

ACID RED 27 DECOLOURISATION AND SIMULTANEOUS ELECTRICITY
GENERATIONS BY MIXED BACTERIAL CULTURES IN MICROBIAL FUEL
CELL

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DEDICATION

I would like to dedicate my deepest appreciation to

my mom Baizurah binti Abdul Kadir and my dad Abdul Aziz bin Omar for endless support and word of motivation that give me great courage;

my beloved husband Wan Mohd Irwan Syazwan thank you for every single moment you spent with me just gives me assurance, feeling secure and appreciated;

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ABSTRACT

Azo dyes are widely used in industries although poorly biodegradable and highly toxic. Poor existing wastewater treatment caused dye residues in industrial effluent to be discharged into neighbouring water bodies causing major distress to the environment. This study was carried out to identify the potential of using mixed bacterial culture system for efficient biodegradation of azo dye in single batch system and for potential electricity generation in microbial fuel cell (MFC). This study used single and mixed bacterial consortium in batch shake flask experiment to decolourize Acid Red azo dye (AR27) and for electricity generation in a dual-chamber MFC. Four bacterial strains coded as MF1, B2, ZL and CO were used. Morphological, biochemical and 16S rDNA sequence analyses of these bacteria were carried out. Phylogenetic analysis successfully confirmed the identity of strains MF1 and B2 as *Klebsiella pneumoniae* (MF1 and B2), CO as *Bacillus cereus*. However, ZL identification was inconclusive. Flask experiment of single strain showed that all single strains were able to degrade AR27 dye, where strain CO displayed 2 to 4-fold higher rate of dye removal compared to other strains. Compatibility test successfully showed that strains CO, MF1 and ZL were able to grow without antagonism effect whereas CO and B2 were competing with each other. Selection of the best mixed bacterial culture based on AR27 dye degradation showed that combination of strains CO, MF1 and ZL produced the best dye decolourisation rate and Chemical Oxygen Demand (COD) removal. Optimization study revealed that mixed culture of CO+MF1+ZL performed the best AR27 dye degradation in CDM medium when adjusted to pH 5.0-7.0, supplemented with 3 g/L yeast extract, 100 mg/L AR27 dye and incubated under static condition at 32 – 37°C. Under optimal conditions, 99% of 100 mg/L AR27 was removed at the rate of 4.21 mg/h. Sequential facultative anaerobic-aerobic condition demonstrated that the COD removal percentage increased from 57% to 69% when aerobic condition was introduced to the system. However, the amount of Total Polyphenolic Content (TPP) removed showed no significant difference. Assessment of potential electricity generation by CO+MF1+ZL mixed culture during AR27 dye removal in dual-chamber MCF demonstrated that the maximum generated open circuit voltage (OCV) was 567.7mV. Closed circuit voltage (CCV) across 5000 Ω external resistance generated was 39.7mV with maximum power density and current density of 0.26 mW/m² and 6.6 mA/m², respectively. In the application of mixed culture for dye removal in real textile wastewater, the voltage generated dropped to 232.38 mV for OCV and 13.9 mV for CCV with maximum power and current density generated 0.03 mW/m² and 2.3 mA/m², respectively. This was due to high pH value of textile wastewater. When textile wastewater was adjusted to pH 7, the voltage generated improved to 489.9 mV for OCV and 32.9 mV for CCV with maximum power and current density generated 0.18 mW/m² and 5.78 mA/m², respectively. This study showed that mixed culture of CO+MF1+ZL has a good potential for treating wastewater containing azo dye for simultaneous decolourization and electricity generation.

ABSTRAK

Pewarna Azo digunakan secara meluas dalam industri walaupun tidak terbiodegradasikan dan sangat toksik. Rawatan air kumbahan sedia ada yang tidak sempurna menyebabkan residu pewarna dalam efluen perindustrian dilepaskan ke dalam sistem saliran air menyebabkan kesan pencemaran buruk kepada alam sekitar. Kajian ini dijalankan untuk mengenal pasti potensi menggunakan campuran bakteria untuk biodegradasi pewarna azo yang berkesan dalam sistem kelompok tunggal dan potensi penjana tenaga elektrik dalam sistem sel bahan api mikrob (MFC). Kajian ini menggunakan goncangan bakteria tunggal dan campuran dalam kelompok, kelalang goncang untuk penyahwarna pewarna azo *Acid Red* (AR27) dan penjana tenaga elektrik di dalam sel bahan api mikrob (MFC) dwi-ruang. Empat jenis bakteria yang diberi kod MF1, B2, ZL dan CO digunakan. Analisis morfologi, biokimia dan jujukan 16S rDNA bagi bakteria ini telah dijalankan. Analisis filogeni berjaya mengesahkan identiti strain MF1 dan B2 sebagai *Klebsiella pneumoniae* (MF1 dan B2) dan CO sebagai *Bacillus cereus*. Walau bagaimanapun, pengenalan strain ZL tidak dapat disimpulkan. Eksperimen kelalang bagi bakteria tunggal menunjukkan bahawa semua strain tunggal mampu untuk menyahwarna AR27, di mana strain CO menunjukkan kadar penyahwarna 2 hingga 4 kali ganda lebih tinggi berbanding dengan strain lain. Ujian keserasian berjaya menunjukkan bahawa strain CO, MF1 dan ZL dapat membiak tanpa kesan antagonisme tetapi CO dan B2 bersaing dengan satu sama lain. Pemilihan campuran bakteria terbaik berdasarkan degradasi pewarna AR27 menunjukkan kombinasi strain CO, MF1 dan ZL mempunyai kadar penyahwarna pewarna dan penyingkiran Permintaan Oksigen Kimia (COD) terbaik. Kajian pengoptimuman kultur campuran CO + MF1 + ZL menunjukkan penyahwarna AR27 terbaik dalam medium CDM yang diselaraskan kepada pH 5.0-7.0, ditambah dengan 100 mg / L pewarna AR27 dan 3 g/L ekstrak yis yang dieram di bawah keadaan statik pada suhu 32-37 °. Di bawah keadaan yang optimum, 99% daripada 100 mg / L AR27 dinyahwarnakan pada kadar 4.21 mg/j. Keadaan anaerobik-aerobik berturutan menunjukkan bahawa peratus jumlah penyingkiran COD meningkat daripada 57% kepada 69% namun jumlah kandungan Pengeluaran Polifenolik (TPP) (22 mg/mL) tidak menunjukkan perbezaan yang ketara. Penaksiran potensi penjana tenaga elektrik oleh campuran CO + MF1 + ZL semasa penyahwarna AR27 dalam MCF dwi-ruang menunjukkan bahawa voltan litar terbuka maksimum (OCV) terjana adalah 567.7mV. Voltan litar tertutup (CCV) merentasi 5000 Ω rintangan luar yang dijana ialah 39.7mV dengan ketumpatan kuasa maksimum dan kepadatan arus 0.26 mW/m² dan 6.6 mA / m². Aplikasi campuran bakteria untuk penyahwarna pewarna dalam sisa kumbahan tekstil, voltan yang dijana menurun kepada 232.38 mV untuk OCV dan 13.9 mV untuk CCV dengan kuasa maksimum dan ketumpatan arus masing-masing menghasilkan 0.03 mW / m² dan 2.3 mA / m². Apabila sisa kumbahan tekstil diselaraskan kepada pH 7, voltan yang dihasilkan meningkat kepada 489.9 mV untuk OCV dan 32.9 mV untuk CCV dengan kuasa maksimum dan ketumpatan arus yang dihasilkan 0.18 mW/m² dan 5.78 mA/m². Kajian ini memperlihatkan bahawa konsortium bakteria CO + MF1 + ZL mempunyai potensi yang baik untuk merawat air kumbahan mengandungi pewarna azo bagi penyahwarna dan penjana tenaga elektrik secara serentak.

