

OPTIMIZATION OF CASCADE LOW-PRESSURE STEAM HEATING AND
ORGANIC ACID PRE-TREATMENT ON PINEAPPLE WASTES FOR
FERMENTABLE SUGARS PRODUCTION

NORHAFIZA BINTI NORDIN

A thesis submitted in fulfilment of the
requirements for the award of the degree of
Master of Philosophy

School of Chemical and Energy Engineering
Faculty of Engineering
Universiti Teknologi Malaysia

MARCH 2022

DEDICATION

Special dedicated to

My beloved father and mother

Nordin bin Othman

Haslina binti Hussein

My beloved husband

Mohammad Amir Zharif bin Mohamad Azilan

My Siblings

Norhana binti Nordin

Norhaida binti Nordin

Norhazwani binti Nordin

Muhammad Azammuddin Nordin

Muhammad Azimmuddin Nordin

My fellow friends

Nurfadhila Nasya Binti Ramlee

Nur Diana binti Abdul Razak

Nur Hidayah Kumar binti Firdaus Kumar

ACKNOWLEDGEMENT

First and foremost, I would like to convey my gratefulness to The Almighty Allah S.W.T. with His blessings for giving me strength and the ability that has been given to me to complete my master project within the planned time.

I would like to express my deepest appreciation to my supervisor, Dr. Nur Izyan binti Wan Azelee for her invaluable patience, guidance, advice, and encouragement throughout the project and completion of the thesis. I also wish to acknowledge the help and the information sharing of all staff members of School of Chemical & Energy Engineering, Faculty of Engineering, and the other post-graduates colleagues, who have been provided me their kind assistance, either directly or indirectly.

However, it would not be possible without my parent's endless moral support, financial support and encouragement, which makes me very grateful. Last but not least, Thanks to all my fellow laboratory mates and colleagues, post-graduate senior from Genetic Laboratory for supervision and continuous encouragement in making all the tasks possible. They helped me by giving their continuous information and mentally support throughout my two years of master's study in UTM.

ABSTRACT

Sugar synthesized from lignocellulosic biomass can potentially supplement the increasing demand of various applications in industries. Pre-treatment of lignocellulosic biomass is challenging due to the need to break the rigid and compact structure to produce sugar. In this study, pineapple waste (PW) were subjected to a cascade pre-treatment of (i) low pressure steam heating (LPSH) and (ii) maleic acid (MA), which aims to improve delignification, increase enzyme accessibility to cellulose and hemicellulose in the PW while reducing the production of inhibitors. The best conditions for pressure (kPa), solid loading (% w/v) and time (min) of LPSH were screened. The resulting solid biomass from LPSH were proceeded to subsequent pre-treatment of MA and optimized by response surface methodology-based Box-Behnken design, where the influence of pre-treatment temperature, acid concentration and time were studied. A total of 68 % w/w delignification with high hemicellulose removal (79.5 %) were achieved while 77.6 % cellulose was retained in the solid residue after cascading both pre-treatments. No 5-hydroxymethyl furfural (5-HMF) and acceptable amount of furfural (1.8 g/L) were detected in the hydrolysate by High Performance Liquid Chromatography (HPLC) analysis, with negligible amount of phenolic content (0.01 g/L) was observed. In comparison, the PW pre-treatment with combined LPSH and conventional sulphuric acid (H_2SO_4) produced 3.6 g/L of furfural and 0.4 g/L of 5-HMF, at similar optimized conditions. The pre-treated PW were further characterized by scanning electron microscopy and Fourier transform infrared spectroscopy to elucidate structural morphology and functional group changes. The optimized cascade pre-treatments can provide up to 54.79 % of glucose yield and 69.23 % of xylose yield. Furthermore, 67.87 % reduction of lignin content from cascade pre-treatment can substantially enhance the glucose yield up to 95.76 % and xylose yield up to 99.07 % during enzymatic hydrolysis using the mixture of cellulase and hemicellulose. This study shows that the cascade pre-treatment of LPSH and MA can decompose the lignin structure of the biomass with a negligible amount of inhibitors and enhance the effectiveness of enzymatic treatment.

ABSTRAK

Gula yang disintesis daripada biojisim lignoselulosa boleh digunakan untuk mengatasi keperluan permintaan yang semakin meningkat dalam pelbagai aplikasi di dalam industri. Proses prarawatan biojisim lignoselulosa adalah proses yang mencabar disebabkan keperluan untuk memecahkan struktur yang padat dan kukuh untuk menghasilkan gula. Dalam kajian ini, sisa nanas (PW) telah melalui proses kombinasi prarawatan secara turutan yang telah dioptimumkan iaitu (i) pemanasan stim bertekanan rendah (LSPH) dan asid maleik (MA), bertujuan untuk menambah baik pelunturan lignin, meningkatkan kebolehcapaian enzim ke selulosa dan hemiselulosa di dalam sisa nanas sekaligus mengurangkan pengeluaran perencat. Keadaan yang terbaik untuk tekanan (kPa), pemuatan pepejal (% w/v) dan masa (minit) untuk LPSH telah disaring. Kemudian, hasil biojisim pepejal yang dirawat dengan LPSH telah dilanjutkan ke prarawatan kedua iaitu prarawatan dengan MA dan proses ini dioptimumkan dengan kaedah permukaan tindakbalas berdasarkan reka bentuk Box-Behnken, di mana pengaruh suhu prarawatan, kepekatan asid dan masa telah dikaji. Sebanyak 68 % w/w delignifikasi dengan penyingkiran hemiselulosa yang tinggi (79.5 %) telah dicapai sementara 77.6 % selulosa dikekalkan dalam sisa pepejal PW, setelah melalui kedua-dua proses prarawatan. Tiada kandungan *5-hidroksimetil* furfural (5-HMF) dan sedikit kandungan furfural (1.8 g/L) yang dikesan dalam hidrolisat melalui analisis kromatografi cecair prestasi tinggi (HPLC), manakala jumlah kandungan fenolik (0.01 g/L) boleh diabaikan. Secara perbandingan, prarawatan PW dengan gabungan LPSH dan asid sulfurik konvensional (H_2SO_4) telah berjaya menghasilkan 3.6 g/L furfural dan 0.4 g/L 5-HMF pada keadaan optimum yang sama. PW yang sudah dirawat selanjutnya dicirikan oleh mikroskopi elektron imbasan dan spektroskopi inframerah transformasi Fourier untuk menentukan perubahan morfologi struktur dan kumpulan berfungsi. Kombinasi prarawatan secara turutan yang telah dioptimumkan boleh menghasilkan sehingga 54.79 % glukosa dan 69.23 % xilosa. Tambahan pula, pengurangan 67.87 % kandungan lignin daripada pra-rawatan ini boleh meningkatkan hasil glukosa dengan ketara sehingga 95.76 % dan hasil xilosa sehingga 99.07% semasa hidrolisis enzimatik yang menggunakan campuran selulosa dan hemiselulosa. Kajian ini menunjukkan bahawa prarawatan secara turutan ini boleh menguraikan

struktur lignin biojisim dengan jumlah perencat yang boleh diabaikan serta meningkatkan keberkesanan rawatan enzimatik.

.

TABLE OF CONTENT

	TITLE	PAGE
	DECLARATION	iii
	DEDICATION	iv
	ACKNOWLEDGEMENT	v
	ABSTRACT	vi
	ABSTRAK	vii
	TABLE OF CONTENT	viii
	LIST OF TABLES	xiii
	LIST OF FIGURES	xv
	LIST OF SYMBOLS	xvii
	LIST OF ABBREVIATION	xviii
	LIST OF APPENDICES	xix
CHAPTER 1	INTRODUCTION	1
	1.1 Research Background	1
	1.2 Problem Statement	4
	1.3 Objectives of the Study	6
	1.4 Scope of the Study	6
	1.5 Significance of the Study	8
CHAPTER 2	LITERATURE REVIEW	9
	2.1 Lignocellulosic Biomass for Fermentable Sugar Production	9
	2.1.1 Lignin	9
	2.1.2 Cellulose	11
	2.1.3 Hemicellulose	12
	2.2 Pineapple (<i>Ananas Comosus</i>) Waste	14
	2.3 Pre-treatment of Biomass	15
	2.3.1 Physical pre-treatment	15

2.3.2	Chemical pre-treatment	16
2.3.2.1	Acid Pre-treatment	17
2.3.2.1.1	Inorganic Acid Pre-treatment	18
2.3.2.1.2	Organic Acid Pre-treatment	20
2.3.2.2	Alkaline Pre-treatment	23
2.3.3	Physio-chemical Pre-treatment	24
2.3.4	Effects of Biomass Loading on Biochemical Conversion of Biomass	26
2.3.5	Effect of temperature on Biochemical Conversion of Biomass	26
2.3.6	Enzyme Hydrolysis	27
CHAPTER 3	MATERIAL AND METHOD	31
3.1	Research Methodology	31
3.2	Detailed Methodology	33
3.2.1	Preparation of Raw Materials	33
3.2.2	Compositional Analyses of Untreated and Treated Pineapple Waste	33
3.2.2.1	Moisture Content	33
3.2.2.2	Ash Content	34
3.2.2.3	Extractives Content	34
3.2.2.4	Acid Insoluble Lignin Content	35
3.2.2.5	Holocellulose Content	36
3.2.2.6	Cellulose and Hemicellulose Content	37
3.2.3	Hydrolysate Analyses	38
3.2.3.1	Total Phenolic Content Assay	38
3.2.3.2	5-HMF and Furfural content	39
3.2.3.3	Glucose Content	40
3.2.3.4	Xylose Content	41
3.2.4	Low Pressure Steam Heating Pre-treatment (1 st stage)	42
3.2.4.1	Screening of Biomass Loading	42

