECH Open Access Publisher.
- D'Alessandro, A. and Zolla, L. (2012). We are what we eat: Food safety and proteomics. *Journal of Proteome Research*, 11(1), 26-36.
- Daliri, E., B., M., Oh, D., H. and Lee, B. H. (2017). Bioactive Peptides. *Foods*, 6(32), 1-21.
- Dall-Donne, I., Scaloni, A., Giustarini, D., Cavarra, E., Tell, G., Lungarella, G., Colombo, R., Rossi, R. and Milzani, A. (2005). Proteins as biomarkers of oxidative/nitrosative stress in diseases: the contribution of redox proteomics. *Mass Spectrometry Reviews*, 24(1), 55-99.
- Damerval, C., De Vienne, D., Zivy, M. and Thiellement H. (1986). Technical improvements in two-dimensional electrophoresis increase the level of genetic variation detected in wheat seedling proteins. *Electrophoresis*, 7(1), 52-54.
- Danaei, G., Lawes, C. M., Vander, H. S., Murray, C. J. and Ezzati, M. (2006). Global and regional mortality from ischaemic heart disease and stroke attributable to higher than optimum blood glucose concentration: comparative risk assessment. *Lancet*, 368(9548), 1651-1659.
- Danwilai, K., Konmun, J., Sripanidkulchai, B. O. and Subongkot, S. (2017). Antioxidant activity of ginger extract as a daily supplement in cancer patients receiving adjuvant chemotherapy: a pilot study. *Cancer Management and Research*, 9(1), 11-18.
- Dasgupta, A. (2011). *Effects of herbal supplements on clinical laboratory test results* (Vol. 2). Walter de Gruyter.
- Dawid-Pač, R. (2013). Medicinal plants used in treatment of inflammatory skin diseases. *Advances in Dermatology and Allergology*, 30(3), 170-177.
- De Smet, P. A. (1995). Health risks of herbal remedies. *Drug Safety*, 13(2), 81-93.
- De Smet, P. A. (1997). Adverse effects of herbal remedies. *Adverse Drug Reaction Bulletin*, 183(1), 695-698.

- Del Guerra, S., Lupi, R., Marselli, L., Masini, M., Bugliani, M., Sbrana, S., Torri, S., Pollera, M., Bogi, U., Mosca, F. and Del Prato, S. (2005). Functional and molecular defects of pancreatic islets in human type 2 diabetes. *Diabetes*, 54(3), 727-735.
- Delahunty, C. and Yates Iii, J. R. (2005). Protein identification using 2d-lc-ms/ms. *Methods*, 35(3), 248-255.
- Dellafiora, L., Paoletta, S., Dall'Asta, C., Dossena, A., Cozzini, P. and Galaverna, G. (2015). Hybrid *in silico/in vitro* approach for the identification of angiotensin I converting enzyme inhibitory peptides from parma dry-cured ham. *Journal of Agricultural and Food Chemistry*, 63(28), 6366–6375.
- Dev, S. R. and Raghavan, V. G. (2012). Advancements in drying techniques for food, fiber, and fuel. *Drying Technology*, 30(11-12), 1147-1159.
- Dharmaraj, S., Hossain, M. A., Zhari, S., Harn, G. L. and Ismail, Z. (2006). The use of principal component analysis and self-organizing map to monitor inhibition of calcium oxalate crystal growth by *Orthosiphon stamineus* extract. *Chemometrics and Intelligent Laboratory Systems*, 81(1), 21-28.
- Di Simplicio, P., Giorgio, L. D., Cardaioli, E., Lecis, R., Miceli, M., Rossi, R., Anichini, R., Mian, M., Seghieri, G. and Franconi, F. (1995). Glutathione, glutathione utilizing enzymes and thioltransferase in platelets of insulin-dependent diabetic patients: relation with platelet aggregation and with microangiopathic complications. *European journal of clinical investigation*, 25(9), 665-669.
- Dinkova-Kostova, A. T. and Kostov, R. V. (2012). Glucosinolates and isothiocyanates in health and disease. *Trends in Molecular Medicine*, 18(6), 337-347.
- Dohare, P., Garg, P., Jagannathan, N. and Ray, M. (2008). Neuroprotective efficacy and therapeutic window of curcuma oil: in rat embolic stroke model. *BMC Complementary and Alternative Medicine*, 8(55), 1-20.
- Dominy, B. N., Minoux, H. and Brooks, C. L. (2004). An electrostatic basis for the stability of thermophilic proteins. *Proteins: Structure, Function, and Bioinformatics*, 57(1), 128-141.
- Dong, Y. W., Liao, M. L., Meng X. L. and Somero, G. N. (2017). Structural flexibility and protein adaptation to temperature: Molecular dynamics analysis of malate dehydrogenases of marine molluscs. *Proceedings of the National Academy of Sciences*, 115(6), 1274-1279.

- Dorn, M., E Silva, M. B., Buriol, L. S. and Lamb, L. C. (2014). Three-dimensional protein structure prediction: methods and computational strategies. *Computational Biology and Chemistry*, 53(1), 251-276.
- Drew, A. K. and Myers, S. P. (1997). Safety issues in herbal medicine: implications for the health professions. *The Medical Journal of Australia*, 166(10), 538-541.
- Du, X., Sang, P., Xia, Y. L., Li, Y., Liang, J., Ai, S. M., Ji, X. L., Fu, Y. X. and Liu, S. Q. (2017). Comparative thermal unfolding study of psychrophilic and mesophilic subtilisin-like serine proteases by molecular dynamics simulations. *Journal of Biomolecular Structure and Dynamics*, 35(7), 1500-1517.
- Dumas, M., E., Canlet, C., Debrauwer, L., Martin, P. and Paris, A. (2005). Selection of biomarkers by a multivariate statistical processing of composite metabonomic data sets using multiple factor analysis. *Journal of Proteome Research*, 4(5), 1485–1492.
- Dunker, A. K., Brown, C. J., Lawson, J. D., Iakoucheva, L. M., and Obradovic, Z. (2002). Intrinsic disorder and protein function. *Biochemistry*, 41(21), 6573-6582.
- Dunnick, J. K. and Nyska, A. (2013). The toxicity and pathology of selected dietary herbal medicines. *Toxicologic Pathology*, 41(2), 374-386.
- Edgar, J., Colegate, S., Boppre, M. and Molyneux, R. (2011). Pyrrolizidine alkaloids in food: a spectrum of potential health consequences. *Additives & Contaminants: Part A*, 28(3), 308-324.
- Edwards, I. R. and Aronson, J. K. (2000). Adverse drug reactions: definitions, diagnosis, and management. *The lancet*, 356(9237), 1255-1259.
- Efferth, T. and Kaina, B. (2011). Toxicities by herbal medicines with emphasis to traditional chinese medicine. *Current Drug Metabolism*, 12(10), 989-996.
- Efferth, T., Romero, M., R., Wolf, D., G., Stamminger, T., Marin, J., J. and Marschall, M. (2008). The antiviral activities of artemisinin and artesunate. *Clinical Infectious Diseases*, 47(6), 804–808.
- Ekor, M. (2014). The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in Pharmacology*, 4(1), 177-187.
- Elcock, A. H. (1998). The stability of salt bridges at high temperatures: implications for thermophilic proteins. *Journal of Molecular Biology*, 284(2), 489-502.

- Emerging Risk Factors Collaboration, Sarwar, N., Gao, P., Seshasai, S. R., Gobin, R., Kaptoge, S., Di Angelantonio, E., Ingelsson, E., Lawlor, D. A., Selvin, E., Stampfer, M., Stehouwer, C. D., Lewington, S., Pennells, L., Thompson, A., Sattar, N., White, I. R., Ray, K. K. and Danesh, J. (2010). Diabetes mellitus, fasting blood glucose concentration, and risk of vascular disease: a collaborative meta-analysis of 102 prospective studies. *Lancet*, 375(9733), 2215-2222.
- Ernst, E. (2002). Heavy metals in traditional Indian remedies. *European Journal of Clinical Pharmacology*, 57(12), 891-896.
- Ernst, E. (2006). Review article: Methodological aspects of Traditional Chinese Medicine (TCM). *Annals Academy of Medicine Singapore*, 35(11), 773–774.
- Espinoza, E., O., Mann, M., J. and Bleasdel, B. (1995). Arsenic and mercury in traditional Chinese herbal balls. *The New England Journal of Medicine*, 333(12), 803-804.
- Evans, J. M., Newton, R. W., Ruta, D. A., MacDonald, T. M. and Morris, A D. (2000). Socio-economic status, obesity and prevalence of Type 1 and Type 2 diabetes mellitus. *Diabetes Medicine: A Journal of the British Diabetic Association*, 17(6), 478-480.
- Fabek, H., Messerschmidt, S., Brulport, V. and Goff, H. D. (2014). The effect of *in vitro* digestive processes on the viscosity of dietary fibres and their influence on glucose diffusion. *Food Hydrocolloids*, 35(1), 718-726.
- Fahey, J. W., Zalcmann, A. T. and Talalay, P. (2001). The chemical diversity and distribution of glucosinolates and isothiocyanates among plants. *Phytochemistry*, 56(1), 5-51.
- Fahey, J. W., Wehage, S. L., Holtzclaw, W. D., Kensler, T. W., Egner, P. A., Shapiro, T. A. and Talalay, P. (2012). Protection of humans by plant glucosinolates: efficiency of conversion of glucosinotes into isothiocyanates by the gastrointestinal microflora. *Cancer Prevention Research*, 5(4), 603-611.
- Fakhruddin, S., Alanazi, W. and Jackson, K. E. (2017). Diabetes-induced reactive oxygen species: mechanism of their generation and role in renal injury. *Journal of Diabetes Research*, 2017(1), 8379327-8379357.
- Falade, K. O. and Igbeka, J. C. (2007). Osmotic dehydration of tropical fruits and vegetables. *Food Reviews International*, 23(4), 373-405.

- Farre, G., Twyman, R. M., Christou, P., Capell, T. and Zhu, C. (2015). Knowledge-driven approaches for engineering complex metabolic pathways in plants. *Current Opinion in Biotechnology*, 32(1), 54-60.
- Faurobert, M., Pelpoir, E., & Chaïb, J. (2007). Phenol extraction of proteins for proteomic studies of recalcitrant plant tissues. In *Plant Proteomics* (pp. 9-14). Humana Press.
- Ferreira, L. G., dos Santos, R. N., Oliva, G. and Andricopulo, A. D. (2015). Molecular docking and structure-based drug design strategies. *Molecules*, 20(7), 13384-13421.
- Firenzuoli, F. and Gori, L. (2007). Herbal medicine today: clinical and research issues. *Evidence-Based Complementary and Alternative Medicine*, 4(S1), 37-40.
- FitzGerald, R. J., Murray, B. A. and Walsh, D. J. (2004). Hypotensive peptides from milk proteins. *The Journal of Nutrition*, 134(4), 980-988.
- Florence, C. and Stephan, G. (2002). Temperature-dependence of protein hydrogen bond properties as studied by high-resolution NMR. *Journal of Molecular Biology*, 317(5), 739-752.
- Franco, O. L., Rigden, D. J., Melo, F. R. and Grossi-De-Sá, M. F. (2002). Plant alpha-amylase inhibitors and their interaction with insect alpha-amylases. *European Journal of Biochemistry*, 269(2), 397-412.
- Fogelman, Y., Kitai, E., Blumberg, G., Golan-Cohen, A., Rapoport, M. and Carmeli, E. (2017). Vitamin B12 screening in metformin-treated diabetics in primary care: were elderly patients less likely to be tested?. *Aging Clinical and Experimental Research*, 29(2), 135-139.
- Fred-Jaiyesimi, A., Kio, A. and Richard, W. (2009). α -Amylase inhibitory effect of 3 β -olean-12-en-3-yl (9Z)-hexadec-9-enoate isolated from *Spondias mombin* leaf. *Food Chemistry*, 116(1), 285–288.
- Fullam, E., Pojer, F., Bergfors, T., Jones, T. A. and Cole, S. T. (2012). Structure and function of the transketolase from *Mycobacterium tuberculosis* and comparison with the human enzyme. *Open Biology*, 2(1), 110026-110038.
- Gallagher, A., Flatt, P., Duffy, G. and Abdel-Wahab, Y. (2003). The effects of traditional antidiabetic plants on *in vitro* glucose diffusion. *Nutrition Research*, 23(3), 413-424.

- Galvani, M., Hamdan, M. and Righetti, P., G. (2001a). Two-dimensional gel electrophoresis/ matrix-assisted laser desorption ionization mass spectrometry of commercial bovine milk. *Rapid Communication in Mass Spectrometry*, 15, 258–264.
- Galvani, M., Rovatti, L., Hamdan, M., Herbert, B. and Righetti, P. G. (2001b). Protein alkylation in the presence/absence of thiourea in proteome analysis: A matrix assisted laser desorption/ionization-time of flight-mass spectrometry investigation. *Electrophoresis*, 22(10), 2066-2074.
- Gan, R. Y., Xu, X. R., Song, F. L., Kuang, L. and Li, H. B. (2010). Antioxidant activity and total phenolic content of medicinal plants associated with prevention and treatment of cardiovascular and cerebrovascular diseases. *Journal of Medicinal Plants Research*, 4(22), 2438-2444.
- Garcia, M. C., Puchalska, P., Esteve, C. and Marina, M. L. (2013). Vegetable foods: A cheap source of proteins and peptides with antihypertensive, antioxidant, and other less occurrence bioactivities. *Talanta*, 106(2013), 328-349.
- Gaso-Sokac, D., Kovac, S. and Josic, D. (2010). Application of proteomics in food technology and food biotechnology: process development, quality control and product safety. *Food Technology and Biotechnology*, 48(3), 284-295.
- Gasteiger, E., Hoogland, C., Gattiker, A., Wilkins, M. R., Appel, R. D., & Bairoch, A. (2005). Protein identification and analysis tools on the ExPASy server. In *The proteomics protocols handbook* (pp. 571-607). Humana press.
- Giles, G. I. (2006). The redox regulation of thiol dependent signaling pathways in cancer. *Current pharmaceutical design*, 12(34), 4427-4443.
- Gills, J. J., Jeffery, E. H., Matusheski, N. V., Moon, R. C., Lantvit, D. D. and Pezzuto, J. M. (2006). Sulforaphane prevents mouse skin tumorigenesis during the stage of promotion. *Cancer Letters*, 236(1), 72-79.
- Gobbetti, M., Minervini, F., & Rizzello, C. G. (2007). Bioactive peptides in dairy products. *Handbook of Food Products Manufacturing*, 489-517.
- Gorg, A., Weiss, W. and Dunn, M. J. (2004). Current two-dimensional electrophoresis technology for proteomics. *Proteomics*, 4(12), 3665-3685.
- Görg, A., Klaus, A., Lück, C., Weiland, F., & Weiss, W. (2007). Two-dimensional electrophoresis with immobilized pH gradients for proteome analysis. *A laboratory manual*.

