LIGNOCELLULOSICS BIOMASS BIODEGRADATION OF WATER HYACINTH AND WATER LETTUCE BY WHITE ROT FUNGI FOR BIOETHANOL PRODUCTION

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DEDICATION

This thesis is dedicated to all my beloved family and friends.

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ABSTRACT

Nowadays, renewable energy has become alternative energy to reduce the consumption of fossil fuels. Therefore, lignocellulosic materials such as crop residues, grass and wood, and aquatic plants that are inedible has become potential sources for bioethanol production. In this study, water hyacinth (WH) and water lettuce (WL) were selected as potential resources of their abundance in nature and can be easily propagated and cultivated. Although these floating aquatic plants are considered as the most problematic plants due to their uncontrollable growth in water bodies worldwide, their ability to remove pollutants from wastewater has created a sustainable approach for their use in phytoremediation and further use as biomass substrates for bioethanol production. The use of phytoremediation by implementing invasive floating aquatic plants can support the sustainable management of wastewater treatment in the future. This study aims to determine the potential of WH and WL as bioindicators for phytoremediation and at same time to produce a high amount of sugar consumption for bioethanol production. In addition, this study emphasizes the biodegradation of WH and WL by white-rot fungi collected from decayed wood and soil. White-rot fungi have the ability to degrade lignin, hydrolyze cellulose, and hemicellulose, and ferment alcohols for bioethanol production. Trichoderma citrinoviride M3, Schizophyllum commune M8, and Pestalotiopsis sp. M12 were selected from twelve fungal species on the basis of rapid growth rate after five days of incubation and further use for degradation of lignocellulosic materials from water hyacinth and water lettuce and for bioethanol production. These fungal species were identified by morphological characterization and 18S rRNA sequence analysis. To date, the use of biological pretreatment using T. citrinoviride M3, S. commune M8, and Pestalotiopsis sp. M12 with regard to water hyacinth and water lettuce substrates as well as its further use for the fermentation process to produce bioethanol has not been explored before. The parameters involved are sugar content by the dinitrosalicylic acid (DNS) Method, the determination of lignin by the Klason Method, the determination of cellulose and hemicellulose by the Chesson Method, and the determination of bioethanol by Gas Chromatography (GC). The results showed that both WH and WL indicated the same correlation trend between biomass growth and sugar content, as the sugar content increased when the plants reached the highest growth. However, WH has more extractable sugar than WL, which is more significant because fermentable sugar is needed for the fermentation process to produce bioethanol. The results also showed that T. citrinoviride M3 has the highest rate of degradation of lignocellulosic materials compared to S. commune M8 and Pestalotiopsis sp. M12. Therefore, T. citrinoviride M3 was selected for further investigation to evaluate the simultaneous saccharification and fermentation process for bioethanol production. The results showed that WH and WL produced 8.6 g/L and 7.4 g/L yield of ethanol, respectively, proportional to the fermentation time, with an increasing time of up to 96 hours. Overall, it can be concluded that WH is a promising biomass when the simultaneous co-cultivation of T. citrinoviride M3 with S. cerevisiae is used for bioethanol production and the fermentation process is fully optimized. The findings of this study would be beneficial for future investigation, especially in exploring the potential production of bioethanol from phytoremediation technology systems.

ABSTRAK

Pada masa kini, tenaga boleh diperbaharui telah menjadi tenaga alternatif untuk mengurangkan penggunaan bahan bakar fosil. Oleh itu, bahan lignoselulosa seperti sisa tanaman, rumput dan kayu, dan tumbuhan akuatik yang tidak boleh dimakan telah menjadi sumber yang berpotensi untuk pengeluaran bioetanol. Dalam kajian ini, keladi bunting (WH) dan pokok kiambang (WL) dipilih sebagai sumber daya yang berpotensi kerana sifatnya mudah ditanam dan boleh membiak dengan banyaknya. Walaupun tumbuhan akuatik terapung ini dianggap sebagai tumbuhan yang paling bermasalah kerana pertumbuhannya yang tidak terkawal di atas air sisa, namun keupayaan mereka untuk membuang bahan pencemar daripada air sisa telah mewujudkan pendekatan yang mampan untuk kegunaannya dalam fitoremediasi dan penggunaan selanjutnya sebagai bahan untuk pengeluaran bioetanol. Oleh itu, penggunaan pokok akuatik terapung ini pastinya dapat menyokong pengurusan rawatan air sisa yang mampan pada masa hadapan. Kajian ini bertujuan untuk menentukan potensi WH dan WL sebagai bioindikator untuk fitoremediasi dan pada masa yang sama menghasilkan jumlah penggunaan gula yang tinggi untuk pengeluaran bioetanol. Di samping itu, kajian ini menekankan pada biodegradasi WH dan WL oleh kulat yang dikumpulkan dari kayu dan tanah yang reput. Kulat ini mempunyai kemampuan untuk menurunkan kandungan lignin, menghidrolisis selulosa dan hemiselulosa, dan fermentasi alkohol untuk pengeluaran bioetanol. Trichoderma citrinoviride M3, Schizophyllum commune M8 dan Pestalotiopsis sp M12 dipilih dari dua belas spesies kulat berdasarkan kadar pertumbuhan pesat setelah lima hari inkubasi. Spesies kulat dikenal pasti dengan ciri morfologi dan analisis urutan rRNA 18S. Sehingga kini, penggunaan prarawatan biologi menggunakan T. citrinoviride M3, S. commune M8, dan Pestalotiopsis sp. M12 menggunakan substrat WH dan WL serta penggunaan selanjutnya untuk proses penapaian untuk menghasilkan bioetanol belum pernah diterokai sebelum ini. Tiga spesies ini seterusnya digunakan untuk mendegradasi bahan lignoselulosa dari WH dan WL dan parameter yang terlibat adalah kandungan gula dengan Kaedah asid dinitrosalicylic (DNS), penentuan lignin dengan Kaedah Klason, penentuan selulosa dan hemiselulosa dengan Kaedah Chesson dan penentuan bioetanol oleh Kromatografi Gas (GC). Hasil kajian menunjukkan bahawa kedua-dua pokok menunjukkan korelasi trend yang sama antara kadar pertumbuhan dan kandungan gula, ketika kandungan gula meningkat maka tanaman akuatik juga mencapai peningkatan pertumbuhan tertinggi. Namun, WH mempunyai lebih banyak gula yang dapat diekstrak berbanding dengan WL, kerana gula fermentasi diperlukan untuk proses fermentasi untuk menghasilkan bioetanol. Hasil kajian menunjukkan bahawa T. citrinoviride M3 mempunyai kadar degradasi tertinggi bahan lignoselulosa berbanding dengan S. commune M8 dan Pestalotiopsis sp M12. Oleh itu, T. citrinoviride M3 dipilih untuk penyelidikan seterusnya untuk menilai proses sakarifikasi dan fermentasi serentak untuk pengeluaran bioetanol. Selain itu, keputusan menunjukkan bahawa WH dan WL menghasilkan 8.6 g/L dan 7.4 g/L hasil bioetanol, masing-masing, berkadar dengan masa fermentasi, dengan masa yang meningkat sehingga 96 jam. Ini dapat disimpulkan bahawa WH adalah bahan biomas yang berpotensi untuk digunakan apabila digabungkan T. citrinoviride M3 dengan S. cerevisiae untuk pengeluaran bioetanol dan proses fermentasi dioptimumkan sepenuhnya. Hasil kajian ini akan bermanfaat untuk kajian masa hadapan, terutamanya dalam menerokai potensi penghasilan bioetanol daripada sistem teknologi fitoremediasi.