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LIST OF ABBREVIATIONS / SYMBOLS

%	-	Percent
μA	-	Microampere
μL	-	Microlitre
μV	-	Microvolt
A	-	Surface area
Abs	-	Absorbance
Abs _{600nm}	-	Absorbance at the wavelength of 600 nm
AR-27	-	Acid Red 27
CDM	-	Chemically Defined Medium
C/N	-	Carbon per nitrogen
CCV	-	Close circuit voltage
Cm	-	Centimeter
COD	-	Chemical oxygen demand
CuSO ₄	-	Copper sulphate
DET	-	Direct electron transfer
e ⁻	-	Electrons
FTIR	-	Fourier Transform Infrared spectroscopy
G	-	Gram
g/L	-	Gram per liters
h	-	Hours
H ⁺	-	Hydrogen ion
H ₂ SO ₄	-	Sulphuric acid
I	-	Current
J	-	Current density
K ₂ Cr ₂ O ₇	-	Potassium dichromate
K ₂ HPO ₄	-	Dipotassium hydrogen phosphate

L	-	Litre
M	-	Molar
mA	-	Miliampere
MFC	-	Microbial fuel cell
Mg/mL	-	Milligram per millilitre
mL	-	Milliliters
mM	-	Millimolar
mm	-	Millimeter
mV	-	Millivolt
mV/s	-	Millivolt per second
mW/m ²	-	Milliwatts per meter square
Na ₂ CO ₃	-	Sodium carbonate
°C	-	Degree Celsius
OCV	-	Open circuit voltage
P	-	Power density
PEM	-	Proton exchange membrane
R _{ext}	-	External resistance
TPP	-	Total polyphenol
UV-Vis	-	Ultra violet visible
V	-	Volt
v/v	-	Volume to volume
w/v	-	Weight to volume
λ	-	Wavelength
λ _{max}	-	Maximum waveleght
Ω	-	Ohm
λ _{max}	-	Maximum waveleght
Ω	-	Ohm

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CHAPTER 1

INTRODUCTION

1.1 Background of study

Textile industry generates a large volume of wastewater containing mixtures of azo dyes as the main source of pollutant (Solis *et al.*, 2012). Azo dye is a synthetic dye that is extensively used in the food, cosmetic, textile, pharmaceutical and paper industries due to its colour varieties and fastness in production as compared to natural dyes (Chang and Lin, 2001; Carneiro *et al.*, 2007). Around 60 – 70 % of total synthetic dye that produced annually consists of azo dyes (Mc Mullan *et al.*, 2001). Azo dye was characterized by the presence of double bonds (-N=N-) are the largest and most versatile class of dyes and are the most common chromophore in azo dyes (Stolz, 2001). During the dye processing approach 2 % of dyes are directly discharged into aqueous effluent (Pearce *et al.*, 2003) and is estimated that about 2 – 50 % of various applied dyes can be lost in the effluent during the textile colouring process (Selvakumar *et al.*, 2013). Even at the low concentration the azo dye that dissolved in the water stream can cause it to become highly coloured. In a clean river water, dye concentration as low as 0.005 ppm are visible (Saratale *et al.*, 2011)

O' Neill *et al.*, (1999) reported that textile processing wastewater usually contains dye concentration between 10 - 200 mg/L. When the coloured wastewater flow into the river it will cause the intense colour to prevent the absorbance of light entering the water thus, greatly reducing photosynthesis of aquatic flora (Slokar *et al.*, 1998). Rai *et al.*, (2005) states that such dyes are defined as colored substances that give a permanent colour when it was applied to fibers and resist fading upon exposure to sweat, light, water and many chemicals, including oxidizing agents and microbial attack. In the intestinal bacteria, liver cells and skin surface micro flora contain azoreductase that would reduce azo bonds in azo dyes to colourless aromatic amines (Xu *et al.*, 2007). Most azo dyes are either inert or toxic but as the azo bond

is reduced it form aromatic amines that are toxic, mutagenic and carcinogenic (Kodam *et al.*, 2006). From the study conducted by several researchers related to the risk assessment of aromatic amines, it was proven that aromatic amines are carcinogenic to the human body (IARC, 2012).

Finding an alternative way to treat azo dyes contaminant particularly for small scale textile industries are major concern since textile dye effluent contained azo dye and the derived metabolites from the improper discharge is aesthetically unpleasant (Vandevivere *et al.*, 1998). These leads to a reduction in sunlight penetration, which in turn decreases photosynthetic activity that cause from the increase in dissolved oxygen concentration and water quality and consequently, acute toxic effects on aquatic flora and fauna, causing severe environmental problems worldwide (Vandevivere *et al.*, 1998). Pang and Abdullah, (2013) reported that in Malaysia textile industry is one of the fastest growing industries and significantly contributes to the economic growth and consequently, produces high discharge rate of wastewater with high load of contaminants that caused main source of water pollution when the release of dyes into the environment during textile fiber dyeing and finishing processes. The most common treatment plan used are conventional treatment systems such as biological treatment alone or physicochemical treatments followed by a biological treatment due to their simplicity and lower treatment cost as compared to some physical and chemical methods (Pang and Abdullah, 2013; You and Teng, 2009).

Biological treatment use bacterial species that are capable of decolourising, and completely mineralise many reactive dyes under certain conditions (Kumar *et al.*, 2012). The degradation process of bacteria is often stimulated under static/anaerobic conditions by an enzymatic transformation reaction for dye decolourisation which results in the formation of aromatic amines (Kumar *et al.*, 2012). The aromatic amines produced are then further oxidised and mineralised to form a simpler non-toxic by-product under aerobic conditions (Chan *et al.* 2012c; Kumar *et al.*, 2012). Decolourization of azo dye by microbial fuel cell has received attention as new technology to treat textile wastewater since it derives bioelectricity generations simultaneously with wastewater treatment (Sun *et al.*, 2009; Chen *et al.*,

2010; Liu *et al.*, 2011a). In a microbial fuel cell electricity is produced through the reaction between the fuel (anode) and oxidant (cathode) in the presence of an electrolyte (Evans *et al.*, 2012). In the anode compartment, active microorganism will oxidise the organic co-substrate that eventually generate electron (Logan *et al.*, 2006; Lai *et al.*, 2017). The current generation is produced as the electrons are transferred to the eventually generate electron (Logan *et al.*, 2006; Lai *et al.*, 2017). The cathode and the anode compartment was separated by a proton exchanges membrane to allow proton migration (Logan *et al.*, 2006; Solanki *et al.*, 2013; Lai *et al.*, 2017).

1.2 Problem Statement

Current situation in Malaysia shows that amount of scheduled waste generated by textile industry increased sharply. However, there is still no centralized system for the textile industry factory to treats the wastewater due to the cost and time consuming process. Meanwhile, high cost needed to supply high quality water for dyeing and finishing process. Due to the high solubility of Azo dye in water and poor wastewater treatment system, the dye residues residing in the effluent discharged from the industry to the neighbouring water bodies is causing major distress to the environment. Findings an economic ways to treat the wastewater effluent was crucial to prevent water pollution from getting worse.

Biological process using bacteria provide a low-cost and an efficient ways for colour removal and degradation of dye. However, several problem need to overcome such as the concentration of co-substrate use by azo degrading bacteria must be sufficient to treat the textile wastewater in an ideal and economical operating system. Other than that, the practically of the possible implementations the optimized system to the real textile wastewater should also be considered. Therefore, it is important to explore the prospect of effective dye degradation by bacterial culture so that these method can be sufficient to treat the textile wastewater in an ideal operating system and subsequently provide an economic ways to treat water pollution problem.

1.3 Significance of study

This study focused on developing bacterial mixed culture capable of removal model azo dye AR-27 and further application to treat Batik textile wastewater in glass fabricated microbial fuel cell with proton exchange membrane (nylon membrane). The main idea for the study was to characterize and identified four unknown strain on their ability to remove azo dye AR-27. Then, developing bacterial mixed culture that are capable of degrading azo dye AR-27. The best combination of the mixed culture bacteria in removal and degrading azo dye AR-27 will be selected for further optimization on the physicochemical conditions. Then, the optimized combination will be applied into the synthetic wastewater under sequential facultative anaerobic-aerobic MFC system to assess the potential of electricity generation by the selected bacterial combination under optimized condition. Finally, the selected mixed bacterial culture was introduced to the real batik textile wastewater to evaluate the potential of electricity generations. This study could provide new idea for the current treatment of textile effluent and as an alternative green energy in the future.

1.4 Objectives

This study was performed to assess the potential of azo dye AR-27 removal and simultaneous electricity generation in microbial fuel cell by selected bacterial combination. Hence, these objectives were established in the research to achieve the research aim:

- (a) To characterize and identify of the ability of pure bacterial strain to decolourise AR-27 dye
- (b) To select and optimization of bacterial mixed culture capable of degrading azo dye AR-27
- (c) To assess the potential of electricity generation by selected bacterial mixed culture under optimized conditions in a Microbial Fuel Cell.