- Gorjanović, S., Beljanski, M. V., Gavrović-Jankulović, M., Gojgić-Cvijović, G., Pavlović, M. D. and Bejosano, F. (2007). Antimicrobial activity of malting barley grain thaumatin-like protein isoforms, S and R. *Journal of the Institute of Brewing*, 113(2), 206-212.
- Gray, A. M. and Flatt, P. R. (1997). Pancreatic and extra-pancreatic effects of the traditional anti-diabetic plant, *Medicago sativa* (lucerne). *The British Journal of Nutrition*, 78(2), 325-334.
- Gray, A. M. and Flatt, P. R. (1998). Actions of the traditional antidiabetic plant, *Agrimony eupatoria* (agrimony): effects on hyperglycaemia, cellular glucose metabolism and insulin secretion. *The British Journal of Nutrition*, 80(1), 109-114.
- Gray, A. M., Abdel-Wahab, Y. H. and Flatt, P. R. (2000). The traditional plant treatment, *Sambucus nigra* (elder), exhibits insulin-like and insulin-releasing actions *in vitro*. *The Journal of Nutrition*, 130(1), 15-20.
- Grebe, S. K. and Singh, R. J. (2011). LC-MS/MS in the clinical laboratory—Where to from here?. *The Clinical Biochemist Reviews*, 32(1), 5-31.
- Grinter, S. Z. and Zou, X. (2014). Challenges, applications, and recent advances of protein-ligand docking in structure-based drug design. *Molecules*, 19(7), 10150-10176.
- Gross, K. C. and Wallner, S. J. (1979). Degradation of cell wall polysaccharides during tomato fruit ripening. *Plant Physiology*, 63(1), 117-120.
- Gross, K. C. (1984). Fractionation and partial characterization of cell walls from normal and non-ripening mutant tomato fruit. *Physiologia Plantarum*, 62(1), 25-32.
- Guine, R. (2018). The drying of foods and its effect on the physical-chemical, sensorial and nutritional properties. *International Journal of Food Engineering*, 2(4), 93-100.
- Gupta, R., Gigras, P., Mohapatra, H., Goswami, V. K. and Chauhan, B. (2003). Microbial α -amylases: A biotechnological perspective. *Process Biochemistry*, 38(11), 1599-1616.

- Gurgel, A. P. A. D., da Silva, J. G., Grangeiro, A. R. S., Oliveira, D. C., Lima, C. M., da Silva, A. C., Oliveira, R. A. and Souza, I. A. (2009). *In vivo* study of the anti-inflammatory and antitumor activities of leaves from *Plectranthus amboinicus* (Lour.) Spreng (Lamiaceae). *Journal of Ethnopharmacology*, 125(2), 361-363.
- Guruprasad, K., Reddy, B. V. P. and Pandit, M. W. (1990). Correlation between stability of a protein and its dipeptide composition: a novel approach for predicting *in vivo* stability of a protein from its primary sequence. *Protein Engineering*, 4(2), 155-161.
- Gururani, M. A., Venkatesh, J., Upadhyaya, C. P., Nookaraju, A., Pandey, S. K. and Park, S. W. (2012). Plant disease resistance genes: Current status and future directions. *Physiology and Molecular Plant Pathology*, 78(1), 51-65.
- Hajduch, M., Ganapathy, A., Stein, J. W. and Thelen, J. J. (2005). A systematic proteomic study of seed filling in soybean. Establishment of high-resolution two-dimensional reference maps, expression profiles, and an interactive proteome database. *Plant Physiology*, 137(4), 1397-1419.
- Hajhashemi, V., Vaseghi, G., Pourfarzam, M. and Abdollahi, A. (2010). Are antioxidants helpful for disease prevention?. *Research in Pharmaceutical Sciences*, 5(1), 1-8.
- Hajheidari, M., Abdollahian-Noghabi, M., Askari, H., Heidari, M., Sadeghian, S.Y., Ober, S. E. and Salekdh, G. H. (2005). Proteome analysis of sugar beet leaves under drought stress. *Proteomics*, 5(4), 950-960.
- Halliwell, B. and Gutteridge, J.M.C. (1995). The definition and measurement of antioxidants in biological systems. *Free Radical Biology and Medicine*, 18(1), 125-126.
- Halliwell, B. (2007). Biochemistry of oxidative stress. *Inflammation*, 1147-1150.
- Hammes, H. P., Du, X., Edelstein, D., Taguchi, T., Matsumura, T., Ju, Q., Lin, J., Bierhaus, A., Nawroth, P. and Hannak, D. (2003). Benfotiamine blocks three major pathways of hyperglycemic damage and prevents experimental diabetic retinopathy. *Nature Medicine*, 9(3), 294-299.
- Hanafusa, N. (1972). Denaturation of enzyme protein by freeze-thawing and freeze-drying. *Contributions from the Institute of Low Temperature Science*, 17(1), 1-20.

- Harnedy, P. A., O’Keeffe, M. B. and FitzGerald, R. J. (2015). Purification and identification of dipeptidyl peptidase (DPP) IV inhibitory peptides from the macroalga *Palmaria palmata*. *Food Chemistry*, 172(1), 400-406.
- Hasima, N. and Aggarwal, B. B. (2014). Targeting proteasomal pathways by dietary curcumin for cancer prevention and treatment. *Current Medicinal Chemistry*, 21(14), 1583-1594.
- Hatanaka, T., Uraji, M., Fujita, A. and Kawakami, K. (2015). Anti-oxidation activities of rice-derived peptides and their inhibitory effects on dipeptidylpeptidase-IV. *International Journal of Peptide Research and Therapeutics*, 21(4), 479-485.
- Henkes, S., Sonnewald, U., Badur, R., Flachmann, R. and Stitt, M. (2001). A small decrease of plastid transketolase activity in antisense tobacco transformants has dramatic effects on photosynthesis and phenylpropanoid metabolism. *The Plant Cell*, 13(3), 535-551.
- Hew, C. S. and Gam, L. H. (2011). Proteome analysis of abundant proteins extracted from the leaf of *Gynura procumbens* (Lour.) Merr. *Applied Biochemistry and Biotechnology*, 165(7-8), 1577-1586.
- Hew, C. S., Khoo, B. Y. and Gam, L. H. (2013). The anti-cancer property of proteins extracted from *Gynura procumbens* (Lour.) Merr. *PloS one*, 8(7), 1-10.
- Higdon, J. V., Delage, B., Williams, D. E. and Dashwood, R. H. (2007). Cruciferous vegetables and human cancer risk: epidemiologic evidence and mechanistic basis. *Pharmacological Research*, 55(3), 224-236.
- Hilou, A., Nacoulma, O. and Guiguemde, T. (2006). *In vivo* antimalarial activities of extracts from *Amaranthus spinosus* L. and *Boerhaavia erecta* L. in mice. *Journal of Ethnopharmacology*, 103(2), 236-240.
- Holman, N., Young, B. and Gadsby, R. (2015). Current prevalence of Type 1 and Type 2 diabetes in adults and children in the UK. *Diabetes Medicine: A Journal of the British Diabetic Association*, 32(9), 1119-1120.
- Holmgren, A. (1978). Glutathione-dependent enzyme reactions of the phage T4 ribonucleotide reductase system. *Journal of Biological Chemistry*, 253(20), 7424-7430.
- Hossain, M. A., Ismail, Z., Rahman, A. and Kang, S. C. (2008). Chemical composition and anti-fungal properties of the essential oils and crude extracts of *Orthosiphon stamineus* Benth. *Industrial Crops and Products*, 27(3), 328-334.

- Hossain, M. M. (2011). Therapeutic orchids: traditional uses and recent advances-an overview. *Fitoterapia*, 82(2), 102-140.
- Hsu, C. C., Ward, C. A., Pearlman, R., Nguyen, H. M., Yeung, D. A. and Curley, J. G. (1992). Determining the optimum residual moisture in lyophilized protein pharmaceuticals. *Developments in Biological Standardization*, 74(1), 255-270.
- Huang, W. F., Wen, K. C. and Hsiao, M. L. (1997). Adulteration by synthetic therapeutic substances of traditional chinese medicines in Taiwan. *The Journal of Clinical Pharmacology*, 37(4), 344-350.
- Humberston, C., Akhtar, J. and Krenzelok, E. (2003). Acute hepatitis induced by Kava Kava: ARTICLE. *Journal of Toxicology: Clinical Toxicology*, 41(2), 109-113.
- Humphrey, W., Dalke, A. and Schulten, K. (1996). VMD - Visual Molecular Dynamics. *Journal of Molecular Graphics*, 14(1), 33-38.
- Hurkman, W. J. and Tanaka, C. K. (1986). Solubilization of plant membrane proteins for analysis by two-dimensional gel electrophoresis. *Plant Physiology*, 81(3), 802-806.
- Hyun, D. H., Hernandez, J. O., Mattson, M. P. and de Cabo, R. (2006). The plasma membrane redox system in aging. *Ageing Research Reviews*, 5(2), 209-220.
- Inada, T., Nozaki, S., Inagaki, A. and Furukawa, T. A. (2003). Efficacy of diazepam as an anti-anxiety agent: meta-analysis of double-blind, randomized controlled trials carried out in Japan. *Human Psychopharmacology*, 18(6), 483-487.
- Iqbal, E., Salim, K. A. and Lim, L. B. (2015). Phytochemical screening, total phenolics and antioxidant activities of bark and leaf extracts of *Goniothalamus velutinus* (Airy Shaw) from Brunei Darussalam. *Journal of King Saud University-Science*, 27(3), 224-232.
- Irar, S., Brini, F., Masmoudi, K. and Pagès, M. (2014). Combination of 2DE and LC for plant proteomics analysis. In *Plant Proteomics* (pp. 131-140). Humana Press, Totowa, NJ.
- Irwin, J., Lorber, D., McGovern, S., Wei, B. and Shoichet, B. (2002). Docking and drug discovery. *Computational Nanoscience and Nanotechnology*, 2(1), 50-51.
- Isaacson, T., Damasceno, C. M., Saravanan, R. S., He, Y., Catalá, C., Saladié, M. and Rose, J. K. (2006). Sample extraction techniques for enhanced proteomic analysis of plant tissues. *Nature Protocols*, 1(2), 769-774.
- Ishtiaq, M. C. (2013). Role of proteomics in plant taxonomy: Drug quality control for pharmaceutical industries. *Research Journal of Biotechnology*, 8(4), 1-1.

- Ishtiaq, M., Maqbool, M., Hussain, T., Azam, S. and Wang, Y. (2014). Leaf proteome analysis of *Clematis chinensis*: A traditional chinese medicine (TCM) by two-dimensional electrophoresis technique. *Pakistan Journal of Botany*, 46(3), 789-801.
- Jacobs, D. I., van der Heijden, R. and Verpoorte, R. (2000). Proteomics in plant biotechnology and secondary metabolism research. *Phytochemical Analysis*, 11(5), 277-287.
- Jagadeesh, D., S., Kannegundla, U. and Reddy, R., K. (2017). Application of proteomics tools in food quality and safety. *Advances in Animal and Veterinary Sciences*, 5(5), 213-225.
- James, P. (1997). Protein identification in the post-genome era: the rapid rise of proteomics. *Quarterly Reviews of Biophysics*, 30(4), 279-331.
- Jelesarov, I. and Karshikoff, A. (2009). Defining the role of salt bridges in protein stability. In *Protein Structure, Stability, and Interactions* (pp. 227-260). New York, US: Humana Press.
- Jeon, C. Y. and Murray, M. B. (2008). Diabetes mellitus increases the risk of active tuberculosis: a systematic review of 13 observational studies. *PLoS Medicine*, 5(7), 1091-1101.
- Jones, G., Willett, P., Glen, R. C., Leach, A. R. and Taylor, R. (1997). Development and validation of a genetic algorithm for flexible docking. *Journal of Molecular Biology*, 267(3), 727-748.
- Jones, J. T., Qian, X., Jos, van der Velden, J. L., Chia, S. B., McMillan, D. H., Flemer, S., Hoffman, S. M., Lahue, K. G., Schneider, R. W., Nolin, J. D., Anathy, V., van der Vliet, A., Townsend, D. M., Tew, K. D. and Janssen-Heininger, M. W. (2016). Glutathione S-transferase pi modulates NF-kappaB activation and pro-inflammatory responses in lung epithelial cells. *Redox Biology*, 8(1), 375-382.
- Jorgensen, W., L., Chandrasekhar, J., Madura, J., D., Impey, R., W. and Klein, M., L. (1983). Comparison of simple potential functions for simulating liquid water. *The Journal of Chemical Physics*, 79(2), 926-935.