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LIST OF ABBREVIATIONS

DNS - Dinitrosalicylic acid

GC - Gas Chromatography

PDA - Potato Dextrose Agar

RBBR - Brilliant Blue R

SEM - Scanning Electron Microscopy

SSF - Simultaneous Saccharification and Fermentation

UTM - Universiti Teknologi Malaysia

WH - Water Hyacinth

WL - Water Lettuce

LIST OF SYMBOLS

h - hour

g - gram

g/L - gram per litre

g/g - gram per gram

mg/g - milligram per gram

°C - degree Celsius

μm - micrometer

PO₄³⁻ - phosphate

NO₃ - nitrate

NO₂ - nitrite

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CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Higher demand for energy and concern for the environment is the crucial factor leading to renewable energy sources. Energy demand in Malaysia is constantly increasing due to advancements in the industrial and transportation sectors (Salleh et al., 2020; Ayodele et al., 2021), and the global depletion of energy resources due to energy consumption and environmental degradation caused by fossil fuels has prompted research on alternative renewable energy sources. As a result, the demand for petroleum-derived fuels has not decreased but increased sharply in recent decades (Shamsuddin, 2012; Li et al., 2021). In contrast, the challenges in finding a long-term solution for a reliable and infinite source of clean energy supply in the future are enormous. The Five-Fuel Diversification Policy Malaysia has consistently promoted renewable energy (RE) over non-renewable energy sources in its energy policy, along with fossil fuels and hydropower (Chin and H'ng, 2013). Malaysia has formulated the National Biofuel Policy to promote the prosperity and sustainable development of the country, and the Sustainable Energy Development Authority (SEDA) has proposed the National Renewable Energy Policy, setting an ambitious target by aiming for 20% of the capacity mix constituting renewable energy sources to reduce greenhouse gas emissions and creating a more conducive environment (Abdullah et al., 2019). Therefore, renewable bioethanol becomes a potential energy source for Malaysia's future renewable energy development prospects.

Bioethanol production deals with biotechnological production from various feedstock sources. The selection of the most suitable feedstock for bioethanol production depends on local conditions. Conventional bioethanol production has been reported from various feedstocks such as sugarcane, corn, and starchy materials; however, there has been a rising interest in low-cost, readily available, and abundant

lignocellulosic biomass that could serve as a potential feedstock for bioethanol production (Wyman et al., 2019; González-Gloria et al., 2022). Lignocellulosic biomass, which includes agricultural and forestry residues and wastes, provides a more excellent choice of potential feedstocks that do not conflict with land use for food production and is also cheaper than conventional bioethanol sources. Lignocellulosic biomass typically consists of carbohydrates that can be derived from monosaccharides and disaccharides (Singh et al., 2019).

Conversely, carbohydrates comprise cellulose and hemicellulose that are tightly wrapped by lignin. Many researchers worldwide are working hard to convert lignocellulosic biomass, such as straws and other plant wastes, into bioethanol production (Arefin et al., 2021). Although the biological conversion of lignocellulosic biomass into monomeric sugars is more complicated than the conversion of starch that is currently used for bioethanol production, many countries (Sweden, Australia, Canada, and Japan) are making efforts to utilize this lignocellulosic biomass into ethanol (Singh and Satapathy, 2018; Singh et al., 2022).

Floating aquatic plants are potential lignocellulosic biomass when it comes to bioethanol production. Currently, the extensive focus has been placed on developing methods to produce bioethanol from aquatic plants with high cellulose-hemicellulose contents. Water hyacinth (WH) and water lettuce (WL) are potential feedstocks since they constitute abundant floating aquatic plants and are quick to propagate for bioethanol production. Moreover, water hyacinth and water lettuce also act as bioindicators for phytoremediation technology to remediate contaminated domestic wastewater. Phytoremediation refers to a new plant-based technology that can be used as an alternative option to purify contaminated water. This technology focuses on using floating aquatic plants to eliminate pollutants from wastewater (Rezania et al., 2015). The selection of aquatic plants is essential as it is the main tool for removing contaminants (De Stefani et al., 2011; Ansari et al., 2020). Thus, the selection is based on the high contaminant uptake as well as fast and easy growth (Roongtanakiat et al., 2007; Mustafa and Hayder, 2021). Microorganisms attached to plant roots can oxidize biodegradable materials in contaminated wastewater (Nayanthara and Bindu, 2017).

Water hyacinth (WH), which is also known as Eichhornia crassipes, is a fast-growing invasive plant that can double its reproduction in less than 13 days (Rezania et al., 2015; Arefin et al., 2021). As a result, this plant can disturb aquatic ecosystems and activities, thus causing problems for navigation. However, due to their high hemicellulose and low lignin content, WH is feasible for bioethanol production (Arefin et al., 2021).

Water lettuce (WL), also known as Pistia stratiotes L., has been widely used to mitigate pollutants from contaminated water (Gupta et al., 2012; Karmakar et al., 2018; Amalia et al., 2019) and belongs to the Araceae family, which can reproduce rapidly and vegetatively on the water surface (Hussnera et al., 2014; Walsh and Maestro, 2014; Lien et al., 2019). WL floats on the water surface and its roots submerse beneath water bodies. The size of WL leaves can reach up to 13 cm long (Dipu et al., 2011). WL also has a soft body that can increase biochemical responsiveness and low labor harvesting due to its small body size (Mishima et al., 2008). Therefore, an emphasis has been placed on exploring WH and WL as bioindicators of phytoremediation technology and as substrates for bioethanol production in future works.

1.2 Problem Statement

The excess concentration of nutrients released from domestic wastewater has become of concern because it has triggered eutrophication, harmful algal blooms, and fish kills in water bodies. Eutrophication typically causes excessive growth of algae on the water surface and subsequently leads to the deterioration of water quality (Sabeena et al., 2018; Feng et al., 2023). Due to the accumulation of organic and nutrients, such as nitrogen and phosphorus, which come from the nearby wastewater domestic ponds, this condition is relatively similar to the selected location of Desa Bakti wastewater. As a result, discharge from the drainage system, encompassing residential and gardening areas as well as nonpoint surface runoff on rainy days, had an impact on it. Therefore, it is essential to remove the wastewater's nutrients to achieve environmental sustainability (Mulling et al., 2014; Qv et al., 2023).