	3.2.4.2	Screening of Pressure Level	43
	3.2.4.3	Screening of Reaction Time	43
	3.2.5	Acid Pre-treatment (2 nd Stage)	43
	3.2.5.1	Screening of Type of Organic Acid used in the Pre-treatment and Solid Loading (%w/v)	
	3.2.5.2	Comparison of Selected Organic Acid with Conventional Sulphuric Acid Pre-treatment	44
	3.2.5.3	Optimization of Selected Organic Acid Pre-treatment using Statistical Analysis (RSM)	45
	3.2.6	Characterization Analysis	46
	3.2.6.1	Surface Morphology Analysis	46
	3.2.6.2	Functional Group Analysis	47
	3.2.7	Enzymatic Activity Determination	47
	3.2.7.1	Preparation of Dinitrosalicylic Acid (DNS) Reagent	47
	3.2.7.2	Cellulase Activity (Filter Paper Assay)	48
	3.2.7.3	Hemicellulase Activity Assay	50
	3.2.8	Enzyme Hydrolysis	52
	3.2.8.1	Screening of Biomass Solid Loading	52
	3.2.8.2	Optimization of Single Cellulase and Hemicellulase Hydrolysis	53
	3.2.8.3	Optimization of Enzyme Mixture (Cellulase and Hemicellulase Hydrolysis)	53
CHAPTER 4		RESULTS AND DISCUSSION	55
	4.1	Chemical Composition of Raw PW	55
	4.2	Optimization of Low-Pressure Steam Heating Pre-treatment (LPSH) (1 st stage)	56
	4.2.1	Effect of the Pre-treatment with Different Biomass Loadings	57
	4.2.2	Effect of the Pre-treatment with Different Pressure Level	59

4.2.3	Effect of the Pre-treatment with Different Reaction Time	62
4.2.4	Glucose and Xylose Yield from Optimized LPSH Pre-treatment Condition	65
4.3	Optimization of Acid Pre-treatment (2 nd stage)	67
4.3.1	Screening of Type of Organic Acid used in the Pre-treatment and Solid Loading (%w/v)	67
4.3.2	Comparison of Maleic Acid pre-treatment Hydrolysate with Conventional Sulphuric Acid Pre-treatment	69
4.3.3	Optimization of Maleic Acid Pre-treatment using Statistical Analysis (RSM)	71
4.4	Hydroxymethyl Furfural (5-HMF) and Furfural Analysis using HPLC after 1 st and 2 nd Pre-treatment	76
4.5	Compositional changes after 1 st and 2 nd Pre-treatment	77
4.6	Characterization Analysis	80
4.6.1	Surface Morphology Analysis	80
4.6.2	Functional Group Analysis	83
4.7	Enzyme Hydrolysis of Pre-treated PW	86
4.7.1	Screening of Biomass Solid Loading	86
4.7.2	Optimization of Single Cellulase and Hemicellulase Hydrolysis	88
4.7.3	Optimization of Enzyme Mixture (Cellulase and Hemicellulase Hydrolysis)	89
CHAPTER 5	CONCLUSIONS AND RECOMMENDATIONS	93
	REFERENCES	95
	APPENDICES	115
	LIST OF PUBLICATION	127

LIST OF TABLES

TABLE NO.	TITLE	PAGE
Table 2.1	Chemical compositions from different part of pineapple waste	13
Table 2.2	Chemical compositions of different organic waste materials	13
Table 2.3	Dilute sulphuric acid pre-treatments in various feedstock	19
Table 2.4	Summary of the organic acid pre-treatment without any catalyst in various feedstock	22
Table 2.5	Comparison of sugar yield and total inhibitors under different pre-treatments and enzymatic hydrolysis of different parts of PW waste used.	29
Table 3.1	Preparation of gallic acid standard curve for total polyphenolic content (TPC)	39
Table 3.2	Retention time of furfural and 5-HMF	40
Table 3.3	Glucose Assay	40
Table 3.4	D-xylose Assay	41
Table 3.5	Maximum temperature chart at given pressure level inside the pressure cooker	42
Table 3.6	Details of the low and upper limit for each parameter used in the BBD design	46
Table 3.7	Preparation of DNS reagent.	48
Table 3.8	Chemical and reagent preparation for cellulase assay	49
Table 3.9	Chemical and reagent preparation for xylanase assay	50

Table 3.10	Series of xylose dilution	51
Table 4.1	Chemical composition of whole PW in % total dry weight (w/w)	55
Table 4.2	Statistical comparison of lignin, cellulose and hemicellulose composition of certain biomass loading condition to other biomass loading (% w/v)	59
Table 4.3	Statistical comparison of lignin, cellulose and hemicellulose composition in certain pressure levels to other pressure level (kPa)	61
Table 4.4	The actual weight of chemical compositions (g) and the percentage of compositional removal (w/w%) of PW before and after LPSH pre-treatment	64
Table 4.5	Summary on the analysis of variance (ANOVA) for different response surface models.	75
Table 4.6	The percentage of composition removal (% w/w) of PW after 1 st and 2 nd stage pre-treatment	78
Table 4.7	The percentage of composition removal (% w/w) of PW after 1 st and 2 nd stage pre-treatment	78

LIST OF FIGURES

FIGURE NO.	TITLE	PAGE
Figure 2.1	Schematic framework of lignocellulose which containing lignin, cellulose and hemicellulose	9
Figure 2.2	Chemical structure of lignin	11
Figure 3.1	The conceptual framework of research methodology	32
Figure 4.1	Screening of biomass loading (%) of 1 st pre-treatment	58
Figure 4.2	Screening of pressure level (kPa) of 1 st pre-treatment	60
Figure 4.3	Screening of reaction time (min) of 1 st pre-treatment	63
Figure 4.4	Actual weight (g) of lignin, cellulose and hemicellulose before and after pre-treat with optimized LPSH	65
Figure 4.5	Glucose and xylose yield at different condition of LPSH	66
Figure 4.6	Glucose yield (mg/g) from different organic acid pre-treatment and solid loading	67
Figure 4.7	Xylose yield (mg/g) from different organic acid pre-treatment and solid loading	68
Figure 4.8	Glucose yield (g/L) measured after 2 nd pre-treatment	69
Figure 4.9	Xylose yield (g/L) measured after 2 nd pre-treatment	70
Figure 4.10	3D Response surface plot for overall desirability function with the actual factor	73

Figure 4.11	Actual weight (g) of lignin, cellulose and hemicellulose before and after pre-treated with optimized LPSH and organic acid	79
Figure 4.12	Significant FTIR spectra in the 4000 to 900 cm^{-1} region for PW samples under different conditions of PW	82
Figure 4.13	Scanning electron micrographs of morphological changes at 1000x magnification	84
Figure 4.14	Scanning electron micrographs of morphological changes at 5000x magnification	85
Figure 4.15	Glucose concentration at different solid loading	87
Figure 4.16	Glucose concentration at different solid loading	87
Figure 4.17	Glucose yields (g/L) of PW at varying hydrolysis time and cellulase activity	88
Figure 4.18	Xylose yields (g/L) of pre-treated PW at varying hydrolysis time and hemicelluase activity	89
Figure 4.19	Glucose and xylose yield of pre-treated PW at varying mixture ratio of cellulase and hemicellulase at 2 hr of hydrolysis time.	90
Figure 4.20	Summary of the optimized cascade pre-treatment of LPSH - MA and subsequent enzymatic hydrolysis on the pineapple waste	92

LIST OF SYMBOLS

%	-	Percentage
Δ	-	Absorbance
$^{\circ}\text{C}$	-	Degree Celcius
% w/w	-	Weight per weight
% v/w	-	Volume per weight
kPa	-	Pressure
U/g	-	Units per gram
U/mL	-	Units per millilitre
M	-	Molar
mM	-	Millimolar
min	-	Minutes
hr	-	Hour
mL	-	Millilitre
rpm	-	Revolution per min
μL	-	Microlitre
μm	-	Micrometre
g	-	Gram
μmole	-	Micromole
β	-	Beta
mg/L	-	Milligram per litre
mg/mL	-	Milligram per millilitre
g/L	-	Gram per litre
A	-	Acid Concentration
B	-	Temperature
C	-	Time
FPU	-	Filter Paper Units
U	-	Units
W	-	Watt

LIST OF ABBREVIATION

FFTC	-	Food and Fertilizer Technology Centre
FAO	-	Food and Agriculture Organization
LPSH	-	Low-Pressure Steam Heating
MA	-	Maleic Acid
AA	-	Acetic Acid
CA	-	Citric Acid
PW	-	Pineapple waste
AIL	-	Acid Insoluble Lignin
DF	-	Dilution Factor
OFAT	-	One -Factor-at-a-Time
RSM	-	Response Surface Methodology
ANOVA	-	Analysis of Variance
3D	-	3 Dimensional
BBD	-	Box-Behnken Design
FTIR	-	Fourier Transform Infrared Spectroscopy
SEM	-	Scanning Electron Microscope
5-HMF	-	5-Hydroxymethyl Furfural
SE	-	Steam Explosion
pH	-	Potential of hydrogen
m	-	slope
DNS	-	3,5-dinitrosalicylic acid
UV-Vis	-	Ultraviolet-visible
ND	-	Not determined

LIST OF APPENDICES

	TITLE	PAGE
APPENDIX A	Determination of cellulase and xylanase activity	115
APPENDIX B	Glucose and xylose standard curve for enzymatic hydrolysis.	119
APPENDIX C	Analysis of variance (ANOVA) for different response surface	120
APPENDIX D	Standard curves for HMF and furfural from HPLC analysis and gallic acid standard curves for total phenolic content assay	125

CHAPTER 1

INTRODUCTION

1.1 Research Background

According to the Food and Agriculture Organization (FAO, 2021), Malaysia is one of the most important producers of canned pineapples in Asian countries in 2017 and recorded a staggering 299912 tonnes of pineapple crops produced in 2019. Pineapple (*Ananas Comosus*) also ranks the 4th among the fruit crops exported from Malaysia after watermelon, banana and papaya (FFTC, 2019), which plays an important role for the country's socio-economic development either domestic or export market.