- Jorin-Novo, J. V., Pascual, J., Sanchez-Lucas, R., Romero-Rodriguez, M. C., Rodriguez-Ortega, M. J., Lenz, C. and Valledor, L. (2015). Fourteen years of plant proteomics reflected in Proteomics: Moving from model species and 2DE—based approaches to orphan species and gel-free platforms. *Proteomics*, 15(1), 1089-1112.
- Juers, D. H., Matthews, B. W. and Huber, R. (2013). LacZ β -galactosidase: Structure and function of an enzyme of historical and molecular biological importance. *Protein Science*, 21(12), 1792-1807.
- Jung, R. E., Zembic, A., Pjetursson, B. E., Zwahlen, M. and Thoma, D. S. (2012). Systematic review of the survival rate and the incidence of biological, technical, and aesthetic complications of single crowns on implants reported in longitudinal studies with a mean follow-up of 5 years. *Clinical Oral Implants Research*, 23(6), 2-21.
- Kahyaoglu, L. N., Sahin, S. and Sumnu, G.. (2012). ‘Spouted bed and microwave assisted spouted bed drying of parboiled wheat. *Food and Bioprocess Processing*, 90(2), 301-308.
- Kalimeri, M., Rahaman, O., Melchionna, S. and Sterpone, F. (2013). How conformational flexibility stabilizes the hyperthermophilic elongation factor G-domain. *Journal of Physical Chemistry B*, 117(1), 13775-13785.
- Kalhari, N., Nulit, R. and Go, R. (2013). Cloning, expression and characterization of sugarcane (*Saccharum officinarum* L.) transketolase. *The Protein Journal*, 32(7), 551-559.
- Kamo, M., Kawakami, T., Miyake, N. and Tsugita, A. (1995). Separation and characterization of *Arabidopsis thaliana* proteins by two-dimensional gel electrophoresis. *Electrophoresis*, 16(3), 423-300.
- Kanda, T., Yokosuka, O., Tada, M., Kurihara, T., Yoshida, S., Suzuki, Y., Nagao, K. and Saisho, H. (2003). N-nitroso-fenfluramine hepatotoxicity resembling chronic hepatitis. *Journal of Gastroenterology and Hepatology*, 18(8), 999-1000.
- Kang, Y., Chong, E. and Oransky, S. H. (1992). Chinese patent medicine as a potential source of mercury poisoning. *Veterinary and Human Toxicology*, 34(3), 235-238.

- Kang, K. J. (2002). Mechanism of Hepatic Ischemia/Reperfusion Injury and Protection Against Reperfusion Injury. In *Transplantation Proceedings*, 34(7), 2659-2661.
- Kazafeos, K. (2011). Incretin effect: GLP-1, GIP, DPP4. *Diabetes Research and Clinical Practice*, 93(Suppl 1), 32-36.
- Keng, C. L. and Siong, L. P. (2006). Morphological similarities and differences between the two varieties of Cat's Whiskers (*Orthosiphon stamineus* Benth.) grown in Malaysia. *International Journal of Botany*, 2(1), 1-6.
- Keum, Y. S., Jeong, W. S. and Kong, A. N. (2005). Chemopreventive functions of isothiocyanates. *Drugs News and Perspectives*, 18(7), 445-451.
- Khan, S. A., Khan, L., Hussain, I., Marwat, K. B. and Akhtar, N. (2008). Profile of heavy metals in selected medicinal plants. *Pakistan Journal of Weed Science Research*, 14(1-2), 101-110.
- Khan, S., Farooq, U. and Kurnikova, M. (2016a). Exploring protein stability by comparative molecular dynamics simulation of homologous hyperthermophilic, mesophilic and psychrophilic proteins. *Journal of Chemical Information and Modeling*, 56(11), 2129-2139.
- Khan, M. I. H., Wellard, R. M., Nagy, S. A., Joardder, M. U. H. and Karim, M. A. (2016b). Investigation of bound and free water in plant-based food material using NMR T2 relaxometry. *Innovative Food Science & Emerging Technologies*, 38(1), 252-261.
- Khider, K., Akretche, D. E. and Larbot, A. (2004). Purification of water effluent from a milk factory by ultrafiltration using Algerian clay support. *Desalination*, 167(1), 147-151.
- Khor, B. Y., Tye, G. J., Lim, T. S. and Choong, Y. S. (2015). General overview on structure prediction of twilight-zone proteins. *Theoretical Biology and Medical Modelling*, 12(15), 1-11.
- Kidane, Y., Bokrezion, T., Mebrahtu, J., Mehari, M., Gebreab, Y. B., Fessehaye, N. and Achila, O. O. (2018). *In Vitro* Inhibition of α -Amylase and α -Glucosidase by Extracts from *Psiadia punctulata* and *Meriandra bengalensis*. *Evidence-based Complementary and Alternative Medicine*, 2018(1), 1-9.
- Klein, J. B. and Thongboonkerd, V. (2004). Overview of proteomics. *Contributions to Nephrology*, 141(1), 1-10.

- Kofler, S., Asam, C., Eckhard, U., Wallner, M., Ferreira, F. and Brandstetter, H. (2012). Crystallographically mapped ligand binding differs in high and low IgE binding isoforms of birch pollen allergen bet v 1. *Journal of Molecular Biology*, 422(1), 109-123.
- Kohlmeier, M. (2013). How nutrients are affected by genetics. In *Nutrigenetics* (pp. 103-221). Amsterdam, US: Elsevier,.
- Koller, A., Washburn, M., P., Lange, B., M., Andon, N., L., Deciu, C. and Haynes, P., A. (2002). Proteomics survey of metabolic pathways in rice. *Proceedings of the National Academy of Sciences of the United States of America*, 99(18), 11989-11974.
- Kong, B. and Xiong, Y. L. (2006). Antioxidant activity of zein hydrolysates in a liposome system and the possible mode of action. *Journal of Agricultural and Food Chemistry*, 54(16), 6059-6068.
- Koyama, M., Naramoto, K., Nakajima, T., Aoyama, T., Watanabe, M. and Nakamura, K. (2013). Purification and identification of antihypertensive peptides from fermented buckwheat sprouts. *Journal of Agricultural and Food Chemistry*, 61(12), 3013–3021.
- Koyama, M., Hattori, S., Amano, Y., Watanabe, M. and Nakamura, K. (2014). Blood pressure-lowering peptides from neo-fermented buckwheat sprouts: A new approach to estimating ACE-inhibitory activity. *PLoS one*, 9(9), 1-8.
- Kreydiyyeh, S. I. and Usta, J. (2002). Diuretic effect and mechanism of action of parsley. *Journal of Ethnopharmacology*, 79(3), 353-357.
- Krishnaiah, D., Sarbatly, R. and Nithyanandam, R. (2011). A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing*, 89(3), 217-233.
- Krul, W. R. and Colclasure, G. C. (1977). Effect of galactose and other monosaccharides on IAA movement in bean hypocotyl segments. *Physiologia Plantarum*, 41(4), 249-253.
- Kumar, S., Tsai, C. J. and Nussinov, R. (2000). Factors enhancing protein thermostability. *Protein Engineering*, 13(3), 179-191.
- Kumar, P. S., Sucheta, S., Deepa, V.S., Selvamani, P. and Latha, S. (2008). Antioxidant activity in some selected Indian medicinal plants. *African Journal of Biotechnology*, 7(12), 1826-1828.

- Kuroiwa, Y., Nishikawa, A., Kitamura, Y., Kanki, K., Ishii, Y., Umemura, T. and Hirose, M. (2006). Protective effects of benzyl isothiocyanate and sulforaphane but not resveratrol against initiation of pancreatic carcinogenesis in hamsters. *Cancer Letters*, 241(2), 275-280.
- Kvasnicka, F. (2003). Proteomics: general strategies and application to nutritionally relevant proteins. *Journal of Chromatography B*, 787(1), 77-89.
- Kyte, J. and Doolittle, R. F. (1982). A simple method for displaying the hydropathic character of a protein. *Journal of Molecular Biology*, 157(1), 105-132.
- Laavola, M., Nieminen, R., Yam, M. F., Sadikun, A., Asmawi, M. Z., Basir, R., Weiling, J., Vapaatalo, H., Korhonen, R. and Moilanen, E. (2012). Flavonoids eupatorin and sinensetin present in *Orthosiphon stamineus* leaves inhibit inflammatory gene expression and STAT1 activation. *Planta Medica*, 78(08), 779-786.
- Lacroix, I., M., Chen, X., M., Kitts, D. and Li-Chan, E., C. (2017). Investigation into the bioavailability of milk protein-derived peptides with dipeptidyl-peptidase IV inhibitory activity using Caco-2 cell monolayers. *Food & Function*, 8(2), 701-709.
- Lafarga, T., O'Connor, P. and Hayes, M. (2014). Identification of novel dipeptidyl peptidase-IV and angiotensin-I-converting enzyme inhibitory peptides from meat proteins using *in silico* analysis. *Peptides*, 59(1), 53–62.
- Lagasse, H. D., Alexaki, A., Simhadri, V. L., Katagiri, N. H., Jankowski, W., Sauna, Z. E. and Kimchi-Sarfatyb, C. (2017). Recent advances in (therapeutic protein) drug development. *F1000Research*, 6(1), 1-17.
- Lai, M. C. and Topp, E. M. (1999). Solid state chemical stability of proteins and peptides. *Journal of Pharmaceutical Sciences*, 88(5), 489-500.
- Laing, W. and Christeller, J. (2004). Extraction of proteins from plant tissues. *Current Protocols in Protein Science*, 38(1), 4-7.
- Lao, Y., Wang, X., Xu, N., Zhang, H., and Xu, H. (2014). Application of proteomics to determine the mechanism of action of traditional chinese medicine remedies. *Journal of Ethnopharmacology*, 155(1), 1-8.
- Laskowski, R. A., MacArthur, M. W., Moss, D. S. and Thirnton, J. M. (1993). PROCHECK – a program to check the stereochemical quality of protein structures. *Journal of Applied Crystallography*, 26(2), 283-291.

- Laskowski, R. A., Rullmann, J. A., MacArthur, M. W., Kaptein, R. and Thornton, J. M. (1996). AQUA and PROCHECK-NMR: programs for checking the quality of protein structures solved by NMR. *Journal of Biomolecular NMR*, 8(4), 477-486.
- Lazaridis, T. and Karplus, M. (1999). Heat capacity and compactness of denatured proteins. *Biophysical Chemistry*, 78(1-2), 207-217.
- Lee, K. J. (2011). Molecular dynamics simulations of a hyperthermophilic and a mesophilic protein L30e. *Journal of Chemical Information and Modeling*, 52(1), 7-15.
- Lekli, I., Mukherjee, S., Ray, D., Gurusamy, N., Kim, Y. H., Tosaki, A., Engelman, R. M., Ho, Y. S., and Das, D. K. (2010). Functional recovery of diabetic mouse hearts by glutaredoxin-1 gene therapy: role of Akt-FoxO-signaling network. *Gene therapy*, 17(4), 478-485.
- Lewis, J. K., Wei, J., & Siuzdak, G. (2006). *Matrix-assisted laser desorption/ionization mass spectrometry in peptide and protein analysis*, in Meyers R. A. (eds.) *Encyclopedia of Analytical Chemistry, Theory and Instrumentation*, pp. 5880-5894.
- Li, J., Matsumura, Y., Shinjo, M., Kojima, M. and Kihara, H. (2007). A stable α -helix-rich intermediate is formed by a single mutation of the β -sheet protein, src SH3, at pH 3. *Journal of Molecular Biology*, 372(3), 747-755.
- Li, S., Yuan, W., Yang, P., Antoun, M., Balick, M. and Cragg, G. (2010). Pharmaceutical crops: an overview. *Pharmaceutical Crops*, 1(1), 1-17.
- Li, J., W. and Bordelon, P. (2011). Hydroxycitric acid dietary supplement-related herbal nephropathy. *The American Journal of Medicine*, 124(11), 5-6.
- Li, E. C., Hunag, S. L., Jao, C. L., Ho, K. P. and Hsu, K. C. (2012a). Peptides derived from atlantic salmon skin gelatin as dipeptidyl-peptidase IV inhibitors. *Journal of Agricultural and Food Chemistry*, 60(4), 973-978.
- Li, Y., Zhou, J., Huang, K., Sun, Y. and Zeng, X. (2012b). Purification of a novel angiotensin I-converting enzyme (ACE) inhibitory peptide with an antihypertensive effect from loach (*Misgurnus anguillicaudatus*). *Journal of Agricultural and Food Chemistry*, 60(5), 1320-1325.

- Li, J., Cai, J., Su, H., Du, H., Zhang, J., Ding, S., Liu, G., Tang, Y. and Li, W. (2016). Effects of protein flexibility and active site water molecules on the prediction of sites of metabolism for cytochrome P450 2C19 substrates. *Molecular BioSystems*, 12(3), 868-878.
- Liao, M., Li, Y. and Wang, Z. (2009). Identification of elicitor-responsive proteins in rice leaves by a proteomic approach. *Proteomics*, 9(10), 2809-2819.
- Lima, R. N., Faheem, M., Barbosa, J. A. R. G., Polêto, M. D., Verli, H., Melo, F. L. and Resende, R. O. (2016). Homology modeling and molecular dynamics provide structural insights into tospovirus nucleoprotein. *BMC Bioinformatics*, 17(Suppl 18), 489-495.
- Lin, S. H., Yang, S. S., Chau, T. and Halperin, M. L. (2003). An unusual cause of hypokalemic paralysis: chronic licorice ingestion. *American Journal of the Medical Sciences*, 325(3), 153-156.
- Lin, G., Wang, J., Y., Li, N., Li, M., Gao, H., Ji, Y., Zhang, F., Wang, H., Zhou, Y., Ye, Y., Xu, H. X. and Zheng, J. (2011). Hepatic sinusoidal obstruction syndrome associated with consumption of *Gynura segetum*. *Journal of Hepatology*, 54(4), 666-673.
- Liu, Z., Xin, M., Qin, J., Peng, H., Ni, Z., Yao, Y. and Sun, Q. (2015). Temporal transcriptome profiling reveals expression partitioning of homeologous genes contributing to heat and drought acclimation in wheat (*Triticum aestivum* L.). *BMC Plant Biology*, 15(1), 1-20.
- Liu, C., Fan, H., Li, Y. and Xiao, X. (2016). Research advances on hepatotoxicity of herbal medicines in China. *BioMed Research International*, 2016(1), 1-14.
- Lobanov, M., Bogatyreva, N. S. and Galzitskaia, O. V. (2008). Radius of gyration is indicator of compactness of protein structure. *Molekuliarnaia Biologiia*, 42(4), 701-706.
- Loguerco, C. and Federico, A. (2003). Oxidative stress in viral and alcoholic hepatitis. *Free Radical Biology and Medicine*, 34(1), 1-10.
- Loizzo, M. R., Said, A., Tundis, R., Rashed, K., Statti, G. A., Hufner, A. and Menichini, F. (2007). Inhibition of angiotensin converting enzyme (ACE) by flavonoids isolated from *Ailanthus excelsa* (Roxb)(Simaroubaceae). *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 21(1), 32-36.