Nowadays, several treatment processes, including physical, chemical, and biological processes, have been used to remove nutrients from wastewater (Nizam et al., 2020; Nidheesh et al., 2022). However, these treatment methods require high operational and maintenance costs (Ali et al., 2020). Among all the treatment methods, phytoremediation is a promising technique for removing nutrients from wastewater owing to the eco-friendly, cost-effective, and efficient process it offers. This technology focuses on the use of floating aquatic macrophytes as a tool to eliminate pollutants from wastewater (Rezania et al. 2015; Ansari et al., 2020). To date, comprehensive information on how to choose plants that will maximize the contribution of each phytoremediation mechanism and how to improve phytoremediation technology performance are still lacking (Qin et al., 2020). Several studies have described the effectiveness of aquatic plants, such as water hyacinth (E. crassipes), water lettuce (P. stratiotes), duckweed (L. minor), and giant salvinia (S. molesta) to remove various pollutants from wastewater (Rezania et al., 2015; Qin et al. 2016; Ting et al., 2018; Ansari et al., 2020; Mustafa & Hayder, 2021). Nevertheless, very few studies have been performed to determine the effectiveness of aquatic plants for removing nutrients from wastewater (Qin et al. 2016; Nayanthara & Bindu, 2017). However, the use of WH and WL is proven superior to other plants as a bio-indicator for phytoremediation in terms of nutrient removal efficiency and sugar recovery for bioethanol production (Dixit et al., 2011; Akinbile and Yusoff, 2012; Nivetha et al., 2016; Qin et al., 2016). Hence, it is needed to evaluate the correlation between sugar content for bioethanol production, biomass growth rate, and nutrients recovered from WH and WL from wastewater.

Thus, the utilization of WH and WL substrates for bioethanol production is explored in this study. Moreover, since the use of WH and WL as substrates for bioethanol production can remove various pollutants in the wastewater and proliferate even under extreme conditions without requiring the land area, several studies have described the potential of WH and WL as substrates for bioethanol production, which consist of significant components such as lignin, cellulose, and hemicellulose. The high percentage of cellulose and hemicellulose in these aquatic plants is the main advantage of their use in bioethanol production (Arefin et al., 2021). Therefore, extensive interest has been emphasized in exploring water hyacinth and water lettuce as a substrate for bioethanol production in the present study and in future studies.

Previously, conventional bioethanol production was reported from various carbonaceous feedstocks such as sugarcane, corn, and starchy (Kassim et al., 2022). However, the main problem in converting carbonaceous feedstocks to bioethanol is the substrate cost and conflict of interest in agricultural products. For example, growing sugarcane for bioethanol production can increase the price of sugar production and tighten food supplies.

Consequently, there has been an emphasis on utilizing lignocellulosic biomass as a source of fermentable sugars for bioethanol production. Since lignocellulosic biomass is readily available, low-cost, and abundant, it could serve as a potential feedstock for bioethanol production (Bilal et al., 2020). Lignocellulosic biomass, which includes agricultural and forestry residues and wastes, also provides a more excellent choice of potential feedstocks that do not conflict with land use for food production and is, in fact, cheaper than conventional bioethanol feedstocks (Krishnan et al., 2020). Hence, in the present study, it is crucial to realize the potential of lignocellulosic biomass of WH and WL, which are rich in fermentable carbohydrates, as they are abundant in nature and cheaper than others in Malaysia.

Generally, bioethanol production relies on the following processes: pretreatment, fermentation, and distillation. However, the major limitation in converting substrates to sugar is the resistance to enzymatic hydrolysis (Li et al., 2018). The low solubilization of the cellulose and hemicellulose components during hydrolysis is another problem in bioethanol production (Arefin et al., 2021). During this process, the sugar is converted for bioethanol production. Therefore, pretreatment is essential for breaking down the lignocellulosic components and improving the cost-effective hydrolysis process (Eshtigahi et al., 2012; Krishnan et al., 2020). To convert lignocellulosic material into bioethanol, using chemicals in the pretreatment method is one of the significant problems in terms of cost and its hazardous effects on the environment. Previous studies have shown that most physical and chemical pretreatments using acid, alkali, microwave, steam explosion, ionizing radiation, or combined processes require specialized instrumentation, consume too much energy, and produce inhibitors that interfere with enzymatic hydrolysis and fermentation (Abo et al., 2019; Krishnan et al., 2020). Hence, biological pretreatment using the

metabolites of a microorganism in nature for ethanol production from biomass is a promising technology due to its numerous advantages, such as its environmentally friendly benefit and economically viable strategy for increasing the enzymatic saccharification rate (Zabed et al., 2019). Several studies have indicated that fungal pretreatment could improve hydrolysis efficiency, resulting in energy consumption limitations (Zabed et al., 2019; Krishnan et al., 2020). Hence, this study explores the suitability of naturally occurring fungal species for the biodegradation process of lignocellulosic plants of water hyacinth and water lettuce as biomass substrates. Since no chemicals are used in this process, no recycling of chemicals is required and, thus, no toxic compounds are released into the environment.

Therefore, biological pretreatment with certain isolated white-rot fungi has been used, which seems promising as an environmentally friendly process. In contrast, no inhibitors are generated during the pretreatment process (Kumar et al., 2020. In this study, *Trichoderma citrinoviride* M3, *Schizophyllum commune* M8, and *Pestalotiopsis* sp. M12 were adopted as an indicator of the biological pretreatment of WH and WL and to be used for the fermentation process to produce bioethanol. The current study focuses on the utilization of biological pretreatment using *Trichoderma citrinoviride*, *Schizophyllum commune*, and *Pestalotiopsis* sp. on WH and WL substrates for bioethanol production, which has not been explored before. However, large-scale use leads to high operating costs as the pretreatment is carried out under sterile conditions, thus increasing the processing costs. Therefore, biological pretreatment is too slow and not recommended for industrial purposes (Chaturvedi and Verma, 2013).

Another bottleneck is the feedback inhibition of cellobiose in the fermentation process after hydrolysis during bioethanol production (Cheng et al., 2015). The most effective method to solve the feedback inhibition problem is simultaneous saccharification and fermentation (SSF) where an enzymatic process hydrolyzes lignocelluloses to sugars and ferments to bioethanol simultaneously, which has been used in many lignocelluloses fermentation systems (Huang et al., 2013; Soares and Gouveia, 2013). To date, the SSF process using *Trichoderma citrinoviride*, *Schizophyllum commune*, and *Pestalotiopsis* sp. with yeast (Saccharomyces

cerevisiae) on WH and WL substrates for bioethanol production has yet to be discovered.