1.5 Scopes

The main scope of this research was to assess the potential of azo dye AR-27 removal and simultaneous electricity generation in microbial fuel cell with nylon membrane by selected bacterial culture combination. Hence, these scope were established to accomplish the azo dye decolourization and the electricity generations.

This study characterized the capability of pure bacterial strain to remove azo dye AR-27 by using CDM medium supplemented with 1.0 g/L glycerol and AR-27 dye as the model dye under facultative anaerobic conditions. The identification was performed by using 16S rRNA Gene Sequence Analysis and biochemical test. Then, the phylogenetic tree was constructed using MEGA6 application.

For the development of combination from the pure bacterial strain, the compatibility test was conducted to study the antagonistic effect between the strains to grow as mixed culture without inhibiting each other. Then 7 type of combination was developed and investigation of the ability for azo dye removal and degradation in CDM medium supplemented with 1.0 g/L glycerol and AR-27 dye as the model dye under facultative anaerobic conditions. The selected combination was then optimized under several physicochemical such as effect of carbon and nitrogen source, effect of shaking vs static, effect of temperature, effect of initial dye concentrations and effect of pH. Next, the selected bacterial combination with optimized condition was inoculated into a fabricated microbial fuel cell with proton exchanges membrane (nylon membrane) to study the potential of electricity generations.

REFERENCES

- Abubacker, M.N., and Ayesha, A. (2011). Decolourization of Azo-Dyes by Bacterial Consortium. *Biosciences Biotechnology Research Asia*, Vol. 8(2), p. 741-746.
- Adedayo, O., Javadpour, S., Taylor, C., Anderson, W.A. and Moo-Young, M. (2004). Decolourization and detoxification of methyl red by aerobic bacteria from a wastewater treatment plant. *World Journal of Microbiology and Biotechnology*, 20: 545 - 550.
- Aelterman, P., Rabaey, K. and Verstraete, W. (2006). Continuous electricity generation at high voltages and currents using stacked microbial fuel cells. *Environmental Science Technology*, 40(10):3388–3394.
- Ali, Y., and Ameta, A. (2013). Degradation and decolouration of amaranth dye by photo-fenton and fenton reagents: a comparative study. *International Journal of Chemistry Science*, 11(3): 1277-1285.
- Ali, R., Daut, I., and Taib, S. (2012). A Review on Existing and Future Energy Sources for Electrical Power Generation in Malaysia. *Renewable and Sustainable Energy Reviews*, 16(6), 4047-4055.
- Anjaneya, O., Shrishailnath, S. S., Guruprasad, K., Nayak, A. S., Mashetty, S. B., and Karegoudar, T. B. (2013). Decolourization of Amaranth dye by bacterial biofilm in batch and continuous packed bed bioreactor. *International Biodeterioration and Biodegradation*, 79, 64-72.
- Anjaneya, O., Souche, S. Y., Santoshkumar, M., and Karegoudar, T. B. (2011). Decolorization of sulfonated azo dye Metanil Yellow by newly isolated bacterial strains: *Bacillus* sp. strain AK1 and *Lysinibacillus* sp. strain AK2. *Journal of Hazardous Materials*, 190(1), 351-358.
- Asgher, M., Azim, N. and Bhatti, H.N. (2009). Decolorization of practical textile industry effluents by white rot fungus *Coriolus versicolor* IBL-04. *Biochemical Engineering Journal*, 47:61-5.
- Asad, S., Amoozegar, M.A., Pourbabae, A.A., Sarbolouki, M.N. and Dastgheib, S.M.M. (2007). Decolorization of textile azo dyes by newly isolated halophilic and halotolerant bacteria. *Bioresources Technology*, 98: 2082-2088.

- Aly, M.M., Khalil, S. and Metwaly, A. (2014). Isolation and Molecular Identification of Klebsiella Microbe Isolated from Chicks. *Alexandria Journal of Veterinary Sciences*, 43: 97-103, 1110-2407.
- Ayed, L., Cheriaa, J., Laadhari, N., Cheref, A. and Bakhrouf, A. (2009). Biodegradation of crystal violet by an isolated *Bacillus* sp. *Annals of Microbiology*, 59(2): 267 - 272.
- Banat, I.M., Nigam, P., Singh, D. and Marchant, R. (1996). Microbial decolorization of textile-dye-containing effluents: A review. *Bioresource Technology*, 217 - 227.
- Bafana, A., Chakrabarti, T., Muthal, P. and Kanade, G. (2009). Detoxification of Benzidine-Based Azo Dye by *E. gallinarum*: Time-Course Study. *Ecotoxicology Environment Safety*, 72, 960.
- Balamurugan, B., Thirumarimurugan, M. and Kannadasan, T. (2011). Anaerobic degradation of textile dye bath effluent using *Halomonas* sp. *Bioresource Technology*, 102(10):6365–6369.
- Barsing, P., Tiwari, A., Joshi, T. and Garg, S.(2011). Application of a novel bacterial consortium for mineralization of sulphonated aromatic amines. *Bioresource Technology*, 102: 765 - 771.
- Bay, H. H., Lim, C. K., Kee, T. C., Ware, I., Chan, G. F., Shahir, S. and Ibrahim, Z. (2013). Decolourisation of acid orange 7 recalcitrant auto oxidation coloured by-products using an acclimatised mixed bacterial culture. *Environmental Science and Pollution Research*, 1-16.
- Blaga, A., Tatyana, A., Lilyana, S. and Sava, M. (2008). Temperature Effect on Bacterial Azo Bond Reduction Kinetics: An Arrhenius Plot Analysis, *Biodegradation*, 19, 387.
- Brás R., Ferra I.A., Pinheiro H.M. and Gonçalves I.C. (2001). Batch tests for assessing decolourisation of azo dyes by methanogenic and mixed cultures. *Journal of Biotechnology*, 89(2), 155-162.
- Bose, P. and Anitha, R. (2015). Decolourization of Textile Dyes using Bacterial Consortium. *Journal of Plant and Biotechnology*, 53: 33-37.
- Bonakdarpour, B., Vyrides, I. and Stuckey, D.C. (2011). Comparison of the performance of one stage and two stage sequential Anaerobic-aerobic biological processes for the treatment of reactive-azo-dye-containing

- synthetic wastewaters. *International Biodeterioration and Biodegradation*, 65: 591 - 599.
- Carliell, C.M., Barclay, S.J., Naidoo, N., Buckley, C.A., Mulholland, D.A. and Senior, E. (1995). Microbial decolourisation of a reactive azo dye under anaerobic conditions. *Water SA*, 21(1): 61 – 69.
- Carneiro, P.A., Nogueira, R.F.P. and Zanoni, M.V.B. (2007). Homogeneous photodegradation of C.I. Reactive Blue 4 using a Photo-Fenton process under artificial and solar irradiation. *Dyes and Pigments*, 74: 127 -132.
- Cao, Y., Hu, Y., Sun, J. and Hou, B. (2010). Explore various co-substrates for simultaneous electricity generation and Congo red degradation in air-cathode single-chamber microbial fuel cell. *Bioelectrochemistry*, 79:71-6.
- Chan, G. F., Gan, H. M., and Rashid, N. A. A. (2012a). Genome sequence of *Citrobacter* sp. strain A1, a dye-degrading bacterium. *Journal of Bacteriology*, 194(19), 5485-5486.
- Chan, G. F., Gan, H. M., and Rashid, N. A. A. (2012b). Genome sequence of *Enterococcus* sp. strain C1, an Azo dye decolorizer. *Journal of Bacteriology*, 194(20), 5716-5717.
- Chan, G. F., Rashid, N. A. A., Chua, L. S., Nasiri, R., and Ikubar, M. R. M. (2012c). Communal microaerophilic–aerobic biodegradation of Amaranth by novel NAR-II bacterial consortium. *Bioresource Technology*, 105, 48-59.
- Chang, S.J. and Lin, Y.C. (2001). Decolorization kinetics of recombinant *Escherichia coli* strain harboring azo dye decolorization determinants for *Rhodococcus* sp. *Biotechnology Letters*, 23: 631 - 636.
- Chang, J. S. and Kuo, T.S. (2000). Kinetics of Bacterial Decolorization of Azo Dye with *Escherichia coli* NO3. *Bioresource Technology*, 75, 107.
- Chae, K.J., Choi, M.J., Kim, K.Y., Ajayi, F.F., Park, W., Kim, C.W. and Kim, I.S. (2010). Methanogenesis control by employing various environmental stress conditions in two-chambered microbial fuel cells. *Bioresource Technology*, 101, 5350–5357.
- Chen, Y.B., Zhang, M.M., Chang, C.T., Ding, Y., Lin, K.L., Chiou, C.S., Hsueh, C.C. and Xu, H. (2010). Assessment upon azo dye decolorization and bioelectricity generation by *Proteus hauseri*. *Bioresource Technology*, 101:4737-41.