- Lordan, S., Smyth, T. J., Soler-Vila, A., Stanton, C. and Ross R. P. (2013). The α -amylase and α -glucosidase inhibitory effects of Irish seaweed extracts. *Food Chemistry*, 141(3), 2170-2176.
- Lu, F. P., Lin, K. P. and Kuo, H. K. (2009). Diabetes and the risk of multi-system aging phenotypes: a systematic review and meta-analysis. *PLoS one*, 4(1), 1-12.
- Lu, Y., Qi, Y., Zhang, H., Zhang, H., Pu, J. and Xie, Y. (2013). Separation and identification of *Musa acuminata* Colla (banana) leaf proteins by two-dimensional gel electrophoresis and mass spectrometry. *Genetics and Molecular Research: GMR*, 12(4), 6871-6881.
- Lu, Y., Li, R., Wang, R., Wang, X., Zheng, W., Sun, Q., Tong, S., Dai, S. and Xu, S. (2017). Comparative proteomic analysis of flag leaves reveals new insight into wheat heat adaptation. *Frontiers in Plant Science*, 8(1), 1-11.
- Luís, I. M., Alexandre, B. M., Oliveira, M. M. and Abreu, I. A. (2016). Selection of an appropriate protein extraction method to study the phosphoproteome of maize photosynthetic tissue. *PLoS one*, 11(10), 1-16.
- Luo, L., Wang, Y., Feng, Q., Zhang, H., Xue, B., Shen, J., Ye, Y., Han, X., Ma, H., Xu, J., Chen, D. and Yin, Z. (2009). Recombinant protein glutathione S-transferases P1 attenuates inflammation in mice. *Molecular Immunology*, 46(5), 848-857.
- Luthy, R., Bowie, J. U. and Eisenberg, D. (1992). Assessment of protein models with three-dimensional profiles. *Nature*, 356(6364), 83-85.
- Lutz, S. (2010). Beyond directed evolution-semi-rational protein engineering and design. *Current Opinion in Biotechnology*, 21(6), 734-743.
- Lyckander, I. M. and Malterud, K. (1996). Lipophilic flavonoids from *Orthosiphon spicatus* prevent oxidative inactivation of 15-lipoxygenase. *Prostaglandins, Leukotrienes and Essential Fatty Acids*, 54(4), 239-246.
- Ma, Z. A., Zhao, Z. and Turk, J. (2011). Mitochondrial dysfunction and β -cell failure in type 2 diabetes mellitus. *Experimental diabetes research*, 2012(1), 1-11.
- Mahalingam, R. (2017). Shotgun proteomics of the barley seed proteome. *BMC genomics*, 18(1), 1-11.
- Maheswari, C., Sajna, V. and Venkatnarayanan, R. (2016). *In silico* docking analysis of the compounds of *Orthosiphon stamineus* for the anticancer activity. *International Research Journal of Pharmacy*, 7(4), 17-23.

- Maiga, A., Malterud, K. E., Diallo, D. and Paulsen, B. S. (2006). Antioxidant and 15-lipoxygenase inhibitory activities of the Malian medicinal plants *Diospyros abyssinica* (Hiern) F. White (Ebenaceae), *Lannea velutina* A. Rich (Anacardiaceae) and *Crossopteryx febrifuga* (Afzel) Benth. (Rubiaceae). *Journal of Ethnopharmacology*, 104(1), 132-137.
- Maisnam, D., Rasane, P., Dey, A., Kaur, S. and Sarma, C. (2017). Recent advances in conventional drying of foods. *Journal of Food Technology and Preservation*, 1(1), 25-34.
- Malcevski, A. and Marmiroli, N. (2012). Plant protein analysis. In *Proteomic applications in biology*. London, United Kingdom: InTech Open.
- Mamone, G., Picariello, G., Caira, S., Addeo, F. and Ferranti, P. (2009). Analysis of food proteins and peptides by mass spectrometry-based techniques. *Journal of Chromatography A*, 1216(43), 7130–7142.
- Manan, F. A., Chai, T. T., SAMAD, A. A. and Mamat, D. D. (2015). Evaluation of the phytoremediation potential of two medicinal plants. *Sains Malaysiana*, 44(4), 503-509.
- Manavalan, A., Ramachandran, U., Sundaramurthi, H., Mishra, M., Sze, S. K., Hu, J. M., Feng, Z. W. and Heese, K. (2012). *Gastrodia elata* Blume (tianma) mobilizes neuro-protective capacities. *International Journal of Biochemistry and Molecular Biology*, 3(2), 219-241.
- Mangelsen, E., Kilian, J., Harter, K., Jansson, C., Wanke, D. and Sundberg, E. (2011). Transcriptome analysis of high-temperature stress in developing barley caryopses: early stress responses and effects on storage compound biosynthesis. *Molecular Plant*, 4(1), 97-115.
- Mann, M. and Jensen, O. N. (2003). Proteomic analysis of post-translational modifications. *Nature Biotechnology*, 21(3), 255-261.
- Marcus, D. M. and Grollman, A. P. (2002). Botanical medicines-the need for new regulations. *The New England Journal of Medicine*, 347(25), 2073-2076.
- Marouga, R., David, S. and Hawkins, E. (2005). The development of the DIGE system: 2D fluorescence difference gel analysis technology. *Analytical and bioanalytical chemistry*, 382(3), 669-678.
- Martin, P. R., Singleton, C. K., Hiller-Sturmhofel, S. (2003). The role of thiamine deficiency in alcoholic brain disease. *Alcohol Research and Health*, 27(2), 134-142.

- Martínez-Esteso, M. J., Casado-Vela, J., Sellés-Marchart, S., Elortza, F., Pedreño, M. A. and Bru-Martínez, R. (2011). iTRAQ-based profiling of grape berry exocarp proteins during ripening using a parallel mass spectrometric method. *Molecular BioSystems*, 7(3), 749-765.
- Martínez-Esteso, M. J., Vilella-Antón, M. T., Pedreño, M. Á., Valero, M. L. and Bru-Martínez, R. (2013). iTRAQ-based protein profiling provides insights into the central metabolism changes driving grape berry development and ripening. *BMC Plant Biology*, 13(167), 1-20.
- Martí-Renom, M. A., Stuart, A. C., Fiser, A., Sánchez, R., Melo, F. and Šali, A. (2000). Comparative protein structure modeling of genes and genomes. *Annual Review of Biophysics and Biomolecular Structure*, 29(1), 291-325.
- Maskan, M. (2001). Kinetics of colour change of kiwifruits during hot air and microwave drying. *Journal of Food Engineering*, 48(2), 169-175.
- Masuda, T., Masuda, K., Shiragami, S., Jitoe, A. and Nakatani, N. (1992). Orthosiphon A and B, novel diterpenoid inhibitors of TPA (12-O-tetradecanoylphorbol-13-acetate)-induced inflammation, from *Orthosiphon stamineus*. *Tetrahedron*, 48(33), 6787-6792.
- Matharu, G. K., Thappa, D., Kamble, V., Krishnan, A. and Sane, R. T. (2010). Use of genetic and protein markers for characterization of medicinal ayurvedic plants. *International Journal of Pharmaceutical Sciences and Research*, 1(12), 95-110.
- Matsubara, T., Bohgaki, T., Watarai, M., Suzuki, H., Ohashi, K. and Shibuya, H. (1999). Antihypertensive actions of methylripariochromene A from *Orthosiphon aristatus*, an Indonesian traditional medicinal plant. *Biological and Pharmaceutical Bulletin*, 22(10), 1083-1088.
- Matsuura, S., Kunii, T. and Inuma, M. (1973). Studies on the flavonoid of Kumis-kuching (*Orthosiphon stamineus* Benth.)(author's transl). *Yakugaku zasshi: Journal of the Pharmaceutical Society of Japan*, 93(11), 1517-1519.
- McClellan, S., Beggs, L., B. and Welch, R., W. (2014). Antimicrobial activity of antihypertensive food-derived peptides and selected alanine analogues. *Food Chemistry*, 146(1), 443-447.
- McHugh, L. and Arthur, J. W. (2008). Computational methods for protein identification from mass spectrometry data. *PLOS Computational Biology*, 4(2), 1-12.

- Meier, J. J. and Nauck, M. A. (2014). Risk of pancreatitis in patients treated with incretin-based therapies. *Diabetologia*, 57(7), 1320–1324.
- Mendes, R. H., Hagen, M. E. K., Barp, J., Jong, E. V. D., Moreira, J. D., Oliveira, Á. R. D., Irigoyen, M. C. C. and Belló-Klein, A. (2014). Isolated soy protein-based diet ameliorates glycemia and antioxidants enzyme activities in streptozotocin-induced diabetes. *Food and Nutrition Sciences*, 5(21), 2089-2096.
- Meyfour, A., Tavirani, M. R. and Sadeghi, M. R. (2013). Common proteomic technologies, applications, and their limitations. *Journal of Paramedical Sciences*, 4(1), 115-125.
- Michalak, S., Michalowska-Wender, G., Adamcewicz, G. and Wender, M. B. (2013). Erythrocyte transketolase activity in patients with diabetic and alcoholic neuropathies. *Folia Neuropathologica*, 51(3), 222-226.
- Mika, A. and Lüthje, S. (2003). Properties of guaiacol peroxidase activities isolated from corn root plasma membranes. *Plant Physiology*, 132(3), 1489-1498.
- Miroshnichenko, S., Tripp, J., Nieden, U., Neumann, D., Conrad, U. and Manteuffel, R. (2005). Immunomodulation of function of small heat shock proteins prevents their assembly into heat stress granules and results in cell death at sublethal temperatures. *The Plant Journal*, 41(2), 269-281.
- Mirzaei, M., Wu, Y., Handler, D., Maher, T., Pascovici, D., Ravishankar, P., Moghaddam, M. Z., Haynes, P. A., Salekdeh, G. H., Chick, J. M. and Willows, R. D. (2016). Applications of quantitative proteomics in plant research. In Salekdeh, G. (eds). *Agricultural Proteomics Volume 1* (pp.1-29). Springer, Cham.
- Mitschke, L., Parthier, C., Schröder-Tittmann, K., Coy, J., Lüdtke, S. and Tittmann, K. (2010). The crystal structure of human transketolase and new insights into its mode of action. *Journal of Biological Chemistry*, 285(41), 31559-31570.
- Mittler, R. (2002). Oxidative stress, antioxidants and stress tolerance. *Trends in Plant Science*. 7(9), 405-410.
- Mittler, R. (2004). Reactive oxygen gene network of plants. *Trends in Plant Science*. 9(10), 490-498.
- Mlichova, Z. and Rosenberg, M. (2006). Current trends of β -galactosidase application in food. *Journal of Food and Nutrition Research*, 45(2), 47-54.

- Moawad, F. J., Hartzell, J. D., Biega, T. J. and Lettieri, C. J. (2006). Transient blindness due to posterior reversible encephalopathy syndrome following ephedra overdose. *Southern Medical Journal*, 99(5), 511-515.
- Mohamed, E. A. H., Mohamed, A. J., Asmawi, M., Sadikun, A., Ebrika, O. S., & Yam, M. F. (2011). Antihyperglycemic effect of *Orthosiphon stamineus* benth leaves extract and its bioassay-guided fractions. *Molecules*, 16(5), 3787-3801.
- Mohamed, E. A. H., Siddiqui, M. J. A., Ang, L. F., Sadikun, A., Chan, S. H., Tan, S. C, Asmawi, M. Z. and Yam, M. F. (2012). Potent α -glucosidase and α -amylase inhibitory activities of standardized 50 % ethanolic extracts and sinensetin from *Orthosiphon stamineus* Benth as anti-diabetic mechanism. *BMC Complementary and Alternative Medicine*. 12(1), 176-182.
- Mohamed, E. A., Ahmad, M., Lee, F. A., Asmawi, M. Z. and Yam, M. F. (2015). Evaluation of α -glucosidase inhibitory effect of 50% ethanolic standardized extract of *Orthosiphon stamineus* Benth in normal and streptozotocin-induced diabetic rats. *Evidence-Based Complementary and Alternative Medicine*, 2015(1), 1-6.
- Morimoto, R. I. (1998). Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaperones, and negative regulators. *Genes & Development*. 12(24), 3788–3796.
- Morre, D. J. (1968). Cell wall dissolution and enzyme secretion during leaf abscission. *Plant Physiology*, 43(9), 1545-1559.
- Morris, G. M., Goodsell, D. S., Halliday, R. S., Huey, R., Hart, W. E., Belew, R. K. and Olson, A. J. (1998). Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function. *Journal of Computational Chemistry*, 19(14), 1639-1662.
- Morse, M. A., Wang, C. X., Stoner, G. D., Mandal, S., Conran, P. B., Amin, S. G., Hecht, S. S. and Chung, F. L. (1989). Inhibition of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone-induced DNA adduct formation and tumorigenicity in the lung of F344 rats by dietary phenethyl isothiocyanate. *Cancer Research*, 49(3), 549-553.
- Mosihuzzaman, M. and Choudhary, M. I. (2008). Protocols on safety, efficacy, standardization, and documentation of herbal medicine (IUPAC Technical Report). *Pure and Applied Chemistry*, 80(10), 2195-2230.