1.3 Research Objectives

The objectives of this study are as follow:

- a) To determine the critical harvesting time of aquatic plants (water hyacinth and water lettuce) based on the correlations among biomass growth, nutrient removal rate, and sugar content from domestic wastewater in the batch system.
- b) To identify the isolated white-rot fungi collected from nature that can degrade the lignocellulosic biomass of water hyacinth and water lettuce.
- c) To determine the efficiency of the lignocellulosic degradation rate of biological pretreatment of aquatic plants (water hyacinth and water lettuce) using isolated white-rot fungi.
- d) To evaluate the efficiency of recombinant isolated white-rot fungi and yeast (Saccharomyces cerevisiae) using simultaneous saccharification and fermentation (SSF) modes from aquatic plants (water hyacinth and water lettuce) to produce bioethanol.

1.4 Scope of Study

The scope of this study entails evaluating the ability of WH and WL to remove nutrient pollutants from Desa Bakti domestic wastewater that lead to the production of a high amount of sugar, which is a crucial parameter for the simultaneous saccharification and fermentation process to produce bioethanol. The two floating aquatic plants (WH and WL) were placed in a tank of a batch system and one tank without plants to serve as a control. The plants were grown in a tank using a batch flow

system. The wastewater samples were collected from the tank, and the plants were harvested from the tank with 3 days of interval till the plants became matured and reached the highest growth of plants. The efficiency of biomass growth and nutrient removal were carried out based on of phosphate (PO₄³⁻), nitrate (NO₃⁻), and nitrite (NO₂) and were compared to determine the best harvested time of WH and WL to produce the highest sugar content for the phase.

12 types of fungi were collected from UTM tropical rainforest. Fungi samples were cultivated and maintained on potato dextrose agar (PDA) and observed at the time interval of 7 days by measuring the diameter of fungal colonies. The screening method was used to test fungi based on speed to choose the preferable fungi for the degradation of aquatic plants. White-rot fungi were identified using 18S rRNA sequence and microscopic observation. The best preferred white-rot fungi were used further for biological pretreatment to degrade lignocellulosic biomass of WH and WL in order to determine the efficiency of isolated white-rot fungi based on the highest sugar content for bioethanol production. The lignocellulosic components and the sugar content of WH and WL were estimated using the Klason Method and Dinitrosalicyclic (DNS) Method, respectively.

The determination of bioethanol production from WH and WL as carried out using the simultaneous saccharification and fermentation (SSF) method. This method conducted the enzymatic hydrolysis and fermentation process simultaneously. The significant advantage of SSF is the released sugar will be instantly consumed by the fermenting agent (yeast), hence indirectly reducing the inhibition of the enzyme due to the low sugar concentration for the bioethanol production inhibition. This study evaluated the feasibility of using monoculture and co-cultures of isolated white-rot fungi and yeast (S. cerevisiae) on WH and WL biomass for bioethanol production. The bioethanol production was determined using Gas Chromatography (GC).

1.5 Significance of the Study

The significance of this study entails the ability of abundant WH and WL as substrates for bioethanol production. Simultaneously, they were used as bioindicators for the phytoremediation process, offering an alternative way to replace the high-cost treatment by utilizing the white-rot fungi in nature to degrade lignocellulosic biomass for bioethanol production. Moreover, in the current study, reducing waste wealth is significant to substitute fossil fuels with more environmentally friendly lignocellulosic biomass. The findings would be beneficial for future investigation in exploring the potential production of bioethanol from phytoremediation technology systems. As such, this study is significant in several ways:

- (a) The aquatic plants that possess the potential for phytoremediation and bioethanol production can ideally be used to redeem wastewater pollutants. After the phytoremediation process, the aquatic plants' biomass can be utilized as a raw material for bioethanol production.
- (b) The ability of fungi to degrade lignocellulose biomass of WH and WL due to their highly effective enzymatic system.
- (c) Biological pretreatment using white-rot fungi in nature for bioethanol production from biomass is a promising technology due to its numerous advantages, such as its environmentally friendly benefits and economically viable strategy for enhancing the enzymatic saccharification rate.
- (d) Studies on the fungal degradation of lignocellulosic material together with yeast as fermenters could yield promising candidate strains that could subsequently be used for bioethanol production.
- (e) Bioethanol can reduce the emissions released when it is burned as a fuel. Therefore, sustainable energy efficiency is essential in addressing the high demand for future low-carbon strategies supported by renewable resources.

REFERENCES

- Abdullah, W.S.W., Osman M., Ab Kadir M.Z.A. and Verayiah R. (2019). The potential and status of renewable energy development in Malaysia. *Energies*. 12(12):2437.
- Abo, B.O., Gao, M., Wang, Y., Wu, C., Ma, H. and Wang, Q. (2019). Lignocellulosic biomass for bioethanol: an overview on pretreatment, hydrolysis and fermentation processes. Reviews on Environmental Health. 34(1):57-68.
- Aftab, M. N., Iqbal, I., Riaz, F., Karadag, A., and Tabatabaei, M. (2019). Different pretreatment methods of lignocellulosic biomass for use in biofuel production. Biomass for Bioenergy: *Recent Trends and Future Challenges*.
- Akinbile C.O. and Yusoff, M. S. (2012). Assessing Water Hyacinth (Eichhornia Crassipes) and Lettuce (Pistia Stratiotes) Effectiveness in Wastewater Treatment. *International Journal of Phytoremediation*. 14(1): 201-211.
- Akinbile, C., Ogunrinde, T.A., Che Bt Man, H. and Aziz, H.A. (2016). Phytroremediation of domestic wastewaters in free water surface constructed wetlands using Azolla pinnata. *International Journal Phytoremediation*. 18(1):54-61.
- Ali, H., Khan, E. and Sajad, M.A. (2013). Phytoremediation of heavy metals concepts and applications. *Chemosphere*. 91:869-881.
- Ali, S., Abbas, Z., Rizwan, M., Zaheer, I.E., Yavas, I., Unay, A., Abdel-DAIM, M.M., Bin-Jumah, M., Hasanuzzaman, M. and Kalderis, D. (2020). Application of floating aquatic plants in phytoremediation of heavy metals polluted water: A review. *Sustainability*. 12(5):19-27.
- Alvira, P., Tomas-Pejo, E., Ballesteros, M. and Negro, M.J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. *Bioresource Technology*. 101(13):4851-4861.
- Amalia, A.A.D., Rahardja, B.S., and Triastuti, R.J (2019). The Use of Water Lettuce (Pistia stratiotesas) as Phytoremediator for Concentration and Deposits of Heavy Metal Lead (Pb) Tilapia (Oreochromis niloticus) Gills. *IOP Conference Series: Earth and Environmental Science*. 236(1):1-7.