PW produced from industrial processing and cultivation comprises of crown, peel and core of the fruit and about 30 – 50% of the total fruit weight is discarded after juice extraction. There are practices for sustainable development of pineapple in Malaysia have been documented in publications such as application for downstream value-added utilization of biomass and waste for further industrial processes like fermentation of sugar (Clauser et al., 2021). The high carbohydrate content in the pineapple waste (PW) makes it a good source for fermentable sugars supply. Extremely significant applications of sugars make it worth to synthesize sugars from biomass-derived resources for the application in fine chemicals (Jarosz, 2015), in replacing petroleum-derived chemicals that used in pharmaceuticals and cosmetics industries (Bhaumik & Dhepe, 2015; Wang et al., 2019) and also to the agriculture industries to increase the crop quality and yield (Dotaniya et al., 2016). The highly efficient conversion of lignocellulose to fermentable sugars usually comprises of two major series of biochemical reactions which is pre-treatment and enzyme hydrolysis (Choi et al., 2019). However, the main contributors to the recalcitrance of biomass such as the cellulose crystallinity, the structural heterogeneity, and high lignin content must be overcome to reduce the resistance of the lignocellulosic biomass from sugar

hydrolysis (Baruah et al., 2018). To reduce biomass recalcitrance and increase enzyme accessibility to cellulose, pre-treatment that disrupts the biomass cell walls is necessary.

The first series of biochemical reaction which is the pre-treatment is typically required to break down the protective sheath surrounding the lignocellulose so that it becomes suitable to enzymatic hydrolysis which further enhance the accessibility to cellulose and hemicellulose. Numbers of pre-treatment methods have been used by several researchers and most of them are not environmental friendly due to large amount of chemicals used during the process (Baruah et al., 2018; Kumari & Singh, 2018). Consequently, cascade processes that combine acid, alkaline, hydrothermal and/or other pre-treatments have been suggested to maximize sugar recovery and give higher efficiency in lignocellulose digestibility compared to one-step pre-treatment (Moodley & Kana, 2017; Sun et al., 2016; Tang et al., 2020). Guo et al. (2013b) observed a significantly higher sugar recovery (glucose (>80 %) and xylose (>70 %)) from dried *Miscanthus* obtained and by-products formation were remarkably reduced after cascading the low severity acid with alkaline pre-treatment compared to single stage pre-treatment. This probably because of the different structural features of the biomass wastes (Tang et al., 2020). The level of glucose in the enzymatic hydrolysate pre-treated cornstalk with two-step hydrothermal processes, dilute acid pre-treatment and oxidated alkali pre-treatment at such mild hydrolysis were reached 57.0% , the internal pore volume and porosity also enhanced which created more active accessibilities to enzymes in the residues compared to the single pre-treatment process (Xia et al., 2021).

Method using physiochemical pre-treatment alone such as steam explosion (Zhiqiang et al., 2015), ammonia fiber explosion (Lee et al., 2010), liquid hot water (LHW) (Zhuang et al., 2016), and superheat steam (SS) (Barchyn & Cenkowski, 2014) have been suggested to minimize formation of inhibitory compounds, reduced energy usage without the use of any chemical catalyst, and result in increasing fermentative sugar. Due to the combination of heat and pressure, it prompts the substrate for further breakdown as it is possible to open the bundles of lignocelluloses and make enzymatic reactions easily accessible to the polymer chain of cellulose and hemicellulose (Baruah

et al., 2018). Steam pre-treatment without the explosion by using equipment like pressurized vessel (pressure cooker) can be effective in cellulose accessibility as steam with explosion. However to give the essentially the same yields of glucose and total reducing sugars, the treatment must be with saturated steam at high temperature (>240 °C) (Brownell & Saddler, 1987). Due to the cellulose and hemicellulose structure having different thermal stabilities and endurance, enough pressure should be applied to keep the water in liquid state within the appropriate time, so that the lignin will be degraded the most while the hemicellulose and cellulose remain intact (Cao et al., 2012b). High temperature (220-260°C) are too harsh for the pre-treatment of biomass as low glucose yield was obtained from the supernatant despite the large amount of cellulose and hemicellulose being solubilized and eventually degraded (Weil et al., 1998).

Recently, increasing attention has been paid to acid pre-treatment with respect to industrial implementation, because of their efficiency in not only maximisation of the sugar yields, but also the minimisation of the production costs (Bensah & Mensah, 2013; Carregari Polachini, 2019). However, it gives some drawbacks such as equipment corrosion, gypsum formation during neutralization, sugar degradation due to the high reactivity and formation of inhibitory by-products (Kumar & Sharma, 2017). Compared to alkaline pre-treatment, acid pre-treatment released higher amount of sugar (Baadhe, 2014). Organic acids such as maleic acid, citric acid, formic acid, oxalic acid, acetic etc. which are defined as organosolv system were often applied at higher concentration and their ability has been proven to replace the inorganic acid. It is less toxic and more efficient in carrying out the hydrolysis over a range of pHs and temperatures (Lee & Jeffries, 2011), resulting in minimal xylose and glucose loss because of its superior selectivity to be able to mimic the structure of the specific active sites of enzyme which help to reduce the degradation of glucose (Guo et al., 2013a; Lu & Mosier, 2008; Mosier et al., 2002b) and produce lesser amount of inhibitors (Deng et al., 2016).

Lastly, the second series of biochemical reaction is enzymatic hydrolysis which totally depends on the biocatalysts capability to convert biopolymers into monomers (Wu et al., 2021). Majority of the results obtained from different studies have proved

that the combinations of cellulase and hemicellulase enzymes for the hydrolysis of pre-treated lignocellulosic biomass successfully produced higher reducing sugar yield compared to single cellulase or hemicellulase hydrolysis (Amani et al., 2018; Conesa et al., 2016). These two types of enzymes need to work in synergy at optimized hydrolysis conditions to effectively break apart the complex biomass structure and releasing high sugar yield.

1.2 Problem Statement

Many research have been conducted to increase the effectiveness of pre-treatment methods for maximal fermentable sugars production from the lignocellulosic biomass due to the recalcitrant structure. However, efficiency of the single pre-treatment process was often not satisfactory compared to cascade pre-treatment (Kumar & Sharma, 2017).

Pre-treatment with pressure cooker consumed less energy (< 143.27 kPa and $< 110^{\circ}\text{C}$ compared to steam explosion (SE) ($170^{\circ}\text{--}210^{\circ}\text{C}$) and autoclaving method (121°C) (Roda et al., 2016b). SE and autoclaving method usually use pressure above 15 psi and that will increase glucose significantly during the enzyme hydrolysis. Unfortunately, xylose were decreased if the pressure used is too high which likely due to solubilization of hemicellulose during the hot water phase (Barchyn & Cenkowski, 2014). Amorphous hemicellulose is known as thermosensitive hemicellulose, where it is sensitive to extreme temperatures ($> 150^{\circ}\text{C}$) and longer residence times, easily degrade 5-carbon xylose sugar into their inhibitory by-products compared to cellulose and lignin (Shaoni Sun et al., 2014) and the hydrolysate need extra detoxification steps after the hydrothermal pre-treatment. The use of water, heat and vapor leads to hemicellulose degradation and lignin transformation owing to the high temperatures approximately $110\text{--}300^{\circ}\text{C}$. During biomass hydrolysis, hot water cleaves hemiacetal links, releasing acids and facilitating the breaking of ether bonds in biomass (Maneeintr et al., 2018). Higher severity conditions caused accumulation of acid insoluble material that led to recondensation reaction of Klason lignin causing high

molecular weight lignin (Kim et al., 2014). These might inhibit the accessibility of cellulose to the enzymatic hydrolysis. In this study, due to the low lignin content of the PW, low pressure steam heating of pressure cooker were used for the 1st stage pre-treatment and optimized with the aim to achieve high delignification but still able to retain high hemicellulose and cellulose content by maintaining the liquid phase (under pressure). The valve regulates the pressure inside the pressure cooker to a pre-set level: typically ~10 psi above atmospheric pressure were able to boil water up to 114°C.

There are studies that investigate the effectiveness of organic acids such as maleic acid, citric acid, acetic acid, oxalic acid and etc. as alternative to the conventional sulphuric acid in order to delignify the lignocellulosic biomass, enhance cellulose and hemicellulose digestibility while reducing the amount of toxic by-products such as furfural and 5-HMF (Cao et al., 2012a; Kootstra et al., 2009a; Sahu & Pramanik, 2018b). By using harsh chemicals from the conventional methods, it required extra detoxification steps due to the production of high total furans (>4.5 g/L) and phenolic content which can also contributed to some sugar losses (Khedkar et al., 2018b). Due to the slightly expensive cost of organic acid pre-treatment compared to inorganic acid, it is suggested to cascade the LSPH with organic acid pre-treatment to reduce the cost.

Hence, this project is focusing on assessing the efficiency of cascade pre-treatment of low-pressure steam heating (LPSH) using simple and economical-value (Roda et al., 2016a) equipment such as commercial pressure cooker (LPSH) and organic acid as means of pre-treating a mixture of whole pineapple wastes parts. The operating conditions of LPSH combined with several organics acids pre-treatment were optimized and characterized together with further optimization of enzymatic hydrolysis conditions for efficient saccharification.

1.3 Objectives of the Study

In general, this work is aimed to improve the pre-treatment process's performance for the conversion of pineapple waste (PW) into fermentable sugars by introducing the cascade of low-pressure steam heating (LSPH) and organic acid pre-treatment. It can be further sub-divided into the following specific objectives:

- i. To optimize the LSPH pre-treatment parameters on pineapple waste with the highest lignin removal while retaining the most hemicellulose and cellulose.
- ii. To optimize the 2nd stage organic acid pre-treatment parameters of the LPSH-pre-treated pineapple waste for optimum hemicellulose degradation with negligible yield of inhibitors.
- iii. To determine the best enzyme mixture ratio for optimum sugar yields and evaluate physical and chemical characterization of the cascade pre-treated PW.