- Moxey, P. W., Gogalniceanu, P., Hinchliffe, R. J., Loftus, I. M., Jones, K. J., Thompson, M. M. and Holt, P. J. (2011). Lower extremity amputations - a review of global variability in incidence. *Diabetic Medicine*, 28(10), 1144-1153.
- Mubassir, M., H., M., Naser, M., A., Abdul Wahab, M. F. and Hamdan, S. (2017). *In silico* structural modeling and molecular dynamics simulation of pathogen-associated molecular pattern RAXX21. *Journal of Chemical and Pharmaceutical Sciences*, 10(1), 121-126.
- Mukesh, B. and Rakesh, K. (2011). Review on Molecular docking. *International Journal of Research in Ayurveda & Pharmacy*, 2(6), 1746-1751.
- Mullins, R. J. and Heddle, R. (2002). Adverse reactions associated with echinacea: the Australian experience. *Annals of Allergy, Asthma & Immunology*, 88(1), 42-51.
- Murphy, S., Dowling, P. and Ohlendieck, K. (2016). Comparative skeletal muscle proteomics using two-dimensional gel electrophoresis. *Proteomes*, 4(3), 27.
- Nail, A. M. M. and Zin, N. H. M. (2015). Protein profiling of different plant tissues from herb *Phyllanthus niruri*. *Jurnal Teknologi*, 77(24), 95-99.
- Nair, S. S., Kavrekar, V. and Mishra, A. (2013). *In vitro* studies on alpha amylase and alpha glucosidase inhibitory activities of selected plant extracts. *European Journal of Experimental Biology*, 3(1), 128-132.
- Nandi, P. K., English, N. J., Zdenek, F. and Antonio, B. (2017). Hydrogen-bond dynamics at the bio-water interface in hydrated proteins: a molecular-dynamics study. *Physical Chemistry Chemical Physics*, 19(1), 318-329.
- Narayanan, S., Prasad, P., Fritz, A., Boyle, D. and Gill B. (2015). Impact of high night-time and high daytime temperature stress on winter wheat. *Journal of Agronomy and Crop Science*, 201(3), 206-218.
- Nasir, N. N., Khandaker, N. M., Moneruzzaman, M. and Mat, N. (2015). Bioactive compound and therapeutic value of the some malaysia medicinal plants: A review. *Journal of Agronomy*, 14(4), 319-330.
- Neilson, K. A., Mariani, M. and Haynes, P. A. (2011). Quantitative proteomic analysis of cold-responsive proteins in rice. *Proteomics*, 11(9), 1696-1706.
- Neilson, K. A., George, I. S., Emery, S. J., Muralidharan, S., Mirzaei, M. and Haynes, P. A. (2014). Analysis of rice proteins using SDS-PAGE shotgun proteomics. In *Plant Proteomics* (pp. 289-302). Humana Press, Totowa, NJ.

- Neta-Sharir, I., Isaacson, T., Lurie, S. and Weiss, D. (2005). Dual role for tomato heat shock protein 21: protecting photosystem II from oxidative stress and promoting color changes during fruit maturation. *The Plant Cell*, 17(6), 1829-1838.
- Neupert, W. and Brunner, M. (2002). The protein import motor of mitochondria. *Nature reviews Molecular cell biology*, 3(8), 555-565.
- Ng, M. L. (2016). *Gel-based Proteome Analysis of Orthosiphon stamineus*. Undergraduate Thesis. Universiti Teknologi Malaysia, Skudai.
- Ngoh, Y. Y. and Gan, C. Y. (2016). Enzyme-assisted extraction and identification of antioxidative and α -amylase inhibitory peptides from Pinto beans (*Phaseolus vulgaris* cv. Pinto). *Food Chemistry*, 190(1), 331-337.
- Nichols, S. E., Baron, R., Ivetac, A. and McCammon, J. A. (2011). Predictive power of molecular dynamics receptor structures in virtual screening. *Journal of Chemical Information and Modeling*, 51(6), 1439-1446.
- Nivetha, A. and Mohanasrinivasan, V. (2017). Mini review on role of β -galactosidase in lactose intolerance. *Materials Science and Engineering*, 263(2), 1-6.
- Nongonierma, A. B. and FitzGerald, R. J. (2014). Susceptibility of milk protein-derived peptides to dipeptidyl peptidase IV (DPP-IV) hydrolysis. *Food Chemistry*, 15(145), 845-852.
- Obiol-Pardo, C. and Rubio-Martinez, J. (2009). Homology modeling of human Transketolase: Description of critical sites useful for drug design and study of the cofactor binding mode. *Journal of Molecular Graphics and Modelling*, 27(6), 723-734.
- Odhav, B., Beekrum, S., Akula, U. and Baijnath, H. (2007). Preliminary assessment of nutritional value of traditional leafy vegetables in KwaZulu-Natal, South Africa. *Journal of Food Composition and Analysis*, 20(5), 430-435.
- Oh, S. H., Witek, R. P., Bae, S. H., Darwiche, H., Jung, Y., Pi, L., Brown, A. and Petersen, B. E. (2009). Detection of transketolase in bone marrow-derived insulin-producing cells: benfotiamine enhances insulin synthesis and glucose metabolism. *Stem Cells and Development*, 18(1), 37-46.

- Ohashi, K., Bohgaki, T., Matsubara, T. and Shibuya, H. (2000a). Chemical structures of two new migrated pimarane-type diterpenes, neoorthosiphols A and B, and suppressive effects on rat thoracic aorta of chemical constituents isolated from the leaves of *Orthosiphon aristatus* (Lamiaceae). *Chemical and Pharmaceutical Bulletin*, 48(3), 433-435.
- Ohashi, K., Bohgaki, T. and Shibuya, H. (2000b). Antihypertensive substance in the leaves of kumis kucing (*Orthosiphon aristatus*) in Java Island. *Yakugaku zasshi: Journal of the Pharmaceutical Society of Japan*, 120(5), 474-482.
- Olaokun, O. O., McGaw, L. J., Rensburg, I. J., Eloff, J. N. and Naidoo, V. (2016). Antidiabetic activity of the ethyl acetate fraction of *Ficus lutea* (Moraceae) leaf extract: comparison of an *in vitro* assay with an *in vivo* obese mouse model. *BMC Complementary and Alternative Medicine*, 16(1), 1-12.
- Olubomehin, O. O., Abo, K. A. and Ajaiyeoba, E. O. (2013). α -amylase inhibitory activity of two *Anthocleista* species and *in vivo* rat model anti-diabetic activities of *Anthocleista djalensis* extracts and fractions. *Journal of Ethnopharmacology*, 146(3), 811-814.
- Ott M, Gogvadze V, Orrenius, S. and Zhivotovsky, B. (2007). Mitochondria, oxidative stress and cell death. *Apoptosis*, 12(5), 913–922.
- Pääkkönen, K., Havento, J., Galambosi, B. and Pyykkönen, M. (1999). Infrared drying of herbs. *Agricultural and Food Science in Finland*, 8(1), 19-27.
- Pácal, L., Tomandl, J., Svojanovský, J., Krusová, D., Štěpánková, S., Řehořová, J., Olšovský, J., Bělobrádková, J., Tanhäuserová, V., Tomandlová, M. and Mužík, J. (2010). Role of thiamine status and genetic variability in transketolase and other pentose phosphate cycle enzymes in the progression of diabetic nephropathy. *Nephrology Dialysis Transplantation*, 26(4), 1229-1236.
- Pace, C.N., Fu, H., Lee Fryar, K., Landua, J., Trevino, S.R., Schell, D., Thurlkill, R.L., Imura, S., Scholtz, J.M., Gajiwala, K. and Sevcik, J. (2014). Contribution of hydrogen bonds to protein stability. *Protein Science*, 23(5), 652-661.
- Pandey, R., Tiwari, R., K. and Shukla, S. S. (2016). Omics: A newer technique in herbal drug standardization and quantification. *Journal of Young Pharmacists*, 8(2), 76-81.
- Pang, G., Xie, J., Chen, Q. and Hu, S. (2012). How functional foods play critical roles in human health. *Food Science and Human Wellness*, 1(1), 26-60.

- Panyawong, S. and Devahastin, S. (2007). Determination of deformation of a food product undergoing different drying methods and conditions via evolution of a shape factor. *Journal of Food Engineering*, 78(1), 151-161.
- Patodia, S., Bagaria, A. and Chopra, D. (2014). Molecular dynamics simulation of proteins: a brief overview. *Journal of Physical Chemistry & Biophysics*, 4(6), 1-4.
- Patterson, S. D. and Aebersold, R. H. (2003). Proteomics: the first decade and beyond. *Nature genetics*, 33(3s), 311-323.
- Patterson, S. D. (2004). How much of the proteome do we see with discovery-based proteomics methods and how much do we need to see? *Current Proteomics*, 1(1), 3-12.
- Percival, M. (1998). Antioxidants. *Clinical Nutrition Insights*, 1(1), 1-4.
- Perez-Gutierrez, R. M. and Damian-Guzman, M. (2012). Meliacinolin: a potent α -glucosidase and α -amylase inhibitor isolated from *Azadirachta indica* leaves and *in vivo* antidiabetic property in streptozotocin-nicotinamide-induced type 2 diabetes in mice. *Biological and Pharmaceutical Bulletin*, 35(9), 1516-1524.
- Periche, A., Castelló, M. L., Heredia, A. and Escriche, I. (2015). Effect of different drying methods on the phenolic, flavonoid and volatile compounds of *Stevia rebaudiana* leaves. *Flavour and Fragrance Journal*, 31(2), 173-177.
- Petronczki, M., Griffith, J. D. and West, S. C. (2010). The breast cancer tumor suppressor BRCA2 promotes the specific targeting of RAD51 to single-stranded DNA. *Nature Structural and Molecular Biology*, 17(10), 1263-1265.
- Pettersen, E. F., Goddard, T. D., Huang, C. C., Couch, G. S., Greenblatt, D. M., Meng, E. C. and Ferrin, T. E. (2004). UCSF Chimera-A visualization system for exploratory research and analysis. *Journal of Computational Chemistry*, 25(13), 1605-1612.
- Pietta, P., Mauri, P., Gardana, C. and Bruno, A. (1991). High-performance liquid chromatography with diode-array ultraviolet detection of methoxylated flavones in *Orthosiphon* leaves. *Journal of Chromatography A*, 547(1), 439-442.
- Piñeiro, C., Barros-Velázquez, J., Vázquez, J., Figueras, A. and Gallardo, J., M. (2003). Proteomics as a tool for the investigation of seafood and other marine products. *Journal of Proteome Research*, 2(2), 127-135.

- Pirovani, C. P., Carvalho, H. A. S., Machado, R. C. R., Gomes, D. S., Alvim, F. C., Pomella, A. W. V., Gramacho, K.P., Cascardo, J.C.D.M., Pereira, G.A.G. and Micheli, F. (2008). Protein extraction for proteome analysis from cacao leaves and meristems, organs infected by *Moniliophthora perniciosa*, the causal agent of the witches' broom disease. *Electrophoresis*, 29(11), 2391-2401.
- Poos, T. and Varju, E. (2017). Drying characteristics of medicinal plants. *International Review of Applied Sciences and Engineering*, 8(1), 83-91.
- Poovitha, S. and Parani, M. (2016). *In vitro* and *in vivo* α -amylase and α -glucosidase inhibiting activities of the protein extracts from two varieties of bitter melon (*Momordica charantia* L.). *BMC Complementary and Alternative Medicine*, 16(1), 1-8.
- Poovitha, S., Sai, M. S. and Parani, M. (2017). Protein extract from the fruit pulp of *Momordica dioica* shows anti-diabetic, anti-lipidemic and antioxidant activity in diabetic rats. *Journal of Functional Foods*, 33(2017), 181-187.
- Psotová, J., Kolář, M., Soušek, J., Švagera, Z., Vičar, J. and Ulrichová, J. (2003). Biological activities of *Prunella vulgaris* extract. *Phytotherapy Research*, 17(9), 1082-1087.
- Puchta, H., Trapp, O. and Seeliger, K. (2011). Homologs of breast cancer genes in plants. *Frontiers in Plant Science*, 2(1), 1-17.
- Pulido, R., Bravo, L. and Saura-Calixto, F. (2000). Antioxidant activity of dietary polyphenols as determined by a modified ferric reducing/antioxidant power assay. *Journal of Agricultural and Food Chemistry*, 48(8), 3396-3402.
- Qin, D., Wu, H., Peng, H., Yao, Y., Ni, Z., Li, Z., Zhou, C. and Sun, Q. (2008). Heat stress-responsive transcriptome analysis in heat susceptible and tolerant wheat (*Triticum aestivum* L.) by using wheat genome array. *BMC Genomics*, 9(1), 1-19.
- Rabilloud, T., Adessi, C., Giraudel, A. and Lunardi, J. (1997). Improvement of the solubilization of proteins in two-dimensional electrophoresis with immobilized pH gradients. *Electrophoresis*, 18(3-4), 307-316.
- Rabilloud, T. and Lelong, C. (2011). Two-dimensional gel electrophoresis in proteomics: a tutorial. *Journal of proteomics*, 74(10), 1829-1841.
- Rahman, M. M., Joardder, M. U., Khan, M. I. H., Pham, N. D. and Karim, M. A. (2018). Multi-scale model of food drying: Current status and challenges. *Critical Reviews in Food Science and Nutrition*, 58(5), 858-876.

- Rajan, T. S., De Nicola, G. R., Lori, R., Rollin, P., Bramanti, P. and Mazzon, E. (2016). Anticancer activity of glucomoringin isothiocyanate in human malignant astrocytoma cells. *Fitoterapia*, 110(1), 1-7.
- Rajasekhar, M. D., Badri, K. R., Kumar, K. V., Kasseti, R. B., Fatima, S. S., Kumar, M. T. S. and Rao, C. A. (2010). Isolation and characterization of a novel antihyperglycemic protein from the fruits of *Momordica cymbalaria*. *Journal of Ethnopharmacology*, 128(1), 58-62.
- Ramakrishna, H., Murthy, S. S., Divya, R., MamathaRani, D. R. and Panduranga, M. G. (2012). Hydroxyl radical and DPPH scavenging activity of crude protein extract of *Leucas linifolia*: A folk medicinal plant. *Asian Journal of Plant Science and Research*, 2(1), 30-35.
- Ramli, A. N. M., Mahadi, N. M., Shamsir, M. S., Rabu, A., Joyce, T. K. H., Murad, A. M. A. and Illias, R. M. (2012). Structural prediction of a novel chitinase from the psychrophilic *Glaciozyma antarctica* PI12 and an analysis of its structural properties and function. *Journal of Computer-Aided Molecular Design*, 26(8), 947-961.
- Ramsak, Z., Baebler, S., Rotter, A., Korbar, M., Mozetic, I., Usadel, B. and Gruden, K. (2013). GoMapMan: intergration, consolidation and visualization of plant gene annotations within the MapMan ontology. *Nucleic Acids Research*, 42(1), 1167-1175.
- Rangaraju, A. A. and Rao, A. V. (2013). A review on molecular docking - Novel tool in drug design and analysis. *Journal of Harmonized Research in Pharmacy*, 2(4), 215-221.
- Rao, V. S., Das, S. K., Rao, V. J. and Srinubabu, G. (2008). Recent developments in life sciences research: Role of bioinformatics. *African Journal of Biotechnology*, 7(5), 495-503.
- Rao, N. K., Bethala, K., Sisinthy, S. P. and Rajeswari, K. S. (2013). Antidiabetic activity of *Orthosiphon stamineus* Benth roots in streptozotocin induced type 2 diabetic rats. *Asian Journal of Pharmaceutical and Clinical Research*, 7(1), 149-153.
- Ravindran, J., Subbaraju, G. V., Ramani, M. V., Sung, B. and Aggarwal, B. B. (2010). Bisdemethylcurcumin and structurally related hispolon analogues of curcumin exhibit enhanced prooxidant, anti-proliferative and anti-inflammatory activities *in vitro*. *Biochemical Pharmacology*, 79(11), 1658-1666.