- Amornraksa, S., Subsaipin, I., Simasatitkul, L., and Assabumrungat, S. (2020). Systematic design of separation process for bioethanol production from corn stover. *BMC Chemical Engineering*. 2(10): https://doi.org/10.1186/s42480-020-00033-1
- Ansari, A.A., Naeem, M. Gill, S.S. and AlZuaibr, F.M. (2020). "Phytoremediation of contaminated waters: An eco-friendly technology based on aquatic macrophytes application." *Egyptian J. of Aquatic Research*. 46:371-376.
- APHA. (2005). Standard Methods for the Examination of water and waste water. 21st *Edition*. American Public Health Association, Washington, DC. 10-15.
- Aransiola, E., Shittu, T.D., Oyewusi, T.F., and Adetoyese, A.O. (2020). Lignocellulosic Pretreatment Methods for Bioethanol Production. *In book:* Valorization of Biomass to Value-Added Commodities. 135-162.
- Arefin, M. A., Rashid, F. and Islam, A. (2021). A review of biofuel production from floating aquatic plants: an emerging source of bio-renewable energy. *Biofuels Bioproducts Biorefining*. 15(2): 574-591.
- Aswathy, U.S., Sukumaran, R.K., Gottumukkala, L.D., Rajasree, K.P., Singhania, R.T. and Pandey, A. (2010). Bioethanol from water hyacinth biomass: An evaluation of enzymatic saccharification strategy. *Bioresource Technology*. 101(3):925-930.
- Ayodele, B.V., Mustafa, S.I., Mohammad, N., and Shakeri, M. (2021). Long-term energy demand in Malaysia as a function of energy supply: A comparative analysis of non-linear autoregressive exogenous neural networks and multiple non-linear regression models. Energy Strategy Reviews. 38:10-25.
- Balata, M., Balata, H, and Oz, C. (2008). Progress in bioethanol processing. *Prog Energy Combust Sci.* 34:551–73.
- Banch, T.J., Hanafiah, M.M., Alkarkhi, A.F.M., Amr, S.S.A., and Nizam, N.U.M. (2020). Evaluation of different treatment processes for landfill leachate using low-cost agro-industrial materials. *Processes*. 8(111).
- Baruah, J., Nath, B.K., Sharma, R., Kumar, S., Deka, R.C., Baruah, D.B and Kalita, E. (2018). Recent trends in the pretreatment of lignocellulosic biomass for value-added products. *Front. Energy Res.* 6(4).
- Bhattacharya, A., and Kumar, P. (2010). Water hyacinth as a potential biofuel crop. *Electron Journal of Environmental Agricultural Food Chemistry*. 9(1): 112-122.

- Bilal, M., and Iqbal, H.M.N. (2020). Recent Advancements in the Life Cycle Analysis of Lignocellulosic Biomass. *Current Sustainable/Renewable Energy Reports*. 7: 100-107.
- Bolenz, S., Omran, H. and Gierschner, K. (1990). Treatments of water hyacinth tissue to obtain useful products. *J. Biological Wast*. 33:263-274.
- Carman, E. and Crossman, T. (2001). Phytoremediation Chapter 9 in In situ treatment technology. *Environmental Science and Engineering Series*. Lewis Publisher.
- Chandra, M., Kalra, A., Sharma, P.K. and Sangwan, R.S. (2009). Cellulose production by six Trichoderma sp. fermented on medicinal plant processing. Journal of Industrial Microbiology & Biotechnology. 36: 605-609.
- Chaturvedi, V. and Verma, P. (2013). An overview of key pretreatment processes employed for bioconversion of lignocellulosic biomass into biofuels and value added products. *3 Biotech*. 3:415-431.
- Chen, X., Wan, X., Weng, B. and Huang, Q. (2010). Disruption of sugarcane bagasse lignocellulosic structure by means of dilute sulfuric acid pretreatment with microwave-assisted heating. *Applied Energy*. 88(8):2726=2734.
- Cheng, Y.S., Chen, K.Y. and Chou, T.H. (2015). Concurrent calcium peroxide pretreatment and wet storage of water hyacinth for fermentable sugar production. *Bioresource Technology*. 176:267-272.
- Chin, K.L. and H'ng, P.S. (2012). A real story of bioethanol from biomass: Malaysia perspective. *Biomass Now Sustainable Growth and Use*. Available at: https://www.intechopen.com/chapters/44391_(Accessed: 2 September 2021).
- Das, S., Bhattacharya, A., Haldar, S., Ganguly, A., Gu, S., Ting, Y.P. and Chatterjee, P.K. (2015). Optimization of enzymatic saccharification of water hyacinth biomass for bio-ethanol: Comparison between artificial neural network and response surface methodology. Sustainable Materials and Technologies. 3:17-28.
- Das, S., Ghosh, P., Paul, T., Ghosh, U., Pati, B.R. and Mondal, K.C. (2016). Production of bioethanol as useful biofuel through the bioconversion of water hyacinth (Eichhornia crassipes). *3 Biotech*.
- De Stefani, G., Tocchetto, D., Salvat, M. and Borin, M. (2011). Performance of a Floating Treatment Wetland for In-Stream Water Amelioration in NE Italy. *Hydrobiologia*. 674(1): 157-167.

- Dhir, B. (2013). Wastewater treatment by phytoremediation methods. *Wastewater Engineering: Types, Characteristics and Treatment Technologies*.
- Dimos, K., Paschos, T., Louloudi, A., Kalogiannis, K.G., Lappas, A.A., Papayannakos, N., Kekos, D., and Mamma, D. (2019). Effect of Various Pretreatment Methods on Bioethanol Production from Cotton Stalks. *Fermentation*. 5(1): 5. https://doi.org/10.3390/fermentation5010005.
- Ding, C., Wang, X., and Li, M. (2019). Evaluation of six white-rot fungal pretreatments on corn stover for the production of cellulolytic and ligninolytic enzymes, reducing sugars, and ethanol. *Applied Microbiology and Biotechnology*. 103: 5641–5652.
- Dipu, S., Kumar, A.A. and Thanga, V.S.G. (2011). Phytoremediation of dairy effluent by constructed wetland technology. *Environmentalist*. 31:263-278.
- Dixit A., Dixit, S. and Goswamy. D.S. (2011). Process and Plants for Wastewater Remediation A Review. *Science reviews and chemical communication*. 1(1): 71-77.
- Eshtiaghi, M. N., Yoswathana, N., Kuldiloke J. and Ebadi, A. G. (2012). Preliminary study for bioconversion of water hyacinth (Eichhornia crassipes) to bioethanol. *African Journal of Biotechnology*. 11(21):4921-4928.
- Farraji, H., Zaman, N.Q., Ramlah, M. and Farajji, H. (2016). Advantages and disadvantages of phytoremediation: A concise review. *International Journal of Environmental and Technology Sciences*. 2:69-75.
- Fazal, S., Zhang, B. and Mehmood, Q. (2015). Biological treatment of combined industrial wastewater. *Ecol. Eng.* 84:551-558.
- Feng, K., Deng, W., Zhang, Y., Tao, K., Yuan, J., Liu, J., Hugueny, B. (2023). Eutrophication induces function homogenization and traits filtering in chinese lacustrine fish communities. *Science of the Total Environment*. 857.
- Fletcher, J., Willby, N., Oliver, D.M. and Quilliam, R.S. (2020). Chapter 7: Phytoremediation using aquatic plants. *Phytoremediation*. 205-260.
- Galal, T.M., Eid, E.M., Dakhil, M.A. and Hassan, L.M. (2018). Bioaccumulation and rhizolfiltration potential of pistia stratiotes L. for mitigating water pollution in the Egyptian wetlands. *Int. Journal Phytoremediation*. 20: 440-447.
- Ganguly, A., Halder, S., Laha, A., Saha, N., Chatterjee, P.K. and Dey, A. (2013). Effect of Alkali pretreatment on water hyacinth biomass for production of ethanol. *Advanced Chemical Engineering Research*. 2(2)- 40-44.