1.4 Scope of the Study

The following are the scopes of the study in order to achieve the objectives:

- a) The biomass used in this study consist of the mixed part of pineapple waste such as their leaves, peels, crown, core, and other non-fruit parts with a fixed particle size at 500 μm .
- b) The parameters chosen for the optimization of the 1st stage pre-treatment (LPSH) are:
 - i) Biomass loading (0.5, 1.5, 2.5, 3.5, and 5 % w/v)
 - ii) Pressure (40,50, 60, 70 and 80 kPa)
 - iii) Time (15, 30, 45, 60 and 75 min)

- c) For the 2nd stage pre-treatment, selection of the best organic acids (maleic acid, citric acid, and acetic acid) and solid loadings (% v/w) were carried out based on one-factor-at-a-time 'OFAT' analysis based on the highest xylose and glucose yields released. The other three pre-treatment parameters (acid concentration, treatment time and reaction temperature) were statistically optimized using 3-level-Box-Behnken design (BBD) matrix in the Response Surface Methodology (RSM), Design Expert Software (version 11.0.) (State-Ease, USA). The ranges of the three parameters are shown below:
- i) Acid concentration (0.1-1.0 M)
 - ii) Treatment time (30-120 min)
 - iii) Reaction temperature (50-180°C)
- d) The yield of 5-Hydroxymethyl furfural (g/L) and furfural (g/L) inhibitors were analyzed using High Performance Liquid Chromatography (HPLC), while phenolic content (g/L) were analyzed using total phenolic content assay.
- e) The efficiency of the pre-treatments were characterized by using Scanning Electron Microscope (SEM) for morphological analysis, Fourier Transform Infrared Spectroscopy (FTIR) for the functional group analysis, Glucose and Xylose Assay Kits for the determination of xylose and glucose yield and DNS method for reducing sugar yield.
- f) Screening and optimization of enzyme mixture (cellulase and hemicellulose) hydrolysis of the pre-treated PW were carried out using OFAT with the target of achieving maximum sugar yield. The studied parameters were:
- i) Solid loading (5%, 10%, 15% w/v)
 - ii) Time (2, 4, 6, 8, 24 hr)
 - iii) Single enzyme loading of cellulase (2, 2.5, 3, 4 FPU/mL) and hemicellulose (4, 12, 20, 40, 60, 80 U/mL)
 - iv) Enzyme mixture ratio (1:1, 1:2, 2:1, 3:1, 1:3)

1.5 Significance of the Study

Overall, this study could significantly provide explanation on ecological alternative to reduce fruit processing waste from being directly discarded to the environment. Since detail investigation on the relationship between parameters and aspect ratio will be done in this research, it will provide good mechanism on the technique on producing high yield of reducing sugar with the optimum reaction during hydrolysis of sugar. The process sequence that will be developed from this study can be potentially adapted onto larger scale for further study. The pineapple biorefinery can be used as a source for edible sugar production which can be employed by the nutraceutical and functional food industries and thus may give economic benefit to the whole community.

REFERENCES

- Abd Hamid. (2015). *Combination of low pressure steam heating and dilute acid pretreatment of palm biomass for fermentable sugar production*. Retrieved from <http://eprints.utm.my/id/eprint/53962/>
- Adeboye, Bettiga, Aldaeus, Larsson, & Olsson. (2015). Catabolism of coniferyl aldehyde, ferulic acid and p-coumaric acid by *Saccharomyces cerevisiae* yields less toxic products. *Microbial cell factories*, 14(1), 149.
- Adney, Baker, & Laboratory. (2008). Measurement of Cellulase Activities: Laboratory Analytical Procedure (LAP) : Issue Date, 08/12/1996. In: National Renewable Energy Laboratory.
- Adrizar, Amizar, & Mahata. (2017). Evaluation of pineapple (*Ananas comosus* [L.] Merr) waste fermented using different local microorganisms solutions as poultry feeds. *Pakistan journal of nutrition*, 16(2), 84-89.
- Agarwal, J. Y. Zhu, & Ralph. (2011, 8-10 June 2011). *Enzymatic Hydrolysis of biomass: Effects of crytallinity, particle size, and lignin removal*. Paper presented at the International symposium on wood, fiber and pulping chemistry, Tianjin, China. Beijing, China.
- Ahmed, Aboudi, Tyagi, Álvarez-Gallego, Fernández-Güelfo, Romero-García, & Kazmi. (2019). Improvement of Anaerobic Digestion of Lignocellulosic Biomass by Hydrothermal Pretreatment. *Applied Sciences*, 9(18), 3853.
- Allard-Massicotte, Chadjaa, & Marinova. (2017). Phenols removal from hemicelluloses pre-hydrolysate by laccase to improve butanol production. *Fermentation*, 3(3), 31. doi: <https://doi.org/10.3390/fermentation3030031>
- Alonso, Wettstein, & Dumesic. (2012). Bimetallic catalysts for upgrading of biomass to fuels and chemicals. *Chemical Society Reviews*, 41(24), 8075-8098.
- Amani, Toh, Tan, & Lee. (2018). The efficiency of using oil palm frond hydrolysate from enzymatic hydrolysis in bioethanol production. *Waste Biomass Valori*, 9(4), 539-548. doi:<https://doi.org/10.1016/j.jclepro.2013.10.007>
- Amnuaycheewa, Hengaroonprasan, Rattanaporn, Kirdponpattara, Cheenkachorn, & Sriariyanun. (2016). Enhancing enzymatic hydrolysis and biogas production from rice straw by pretreatment with organic acids. *Industrial Crops and Products*, 87, 247-254.

- Anoop. (2014). *Utilization of pineapple (Ananas comosus (L) Merr.) biomass for biofuel production*. College of Agriculture, Vellayani,
- Antunes, dos Santos, da Cunha, Brumano, dos Santos Milessi, Terán-Hilares, Peres, Medina, da Silva, & Dalli. (2017). Biotechnological production of xylitol from biomass. In *Production of Platform Chemicals from Sustainable Resources* (pp. 311-342): Springer.
- Ariffin, Masngut, Seman, Saufi, Jamek, & Sueb. (2020). *Dilute acid hydrolysis pretreatment for sugar and organic acid production from pineapple residues*. Paper presented at the IOP Conference Series: Mater Sci Eng.
- Auxenfans, Crônier, Chabbert, & Paës. (2017). Understanding the structural and chemical changes of plant biomass following steam explosion pretreatment. *Biotechnology for biofuels*, 10(1), 36.
- Azelee, Adnan, Manas, Dailin, Ramli, & Illias. (2019). *Assessment of microwave-assisted pretreatments for enhancing pineapple waste delignification*. Paper presented at the AIP Conference Proceedings.
- Azelee, Jahim, Rabu, Murad, Bakar, & Illias. (2014). Efficient removal of lignin with the maintenance of hemicellulose from kenaf by two-stage pretreatment process. *Carbohydrate polymers*, 99, 447-453.
- Baadhe. (2014). Influence of dilute acid and alkali pretreatment on reducing sugar production from corncobs by crude enzymatic method: A comparative study. *Bioresource Technology*, 162, 213–217. doi:10.1016/j.biortech.2014.03.117
- Ballesteros, Negro, Oliva, Cabañas, Manzanares, & Ballesteros. (2006). *Ethanol production from steam-explosion pretreated wheat straw*. Paper presented at the Twenty-seventh symposium on biotechnology for fuels and chemicals.
- Banoth, Sunkar, Tondamanati, & Bhukya. (2017). Improved physicochemical pretreatment and enzymatic hydrolysis of rice straw for bioethanol production by yeast fermentation. *3 Biotech*, 7(5), 1-11. doi: <https://doi.org/10.1007/s13205-017-0980-6>
- Baral, & Shah. (2014). Microbial inhibitors: formation and effects on acetone-butanol-ethanol fermentation of lignocellulosic biomass. *Applied microbiology and biotechnology*, 98(22), 9151-9172. doi:<https://doi.org/10.1007/s00253-014-6106-8>

- Barchyn, & Cenkowski. (2014). Process analysis of superheated steam pre-treatment of wheat straw and its relative effect on ethanol selling price. *Biofuel Research Journal*, 1(4), 123-128.
- Barisik, Isci, Kutlu, Bagder Elmaci, & Akay. (2016). Optimization of organic acid pretreatment of wheat straw. *Biotechnology progress*, 32(6), 1487-1493.
- Baruah, Nath, Sharma, Kumar, Deka, Baruah, & Kalita. (2018). Recent trends in the pretreatment of lignocellulosic biomass for value-added products. *Frontiers in Energy Research*, 6, 141.
- Basu. (2018). *Biomass gasification, pyrolysis and torrefaction: practical design and theory*: Academic press.
- Behera, Meena, Chakraborty, & Meikap. (2018). Application of response surface methodology (RSM) for optimization of leaching parameters for ash reduction from low-grade coal. *Int J Min Sci Technol*, 28(4), 621-629. doi:<https://doi.org/10.1016/j.ijmst.2018.04.014>
- Bensah, & Mensah. (2013). Chemical pretreatment methods for the production of cellulosic ethanol: technologies and innovations. *International Journal of Chemical Engineering*, 2013.
- Bhaumik, & Dhepe. (2015). Conversion of biomass into sugars. *Royal Society of Chemistry*.
- Borand, & Karaosmanoğlu. (2018). Effects of organosolv pretreatment conditions for lignocellulosic biomass in biorefinery applications: a review. *Journal of Renewable and Sustainable Energy*, 10(3), 033104.
- Brownell, & Saddler. (1987). Steam pretreatment of lignocellulosic material for enhanced enzymatic hydrolysis. *Biotechnology and bioengineering*, 29(2), 228-235.
- Bugg, Ahmad, Hardiman, & Rahmanpour. (2011). Pathways for degradation of lignin in bacteria and fungi. *Natural product reports*, 28(12), 1883-1896.
- Cao, Sun, Liu, Yin, & Wu. (2012a). Comparison of the effects of five pretreatment methods on enhancing the enzymatic digestibility and ethanol production from sweet sorghum bagasse. *Bioresour Technol*, 111, 215-221. doi:<https://doi.org/10.1016/j.biortech.2012.02.034>
- Cao, Sun, Liu, Yin, & Wu. (2012b). Comparison of the effects of five pretreatment methods on enhancing the enzymatic digestibility and ethanol production from sweet sorghum bagasse. *Bioresource Technology*, 111, 215-221.