- Chen, K. C., Wu, J. Y., Liou, D. J., and Hwang, S. C. J. (2003). Decolorization of the textile dyes by newly isolated bacterial strains. *Journal of Biotechnology*, 101(1), 57-68.
- Cui, D., Li, G., Zhao, M., & Han, S. (2014). Decolourization of azo dyes by a newly isolated *Klebsiella* sp. strain Y3, and effects of various factors on biodegradation. *Biotechnology, biotechnological equipment*, 28(3), 478-486.
- Cui, D, Li G, Zhao D, Gu X, Wang C, Zhao M. (2012). Microbial community structures in mixed bacterial consortia for azo dye treatment under aerobic and anaerobic conditions. *Journal of Hazard Materials*, 221-222:185-92.
- Daizong, C., Guofang, L., Dan, Z., Xiaoxu, G., Chunlei W. and Min, Z. (2012). Microbial community structures in mixed bacterial consortia for azo dye treatment under aerobic and anaerobic conditions. *Journal of Hazardous Materials*, Volumes 221–222, Pages 185-192, 0304-3894
- del Campo, A. G., Cañizares, P., Lobato, J., Rodrigo, M., and Morales, F. F. (2014). Effects of External Resistance on Microbial Fuel Cell's Performance. *In Environment, Energy and Climate Change II*, pp. 175-197.
- Dos Santos, A.B., Cervantes, F.J., Yaya-Beas, R.E., Van Lier, J.B. (2003). Effect of redox mediator, AQDS, on the decolourisation of a reactive azo dye containing triazine group in a thermophilic anaerobic EGSB reactor. *Enzyme and Microbial Technology*, 33: 942 - 51.
- Dos Santos, A.B., Bisschops, I.A.E., Cervantes, F.J. and van Lier, J.B. (2004). Effect of different redox mediators during thermophilic azo dye reduction by anaerobic granular sludge and comparative study between mesophilic (30°C) and thermophilic (55°C) treatments for decolourisation of textile wastewaters. *Chemosphere*, 55: 1149 - 1157.
- Dos Santos, A.B., Cervantes, F.J. and Lier van, J.B.(2007). Review paper on current technologies for decolourisation of textile wastewaters: Perspectives for anaerobic biotechnology. *Bioresource Technology*, 98: 2369 - 2385.
- Elisangela, F., Andrea, Z., Fabio, G.F., Cristiano, R.M., Regina, L.D. and Artu, C.P. (2009). Biodegradation of textile azo dyes by a facultative *Staphylococcus arlettae* strain VN-11 using a sequential microaerophilic/aerobic process. *International Biodeterioration & Biodegradation*, Volume 63, Issue 3, Pages 280-288.

- El-Kabbany, F., Taha, S., & Hafez, M. (2010). IR spectroscopic analysis of polymorphism in diphenyl carbazide. *The Journal of American Science*, 6, 263–270.
- Evans, A., Strezov, V., and Evans, T. J. (2012). Assessment of utility energy storage options for increased renewable energy penetration. *Renewable and Sustainable Energy Reviews*, 16(6), 4141-4147.
- Fang, Z., Song, H.L., Cang, N. and Li, X.N. (2015). Electricity production from Azo dye wastewater using a microbial fuel cell coupled constructed wetland operating under different operating conditions. *Biosensor Bioelectron*, 68:135–141.
- Fanchiang, J. M., & Tseng, D. H. (2009). Degradation of anthraquinone dye C.I. Reactive blue 19 in aqueous solution by ozonation. *Chemosphere*, 77, 214–221.
- Forato, L. A., Bernardes Filho, R., & Colnago, L. A. (1998). Estudo de métodos de aumento de resolução de espectros de FTIR para análise de estruturas secundárias de proteínas. *Quimica Nova*, 21, 146–150.
- Gan, H.M., Shahir, S., Ibrahim, Z. and Yahya, A. (2011). Biodegradation of 4-aminobenzenesulfonate by *Ralstonia* sp. PBA and *Hydrogenophaga* sp. PBC isolated from textile wastewater treatment plant. *Chemosphere* 82: 507 - 513.
- Ghodake, G., Jadhav, U., Tamboli, D., Kagalkar, A. and Govindwar, S. (2011). Decolorization of textile dyes and degradation of mono-azo dye Amaranth by *Acinetobacter calcoaceticus* NCIM 2890. *Indian Journal of Microbiology* 51(4): 501 - 508.
- Guadie, A., Tizazu, S., Melese, M., Guo, W., Ngo, H. H., & Xia, S. (2017). Biodecolorization of textile azo dye using *Bacillus* sp. strain CH12 isolated from alkaline lake. *Biotechnology reports (Amsterdam, Netherlands)*, 15, 92–100.
- Haroun, M., and Idris, A. (2009). Treatment of textile wastewater with an anaerobic fluidized bed reactor, *Desalination*. 237(1): 357-366.
- Hsueh, C.C., Chen, B.Y. and Yen, C.Y. (2009). Understanding effects of chemical structure on azo dye decolorization characteristics by *Aeromonas hydrophila*. *Journal of Hazard Materials*, 167:995-1001.

- Hong, S. W., Chang, I. S., Choi, Y. S., and Chung, T. H. (2009). Experimental evaluation of influential factors for electricity harvesting from sediment using microbial fuel cell. *Bioresource Technology*, 100(12), 3029-3035.
- Hu, T. L. (2001). Kinetics of Azoreductase and Assessment of Toxicity of Metabolic Products from Azo Dyes by *Pseudomonas luteola*. *Water Science Technology*, 43, 261.
- Hu, C., Yu, J.C., Hao, Z. and Wong, P.K. (2003). Photocatalytic degradation of triazine-containing azo dyes in aqueous TiO₂ suspensions. *Appl Catalysis B Environment*, 42:47–55.
- Ilamathi, R. and Jayapriya, J. (2017). Microbial fuel cell for dye decolourization. *Environmental Chemistry Letters*, 16(1), 239-250.
- Isik, M. and Sponza, D.T. (2003). Effect of oxygen on decolorization of azo dyes by *Escherichia coli* and *Pseudomonas* sp. and fate of aromatic amines. *Process Biochemistry*, 38: 1183 - 1192.
- Isik, M. and Sponza, D.T. (2007). Fate and toxicity of azo dye metabolites under batch long-term anaerobic incubations. *Enzyme and Microbial Technology*, 40:934 - 939.
- Jadhav, G.S. and Ghangrekar, M.M. (2009). Performance of microbial fuel cell subjected to variation in pH, temperature, external load and substrate concentration. *Bioresource Technology*, 100,717–723.
- Jin, X.C., Liu, G.Q., Xu, Z.H. and Tao, W.Y. (2007). Decolorization of a dye industry effluent by *Aspergillus fumigatus* XC6. *Apply Microbiology Biotechnology*, 74:239-43.
- Juárez-Hernández, J., Zavala-Soto, M. E., Bibbins-Martínez, M., Delgado-Macuil, R., Díaz-Godínez, G., & Rojas-Lopes, M. (2008). FTIR spectroscopy applied in remazol blue dye oxidation by laccases. *AIP Conference Proceedings*, 992, 1253–1257.
- Kamboh, A.A., Rajput, N., Rajput, I.R., Khaskheli, M. and Khaskheli, G.B. (2009). Biochemical Properties of Bacterial Contaminants Isolated from Livestock Vaccines. *Pakistan Journal of Nutrition*, 8 (5): 578-581, 1680-5194.
- Karim, E., Dhar, K and Hossain, M.T. (2018). Decolorization of Textile Reactive Dyes by Bacterial Monoculture and Consortium Screened from Textile