- Reinbothe, T. M., Ivarsson, R., Li, D. Q., Niazi, O., Jing, X., Zhang, E., Stenson, L., Bryborn, U., and Renstrom, E. (2009). Glutaredoxin-1 mediates NADPH-dependent stimulation of calcium-dependent insulin secretion. *Molecular Endocrinology*, 23(6), 893-900.
- Rietveld, A. and Wiseman, S. (2003). Antioxidant effects of tea: evidence from human clinical trials. *The Journal of Nutrition*, 133(10), 3285-3292.
- Ripim, N. S. M., Fazil, N., Ibrahim, S. N. K., Bahtiar, A. A., Wai, Y. C., Ibrahim, N. and Nor, N. S. M. (2018). Antiviral Properties of *Orthosiphon stamineus* Aqueous Extract in Herpes Simplex Virus Type 1 Infected Cells. *Sains Malaysiana*, 47(8), 1725-1730.
- Riza, A. L., Pearson, F., Ugarte-Gil, C., Alisjahbana, B., Van de Vijver, S., Panduru, N. M., Hill, P. C., Ruslami, R., Moore, D., Aarnoutse, R., Critchley, J. A. and Van Crevel, R. (2014). Clinical management of concurrent diabetes and tuberculosis and the implications for patient services. *The Lancet Diabetes & Endocrinology*, 2(9), 740-753.
- Rizzello, C. G., Tagliazucchi, D., Babini, E., Rutella, G. S., Saa, D. L. T. and Gianotti, A. (2016). Bioactive peptides from vegetable food matrices: Research trends and novel technologies for synthesis and recovery. *Journal of Functional Foods*, 27(1), 549-569.
- Robert, X. and Gouet, P. (2014). Deciphering key features in protein structures with the new ENDscript server. *Nucleic Acids Research*, 42(1), 320-324.
- Robinson, M. M. and Zhang, X. (2011). *The world medicines situation 2011, traditional medicines: Global situation, issues and challenges*. Geneva, Switzerland: World Health Organization.
- Rodrigues, E. P., Torres, A. R., Batista, J. S. D. S., Huergo, L. and Hungria, M. (2012). A simple, economical and reproducible protein extraction protocol for proteomics studies of soybean roots. *Genetics and Molecular Biology*, 35(1), 348-352.
- Rollins, J. A., Habte, E., Templer, S. E., Colby, T., Schmidt, J. and Korff, von Korff, M. (2013). Leaf proteome alterations in the context of physiological and morphological responses to drought and heat stress in barley (*Hordeum vulgare* L.). *Journal of Experimental Botany*, 64(11), 3201-3212.

- Ross, G. S., Wegrzyn, T., MacRae, E. A. and Redgwel, R. J. (1994). Apple [beta]-galactosidase (Activity against cell wall polysaccharides and characterization of a related cDNA clone). *Plant Physiology*, 106(2), 521-528.
- Rosti, J., Barton, C. J., Albrecht, S., Dupree, P., Pauly, M., Findlay, K. Roberts, K. and Seifert, G. J. (2007). UDP-glucose 4-epimerase isoforms UGE2 and UGE4 cooperate in providing UDP-galactose for cell wall biosynthesis and growth of *Arabidopsis thaliana*. *The Plant Cell*, 19(5), 1565-1579.
- Roy, I. and Gupta, M.N. (2004). Freeze-drying of proteins: some emerging concerns. *Biotechnology and Applied Biochemistry*, 39(2), 165-177.
- Roy, T. and Lloyd, C. E. (2012). Epidemiology of depression and diabetes: a systematic review. *Journal of Affective Disorders*, 142(1), 8-21.
- Sacchetti, G., Maietti, S., Muzzoli, M., Scaglianti, M., Manfredini, S., Radice, M. and Bruni, R. (2005). Comparative evaluation of 11 essential oils of different origin as functional antioxidants, antiradicals and antimicrobials in foods. *Food chemistry*, 91(4), 621-632.
- Sahay, A. and Shakya, M. (2010). *In silico* analysis and homology modelling of antioxidant proteins of spinach. *Journal of Proteomics and Bioinformatics*, 3(5), 148-154.
- Saito, N., Kimura, M., Kuchiba, A. and Itokawa, Y. (1987). Blood thiamine levels in outpatients with diabetes mellitus. *Journal of Nutritional Science and Vitaminology*, 33(6), 421-430.
- Salama, S. M., Abdulla, M. A., AlRashdi, A. S., Ismail, S., Alkiyumi, S. S. and Golbabapour, S. (2013). Hepatoprotective effect of ethanolic extract of *Curcuma longa* on thioacetamide induced liver cirrhosis in rats. *BMC Complementary and Alternative Medicine*, 13(56), 1-17.
- Salem, R. B. S., Ktari, N., Bkhairia, I., Nasri, R., Mora, L., Kallel, R., Hamdi, S., Jamoussi, K., Boudaouara, T., El-Feki, A., Toldra, F. and Nasri, M. (2018). *In vitro* and *in vivo* antidiabetic and anti-hyperlipidemic effects of protein hydrolysates from *Octopus vulgaris* in alloxanic rats. *Food Research International*, 106(1), 952-963.
- Sanchez, M. C., Larrauri, J. A. and Saura, C. F. (1998). A procedure to measure the antiradical efficiency of polyphenols. *Journal of the Science of Food and Agriculture*, 76(1), 270-276.

- Saper, R., B., Phillips, R., S., Sehgal, A., Khouri, N., Davis, R., B., Paquin, J., Thuppil, V. and Kalses, S., N. (2008). Lead, mercury, and arsenic in US- and Indian-manufactured Ayurvedic medicines sold via the internet. *JAMA*, 300(8), 915–923.
- Saravanan, R. S. and Rose, J. K. (2004). A critical evaluation of sample extraction techniques for enhanced proteomic analysis of recalcitrant plant tissues. *Proteomics*, 4(9), 2522-2532.
- Sarkar, N. K., Kim, Y. K. and Grover, A. (2014). Coexpression network analysis associated with call of rice seedlings for encountering heat stress. *Plant Molecular Biology*, 84(1-2), 125-143.
- Sattar, N. A., Hussain, F., Iqbal, T. and Sheikh, M. A. (2012). Determination of *in vitro* antidiabetic effects of *Zingiber officinale* Roscoe. *Brazilian Journal of Pharmaceutical Sciences*, 48(4), 601-607.
- Schenk, G., Duggleby, R. G. and Nixon, P. F. (1998). Properties and functions of the thiamin diphosphate dependent enzyme transketolase. *The International Journal of Biochemistry & Cell Biology*, 30(12), 1297-1318.
- Schmid, N., Eichenberger, A. P., Choutko, A., Riniker, S., Winger, M., Mark, A. E. and van Gunsteren, W. F. (2011). Definition and testing of the GROMOS force-field versions 54A7 and 54B7. *European Biophysics Journal*, 40(7), 843.
- Schoepfer, A. M., Engel, A., Fattinger, K., Marbet, U. A., Criblez, D., Reichen, J., Zimmermann, A. and Oneta, C. M. (2007). Herbal does not mean innocuous: ten cases of severe hepatotoxicity associated with dietary supplements from Herbalife® products. *Journal of Hepatology*, 47(4), 521-526.
- Schuchardt, S. and Sickmann, A. (2007). Protein identification using mass spectrometry: a method overview. In *Plant Systems Biology* (pp. 141-170). Switzerland: Birkhäuser Basel.
- Schulze, W. X. and Usadel, B. (2010). Quantitation in mass-spectrometry-based proteomics. *Annual Review of Plant Biology*, 61(1), 491-516.
- Seghieri, G., Di Simplicio, P., De Giorgio, L. A., Anichini, R., Alberti, L. and Franconi, F. (2000). Relationship between metabolic glycaemic control and platelet content of glutathione and its related enzymes, in insulin-dependent diabetes mellitus. *Clinica chimica acta*, 299(1-2), 109-117.

- Seifert, G. J., Barber, C., Wells, B., Dolan, L. and Roberts, K. (2002). Galactose biosynthesis in *Arabidopsis*: genetic evidence for substrate channeling from UDP-D-galactose into cell wall polymers. *Current Biology*, 12(21), 1840-1845.
- Sensi, E., Mazzuca, S. and Cresti, M. (2003). Protein extraction for two-dimensional electrophoresis from olive leaf, a plant tissue containing high levels of interfering compounds. *Electrophoresis*, 24(14), 2369-2375.
- Sewram, V., Shephard, G. S., van der Merwe, L. and Jacobs, T. V. (2006). Mycotoxin contamination of dietary and medicinal wild plants in the Eastern Cape Province of South Africa. *Journal of Agricultural and Food Chemistry*, 54(15), 5688-5693.
- Shadkam, M. N., Mozaffari-Khosravi, H. and Mozayan, M. R. (2010). A comparison of the effect of honey, dextromethorpan, and diphenhydramine on nightly cough and sleep quality in children and their parents. *Journal of Alternative and Complementary Medicine*, 16(7), 787-793.
- Shafaei, A., Esmaili, K., Farsi, E., Aisha, A. F., Majid, A. M. S. A. and Ismail, Z. (2015). Genotoxicity, acute and subchronic toxicity studies of nano liposomes of *Orthosiphon stamineus* ethanolic extract in Sprague Dawley rats. *BMC Complementary and Alternative Medicine*, 15(1), 360-374.
- Shafaei, A., Sultan Khan, M. S., Aisha, A. F., Abdul Majid, A. M. S., Hamdan, M. R., Mordi, M. N. and Ismail, Z. (2016). Flavonoids-rich *Orthosiphon stamineus* extract as new candidate for angiotensin I-converting enzyme inhibition: A molecular docking study. *Molecules*, 21(11), 1500-1516.
- Shah, P., Gutierrez-Sanchez, G., Orlando, R. and Bergmann, C. (2009). A proteomic study of pectin-degrading enzymes secreted by *Botrytis cinerea* grown in liquid culture. *Proteomics*, 9(11), 3126-3135.
- Shahidi, F., and Zhong, Y. (2008). Bioactive peptides. *Journal of AOAC International*, 91(4), 914-931.
- Shaw, D., Ladds, G., Duez, P., Williamson, E. and Chan, K. (2012). Pharmacovigilance of herbal medicine. *Journal of Ethnopharmacology*, 140(1), 513-518.
- Shim, S. Y. and Gam, L. H. (2012). Analysis of *Piper sarmentosum* proteome using two dimensional gel electrophoresis and mass spectrometry. *Asia-Pacific Journal of Molecular Biology and Biotechnology*, 20(4), 124-139.

- Shishikura, Y. and Khokhar, S. (2005). Factors affecting the levels of catechins and caffeine in tea beverage: estimated daily intakes and antioxidant activity. *Journal of the Science of Food and Agriculture*, 85(12), 2125-2133.
- Siaud, N., Dray, E., Gy, I., Gerard, E., Takvorian, N. and Doutriaux, M. P. (2004). Brca2 is involved in meiosis in *Arabidopsis thaliana* as suggested by its interaction with Dmc1. *The EMBO Journal*, 23(6), 1392-1401.
- Siddiqui, M., J., Hafizoh, S., N., Ismail, Z., Sahib, H., B., Helai, M., H., S. and Abdul Majid, A., M., S. (2009). Analysis of total proteins, polysaccharides and glycosaponins contents of *Orthosiphon stamineus* Benth. In spray and freeze dried methanol: water(1:1) extract and its contribution to cytotoxic and antiangiogenic activities. *Pharmacognosy Research*, 1(5), 320-326.
- Sidhu, A. K., Wani, S. J., Tamboli, P. S. and Patil, S. N. (2014). *In vitro* evaluation of anti-diabetic activity of leaf and callus extracts of *Costus pictus*. *International Journal of Science and Research*, 3(6), 1622-1625.
- Singh, I. and Shah, K. (2012). *In silico* study of interaction between rice proteins enhanced disease susceptibility 1 and phytoalexin deficient 4, the regulators of salicylic acid signalling pathway. *Journal of Biosciences*, 37(3), 563-571.
- Singh, V., Singh, B., Joshi, R., Jaju, P. and Pati, P. K. (2017). Changes in the leaf proteome profile of *Withania somnifera* (L.) Dunal in response to *Alternaria alternata* infection. *PLoS one*, 12(6), 1-19.
- Sreedhar, A., Prakash, S., Sapna, N. and Kumar, S. (2011). Proteomics-the new era of periodontics. *Cancer*, 6(1), 87-90.
- Sriplang, K., Adisakwattana, S., Rungsipipat, A. and Yibchok-Anun, S. (2007). Effects of *Orthosiphon stamineus* aqueous extract on plasma glucose concentration and lipid profile in normal and streptozotocin-induced diabetic rats. *Journal of Ethnopharmacology*, 109(3), 510-514.
- Sterling, H. J., Batchelor, J. D., Wemmer, D. E. and Williams, E. R. (2010). Effects of buffer loading for electrospray ionization mass spectrometry of a noncovalent protein complex that requires high concentrations of essential salts. *Journal of the American Society for Mass Spectrometry*, 21(6), 1045-1049.
- Stivala, L. A., Savio, M., Cazzalini, O., Pizzala, R., Rehak, L., Bianchi, L., Vannini, V. and Prospero, E. (1996). Effect of β -carotene on cell cycle progression of human fibroblasts. *Carcinogenesis*, 17(11), 2395-2401.