- Carregari Polachini. (2019). *Evaluation of the acid pretreatment of agroindustrial waste assisted by high-intensity ultrasound for further bioethanol production*. (Doctoral), Universitat Politècnica de València.
- Cassellis, Pardo, Lopez, & Escobedo. (2014). Structural, physicochemical and functional properties of industrial residues of pineapple (*Ananas comosus*). *Cellulose Chemistry and Technology*, 633-641.
- Castañón-Rodríguez, Welti-Chanes, Palacios, Torrestiana-Sanchez, Ramírez de León, Velázquez, & Aguilar-Uscanga. (2015). Influence of high pressure processing and alkaline treatment on sugarcane bagasse hydrolysis. *CyTA-Journal of Food*, 13(4), 613-620.
- Chambon, Verdía, Fennell, & Hallett. (2021). Process intensification of the ionoSolv pretreatment: effects of biomass loading, particle size and scale-up from 10 mL to 1 L. *Scientific reports*, 11(1), 1-15.
- Chen, Gong, Liu, Liu, Eggert, Guo, Zhao, Zhao, & Zhao. (2018). Postharvest Ultrasound-Assisted Freeze-Thaw Pretreatment Improves the Drying Efficiency, Physicochemical Properties, and Macamide Biosynthesis of Maca (*Lepidium meyenii*). *Journal of food science*, 83(4), 966-974.
- Chen, & Liu. (2015). Steam explosion and its combinatorial pretreatment refining technology of plant biomass to bio-based products. *Biotechnol J*, 10(6), 866-885. doi:<https://doi.org/10.1002/biot.201400705>
- Chen, & Liu. (2017). Enzymatic hydrolysis of lignocellulosic biomass from low to high solids loading. *Engineering in Life Sciences*, 17(5), 489-499.
- Chen, Yan, Liang, Ran, Wu, Wang, Zou, Zhao, Fang, & Shen. (2020). Comparative Evaluation of Organic Acid Pretreatment of Eucalyptus for Kraft Dissolving Pulp Production. *Materials*, 13(2), 361. doi:<https://doi.org/10.3390/ma13020361>
- Choi, Jang, Kim, Park, Kim, Jeong, Kim, & Choi. (2019). Simultaneous production of glucose, furfural, and ethanol organosolv lignin for total utilization of high recalcitrant biomass by organosolv pretreatment. *Renewable energy*, 130, 952-960.
- Chongkhong, & Tongurai. (2019). Optimization of soluble sugar production from pineapple peel by microwave-assisted water pretreatment. *Songklanakarin Journal of Science & Technology*, 41(1).

- Choonut, Saejong, & Sangkharak. (2014). The production of ethanol and hydrogen from pineapple peel by *Saccharomyces cerevisiae* and *Enterobacter aerogenes*. *Energy Procedia*, 52, 242-249. doi:<https://doi.org/10.1016/j.egypro.2014.07.075>
- Clauser, González, Mendieta, Kruyeniski, Area, & Vallejos. (2021). Biomass waste as sustainable raw material for energy and fuels. *Sustainability*, 13(2), 794.
- Conesa, Seguí, Laguarda-Miró, & Fito. (2016). Microwave-assisted alkali pretreatment for enhancing pineapple waste saccharification. *BioResources*, 11(3), 6518-6531. doi:<https://doi.org/10.15376/biores.11.3.6518-6531>
- Cruz, Scullin, Mu, Cheng, Stavila, Varanasi, Xu, Mentel, Chuang, & Simmons. (2013). Impact of high biomass loading on ionic liquid pretreatment. *Biotechnology for Biofuels*, 6(1), 1-10.
- Daud, Hatta, Kassim, & Aripin. (2014). Analysis of the chemical compositions and fiber morphology of pineapple (*Ananas comosus*) leaves in Malaysia. *Journal of Applied Sciences*, 14(12), 1355-1358.
- del Pilar Castillo, Ander, & Stenstrom. (1997). Lignin and manganese peroxidase activity in extracts from straw solid substrate fermentations. *Biotechnology Techniques*, 11(9), 701-706.
- Delbecq, Wang, Muralidhara, El Ouardi, Marlair, & Len. (2018). Hydrolysis of hemicellulose and derivatives—A review of recent advances in the production of furfural. *Frontiers in Chemistry*, 6, 146.
- Deng, Ren, Wang, Li, Lin, Yan, Sun, & Liu. (2016). Production of xylo-sugars from corncob by oxalic acid-assisted ball milling and microwave-induced hydrothermal treatments. *Industrial Crops and Products*, 79, 137-145.
- Deshavath, Mohan, Veeranki, Goud, Pinnamaneni, & Benarjee. (2017). Dilute acid pretreatment of sorghum biomass to maximize the hemicellulose hydrolysis with minimized levels of fermentative inhibitors for bioethanol production. *3 Biotech*, 7(2), 1-12.
- Donohoe, Decker, Tucker, Himmel, & Vinzant. (2008). Visualizing lignin coalescence and migration through maize cell walls following thermochemical pretreatment. *Biotechnol Bioeng*, 101(5), 913-925. doi:<https://doi.org/10.1002/bit.21959>
- Dotaniya, Datta, Biswas, Dotaniya, Meena, Rajendiran, Regar, & Lata. (2016). Use of sugarcane industrial by-products for improving sugarcane productivity and soil

- health. *International Journal of Recycling of Organic Waste in Agriculture*, 5(3), 185-194.
- El Fels, Zamama, & Hafidi. (2015). Advantages and limitations of using FTIR spectroscopy for assessing the maturity of sewage sludge and olive oil waste co-composts. In *Biodegradation and Bioremediation of Polluted Systems-New Advances and Technologies*: IntechOpen.
- Fan, Santomauro, Budarin, Whiffin, Abeln, Chantasuban, Gore-Lloyd, Henk, Scott, & Clark. (2018). The additive free microwave hydrolysis of lignocellulosic biomass for fermentation to high value products. *Journal of cleaner production*, 198, 776-784.
- FAO. (2021). Pineapples Fresh Production. *Food and Agriculture Organization of the United Nations*. Retrieved from <http://www.fao.org/faostat/en/#data/QC>
- Feng, Qin, Liu, Dong, Li, & Yuan. (2014). Combined severity during pretreatment chemical and temperature on the saccharification of wheat straw using acids and alkalis of differing strength. *BioResources*, 9(1), 24-38.
- Ferreira, & Taherzadeh. (2020). Improving the economy of lignocellulose-based biorefineries with organosolv pretreatment. *Bioresour Technol*, 299, 122695. doi:<https://doi.org/10.1016/j.biortech.2019.122695>
- FFTC. (2019). Trends in production trade and consumption of tropical food in malaysia. Retrieved from <https://ap.fftc.org.tw/article/1381>
- Garrido, Reckamp, & Satrio. (2017). Effects of pretreatments on yields, selectivity and properties of products from pyrolysis of *Phragmites australis* (common reeds). *Environments*, 4(4), 96.
- Gellerstedt, & Northey. (1989). Analysis of birch wood lignin by oxidative degradation. *Wood science and technology*, 23(1), 75-83.
- Giachetti, & Hardyniec. (2020). Characterization of the release of heated and pressurized water from a pressure cooker. *Burns*.
- Gonzales, Sivagurunathan, & Kim. (2016). Effect of severity on dilute acid pretreatment of lignocellulosic biomass and the following hydrogen fermentation. *International Journal of Hydrogen Energy*, 41(46), 21678-21684.
- Guo, Zhang, Yu, Lee, Jin, & Morgenroth. (2013a). Two-stage acidic–alkaline hydrothermal pretreatment of lignocellulose for the high recovery of cellulose

- and hemicellulose sugars. *Applied biochemistry and biotechnology*, 169(4), 1069-1087.
- Guo, Zhang, Yu, Lee, Jin, & Morgenroth. (2013b). Two-stage acidic–alkaline hydrothermal pretreatment of lignocellulose for the high recovery of cellulose and hemicellulose sugars. *Applied biochemistry and biotechnology*, 169(4), 1069-1087. doi:<https://doi.org/10.1007/s12010-012-0038-5>
- Hajar, Zainal, Nadzirah, Roha, Atikah, & Elida. (2012). Physicochemical properties analysis of three indexes pineapple (*Ananas comosus*) peel extract variety N36. *APCBEE Procedia*, 4, 115-121.
- Hájková, Bouček, Procházka, Kalous, & Budský. (2021). Nitrate-Alkaline Pulp From Non-Wood Plants. *Materials (Basel)*.
- Harmsen, Huijgen, Bermudez, & Bakker. (2010). *Literature review of physical and chemical pretreatment processes for lignocellulosic biomass* (9085857570). Retrieved from
- He, & Chai. (2016). An efficient method for determining the α -, β -, and γ -cellulose content in fully delignified pulps by reaction-based headspace gas chromatography. *Journal of Wood Chemistry and Technology*, 36(6), 412-417.
- Hendriks, & Zeeman. (2009). Pretreatments to enhance the digestibility of lignocellulosic biomass. *Bioresource technology*, 100(1), 10-18.
- Hodge, Karim, Schell, & McMillan. (2008). Soluble and insoluble solids contributions to high-solids enzymatic hydrolysis of lignocellulose. *Bioresour Technol*, 99(18), 8940-8948. doi:<https://doi.org/10.1016/j.biortech.2008.05.015>
- Ilnicka, & Lukaszewicz. (2015). Discussion remarks on the role of wood and chitin constituents during carbonization. *Frontiers in Materials*, 2, 20.
- Ioelovich. (2015). Methods for determination of chemical composition of plant biomass. *Scientific Israel Technological Advantages*, 17(4), 208-214.
- Ioelovich, & Morag. (2012). Study of enzymatic hydrolysis of pretreated biomass at increased solids loading. *BioResources*, 7(4), 4672-4682.
- Jarosz. (2015). Fine Chemicals with High Added Value from Sucrose: Synthesis of Macrocyclic Receptors from “Normal” Sugar. *Journal of Carbohydrate Chemistry*, 34(7), 365-387.
- Jawaid, Tahir, & Saba. (2017). *Lignocellulosic fibre and biomass-based composite materials: processing, properties and applications*: Woodhead Publishing.