- Dyeing Effluent, *Journal of Genetic Engineering and Biotechnology*, Volume 16, Issue 2, Pages 375-380
- Kalyani, D. C., Telke, A. A., Dhanve, R. S., and Jadhav, J. P. (2009). Ecofriendly biodegradation and detoxification of Reactive Red 2 textile dye by newly isolated *Pseudomonas* sp. SUK1. *Journal of Hazardous Materials*, 163(2), 735-742.
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S. and Chimni, S.S. (2005). Decolorization of various azo dyes by bacterial consortium, *Dyes and Pigments*, Volume 67, Issue 1, Pages 55-61, 0143-7208
- Khalid, A., Arshad, M. and Crowley, D.E. (2008). Decolorization of azo dyes by *Shewanella* sp. under saline conditions. *Applied Microbiology and Biotechnology* 79: 1053 - 1059.
- Khalid, A., Arshad, M. and Crowley, D.E. (2008a). Accelerated decolorization of structurally different azo dyes by newly isolated bacterial strains. *Environmental Biotechnology*, 78: 361 - 369.
- Khalid, A., Arshad, M. and Crowley, D. (2009). Biodegradation potential of pure and mixed bacterial cultures for removal of 4-nitroaniline from textile dye wastewater. *Water Research*, 43(3): 1110 - 1116.
- Khan, R. and Banarjee, U.C. (2010). Decolorization of azo dyes by immobilized bacteria. *Biodegradation of Azo Dyes*, 9: 73 - 84.
- Knapp, J.S. and Newby, P.S. (1995). The microbiological decolorization of an industrial effluent containing a diazo-linked chromophore. *Water Research* 29:1807 - 1809.
- Khouni, I., Marrot, B. and Amar, R.B. (2012). Treatment of reconstituted textile wastewater containing a reactive dye in an aerobic sequencing batch reactor using a novel bacterial consortium. *Sep Purification Technology*, 87:110-9.
- Krishnamoorthy, R & Jose, Polpass & Ranjith, M & Anandham, Rangasamy & Suganya, K & Prabhakaran, J & Thiyageshwari, S & Johnson, Jijo & N O, Gopal & Kumutha, Karunanandham. (2017). Decolourisation and degradation of azo dyes by mixed fungal culture consisted of *Dichotomomyces cejpilii* MRCH 1-2 and *Phoma tropica* MRCH 1-3. *Journal of Environmental Chemical Engineering*. 6. 10.1016

- Kuberan, T., Anburaj, J., Sundaravadivelan, C. and Kumar, P. (2011). Biodegradation of azo dye by *Listeria* sp. *International Journal of Environmental Science*, 1:1760-70.
- Kumar, K., Devi, S.S., Krishnamurthi, K., Gampawar, S., Mishra, N., Pandya, G.H and Chakrabarti, T. (2005). Decolorisation, biodegradation and detoxification of benzidine based azo dye. *Bioresource Technology* 97(3): 407 - 413.
- Kumar, G., Tripathi, M., Singh, S. K., and Tiwari, J. K. (2012). Biodecolorization of textile dye effluent by *Pseudomonas putida* SKG-1 (MTCC 10510) under the conditions optimized for monoazo dye orange II color removal in simulated minimal salt medium. *International Biodeterioration and Biodegradation*, 74, 24-35.
- Kumar, A., Katuri, K., Lens, P., and Leech, D. (2012). Does bioelectrochemical cell configuration and anode potential affect biofilm response? *Biochemical Social Transactions*, 40(6), 1308-1314.
- Kumar, K., Dastidara, M.G. and Sreekrishnan, T.R. (2009). Effect of process parameters on aerobic decolourization of reactive azo dye using mixed culture. *World AcadSci Eng Technology*, 58:962-5.
- Kodam, K.M. and Gawai, K.R. (2006). Decolorisation of Reactive Red 11 and 152 azo dyes under aerobic conditions. *Indian Journal of Biotechnology*, 5: 422 - 424.
- Kodam, K.M., Soojhawon, I., Lokhande, P.D. and Gawai, K.R.(2005). Microbial decolorization of reactive azo dyes under aerobic conditions. *Word Journal of Microbiology Biotechnology*, 21: 367-370.
- Lai, C. Y., Wu, C. H., Meng, C. T., and Lin, C. W. (2017). Decolorization of azo dye and generation of electricity by microbial fuel cell with laccase- producing white-rot fungus on cathode. *Applied Energy*, 188, 392-398.
- Li, G.N., Xia, X.J., Zhao, H.H., Sendegeya, P. and Zhu, Y. (2015). Identification and Characterization of *Bacillus cereus* SW7-1 in *Bombyx mori*. *Journal of Insect Science*, 15(1): 136.
- Li, W. W., Sheng, G. P., Liu, X. W., and Yu, H. Q. (2011). Recent advances in the separators for microbial fuel cells. *Bioresource Technology*, 102(1), 244-252.

- Li, W.-W., Yu, H.-Q., and He, Z. (2014). Towards sustainable wastewater treatment by using microbial fuel cells-centered technologies. *Energy and Environmental Science*, 7(3), 911-924.
- Li, X. M., Cheng, K. Y., Selvam, A., and Wong, J. W. (2013). Bioelectricity production from acidic food waste leachate using microbial fuel cells: effect of microbial inocula. *Process Biochemistry*, 48(2), 283-288.
- Liao, C.S., Hung, H.H. and Chao, S.L.(2012). Decolorization of azo dye reactive black B by *Bacillus cereus* strain HJ-1. *Chemosphere*, Volume 90, Issue 7, P-2109-2114.
- Liu, R.H., Guo, P., Sheng, G.P., Sun, M., Zang, G.L., Li, W.W. and Tong, Z.H. (2011). Enhanced reductive degradation of methyl orange in a microbial fuel cell through cathode modification with redox mediators. *Apply Microbiology Biotechnology*, 89:201-8.
- Lin, J., X. Zhang, Z. Li, and L. Lei. (2010). Biodegradation of Reactive Blue 13 in a Two Stage Anaerobic/Aerobic Fluidized Beds System with a *Pseudomonas* sp. Isolate. *Bioresource Technology*, 101, 34.
- Liu, G., Zhou, J., Lv, H., Xiang, X., Wang, J. and Zhou, M. (2007). Azoreductase from *Rhodobacter sphaeroides* AS1.1737 is a flavodoxin that also functions as nitroreductase and flavin mononucleotide reductase. *Apply Microbiology Biotechnology* (76) pp 1271-1279.
- Liu, G., Zhou, J., Jin, R., Zhou, M., Wang, J. and Lu, H. (2008) Enhancing survival of *Escherichia coli* by expression of azoreductase AZR possessing quinone reductase activity. *Apply Microbiology Biotechnology* (80) pp 409-416.
- Liu, Y.H., Ye, M., Lu, Y., Zhang, X. and Li, G. (2011). Improving the decolorization for textile dyes of a metagenome-derived alkaline laccase by directed evolution. *Apply Microbiology Biotechnology* (91) pp 667-675.
- Logan, B. E., Hamelers, B., Rozendal, R., Schröder, U., Keller, J., Freguia, S., and Rabaey, K. (2006). Microbial fuel cells: methodology and technology. *Environmental Science and Technology*, 40(17), 5181-5192.
- Lyon, D.Y., Buret, F., Vogel, T.M. and Monier, J.M. (2010) Is resistance futile? Changing external resistance does not improve microbial fuel cell performance. *Bioelectrochemistry*, 78:2-7.