- Stoner, G. D., Morrissey, D. T., Heur, Y. H., Daniel, E. M., Galati, A. J. and Wagner, S. A. (1991). Inhibitory effects of phenethyl isothiocyanate on N-nitrosobenzylmethylamine carcinogenesis in the rat esophagus. *Cancer Research*, 51(8), 2063-2068.
- Suganya, J. A. and Mahendran, R. (2016). In Silico qsar and molecular docking studies of selected medicinal plant compounds against ns5 & ns3 protein of dengue virus: a comparative approach. *International Journal of Pharma and Bio Sciences*, 7(3), 1135-1144.
- Sugie, S., Okumura, A., Tanaka, T. and Mori, H. (1993). Inhibitory effects of benzyl isothiocyanate and benzyl thiocyanate on diethylnitrosamine-induced hepatocarcinogenesis in rats. *Japanese Journal of Cancer Research*, 84(8), 865-870.
- Sumaryono, W., Proksch, P., Wray, V., Witte, L. and Hartmann, T. (1991). Qualitative and quantitative analysis of the phenolic constituents from *Orthosiphon aristatus*. *Planta medica*, 57(2), 176-180.
- Sun, W., Zhang, H., Guo, J., Zhang, X., Zhang, L., Li, C. and Zhang, L. (2016). Comparison of the efficacy and safety of different ACE inhibitors in patients with chronic heart failure. *Medicine*, 95(6), 1-8.
- Sundarammal, S., Thirugnanasampandan, R. and Selvi, M. T. (2012). Chemical composition analysis and antioxidant activity evaluation of essential oil from *Orthosiphon thymiflorus* (Roth) Sleesen. *Asian Pacific Journal of Tropical Biomedicine*, 2(1), 112-115.
- Surabhi, G. K., Pattanaik, S. and Mohanty, S. (2016). A comparative method for protein extraction and proteome analysis by two-dimensional gel electrophoresis from banana fruit. *Horticultural Biotechnology Research*, 2(1), 8-13.
- Suzuki, N., Koussevitzky, S., Mittler, R. and Miller, G. (2012). ROS and redox signalling in the response of plants to abiotic stress. *Plant, Cell & Environment*, 35(2), 259-270.
- Switzar, L., Giera, M. and Niessen, W. M. A. (2013). Protein digestion: An Overview of the available techniques and recent developments. *Journal of Proteome Research*, 12(3), 1067-1077.

- Szymańska, S., Płociniczak, T., Piotrowska-Seget, Z., Złoch, M., Ruppel, S. and Hryniewicz, K. (2016). Metabolic potential and community structure of endophytic and rhizosphere bacteria associated with the roots of the halophyte *Aster tripolium* L. *Microbiological Research*, 182(1), 68-79.
- Tabana, Y. M., Al-Suede, F. S. R., Ahamed, M. B. K., Dahham, S. S., Hassan, L. E. A., Khalilpour, S., Taleb-Agha, M., Sandai, D., Majid, A. S. A. and Majid, A. M. S. A. (2016). Cat's whiskers (*Orthosiphon stamineus*) tea modulates arthritis pathogenesis via the angiogenesis and inflammatory cascade. *BMC Complementary and Alternative Medicine*, 16(1), 1-11.
- Tabb, D. L. (2013). Quality assessment for clinical proteomics. *Clinical biochemistry*, 46(6), 411-420.
- Takeda, Y., Matsumoto, T., Terao, H., Shingu, T., Futatsuishi, Y., Nohara, T. and Kajimoto, T. (1993). Orthosiphon D and E, minor diterpenes from *Orthosiphon stamineus*. *Phytochemistry*, 33(2), 411-415.
- Tandon, G., Jaiswal, S., Iquebal, M., Kumar, S., Kaur, S., Rai, A. and Kumar, D. (2015). Evidence of salicylic acid pathway with EDS1 and PAD4 proteins by molecular dynamics simulation for grape improvement. *Journal of Biomolecular Structure and Dynamics*, 33(10), 2180-2191.
- Tang, W., Yuan, H., Zhang, H., Wang, L., Qian, H. and Qi, X. (2015). An antimicrobial peptide screened from casein hydrolyzate by *Saccharomyces cerevisiae* cell membrane affinity method. *Food Control*, 50(1), 413-422.
- Tavasalkar, S. U., Mishra, H. N. and Madhavan, S. (2012). Evaluation of antioxidant efficacy of natural plant extracts against synthetic antioxidants in sunflower oil. *Open Access Scientific Reports*, 1(11), 1-5.
- Tegar, M. and Purnomo, H. (2013). Tea leaves extracted as anti-malaria based on molecular docking PLANTS. *Procedia Environmental Sciences*, 17(1), 188-194.
- Teige, M., Melzer, M. and Süß, K. H. (1998). Purification, properties and in situ localization of the amphibolic enzymes D-ribulose 5-phosphate 3-epimerase and transketolase from spinach chloroplasts. *European Journal of Biochemistry*, 252(2), 237-244.

- Tepkeeva I. I., Moiseeva E. V., Chaadaeva A. V., Zhavoronkova E. V., Kessler Y. V., Semushina S. G. and Demushkin V. P. (2008). Evaluation of antitumor activity of peptide extracts from medicinal plants on the model of transplanted breast cancer in CBRB-Rb (8.17)1Iem mice. *Bulletin of Experimental Biology and Medicine*, 145(4), 464-466.
- Tew, K. D. (1994). Glutathione-associated enzymes in anticancer drug resistance. *Cancer Research*, 54(16), 4313-4320.
- Tezuka, Y., Stampoulis, P., Banskota, A. H., Awale, S., Tran, K. Q., Saiki, I. and Kadota, S. (2000). Constituents of the Vietnamese medicinal plant *Orthosiphon stamineus*. *Chemical and Pharmaceutical Bulletin*, 48(11), 1711-1719.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-zevallos, L. and Hawkins, D. (2006). Comparison of ABTS, DPPH, FRAP and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19(6-7), 669-675.
- Théolier, J., Hammami, R., Labelle, P., Fliss, I. and Jean, J. (2013). Isolation and identification of antimicrobial peptides derived by peptic cleavage of whey protein isolate. *Journal of Functional Foods*, 5(2), 706-714.
- Thong, K. Gupta, P. S., Blann, A. and Ryder, R. (2015). The influence of age and metformin treatment status on reported gastrointestinal side effects with liraglutide treatment in Type 2 diabetes. *Diabetes Research and Clinical Practice*, 109(1), 124-129.
- Tobin, P. H., Richards, D. H., Callender, R. A., and Wilson, C. J. (2014). Protein engineering: a new frontier for biological therapeutics. *Current Drug Metabolism*, 15(7), 743-756.
- Townsend, D. M., Tew, K. D. and Tapiero, H. (2003). The importance of glutathione in human disease. *Biomedicine and Pharmacotherapy*, 57(3-4), 145-155.
- Trainotti, L., Spinello, R., Piovan, A., Spolaore, S. and Casadoro, G. (2001). β -Galactosidases with a lectin-like domain are expressed in strawberry. *Journal of Experimental Botany*, 52(361), 1635-1645.
- Traka, M., and Mithen, R. (2009). Glucosinolates, isothiocyanates and human health. *Phytochemistry Reviews*, 8(1), 269-282.
- Tripathi, A. A. and Misra, K. (2017). Molecular docking: a structure-based drug designing approach. *JSM Chemistry*, 5(2), 1042-1046.

- Trofimiuk, E., Walesiuk, A. and Braszko, J. J. (2005). St John's wort (*Hypericum perforatum*) diminishes cognitive impairment caused by the chronic restraint stress in rats. *Pharmacological Research*, 51(3), 239-246.
- Trott, O. A. and Olson, A. J. (2010). AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *Journal of Computational Chemistry*, 31(2), 455-461.
- Tsugita, A., Kawakami, T., Uchiyama, Y., Kamo, M., Miyatake, N. and Nozu, Y. (1994). Separation and characterization of rice proteins. *Electrophoresis*, 15(5), 708-720.
- Upston, J. M., Kritharides, L. and Stocker, R. (2003). The role of vitamin E in atherosclerosis. *Progress in Lipid Research*, 42(5), 405-422.
- Ushimaru, P. I., Silva, M. T. N. d., Di Stasi, L. C., Barbosa, L. and Fernandes Junior, A. (2007). Antibacterial activity of medicinal plant extracts. *Brazilian Journal of Microbiology*, 38(4), 717-719.
- Usman, K. (2014). *Protein Profiling of Orthosiphon stamineus*. Master Thesis. Universiti Teknologi Malaysia, Skudai.
- Usmanov, R. A. and Kochetov, G. A. (1978). Study of different conformational states of transketolase by the method of perturbation UV spectrophotometry. *Biokhimiia*, 43(1), 1796-1804.
- Uvackova, L., Ondruskova, E., Danchenko, M., Skultety, L., Miernyk, J. A., Hrubik, P. and Hajdich, M. (2014). Establishing a leaf proteome reference map for *Ginkgo biloba* provides insight into potential ethnobotanical uses. *Journal of Agricultural and Food Chemistry*, 62(47), 11547-11556.
- Van den Bergh, G. and Arckens, L. (2004). Fluorescent two-dimensional difference gel electrophoresis unveils the potential of gel-based proteomics. *Current Opinion in Biotechnology*, 15(1), 38-43.
- Van der Werf, M. J., Schuren, F. H. J., Bijlsma, S., Tas, A. C., & Ommen, B. V. (2001). Nutrigenomics: application of genomics technologies in nutritional sciences and food technology. *Journal of Food Science*, 66(6), 772-780.
- Vanderschuren, H., Lentz, E., Zainuddin, I. and Gruissem, W. (2013). Proteomics of model and crop plant species: status, current limitations and strategic advances for crop improvement. *Journal of proteomics*, 93(1), 5-19.

- Vanherweghem, J., L., Depierreux, M., Tielemans, C., Abramowicz, D., Dratwa, M., Jadoul, M., Richard, C., Vandervelde, D., Verbeelen, D. and Jadoul, M. (1993). Rapidly progressive interstitial renal fibrosis in young women: association with slimming regimen including Chinese herbs. *Lancet*, 341 (8842), 387-391.
- Vanherweghem, J. L. (2000). Nephropathy and herbal medicine. *American Journal of Kidney Disease*, 35(2), 330-332.
- Varlibas, F., Delipoyraz, I., Yuksel, G., Filiz, G., Tireli, H. and Gecim, N. O. (2009). Neurotoxicity following chronic intravenous use of “Russian cocktail”. *Clinical Toxicology*, 47(2), 157-160.
- Veeresham, C. (2012). Natural products derived from plants as a source of drugs. *Journal of Advanced Pharmaceutical Technology & Research*, 3(4), 200-201.
- VerBerkmoes, N. C., Hettich, R. L., Bruce, B. D., Nguyen, R. and Savage, T. L. (2002). One-end two-dimensional LC/MS/MS analysis of *Arabidopsis thaliana* proteome. *LC GC NORTH AMERICA*, 20(8), 10-11.
- Verhees, C. H., Denise, G. M., Ettema, T. J. and Dijkema, C. (2002). Biochemical adaptations of two sugar kinases from the hyperthermophilic archaeon *Pyrococcus furiosus*. *Biochemical Journal*, 366(1), 121-127.
- Vierling, E. (1991). The role of heat shock proteins in plant. *Annual Review of Plant Physiology and Plant Molecular Biology*, 42(1), 579-620.
- Vilhena, M. B., Franco, M. R., Schmidt, D., Carvalho, G. and Azevedo, R. A. (2015). Evaluation of protein extraction methods for enhanced proteomic analysis of tomato leaves and roots. *Anais da Academia Brasileira de Ciências*. 87(3), 1853-1863.
- Vitkup, D., Ringe, D., Petsko, G., A. and Karplus, M. (2000). Solvent mobility and the protein 'glass' transition. *Nature Structural and Molecular Biology*, 7(1), 34-38.
- Vogeser, M. and Parhofer, K. (2007). Liquid chromatography tandem-mass spectrometry (LC-MS/MS)-technique and applications in endocrinology. *Experimental and Clinical Endocrinology & Diabetes*, 115(9), 559-570.
- Vogt, G. and Argos, P. (1997). Protein thermal stability: hydrogen bonds or internal packing? *Folding and Design*, 2(1), 40-46.

- Vyankatrao, N. P., Arts, N. T. A. and Commerce, V. S. S. (2014). Effect of drying methods on nutritional value of some vegetables. In *Proceeding of the National Conference on Conservation of Natural Resources & Biodiversity for Sustainable Development*, 6(1), 72-79.
- Wada, T., Sasaki, M., Kataoka, H., Tanida, S., Itoh, K., Ogasawara, N., Oshima, T., Togawa, S., Kubota, E., Yamada, T., Mori, Y., Fujita, F., Ohara, H., Nakao, H., Sobue, S., Joh, T. and Itoh, M. (2005). Efficacy of famotidine and omeprazole in healing symptoms of non-erosive gastro-oesophageal reflux disease: randomized-controlled study of gastro-oesophageal reflux disease. *Alimentary Pharmacology & Therapeutics*, 21(2), 2-9.
- Wahab, N. A. A., Abdullah, N. and Aminudin, N. (2014). Characterisation of potential antidiabetic-related proteins from *Pleurotus pulmonarius* (Fr.) Qué.(grey oyster mushroom) by MALDI-TOF/TOF mass spectrometry. *BioMed Research International*, 2014(1), 1-9.
- Wahid, A., Gelani, S., Ashraf, M. and Foolad, M. R. (2007). Heat tolerance in plants: An overview. *Environmental and Experimental Botany*, 61(3), 199-223.
- Wang, W. (2000). Lyophilization and development of solid protein pharmaceuticals. *International Journal of Pharmaceutics*, 203(1-2), 1-60.
- Wang, L. L. and Xiong, Y. L. (2005). Inhibition of lipid oxidation in cooked beef patties by hydrolyzed potato protein is related to its reducing and radical scavenging ability. *Journal of Agricultural and Food Chemistry*, 53(23), 9186-9192.
- Wang, W., Vignani, R., Scali, M. and Cresti, M. (2006). A universal and rapid protocol for protein extraction from recalcitrant plant tissues for proteomic analysis. *Electrophoresis*, 27(13), 2782-2786.
- Wang, W., Tai, F. and Chen, S. (2008). Optimizing protein extraction from plant tissues for enhanced proteomics analysis. *Journal of Separation Science*, 31(11), 2032-2039.
- Wang, S., Durrant, W. E., Song, J., Spivey, N. W. and Dong, X. (2010). Arabidopsis BRCA2 and RAD51 proteins are specifically involved in defense gene transcription during plant immune responses. *Proceedings of the National Academy of Sciences*, 107(52), 22716-22721.