- Jędrzejczyk, Soszka, Czapnik, Ruppert, & Grams. (2019). Physical and chemical pretreatment of lignocellulosic biomass. In *Second and Third Generation of Feedstocks* (pp. 143-196): Elsevier.
- Jiang, Chang, Li, Oleskowicz-Popiel, & Xu. (2015). Liquid hot water pretreatment on different parts of cotton stalk to facilitate ethanol production. *Bioresource technology*, *176*, 175-180.
- Jin, Hu, Wu, Song, Yue, & Xiang. (2019). Promoting the material properties of xylan-type hemicelluloses from the extraction step. *Carbohydrate polymers*, *215*, 235-245.
- Jin, Zhang, Zhang, Li, Wang, Fan, & Zhou. (2016). Microwave assisted alkaline pretreatment to enhance enzymatic saccharification of catalpa sawdust. *Bioresource technology*, *221*, 26-30.
- Jönsson, & Martín. (2016). Pretreatment of lignocellulose: formation of inhibitory by-products and strategies for minimizing their effects. *Bioresource technology*, *199*, 103-112.
- Kalogiannis, Matsakas, Aspden, Lappas, Rova, & Christakopoulos. (2018). Acid assisted organosolv delignification of beechwood and pulp conversion towards high concentrated cellulosic ethanol via high gravity enzymatic hydrolysis and fermentation. *Molecules*, *23*(7), 1647.
- Kamarludin, Safaai, Azizan, Madzaki, Mamat, Zulkifli, & Zainuddin. (2014). Effect of Mechanical Grinding and Ionic Liquid Pre-treatment on Oil Palm Frond. *Malaysian Journal of Analytical Sciences*, *18*(3), 737-742.
- Karimi, Shafiei, & Kumar. (2013). Progress in physical and chemical pretreatment of lignocellulosic biomass. In *Biofuel technologies* (pp. 53-96): Springer.
- Khalil, Alwani, & Omar. (2006). Chemical composition, anatomy, lignin distribution, and cell wall structure of Malaysian plant waste fibers. *BioResources*, *1*(2), 220-232. doi:<https://doi.org/10.15376/biores.1.2.220-232>
- Khedkar, Nimbalkar, Kamble, Gaikwad, Chavan, & Bankar. (2018a). Process intensification strategies for enhanced holocellulose solubilization: Beneficiation of pineapple peel waste for cleaner butanol production. *J Clean Prod*, *199*, 937-947. doi:<https://doi.org/10.1016/j.jclepro.2018.07.205>
- Khedkar, Nimbalkar, Kamble, Gaikwad, Chavan, & Bankar. (2018b). Process intensification strategies for enhanced holocellulose solubilization:

- Beneficiation of pineapple peel waste for cleaner butanol production. *Journal of Cleaner Production*, 199, 937-947.
- Kim. (2018). Physico-chemical conversion of lignocellulose: Inhibitor effects and detoxification strategies: A mini review. *Molecules*, 23(2), 309.
- Kim, Dien, & Singh. (2016). Promise of combined hydrothermal/chemical and mechanical refining for pretreatment of woody and herbaceous biomass. *Biotechnology for biofuels*, 9(1), 1-15.
- Kim, Hwang, Oh, Kim, Kim, & Choi. (2014). Investigation of structural modification and thermal characteristics of lignin after heat treatment. *International journal of biological macromolecules*, 66, 57-65.
- Kim, Liu, Abu-Omar, & Mosier. (2012). Selective conversion of biomass hemicellulose to furfural using maleic acid with microwave heating. *Energy & fuels*, 26(2), 1298-1304.
- Kim, Park, Song, Wee, & Jeong. (2013). Effect of fermentation inhibitors in the presence and absence of activated charcoal on the growth of *Saccharomyces cerevisiae*. *Bioprocess Biosyst Eng*, 36(6), 659-666. doi:<https://doi.org/10.1007/s00449-013-0888-4>
- Kootstra, Beftink, Scott, & Sanders. (2009a). Comparison of dilute mineral and organic acid pretreatment for enzymatic hydrolysis of wheat straw. *Biochem Eng J*, 46(2), 126-131. doi:<https://doi.org/10.1016/j.bej.2009.04.020>
- Kootstra, Beftink, Scott, & Sanders. (2009b). Optimization of the dilute maleic acid pretreatment of wheat straw. *Biotechnol Biofuels*, 2(1), 1-14. doi:<https://doi.org/10.1186/1754-6834-2-31>
- Kumar, Dheeran, Singh, Mishra, & Adhikari. (2015). Bioprocessing of bagasse hydrolysate for ethanol and xylitol production using thermotolerant yeast. *Bioprocess and biosystems engineering*, 38(1), 39-47.
- Kumar, & Sharma. (2017). Recent updates on different methods of pretreatment of lignocellulosic feedstocks: a review. *Bioresour Bioprocess*, 4(1), 1-19. doi:<https://doi.org/10.1186/s40643-017-0137-9>
- Kumar, Singh, & Singh. (2008). Bioconversion of lignocellulosic biomass: biochemical and molecular perspectives. *Journal of Industrial Microbiology and Biotechnology*, 35(5), 377-391.

- Kumari, & Singh. (2018). Pretreatment of lignocellulosic wastes for biofuel production: a critical review. *Renewable and Sustainable Energy Reviews*, 90, 877-891.
- Ladeira Ázar, Bordignon-Junior, Laufer, Specht, Ferrier, & Kim. (2020). Effect of lignin content on cellulolytic saccharification of liquid hot water pretreated sugarcane bagasse. *Molecules*, 25(3), 623. doi:<https://doi.org/10.3390/molecules25030623>
- Lau, Thoma, Clausen, & Carrier. (2014). Kinetic modeling of xylose oligomer degradation during pretreatment in dilute acid or in water. *Industrial & Engineering Chemistry Research*, 53(6), 2219-2228.
- Lee, Jameel, & Venditti. (2010). A comparison of the autohydrolysis and ammonia fiber explosion (AFEX) pretreatments on the subsequent enzymatic hydrolysis of coastal Bermuda grass. *Bioresource Technology*, 101(14), 5449-5458.
- Lee, & Jeffries. (2011). Efficiencies of acid catalysts in the hydrolysis of lignocellulosic biomass over a range of combined severity factors. *Bioresour Technol*, 102(10), 5884-5890. doi:<https://doi.org/10.1016/j.biortech.2011.02.048>
- Lee, Kazlauskas, & Park. (2017). One-step pretreatment of yellow poplar biomass using peracetic acid to enhance enzymatic digestibility. *Scientific reports*, 7(1), 12216.
- Li, Cao, Meng, Studer, Wyman, Ragauskas, & Pu. (2017). The effect of liquid hot water pretreatment on the chemical–structural alteration and the reduced recalcitrance in poplar. *Biotechnology for biofuels*, 10(1), 1-13.
- Li, Zhang, Guo, Hu, Zhang, Feng, Yi, Zou, Wang, Wu, Tian, Lu, Xie, & Peng. (2015). High-level hemicellulosic arabinose predominately affects lignocellulose crystallinity for genetically enhancing both plant lodging resistance and biomass enzymatic digestibility in rice mutants. *Plant Biotechnol J*, 13(4), 514-525. doi:10.1111/pbi.12276
- Limayem, & Ricke. (2012). Lignocellulosic biomass for bioethanol production: current perspectives, potential issues and future prospects. *Progress in energy and combustion science*, 38(4), 449-467.
- Lin, Waters, Mallinson, Lobban, & Bartley. (2015). Relationships between biomass composition and liquid products formed via pyrolysis. *Frontiers in Energy Research*, 3, 45.

- Liu. (2006). Genomic adaptation of ethanologenic yeast to biomass conversion inhibitors. *Applied microbiology and biotechnology*, 73(1), 27-36.
- Liu, & Hui. (2014). Acetic Acid Catalyzed Steam Explosion for Improving the Sugar Recovery of Wheat Straw. *BioResources*, 9(3), 4703-4709.
- Liu, & Wyman. (2003a). The effect of flow rate of compressed hot water on xylan, lignin, and total mass removal from corn stover. *Industrial & Engineering Chemistry Research*, 42(21), 5409-5416.
- Liu, & Wyman. (2003b). The effect of flow rate of compressed hot water on xylan, lignin, and total mass removal from corn stover. *Ind Eng Chem Res*, 42(21), 5409-5416. doi:<https://doi.org/10.1021/ie030458k>
- Lu, & Mosier. (2007). Biomimetic catalysis for hemicellulose hydrolysis in corn stover. *Biotechnology progress*, 23(1), 116-123.
- Lu, & Mosier. (2008). Kinetic modeling analysis of maleic acid-catalyzed hemicellulose hydrolysis in corn stover. *Biotechnology and bioengineering*, 101(6), 1170-1181.
- Maneeintr, Leewisuttikul, Kerdsuk, & Charinpanitkul. (2018). Hydrothermal and enzymatic treatments of pineapple waste for energy production. *Energy Procedia*, 152, 1260-1265. doi:<https://doi.org/10.1016/j.egypro.2018.09.179>
- Mansora, Lima, Anib, Hashima, & Hoa. (2019). Characteristics of cellulose, hemicellulose and lignin of MD2 pineapple biomass. *CHEMICAL ENGINEERING*, 72(1), 79-84.
- Mardawati, Andoyo, Syukra, Kresnowati, & Bindar. (2018). *Production of xylitol from corn cob hydrolysate through acid and enzymatic hydrolysis by yeast*. Paper presented at the IOP Conference Series: Earth and Environmental Science.
- Marques, Soares, Lomonaco, e Silva, Santaella, de Freitas Rosa, & Leitao. (2021). Steam explosion pretreatment improves acetic acid organosolv delignification of oil palm mesocarp fibers and sugarcane bagasse. *Int J Biol Macromol*, 175, 304-312. doi:<https://doi.org/10.1016/j.ijbiomac.2021.01.174>
- McMillan. (1994). Pretreatment of lignocellulosic biomass. *ACS Publications*.
- Mechmech, Chadjaa, Rahni, Marinova, Akacha, & Gargouri. (2015). Improvement of butanol production from a hardwood hemicelluloses hydrolysate by combined sugar concentration and phenols removal. *Bioresour Technol*, 192, 287-295. doi:<https://doi.org/10.1016/j.biortech.2015.05.012>