- Mahmood, R., Sharif, F., Ali, S., & Hayyat, M. U. (2015). Enhancing the decolorizing and degradation ability of bacterial consortium isolated from textile effluent affected area and its application on seed germination. *The Scientific World Journal*, 2015, 628195.
- Maier, J. Kandelbauer, A., Erlacher, A., Cavaco - Paulo, A. and Gubitzi, M. (2004). A new alkali - thermostable azoreductase from *Bacillus* sp. strain SF. *Applied and Environmental Microbiology*, 70(2): 837 - 844.
- Mathew, S. and Madamwar, D. (2004). Decolorization of ranocid fast blue dye by bacterial consortium SV5. *Applied Biochemistry Biotechnology*, 118: 371-381.
- Malaysia's Environmental Law, ENVIRONMENTAL QUALITY ACT, 1974, the Malaysia Environmental Quality (Industrial Effluents) Regulations 2009 (PU(A) 434).
- Mane, U.V. Gurav, P.N., Deshmukh, A.M. and Govindwar, S.P. (2008). Degradation of textile dye reactive navy - blue Rx (Reactive blue-59) by an isolated Actinomycete *Streptomyces krainskii* SUK - 5. *Malaysian Journal of Microbiology*, 4(2): 1 - 5.
- Menicucci J, Beyenal H, Marsili E, Veluchamy RA, Demir G, and Lewandowski Z (2006). Procedure for determining maximum sustainable power generated by microbial fuel cell. *Environmental Sciences and Technology*, 40,1062–1068.
- McMullan, G., Meehan, C., Conneely, A., Kirby, N., Robinson, T., Nigam, P., Banat, I.M., Marchant, R. and Smyth, W.F. (2001). Microbial decolourisation and degradation of textile dyes. *Applied Microbiology and Biotechnology*, 56: 81-87.
- Mohamed, A. R., and Lee, K. T. (2006). Energy for sustainable development in Malaysia: Energy policy and alternative energy. *Energy Policy*, 34(15), 2388-2397.
- Mohan, S. V., Babu, P. S., and Srikanth, S. (2013). Azo dye remediation in periodic discontinuous batch mode operation: evaluation of metabolic shifts of the biocatalyst under aerobic, anaerobic and anoxic conditions. *Separation and Purification Technology*, 118, 196-208.
- Mohan, S. V., Saravanan, R., Raghavulu, S. V., Mohanakrishna, G., and Sarma, P. N. (2008). Bioelectricity production from wastewater treatment in dual

- chambered microbial fuel cell (MFC) using selectively enriched mixed microflora: effect of catholyte. *Bioresource Technology*, 99(3), 596-603.
- Mohan, Y., and Das, D. (2009). Effect of ionic strength, cation exchanger and inoculum age on the performance of microbial fuel cells. *International Journal of Hydrogen Energy*, 34(17), 7542-7546.
- Mohana, S.V., Shrivastav, S., Divecha, J. and Madamwar, D. (2008). Response surface methodology for optimization of medium for decolorization of textile dye Direct Black 22 by a novel bacterial consortium. *Bioresource Technology*, 99:562-9.
- Moosvi, S., Keharia, H. and Madamwar, D. (2005). Decolourization of textile dye Reactive Violet 5 by a newly isolated bacterial consortium RVM 11.1. *World Journal of Microbiology and Biotechnology* 21: 667 - 672.
- Moosvi, S., Kher, X. and Madamwar, D. (2007). Isolation, characterization and decolorization of textile dyes by a mixed bacterial consortium JW-2. *Dyes and Pigments* 74(3): 723 - 729.
- Moutaouakkil, A., Zeroual, Y., Dzayri, F.Z., Talbi, M., Lee, K. and Blaghen, M. (2003). Bacterial decolorization of the azo dye methyl red by *Enterobacter agglomerans*. *Annals of Microbiology*, 53: 161 - 169.
- Moutaouakkil, A., Zeroual, Y., Dzayri, F.Z., Talbi, M., Lee, K. and Blaghen, M. (2003a). Purification and partial characterization of azoreductase from *Enterobacter agglomerans*. *Archives of Biochemistry and Biophysics* 413: 139 - 146.
- Nam, J.Y., and Logan, B. E. (2011). Enhanced Hydrogen Generation using a Saline Catholyte in a Two Chamber Microbial Electrolysis Cell. *International Journal Of Hydrogen Energy*, 36, 5105-15110.
- Niessen, J., Schröder, U., Rosenbaum, M., and Scholz, F. (2004). Fluorinated polyanilines as superior materials for electrocatalytic anodes in bacterial fuel cells. *Electrochemistry Communications*, 6(6), 571-575.
- Nigam, P., Mc Mullan, G., Banat, I.M. and Marchant, R. (1996). Decolourisation of effluent from the textile industry by a microbial consortium. *Biotechnology letters* 18(1): 117 - 120.
- Nor, M. H. M., Mubarak, M. F. M., Elmi, H. S. A., Ibrahim, N., Wahab, M. F. A., and Ibrahim, Z. (2015). Bioelectricity generation in microbial fuel cell using

- natural microflora and isolated pure culture bacteria from anaerobic palm oil mill effluent sludge. *Bioresource Technology*, 190, 458-465.
- Oturkar, C.C., Nemade, H.N., Mulik, P.M., Patole, M.S., Hawaldar, R.R. and Gawai, K.R. (2011). Mechanistic investigation of decolorization and degradation of Reactive Red 120 by *Bacillus lentus* BI377. *Bioresource Technology* 102: 758 - 764.
- O' Neill, C., Hawkes, F. R., Hawkes, D. L., Lourenco, N. D., Pinheiro, H. M. and Delee, W. (1999). Color in textile effluents-sources, measurement, discharge consents and simulation: a review. *Journal of Chemical Technology and Biotechnology*, 74: 1009 - 1018.
- O'Neill, C., Lopez, A., Esteves, S., Hawkes, F.R., Hawkes, D.L. and Wilcox, S. (2000). Azo-dye degradation in an anaerobic-aerobic treatment system. operating on simulated textile effluent. *Applied Microbiology and Biotechnology*, 53: 249 - 254.
- Ong, S.A., Toorisaka, E., Hirata, M. and Hano, T. (2005). Treatment of azo dye Orange II in a sequential anaerobic and anerobic sequencing batch ractor system. *Environmental Chemistry Letters*, 2(4): 203 - 207.
- Ong, S.A., Toorisaka, E., Hirata, M. and Hano, T. (2005a). Treatment of azo dye Orange II in aerobic and anaerobic-SBR systems. *Process Biochemistry*, 40: 2907 - 2914.
- Patel, S.S., Chauhan, H.C., Patel, A.C., Shrimali, M.D., Patel, K.B., Prajapati, B.I., Kala, J.K., Patel, M.G., Manish, R. and Patel, M.A. (2017). Isolation and Identification of *Klebsiella pneumoniae* from Sheep-Case Report. *International J.Curr.Microbiol.App.Sci.* 6(5): 331-334.
- Pang, Y. L., and Abdullah, A. Z. (2013). Current status of textile industry w astewater management and research progress in Malaysia: a review. *CLEAN–Soil, Air, Water*, 41(8), 751-764.
- Pearce, C.I., Lloyd, J.R. and Guthrie, J.T. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments*, 58(3): 179 - 196.
- Pearce, C.I., Lloyd, J.R. and Guthrie, J.T. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments*, 58(3): 179 - 196.

- Polunin, K. E., Sokolova, N. P., Gorbuno, A. M., Bulgakova, R. A., & Polunina, I. A. (2008). FTIR spectroscopic studies of interactions of stilbenes with silicon dioxide. *Protection of Metals*, 44, 352–357.
- Popli, S. and Patel, U.D. (2015). Destruction of azo dyes by anaerobic–aerobic sequential biological treatment: a review. *International Journal of Environmental Science and Technology*, 12(1):405–420.
- Pricelius, S., Held, C., Sollner, S., Deller, S., Murkovic, M. and Ullrich, R. (2007). Enzymatic reduction and oxidation of fibre-bound azo-dyes. *Enzyme Microbiology Technology*, 40:1732-8.
- Pourbabae, A.A. and Malekzadeh, F. (2005). Decolorization of Methyl Orange (as a model azo dye) by the newly discovered *Bacillus Sp.* *Iranian Journal of Chemistry and Chemical Engineering* 24(3): 41 - 45.
- Pourbabae, A.A., Malekzadeh, F., Sarbolouki, M.N. and Najafi, F. (2006). Aerobic decolorization and detoxification of a disperse dye in textile effluent by a new isolate of *Bacillus sp.* *Biotechnology and Bioengineering* 93(4): 631 - 635.
- Potter, M. C. (1910). On the Difference of Potential Due to the Vital Activity of Microorganisms. *Durham University Philosophy Society*, 3, 245-249.
- Potter, M. C. (1911). Electrical Effects Accompanying the Decomposition of Organic Compounds. *Proceedings of the Royal Society of London*, 84(571), 260-276.
- Prasad, A.S.A. and Rao, K.V.B. (2014). Aerobic biodegradation of azo dye Acid Black-24 by *Bacillus halodurans*, *Journal of Environmental Biology*, Vol. 35, 549-554.
- Rabaey, K., Boon, N., Höfte, M., and Verstraete, W. (2005b). Microbial Phenazine Production Enhances Electron Transfer in Biofuel Cells. *Environmental Science and Technology*, 39(9), 3401-3408.
- Rabaey, K., Boon, N., Siciliano, S. D., Verhaege, M., and Verstraete, W. (2004). Biofuel Cells Select for Microbial Consortia That Self-Mediate Electron Transfer. *Applied And Environmental Microbiology*, 70(9), 5373–5382.
- Rabaey, K., Butzer, S., Brown, S., Keller, J., and Rozendal, R. A. (2010). High Current Generation Coupled to Caustic Production using a Lamellar Bioelectrochemical System. *Environmental Science and Technology*, 44(11), 4315-4321.