- González-Gloria, K. D., Rodríguez-Jasso, R. M., Saxena, R., Sindhu, R., Ali, S. S., Singhania, R. R., Ruiz, H.A. (2022). Bubble column bioreactor design and evaluation for bioethanol production using simultaneous saccharification and fermentation strategy from hydrothermally pretreated lignocellulosic biomass. *Biochemical Engineering Journal*, 187.
- Gunnarsson, C.C. and Petersen, C.M. (2007). Water hyacinth as a resource in agriculture and energy production. A literature review. *Waste Management*. 27(1): 117-129.
- Guo, H. Daroch, M., Liu, L., Qiu, G., Geng, S. and Wang, G. (2013). *Bioresource Technology*. 127:422-428.
- Gupta, P., Roy, S. and Mahindrakar, A.B. (2012). Treatment of water using water hyacinth, water lettuce and vetiver grass A review. *Resources and Environment*. 2(5): 202-215.
- Guragain, Y.N., De Coninck, J., Husson, F., Durand, A. and Rakshit, S.K. (2011). Comparison of some new pretreatment methods for second generation bioethanol production from wheat straw and water hyacinth. *Bioresource Technology*. 102(6): 4416-4424.
- Gírio, F.M., Fonseca, C., Carvalheiro, F., Duarte, L.C., Marques, S., and Bogel-Lukasik, R. (2010). Hemicelluloses for fuel ethanol: A review. *Bioresour Technol*. 101(13):4775–800.
- Harun, M.Y., Radiah, A.D., Abidin, Z.Z. and Yunus R. (2011). Effect of physical pretreatment on dilute acid hydrolysis of water hyacinth (Eichhornia crassipes). *Bioresource Technology*. 102(8): 5193-5199.
- Hendriks, A.T.W.M. and Zeeman, G. (2009). Pretreatment to enhance the digestibility of lignocellulosic biomass. *Bioresource Technology*. 100(1):10-18.
- Horisawa, S., Inoue, A., and Yamanaka, Y. (2019). Direct Ethanol Production from Lignocellulosic Materials by Mixed Culture of Wood Rot Fungi Schizophyllum commune, Bjerkandera adusta, and Fomitopsis palustris. *Fermentation*. 5:21.
- Hussain, S.T., Mahmood, T. and Malik, S.A. (2010). Phytoremediation technologies for Ni(++) by water hyacinth. *Afr. J. Biotechnol.* 9:8648-8660.
- Hussnera, A., Heidbuechela, P. and Heiligtagb, S. (2014). Vegetative overwintering and viable seed production explain the establishment of invasive pistia stratiotes in the thermal abnormal. *Aquat. Bot.* 119:28-32.

- Ingle, A.P., Chandel, A. K., Antunes, F. A. F., Rai, M., and Silva, S. S. D. (2019). New trends in application of nanotechnology for the pretreatment of lignocellulosic biomass. *Biofuels Bioproducts & Biorefining*. 13(3): 776-788.
- Isroi, Millati, R., Syamsiah, S., Niklasson, C., Cahyanto, M.N., Lundquist, K., and Taherzadeh, M.J. (2011). Biological pretreatment of lignocelluloses withwhite-rot fungi and its applications: a review. *Bioresource*. 6(4): 5224-5259.
- Jamuna, S. and Noorjahan, C.M. (2009). Treatment of sewage waste water using water hyacinth – Eichhornia sp and its reuse for fish culture. *Toxicology International*. 16(2):103-106.
- Kafle, A., Timilsina, A., Gautam, A., Adhikari, K., Bhattarai, A., and Aryal, N. (2022).
 Phytoremediation: Mechanisms, plant selection and enhancement by natural and synthetic agents. *Environmental Advances*. 8: 100-2023.
- Karmakar, S., Mukherjee, J., and Mukherjee, S. (2018). Biosorption of fluoride by water lettuce (Pistia stratiotes) from contaminated water. *International Journal of Environmental Science and Technology*. 15(4):801–810.
- Kinidi, L. and Salleh, S. (2017). Phytoremediation of nitrogen as green chemistry for wastewater treatment system. *International Journal of Chemical Engineering*.
- Kumar, A., Singh, and Ghosh, s. (2009). Bioconversion of lignocellulosic fraction of water hyacinth (Eichhornia crassipes) hemicellulose acid hydrolysate to ethanol by Pichia stiptis. *Bioresource Technology*. 100(13):3293-3297.
- Kumar, D. and Murthy, G.S. (2011). Impact of pretreatment and downstream processing technologies on economics and energy in cellulosic ethanol production. *Biotechnology for Biofuels*. 4(1): 27.
- Lamichhane, G., Acharya, A., Poudel, D.K., Aryal, B., Gyawali, N. and Niraula, P. (2020). Recent advances in bioethanol production from lignocellulosic biomass. *International Journal of Green Energy*. 18(7):731-744.
- Li, X., Shi, Y., Kong, W., Wei, J., Song, W. and Wang S. (2022). Improving enzymatic hydrolysis of lignocellulosic biomass by bio-coordinated physicochemical pretreatment A review. *Energy Reports*. 696-709.
- Lien, Y. H., Liu, F. Y., Chen, J. N., Huang, Y. S., Wei, Y. H., Yu, C., and Shu, C. C. (2019). Using the Juice of Water Lettuce (Pistia stratiotes) as Culture Medium to Increase the Cell Density and the Production of Microbial Lipid. *Biotechnology and Bioprocess Engineering*. 24(2):395–400.