- Megashah, Ariffin, Zakaria, & Hassan. (2018). *Properties of cellulose extract from different types of oil palm biomass*. Paper presented at the IOP Conference Series: Materials Science and Engineering.
- Miller. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical chemistry*, 31(3), 426-428.
- Modenbach, & Nokes. (2012). The use of high-solids loadings in biomass pretreatment—a review. *Biotechnology and Bioengineering*, 109(6), 1430-1442.
- Moodley, & Kana. (2017). Comparison of a two-stage and a combined single stage salt-acid based lignocellulosic pretreatment for enhancing enzymatic saccharification. *Industrial Crops and Products*, 108, 219-224.
- Mosier, Ladisch, & Ladisch. (2002a). Characterization of acid catalytic domains for cellulose hydrolysis and glucose degradation. *Biotechnol Bioeng*, 79(6), 610-618. doi:<https://doi.org/10.1002/bit.10316>
- Mosier, Ladisch, & Ladisch. (2002b). Characterization of acid catalytic domains for cellulose hydrolysis and glucose degradation. *Biotechnology and bioengineering*, 79(6), 610-618.
- Mosier, Sarikaya, Ladisch, & Ladisch. (2001). Characterization of dicarboxylic acids for cellulose hydrolysis. *Biotechnology progress*, 17(3), 474-480.
- Mosier, Wyman, Dale, Elander, Lee, Holtzapple, & Ladisch. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource technology*, 96(6), 673-686.
- Mouthier, Appeldoorn, Pel, Schols, Gruppen, & Kabel. (2018). Corn stover lignin is modified differently by acetic acid compared to sulfuric acid. *Industrial Crops and Products*, 121, 160-168.
- Nakagame, Chandra, Kadla, & Saddler. (2011). The isolation, characterization and effect of lignin isolated from steam pretreated Douglas-fir on the enzymatic hydrolysis of cellulose. *Bioresource technology*, 102(6), 4507-4517.
- Oktaviani, Hermiati, Thontowi, Laksana, Kholida, Andriani, & Mangunwardoyo. (2019). *Production of xylose, glucose, and other products from tropical lignocellulose biomass by using maleic acid pretreatment*. Paper presented at the IOP Conference Series: Earth and Environmental Science.

- Quatmane, Provenzano, Hafidi, & Senesi. (2000). Compost Maturity Assessment Using Calorimetry, Spectroscopy and Chemical Analysis. *Compost Science & Utilization*, 8(2).
- Pal, Joy, Trimukhe, Kumbhar, Varma, & Padmanabhan. (2016). Pretreatment and enzymatic process modification strategies to improve efficiency of sugar production from sugarcane bagasse. *3 Biotech*, 6(2), 1-14. doi:<https://doi.org/10.1007/s13205-016-0446-2>
- Palmqvist, & Hahn-Hägerdal. (2000). Fermentation of lignocellulosic hydrolysates. I: inhibition and detoxification. *Bioresour Technol*, 74(1), 17-24. doi:[https://doi.org/10.1016/S0960-8524\(99\)00160-1](https://doi.org/10.1016/S0960-8524(99)00160-1)
- Pattiya, Chaow-u-Thai, & Rittidech. (2013). The influence of pretreatment techniques on ash content of cassava residues. *International Journal of Green Energy*, 10(5), 544-552.
- Paz, Outeiriño, Guerra, & Domínguez. (2019). Enzymatic hydrolysis of brewer's spent grain to obtain fermentable sugars. *Bioresource Technology*, 275, 402-409.
- Pecha, & Garcia-Perez. (2015). *Pyrolysis of lignocellulosic biomass: oil, char, and gas*. Paper presented at the Bioenergy.
- Pourmorad, Hosseinimehr, & Shahabimajd. (2006). Antioxidant activity, phenol and flavonoid contents of some selected Iranian medicinal plants. *African journal of biotechnology*, 5(11).
- Pradhan. (2015). *Physical treatments for reducing biomass ash and effect of ash content on pyrolysis products*. Auburn University,
- Qiao, Cui, Ouyang, Shi, & Ouyang. (2019). Comparison of dilute organic acid pretreatment and a comprehensive exploration of citric acid pretreatment on corn cob. *Journal of Renewable Materials*, 7(11), 1197-1207.
- Radotić, & Mičić. (2016). Methods for extraction and purification of lignin and cellulose from plant tissues. In *Sample preparation techniques for soil, plant, and animal samples* (pp. 365-376): Springer.
- Rahimi, Ulbrich, Coon, & Stahl. (2014). Formic-acid-induced depolymerization of oxidized lignin to aromatics. *Nature*, 515(7526), 249-252.
- Rattanaporn, Tantayotai, Phusantisampan, Pornwongthong, & Sriariyanun. (2018). Organic acid pretreatment of oil palm trunk: effect on enzymatic saccharification and ethanol production. *Bioprocess and biosystems engineering*, 41(4), 467-477.

- Ravindran, & Jaiswal. (2016). A comprehensive review on pre-treatment strategy for lignocellulosic food industry waste: challenges and opportunities. *Bioresource technology*, 199, 92-102.
- Reza, Emerson, Uddin, Gresham, & Coronella. (2015). Ash reduction of corn stover by mild hydrothermal preprocessing. *Biomass Conversion and Biorefinery*, 5(1), 21-31.
- Roche, Dibble, & Stickel. (2009). Laboratory-scale method for enzymatic saccharification of lignocellulosic biomass at high-solids loadings. *Biotechnol Biofuels*, 2(1), 1-11. doi:<https://doi.org/10.1186/1754-6834-2-28>
- Roda, De Faveri, Dordoni, & Lambri. (2014). Vinegar production from pineapple wastes—Preliminary saccharification trials. *Chem Eng*, 37. doi:<https://doi.org/10.3303/CET1437102>
- Roda, De Faveri, Giacosa, Dordoni, & Lambri. (2016a). Effect of pre-treatments on the saccharification of pineapple waste as a potential source for vinegar production. *J Clean Prod*, 112, 4477-4484. doi:<https://doi.org/10.1016/j.jclepro.2015.07.019>
- Roda, De Faveri, Giacosa, Dordoni, & Lambri. (2016b). Effect of pre-treatments on the saccharification of pineapple waste as a potential source for vinegar production. *Journal of cleaner production*, 112, 4477-4484.
- Rosdee, Masngut, Shaarani, Jamek, & Sueb. (2020). *Enzymatic hydrolysis of lignocellulosic biomass from pineapple leaves by using endo-1, 4-xylanase: Effect of pH, temperature, enzyme loading and reaction time*. Paper presented at the IOP Conference Series: Materials Science and Engineering.
- Rosli, Harun, Jahim, & Othaman. (2017a). Chemical and physical characterization of oil palm empty fruit bunch. *Malays J Anal Sci*, 21(1), 188-196. doi:<https://doi.org/10.17576/mjas-2017-2101-22>
- Rosli, Harun, Jahim, & Othaman. (2017b). Chemical and physical characterization of oil palm empty fruit bunch. *Malaysian Journal of Analytical Sciences*, 21(1), 188-196.
- Rowell, Pettersen, Han, Rowell, & Tshabalala. (2005). Cell wall chemistry. *Handbook of wood chemistry and wood composites*, 2.
- Rowell, & Rowell. (1996). *Paper and composites from agro-based resources*: CRC press.

- Roy, Rahman, & Raynie. (2020). Recent advances of greener pretreatment technologies of lignocellulose. *CRGSC*, 100035. doi:<https://doi.org/10.1016/j.crgsc.2020.100035>
- Saha, Iten, Cotta, & Wu. (2005). Dilute acid pretreatment, enzymatic saccharification, and fermentation of rice hulls to ethanol. *Biotechnology progress*, 21(3), 816-822.
- Saha, Kurade, El-Dalatony, Chatterjee, Lee, & Jeon. (2016). Improving bioavailability of fruit wastes using organic acid: An exploratory study of biomass pretreatment for fermentation. *Energy conversion and management*, 127, 256-264.
- Sahu, & Pramanik. (2018a). Evaluation and optimization of organic acid pretreatment of cotton gin waste for enzymatic hydrolysis and bioethanol production. *Applied biochemistry and biotechnology*, 186(4), 1047-1060.
- Sahu, & Pramanik. (2018b). Evaluation and optimization of organic acid pretreatment of cotton gin waste for enzymatic hydrolysis and bioethanol production. *Applied biochemistry and biotechnology*, 186(4), 1047-1060. doi:<https://doi.org/10.1007/s12010-018-2790-7>
- Samuel, Foston, Jiang, Allison, & Ragauskas. (2011). Structural changes in switchgrass lignin and hemicelluloses during pretreatments by NMR analysis. *Polymer degradation and stability*, 96(11), 2002-2009.
- Sánchez, Sierra, & Alméciga-Díaz. (2011). Delignification process of agro-industrial wastes an alternative to obtain fermentable carbohydrates for producing fuel. In *Alternative fuel*: IntechOpen.
- Sannigrahi, Pu, & Ragauskas. (2010). Cellulosic biorefineries—unleashing lignin opportunities. *Current Opinion in Environmental Sustainability*, 2(5-6), 383-393.
- Santos, Neiva Correia, Mateus, Saraiva, Vicente, & Moldão. (2019). Fourier transform infrared (FT-IR) spectroscopy as a possible rapid tool to evaluate abiotic stress effects on pineapple by-products. *Appl Sci*, 9(19), 4141. doi:<https://doi.org/10.3390/app9194141>
- Savelkoul, Chanput, & Wichers. (2013). Immunomodulatory effects of mushroom β -glucans. In *Diet, immunity and inflammation* (pp. 416-434): Elsevier.