- Rabaey, K., Lissens, G., Siciliano, S. D., and Verstraete, W. (2003). A Microbial Fuel Cell Capable of Converting Glucose to Electricity at High Rate and Efficiency. *Biotechnology Letters*, 25, 1531-1535.
- Rabaey, K., Rodriguez, J., Blackall, L. L., Keller, J., Gross, P., Batstone, D., Verstraete, W., and Neelson, K. H. (2007). Microbial Ecology Meets Electrochemistry: Electricity-Driven and Driving Communities. *The ISME Journal*, 1(1), 9-18.
- Rabaey, K., and Verstraete, W. (2005). Microbial fuel cells: novel biotechnology for energy generation. *TRENDS in Biotechnology*, 23(6), 291-298.
- Rahimnejad, M., Adhami, A., Darvari, S., Zirepour, A., and Oh, S. E. (2015). Microbial fuel cell as new technology for bioelectricity generation: a review. *Alexandria Engineering Journal*, 54(3), 745-756.
- Rajee, O. and Patterson, J. (2011). Decolorization of azo dye (Orange MR) by an autochthonous bacterium, *Micrococcus* sp. DBS 2. *Indian Journal of Microbiology*, 51(2): 159 - 163.
- Rajaguru, P., Kalaiselvi, K., Palanivel, M. and Subburam, V. (2000). Biodegradation of azo dyes in a sequential anaerobic-aerobic system. *Applied Microbiology and Biotechnology*, 54: 268 - 273.
- Rai, H., Bhattacharya, M., Singh, J., Bansal, T.K., Vats, P., and Banerjee, U.C. (2005). Removal of Dyes from the Effluent of Textile and Dyestuff Manufacturing Industry: A Review of Emerging Techniques with Reference to Biological Treatment, *Crit. Rev. Environmental. Science Technology*, 35, 219.
- Rashidi, H.R., Sulaiman, N.M.N., Hashim, N.A. and Hassan, C.R.C. (2013). Synthetic batik wastewater pretreatment progress by using physical treatment. *Malaysia Advanced Materials Research*, Vol. 627.,pp 394-398.
- Rehorek, A. and Plum, A. (2006). Online LC-MS-MS process monitoring for optimization of biological treatment of wastewater containing azo dye concentrates. *Analytical and Bioanalytical Chemistry*, 384: 1123 - 1128.
- Sandhya, S. (2010). Biodegradation of azo dyes under anaerobic condition: Role of azoreductase. *Biodegradation of Azo dyes*. H. Atacag Erkurt (ed.). *The Handbook of Environmental Chemistry*, 9: 39 - 57.

- Sani, R.K. and Banerjee, U.C. (1999). Decolorization of triphenylmethane dyes and textile and dye-stuff effluent by *Kurthia sp.* *Enzyme Microbiology Technology*, 24, pp. 433-437.
- Santos, P. and Corso, C.R. (2014). Comparative Analysis of Azo Dye Biodegradation by *Aspergillus oryzae* and *Phanerochaete chrysosporium*. *Water Air and Soil Pollution*, 2026-6.
- Saratale, R. G., Saratale, G. D., Chang, J. S., and Govindwar, S. P. (2011). Bacterial decolorization and degradation of azo dyes: a review. *Journal of the Taiwan Institute of Chemical Engineers*, 42(1), 138- 157.
- Saratale, R. G., Saratale, G. D., Chang, J. S., and Govindwar, S. P. (2009). Ecofriendly degradation of sulfonated diazo dye CI Reactive Green 19A using *Micrococcus glutamicus* NCIM-2168. *Bioresource Technology*, 100(17), 3897-3905.
- Sarayu, K. and Sandhya, S. (2010). Aerobic Biodegradation Pathway for Remazol Orange by *Pseudomonas aeruginosa*. *Apply Biochemical Biotechnology*, 160, 1241.
- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of azo dyes. *Applied Microbiology and Biotechnology*, 56(1-2): 69-80.
- Solanki, K., Subramanian, S., and Basu, S. (2013). Microbial fuel cells for azo dye treatment with electricity generation: a review. *Bioresource Technology*, 131, 564-571.
- Sarayu, K., and Sandhya, S. (2012). Current technologies for biological treatment of textile wastewater—a review. *Applied Biochemistry and Biotechnology*, 167(3), 645-661.
- Sarkar P., Rai A. R., Ghosh S. (2013). Degradation of aromatic petroleum hydrocarbons (BTEX) by a solvent tolerant bacterial consortium. *Journal of Urban Environmental Engineering*, 7 274–279.
- Shah, M.P., Patel, K.A., Nair, S.S., and Darji, A.M. (2013). Potential Effect of Two *Bacillus* spp on Decolorization of Azo dye. *J Bioremed Biodeg* 4: 199.
- Shailesh, R. D. and Riddhi, H. D. (2009). Isolation and characterization of *Bacillus thuringiensis* for Acid red 119 dye decolourisation. *Bioresource Technology*, Volume 100, Issue 1, Pages 249-253; 0960-8524.

- Sharma, S., Munjal, A. and Gupta, S. (2011). Comparative studies on decolorization of textile Azo dyes by different bacterial consortia and pure bacterial isolate, *Journal of Pharmacy Research*,4(9),3180-3183
- Sheth, N.T. and Dave, S.R.(2009). Optimisation for enhanced decolourization and degradation of Reactive Red BS C.I. 111 by *Pseudomonas aeruginosa* NGKCTS. *Journal Of Microbiology*, 0923-9820, 1572-9729.
- Scott, K., and Shukla, A. K. (2004). Polymer electrolyte membrane fuel cells: Principles and advances. *Reviews in Environmental Science and Bio/Technology*, 3(3), 273-280.
- Shah, K. (2014). Biodegradations of azo dye compounds. *International Research Journal of Biochemistry and Biotechnology* ,Vol. 1(2), pp. 005-013
- Solís, M., Solis, A., Perezb, H.I., Manjarrezb, N. and Floresa, M. (2012). Microbial decolouration of azo dyes: A review. *Process Biochemistry*, 47, pp. 1723–1748
- Solanki, K., Subramanian, S., and Basu, S. (2013). Microbial fuel cells for azo dye treatment with electricity generation: a review. *Bioresource Technology*, 131, 564-571.
- Song, T.S., Yan, Z.S., Zhao, Z.W. and Jiang, H.L. (2010). Removal of organic matter in freshwater sediment by microbial fuel cells at various external resistances. *Journal of Chemical Technology and Biotechnology*, 85, 1489–1493.
- Stolz, A. (2001). Basic and applied aspects in the microbial degradation of azo dyes. *Applied Microbiology and Biotechnology*, 56(1 – 2): 69 – 80.
- Slokar, Y.M. and Le Marechal, A.M. (1998). Methods of decoloration of textile wastewater. *Dyes and Pigments*, 37(4): 355 – 356.
- Sun, J., Hu, Y., Bi, Y.Z., and Cao, Y.Q. (2009a). Simultaneous decolorization of azo dye and bioelectricity generation using a microfiltration membrane air-cathode single-chamber microbial fuel cell. *Bioresource Technology*, 100:3185–3192.
- Sun, J., Hu, Y., Bi, Y.Z., and Cao, Y.Q. (2009b). Improved performance of air-cathode single-chamber microbial fuel cell for wastewater treatment using microfiltration membranes and multiple sludge inoculation. *Journal of Power Sources*, 187:471–479.
- Sun, J., Hu, Y. and Hou, B. (2011). Electrochemical characterization of the bioanode during simultaneous azo dye decolorization and bioelectricity generation in