- Lin, Y.L. and Li, B.K. (2016). Removal of pharmaceuticals and personal care products by Eichhornia crassipes and pistia stratiotes. *Journal of Taiwan Inst. Chem. Eng.* 58:318-323.
- Lin, Y. and Tanaka, S. (2006). Ethanol fermentation from biomass resources: current state and prospect. *Appl. Microbiology Biotechnology*. 69:627-642.
- Lu, Q., He, Z.L., Graetz, D.A., Stofella, P.J. and Yang, X. (2011). Uptake and distribution of metals by water lettuce (Pistia stratiotes L.). *Environ. Sci. Pollut. Res.* 18(6):978-986.
- Ma, F., Yang, N., Xu, C., Yu, H., Wu, J. and Zhang, X. (2010). Combination of biological pretreatment with mild acid pretreatment for enzymatic hydrolysis and ethanol production from water hyacinth. *Bioresource Technology*. 101(24):9600-9604.
- Magar, R.B., Honnutgi, A. and Khan, A.N. (2017). Waste Water Treatment Using Water Hyacinth" 32ndIndian Engineering Congress, The Institution of Engineers (India).
- Mahamadi, C. (2011). Water hyacinth as a biosorbent: A review. *Afr. J. Environ. Sci. Technol.* 5:1137-1145.
- Mahmood, H., Moniruzzaman, M., Iqbal, T. and Khan, M.J. (2019). Recent advances in the pretreatment of lignocellulosic biomass for biofuels and value-added products. *Current opinion in green and sustainable chemistry*. 20:18-24.
- Maurya, D.P., Singla, A. and Negi, S. (2015). An overview of key pretreatment processes for biological conversion of lignocellulosic biomass to bioethanol. *3 Biotech.* 5:597-609.
- Mazaheri, D., Orooji, Y., Mazaheri, M., Moghaddam, M.S., and Karimi-Maleh, K. (2021). Bioethanol production from pomegranate peel by simultaneous saccharification and fermentation process. *Biomass Conversion and Biorefinery*.
- Menon, V. and Rao, M. (2012). Trends in bioconversion of lignocelluloses: Biofuels, platform chemicals & biorefinery concept. *Progress in Energy and Combustion Science*. 38:522-550.
- Miller, G.L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*. 31(3): 426-428.
- Mishima, D., Tateda, M., Ike, M. and Fujita, M. (2006). Comparative study on chemical pretreatments to accelerate enzymatic hydrolysis of aquatic

- macrophyte biomass used in water purification processes. *Bioresource Technology*. 97:2166-2172.
- Mishima, D., Kuniki, M., Sci, K. Soda, S., Ike, M. and Fujita, M. (2008). Ethanol production from candidate energy crops: water hyacinth (Eichhornia crassipes) and water lettuce (Pistia stratiotes L.). *Bioresource Technology*. 2495-2500.
- Mkandawire, M. and Dudel. E.G. (2007). Are Lemna spp. effective phytoremediation agents. *Bioremediat. Biodivers. Bioavailab.* 1:56-71.
- Mulling, B.T.M., Soeter, A.M., van der Geest, H.G. and Admiraal, W. (2014). Changes in the Planktonic Microbial Community during Residence in a Surface Flow Constructed Wetland Used for Tertiary Wastewater Treatment. *Science Total Environmental*. 466-467: 881-887.
- Mustafa, H.M. and Hayder. G. (2020). Recent studies on application of aquatic weed plants in phytoremediation of wastewater: A review article. *Ain Shams Engineering Journal*. 12:355-365.
- Mustafa, H.M. and Hayder, G. (2021). Evaluation of water lettuce, giant salvinia and water hyacinth systems in phytoremediation of domestic wastewater. *H2Open Journal*. 4(1): 167-181.
- Nayanthara, O.S. and Bindu. A.G. (2017). Effectiveness of Water Hyacinth and Water Lettuce for the Treatment of Greywater A Review. *International Journal of Innovative Research in Science and Engineering*. 3(1): 349-355.
- Nidheesh, P. V., Khan, F. M., Kadier, A., Akansha, J., Bote, M. E., and Mousazadeh, M. (2022). Removal of nutrients and other emerging inorganic contaminants from water and wastewater by electrocoagulation process. *Chemosphere*. 307(P2): 135756.
- Nigam, J.N. (2002). Bioconversion of water hyacinth (Eichhornia crassipes) hemicellulose acid hydrolysate to motor fuel ethanol by xylose-fermenting yeast. *Journal Biotechnology*. 97:107-116.
- Nivetha, C., Subraja, S., Sowmya, R. and Induja, N.M. (2016). Water lettuce for removal of nitrogen and phosphate from sewage. *Journal of Mechanical and Civil Engineering*. 13(2):104-107.
- Nizam, N.U.M., Hanafiah, M.M., Noor, I.M. and Karim. H.IA. (2020). Efficiency of five selected aquatic plants in phytoremediation of aquaculture wastewater. *Applied Science*. 10(8):2712.

- Priya, E.S. and Selvan, P.S. (2014). Water hyacinth (Eichhornia crassipes) An efficient and economic adsorbent for textile effluent treatment A review. *Arab J. Chem.* 1-11.
- Qin, H., Zhang, Z., Liu, M., Liu, H., Wang, Y., Wen, X., Zhang, Y. and Yan, S. (2016). Site test of phytoremediation of an open pond contaminated with domestic sewage using water hyacinth and water lettuce. *Ecological Engineering*. 95(1):753-762.
- Qv, M., Dai, D., Liu, D., Wu, Q., Tang, C., Li, S., and Zhu, L. (2023). Towards advanced nutrient removal by microalgae-bacteria symbiosis system for wastewater treatment. *Bioresource Technology*: 370(January): 128574.
- Rahman, M.A. and Hasegawa, H. (2011). Aquatic arsenic: Phytoremediation using floating macrophytes. *Chemosphere*. 83:633-646.
- Rezania, S., Ponraj, M., Md Din, M.F., Songip, A.R. and Chelliapan, S. (2014). True potential of aquatic plants (Eichhornia crassipes, Pistia stratiotes) in the production of bio-ethanol. *International Young Water Profession Conference*. 267-277.
- Rezania, S., Ponraj, M., Talaiekhozani, A., Mohamad, S.E., Md Din, M.F., Taib, S.
 M., Sabbagh, F. and Sairan., F.M. (2015). Perspectives of Phytoremediation
 Using Water Hyacinth for Removal of Heavy Metals, Organic and Inorganic
 Pollutants in Wastewater. *Journal of Environmental Management*. 163: 125-133.
- Roongtanakiat, N., Tangruangkiat, S. and Meesat, R. (2007). Utilization of vetiver grass (vetiveria zizanioides) for removal heavy metals from industrial wastewater. *ScienceAsia*. 33:397-403.
- Sabeena H., Ngadia, N., Noora, Z.Z., Raheema, A.B., Aqouillalc, F., Mohammeda, A.A. and Abdulkarima, B.I. (2018). Characteristics of the Effluent Wastewater in Sewage Treatment Plants of Malaysian Urban Areas. *The Italian Association of Chemical Engineering*. 63: 691-696.
- Shamsuddin, A.H. (2012). Development of renewable energy in Malaysia strategic initiatives for carbon reduction in the power generation sector. *Proc. Eng.* 49:384-391.
- Shuvaeva, O.V., Belchenko, L.A. and Romanova, T.E. (2013). Studies on cadmium accumulation by some selected floating macrophytes. Int. J. *Phytoremediation*. 15:979-990.