- Schoenherr, Ebrahimi, & Czermak. (2017). Lignin Degradation Processes and the Purification of Valuable Products. In *Lignin-Trends and Applications: IntechOpen*.
- Sen, Chou, Wu, & Liu. (2016). Pretreatment conditions of rice straw for simultaneous hydrogen and ethanol fermentation by mixed culture. *International Journal of Hydrogen Energy*, 41(7), 4421-4428.
- Shimizu, de Azevedo, Coelho, Pagnocca, & Brienzo. (2020). Minimum Lignin and Xylan Removal to Improve Cellulose Accessibility. *BioEnergy Research*, 1-11.
- Singh, Matsagar, & Dhepe. (2021). Determination of Alpha-, Beta-and Gamma-Cellulose in Bagasse and Wheat Straw: Lignin Recovery, Characterization and Depolymerization.
- Singh, Sarangi, & Singh. (2018). Tenderisation of meat by bromelain enzyme extracted from pineapple wastes. *International Journal of Current Microbiology and Applied Sciences*, 7(9), 3256-3264.
- Sluiter, Hames, Hyman, Payne, Ruiz, Scarlata, Sluiter, Templeton, & Wolfe. (2008). Determination of total solids in biomass and total dissolved solids in liquid process samples. *National Renewable Energy Laboratory, Golden, CO, NREL Technical Report No. NREL/TP-510-42621*, 1-6.
- Sluiter, Hames, Ruiz, Scarlata, Sluiter, & Templeton. (2005). Determination of ash in biomass laboratory analytical procedure. *National Renewable Energy Laboratory Analytical Procedure, Golden, CO*.
- Solikhin, Hadi, Massijaya, & Nikmatin. (2016). Basic Properties of oven-heat treated oil palm empty fruit bunch stalk fibers. *BioResources*, 11(1), 2224-2237.
- Soontornchaiboon, Kim, & Pawongrat. (2016). Effects of alkaline combined with ultrasonic pretreatment and enzymatic hydrolysis of agricultural wastes for high reducing sugar production. *Sains Malaysiana*, 45(6), 955-962.
- Sudiyani, Sembiring, Hendarsyah, & Alawiyah. (2010). Alkaline pretreatment and enzymatic saccharification of oil palm empty fruit bunch fiber for ethanol production. *Menara Perkebunan*, 78(2), 70-74.
- Sukkaew, Boonsong, Thongpradistha, & Intan. (2017). *Physical and chemical pretreatment of lignocellulosics in pineapple (ananas comosus) peels dried for investment*. Paper presented at the AIP Conference Proceedings.

- Sukruansuwan, & Napathorn. (2018a). Use of agro-industrial residue from the canned pineapple industry for polyhydroxybutyrate production by *Cupriavidus necator* strain A-04. *Biotechnol Biofuels*, *11*(1), 1-15. doi:<https://doi.org/10.1186/s13068-018-1207-8>
- Sukruansuwan, & Napathorn. (2018b). Use of agro-industrial residue from the canned pineapple industry for polyhydroxybutyrate production by *Cupriavidus necator* strain A-04. *Biotechnology for biofuels*, *11*(1), 202.
- Sun, Cao, Sun, Xu, Song, Sun, & Jones. (2014). Improving the enzymatic hydrolysis of thermo-mechanical fiber from *Eucalyptus urophylla* by a combination of hydrothermal pretreatment and alkali fractionation. *Biotechnology for biofuels*, *7*(1), 116.
- Sun, Foston, Meng, Sawada, Pingali, O'Neill, Li, Wyman, Langan, & Ragauskas. (2014). Effect of lignin content on changes occurring in poplar cellulose ultrastructure during dilute acid pretreatment. *Biotechnology for Biofuels*, *7*(1), 1-14.
- Sun, Sun, Cao, & Sun. (2016). The role of pretreatment in improving the enzymatic hydrolysis of lignocellulosic materials. *Bioresource technology*, *199*, 49-58.
- Tang, Cao, Xu, Wu, Li, Ye, Luo, Gao, Liao, & Yan. (2020). One-Step or Two-Step Acid/Alkaline Pretreatments to Improve Enzymatic Hydrolysis and Sugar Recovery from *Arundo Donax* L. *Energies*, *13*(4), 948.
- Tappi. (2004). 204 cm-97, Solvent extractives of wood and pulp. *TAPPI test methods*, 2005.
- Upadhyay, Lama, & Tawata. (2010). Utilization of pineapple waste: a review. *Journal of Food Science and Technology Nepal*, *6*, 10-18.
- Valdez-Fragoso, Mújica-Paz, Welti-Chanes, & Torres. (2011). Reaction kinetics at high pressure and temperature: effects on milk flavor volatiles and on chemical compounds with nutritional and safety importance in several foods. *Food and Bioprocess Technology*, *4*(6), 986-995.
- Van Der Maas, Scott, & Van Haasterecht. (2016). *The effect of dilute-acid pretreatment of cellulose crystallinity and digestibility*. Netherlands: Wageningen University,
- van der Pol, Vaessen, Weusthuis, & Eggink. (2016). Identifying inhibitory effects of lignocellulosic by-products on growth of lactic acid producing micro-

- organisms using a rapid small-scale screening method. *Bioresource Technology*, 209, 297-304.
- Vanholme, Demedts, Morreel, Ralph, & Boerjan. (2010). Lignin biosynthesis and structure. *Plant physiology*, 153(3), 895-905.
- Wang, Beine, & Palkovits. (2019). 1, 2-Propylene Glycol and Ethylene Glycol Production From Lignocellulosic Biomass. In *Studies in Surface Science and Catalysis* (Vol. 178, pp. 173-193): Elsevier.
- Wang, Wei, Li, Sun, He, & He. (2016). Comparative study of alkali and acidic cellulose solvent pretreatment of corn stover for fermentable sugar production. *BioResources*, 11(1), 482-491. doi:<https://doi.org/10.15376/biores.11.1.482-491>
- Weil, Sarikaya, Rau, Goetz, Ladisch, Brewer, Hendrickson, & Ladisch. (1998). Pretreatment of corn fiber by pressure cooking in water. *Applied biochemistry and biotechnology*, 73(1), 1-17.
- Wu, Snajdrova, Moore, Baldenius, & Bornscheuer. (2021). Biocatalysis: enzymatic synthesis for industrial applications. *Angewandte Chemie International Edition*, 60(1), 88-119.
- Xia, Liu, Hu, Li, Li, & Ma. (2021). Macromolecules Evolution in Sequential Acid-Alkali Pretreatment of Corn Stalk. Available at SSRN 3919664.
- Xiao, Sun, Shi, Xu, & Sun. (2011). Impact of hot compressed water pretreatment on the structural changes of woody biomass for bioethanol production. *BioResources*, 6(2), 1576-1598. doi:<https://doi.org/10.15376/BIORES.6.2.1576-1598>
- Ximenes, Kim, Mosier, Dien, & Ladisch. (2011). Deactivation of cellulases by phenols. *Enzyme Microb Technol*, 48(1), 54-60. doi:<https://doi.org/10.1016/j.enzmictec.2010.09.006>
- Xin, Dong, Zhang, Ma, & Jiang. (2019). Biobutanol production from crystalline cellulose through consolidated bioprocessing. *Trends in biotechnology*, 37(2), 167-180.
- Xue, Li, An, Li, Li, Wu, & Wei. (2021). Ethylene glycol based acid pretreatment of corn stover for cellulose enzymatic hydrolysis. *RSC Adv*, 11(23), 14140-14147. doi:<https://doi.org/10.1039/D0RA10877D>
- Yang, Dai, Ding, & Wyman. (2011). Enzymatic hydrolysis of cellulosic biomass. *Biofuels*, 2(4), 421-449.

- Yi. (2021). Tiny bugs play big role: microorganisms' contribution to biofuel production. In *Advances in 2nd Generation of Bioethanol Production* (pp. 113-136): Elsevier.
- Youcai, & Guangyin. (2016). *Pollution Control and Resource Recovery: Sewage Sludge*: Butterworth-Heinemann.
- Youssefian, & Rahbar. (2015). Molecular origin of strength and stiffness in bamboo fibrils. *Scientific reports*, 5, 11116.
- Zeng, Zhao, Wei, Tucker, Himmel, Mosier, Meilan, & Ding. (2015). In situ micro-spectroscopic investigation of lignin in poplar cell walls pretreated by maleic acid. *Biotechnol Biofuels*, 8(1), 1-12. doi:<https://doi.org/10.1186/s13068-015-0312-1>
- Zhang, Choi, Yoo, Kim, Brown, & Shanks. (2015). Cellulose–hemicellulose and cellulose–lignin interactions during fast pyrolysis. *ACS Sustainable Chemistry & Engineering*, 3(2), 293-301.
- Zheng, & Rehmann. (2014). Extrusion pretreatment of lignocellulosic biomass: a review. *International journal of molecular sciences*, 15(10), 18967-18984.
- Zhiqiang, FeiB enhua, & Zehui. (2015). Effect of steam explosion pretreatment on bamboo for enzymatic hydrolysis and ethanol fermentation. *BioResources*, 10(1), 1037-1047.
- Zhou, Li, Mabon, & Broadbelt. (2017). A critical review on hemicellulose pyrolysis. *Energy Technology*, 5(1), 52-79.
- Zhuang, Wang, Yu, Qi, Wang, Tan, Zhou, & Yuan. (2016). Liquid hot water pretreatment of lignocellulosic biomass for bioethanol production accompanying with high valuable products. *Bioresour Technol*, 199, 68-75. doi:doi: 10.1016/j.biortech.2015.08.051

LIST OF PUBLICATION

Indexed Conference Proceedings

1. Nordin, N., Illias, R. M., Manas, N. H. A., Nor, A., Ramli, M., & Azelee, N. I. W. (2020, December). Efficient Delignification of Pineapple Waste by Low-Pressure Steam Heating Pre-Treatment. In *Third International Conference on Separation Technology 2020 (ICoST 2020)* (pp. 10-16). Atlantis Press. <https://doi.org/10.2991/aer.k.201229.002>. **(Indexed by SCOPUS)**

Indexed Journal

1. Azelee, N. I. W., Nordin, N., Illias, R. M., Hasmaliana, N., Manas, A., & Ghazali, M. N. F. M. Enzyme Kinetics Study for Heterogeneous System of Pretreated Kenaf Hydrolysis **(Indexed by ISI)**