- Wang, X., Dinler, B. S., Vignjevic, M., Jacobsen, S. and Wollenweber B. (2015). Physiological and proteome studies of responses to heat stress during grain filling in contrasting wheat cultivars. *Plant Science*, 230(1), 33-50.
- Waterman, C., Cheng, D. M., Rojas-Silva, P., Poulev, A., Dreifus, J., Lila, M. A. and Rskin, I. (2014). Stable, water extractable isothiocyanates from *Moringa oleifera* leaves attenuate inflammation *in vitro*. *Phytochemistry*, 103(1), 114-122.
- Wattanasiritham, L., Kubglomsong, S. and Theerakulkait, C. (2015). Antioxidant activity of rice bran protein extract, its enzymatic hydrolysates and its combination with commercial antioxidants. *Pakistan Journal of Nutrition*, 14(10), 647-52.
- Wickenberg, J., Ingemansson, S. L. and Hlebowicz, J. (2010). Effects of *Curcuma longa* (turmeric) on postprandial plasma glucose and insulin in healthy subjects. *Nutrition Journal*, 9(1), 1-5.
- Wilm, M. (2011). Principles of electrospray ionization. *Molecular & Cellular Proteomics*, 10(7), 1-30.
- Wiriya, P., Paiboon, T. and Somchart, S. (2009). Effect of drying air temperature and chemical pretreatments on quality of dried chilli. *International Food Research Journal*, 16(3), 441-454.
- Witchel, S. F. and Plant, T. M. (2009). Puberty: Gonadarche and Adrenarche. In *Yen & Jaffe's Reproductive Endocrinology* (pp. 395-431). London, United Kingdom: Elsevier.
- Wong, E., Backholer, K., Gearon, E., Harding, J., Freak-Poli, R., Stevenson, C. and Peeters, A. (2013). Diabetes and risk of physical disability in adults: a systematic review and meta-analysis. *The Lancet Diabetes & Endocrinology*, 1(2), 106-114.
- Woo, C. S. J., Lau, J. S. H., & El-Nezami, H. (2012). Herbal medicine: toxicity and recent trends in assessing their potential toxic effects. In *Advances in Botanical Research*, 62(1), 365-384.
- World Health Organization. (2015). Global Health Observatory Data Repository. World Health Organization. Geneva, Switzerland.
- World Health Organization. (2016). Global Tuberculosis Report 2016. World Health Organization. Geneva, Switzerland.

- Wresdiyati, T., Sa'diah, S. I. T. I., Winarto, A. and Febriyani, V. (2015). Alpha-glucosidase inhibition and hypoglycemic activities of *Sweitenia mahagoni* seed extract. *HAYATI Journal of Biosciences*, 22(2), 73-78.
- Wu, W., Tang, X., Hu, W., Lotan, R., Hong, W. K. and Mao, L. (2002). Identification and validation of metastasis-associated proteins in head and neck cancer cell lines by two-dimensional electrophoresis and mass spectrometry. *Clinical & Experimental Metastasis*, 19(4), 319-326.
- Wu, H. C., Chen, H. M. and Shiau, C. Y. (2003). Free amino acids and peptides as related to antioxidant properties in protein hydrolysates of mackerel (*Scomber austriasicus*). *Food Research International*, 36(9-10), 949-957.
- Wu, Y., Fan, Y., Xue, B., Luo, L., Shen, J., Zhang, S., Jiang, Y. and Yin, Z. (2006). Human glutathione S-transferase P1-1 interacts with TRAF2 and regulates TRAF2-ASK1 signals. *Oncogene*, 25(42), 5787-5800.
- Wu, X., Xiong, E., Wang, W., Scali, M. and Cresti, M. (2014). Universal sample preparation method integrating trichloroacetic acid/acetone precipitation with phenol extraction for crop proteomic analysis. *Nature Protocols*, 9(2), 362-374.
- Xiao, H. W., Gao, Z. J., Lin, H. and Yang, W. X. (2010). Air impingement drying characteristics and quality of carrot cubes. *Journal of Food Process Engineering*, 33(5), 899-918.
- Xie, H., Li, Y., Yu, F., Xie, X., Qiu, K. and Fu, J. (2015). An investigation of molecular docking and molecular dynamic simulation on imidazopyridines as b-raf kinase inhibitors. *International Journal of Molecular Sciences*, 16(11), 27350-27361.
- Xu, J., Jiao, F. and Yu, L. (2008). Protein structure prediction using threading. In *Protein Structure Prediction* (pp. 91-121). Totowa: Humana Press.
- Xu, J., Lan, H., Fang, H., Huang, X., Zhang, H. and Huang, J. (2015). Quantitative proteomic analysis of the rice (*Oryza sativa* L.) salt response. *PLoS One*, 10(3), 1-19.
- Xue, B., Wu, Y., Yin, Z., Zhang, H., Sun, S., Yi, T. and Luo, L. (2005). Regulation of lipopolysaccharide-induced inflammatory response by glutathione S-transferase P1 in RAW264.7 cells. *FEBS Letters*, 579(19), 4081-4087.

- Yamamoto, R., Inouhe, M. and Masuda, Y. (1988). Galactose inhibition of auxin-induced growth of mono- and dicotyledonous plants. *Plant Physiology*, 86(4), 1223-1227.
- Yang, Y., Yue, Y., Runwei, Y. and Guolin, Z. (2010). Cytotoxic, apoptotic and antioxidant activity of the essential oil of *Amomum tsao-ko*. *Bioresource Technology*, 101(11), 4205-4211.
- Yang, C. Q., Fang, X., Wu, X. M., Mao, Y. B., Wang, L. J. and Chen, X. Y. (2012). Transcriptional regulation of plant secondary metabolism F. *Journal of Integrative Plant Biology*, 54(10), 703-712.
- Yang, D., Du, X., Yang, Z., Liang, Z., Guo, Z. and Liu, Y. (2014). Transcriptomics, proteomics, and metabolomics to reveal mechanisms underlying plant secondary metabolism. *Engineering in Life Sciences*, 14(5), 456-466.
- Yehya, A. H. S., Asif, M., Kaur, G., Hassan, L. E. A., Al-Suede, F. S. R., Abdul Majid, A. M. S. and Oon, C. E. (2018). Toxicological studies of *Orthosiphon stamineus* (Misai Kucing) standardized ethanol extract in combination with gemcitabine in athymic nude mice model. *Journal of Advanced Research*, 15(1), 59-68.
- Yin, Z., Ivanov, V. N., Habelhah, H., Tew, K and Ronai, Z. (2000). Glutathione S-transferase p elicits protection against H₂O₂-induced cell death via coordinated regulation of stress kinases. *Cancer Research*, 60(15), 4053-4057.
- Yoon, Y., Galloway, C. A., Jhun, B. S. and Yu, T. (2011). Mitochondrial dynamics in diabetes. *Antioxidants & redox signaling*, 14(3), 439-457.
- Yoshida, K., Kaothien, P., Matsui, T., Kawaoka, A. and Shinmyo, A. (2003). Molecular biology and application of plant peroxidase. *Applied Microbiology and Biotechnology*, 60(6), 665-670.
- You, W. P. and Henneberg, M. (2016). Type 1 diabetes prevalence increasing globally and regionally: the role of natural selection and life expectancy at birth. *BMJ Open Diabetes Research & Care*, 4(1), 1-7.
- Yu, H., Zhong, S., Chan, K. and Berry, M., I. (1995). Pharmacognostical investigation on traditional Chinese medicinal herbs: identification of four herbs from the UK market. *Journal of Pharmacy and Pharmacology*, 47(12), 1129-1129.
- Yu, H., Zhong, S., Berry, M., I. and Chan, K. (1997). Pharmacognostical investigation on traditional Chinese medicinal herbs: identification of six herbs from the UK market. *Journal of Pharmacy and Pharmacology*, 49(1), 212-212.

- Yuan, Z., Bailey, T. L. and Teasdale, R. D. (2005). Prediction of protein B-factor profiles. *Proteins: Structure, Function, and Bioinformatics*, 58(4), 905-912.
- Yue, Y. T., Zhang, X. F., Pan, J., Ou-Yang, Z., Wu, J. and Yang, M. H. (2010). Determination of deoxynivalenol in medicinal herbs and related products by GC–ECD and confirmation by GC–MS. *Chromatographia*, 71(5-6), 533-538.
- Yuliana, N. D., Khatib, A., Link-Struensee, A. M., Ijzerman, A. P., Rungkat-Zakaria, F., Choi, Y. H. and Verpoorte, R. (2009). Adenosine A1 receptor binding activity of methoxy flavonoids from *Orthosiphon stamineus*. *Planta Medica*, 75(2), 132-136.
- Zadraznik, T., Hollung, K., Egge-Jacobsen, W., Meglic, V. and Sustar-Vozlic, J. (2013). Differential proteomic analysis of drought stress response in leaves of common bean (*Phaseolus vulgaris* L.). *Journal of Proteomics*, 78(1), 254-272.
- Zahir, F., Rizwi, S. J., Haq, S. K. and Khan, R. H. (2005). Low dose mercury toxicity and human health. *Environmental Toxicology and Pharmacology*, 20(2), 351-360.
- Zakaria, Z., Aziz, R., Lachimanan, Y. L., Sreenivasan, S. and Rathinam, X. (2008). Antioxidant activity of *Coleus blumei*, *Orthosiphon stamineus*, *Ocimum basilicum* and *Mentha arvensis* from Lamiaceae family. *Internatioanl Journal of Natural and Engineering Sciences*, 2(1), 93-95.
- Zambrowicz, A., Eckert, E., Pokora, M., Bobak, Ł., Dąbrowska, A., Szoltysik, M., Trziszka, T. and Chrzanowska, J. (2015). Antioxidant and antidiabetic activities of peptides isolated from a hydrolysate of an egg-yolk protein by-product prepared with a proteinase from Asian pumpkin (*Cucurbita ficifolia*). *RSC Advances*, 5(14), 10460-10467.
- Zhang, J., Zhang, H., Wang, L., Guo, X., Wang, X. and Yao, H. (2009). Antioxidant activities of the rice endosperm protein hydrolysate: identification of the active peptide. *European Food Research and Technology*, 229(4), 709-719.
- Zhang, G., Annan, R. S., Carr, S. A. and Neubert, T. A. (2010). Overview of peptide and protein analysis by mass spectrometry. *Current protocols in protein science*, 62(1), 1-16.
- Zhang, H. and Ge, Y. (2011). Comprehensive analysis of protein modifications by top-down mass spectrometry. *Circulation: Genomic and Precision Medicine*, 4(6), 711-711.

- Zhang, L., Yan, J., Liu, X., Ye, Z., Yang, X., Meyboom, R., Chan, K., Shaw, D. and Duez, P. (2012). Pharmacovigilance practice and risk control of Traditional Chinese Medicine drugs in China: Current status and future perspective. *Journal of Ethnopharmacology*, 140(3), 519–525.
- Zhao J. and Zhong C. J. (2009). A review on research progress of transketolase. *Neuroscience Bulletin*, 25(2), 94-99.
- Zhuang, J. P., Su, J., Li, X. P. and Chen, W. X. (2006). Cloning and expression analysis of beta-galactosidase gene related to softening of banana (*Musa* sp.) fruit. *Journal of Plant Physiology and Molecular Biology*, 32(4), 411-419.
- Zhuo, Y., Cao, X., Yang, Y., Wang, J., Yang, W., Ben, P., Shen, L., Cao, P., Luo, L. and Yin, Z. (2018). Glutathione S-transferase Pi prevents sepsis-related high mobility group box-1 protein translocation and release. *Frontiers in Immunology*, 9(1), 1-17.
- Zuo, X. and Speicher, D. W. (2000). Quantitative evaluation of protein recoveries in two-dimensional electrophoresis with immobilized pH gradients. *Electrophoresis*, 21(14), 3035-3047.

LIST OF PUBLICATIONS

Journal with Impact Factor

1. **Ng, M. L.**, Shahir, M. S. & Zaidah, R. (2019). Molecular modeling and simulation of transketolase from *Orthosiphon stamineus*. *Current Computer-Aided Drug Design*. 10.2174/1573409914666181022141753. (**Q4, IF: 0.77**)

Non-Indexed Conference Proceedings

1. **Ng, M. L.**, Usman, K. & Zaidah, R. (2016). Leaf proteome and simulation of transketolase from *Orthosiphon stamineus*. In *International Conference on Biosciences & Medical Engineering* (pp. 125). ICBME2016.
2. **Ng, M. L.**, Shahir, M. S & Zaidah, R. (2017). Molecular docking and simulation of transketolase from *Orthosiphon stamineus*. In *International Postgraduate Symposium in Biotechnology 2017* (pp. 1-5). Universiti Teknologi Malaysia.
3. **Ng, M. L.**, Shahir, M. S & Zaidah, R. (2018). Optimization of two-dimensional gel electrophoresis for *Orthosiphon stamineus* leaf proteome analysis. In *7th International Graduate Conference of Engineering, Science and Humanities* (pp.482-484). Universiti Teknologi Malaysia.
4. **Ng, M. L.**, Shahir, M. S. & Zaidah, R. (2019). Effect of hot and cold infusion of *Orthosiphon stamineus* on the antioxidant activity. In *2nd International Conference on Biosciences and Medical Engineering* (pp. 15). Universiti Teknologi Malaysia.

Book Chapter

1. **Ng, M. L. & Zaidah, R.** (2018). Impact of various drying methods on plant nutrient. In *Plant Science and Its Application* (167-190). Universiti Teknologi Malaysia.