- Sindhu, R., Binod, P. and Pandey, A. (2016). Biological pretreatment of lignocellulosic biomass An overview. *Bioresource Technology*. 199: 76-82.
- Singh, D., Tiwari, A. and Gupta, R. (2012). Phytoremediation of lead from wastewater using aquatic plants. *J. Agric. Technol.* 8: 1-11.
- Singh, J. K., Chaurasia, B., Dubey, A., Noguera, A.M.F., Gupta, A., Kothari, R., Upadhyaya, C.P., Kumar, A., Hashem, A., Alqarawi, A.A., and Abd Allah, E.F. (2020). Biological Characterization and Instrumental Analytical Comparison of Two Biorefining Pretreatments for Water Hyacinth (Eichhornia crassipes) Biomass Hydrolysis. *Sustainability*. 13(1):245.
- Singh, S., Kumar, A., Sivakumar, N., and Verma, J. P. (2019). Deconstruction of lignocellulosic biomass for bioethanol production: *Recent advances and future prospects*. Fuel. 327: 125109.
- Singh, Y.D. and Satapathy, K.B. (2018). Conversion of lignocellulosic biomass to bioethanol: An overview with a focus on pretreatment. *International Journal of Engineering and Technologies*. 15:17-43.
- Soda, S., Mishima, D., Inoue, D. and Ike, M. (2013). A co-beneficial system using aquatic plants: bioethanol production from free-floating aquatic plants used for water purification. *Water Sci. Technol.* 67(11):2637-2644.
- Tran, T.T.A., Le, T.K.P., Mai, T.P., and and Nguyen, D.Q. (2019). Bioethanol Production from Lignocellulosic Biomass. *Alcohol Fuels*. DOI: 10.5772/intechopen.86437.
- Tye, Y.Y., Lee, K.T., Abdullah, W.N.W. and Leh, C.P. (2011). Second-generation bioethanol as a sustainable energy source in Malaysia transportation sector: status, potential and future prospects. *Renewable and Sustainable Energy Reviews*. 15:4521-4536.
- Waghunde, R.R. and Patil, R.K. (2010). Physiological studies of the alternaria fruit rot (Alternaria Alternata) of aonla. Journal of Plant Disease Sciences. 5(1): 73-75.
- Walsh, D.C. and Maestro, M. (2014). Evaluation of intraguild interactions between two species of insect herbivores on Pistia stratiotes. *Biological Control*. 76: 74-78.
- Watson, S.B., Miller, C., Arhonditsis, G., Boyer, G.L., Carmichael, W., Charlton, M.N., Confesor, R., Depew, D.C., Hook, T.O., Ludsin, S.A., Matisoff, G., McElmurry, Murray, S.P., Richards, M.W., Rao, R.P., Steffen, M.M. and

- Wilhelm, S.W. (2016). The Re-eutrophication of Lake Erie: Harmful Algal Blooms and Hypoxia. *Harmful Algae*. 56: 44-66.
- Whangchai, K., Inta, W., Unpaprom, Y., Bhuyar, P., Adoonsook, D. and Ramaraj, R. (2021). Comparative analysis of fresh and dry free-floating aquatic plant Pistia stratiotes via chemical pretreatment for second-generation (2G) bioethanol production. *Bioresource Technology Reports*. 14:100651.
- Wyman, C.E., Cai, C.M. Kumar, R. (2019). Bioethanol from lignocellulosic biomass. *In:Meyers R. (eds) Encyclopedia of Sustainability Science and Technology*. Available at: https://doi.org/10.1007/987-1-4939-2493-6_521-3 (Accessed: 28 August 2021).
- Xia, H. and Ma, X. (2006). Phytoremediation of ethion by water hyacinth (Eichhornia crassipes) from water. *Bioresource Technology*. 97:1050-1054.
- Yoosin, S. and Sorapipatana, C. (2007). A study of ethanol production cost for gasoline substitution in Thailand and its competitiveness. *Thammasat International Journal of Science and Technology*. 12:69-80.
- Zhang, Q., Weng, C., Huang, H., Achal, V. and Wang, D. (2015). Optimization of bioethanol production using whole plant of water hyacinth as substrate in simultaneous saccharification and fermentation process. *Frontiers in Microbiology*. 6:1411.
- Zhao, H., Jones, C.L., Baker, G.A., Xia, S., Olubajo, O. and Person, V.N. (2009).
 Regenerating cellulose from ionic liquids for an accelarated enzymatic hydrolysis. *Journal of Biotechnology*. 139(1): 47-54

LIST OF PUBLICATIONS

Journal

- Zainuddin, N.A., Md Din, M.F., Ponraj, M., Hassim, M.H., Mat Taib, S., Sanjaya, E.H. (2022). Biodegradation Efficiency of Fungi for Lignocellulosic Biomass of Water Hyacinth (Eicchornia crassipes). *International Journal of Advanced Research in Science, Communication and Technology*. 2(8): 46-56.
- Zainuddin, N.A., Md Din, M.F., Nuid, M., Abdul Halim, K., Salim, N.A.A., Elias, S.H., and Mat Lazim, Z. (2022). The Phytoremediation using Water Hyacinth and Water Lettuce: Correlation between Sugar Content, Biomass Growth Rate, Nutrients. *Jurnal Kejuruteraan*. 34(5): 915-924.
- 3. Krishnan, S., Ahmad, M.F., **Zainuddin, N.A.,** Md Din, M.F., Rezania, S., Chelliapan, S., Mat Taib, S., Nasrullah, M. And Wahid, Z.A. (2020). Bioethanol production from lignocellulosic biomass (water hyacinth): a biofuel alternative. *Bioreactors: Sustainable Design and Industrial Applications in Mitigation of GHG Emissions*. 123-143.

Book Chapter

- 1. **Zainuddin, N.A.,** Md Din, M.F. and Aminudin, E. (2020). Water hyacinth (eichhornia crassipes) and water lettuce (pistia stratiotes L.) as nutrient uptake from domestic wastewater. *Issues and Technologies in Water Contaminants*. UTM Press. Universiti Teknologi Malaysia.
- Zainuddin, N.A., Md Din, M.F., Hassim, M.H. and Mat Taib, S. (2022). Application of floating aquatic macrophytes for Phytoremediation of wastewater. Water and Wastewater Treatment Technology. UTM Press. Universiti Teknologi Malaysia.