- an air-cathode single chambered microbial fuel cell. *Electrochim Acta*, 56(19):6874–6879
- Tan, N.C.G., van Leeuwen, A., van Voorthuizen, E.M., Slenders, P., Prenafeta-Boldu, F.X., Temmink, H., Lettinga, G. and Field, J.A. (2005). Fate and biodegradability of sulfonated aromatic amines. *Biodegradation*, 16: 527 – 537.
- Telke, A.A., Kalyani, D.C., Jadhav, J.P. and Govindwar, S.P. (2008). Kinetics and mechanism of reactive red 141 degradation by a bacterial isolate *Rhizobium radiobacter* MTCC 8161. *Acta Chimica Slovenica*, 55: 320 – 329.
- Telke, A.A., Kalyani, D.C., Dawkar, V.V. and Govindwar, S.P. (2009). Influence of organic and inorganic compounds on oxidoreductive decolorization of sulfonated azo dye C.I. Reactive Orange 16. *Journal of Hazardous Materials*, 172(1): 298 – 309.
- Telke, A.A., Joshi, S.M., Jadhav, S.U., Tamboli, D.P. and Govindwar, S.P. (2010). Decolorization and detoxification of Congo Red and textile industry effluent by an isolated bacterium *Pseudomonas* sp. SU-EBT. *Biodegradation*, (21): 283 – 296.
- Thung, W.E., Ong, S.A., Ho, L.N., Wong, Y.S., Ridwan, F., Oon, Y.L., Oon, Y.S. and Lehl, H.K. (2015). A highly efficient single chambered up-flow membrane-less microbial fuel cell for treatment of azo dye Acid Orange 7-containing wastewater. *Bioresource Technology*, 197:284–288.
- Tripathi, A. and Srivastava, S.K. (2011). Ecofriendly Treatment of Azo Dyes: Biodecolorization using Bacterial Strains. *International Journal of Bioscience, Biochemistry and Bioinformatics*, Vol. 1, No. 1
- Tony, B.D., Goyal, D., Khanna, S. (2009a). Decolorization of Textile Azo Dyes by Aerobic Bacterial Consortium. *International Biodeter. Biodegradation*, 63 , p. 462
- Tony, B.D., Goyal, D., Khanna, S. (2009b). Decolorization of Direct Red 28 by Mixed Bacterial Culture in an Up-Flow Immobilized Bioreactor. *Journal of Indian Microbiology Biotechnology*, 36, p. 955
- Vandevivere, P. C., Bianchi, R., and Verstraete, W. (1998). Treatment and Reuse of Wastewater from the Textile Wet-processing Industry: Review of Emerging Technologies. *Journal of Chemical Technology Biotechnology*, 72, 289 (1998).

- Venkata Mohan, S., Suresh Babu, P. and Srikanth, S. (2013). Azo dye remediation in periodic discontinuous batch mode operation: evaluation of metabolic shifts of the biocatalyst under aerobic, anaerobic and anoxic conditions. *Separation and Purification Technology*.
- Venugopal, R., Tollefson, L., Hyman, F. N., Timbo, B. and Joyce, R. E. (1996). Recalls of Foods and Cosmetics by the US Food and Drug Administration. *Journal of Food Protocol*, 59 (8), 876–880.
- Vijaykumar, M.H., Vaishampayan, P.A., Shouche, Y.S. and Karegoudar, T.B. (2007). Decolourization of naphthalene-containing sulfonated azo dyes by *Kerstersia* sp. strain VKY1. *Enzyme and Microbial Technology*, 40: 204 – 211.
- Waghmode, T.R., Kurade, M.B. and Govindwar, S.P. (2011). Time dependent degradation of mixture of structurally different azo and non azo dyes by using *Galactomyces geotrichum* MTCC 1360. *International Biodeter Biodegradation*, 65:479–86
- Wong, P.K. and Yuen, P.Y. (1996) Decolorization and biode-gradation of methyl red by *Klebsiella pneumoniae* RS-13. *Water Research*, 30, 1736–1744
- Wong, P.K. and Yuen, P.Y. (1998). Decolourization and biodegradation of N,N-dimethyl-p-phenylenediamine by *Klebsiella pneumoniae* RS-13 and *Acetobacter liquefaciens* S-1. *Journal of Applied Microbiology*, 85, 79–87
- Wharfe, E. S., Jarvis, R. M., Winder, C. L., Whiteley, A. S., & Goodacre, R. (2010). Fourier transform infrared spectroscopy as a metabolite fingerprinting tool for monitoring the phenotypic changes in complex bacterial communities capable of degrading phenol. *Environmental Microbiology*, 12, 3253–3263
- Xu, H., Heinze, T.M., Chen, S., Cerniglia, C.E. and Chen, H. (2007). Anaerobic metabolism of 1-amino-2-naphthol-based azo dyes (Sudan Dyes) by human intestinal microflora. *Applied and Environmental Microbiology*, 73(23): 7759 – 7762.
- Xu, M.Y., Guo, J. and Sun, G.P. (2007). Biodegradation of textile azo dye by *Shewanella decolorationis* S12 under microaerophilic conditions. *Applied Microbiology and Biotechnology*, 76: 719 – 726.

- Yang, Q., Li, C., Li, H., Li, Y. and Yu, N. (2009). Degradation of synthetic reactive azo dyes and treatment of textile wastewater by a fungi consortium reactor. *Biochemical Engineering Journal*, 43:225–30.
- You, S.J. and Teng, J.Y. (2009). Anaerobic Decolorization Bacteria for the Treatment of Azo Dye in a Sequential Anaerobic and Aerobic Membrane Bioreactor. *Journal of Taiwan Institute Chemical Engineering*, 40(5), pp 500-504
- Yoo, E.S., Libra, J. and Adrian, L. (2001). Mechanism of decolorization of azo dyes in anaerobic mixed culture. *Journal of Environmental Engineering*, 127(9): 844 – 849.
- Yu, J., Wang, X., and Yue, P. (2001). Optimal Decolorization and Kinetic Modeling of Synthetic Dyes by Pseudomonas Strains, *Water Research*, 35, 3579
- Wang, Y., Zhang, J., Wang, Y., Wang, K., Wei, H. and Shen, L. (2018). Isolation and characterization of the *Bacillus cereus* BC7 strain, which is capable of zearalenone removal and intestinal flora modulation in mice, *Toxicon*, Volume 155, Pages 9-20, 0041-0101
- Zee van der, F.P., Lettinga, G. and Field, J.A. (2000). The role of (auto)catalysis in the mechanism of anaerobic azo reduction. *Water Science and Technology*, 42: 301 – 308.
- Zee van der, F.P., Lettinga, G. and Field, J.A. (2001). Azo dye decolourisation by anaerobic granular sludge. *Chemosphere*, 44: 1169 – 1176.
- Zee van der, F.P. (2002). Anaerobic azo dye reduction. *Environmental Science Technology*. 37(2): 402-408.
- Zee van der, F.P., Bisschops, I.A.E., Blanchard, V.G., Lettinga, G., and Field, J.A. (2003a). Characterization of azo reduction activity in a novel Ascomycete yeast strain. *Water Science Technology* : 97-104.
- Zee van der, F.P., Bisschops, I.A.E., Blanchard, V.G., Bouwman, R.H.M., Letting, G. and Field, J.A. (2003). The contribution of biotic and abiotic processes during azo dye reduction in anaerobic sludge. *Water Research*, 37: 3098 – 3109.
- Zee van der, F.P. and Villaverde, S. (2005). Combined anaerobic–aerobic treatment of azo dyes – A short review of bioreactor studies. *Water Research*, 39: 1425 – 1440.

- Zhang, L., Zhou, S., Zhuang, L., Li, W., Zhang, J., Lu, N. and Deng, L. (2008).
Microbial Fuel cell based on Klebsiella Pneumoniae biofilm.
Electrochemistry Communications 10, pg 1641–1643
- Zollinger, H. (1987). Azo dyes and pigments. Colour chemistry-synthesis, properties
and applications of organic Dyes and pigments. Color Chemistry, pg. 92-102.